

UNIVERSITI PUTRA MALAYSIA

DEVELOPMENT OF ENCAPSULATED Trichoderma harzianum UPM40 AGAINST Sclerotium rolfsii ON CHILI PLANT

FARIZ BIN ADZMI

IPTSM 2019 11



DEVELOPMENT OF ENCAPSULATED Trichoderma harzianum UPM40 AGAINST Sclerotium rolfsii ON CHILI PLANT



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

September 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

DEVELOPMENT OF ENCAPSULATED Trichoderma harzianum UPM40 AGAINST Sclerotium rolfsii ON CHILI PLANT

By

FARIZ BIN ADZMI

September 2018

Chairman : Professor Mohamed Hanafi Musa, PhD Institute : Tropical Agriculture and Food Security

Trichoderma as a biological control agent is widely used in sustainable agriculture. Commercialisation of this product in large scale with suitable formulation is still remains a problem especially to ensure the viability and efficiency of the formulation upon application in the field. Encapsulation method has emerged as sophisticated technique to develop the formulation of biological control agents. Encapsulation is the process of entrapping an active ingredient to shield it from adverse environmental conditions. Encapsulation of biological control agents (BCA) enables the creation of a microenvironment wherein the viability of the cells is maintained for storage, controlled release, and easy delivery. Encapsulation technique using extrusion method was used to prepare Trichoderma harzianum UPM40 beads. Based on the physical characterisation, a formulation with 2% (w/v) alginate, 1% (w/v) montmorillonite clay (MMT), and 5% (w/v) starch was able to produce T. harzianum UPM40 beads. This formulation produced almost perfectly shaped beads with SF 0.041 ± 0.006. It also offered a good swelling ability (62.11%) and less shrinkage (48.48%) during the drying process. Chemical characterization using the Fourier Transform Infrared Spectroscopy (FTIR) showed the interaction between the functional groups of alginate, MMT, and starch in the alginate-MMT-starch beads. It was shown by the shifting characteristic peaks of COOfrom starch at 1602 to 1610 cm⁻¹. Next C-O-C of alginate shifted from 1024 to 1002 cm⁻¹. Lastly, Si-O bending of MMT shifted from 405 to 453 cm⁻¹. Thermogravimetric analysis (TGA) showed an improvement in the thermal stability of the alginate-MMT-starch compared to alginate-MMT beads. X-ray Diffraction Analysis (XRD) shows the intercalation and exfoliation between starch and MMT. Peak for starch at $2\theta^{\circ}$ (15.06°) was shifting to $2\theta^{\circ}$ (16.64°) while peak at $2\theta^{\circ}$ (8.8°) for MMT was disappear in alginate-MMT-starch. Scanning electron microscopy (SEM) revealed a homogeneous distribution of MMT and starch particles throughout the alginate linkage. The surface area and pore diameter were 4.46 m²/g and 38.2 Å, respectively. Thus, *T. harzianum* UPM40 was successfully encapsulated in the alginate–MMT–starch beads. Storage analysis of the encapsulated *T. harzianum* UPM40 showed that low storage temperature (5 °C) was significantly better (p < 0.05) compared to at room temperature (30 °C). At low temperature, *T. harzianum* UPM40 beads maintained its viability of 6.59 log cfu/g up to seven months. The *T. harzianum* UPM40 beads were tested as a biological control agent against a soil-borne pathogen, *S. rolfsii*. In a dual culture, *T. harzianum* UPM40 displayed strong antagonistic activity against *S. rolfsii*. The percentage of inhibition of radial growth (PIRG) was 58.12%. Efficacy test of *T. harzianum* UPM40 for suppressing soil-borne diseases caused by *S. rolfsii* was conducted on chilli plants. Disease assessment results showed significant differences in disease incidence and disease reduction of DI at 88.46% and DSI at 72.80%.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PEMBANGGUNAN PENGKAPSULAN Trichoderma harzianum UPM40 KESAN TERHADAP Sclerotium rolfsii PADA POKOK CILI

Oleh

FARIZ BIN ADZMI

September 2018

Pengerusi : Profesor Mohamed Hanafi Musa, PhD Institut : Pertanian Tropika dan Keselamatan Makanan

Trichoderma sebagai agen kawalan biologi merupakan salah produk yang diguanakan dalam sektor pertanian mampan. Pengkomersilan produk ini pada sekala yang lebih besar dengan formulasi yang tebaik adalah menjadi salah satu masalah utama, terutamanya dalam memastikan kadar penyimpanan dan efisensi dalam penggunaan di lapangan. Kaedah pengkapsulan muncul sebagai satu teknik yang terbaik dalam pembanggunan formulasi agen kawalan biologi. Pengkapsulan adalah proses memerangkap suatu bahan aktif untuk melindunginya daripada persekitaran yang tidak diingini. Pengkapsulan agen kawalan biologi membolehkan pembentukan mikropersekitaran yang boleh mengekalkan keboleh hidupan sel-sel untuk penyimpanan, pelepasan terkawal, dan memudahkan penghantaran. Teknik pengkapsulan yang dilakukan melalui proses penyemperitan digunakan untuk menyediakan manik Trichoderma harzianum UPM40. Berdasarkan pencirian fizikal, formulasi dengan 2% (w/v) alginat, 1% (w/v) tanah liat montmorilonit (MMT), dan 5% (w / v) kanji merupakan formulasi terbaik untuk menghasilkan manik T. harzianum UPM40. Formulasi ini menghasilkan bentuk manik yang hampir sempurna dengan SF 0.041±0.006. Ia juga menunjukkan keupayaan mengembang sebanyak 62.11% dan kadar pengucupan 48.48%. Pencirian kimia menggunakan Fourier Transformasi Spektroskop Inframerah (FTIR) menunjukkan interaksi antara kumpulan-kumpulan berfungsi alginat, MMT, dan kanji dalam biji-biji kapsul alginat-MMT-kanji. Ini telah ditunjukkan pada pencirian perubahan puncak-puncak pada kumpulan COO- daripada kanji dari 1602 kepada 1610 cm⁻¹. Seterusnya, kumpulan berfungsi C-O-C daripada alginat dari 1024 to 1002 cm⁻¹. Akhir sekali kumpulan berfungsi Si-O lengkokan dari 405 to 453 cm⁻¹. Analisis Termogravimetrik (TGA) menunjukkan peningkatan dalam kestabilan terma manik alginat-MMT-kanji adalah lebih baik daripada manik alginat-MMT. Analisis Difraksi Sinar-X menunjukkan berlakunya interkelasi dan pengulupasan di antara zarah kanji dan MMT. Puncak untuk zarah kanji pada 20° (15.06°) telah berubah, kepada 20° (15.64°) sementara puncak 20° (8.8°) untuk MMT telah hilang. Mikroskopi Elektron Pengimbas (SEM) mendedahkan taburan secara sekata zarah-zarah MMT dan kanji di dalam jaringan alginat. Keluasan permukaan dan diameter adalah 4.46 m²/g and 38.2 Å. Maka, T. harzianum UPM40 berjaya diperangkap di dalam biji-biji manik alginat-MMTkanji. Analisis penyimpanan manik T. harzianum UPM40 menunjukkan bahawa suhu penyimpanan yang rendah pada 5 °C adalah lebih baik (P<0.05) berbanding pada suhu bilik (30 °C). Pada suhu rendah keupayaan hidup T. harzianum UPM40 dapat dikekalkan pada 6.59 log cfu/ g selama 7 bulan. Manik T. harzianum UPM40 diuji sebagai agen kawalan biologi untuk melawan patogen bawaan tanah, S. rolfsii. T. harzianum UPM40 menunjukkan aktiviti antagonistik yang kuat terhadap S. rolfsii di dalam kultur berganda. Analisis Peratusan Kerencatan Jejari (PIRG) dicatatkan pada 58.12%. Ujian keberkesanan T. harzianum UPM40 untuk merencat penyakit bawaan tanah oleh S. Rolfsii terhadap pokok cili. Keputusan penilaian penyakit menunjukkan perbezaan yang ketara dalam insidens penyakit dan indeks keterukan penyakit. Biji kapsul T. harzianum UPM40 boleh merencatkan S. rolfsii dengan pengurangan penyakit pada DI pada 88.46% dan DSI pada 72.80%.

ACKNOWLEDGEMENTS

Alhamdulillah to Almighty God

First and foremost, I would like to express my deepest appreciation to my supervisor, Prof. Dr. Mohamed Hanafi Musa, my previous supervisor Prof. Dr. Sariah Meon and members of the committee, Assoc. Prof. Dr. Radziah Othman and Prof. Dr. Nor Azah Yusuf for their guidance, advice and suggestion throughout the duration of this study.

I would like to extend my tremendous thanks and greatest gratitude to the lecturers and supported staff at Universiti Putra Malaysia who have directly or indirectly contributed their part. My sincere thanks also to my office and laboratory mate. I'm also very thankful to the financial support from Universiti Putra Malaysia.

My special thanks to my family especially my parent Tuan Haji Adzmi Othman and Hajjah Maznah Yaacob and also my wife Norhafeza Hamzah together with my children Fara Ellysha, Muhammad Iman and Muhammad Anas for their support and encourage.

Last but not least, I'm also indebted to all my beloved friend around the world, whose friendship I'll forever treasure.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohamed Hanafi Musa, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Radziah Othman, PhD

Associate Professor Faculty of Human Ecology Universiti Putra Malaysia (Member)

Nor Azah Yusuf, PhD

Professor Faculty of Science Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

Date:

Name and Matric No.: Fariz bin Adzmi, GS29584

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Professor Dr. Mohamed Hanafi Musa
Signature:	
Name of Member	
of Supervisory	Associate Professor Dr. Padziah Othman
Commutee.	Associate Professor DI. Nadzian Otinnan
Signature:	
Name of Member of Supervisory	
Committee:	Professor Dr. Nor Azah Yusuf

TABLE OF CONTENTS

			i age
ABST ABST ACKN APPR DECL LIST (LIST (RACT <i>RAK</i> IOWLEDGEMEN OVAL ARATION OF TABLES OF FIGURES OF ABBREVIAT	ITS	i iii v vi viii xiii xiii xiv xvi
CHAP 1	TER INTRODUCTIO		1
2	LITERATURE 2.1 Plant II 2.2 Sclero 2.3 Biologi 2.4 Tricho 2.5 Formu 2.6 Encapi 2.7 Encapi 2.7.1 2.7.2 2.7.3 2.8 Encapi 2.8.1 2.8.2 2.8.3 2.8.4 2.8.5	REVIEW Disease on Chilli tium rolfsii cal Control of Plant Disease derma as a Biological Control Agent lation of Biological Control Agent sulation sulation Method Spray Drying Freeze-Drying Complex Coacervation sulation Carrier Alginate Starch Montmorillonite Clay Chitosan Gelatins	4 4 5 6 7 7 8 9 10 10 11 11 12 12 13 13
3	MATERIALS / 3.1 Materia 3.2 Genera 3.2.1 3.2.2 3.2.3 3.2.4 3.2.5 3.2.6 3.2.7 3.2.8 3.2.9 3.2.10	AND METHOD als al Methodology Size Measurement Shape Measurement Swelling Ability of Beads Shrinkage Fourier Transform Infrared Thermogravimetric Analysis Scanning Electron Microscopy Analysis of Surface Area and Porosity X-ray Diffraction Analysis Preparation of <i>T. harzianum</i> UPM40 Culture	15 15 15 15 15 16 16 16 17 17

6

	3.2.11	Entrapment Efficacy of <i>T. harzianum</i> UPM40	10			
	3 2 12	Stability of Encapsulated T harzianum	10			
	0.2.12	UPM40	18			
	3.2.13	Statistical Analysis	19			
3.3	Prepar	ation, Characterisation, and Stability of				
	Encap	sulated <i>T. harzianum</i> UPM40 in Alginate–MMT	19			
	3.3.1	Preparation of Alginate Beads	19			
	3.3.2	Preparation of Alginate–MMT Beads	20			
	3.3.3	Encapsulation of <i>T. harzianum</i> UPM40 in				
2.4	Dronor	Alginate–MMI	20			
3.4	Encon	allon, Characlensallon, and Stability of subtod T harrignum LIPM40 in Alginate MMT				
	starch		20			
	341	Preparation of Alginate-MMT-starch Beads	20			
	342	Encapsulation of <i>T</i> harzianum UPM40 in	21			
		alginate-MMT-starch	21			
3.5	Biologi	ical Control of Soil-borne Disease caused by				
	Sclero	tium rolfsii in Chilli Plant using T. harzianum				
	UPM4	0 beads	21			
	3.5.1	Preparation of S. rolfsii Inoculum	21			
	3.5.2	Percentage Inhibition of Radial Growth	21			
	3.5.3	Slide Culture Method	22			
	3.5.4	Scanning Electron Microscopy Analysis	22			
	3.5.5	LIPM40 in Planting Modium	22			
	356	Plant Materials and Growth Condition	23			
	357	Disease Assessment	23			
	0.0.1		21			
RESU		DISCUSSION	26			
4.1	Prepar	ation. Characterisation. and Stability of	20			
	Encap	sulated T. harzianum UPM40 in Alginate-MMT				
	Beads	Ů	26			
	4.1.1	Physical Characterisation	26			
	4.1.2	Chemical Characterisation of Alginate-MMT				
		Beads	27			
	4.1.3	Characterisation of Encapsulated <i>I</i> .	00			
	111	Nichility and Stability of Enconculated T	32			
	4.1.4	harzianum LIPM40 boods	24			
12	Prenar	ration Characterisation and Stability of	54			
4.2	Encan	Encapsulated <i>T</i> harzianum UPM40 in Alginate–MMT–				
	starch		36			
	4.2.1	Physical Characterisation of the Alginate-				
		MMT-Starch	36			
	4.2.2	Chemical Characterisation of Alginate-MMT-				
		starch Beads	37			
	4.2.3	Viability of Encapsulated <i>T. harzianum</i> UPM40				
		Beads	43			

4

G

		4.2.4	Stability o UPM40 Bea	of ads	Encapsulated	Т.	harzianum	44
	4.3	Biologic Scleroti	al Control (<i>um rolfsii</i> ir	of 1 C	Soil-borne Dise Chilli Plant using	ease g <i>T.</i>	caused by <i>harzianum</i>	
		UPM40	Beads					46
		4.3.1	In vitro Ant Beads again	ago nst	onism by <i>T. ha</i> . <i>S. rolfsii</i>	rzianı	<i>ım</i> UPM40	46
		4.3.2	Viability of Planting Me	<i>T.</i> ediu	<i>harzianum</i> UF Im	PM40	Beads in	48
		4.3.3	Evaluation harzianum	of UP	f Biological (M40 Beads ag	Contr ainst	ol by <i>T.</i> Soil-borne	
			Pathogen S	5. rc	olfsii			48
5	SUMMA	ARY. CO	NCLUSION	J. A	ND RECOMME	NDA	TION FOR	
	FUTUR	E RÉSE	ARCH	<i>.</i>				52
								54 63
BIODA	TA OF S	TUDEN	т					75
PUBLI	CATION							76

5

 \bigcirc

LIST OF TABLES

Table		Page
3.1	Percentage of acetone use and timing for dehydration process	22
3.2	List of samples and treatments	23
3.3	Observation of signs and symptoms of infection	25
4.1	Physical properties of beads formulated with different percentages of MMT	26
4.2	Analysis of BET surface areas for MMT, alginate, and alginate–MMT beads	32
4.3	Comparison of weight, diameter, and shape factor of encapsulated <i>T. harzianum</i> UPM40 in alginate and alginate–MMT beads	33
4.4	Compari <mark>son of</mark> the shrinkage and conidia load of encapsulated <i>T. harzianum</i> UPM40 in alginate and alginate-MMT beads	33
4.5	Comparison of weight, diameter shrinkage, swelling ability, and shape factor of different percentages of starch in the alginate-MMT-starch bead formulation	36
4.6	Thermal decomposition temperature at every 10% of weight loss for alginate-MMT and alginate-MMT-starch beads	39
4.7	Comparison of entrapped conidia in the fresh and dried alginate–MMT–starch beads	43
4.8	Viability of different amounts of <i>T. harzianum</i> UPM40 beads in 1 kg of media (100% coconut dust)	48
4.9	The area under disease progression and disease reduction of chilli plant after 40 days of treatment based on disease incidence	49
4.10	The area under disease progression and disease reduction for chilli plant after 40 days of treatment based on disease severity	50

6

LIST OF FIGURES

Figure		Page
2.1	Schematic diagram of the extrusion encapsulation method	9
2.2	Schematic diagram of the freeze-drying encapsulation method	10
2.3	Alginate consists of linear unbranched polymer containing β -(1,4)-linked D-mannuronic acid (M) and α -(1,4)-linked L-guluronic acid (G) residue	11
2.4	The structure of montmorillonite clay	13
3.1	Serial dilution method of <i>T. harzianum</i> UPM 40 suspension	18
3.2	Graphical methodology for preparation and characterisation of alginate–MMT beads	19
3.3	Graphical methodology for preparation and characterisation of alginate-MMT-starch beads	20
4.1	FTIR spectra of: (a) alginate beads; (b) MMT; and (c) alginate- MMT beads in the spectrum scale of 4000 – 500 cm ⁻¹ shown the interaction between function group of alginate, MMT and alginate-MMT by shifting the wavenumber	29
4.2	TGA and DTG (inset figure) show the comparison of thermal degradation of MMT, alginate, and alginate-MMT	29
4.3a	SEM image of the distribution of MMT particles with irregular shape (1000×)	30
4.3b	SEM image of the cross section of alginate bead shows the dense and packed alginate linkage (1000×)	30
4.3c	SEM image of the cross section of alginate–MMT bead shows the distribution of MMT particles throughout the alginate matrix (1000×)	30
4.4	SEM image of cross section of <i>T. harzianum</i> UPM40 alginate– MMT bead that shows the distribution of <i>T. harzianum</i> UPM40 conidia. Arrow indicates pure conidia with size 2.14 μ m (10,000×)	33

4.5	Comparison between the viability of <i>T. harzianum</i> UPM40 released from the encapsulated beads under different storage conditions at room temperature $(30 \pm 2 \ ^{\circ}C)$ and cold temperature $(5 \pm 2 \ ^{\circ}C)$ from one to six months. Cold temperature $(5 \pm 2 \ ^{\circ}C)$ shows better storage condition for <i>T. harzianum</i> UPM40 beads	35
4.6	FTIR spectra of: (a) alginate (b) MMT (c) starch and alginate- MMT-starch beads in the spectrum scale of 4000 – 500 cm ⁻¹ shown the interaction between function group of alginate, MMT, starch and alginate-MMT-starch by shifting the wavenumber	38
4.7	TGA and DTG shows the comparison thermal degradation of alginate-MMT, alginate and alginate-MMT-starch	39
4.8a	SEM image of the starch particles (1000×)	40
4.8b	SEM image of the cross section of alginate–MMT bead showing the distribution of MMT particles throughout the alginate matrix (1000×)	40
4.8c	SEM image of the cross section of alginate–MMT–starch bead shows the distribution of MMT and starch particles throughout the alginate matrix (1000×)	40
4.9	XRD of alginate, MMT, starch, and alginate–MMT–starch. Red arrow shows the shfting of peak at 20 of 15.06° for starch to 16.64° in alginate–MMT–starch. The peak at 20 of 8.8° for MMT disappeared in alginate–MMT–starch	42
4.10	SEM image of the cross section of <i>T. harzianum</i> UPM40 beads (alginate–MMT–starch), showing the distribution of <i>T. harzianum</i> UPM40 conidia throughout the alginate–MMT–starch matrix (1000×)	44
4.11	Comparison between the viability of <i>T. harzianum</i> UPM40 released from the encapsulated beads under different storage conditions (room temperature $[30 \pm 2 \degree C]$ and cold temperature $[5 \pm 2 \degree C]$) from one to nine months. Cold temperature $(5 \pm 2 \degree C)$ showed better storage condition for <i>T. harzianum</i> UPM40 encapsulated in alginate–MMT–starch	45
4.12	(a) Inhibition zone in PIRG analyses between <i>T. harzianum</i> UPM40 against <i>S. rolfsii</i> in dual culture analysis. (b) SEM micrograph of the inhibition zone between <i>S. rofsii</i> mycelia and <i>T. harzianum</i> UPM40 hyphae (100×)	47

x٧

- 4.13 (a) SEM micrograph of *S. rolfsii* mycelium (1000×) and (b) SEM micrograph of *T. harzianum* UPM40 mycelium and conidia (1000×)
- 4.14 Soil-borne disease incidence for chilli plant, 40 days after treatment.
- 4.15 Soil-borne disease severity index for chilli plant, 40 days after treatment.



50

G

LIST OF ABBREVIATIONS

ANOVA	Analysed One-way of Variance		
ASAP	Surface Area and Porosity Analysis		
AUDPC	Area Under the Disease Progressive		
BCA	Biological Control Agent		
DAT	Day After Transplant		
DI	Disease Incidence		
DSI	Disease Severity Index		
EPA	Environmental Protection Agency		
FTIR	Fourier Transfrom Infared		
LSD	Least Significant Different		
MMT	Montmorillonite Clay		
PDA	Potato Dextrose Agar		
PIRG	Percentage of Inhibition Radial Growth		
RCBD	Randomized Complete Block Design		
SEM	Scanning Electron Microscopy		
SF	Sphericity Factor		
TGA	Thermal Gravimetric Analysis		
XRD	X-ray Diffraction		

6

CHAPTER 1

INTRODUCTION

Plant disease is a serious problem worldwide and has been a major factor that influences food production and human society development over thousand years (Palmgren et al., 2015). On the economic side, plant diseases cause an estimated 40 billion dollars losses worldwide every year, either directly or indirectly. At least 20%–40% of losses in crop yield are caused by pathogenic infections (Savary et al., 2012).

In Malaysia, the agriculture sector is the backbone of the economic strength and contributed 8.1% (RM89.5b) to the gross domestic product in 2016 (Department of Statistics Malaysia, 2016). However, Malaysia's agriculture is in an unsatisfactory condition with low productivity and quality. Several factors have been identified such as the quality of soil and seed, unpredictable weather, limitations in the use of new technologies, labour shortage, and poor maintenance of agriculture infrastructures. Disease outbreaks and insect pests are other contributing factors that cause the low production of yield. Infectious plant diseases are often caused by pathogenic organisms, such as fungi, bacteria, mycoplasma, viruses, viroids, and nematodes. Examples of plant disease include leaf spot, bacterial wilt, fusarium wilt, mosaic virus infection, and soil-borne disease (Nasehi et al., 2012; Vos et al., 2014; Ben-Jabeur et al., 2015).

The current agricultural practice to control plant diseases is by using pesticide. The U.S. Environmental Protection Agency (EPA) defines pesticide as 'any substance intended for preventing, destroying, repelling, or mitigating any pest'. Pesticides cover a wide range of compounds, including fungicides, insecticides, herbicides, rodenticides, molluscicides, nematicides, and plant growth regulators. This practice is uneconomical in the long run because pesticide use can damage agricultural land by harming beneficial insect species, soil microorganisms, and worms that naturally limit pest populations and maintain soil health; weakening plant root systems and immune systems; and reducing concentrations of essential plant nutrients in the soil such as nitrogen and phosphorus.

Biological control agents (BCA) is sociologically, commercially, and environmentally accepted as a legitimate tool to control plant pathogen. It can be enhanced by cultural practices such as <u>crop rotation</u> and <u>soil amendments</u>, priming plants to be disease-resistant, or genetically altering them to control pathogens (Lazarovits et al., 2014). Nowadays, there are several commercial BCA available in the market, such as *Agrobacteria*, *Pseudomonas*, *Streptomyces*, *Bacillus*, *Gliocladium*, *Trichoderma*, *Ampelomyces*, and *Coniothyrium*. The shelf life and effectiveness are the major difficulties of BCAs to compete with chemical fungicides. Both problems can be solved with scientific development of BCA formulations. The advantages of formulation include greater efficacy, increased shelf life, ease of handling, increased safety, lower production costs, and compatibility with agricultural practices (Vemmer & Patel, 2013).

It is well established that encapsulation technique has emerged as a sophisticated method in the formulation of biological control agents. Encapsulation within a matrix protects the microbial agents from biotic and abiotic stress factors (contaminations, soil antagonist, temperature, dryness, UV light, or mechanical stress) by providing a beneficial microenvironment (Rathore et al., 2013). This leads to extended shelf-life and maintains the metabolic activity for extended periods of time, not only during storage, but also after application, which results in fewer applications, dose reduction, as well as reduced biomass content (Szczech & Maciorowski, 2016).

Furthermore, one of the properties of encapsulated beads is the controlled release of the entrapped or encapsulated cell. The cells are released slowly by the factor of osmosis from the bead matrix (He at al., 2015). Cells survive longer in the soil and have extended persistence, which results in a reduced number of applications.

This study adopted an encapsulation technique to improve the viability and shelf life of *Trichoderma harzianum* UPM40, which was used as a biocontrol agent. Depending on the strain, the use of *Trichoderma* in agriculture provides numerous advantages: (i) colonisation of the rhizosphere by the BCA (rhizosphere competence) and allowing rapid establishment within the stable microbial communities in the rhizosphere, (ii) controlling pathogenic and competitive/deleterious microflora by using a variety of mechanisms, (iii) improving plant's health, and (iv) stimulating root growth (Soresh & Harman, 2008). For the encapsulated *Trichoderma* to be a success, the choice of carrier for the bead matrix is an important factor. Biodegradable polymer materials are usually used as a capsule matrix, as well as more specific, natural polysaccharides.

The characteristics of alginate, such as its biodegradability, non-toxicity, and biocompatibility, make it a preferred carrier material (Lee et al., 2012). It consists of α -L-guluronic acid (G) and β -D-mannuronic acid (M) residues, linearly linked by 1,4-glycosidic linkages (Paques et al., 2014).

However, the physical and chemical properties of alginate as a carrier are insufficient to achieve long-term stability of the encapsulated cells. Thus, to overcome this limitation, a combination carrier with different characteristics should be added into the formulation. It can be soluble, insoluble, or a combination of both. Several carriers, such as clay and starch, have been used in the formulation of encapsulated beads (Roy et al., 2009). The incorporation of clay as a carrier attracted great attention because it entrapped higher amount and increased encapsulation efficiency (Bušić et al., 2016). Moreover, the starch acted as a structural support to control the extent of shrinkage and maintain the shape of the alginate backbone during the drying process.

This study reports the preparation of alginate–MMT–starch beads through the extrusion technique. The resulting beads were characterised using Fourier transform infrared spectroscopy, thermogravimetric analysis, X-ray diffraction analysis, and scanning electron microscopy. Then, the encapsulation technique was used for *T. harzianum* UPM40. The viability and stability of *T. harzianum* UPM40 in different storage conditions were tested. The efficiency of *T. harzianum* UPM40 beads as a biological control agent against *S. rolfsii* was studied in a dual culture analysis and a shelter house condition.

Objectives:

- 1. To prepare and characterise encapsulated *T. harzianum* UPM40 in alginate–MMT formulation.
- 2. To evaluate the viability of encapsulated *T. harzianum* UPM40 in alginate–MMT during storage period.
- 3. To study the optimum content of starch in the alginate–MMT–starch formulation.
- 4. To investigate the bio-efficacy of the encapsulated *T. harzianum* UPM40 beads in alginate–MMT–starch as a biological control agent against *S. rolfsii* in chilli plants.

REFERENCES

- Aguiar, J., Estevinho, B. N., and Santos, L. 2016. Microencapsulation of natural antioxidants for food application – the specific case of coffee antioxidants – a review. *Trends in Food Science & Technology* 58: 21– 39.
- Akolade, J. O., Oloyede, H. O. B., and Onyenekwe, P. C. 2017. Encapsulation in chitosanbased polyelectrolyte complexes enhances antidiabetic activity of curcumin. *Journal of Functional Foods* 35: 584–594.
- Almasi, H., B. Ghanbarzadeh and A. A. Entezami, A. A. 2010. Physicochemical properties of starch—CMC–nanoclay biodegradable films. *International Journal of Biological Macromolecules* 46: 1–5.
- Al-Saleh M. M., Ibrahim, Y. E., Abo-Elyousr, K. A. M. and Alibrahim, J. S. 2011. Population dynamics of *Xanthomonas cmpestris pv. Vitians* on different plant species and management of bacterial leaf spot of lettuce under greenhouse conditions. *Crop Protection* 30: 88-887.
- Anal, A. and Singh H. 2007. Recent advances in microencapsulation of probiotics for industrial applications and targeted delivery. *Trends in Food Science Technology* 18: 240-251.
- Azevedo, M. A., Bourbon, A. I., Vicente, A. A., and Cerqueira, M. A. 2014. Alginate/ chitosan nanoparticles for encapsulation and controlled release of vitamin B2. *International Journal of Biological Macromolecules* 71: 141–146.
- Bailey, B. A., Bae, H., Strem, M. D., Crozier, J. and Holmes, K A. 2008. Antibiosis, mycoparasitism, and colonization success for endophytic *Trichoderma* isolates with biological control potential in *Theobroma cacao*. *Biological Control* 46: 24-35.
- Bakry, A. M., Abbas, S., Ali, B., Majeed, H., Abouelwafa, M. Y. and Mousa, A., 2016. Microencapsulation of oils: A comprehensive review of benefits, techniques, and applications. *Comprehensive Reviews in Food Science and Food Safety* 15: 143–182.
- Bashan, Y., Hernandes, J. P., Leyva, L. A. and Bacilio, M. 2002. Alginate microbeads as inoculants carrier for plant growth-promoting bacteria. *Biology and Fertility of Soils* 35: 359-368.
- Belscak-Cvitanovic, A., Komes, D., Karlović, S., Djaković, S., Špoljarić, I. and Mršić, G. 2015. Improving the controlled delivery formulations of caffeine in alginate hydrogel beads combined with pectin, carrageenan, chitosan and psyllium. *Food Chemistry* 167: 378–386.

- Benitez, T., Rincon, A. M., Limon, M. C. and Codon, A. C. 2004. Biocontrol mechanisms of Trichoderma strains. *International Microbiology* 7: 249-260.
- Ben-Jabeur, M., Ghabri, E., Myriam M. and Hamada W. 2015. Thyme essential oil as a defense inducer of tomato against gray mold and Fusarium wilt. *Plant Physiology and Biochemistry* 94: 3-40.
- Bergaya, F., Lagaly, G., 2013. Chapter 1–introduction to clay science: techniques and applications. In: Developments in Clay Science. 5. pp. 1–7. Elsevier Science.
- Berta, M., Lindsay, C., Pans, G. and Camino, G. 2006. Effect of chemical structure on combustion and thermal behavior of polyurethane elastomer layered silicate nanocomposites. *Polymer Degradation and Stability* 91: 1179–91.
- Cai, F., Yu. G., Wang, P., Wei, Z., Fu, L., Shen, Q. and Chen, W. 2013. Harzianolide, a novel plant growth regulator and systematic resistance elicitor from Trichoderma harzianum. *Plant Physiology and Biochemistry* 73: 106-113.
- Castro, H. P., Teixiera, P.M and Kirby, R. 1997. Evidence of membrane damange in Lactobacillus bulgaricus following freeze drying. *Journal of Applied Microbiology* 82: 87-94.
- Champagne, C.P., Mondou, F. Raymond, Y. and Roy, D. 1996. Effect of polymers and storage temperature on the stability of freezed-dried lactic acid bacteria. *Food Research International* 29: 555-562.
- Chan, E. S., Wong, S. L., Lee, P. P., Lee, J. S., Ti, T. B., Zhang, Z., Poncelet, D., Ravindra, P., Phan, S. H. and Yim, Z. H. 2011. Effects of starch filler on the physical properties of lyophilized calcium-alginate beads and the viability of encapsulated cells. *Carbohydrate Polymer* 83: 225-232.
- Cigdem K, Merih K. Effect of formulation on the viability of biocontrol agent, *Trichoderma harzianum* conidia. African J Biotechnol 2005; 85:483-486.
- Connick, W. J., Jackson, M. A., Williams, K.S. and Boyette, C. D. 1997. Stability of microsclerotial inoculums of *Colletotrichum truncatum* encapsulated in wheat flour-koalin granules. World Journal of Microbiology and Biotechnology 13: 549-554.
- Curtis, F. D., Lima, G. Vitullo, D. and De Cicco, V. 2010. Biocontrol of Rhizoctonia solani and Sclerotium rolfsii on tomato by delivering antagonistic bacteria through a drip irrigation system. *Crop Protection* 29: 663-670.

- Dang, X., Yang, M., Shan, Z., Mansouri, S., May, B. K. and Chen, X. 2017. On spray drying of oxidized corn starch cross-linked gelatin microcapsules for drug release. *Materials Science and Engineering: C* 74: 493–500.
- de Souza Simões, L., Madalena, D. A., Pinheiro, A. C., Teixeira, J. A., Vicente, A. A., and Ramos, Ó. L. 2017. Micro- and nano bio-based delivery systems for food applications: In vitro behavior. *Advances in Colloid* and Interface Science 243: 23–45.
- Department of Statistics Malaysia, 2016https://www.dosm.gov.my/v1/index.php?r=column/cthemeByCat& cat=72&bul_id=MDNYUitINmRKcENRY2FvMmR5TWdGdz09&menu_i d=Z0VTZGU1UHBUT1VJMFIpaXRR0xpdz09. Access October 2018
- Díaz, A., L. Franco, M.T. Casas, L.J. del Valle, J. Aymamí, C. Olmo and J. Puiggalí. 2014. Preparation of micro-molded exfoliated clay nanocomposites by means of ultrasonic technology. *Journal of Polymer Research* 24: 584-596.
- Druzhinina, I.S., Kopchinskiy, A.G. and Kubicek, C.P. 2006. The first 100 *Trichoderma* species characterized by molecular data. *Mycoscience* 47: 55–64.
- Ennajih, H., Bouhfid, R., Essassi, E. M., Bousmina, M. and El Kadib, A. 2012. Chitosan montmorillonite bio-based aerogel hybrid microspheres. *Microporous and Mesoporous Materials* 152: 208–213.
- Evidente, A., Cabras, A., Maddau, L., Serra, S., Andolfi, A. and Motta, A. 2003. Viridepyronone, a new antifungal 6-substitute 2H-pyran-2-one produce by Trichoderma viride. *Journal of Agriculture Food Chemistry* 51: 6957-6960.
- Fang, J., Fowler, P., Tomkinson, J. and Hill, C. A. 2002. The preparation and characterization of a series chemically modified patato starches. *Carbohydrate Polymer* 47: 245-252.
- Fathi, M., Martín, Á. and McClements, D. J. 2014. Nanoencapsulation of food ingredients using carbohydrate based delivery systems. *Trends in Food Science & Technology* 39: 18–39.
- Fuciños, C., Míguez, M., Fuciños, P., Pastrana, L. M., Rúa, M. L. and Vicente, A. A. 2017. Creating functional nanostructures: Encapsulation of caffeine into α-lactalbumin nanotubes. *Innovative Food Science & Emerging Technologies* 40: 10–17.
- Ganesan, S. Kuppusamy, R. G. and Sekar, R. 2007. Intergrated management of stem tot disease (*Sclerotium rolfsii*) of groundnut (*Arachis hypogeasa L.*) using *Rhizobium* and *Trichoderma harzianum. Turkey Journal Agriculture Forestry* 31: 103-108.

- Giovanni, S., Franca, B., Andrea, P., Cecilia, .P, Attilio, A., Raimondo, M., Nicoletta, M. and Guglielmina, G. 1995. Electrophilic alkenylation of aromatics with phenylacetylne over zeolite HSZ-360. *Tetrahedron Letter* 36: 9177-9180.
- Hani, N. M., Torkamani, A. E., Azarian, M. H., Mahmood, K. W. A., and Ngalim, S. H. 2017. Characterisation of electrospun gelatine nanofibres encapsulated with Moringa oleifera bioactive extract. *Journal of the Science of Food and Agriculture* 97: 3348–3358.
- He, Y. H., Wu, Z., Tu, L., Han, Y., Zhang, G. and Li, C. 2015. Encapsulation and characterization of slow-release microbial fertilizer from the composites of bentonite and alginate. *Applied Clay Science* 109–110: 68–75.
- Hosseini, S., Hosseini, H., Mohammadifar, M. A., German, J. B., Mortazavian, A. M., Mohammadi, A., Khosravi-Darani, K., Shojaee-Aliabadi, S. and Khaksar R. 2014. Preparation and characterization of alginate and alginate-resistant starch microparticles containing nisin. *Carbohydrate Polymers* 103: 573-580. http://www.fao.org/faostat/en/#data/QC. Access October 2018
- Huang, H. C. and Chang, C. 2003. Effect of relative humidity on myceliogenic germination of sclerotia of sclerotinia minor. Plant Pathology Bulletin 12: 65- 68.
- Hyakumachi, M., Mondal, S. N., Elsharkawy M. M. and Hassan N. 2014. Carbon loss by sclerotia of *Sclerotium rolfsii* under the influence of soil pH, temperature and matric potential and its effect on sclerotial germination and virulence. *Applied Soil Ecology* 77:_34-41.
- Ishwarya, S. P., Anandharamakrishnan, C., and Stapley, A. G. F. 2015. Sprayfreezedrying: A novel process for the drying of foods and bioproducts. *Trends in Food Science & Technology* 41: 161–181.
- Jia, Z., Dumont, M. J. and Orsat, V. 2016. Encapsulation of phenolic compounds present in plants using protein matrices. *Food Bioscience* 15: 87–104.
- Junaid, J. M., Nisar, A. D., Tariq, A. B., Arif, A. B. and Mudasir, A.B. 2013. Comicial biocontrol agents and their mechanism of action in the management of plant pathogens. *International Journal of Modern Plant* & *Animal Science* 1: 39-57.
- Kittur, F. S., Prashanth, K. H., Sankar, K.U. and Tharanathan, R. N. 2002. Characterization of chitin, chitosan and their carboxymethyl derivatives by differential scanning calorimetry. *Carbohydrate polymer* 49: 185-193.

- Komes, D. 2016. Application of whey protein isolates and zein for the formulation of alginate-based delivery systems encapsulating Ganoderma lucidum polyphenols. *Croatian Journal of Food Science and Technology*, 8(2): 99–106.
- Larena, I., Sabuquillo, P., Melgarejo, P. and De Cal, A. 2003. Biocontrol of *Fusarium* and *Verticillium* wilt of tomato by *Penicillium oxalicum* under greenhouse and field conditions. *Journal of Phytopathology* 151: 507-512.
- Lazarovits, G., Turnbull, A., Johnston-Monje, D. 2014. Encyclopedia of Agriculture and Food Systems. *Reference Module in Food Science* 388-399.
- Lee, K. Y. and Mooney, D. J. 2012. Alginate: Properties and biomedical applications. *Progress in polymer Science* 37: 106-126.
- Leoni, C., Braak, C. J. F., Golsanz, J. C., Dogliotti, S., Rossing, W. A. H. and van Bruggen, A. H. C. 2014. Sclerotium rolfsii dynamics in soil as affected by crop sequences. *Applied Soil Ecology* 75: 95-105.
- Liu, Y. and Bell-Pederson, D. 2006. Circadin rhythms in *Neurospora crasa* and other filamentous fungi. *Eukaryotic Cell* 5: 1184-1193.
- Lozano-Vazquez, G. Lobato-Calleros, C. Escalona-Buendia, H. Chavez, G. Alvarez-Ramirez, J. and Vernon-Carter, E.J. 2015. Effect of the weight ratio of alginate-modified tapioca starch on the physicochemical properties and release kinetics of chlorogenic acid containing beads. *Food Hydrocolloids* 48: 301-311.
- Luo, Y. and Wang, Q. 2014. Recent development of chitosan-based polyelectrolyte complexes with natural polysaccharides for drug delivery. *International Journal of Biological Macromolecules* 64: 353– 367.
- Mansoori, Y., Atghia, S. V., Zamanloo, M. R., I Manzadeh, G. h., and Sirousazar, M. 2010. Polymer–clay nanocomposites:Free-radical grafting of polyacrylamide onto organophilic montmorillonite. *European Polymer Journal* 46: 1844–1853.
- Markovish, N. A. And Kononova G. L. 2003. Lytic enzymes of Trichoderma and their role in plant defences from fungal disease: A review. *Applied Biochemistry and Microbiology* 34: 341-351.
- Muller, P., Kapin, E. and Fekete, E. 2014. Effects of preparation method on the structure and mechanical properties of wet conditioned starch/montmorillonite nanocomposite films. *Carbohydrate Polymer* 113: 569-576.

- Nasehi, J., Kadir, J. B., Zainal Abidin, M. A., Wong, M. Y. and Mahmodi, F. 2012. First report of Alternaria tenuissima causing leaf spot on eggplant in Malaysia. *American Phytopathological Society* 96: 1226.
- Nesterenko, A., Alric, I., Silvestre, F., and Durrieu, V. 2013. Vegetable proteins in microencapsulation: A review of recent interventions and their effectiveness. *Industrial Crops and Products* 42: 469–479.
- Ogrodowska, D., Tańska, M., and Brandt, W. 2017. The influence of drying process conditions on the physical properties, bioactive compounds and stability of encapsulated pumpkin seed oil. *Food and Bioprocess Technology* 10: 1265–1280.
- Palmgren, M., G., Edenbrandt, A., K., Vedel, S., E., Andersen, M., M., Landes, X., Østerberg, J., T., Falhof, J., Olsen, L., I, Christensen, S., B., Sandøe, P., Gamborg, C., Kappel, K., Thorsen, B., J and Pagh, P. 2015. Are we ready for back-to-nature crop breeding? *Trends in Plant Science* 20: 155–164.
- Paques, J. P., Linden E. V. D., Rijn C. J. M. V. and Sagis L. M. C. 2014. Preparation methods of alginate nanoparticles. Advances in Colloid and Interface Science 209: 163-171.
- Park, M.J., Choi, Y.J., Hong, S.B. and Shin, H.D. 2010. Genetic variability and mycohost association of *Ampelomyces quisqualis* isolates inferred from phylogenetic analyses of ITS rDNA and acting gene sequences. *Fungal Biology* 114: 235-247.
- Pasparakis, G., and Bouropoulos, N. 2006. Swelling studies and in vitro release of verapamil from calcium alginate and calcium alginate chitosan beads. *International Journal of Pharmaceutics* 323: 34-42.
- Paul, K. C., Chuang, Y.-H., Cockburn, M., Bronstein, Horvath, S. and Ritz, B. 2018. Organophosphate pesticide and differential genome-wide DNA methylation. *Science of Total Environment* 645: 1135-1143.
- Pavlidou, S.and Papaspyrides, C. D. 2008. A review on polymer-layered silicate nanocomposites. *Progress in Polymer Science* 33(12): 1119–1198.
- Perez, Z. C., Chet, I. and Nussinovitch, A. 2004. Irregular textual features of dried alginate-filler beads. *Food Hydrocolloids* 18: 249-258.
- Persoon, C.H. 1794. Disposita methodica fungorum. *Römer's Neues Magazin Botanik* 1: 81–128.
- Pill, W.G., Collins, C. M., Goldberger, B. and Gregory, N. 2009. Responses of non-primed or prime seed of 'marketmore 76' cucumber (*cucumis sativus L.*) slurry coated with trichoderma sprcies to planting in growth

infested with *phytium aphanidermatum*. Science Horticulture-Amsterdam 121: 54-62.

- Priyanthi, M. A., Kalpana, S. K. and Dinesh, R. K. 2009. Nature of organic fluidmontmorillonite interaction: An FTIR spectroscopy study. *Journal of Colloid and Interface Science* 337: 95-105.
- Punja, Z. K., 1985. The biology, ecology and control of *Sclerotium rolfsii*. *Annual Revision Phytopathology* 23: 97-127.
- Rassis, D. K., Saguy, I. S.and Nussinovitch, A. 2002. Collapses shrinkage and structural changes in dried alginate gels containing fillers. *Food Hydrocolloids* 16: 139-151.
- Rathore, S., Desai, P. M. Liew, C. V., Chan, L. W. and Heng, P. W. S. 2013. Microencapsulation of microbial cells. *Journal of Food Engineering*, 116: 369-381.
- Ray, S., Raychaudhuri, U. and Chakraborty, R. 2016. An overview of encapsulation of active compounds used in food products by drying technology. *Food Bioscience* 13: 76–83.
- Rekha, P.D., Wai, A.L., Arun, A.B. and Chiu, C.Y. 2007. Effect of free and encapsulated *Pseudomonas putida* CC-FR2-4 and *Bacillus subtilis* CC-pg104 on plant growth under gnotobiotic conditions. *Bioresource Technology* 80: 447-451.
- Rojan P. J., Tyagi, R. D., Prévost, D., Brar, S. K. and Surampalli, R.Y. 2010. Mycoparasitic *Trichoderma viride* as a biocontrol agent against Fusarium oxysporum f. sp. adzuki and *Pythium arrhenomanes* and as a growth promoter of soybean. *Crop Protection* 29: 1452-1459.
- Rosa, D. R. and Herrera, C. J. 2009. Evaluation of *Trichoderma* spp. as biocontrol agents against avocado white root rot. *Biological Control* 51: 66-71.
- Roy, A., Bajpai, J. and Bajpai, A. K. 2009. Dynamics of controlled release of chlorpyrifos from swelling and eroding biopolymeric microspheres of calcium alginate and starch. *Carbohydrate Polymers*, 76: 222-231.
- Ruckert, C., Blom, J., Chen, X.H., Reva, O. and Borriss, R., 2011. Genome sequence of Bacillus amyloliquefaciens type strain DSM7T reveals differences to plant associated Bacillus amyloliquefaciens FZB42. *Journal of Biotechnology* 155: 78–85.
- Sarhy-Bagnon, V., Lozano, P., Sacedo-Castaneda, G. and Roussos, S. 2000. Production of 6-pentyl-a-pyrone by *Trichoderma harzianum* in liquid and solid state cultures. *Process Biochemistry* 36: 103-109.

- Savary, S., Ficke, A., Aubertot, J.N. and Hollier, C. 2012. Crop losses due to diseases and their implications for global food production losses and food security. *Food Security* 4: 519-537.
- Sennoi, R., Singkham, N., Jogloy, S., Boonlue, S., Saksirirat, W., Kesmala, T. and Patanothai. 2013. Biological control southern stem rot caused by *Sclerotium rolfsii* using *Trichoderma harzianum* and arbcular mycorrhizalfungi on Juresalem Artichoke (*Helianthus tuberosus* L.). *Crop Protection* 54: 148-153.
- Shabeer, T.P.A., Banerjee, K., Jadhav, M., Girame, R., Utture, S., Hingmire, S. and Oulkar, D., 2015. Residue dissipation and processing factor for dimethomorph, famoxadone and cymoxanil during raisin preparation. *Food Chemi*stry 170: 180–185.
- Sheu, T.Y. and Marshall, R.T. 1993. Microencapsulation of lactobacilli in calcium alginate gels. *Journal of Food Science* 54: 557-561.
- Siddiqui, Y. and Sariah, M, S. 2009. Effect of Seed Bacterization on Plant Growth Response and Induction of Disease Resistance in Chilli. *Agricultural Sciences in China* 8:963-971.
- Soresh, M. and Harman, G.E. 2008. The relationship between increased growth and resistance induced in plants by root colonizing microbes. *Plant Signaling & Behaviors* 3(9): 237-239.
- Sun, D., Zhuo, T., Hu, X., Fan, X. and Zou H. 2017. Identification of Pseudomonas putida as biocontrol agent for tomato bacterial wilt disease. *Biological Control* 114:45-50.
- Szczech, M. and Maciorowski, R. 2016. Microencapsulation technique with organic additives for biocontrol agents. *Journal of Horticultural Research* 24: 111-112.
- Taha, M.O. Aiedeh, K.M. Al-Hiari, Y. and Al-Khatib, H. 2005. Synthesis of zinccrosslinked thiolated alginic acid beads and their in vitro evaluation as potential enteric delivery system utilizing folic acid as model drug. *Pharmazie* 60: 736–742.
- Tavassoli-Kafrani, E., Shekarchizadeh, H. and Masoudpour-Behabadi, M. 2016. Development of edible films and coatings from alginates and carrageenans. *Carbohydrate Polymers* 137:360–374.
- Toghueo, R. M. K., Eke, P., Zabalgogeazcoa, I., Aldana, B.R.V., Nana, L. W. And Boyom, F. F. 2016. Biocontrol and growth enhancement potential of two endophytic Trichoderma spp. From Terminalia catappa against the causative agent of Common Bean Root Rot (Fusarium solani). *Biological Control* 96:8-20.

- Vargas, W. A., Mukkherjee, P. K., Laughlin, D., Wiest, A., Maran-Diez, M. E. And Kenerley, C. M. 2014. Role of gliotoxin unthe symbiotic and pathogenic interaction of Trichoderma virens. *Microbiology* 160: 2319-2330.
- Vemmer, M. and Patel, A.V. 2013. Review of encapsulation methods suitable for microbial biological control agents. *Biological Control* 67: 380-389.
- Vinale, F., Sivasitharamparam, K., Ghisalberti, E. L., Marra, R., Woo, S. L. and Lorito, M. 2008. Trichoderma-plant-pathogent interaction. *Soil Biology* & *Biochemistry* 40: 1-10.
- Vos, C.M. Yang, Y., De Coninck, B. and Cammue B. P.A. 2014. Fungal (-like) biocontrol organisms in tomato disease control. *Biological Control* 84: 65-81.
- Wandrey, C., Bartkowiak, A. and Harding, S. E. 2010. Materials for encapsulation. In N. J. Zuidam, & V. A. Nedovic (Eds.). Encapsulation technologies for food active ingredients and food processing (pp. 31– 100). Dordrecht: Springer.
- Wang, S., Chen, Y., Liang, H., Chen, Y., Shi, M. and Wu, J. 2015. Intestinespecific delivery of hydrophobic bioactives from oxidized starch microspheres with an enhanced stability. *Journal of Agricultural and Food Chemistry* 63: 8669–8675.
- Yaseer, S. M., Mohamad, A. M., Suwardi, A. A., Adzemi, M. A., Nur Farah, H. M. and Norrizah, J. S. 2016. Economic of chili cultivation using fertigation technology in Malaysia. *Economic and Technology Management Review* 11a: 19-26.
- Zachaw, C., Berg, C., Muller, H., Monk, J., and Berg, G. 2016. Endemic plants harbour specific trichoderma communities with an exceptional potential for biocontrol phytopathogens. *Journal of Biotechnology* 235: 162-170.
- Zhang, J., Wang, Q., Xie, X., Li, X. and Wang, A. 2010. Preparation and swelling properties of pH-sensitive sodium alginate/layered double hydroxides hybrid beads for controlled release of diclofenac sodium. *Journal of Biomedical Materials Research Part A B* 92B: 205–214.
- Zhao, X., Han, Y., Tan, X.Q., Wang, J. and Zhou, Z.J. 2014. Optimization of antifungal lipopeptide production from Bacillus sp. BH072 by response surface methodology. Journal Microbiology 52 (4): 324–332.

BIODATA OF STUDENT

Fariz bin Adzmi was born on 23rd April 1981 in Gurun Kedah. He is the first of six siblings. He started his first education at Sekolah Kebangsaan Gurun (P) and secondary school at MRSM PDRM, Kulim, Kedah. He then furthered his study at Kolej Mara Kulim under UPM Matriculation and later continued his study at Universiti Putra Malaysia. He graduated in Bachelor of Science – Industrial Chemistry in 2003 and later Master of Science – Materials Science in 2007. Now he enrolled as Doctoral of Philosophy in Plant Disease Management at Institute of Tropical Agriculture and Food Security, Universiti Putra Malaysia.



PUBLICATION

Fariz A., Sariah, M., Hanafi, M. M., Azah, Y. (2012). Preparation, characterisation and viability of encapsulated *Trichoderma harzianum* UPM40 in alginate-montmorillonite clay. *Journal of Microencapsulation* 29 (3): 205-210.

Patent

6

Encapsulation of Bio-control Agent, PI2013700219, Malaysia Patent





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION :

TITLE OF THESIS / PROJECT REPORT :

DEVELOPMENT OF ENCAPSULATED Trichoderma harzianum UPM40 AGAINST

Sclerotium rolfsii ON CHILI PLANT

NAME OF STUDENT: FARIZ BIN ADZMI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (V)



(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from		until		
	(date)		(date)	

Approved by:

(Signature of Student) New IC No/ Passport No.: (Signature of Chairman of Supervisory Committee) Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]