



UNIVERSITI PUTRA MALAYSIA

***EVALUATION OF THE EFFECTIVE TIME FOR GSK-3 INHIBITION IN
EXPERIMENTAL AUTOIMMUNE ENCEPHALOMYELITIS MOUSE
MODEL BY TIDEGLUSIB (NP-12) IN PREVENTING
NEUROINFLAMMATION***

AHMED SALAM AL-ZAIDI

FPSK(m) 2019 33



**EVALUATION OF THE EFFECTIVE TIME FOR GSK-3 INHIBITION IN
EXPERIMENTAL AUTOIMMUNE ENCEPHALOMYELITIS MOUSE
MODEL BY TIDEGLUSIB (NP-12) IN PREVENTING
NEUROINFLAMMATION**

By

AHMED SALAM AL-ZAIDI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

December 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**EVALUATION OF THE EFFECTIVE TIME FOR GSK-3 INHIBITION IN
EXPERIMENTAL AUTOIMMUNE ENCEPHALOMYELITIS MOUSE
MODEL BY TIDEGLUSIB (NP-12) IN PREVENTING
NEUROINFLAMMATION**

By

AHMED SALAM AL-ZAIDI

December 2018

Chairman : Masriana Hassan, PhD
Faculty : Medicine and Health Sciences

Glycogen synthase kinase-3 (GSK-3) is an enzyme involved in various neurodegenerative and neuro-inflammatory diseases, including multiple sclerosis (MS) and its animal model of experimental autoimmune encephalomyelitis (EAE). The severity of the disease could be characterized by the degree of cellular invasion to the central nervous system (CNS) and demyelination that partly mediated by Th1 and Th17 effector cells. The present study is to investigate the efficacy of GSK-3 inhibition at a different course of treatment in ameliorating disease progression. This study also aims to evaluate the effects of GSK-3 inhibition to central neuro-invasion and demyelination. Female C57BL/6 mice were induced with myelin oligodendrocyte glycoprotein (MOG35-55) in conjunction with complete Freund's adjuvant (CFA) and pertussis toxin. Inhibition of GSK-3 is performed by the administration of NP-12, a small heterocyclic Thiadiazolidinones (TDZD), intraperitoneally during pre-EAE induction, on the same day, and post-EAE. Data revealed that NP-12 delivery during pre-EAE induction greatly protected the mice from EAE, delayed the onset of EAE symptoms by 7 days from day-14.3 ± 0.5 (in EAE mice) until day-21.3 ± 3.2 (in the treated mice). Furthermore, NP-12 treated-EAE mice had notably reduced the inflammatory cells infiltration and axonal damage (demyelination) in the spinal cord. Inhibition of GSK-3 also abrogated the production of Th1, Th9 and Th17 associated cytokines but, increases the production of IL-4 level to 1790 ± 95.0 pg/mL compare to EAE 480 ± 38.0 pg/mL and in IL-10 780 ± 22.0 pg/mL compared to EAE 410 ± 16.0 pg/mL. These data demonstrate the effectiveness of NP-12 at a different time course of administration to reduce the severity of EAE disease and protect the CNS environment from potential cellular invasion or demyelination, thus, suggests the potential use of GSK-3 inhibitor in the treatment of EAE.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENILAIAN MASA EFEKTIF BAGI PERENCATAN GSK-3 DI DALAM
EAE OLEH TIDEGLUSIB UNTUK PENCEGAHAN NEUROINFLAMASI**

Oleh

AHMED SALAM AL-ZAIDI

Disember 2018

Pengerusi : Masriana Hassan, PhD
Fakulti : Perubatan dan Sains Kesihatan

Glycogen synthase kinase-3 (GSK-3) adalah enzim yang terlibat dalam pelbagai penyakit neurodegeneratif dan neuro-inflamatori, seperti multiple sclerosis (MS) dan model heliks eksperimental autoimmune encephalomyelitis (EAE). Keparahan penyakit tersebut boleh dicirikan oleh tahap pencerobohan selular kepada sistem saraf pusat (CNS) dan nyah-myelinasi yang melibatkan effektor-effektor sel Th1 dan Th17. Kajian ini dijalankan untuk mengkaji keberkesanan perencatan GSK-3 pada waktu rawatan yang berbeza di dalam membaikpulih kondisi penyakit. Kami juga bertujuan untuk menentukan kesan perencatan GSK-3 kepada pencerobohan neuro pusat dan nyah-myelinasi. Model tikus betina C57BL/6 dirangsang oleh glikoprotein myelin oligodendrocyte (MOG35-55) bersama toksin pertusis lengkap (CFA) dan toksin pertusis. Inhibisi GSK-3 dilakukan melalui Tideglusib, heterosiklik kecil Thiadiazolidinones (TDZD), dengan cara 'intraperitoneal' semasa induksi pra-EAE, pada hari yang sama, dan selepas induksi EAE. Data menunjukkan bahawa penghantaran Tideglusib semasa induksi pra-EAE sangat melindungi tikus dari kondisi EAE, hal ini telah menunda permulaan simptom EAE sebanyak 7 hari dari hari-14.3 ± 0.5 (dalam tikus EAE) hingga hari-21.3 ± 3.2 (dalam tikus yang dirawat). Tambahan pula, tikus EAE yang dirawat Tideglusib telah mengurangkan pencerobohan sel-sel radang dan kerosakan aksonal (nyah-myelinasi) dalam kord rahim. Inhibisi GSK-3 juga membatalkan penghasilan sitokin Th1, Th9 dan Th17 yang berkaitan, tetapi meningkatkan tahap IL-4 kepada 1790 ± 95.0 pg / mL berbanding EAE 480 ± 38.0 pg / mL dan IL-10 780 ± 22.0 pg / mL berbanding EAE 410 ± 16.0 pg / mL. Data telah menunjukkan keberkesanan Tideglusib pada masa yang berlainan dapat mempengaruhi keparahan penyakit EAE, dan dapat melindungi persekitaran kanser daripada pencerobohan atau potensi nyah-myelinasi selular. Oleh itu, kajian dengan bukti telah mencadangkan potensi penggunaan GSK-3 di dalam rawatan EAE.

ACKNOWLEDGEMENT

To my parents and family and My deepest gratitude to the best supervisor, Dr. Masriana Hassan for her encouragement and advices.



Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____

Date: _____

Name and Matric No: Ahmed Salam Sadeq, GS44982

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Dr. Masriana Hassan

Signature: _____
Name of Member
of Supervisory
Committee: Associate Professor Dr. Abdah Md. Akim

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		ii
ACKNOWLEDGEMENT		iii
APPROVAL		iv
DECLARATION		vi
LIST OF TABLES		x
LIST OF FIGURES		xi
CHAPTER		
1	INTRODUCTION	1
	1.1 Introduction	1
	1.2 Problem statement	2
	1.3 Objectives	2
	1.4 Research question	2
2	LITERATURE REVIEW	3
	2.1 Introduction	3
	2.2 The Immune System	3
	2.2.1 T Lymphocytes	3
	2.2.1.1 Activation of T cells	3
	2.2.1.2 T cell differentiation	4
	2.2.1.3 CD4+ T cells	4
	2.2.1.4 Helper T cells subtypes	5
	2.3 Autoimmune Disease (Multiple Sclerosis)	6
	2.3.1 Etiology and Pathology of MS	7
	2.4 Experimental Autoimmune Encephalomyelitis (EAE)	7
	2.5 The mechanism of CD4+ mediated Immunopathogenesis of EAE	8
	2.6 Current Therapies for EAE	9
	2.7 Interleukin 10 (IL-10)	9
	2.8 Glycogen Kinase Synthesis 3 (GSK-3)	10
	2.9 GSK-3 Inhibitors	10
	2.9.1 Effects of GSK-3 inhibition on CD4+ T cells differentiation and EAE	11
	2.9.2 Tideglusib (NP-12)	12
3	MATERIALS AND METHODS	13
	3.1 Introduction	13
	3.2 Source of experimental animals (The C57BL/6 mice)	13
	3.3 Media and Buffers	13
	3.4 Experimental Autoimmune Encephalomyelitis (EAE)	13
	3.4.1 EAE Induction	13
	3.4.2 Treatment with GSK-3 inhibitor	14
	3.4.3 EAE Monitoring	14

3.5	Cytometric Beads Array (CBA)	15
3.5.1	Standard Preparation	15
3.5.2	Sample Dilution	15
3.5.3	Assay Procedure	15
3.6	Histopathology	16
3.6.1	Brain and Spinal Cord samples collection	16
3.6.2	Hematoxylin and Eosin (H&E) staining and scoring	17
3.6.3	Luxol Fast Blue (LFB) staining and scoring	17
3.6.3.1	Assay Procedure	17
3.7	Statistical analysis	18
4	RESULTS	19
4.1	Induction of EAE in C57BL/6 mice with MOG ₃₅₋₅₅ peptide	19
4.2	Inhibition of GSK-3 signaling by NP-12 reduced severity and delays the onset of disease	20
4.3	Effect of NP-12 administration at different intervention time on cell infiltration and demyelination in EAE	24
4.4	The effect of Tideglusib NP-12 intervention time on the expression levels of different cytokines	29
5	DISCUSSION	34
5.1	Introduction	34
5.2	NP-12 alleviated clinical severity of EAE	34
5.3	NP-12 attenuated the inflammatory cells infiltrate and demyelination in CNS	35
5.4	NP-12 decreased the inflammatory cytokine and increased the production of anti-inflammatory cytokines in EAE mice	36
6	CONCLUSION	39
6.1	Introduction	39
6.2	Research summary	39
6.3	Recommendations for future research	40
	REFERENCES	41
	APPENDICES	52
	BIODATA OF STUDENT	54

LIST OF TABLES

Table	Page	
2.1	Types of T Helper Cells	6
3.1	Clinical scoring for EAE	14
3.2	Final concentration of each standard after serial dilution	15
3.3	Assessment of inflammation	17
3.4	Assessment of demyelination	17
4.1	Comparison of EAE and Tideglusib (NP-12) treated groups. Combined data from three independent experiments. Results showed the effect of NP-12 on EAE clinical features at different intervention time	21

LIST OF FIGURES

Figure		Page
4.1	Severity score of EAE induction in C57BL/6 mice after immunization with MOG peptide	19
4.2	Administration of GSK3 inhibitor NP-12 delayed the onset of EAE and reduced EAE clinical symptoms	23
4.3	Effect of Tideglusib (NP-12) on cell infiltration in EAE	25
4.4	Effect of Tideglusib (NP-12) on demyelination of spinal cord in EAE	26
4.5	Effect of Tideglusib (NP-12) on inflammation of brain in EAE	27
4.6	Effect of Tideglusib (NP-12) on demyelination of brain in EAE	28
4.7	Quantitative histopathological analysis of spinal cord	29
4.8	Quantitative data showing NP-12 treatment regulated the levels of cytokines production in C57BL/6 EAE mice serum.	33

CHAPTER 1

INTRODUCTION

1.1 Introduction

Multiple sclerosis (MS) is a progressive chronic autoimmune inflammatory and demyelinating disease of the central nervous system (CNS) affecting over 2.5 million people worldwide (Milo & Kahana, 2010). The etiology of MS is uncertain, however, both cellular and environmental factors have an influence on disease demonstration (Comabella & Khoury, 2012). MS is characterized pathologically by multiple areas of CNS white matter demyelination of brain stem (disruption of myelin that insulates and protects nerve cells), axonal injury/loss, inflammation, cerebellum and blood-brain barrier (BBB) disruption (Dhib-Jalbut, 2007).

In order to closely study the pathogenesis and pathophysiology of MS, different *in vivo* models have been developed using experimental animals. Experimental Autoimmune Encephalomyelitis (EAE) is one of the most used animal models to investigate neuro-inflammatory pathways and pathophysiology of MS. It is the most common model because it mimics the features of MS, such as inflammation, demyelination, and axonal injury, although it may not demonstrate all pathophysiological characteristics of the human disease. The pathophysiology of EAE is based on the reaction of the adaptive immune system against brain-specific antigens. This reaction promotes inflammation and demolition of the antigen carrying structures, ending with neurological and pathological characteristics comparable which those noticed in MS patients (Stromnes & Goverman, 2006). Central to the pathophysiology of MS, and by extension the EAE model, is glycogen synthase kinase-3 (GSK3), a serine/threonine kinase which is an essential factor in the inflammatory process (Beurel, Grieco, & Jope, 2015). For this reason, several drugs have been developed targeting GSK-3, as the inhibition of this enzyme results in an impressive anti-inflammatory protection that ameliorates the disease severity and symptoms (Klein & Melton, 1996). One such drug targeting GSK3 is NP-12 which is a synthetic derivative of Thiadiazolidine (TDZD) with potent neuroprotective and anti-inflammatory effects *in vivo* (Luna-Medina et al., 2007). However, treatments with GSK-3 in EAE models with different degrees of inflammatory conditions did not always translate into a decrease of the disease severity as expected. In some cases, no beneficial effects were observed during the disease, whereas others demonstrated an exacerbation of clinical symptoms (Beurel et al., 2013).

In line with this, the present study aimed at evaluating the effect of NP-12 injections associated with different degrees of inflammatory conditions; pre, same time, and after the C57BL/6 mice being induced with EAE. Our data indicate that early treatment with NP-12 is highly effective in reducing the clinical score and delay the onset of EAE. we postulate that the effect of NP-12 in pre and same time of injections were able to diminish the lymphocyte infiltration into the spinal cord and reduce

demyelination in brain and spinal cord. Finally, our study demonstrates that the increase in serum cytokine levels via the NP-12 injection would increase their immunosuppressive capacity by increase the production of anti-inflammatory cytokines.

1.2 Problem statement

Recent studies have only demonstrated the effect of GSK-3 inhibitor, NP-12, before EAE induction. Furthermore, the effect of NP-12 on cell infiltration and demyelination during EAE still unclear. Moreover, cytokine levels after the administration of NP-12 to EAE is not elucidated yet. For these reasons, there is the need for experimental evidence identifying the ideal time to administrate NP-12 in EAE-induced mice, to obtain the most effective treatment result.

1.3 Objectives

1. To optimize the induction of EAE in C57BL/6 mice and determine the effect of GSK-3 inhibition on the severity of clinical symptoms.
2. To evaluate the effects of NP-12 on EAE-induced mice *in vivo*, and in the brain and spinal cord by histopathological analysis.
3. To compare the serum level of cytokines produced in NP-12 treated and control EAE-induced mice.

1.4 Research question

What is the effect of GSK3 inhibition on the *in vivo* EAE mouse model at different time point?

REFERENCES

- Abbas, A. K., Lichtman, A. H., & Pillai, S. (2011). *Cellular and molecular immunology*: Elsevier Health Sciences.
- Alabed, Y. Z., Pool, M., Tone, S. O., Sutherland, C., & Fournier, A. E. (2010). GSK3 β regulates myelin-dependent axon outgrowth inhibition through CRMP4. *The Journal of Neuroscience*, 30(16), 5635-5643.
- Anderson, M. S., Venanzi, E. S., Klein, L., Chen, Z., Berzins, S. P., Turley, S. J., . . . Benoist, C. (2002). Projection of an immunological self shadow within the thymus by the aire protein. *Science*, 298(5597), 1395-1401.
- Aranami, T., & Yamamura, T. (2008). Th17 Cells and autoimmune encephalomyelitis (EAE/MS). *Allergology International*, 57(2), 115-120.
- Asseman, C., Mauze, S., Leach, M. W., Coffman, R. L., & Powrie, F. (1999). An essential role for interleukin 10 in the function of regulatory T cells that inhibit intestinal inflammation. *The Journal of experimental medicine*, 190(7), 995-1004.
- Awasthi, A., Murugaiyan, G., & Kuchroo, V. K. (2008). Interplay between effector Th17 and regulatory T cells. *Journal of clinical immunology*, 28(6), 660-670.
- Azim, K., & Butt, A. M. (2011). GSK3 β negatively regulates oligodendrocyte differentiation and myelination in vivo. *Glia*, 59(4), 540-553.
- Bansal, R., Miyake, H., Nakamura, I., Eto, H., Gotoh, A., Fujisawa, M., . . . Hara, I. (2002). Fibroblast growth factors and their receptors in oligodendrocyte development: implications for demyelination and remyelination. *Developmental neuroscience*, 24(1), 35-46.
- Bettelli, E., Carrier, Y., Gao, W., Korn, T., Strom, T. B., Oukka, M., . . . Kuchroo, V. K. (2006). Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature*, 441(7090), 235-238.
- Beurel, E., Grieco, S. F., & Jope, R. S. (2015). Glycogen synthase kinase-3 (GSK3): regulation, actions, and diseases. *Pharmacology & therapeutics*, 148, 114-131.
- Beurel, E., & Jope, R. S. (2006). The paradoxical pro-and anti-apoptotic actions of GSK3 in the intrinsic and extrinsic apoptosis signaling pathways. *Progress in neurobiology*, 79(4), 173-189.
- Beurel, E., & Jope, R. S. (2009). Lipopolysaccharide-induced interleukin-6 production is controlled by glycogen synthase kinase-3 and STAT3 in the brain. *Journal of neuroinflammation*, 6(1), 9.

- Beurel, E., Kaidanovich-Beilin, O., Yeh, W.-I., Song, L., Palomo, V., Michalek, S. M., . . . Martinez, A. (2013). Regulation of Th1 cells and experimental autoimmune encephalomyelitis by glycogen synthase kinase-3. *The Journal of immunology*, *190*(10), 5000-5011.
- Beurel, E., Michalek, S. M., & Jope, R. S. (2010). Innate and adaptive immune responses regulated by glycogen synthase kinase-3 (GSK3). *Trends in immunology*, *31*(1), 24-31.
- Beurel, E., Yeh, W.-I., Michalek, S., Harrington, L., & Jope, R. (2011). Glycogen synthase kinase-3 is an early determinant in the differentiation of pathogenic Th17 cells.(152.3). *The Journal of immunology*, *186*(1 Supplement), 152.153-152.153.
- Bittner, S., Afzali, A. M., Wiendl, H., & Meuth, S. G. (2014). Myelin oligodendrocyte glycoprotein (MOG35-55) induced experimental autoimmune encephalomyelitis (EAE) in C57BL/6 mice. *Journal of visualized experiments: JoVE*(86).
- Carty, M., & Bowie, A. G. (2011). Evaluating the role of Toll-like receptors in diseases of the central nervous system. *Biochemical pharmacology*, *81*(7), 825-837.
- Chabas, D., Baranzini, S. E., Mitchell, D., Bernard, C. C., Rittling, S. R., Denhardt, D. T., . . . Pedotti, R. (2001). The influence of the proinflammatory cytokine, osteopontin, on autoimmune demyelinating disease. *Science*, *294*(5547), 1731-1735.
- Chappert, P., & Schwartz, R. H. (2010). Induction of T cell anergy: integration of environmental cues and infectious tolerance. *Current opinion in immunology*, *22*(5), 552-559.
- Chitnis, T., & Khoury, S. J. (2003). Cytokine shifts and tolerance in experimental autoimmune encephalomyelitis. *Immunologic research*, *28*(3), 223-239.
- Chomarat, P., Rissoan, M., Banchereau, J., & Miossec, P. (1993). Interferon gamma inhibits interleukin 10 production by monocytes. *Journal of Experimental Medicine*, *177*(2), 523-527.
- Chou, W.-y., Lu, C.-n., Lee, T.-h., Wu, C.-l., Hung, K.-s., Concejero, A. M., . . . Wang, C.-h. (2006). Electroporative interleukin-10 gene transfer ameliorates carbon tetrachloride-induced murine liver fibrosis by MMP and TIMP modulation. *Acta Pharmacologica Sinica*, *27*(4), 469-476.
- Coghlan, M. P., Culbert, A. A., Cross, D. A., Corcoran, S. L., Yates, J. W., Pearce, N. J., . . . Cox, L. R. (2000). Selective small molecule inhibitors of glycogen synthase kinase-3 modulate glycogen metabolism and gene transcription. *Chemistry & biology*, *7*(10), 793-803.

- COHEN, P. (1985). The role of protein phosphorylation in the hormonal control of enzyme activity. *European Journal of Biochemistry*, 151(3), 439-448.
- Comabella, M., & Khoury, S. J. (2012). Immunopathogenesis of multiple sclerosis. *Clinical immunology*, 142(1), 2-8.
- Cua, D. J., Sherlock, J., Chen, Y., Murphy, C. A., Joyce, B., Seymour, B., . . . Churakova, T. (2003). Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. *Nature*, 421(6924), 744.
- Curtsinger, J. M., & Mescher, M. F. (2010). Inflammatory cytokines as a third signal for T cell activation. *Current opinion in immunology*, 22(3), 333-340.
- Cuzzocrea, S., Genovese, T., Mazzon, E., Crisafulli, C., Di Paola, R., Muià, C., . . . Thiernemann, C. (2006). Glycogen synthase kinase-3 β inhibition reduces secondary damage in experimental spinal cord trauma. *Journal of Pharmacology and Experimental Therapeutics*, 318(1), 79-89.
- Dardalhon, V., Korn, T., Kuchroo, V. K., & Anderson, A. C. (2008). Role of Th1 and Th17 cells in organ-specific autoimmunity. *Journal of autoimmunity*, 31(3), 252-256.
- De Sarno, P., Axtell, R. C., Raman, C., Roth, K. A., Alessi, D. R., & Jope, R. S. (2008). Lithium prevents and ameliorates experimental autoimmune encephalomyelitis. *The Journal of immunology*, 181(1), 338-345.
- del Ser, T., Steinwachs, K. C., Gertz, H. J., Andres, M. V., Gomez-Carrillo, B., Medina, M., . . . Leon, T. (2013). Treatment of Alzheimer's disease with the GSK-3 inhibitor tideglusib: a pilot study. *Journal of Alzheimer's Disease*, 33(1), 205-215.
- Dhib-Jalbut, S. (2007). Pathogenesis of myelin/oligodendrocyte damage in multiple sclerosis. *Neurology*, 68(22 suppl 3), S13-S21.
- Edwards, J. P., Zhang, X., Frauwirth, K. A., & Mosser, D. M. (2006). Biochemical and functional characterization of three activated macrophage populations. *Journal of leukocyte biology*, 80(6), 1298-1307.
- El-behi, M., Rostami, A., & Ciric, B. (2010). Current views on the roles of Th1 and Th17 cells in experimental autoimmune encephalomyelitis. *Journal of Neuroimmune Pharmacology*, 5(2), 189-197.
- Eldar-Finkelman, H., & Martinez, A. (2011). GSK-3 inhibitors: preclinical and clinical focus on CNS. *Front. Mol. Neurosci*, 4.
- Elyaman, W., Bradshaw, E. M., Uyttenhove, C., Dardalhon, V., Awasthi, A., Imitola, J., . . . Renaud, J.-C. (2009). IL-9 induces differentiation of TH17 cells and enhances function of FoxP3⁺ natural regulatory T cells. *Proceedings of the National Academy of Sciences*, pnas. 0812530106.

- Fillatreau, S., Sweeney, C. H., McGeachy, M. J., Gray, D., & Anderton, S. M. (2002). B cells regulate autoimmunity by provision of IL-10. *Nature immunology*, 3(10), 944-950.
- Fiorentino, D. F., Bond, M. W., & Mosmann, T. (1989). Two types of mouse T helper cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. *The Journal of experimental medicine*, 170(6), 2081-2095.
- Fitzgerald, D. C., Zhang, G.-X., El-Behi, M., Fonseca-Kelly, Z., Li, H., Yu, S., . . . Rostami, A. (2007). Suppression of autoimmune inflammation of the central nervous system by interleukin 10 secreted by interleukin 27-stimulated T cells. *Nature immunology*, 8(12), 1372.
- Gabryšová, L., Nicolson, K. S., Streeter, H. B., Verhagen, J., Sabatos-Peyton, C. A., Morgan, D. J., & Wraith, D. C. (2009). Negative feedback control of the autoimmune response through antigen-induced differentiation of IL-10-secreting Th1 cells. *Journal of Experimental Medicine*, 206(8), 1755-1767.
- Gazzinelli, R. T., Makino, M., Chattopadhyay, S. K., Snapper, C., Sher, A., Hügin, A., & Morse, H. d. (1992). CD4+ subset regulation in viral infection. Preferential activation of Th2 cells during progression of retrovirus-induced immunodeficiency in mice. *The Journal of immunology*, 148(1), 182-188.
- Gold, R., Lington, C., & Lassmann, H. (2006). Understanding pathogenesis and therapy of multiple sclerosis via animal models: 70 years of merits and culprits in experimental autoimmune encephalomyelitis research. *Brain*, 129(8), 1953-1971.
- Goldenberg, M. M. (2012). Multiple sclerosis review. *Pharmacy and Therapeutics*, 37(3), 175.
- Goverman, J. (2009). Autoimmune T cell responses in the central nervous system. *Nature Reviews Immunology*, 9(6), 393.
- Grossi, J. A., Raulet, D. H., & Allison, J. P. (1992). CD28-mediated signalling co-stimulates murine T cells and prevents induction of anergy in T-cell clones. *Nature*, 356, 607-609.
- Groux, H., Bigler, M., de Vries, J. E., & Roncarolo, M.-G. (1998). Inhibitory and stimulatory effects of IL-10 on human CD8+ T cells. *The Journal of immunology*, 160(7), 3188-3193.
- Haak, S., Croxford, A. L., Kreymborg, K., Heppner, F. L., Pouly, S., Becher, B., & Waisman, A. (2009). IL-17A and IL-17F do not contribute vitally to autoimmune neuro-inflammation in mice. *The Journal of clinical investigation*, 119(1), 61-69.

- Hawkins, C., Munro, P., MacKenzie, F., Kesselring, J., Tofts, P., Du Boulay, E., . . . McDonald, W. (1990). Duration and selectivity of blood-brain barrier breakdown in chronic relapsing experimental allergic encephalomyelitis studied by gadolinium-DTPA and protein markers. *Brain*, *113*(2), 365-378.
- Hill, E. V., Ng, T., Burton, B. R., Oakley, C. M., Malik, K., & Wraith, D. C. (2015a). Glycogen synthase kinase- 3 controls IL- 10 expression in CD4+ effector T-cell subsets through epigenetic modification of the IL- 10 promoter. *European journal of immunology*, *45*(4), 1103-1115.
- Hill, E. V., Ng, T. S., Burton, B. R., Oakley, C. M., Malik, K., & Wraith, D. C. (2015b). Glycogen synthase kinase- 3 controls IL- 10 expression in CD4+ effector T- cell subsets through epigenetic modification of the IL- 10 promoter. *European journal of immunology*, *45*(4), 1103-1115.
- Hobom, M., Storch, M. K., Weissert, R., Maier, K., Radhakrishnan, A., Kramer, B., . . . Diem, R. (2004). Mechanisms and time course of neuronal degeneration in experimental autoimmune encephalomyelitis. *Brain pathology*, *14*(2), 148-157.
- Hofstetter, H., Gold, R., & Hartung, H.-P. (2009). Th17 cells in MS and experimental autoimmune encephalomyelitis. *The International MS Journal*, *16*(1), 12-19.
- Hofstetter, H. H., Ibrahim, S. M., Koczan, D., Kruse, N., Weishaupt, A., Toyka, K. V., & Gold, R. (2005). Therapeutic efficacy of IL-17 neutralization in murine experimental autoimmune encephalomyelitis. *Cellular immunology*, *237*(2), 123-130.
- Høglund, R. A., & Maghazachi, A. (2014). Multiple sclerosis and the role of immune cells. *World journal of experimental medicine*, *4*(3), 27-37.
- Hou, Y., Ryu, C. H., Park, K. Y., Kim, S. M., Jeong, C. H., & Jeun, S.-S. (2013). Effective combination of human bone marrow mesenchymal stem cells and minocycline in experimental autoimmune encephalomyelitis mice. *Stem cell research & therapy*, *4*(4), 77.
- Hsieh, C.-S., Macatonia, S. E., Tripp, C. S., Wolf, S. F., O'GARRA, A., & Murphy, K. M. (2008). Development of TH1 CD4+ T cells through IL-12 produced by listeria-induced macrophages. *The Journal of immunology*, *181*(7), 4437-4439.
- Huang, Y. H., Zozulya, A. L., Weidenfeller, C., Schwab, N., & Wiendl, H. (2009). T cell suppression by naturally occurring HLA- G- expressing regulatory CD4+ T cells is IL- 10- dependent and reversible. *Journal of leukocyte biology*, *86*(2), 273-281.
- Huber, S., Gagliani, N., Esplugues, E., O'Connor Jr, W., Huber, F. J., Chaudhry, A., . . . Rudensky, A. Y. (2011). Th17 cells express interleukin-10 receptor and are controlled by Foxp3⁻ and Foxp3⁺ regulatory CD4⁺ T cells in an interleukin-10-dependent manner. *Immunity*, *34*(4), 554-565.

- Janeway, C., Travers, P., & Walport, M. (2005). *Immuno biology: Immune system in health and disease*.
- Janeway Jr, C. A., & Medzhitov, R. (2002). Innate immune recognition. *Annual review of immunology*, 20(1), 197-216.
- Jiang, H., Guo, W., Liang, X., & Rao, Y. (2005). Both the establishment and the maintenance of neuronal polarity require active mechanisms: critical roles of GSK-3 β and its upstream regulators. *Cell*, 120(1), 123-135.
- Jones, M., Nguyen, T., Deboy, C., Griffin, J., Whartenby, K., Kerr, D., & Calabresi, P. (2008). Behavioral and pathological outcomes in MOG 35–55 experimental autoimmune encephalomyelitis. *Journal of neuroimmunology*, 199(1), 83-93.
- Jope, R. S., & Johnson, G. V. (2004). The glamour and gloom of glycogen synthase kinase-3. *Trends in biochemical sciences*, 29(2), 95-102.
- Jope, R. S., Yuskaitis, C. J., & Beurel, E. (2007). Glycogen synthase kinase-3 (GSK3): inflammation, diseases, and therapeutics. *Neurochemical research*, 32(4-5), 577-595.
- Juhler, M., Barry, D. I., Offner, H., Konat, G., Klinken, L., & Paulson, O. B. (1984). Blood-brain and blood-spinal cord barrier permeability during the course of experimental allergic encephalomyelitis in the rat. *Brain research*, 302(2), 347-355.
- Kelly, S., Zhao, H., Sun, G. H., Cheng, D., Qiao, Y., Luo, J., . . . Yenari, M. A. (2004). Glycogen synthase kinase 3 β inhibitor Chir025 reduces neuronal death resulting from oxygen-glucose deprivation, glutamate excitotoxicity, and cerebral ischemia. *Experimental neurology*, 188(2), 378-386.
- Klein, P. S., & Melton, D. A. (1996). A molecular mechanism for the effect of lithium on development. *Proceedings of the National Academy of Sciences*, 93(16), 8455-8459.
- Komiyama, Y., Nakae, S., Matsuki, T., Nambu, A., Ishigame, H., Kakuta, S., . . . Iwakura, Y. (2006). IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. *The Journal of immunology*, 177(1), 566-573.
- Korn, T., Bettelli, E., Gao, W., Awasthi, A., Jäger, A., Strom, T. B., . . . Kuchroo, V. K. (2007a). IL-21 initiates an alternative pathway to induce proinflammatory T H 17 cells. *Nature*, 448(7152), 484.
- Korn, T., Bettelli, E., Gao, W., Awasthi, A., Jäger, A., Strom, T. B., . . . Kuchroo, V. K. (2007b). IL-21 initiates an alternative pathway to induce proinflammatory TH17 cells. *Nature*, 448(7152), 484-487.

- Kroenke, M. A., Carlson, T. J., Andjelkovic, A. V., & Segal, B. M. (2008). IL-12–and IL-23–modulated T cells induce distinct types of EAE based on histology, CNS chemokine profile, and response to cytokine inhibition. *The Journal of experimental medicine*, 205(7), 1535-1541.
- Langrish, C. L., Chen, Y., Blumenschein, W. M., Mattson, J., Basham, B., Sedgwick, J. D., . . . Cua, D. J. (2005). IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *Journal of Experimental Medicine*, 201(2), 233-240.
- Laurence, A., Tato, C. M., Davidson, T. S., Kanno, Y., Chen, Z., Yao, Z., . . . Hennighausen, L. (2007). Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. *Immunity*, 26(3), 371-381.
- Lobell, A., Weissert, R., Eltayeb, S., de Graaf, K. L., Wefer, J., Storch, M. K., . . . Olsson, T. (2003). Suppressive DNA vaccination in myelin oligodendrocyte glycoprotein peptide-induced experimental autoimmune encephalomyelitis involves a T1-biased immune response. *The Journal of immunology*, 170(4), 1806-1813.
- Lublin, F. D., Knobler, R. L., Kalman, B., Goldhaber, M., Marini, J., Perrault, M., . . . Korngold, R. (1993). Monoclonal anti-gamma interferon antibodies enhance experimental allergic encephalomyelitis. *Autoimmunity*, 16(4), 267-274.
- Lublin, F. D., & Reingold, S. C. (1996). Defining the clinical course of multiple sclerosis results of an international survey. *Neurology*, 46(4), 907-911.
- Luna-Medina, R., Cortes-Canteli, M., Sanchez-Galiano, S., Morales-Garcia, J. A., Martinez, A., Santos, A., & Perez-Castillo, A. (2007). NP031112, a thiadiazolidinone compound, prevents inflammation and neurodegeneration under excitotoxic conditions: potential therapeutic role in brain disorders. *Journal of Neuroscience*, 27(21), 5766-5776.
- Martín-Saavedra, F. M., Flores, N., Dorado, B., Eguiluz, C., Bravo, B., García-Merino, A., & Ballester, S. (2007). Beta-interferon unbalances the peripheral T cell proinflammatory response in experimental autoimmune encephalomyelitis. *Molecular immunology*, 44(14), 3597-3607.
- Martinez, A., Alonso, M., Castro, A., Pérez, C., & Moreno, F. J. (2002). First non-ATP competitive glycogen synthase kinase 3 β (GSK-3 β) inhibitors: thiadiazolidinones (TDZD) as potential drugs for the treatment of Alzheimer's disease. *Journal of medicinal chemistry*, 45(6), 1292-1299.
- Martinez, A., Castro, A., Dorronsoro, I., & Alonso, M. (2002). Glycogen synthase kinase 3 (GSK-3) inhibitors as new promising drugs for diabetes, neurodegeneration, cancer, and inflammation. *Medicinal research reviews*, 22(4), 373-384.
- Matzinger, P. (1994). Tolerance, danger, and the extended family. *Annual review of immunology*, 12(1), 991-1045.

- Mazanetz, M. P., & Fischer, P. M. (2007). Untangling tau hyperphosphorylation in drug design for neurodegenerative diseases. *Nature reviews Drug discovery*, 6(6), 464-479.
- McGuirk, P., McCann, C., & Mills, K. H. (2002). Pathogen-specific T Regulatory 1 Cells Induced in the Respiratory Tract by a Bacterial Molecule that Stimulates Interleukin 10 Production by Dendritic Cells A Novel Strategy for Evasion of Protective T Helper Type 1 Responses by Bordetella pertussis. *The Journal of experimental medicine*, 195(2), 221-231.
- Milo, R., & Kahana, E. (2010). Multiple sclerosis: geoeidemiology, genetics and the environment. *Autoimmunity reviews*, 9(5), A387-A394.
- Milo, R., & Miller, A. (2014). Revised diagnostic criteria of multiple sclerosis. *Autoimmunity reviews*, 13(4), 518-524.
- Miranda-Hernandez, S., Gerlach, N., Fletcher, J. M., Biro, E., Mack, M., Körner, H., & Baxter, A. G. (2011). Role for MyD88, TLR2 and TLR9 but not TLR1, TLR4 or TLR6 in experimental autoimmune encephalomyelitis. *The Journal of immunology*, 187(2), 791-804.
- Mix, E., Meyer-Rienecker, H., & Zettl, U. K. (2008). Animal models of multiple sclerosis for the development and validation of novel therapies—potential and limitations. *Journal of neurology*, 255(6), 7-14.
- Mosmann, T. (1996). Two types of murine helper T cell clones. I. Definition according to profiles of lymphokine activities and secreted proteins. *J. Immunol.*, 136, 2346.
- Mosmann, T. R., Cherwinski, H., Bond, M. W., Giedlin, M. A., & Coffman, R. L. (1986). Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *The Journal of immunology*, 136(7), 2348-2357.
- Nagelkerken, L. (1998). Role of Th1 and Th2 cells in autoimmune demyelinating disease. *Brazilian journal of medical and biological research*, 31(1), 55-60.
- Neefjes, J., Jongstra, M. L., Paul, P., & Bakke, O. (2011). Towards a systems understanding of MHC class I and MHC class II antigen presentation. *Nature Reviews Immunology*, 11(12), 823-836.
- Noble, W., Planel, E., Zehr, C., Olm, V., Meyerson, J., Suleman, F., . . . Feinstein, B. (2005). Inhibition of glycogen synthase kinase-3 by lithium correlates with reduced tauopathy and degeneration in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 102(19), 6990-6995.
- Nowak, E. C., Weaver, C. T., Turner, H., Begum-Haque, S., Becher, B., Schreiner, B., . . . Noelle, R. J. (2009). IL-9 as a mediator of Th17-driven inflammatory disease. *Journal of Experimental Medicine*, 206(8), 1653-1660.

- O'Brien, W. T., & Klein, P. S. (2009). Validating GSK3 as an in vivo target of lithium action. *Biochemical Society Transactions*, 37(5), 1133-1138.
- O'Garra, A., & Vieira, P. (2007). TH1 cells control themselves by producing interleukin-10. *Nature Reviews Immunology*, 7(6), 425-428.
- Opal, S. M., & DePalo, V. A. (2000). Anti-inflammatory cytokines. *Chest Journal*, 117(4), 1162-1172.
- Ouyang, W., Kolls, J. K., & Zheng, Y. (2008). The biological functions of T helper 17 cell effector cytokines in inflammation. *Immunity*, 28(4), 454-467.
- PADILLA, A., HAUER, J. A., TSIGELNY, I., PARELLO, J., & TAYLOR, S. S. (1997). Solution structure of synthetic peptide inhibitor and substrate of cAMP-dependent protein kinase. A study by 2D 1H NMR and molecular dynamics. *The Journal of peptide research*, 49(3), 210-220.
- Ring, D. B., Johnson, K. W., Henriksen, E. J., Nuss, J. M., Goff, D., Kinnick, T. R., . . . Slabiak, T. (2003). Selective glycogen synthase kinase 3 inhibitors potentiate insulin activation of glucose transport and utilization in vitro and in vivo. *Diabetes*, 52(3), 588-595.
- Rösler, K., & Hess, C. (2010). Conduction studies in multiple sclerosis. *Multiple Sclerosis: Recovery of Function and Neurorehabilitation*, 1, 1-10.
- Seira, O., Gavín, R., Gil, V., Llorens, F., Rangel, A., Soriano, E., & Del Río, J. A. (2010). Neurites regrowth of cortical neurons by GSK3 β inhibition independently of Nogo receptor 1. *Journal of neurochemistry*, 113(6), 1644-1658.
- Selenica, M. L., Jensen, H. S., Larsen, A. K., Pedersen, M., Helboe, L., Leist, M., & Lotharius, J. (2007). Efficacy of small- molecule glycogen synthase kinase- 3 inhibitors in the postnatal rat model of tau hyperphosphorylation. *British journal of pharmacology*, 152(6), 959-979.
- Sereno, L., Coma, M., Rodriguez, M., Sanchez-Ferrer, P., Sanchez, M., Gich, I., . . . Guardia-Laguarta, C. (2009). A novel GSK-3 β inhibitor reduces Alzheimer's pathology and rescues neuronal loss in vivo. *Neurobiology of disease*, 35(3), 359-367.
- Sethna, M. P., & Lampson, L. A. (1991). Immune modulation within the brain: recruitment of inflammatory cells and increased major histocompatibility antigen expression following intracerebral injection of interferon- γ . *Journal of neuroimmunology*, 34(2-3), 121-132.
- Setoguchi, R., Hori, S., Takahashi, T., & Sakaguchi, S. (2005). Homeostatic maintenance of natural Foxp3⁺ CD25⁺ CD4⁺ regulatory T cells by interleukin (IL)-2 and induction of autoimmune disease by IL-2 neutralization. *Journal of Experimental Medicine*, 201(5), 723-735.

- Shaw, P. J., Barr, M. J., Lukens, J. R., McGargill, M. A., Chi, H., Mak, T. W., & Kanneganti, T.-D. (2011). Signaling via the RIP2 adaptor protein in central nervous system-infiltrating dendritic cells promotes inflammation and autoimmunity. *Immunity*, 34(1), 75-84.
- Shtrichman, R., & Samuel, C. E. (2001). The role of gamma interferon in antimicrobial immunity. *Current opinion in microbiology*, 4(3), 251-259.
- Sospedra, M., & Martin, R. (2005). Immunology of multiple sclerosis*. *Annu. Rev. Immunol.*, 23, 683-747.
- Stadelmann, C., Wegner, C., & Brück, W. (2011). Inflammation, demyelination, and degeneration—recent insights from MS pathology. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1812(2), 275-282.
- Steinman, L., & Zamvil, S. S. (2005). Virtues and pitfalls of EAE for the development of therapies for multiple sclerosis. *Trends in immunology*, 26(11), 565-571.
- Stromnes, I. M., Cerretti, L. M., Liggitt, D., Harris, R. A., & Goverman, J. M. (2008). Differential regulation of central nervous system autoimmunity by TH1 and TH17 cells. *Nature medicine*, 14(3), 337-342.
- Stromnes, I. M., & Goverman, J. M. (2006). Active induction of experimental allergic encephalomyelitis. *Nature protocols*, 1(4), 1810.
- Sydora, B. C., MacFarlane, S. M., Tavernini, M. M., Doyle, J. S., & Fedorak, R. N. (2008). CD4+ CD25+ regulatory T cells have divergent effects on intestinal inflammation in IL-10 gene-deficient mice. *Digestive diseases and sciences*, 53(6), 1544-1552.
- Szabo, S. J., Kim, S. T., Costa, G. L., Zhang, X., Fathman, C. G., & Glimcher, L. H. (2000). A novel transcription factor, T-bet, directs Th1 lineage commitment. *Cell*, 100(6), 655-669.
- Tipping, P. G., Kitching, A. R., Huang, X. R., Mutch, D. A., & Holdsworth, S. R. (1997). Immune modulation with interleukin- 4 and interleukin- 10 prevents crescent formation and glomerular injury in experimental glomerulonephritis. *European journal of immunology*, 27(2), 530-537.
- VAKNIN-DEMBINSKY, A., BALASHOV, K., & WEINER, H. L. (2006). IL-23 is increased in dendritic cells in multiple sclerosis and down-regulation of IL-23 by antisense oligos increases dendritic cell IL-10 production. *The Journal of immunology*, 176(12), 7768-7774.
- Vicente, R., Swainson, L., Marty-Grès, S., De Barros, S. C., Kinet, S., Zimmermann, V. S., & Taylor, N. (2010). *Molecular and cellular basis of T cell lineage commitment*. Paper presented at the Seminars in immunology.

- Viglietta, V., Baecher-Allan, C., Weiner, H. L., & Hafler, D. A. (2004). Loss of functional suppression by CD4⁺ CD25⁺ regulatory T cells in patients with multiple sclerosis. *The Journal of experimental medicine*, 199(7), 971-979.
- Waldner, H. (2009). The role of innate immune responses in autoimmune disease development. *Autoimmunity reviews*, 8(5), 400-404.
- Wefer, J., Harris, R. A., & Lobell, A. (2004). Protective DNA vaccination against experimental autoimmune encephalomyelitis is associated with induction of IFN β . *Journal of neuroimmunology*, 149(1), 66-76.
- Weiner, H. L. (2009). The challenge of multiple sclerosis: how do we cure a chronic heterogeneous disease? *Annals of neurology*, 65(3), 239-248.
- Wilson, C. B., Rowell, E., & Sekimata, M. (2009). Epigenetic control of T-helper-cell differentiation. *Nature Reviews Immunology*, 9(2), 91-105.
- Wingerchuk, D. M., & Carter, J. L. (2014). *Multiple sclerosis: current and emerging disease-modifying therapies and treatment strategies*. Paper presented at the Mayo Clinic Proceedings.
- Wood, P. J. (2006). *Understanding immunology*: Pearson Education.
- Woodgett, J. R. (1990). Molecular cloning and expression of glycogen synthase kinase-3/factor A. *The EMBO journal*, 9(8), 2431.
- Yang, X.-P., Ghoreschi, K., Steward-Tharp, S. M., Rodriguez-Canales, J., Zhu, J., Grainger, J. R., . . . Vahedi, G. (2011). Opposing regulation of the locus encoding IL-17 through direct, reciprocal actions of STAT3 and STAT5. *Nature immunology*, 12(3), 247.
- Yang, X. O., Panopoulos, A. D., Nurieva, R., Chang, S. H., Wang, D., Watowich, S. S., & Dong, C. (2007). STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. *Journal of Biological Chemistry*, 282(13), 9358-9363.
- Yuskaitis, C. J., & Jope, R. S. (2009). Glycogen synthase kinase-3 regulates microglial migration, inflammation, and inflammation-induced neurotoxicity. *Cellular signalling*, 21(2), 264-273.
- Zain, Z. M., Vidyadaran, S., & Hassan, M. GSK3 Inhibition Reduces Inflammatory Responses of Microglia and Upregulates Il-10 Production.
- Zhu, J., Yamane, H., & Paul, W. E. (2010). Differentiation of effector CD4 T cell populations. *Annual review of immunology*, 28, 445.

BIODATA OF STUDENT

Ahmed Salam Al-Zaidi was born in 1988. In 2012, I finished my bachelor's degree in medical laboratory technology from Baghdad University, Iraq. I worked in different medical laboratories in Baghdad for two years with a focus on the autoimmune diseases, then after I started my master study in UPM in September 2015.





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2018/2019

TITLE OF THESIS / PROJECT REPORT :

EVALUATION OF THE EFFECTIVE TIME FOR GSK-3 INHIBITION IN EXPERIMENTAL
AUTOIMMUNE ENCEPHALOMYELITIS MOUSE MODEL BY TIDEGLUSIB (NP-12) IN
PREVENTING NEUROINFLAMMATION

NAME OF STUDENT: AHMED SALAM AL-ZAIDI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (v)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]