



UNIVERSITI PUTRA MALAYSIA

**PRESERVATION, EXTRACTION AND ANALYTICAL METHODS FOR
FAECAL PROGESTERONE METABOLITE IN COWS AND IMMUNE
RESPONSE TO PREGNANOLONE HEMISUCCINATE IN RABBITS**

INNOCENT DAMUDU PETER

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By

INNOCENT DAMUDU PETER

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

July 2019

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UPM

In all your ways acknowledge Him and He shall direct your paths

Proverbs 3:6

DEDICATION

This thesis is dedicated to:

Kwasku Mada

my paternal grandmother: whose prayers for me are never ending

Prof. Peter M. Bzugu and Mrs. Lucy M. Peter

my parents: who have taught me to do the best I can possibly do and it will be enough and also making me believe that everything is possible as long as i have faith in it

Mary Adarju Innocent and Martha Kwasku Innocent

my children: who always tell me 'daddy, hurry up and return home'

Barr. (Mrs.) Munakur Samiyu Peter

my lovely wife: for her unrelenting daily prayers and priceless patience.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

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July 2019

Chairman : Abd Wahid Haron, PhD
Faculty : Veterinary Medicine

Monitoring reproductive function using faecal progesterone metabolite evaluations is a well-established technique in gazelles, horses, baboon, pigs, rhinoceros, elephants and should be extensively applied in domestic animals. Some factors however, prevent the application of this non-invasive technique to monitor reproduction in animals. These factors include paucity of practical information on stability of faecal progesterone metabolites under different preservation and extraction methods as well as sensitivity and specificity of analytical techniques. Therefore, general objective of this study was to evaluate preservation, extraction and analytical techniques for progesterone metabolite in faeces of cows; determine plasma progesterone and progesterone metabolite profiles in pregnant and non-pregnant cows; modify existing high performance liquid chromatography (HPLC) method for steroid hormones for the simultaneous detection of multiple progesterone metabolites in faeces of cows and to determine the immune responses and cellular changes following administration of low dose of pregnanolone hemisuccinate in rabbits. Progesterone metabolites were extracted from faecal samples using already established procedures and their concentrations together with plasma progesterone levels were determined using radioimmunoassay. A mobile phase made of distilled water and acetonitrile (70:30 v/v) and a stationary phase made up of C₁₈ column coupled to an Agilent HPLC 1100 series module was modified from existing HPLC method of steroid hormones to detect progesterone metabolites in faeces of cows. This modified HPLC method was hypothesized to be able to serve as an alternative to use of immunoassay on the analysis of progesterone metabolite in animals. Male rabbits were parenterally administered with pregnanolone hemisuccinate on day 0 and were given a booster dose on day 14 so as to produce group specific antibody to progesterone metabolites. Antibody response was determined using IgG and IgM evaluations using ELISA. The results show that there is no statistically significant difference in progesterone metabolite concentration in faeces that were dried at 70°C (9.33±5.05 ng/g), 50°C

(5.66±4.19 ng /g) or at 90°C (5.29±2.51 ng/g). Similarly, radioimmunoassay technique was used to determine the stability of progesterone metabolites in oven dried and fresh faecal samples. Faecal progesterone metabolite concentrations were found to be higher when extracted from oven-dried samples (2.06±1.58 ng /g) as compared to extraction from fresh faecal samples (0.49±0.30 ng /g). This difference was found to be statistically significantly (P<0.05). When un-preserved faeces were left exposed to environmental conditions for several days, progesterone metabolite concentrations therein were observed to rise from 11.04±7.68 ng/g at 0 hrs to maximum value of 31.71±10.33 ng /g at 48 hrs and thereafter decline to 36.62±12.30 ng/g after 216 hrs. There was no statistically significant difference (P>0.05) among these values. The relationship between faecal progesterone metabolites concentration and plasma progesterone concentration in cows were also investigated in this study. The results showed a correlation between plasma progesterone and faecal progesterone metabolites in pregnant (r=0.211, n=8, p=0.23) and in non-pregnant cows (r=0.209, n=8, p=0.35). A High-Performance Liquid Chromatography (HPLC) method for steroid hormones commonly employed in other species of animals was modified and used in this study for cows using 5β-pregnan-3α-ol-20-one and 3β-Hydroxy-5α-pregnan-20-one sulphate pyridine salt as internal standards. UV detection was employed to monitor the eluent from a stationary phase at an excitation wavelength of 254 nm and an emission wavelength of 360 nm. The linearity of the calibration curve was obtained in the concentration range of 500-7500 µg/µL for 5β-pregnan-3α-ol-20-one and 3-15 µg/µL for 3β-Hydroxy-5α-pregnan-20-one sulphate pyridine salt respectively. The co-efficient of determination was found to be 0.9977 for 5β-pregnan-3α-ol-20-one and 0.9776 for 3β-Hydroxy-5α-pregnan-20-one sulphate pyridine salt. The production of group specific antibodies for progesterone metabolites using the immunogen in rabbits was not achieved in this study. This could have been due to minute amount of the immunogen used or a defective bioconjugation method or due to inherent molecular properties of 5β-pregnan-3α-ol-20-one. Histopathological, haematological and biochemical parameters did not significantly differ between control and immunised rabbits in this study. This study provides practical and reliable information on faecal progesterone metabolite levels exposed to different storage, preservation and extraction parameters in cows. Best results for the analysis of faecal progesterone metabolites in cows are obtained when faeces are dried at 70°C and are extracted soon after voiding. The correlation observed between faecal progesterone metabolites and plasma progesterone concentrations shows that faecal progesterone metabolites concentration can be used for determination of reproductive status in cows. The HPLC method described in this study is simple, practical and rapid and can be applied for determination of progesterone metabolite concentration. It is therefore recommended that the information provided by this study be utilised in the non-invasive study of reproductive function in cows. These techniques are simple, reliable and can be performed within a short time. Group specific antibodies were not produced using 5β-pregnan-3α-ol-20-one-hemisuccinate, it is recommended that a modification in the bioconjugation method used or immunisation procedure be performed. Furthermore, other progesterone metabolites could be used to produce group specific antibodies and thereafter used in immunoassays to produce a more specific and sensitive analysis.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai menenuhi keperluan untuk ijazah Doktor Falsafah

**PEMULIHARAAN, PENGEKSTRAKAN DAN KAEDAH ANALISIS
METABOLIT PROGESTERON DALAM TINJA LEMBU DAN TINDAK
BALAS IMUN TERHADAP PREGNANOLONE HEMISUCCINATE DALAM
ARNAB**

Oleh

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Pemantauan fungsi pembiakan menggunakan metabolit progesteron tinja adalah teknik yang telah digunakan dalam beberapa spesis haiwan dan seharusnya digunakan secara meluas dalam haiwan domestik. Beberapa faktor menjadi penghalang untuk mengaplikasikan teknik tidak invasif ini dan menyelia pembiakan di kalangan haiwan. Faktor-faktor ini termasuk kekurangan maklumat praktikal mengenai kestabilan metabolit progesteron tinja dari segi kaedah pemeliharaan dan pengekstrakan yang berlainan serta kepekaan dan kekhususan teknik analisis.. Objectif kajian ini adalah: untuk menilai pemeliharaan, pengekstrakan dan teknik analisis metabolit progesteron dalam tinja lembu; menentukan progesteron plasma dan profil progestagen dalam lembu hamil dan tidak hamil; menghasilkan kaedah kromatografi cecair prestasi tinggi (HPLC) yang mudah dari kaedah lama yang telah ada untuk mengesan metabolit progesteron secara serentak dalam bentuk najis lembu dan untuk menentukan tindak balas imun dan perubahan sel selepas pemberian dos rendah ke pregnanolone hemisuccinate dalam arnab. Metabolit progesteron diekstrak daripada sampel tinja menggunakan prosedur yang telah ditetapkan dan kepekatan progesteron plasma dan metabolit ditentukan menggunakan radioimmunoassay. Fasa mudah alih yang diperbuat daripada air sulingan dan asetonitrile (70:30 v / v) dan satu fasa pegun yang terdiri daripada kolum C₁₈ dan modul siri Agilent HPLC 1100 untuk menghasilkan kaedah yang baru untuk mengesan metabolit progesteron dalam tinja lembu. Arnab jantan telah digunakan dengan suntikan parenteral pregnanolone hemisuccinate pada hari 0 dan dos penggalak telah disuntik pada hari ke 14 untuk menghasilkan antibodi khas kumpulan untuk metabolit progesteron. Tindak balas antibodi ditentukan menggunakan penilaian IgG dan IgM menggunakan ELISA. Hasilnya menunjukkan kepekatan metabolit progesterone yang lebih tinggi diperolehi apabila tinja dikeringkan pada suhu 70 ° C (9.33 ± 5.86 ng/g) berbanding suhu pengeringan pada suhu 50 ° C (7.37 ± 6.91 ng/g) atau 90 ° C (5.29 ± 2.51 ng/g). Perbezaan ini

bagaimanapun tidak ketara secara statistik ($P > 0.05$). Teknik radioimmunoassay digunakan untuk menentukan kestabilan metabolit progesteron dalam tinja segar yang dikeringkan dalam ketuhar. Kepekatan metabolit progesteron tinja didapati lebih tinggi apabila diekstrak dari sampel pengeringan oven (3.06 ± 3.12 ng/g) berbanding dengan sampel dari tinja segar (0.49 ± 0.30 ng/g). Perbezaan ini didapati ketara secara statistik ($P > 0.05$). Apabila tinja yang tidak dipulihara terdedah kepada keadaan sekitar, kepekatan metabolit progesterone meningkat dari 29.37 ± 37.07 ng / g pada 0 jam ke nilai maksimum 86.23 ± 73.20 ng / g pada 48 jam dan kemudian menurun ke 46.12 ± 38.04 ng / g selepas 216 jam. Tiada perbezaan ketara secara statistik ($P > 0.05$) di antara nilai-nilai ini. Hubungan antara kepekatan metabolit progesteron tinja dan kepekatan progesteron plasma dalam lembu juga diselidiki dalam kajian ini. Keputusan menunjukkan hubungan antara plasma progesteron dan metabolit progesteron faecal semasa mengandung ($r = 0.211$, $n = 8$, $p = 0.23$) dan dalam lembu tidak hamil ($r = 0.209$, $n = 8$, $p = 0.35$). Kaedah kromatografi cecair prestasi tinggi untuk hormone steroids yang digunakan untuk haiwan species lain telah diubahsuai dengan menggunakan garam pyridine sulfat 5β -pregnan- 3α -ol-20-one dan 3β -Hydroxy- 5α -pregnan-20-one sebagai piawai dalam. Pengesanan UV digunakan untuk memantau bahan pengelusi dari lajur pada panjang gelombang pengujaan 254 nm dan panjang gelombang pelepasan 360 nm. Faktor linear lengkungan yang diperolehi dalam julat kepekatan 500 - 7500 $\mu\text{g} / \mu\text{L}$ untuk 5β -pregnan- 3α -ol-20-one dan 3 - 15 $\mu\text{g} / \mu\text{L}$ untuk 3β -Hydroxy- 5α -pregnan-20-one garam pyridine. Koefisien penentuan didapati 0.9977 untuk 5β -pregnan- 3α -ol-20-one dan 0.9776 untuk 3β -Hidroksi- 5α -pregnan-20-one garam pyridine sulfat. Pengeluaran antibodi spesifik kumpulan untuk metabolit progesteron menggunakan 5β -pregnan- 3α -ol-20-one-hemisuccinate dalam arnab tidak dicapai dalam kajian ini. Ini mungkin disebabkan oleh bilangan kecil imunogen yang digunakan atau kaedah biokonjugasi yang rosak atau disebabkan sifat molekul yang wujud 5β -pregnan- 3α -ol-20-one. Histopatologi, hematologic dan parameter biokimia tidak menunjukkan perbezaan ketara antara kawalan dan arnab yang telah di imunisasikan dalam kajian ini. Kajian ini memberikan maklumat praktikal dan boleh dipercayai mengenai tahap metabolit progesteron tinja yang terpengaruh dengan pelbagai cara penyimpanan, pemeliharaan dan pengekstrakan dalam lembu. Hasil terbaik untuk analisis metabolit progesteron tinja pada lembu diperolehi apabila ia dikeringkan pada suhu 70 °C dan diekstrak secepat mungkin selepas dikeluarkan. Korelasi di antara metabolit progesteron tinja dan konsentrasi progesteron plasma menunjukkan bahawa konsentrasi metabolit tinja boleh digunakan untuk menentukan status pembiakan dalam lembu. Kaedah HPLC yang di terangkan dalam kajian ini adalah mudah, praktikal dan pantas dan boleh digunakan untuk menentukan konsentrasi metabolit progesteron. Oleh yang demikian, maklumat daripada kajian ini boleh digunakan untuk mengkaji fungsi pembiakan lembu secara tidak invasif. Teknik ini mudah, boleh dipercayai dan boleh dilakukan dalam masa yang singkat. Antibodi khas kumpulan tidak dihasilkan menggunakan 5β -pregnan- 3α -ol-20-one-hemisuccinate, dan disarankan agar pengubahsuaian biokonjugasi atau prosedur imunisasi dilakukan. Tambahan pula, metabolit progesteron lain boleh digunakan untuk menghasilkan antibodi spesifik kumpulan dan ia digunakan dalam immunoassays untuk menghasilkan analisis yang lebih spesifik dan sensitif.

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This Thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

%	Percentage
µL	Micro litre
µm	Micro meter
°C	Degree centigrade
¹²⁵ I	Iodine
ALT	Alanine amino transferase
ANOVA	Analysis of variance
APC	Antigen presenting cells
AST	Aspartate amino stransferase
AUP	Animal utilisation protocol
BSA	Bovine serum albumin
CFA	Complete Freunds Adjuvant
CBC	Complete blood count
CL	Corpus luteum
cpm	Count per minute
Da	Dalton
DLC	Differential leucocyte count
E ₂	Estradiol
EIA	Enzyme immuno assay
ELISA	Enzyme linked immunosorbent assay
fl	Femto litre
FSH	Follicle stimulating hormone
G	Guage
Glu	Glucose

GnRH	Gonadotrophin releasing hormone
h	Hour
H&E	Hematoxylin and Eosin
H ₀	Null hypothesis
H _A	Alternate hypothesis
Hb	Hemaglobin
HPLC	High performance liquid chromatography
IACUC	Institutional animal care and use committee
IgA	Immunoglobulin A
IgD	Immunoglobulin D
IgE	Immunoglobulin E
IgG	Immunoglobulin G
IgM	Immunoglobulin M
KDA	Kilo Dalton
KLH	Keyhole Limpet Hemolysin
KPa	Kilo pascal
LC	Liquid chromatography
LOD	Limit of detection
LLOQ	Lower limit of quantification
LH	Luteinizing hormone
M	Molar
MCH	Mean corpuscular haemoglobin
MCHC	Mean corpuscular haemoglobin concentration
MCV	Mean corpuscular volume
mg	Milligram
MHC	Major histocompatibility complex

mL	Milli litre
n	Number
ng	Nanogram
nm	Nano metre
p	Statistical significance level
PBS	Phosphate buffered saline
PCV	Packed cell volume
PGF _{2α}	ProstaglandinF _{2α}
pH	Potential of Hydrogen
PLT	Platelets
r ²	Coefficient of determination of linear regression
RBC	Red blood cells
RIA	Radio immunoassay
Rt	Retention time
SPSS	Statistical package for social scientist
Th	Helper T cells
TP	Total Protein
UV	Ultra violet
v/v	Volume per volume
WBC	White blood cells

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Progesterone, a steroid hormone made up of 21 carbon atoms, is produced by granulosa-lutein cells located within corpus luteum of the ovary. The hormone is released into the circulatory system soon after synthesis where it is transported principally to the uterus (Al-Asmakh, 2007; Barros et al., 2015). Progesterone plays a fundamental role in reproductive events in animals as it is directly associated with events leading to the establishment and maintenance of pregnancy such as ovulation and implantation (Conneely et al., 2002; Heyman et al., 2002; Al-Asmakh, 2007; Kornmatitsuk et al., 2007; Capezzuto et al., 2008; Carter et al., 2008). During pregnancy, additional amounts of progesterone is produced by the syncytiotrophoblast of the placenta and by the *zona fasciculata* and *zona reticularis* of the adrenal cortex (Adashi et al., 1996; Al-Asmakh, 2007; Barros et al., 2015). Plasma progesterone concentrations are often used as a reflection of ovarian function in animals and have been used to classify oestrus cycle into two distinct phases: the luteal (periods of progesterone dominance in plasma) and follicular (periods of declining plasma progesterone) phases (Kornmatitsuk et al., 2007). During luteal phase, progesterone is known to actively cause endometrial cellular proliferation in animals in preparation to implantation following an eventual mating and subsequent fertilisation (Al-Asmakh, 2007). Plasma progesterone levels remain elevated during luteal phase and pregnancy. This elevated progesterone levels have been used to diagnose pregnancy or monitor oestrous cycle in animals (Masunda et al., 1999; Carter et al., 2008).

For a successful determination of ovarian function in animals, steroid hormones such as progesterone are evaluated from serially collected blood samples (Nugraha et al., 2017). However, the repeated collection of blood samples is not always possible in animals (Schwarzenberger et al., 1997). Non-invasive samples such as hair, urine, milk and faeces are reported to have quantifiable amount of steroid hormone that could serve diagnostic purposes (Kumar et al., 2013). However, there are some limitations in utilising such non-invasive samples for monitoring reproductive function in animals. Hair samples are only suitable for determination of long-term hormone levels in animals and are not suitable for short term evaluation of hormone levels (Ventrella et al., 2018). Collection of urine samples in animal will require animal restraint for insertion of catheters and this eventually limit the use of this type of sample (Kumar et al., 2013). Milk samples on the other hand are limited to lactating animals and cannot be used for heifers or dry-off animals (Kumar et al., 2013). Furthermore, animal handling and restraint to obtain samples may potentially because capture induced-stress and could affect results of hormone analysis. Faecal samples however can be collected repeatedly in animals with ease and without causing any danger, pain or distress to the subject (Gholib et al., 2017). Furthermore, progesterone metabolites are mainly excreted through the faeces of animals (Schwarzenberger et al., 1996; Kumar et al., 2013; Gholib et al., 2017). Biological samples collected without restraint

provide precise amounts of faecal steroid hormones. These advantages therefore make faeces to be a practical choice for repeated sample collection for analysing steroid hormones for monitoring reproductive function in animals (Schwarzenberger et al., 1996; Ziegler and Wittwer, 2005).

The terms “faecal progesterone”, “progestogens”, “progestins” and “progestagens” have been used rather loosely in literature when referring to progesterone metabolites in faeces of animals (Van der Goot et al., 2015; Mithileshwari et al., 2016; Cao et al., 2016; Davis, 2017; Gascoigne et al., 2017; Silvestre et al., 2017). The term progesterone is strictly reserved for a natural hormone that is synthesized and released by the corpus luteum in the ovary. Since natural progesterone is not present in feces, it becomes proper that the term faecal progesterone metabolites be used (Schwarzenberger et al., 1997). Progestogens refers to a group of naturally occurring progesterone metabolites while progestins is used to describe synthetic progestational agents that are designed to target and bind to progesterone receptors (Sitruk-Ware, 2008). The term “progestagens” appears to be a better terminology for faecal progesterone metabolites as the extent to which faecal progesterone metabolite possess progestational activity is not fully understood.

The half-life of progesterone is reported to be approximately 5 minutes, after which the hormone is extensively metabolised into several metabolites by the liver (Schumacher and Robert, 2002; Al-Asmakh, 2007). Progesterone is first converted into 5 α -dihydro-progesterone by cytochromes P450 enzyme (Shimada et al., 1994) and is thereafter metabolised by the liver to yield pregnanolone and several other metabolites before their eventual excretion in faeces (Lemoine et al., 1993; Shimada et al., 1994; Schwarzenberger et al., 1997). Earlier studies on progesterone metabolites were performed to elucidate on the metabolism and route of excretion of natural progesterone. Such studies have now paved ways for further research that have now demonstrated the application of faecal progesterone analysis to determine reproductive function in animals (Hirata and Mori, 1995; Capezzuto et al., 2008). Data from several studies have established that faecal progesterone metabolites concentrations exhibit a similar pattern to plasma progesterone in animals (Schwarzenberger et al., 1996; 1997; Masunda et al., 1999; Capezzuto et al., 2008; Yimer et al., 2012). By using immunoassay, several studies have described a positive correlation between plasma progesterone and faecal progesterone metabolite concentrations. This has led several researchers to utilise faecal progesterone metabolites evaluations for characterizing reproductive status of animals (Desaulniers et al., 1989; Schwarzenberger et al., 1992; Masunda et al., 1999; Graham et al., 2001; Isobe et al., 2005a; Gan and Patel, 2013). Faecal progesterone metabolite evaluations have now been successfully used to determine the onset of puberty, corpus luteum function, pregnancy, abortion and in monitoring therapy of the reproductive system in an ever-expanding number of animal species (Desaulniers et al., 1989; Hirata and Mori, 1995; Borjesson et al., 1996; Wasser et al., 1996; Masunda et al., 1999; Pereira et al., 2006; Ncube et al., 2011).

Faecal steroid analysis is a potent tool that can provide information on physiology and reproductive status in a wide range of animal species (Capezzuto et al., 2008). The use of faecal progesterone metabolites for monitoring reproductive function in free ranging animals have been extensively applied in free ranging terrestrial animals such as rhinoceros (Schwarzenberger et al., 2000), cheetah (Terio et al., 2002), lion (French et al., 2003), boar (Macchi et al., 2010), zebra (Ncube et al., 2011), panda (Deng et al., 2014), deer (Mithileshwari et al., 2016; Wang et al., 2016), tiger (Panda et al., 2017), jaguar (Conforti et al., 2017), gazelles (Wojtusik et al., 2017), parrots (Pereira et al., 2018) and even sea animals such as whales (Larson et al., 2003) and otters (Rolland et al., 2005). In these animal species, many techniques for analysing progesterone metabolites have been established and are routinely used. Generally, fresh faecal samples are used in analysis of faecal progesterone metabolites in terrestrial animals. In aquatic animals however, drying of faecal samples usually precede analysis of faecal progesterone. Although analysis of progesterone metabolites in terrestrial animals is done using fresh faecal samples, there is little information on the concentration of faecal progesterone metabolite in fresh and dried faecal samples in domestic ruminants. Studies are required to elucidate on the value of using fresh faecal samples for analysis progesterone metabolites.

Despite the widespread application of non-invasive techniques for monitoring reproduction in many animal species, faecal progesterone metabolites analysis has not been extensively applied to monitor reproductive function in farm animals. This could partly be due to a failure in establishing newer analytical techniques for animals. Use of non-invasive samples for quantifying faecal steroid hormone could greatly improve animal welfare. The development and validation of a new analytical method is the first step before adopting any method in a defined animal species. Traditionally, immunoassay systems using antibodies to progesterone are used to determine the total concentration of progesterone metabolite in faeces of animals. This is due to the widespread availability of antibodies to natural progesterone and because such antibodies have abilities to cross react with progesterone metabolite molecules in feces. Due to this reason, progesterone metabolites are easily quantified. Antibodies against any of several progesterone metabolites have not been produced commercially. The development of immunoassay systems with antibodies to progesterone metabolites would have more specificity and sensitivity as compared to the more readily available immunoassay systems made with antibodies to progesterone. Among several factors to be considered for production of antibodies is immunogen preparation as most steroid hormone metabolites have low molecular weight and are non-immunogenic on their own. High Performance Liquid Chromatography (HPLC) methods for progesterone have been previously described in literature for steroid hormone and their metabolites. HPLC methods for progesterone metabolites using non-faecal samples are earlier being described. Newer and highly specific methods for detection of individual faecal progesterone metabolites therefore become necessary which will improve the methodologies employed in non-invasive methods of monitoring reproductive function in animals.

To fully utilize the faecal progesterone metabolite analysis in farm animals for monitoring reproductive function, optimum preservation, extraction and storage conditions needs to be determined and evaluated (Pappano et al., 2010). One important challenge in the analysis of faecal steroids is the preservation and storage of faecal samples before analysis (Nugraha et al., 2017). Faecal bacterial enzymes have been reported to alter the composition and concentration of steroids hormone metabolite when faeces are left unpreserved (Khan et al., 2002; Nugraha et al., 2017). Post defecation changes in progesterone concentration in faeces of animals have been reported. This has been mainly attributed to the activity of faecal bacteria or bacterial enzymes (Kumar et al., 2013). In cows, there is paucity of information on the fate of progesterone metabolite in voided faeces over time. This therefore necessitate the preservation of faecal samples in preservatives or storage at sub-zero temperatures before analysis is indicated (Ziegler et al., 2000; Nugraha et al., 2017). This is however not always achievable (Khan et al., 2002). Drying of faecal samples using oven or using solar box cooker are alternatives to freezing (Galama et al., 2004). Other alternative methods are storage in ethanol or sodium azide and kept at room temperature (Khan et al., 2002). Several studies have explained the degradative activities of faecal bacteria on progesterone metabolites after voiding (Schwarzenberger et al., 1996; Kumar et al., 2013). However, the short- and long-term degradative effects of faecal bacteria on progesterone metabolites in cow faeces remains to be fully understood. This information may allow an informed decision as to how long can voided faeces from cows be of analytical use.

1.2 Problem statement

The assessment of reproductive function is traditionally performed using blood samples. Blood sample collection in animals requires restraint which is associated with both pain and stress. Animal restraint procedures affects hormone concentration in animals and could give inaccurate results. Because of this, blood sample analysis is now fast giving way to non-invasive sample analysis such as faeces in animals to determine steroid hormone levels. Within the last few decades, there has been an increasing interest in the use of non-invasive faecal steroid analytical methods for monitoring reproduction in animals just as concerns about animal welfare grows. However, before adopting a new non-invasive faecal steroid methodology in a new species, the preservation, extraction and analytical methods for faecal steroids need to be assessed. This is because several methodologies for preservation, extraction and storage methods are available and have been established in some animal species. Equally, multiple types of HPLC and immunoassay methods are utilised in the determination of total progesterone metabolites in biological samples. HPLC methods and immunoassay system determines the concentration of individual progesterone metabolite in faecal samples. Commercially available immunoassay systems prepared with antibodies to natural progesterone are acceptable immune assay systems for quantifying faecal progesterone metabolites. These kits have significant cross reactivity with many progesterone metabolites and are thus rightly used. However, production of antibodies against pregnanolone as a group specific antibody would greatly provide higher analytical precision for a better analytical result in future.

1.3 Justification of the study

The determination of reproductive status of animals by faecal steroid analysis is one of valuable tools for efficient management of animals. This technique is safe, non-invasive, and simple and could be used as an alternative to invasive monitoring of reproductive function in animals. Faecal progesterone metabolite analysis is therefore explored in this study as a non-invasive tool to monitor reproductive status for domestic animals.

1.4 Research hypotheses

The general hypothesis for this study is that under appropriate preservation and extraction methods, faecal progesterone metabolite evaluations can be used to monitor the oestrous cycle and pregnancy in cows.

1.5 General objectives

The general objective of this study is to evaluate and determine best preservation and extraction methods for progesterone metabolites in faeces of cows. The study is also designed to determine immune response towards pregnanolone hemisuccinate using an animal model (rabbit).

1.5.1 Specific objectives

The specific objectives of this study are to:

1. evaluate preservation and extraction methods for progesterone metabolites in faeces of cows
 - i. determine the optimum oven drying temperature for faeces that would be used for progesterone metabolites analysis in cows
 - ii. to determine the post defecation changes in progesterone metabolite levels in unpreserved feces
 - iii compare the concentrations of faecal progesterone metabolites concentrations in fresh and oven-dried faecal samples
2. determine the relationship between plasma progesterone concentration and progesterone metabolite concentration in faeces of pregnant and non-pregnant cows.
3. apply a high-performance liquid chromatography method for the detection of progesterone metabolites in faeces of cows.
 - i. demonstrate the application of this method for the analysis of progesterone metabolites in faecal samples.
4. determine immune response as well as haematological, biochemical and histopathological changes following parenteral administration of pregnanolone hemisuccinate in rabbits.

REFERENCES

- Abdel-Khalik, J., Björklund, E. and Hansen, M. (2013). Simultaneous determination of endogenous steroid hormones in human and animal plasma and serum by liquid or gas chromatography coupled to tandem mass spectrometry. *Journal of Chromatography B*, 928:58-77.
- Actor, J.K. (2014). Assessment of Immune parameters and immunodiagnostics In Introductory Immunology: *Basic Concepts for Interdisciplinary Applications*. Elsevier AP.
- Adachi, S., Yamada, S., Takatsu, Y., Matsui, H., Kinoshita, M., Takase, K. and Tsukamura, H. (2007). Involvement of anteroventral periventricular metastin/kisspeptin neurons in estrogen positive feedback action on luteinizing hormone release in female rats. *Journal of Reproduction and Development*, 53(2):367-378.
- Adamczyk, M., Buko, A., Chen, Y.Y., Fishpaugh, J.R., Gebler, J.C. and Johnson, D.D. (1994). Characterization of protein-hapten conjugates. 1. Matrix-assisted laser desorption ionization mass spectrometry of immuno BSA-hapten conjugates and comparison with other characterization methods. *Bioconjugate Chemistry*, 5(6):631-635.
- Adams, G.P., Matteri, R.L., Kastelic, J.P., Ko, J.C.H. and Ginther, O.J. (1992). Association between surges of follicle-stimulating hormone and the emergence of follicular waves in heifers. *Journal of Reproduction and Fertility*, 94(1): 177-188.
- Adashi, E.Y., Rock, J.A. and Rosenwaks, Z. (1996). Reproductive endocrinology, surgery, and technology. Lippincott-Raven. Philadelphia
- Adeyemo, O. and Heath, E. (1980). Plasma progesterone concentration in *Bos taurus* and *Bos indicus* heifers. *Theriogenology*, 14(6): 411-420.
- Aguilar-Pérez, C., Ku-Vera, J., Centurión-Castro, F. and Garnsworthy, P.C. (2009). Energy balance, milk production and reproduction in grazing crossbred cows in the tropics with and without cereal supplementation. *Livestock Science*, 122(2-3):227-233.
- Ahuja-Aguirre, C., López-deBuen, L., Rojas-Maya, S. and Hernández-Cruz, B.C. (2017). Progesterone and estradiol profiles in different reproductive stages of captive collared peccary (*Pecari tajacu*) females assessed by faecal metabolites. *Animal Reproduction Science* 180: 121-126.
- Al-Asmakh, M. (2007). Reproductive functions of progesterone. *Middle East Fertility Society Journal*, 12: 147-152.

- Alila, H.W. and Hansel, W. (1984). Origin of different cell types in the bovine corpus luteum as characterized by specific monoclonal antibodies. *Biology of Reproduction*, 31(5): 1015-1025.
- Allenmark, S., Hammar, M. and Lindström, E. (1981). Combined paper and reversed-phase high-performance liquid chromatography method for the study of pregnenolone and progesterone metabolites. *Journal of Chromatography B: Biomedical Sciences and Applications*, 224(3): 399-405.
- Alvarez, P., Spicer, L.J., Chase, C.C., Payton, M.E., Hamilton, T.D., Stewart, R.E. and Wettemann, R.P. (2000). Ovarian and endocrine characteristics during an estrous cycle in Angus, Brahman, and Senepol cows in a subtropical environment. *Journal of Animal Science*, 78 (5): 1291-1302.
- Ando, T., Kamimura, S. and Hamana, K. (2004). Estrous synchronization using an intravaginal progesterone device in combination with GnRH or estradiol benzoate characterized by the initial ovarian conditions in Japanese black cows. *Journal of Veterinary Medical Science*, 66 (12): 1497-1502.
- Andréen, L., Spigset, O., Andersson, A., Nyberg, S., and Bäckström, T. (2006). Pharmacokinetics of progesterone and its metabolites allopregnanolone and pregnanolone after oral administration of low-dose progesterone. *Maturitas*, 54(3): 238-244.
- Ashar, B. and Turcotte, L.R. (1981). Analyses of longest IAB implant in human patient (327 days). *ASAIO Journal*, 27(1): 372-379.
- Baltic, M., Jenni- Eiermann, S., Arlettaz, R. and Palme, R. (2005). A non-invasive technique to evaluate human- generated stress in the black grouse. *Annals of the New York Academy of Sciences*: 1046(1): 81-95.
- Baradaran, B., Majidi, J., Hassan, Z. M. and Abdolalizadeh, J. (2006). Large scale production and characterization of anti-human IgG monoclonal antibody in peritoneum of Balb/c mice. *American Journal of Biochemistry and Biotechnology*, 1(4): 190-3.
- Barros, L., Tufik, S. and Andersen, M. (2015). The role of progesterone in memory: an overview of three decades. *Neuroscience and Biobehavioral Reviews*, 49: 193-204.
- Baruselli, P.S., Reis, E.L., Marques, M.O., Nasser, L.F. and Bó, G.A. (2004). The use of hormonal treatments to improve reproductive performance of anoestrus beef cattle in tropical climates. *Animal Reproduction Science*, 82: 479-486.
- Bauminger, S., Kohen, F. and Lindner, H.R. (1975). Steroids as haptens: optimal design of antigens for the formation of antibodies to steroid hormones. In *Recent Advances in Steroid Biochemistry*, (pp. 739-747).
- Beehner, J.C. and Whitten, P.L. (2004). Modifications of a field method for faecal steroid analysis in baboons. *Physiology and Behavior*, 82(2-3): 269-277.

- Beiser, S.M., Erlanger, B.F., Agate, F.J. and Lieberman, S. (1959). Antigenicity of steroid-protein conjugates. *Science*, 129(3348): 564-565.
- Benbow, A.L. and Waddell, B.J. (1995). Distribution and metabolism of maternal progesterone in the uterus, placenta, and fetus during rat pregnancy. *Biology of Reproduction*, 52(6): 1327-1333.
- Bier, H., Hoffmann, T., Hauser, U., Wink, M., Ochler, M., Kovar, A., Muser, M. and Knecht, R. (2001). Clinical trial with escalating doses of the antiepidermal growth factor receptor humanized monoclonal antibody EMD 72000 in patients with advanced squamous cell carcinoma of the larynx and hypopharynx. *Cancer Chemotherapy Pharmacology*, 47: 519–524
- Berkeley, E.V., Kirkpatrick, J.F., Schaffer, N.E., Bryant, W.M. and Threlfall, W.R. (1997). Serum and faecal steroid analysis of ovulation, pregnancy, and parturition in the black rhinoceros (*Diceros bicornis*). *Zoo Biology: Published in affiliation with the American Zoo and Aquarium Association*, 16(2): 121-132.
- Bielohuby, M., Popp, S. and Bidlingmaier, M. (2012). A guide for measurement of circulating metabolic hormones in rodents: Pitfalls during the pre-analytical phase. *Molecular Metabolism* 1: 47-60.
- Blavy, P., Derks, M., Martin, O., Höglund, J.K. and Friggens, N.C (2016). Overview of progesterone profiles in dairy cows. *Theriogenology* 86 (4): 1061-1071.
- Bochskanl, R., Thie, M., and Kirchner, C. (1989). Active immunization of rabbits against progesterone: Increase in hormone levels, and changes in metabolic clearance rates and in genital tract tissues. *Journal of Steroid Biochemistry*, 33(3): 349-355.
- Borjesson, D. L., Boyce, W. M., Gardner, I. A., DeForge, J. and Lasley, B. (1996). Pregnancy detection in bighorn sheep (*Ovis canadensis*) using a faecal -based enzyme immunoassay. *Journal of Wildlife Diseases*, 32(1): 67-74.
- Bosze, P. (1988) Antibodies against steroids. *Human Reproduction*, 3(1): 63-68.
- Brockman, D.K. and Whitten, P.L. (1996). Reproduction in free-ranging *Propithecus verreauxi*: estrus and the relationship between multiple partner matings and fertilization. *American Journal of Physical Anthropology*, 100: 57-69.
- Broes, A., and LeBlanc, S.J. (2014). Comparison of commercial progesterone assays for evaluation of luteal status in dairy cows. *The Canadian Veterinary Journal*, 55(6): 582.
- Brooks, R.M., Denniston, D.J., Bruemmer, J.E., Nett, T.M. and McCue, P.M. (2013). Development of a direct (non-extracted) enzyme immunoassay for measurement of serum progesterone in mares. *Journal of Equine Veterinary Science*, 33(5): 371.

- Brown, J.L., Graham, L.H., Wielebnowski, N., Swanson, W.F., Wildt, D.E. and Howard, J.G. (2001). Understanding the basic reproductive biology of wild felids by monitoring of faecal steroids. *Journal of Reproduction and Fertility. Supplement*, 57: 71-82.
- Brown, J.L., Wasser, S., Wildt, D., Graham, L.H. and Monfort, S. (1997). Faecal steroid analysis for monitoring ovarian and testicular function in diverse carnivore, primate, and ungulate species. *Zeitschrift Fur Säugetierkunde*, 62: 27-31.
- Byszewska- Szpocińska, E. and Markiewicz, A. (2006). New RIA kit for the determination of progesterone in cows' milk. *Journal of Immunoassay and Immunochemistry*, 27(3): 279-288.
- Cao, X., Wei, H., Xue, H., Li, X., Zhao, W., Xu, C., and Xu, B. (2016). Faecal progesterin concentrations as an indicator of reproductive success in American mink. *Animal Reproduction Science*, 165: 11-16.
- Capezzuto, A., M. O. M. Chelini, E. C. G. Felipe, and C. A. Oliveira. (2008). "Correlation between Serum and Faecal Concentrations of Reproductive Steroids throughout Gestation in Goats." *Animal Reproduction Science*, 103(1-2):78-86.
- Carleton, H.M. and Drury, R.A.B. (1957). Histological technique for normal and pathological tissues and the identification of parasites. (3rd edit). London Oxford University Press.
- Carter, F., Forde, N., Duffy, P., Wade, M., Fair, T., Crowe, M., Evans, A., Kenny, D., Roche, J. and Lonergan, P. (2008). Effect of increasing progesterone concentration from Day 3 of pregnancy on subsequent embryo survival and development in beef heifers. *Reproduction, Fertility and Development* 20:368-375.
- Cavigelli, S.A. and Pereira, M.E. (2000). Mating season aggression and faecal testosterone levels in male ring-tailed lemurs (*Lemur catta*). *Hormones and Behavior*, 37: 246-255.
- Čebulj-Kadunc, N., Snoj, T. and Cestnik, V. (2000). Faecal gestagen, serum and milk progesterone concentrations in ewes of the Jezersko-Solchava breed. *Acta Veterinaria Brno*, 69(1): 33-37.
- Chen, K. and Cerutti, A. (2011). The function and regulation of immunoglobulin D. *Current Opinion in Immunology*, 23(3): 345-352.
- Chung, W.B., Cheng, W.F., Wu, L.S. and Yang, P.C. (2002). The use of plasma progesterone profiles to predict the reproductive status of anestrus gilts and sows. *Theriogenology*, 58(6): 1165-1174.

- Colazo, M.G., Ambrose, D.J., Kastelic, J.P. and Small, J.A. (2008). Comparison of 2 enzyme immunoassays and a radioimmunoassay for measurement of progesterone concentrations in bovine plasma, skim milk, and whole milk. *Canadian Journal of Veterinary Research*, 72(1): 32.
- Conforti, V.A., Bravo, N.R.S., Moreira, M.R., Villar, E.C., Paulo, O.L.O.H, Veneziani, R.C.S. and Silva, M.A. (2017). High-performance liquid chromatography as a novel tool for assessing ovarian function in jaguars (*Panthera onca*): development and validation of the method and quantification of ovarian steroids. *Animal Reproduction*, 14: 361-361.
- Conneely, O.M., Mulac-Jericevic, B., Demayo, F., Lydon, J.P. and O'Malley, B.W. (2002). Reproductive functions of progesterone receptors. *Recent progress in Hormone Research*, 57: 339-356.
- Curtis, D.J., Zaramody, A., Green, D.I. and Pickard, A.R. (2000). Non-invasive monitoring of reproductive status in wild mongoose lemurs (*Eulemur mongoz*). *Reproduction, Fertility and Development*, 12: 21-29.
- Czekala, N. M. and Callison, L. (1996). Pregnancy diagnosis in the black rhinoceros (*Diceros bicornis*) by salivary hormone analysis. *Zoo Biology: Published in affiliation with the American Zoo and Aquarium Association*, 15(1): 37-44.
- D'Angelo, V., Tessari, F., Bellagamba, G., De Luca, E., Cifelli, R., Celia, C., and Locatelli, M. (2016). Microextraction by packed sorbent and HPLC–PDA quantification of multiple anti-inflammatory drugs and fluoroquinolones in human plasma and urine. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 31(sup3): 110-116.
- Davidian, E., Benhaiem, S., Courtiol, A., Hofer, H., Höner, O. P. and Dehnhard, M. (2015). Determining hormone metabolite concentrations when enzyme immunoassay accuracy varies over time. *Methods in Ecology and Evolution*, 6(5): 576-583.
- Davis, M. P. (2017). Characterized by non-invasive endocrine monitoring. *The Pygmy Hippopotamus*, 137.
- Delahaut, P. (2017). Immunisation—choice of host, adjuvants and boosting schedules with emphasis on polyclonal antibody production. *Methods*, 116: 4-11.
- Delanghe, J. R. and Speeckaert, M. M. (2011). Creatinine determination according to Jaffe—what does it stand for?. *Nephrology Dialysis Transplantation Plus*, 4(2): 83-86.
- Deng, H., Liu, S., Jin, X., Ge, X., He, L., Liu, G. and Hu, D. (2014). Research on methods of preserving faecal steroid hormones in giant panda (*Ailuropoda melanoleuca*). *North-Western Journal of Zoology*, 10(1): 210-216.

- Desaulniers, D.M., Goff, A.K., Betteridge, K.J., Rowell, J.E. and Flood, P.F. (1989). Reproductive hormone concentrations in faeces during the oestrous cycle and pregnancy in cattle (*Bos taurus*) and muskoxen (*Ovibos moschatus*). *Canadian Journal of Zoology* 67: 1148-1154.
- Deshpande, S.S. (2012). *Enzyme immunoassays: from concept to product development*. Springer Science and Business Media.
- Díaz- Cruz, M. S., López de Alda, M. J., López, R. and Barceló, D. (2003). Determination of estrogens and progestogens by mass spectrometric techniques (GC/MS, LC/MS and LC/MS/MS). *Journal of Mass Spectrometry*, 38(9): 917-923.
- Diskin, M.G., Mackey, D.R., Roche, J.F. and Sreenan, J.M. (2003). Nutrition and ovarian follicle function in cattle. *Animal Reproduction Science*. 78: 345-370.
- Diskin, M. G., and Kenny, D. A. (2016). Managing the reproductive performance of beef cows. *Theriogenology*, 86(1): 379-387.
- Djebaili, M., Hoffman, S. and Stein, D. (2004). Allopregnanolone and progesterone decrease cell death and cognitive deficits after a contusion of the rat pre-frontal cortex. *Neuroscience*, 123: 349-359.
- Dolan, J. (2000). Stainless steel surfaces in LC systems, Part I-Corrosion and erosion. *LC GC North America*, 18(6): 600-608.
- Dolomatov, S.I., Zukow, W., Atmazhov, I.D., Muszkieta, R. and Skaliy, A. (2012). The use of hormones indicators in human saliva in diagnosing parodontitis in pregnant women. *Indian Journal of Human Genetics*, 18(3): 305.
- Drupt, F., Paris, M., Frydman, A. and Leclerc, M. (1974). Serum albumin assay by bromocresol green method: application to different automatic apparatus. In *Annales Pharmaceutiques Francaises*, 32 (5): 249.
- Dubreuil, O., Bossus, M., Graille, M., Bilous, M., Savatier, A., Jolivet, M. and Ducancel, F. (2005). Fine tuning of the specificity of an anti-progesterone antibody by first and second sphere residue engineering. *Journal of Biological Chemistry*, 280(26): 24880-24887.
- Endresen, M. J., Haug, E., Åbyholm, T. and Henriksen, T. (1990). The source of cholesterol for progesterone synthesis in cultured preovulatory human granulosa cells. *Acta Endocrinologica*, 123(3): 359-364.
- Erlanger, B. F. (1980). The preparation of antigenic Hapten-Carrier conjugates: A survey. in *Methods in enzymology*, 70: 85-104. *Academic Press*.
- Fagarasan, S. and Macpherson, A. J. (2015). The regulation of IgA production. In *Mucosal Immunology (Fourth Edition)*. Elsevier, Oxford, GB. pp. 471-484.

- Fahmi, H.A., Williamson, N.B., Tibary, A. and Hegstad, R.L. (1985). The influence of some sample handling factors on progesterone and testosterone analysis in goats. *Theriogenology*, 24(2): 227-233.
- Fantl, V.E., Wang, D.Y. and Whitehead, A.S. (1981). Production and characterisation of a monoclonal antibody to progesterone. *Journal of Steroid Biochemistry*, 14(4): 405-407.
- Farin, P.W. and Estill, C.T. (1993). Infertility due to abnormalities of the ovaries in cattle. *Veterinary Clinics of North America: Food Animal Practice*, 9(2):291-308.
- Fasciglione, G.F., Marini, S., Bannister, J.V. and Giardina, B. (1996). Hapten—Carrier Interactions and Their Role in the Production of Monoclonal Antibodies against Hydrophobic Haptens. *Hybridoma*, 15(1):1-9.
- Fatnassi, M., Hammadi, M. and Khorchani, T. (2013). Effects of storage temperature and time on faecal progesterone concentration in camel (*Camelus dromedarius*). *Emirates Journal of Food and Agriculture* 25: 301.
- Flaherty, D. (2014). *Immunology for Pharmacy-E-Book*. Elsevier Health Sciences.
- Fodey, T.L., Greer, N.M. and Crooks, S.R. (2009). Antibody production: Low dose immunogen vs. low incorporation hapten using salmeterol as a model. *Analytica Chimica Acta*, 637(1-2): 328-332.
- Forde, N., Beltman, M.E., Lonergan, P., Diskin, M., Roche, J.F. and Crowe, M.A. (2011). Oestrous cycles in *Bos taurus* cattle. *Animal Reproduction Science*, 124 (3-4): 163-169.
- Forde, N., Carter, F., Fair, T., Crowe, M.A., Evans, A.C.O., Spencer, T.E. and Lonergan, P. (2009). Progesterone-regulated changes in endometrial gene expression contribute to advanced conceptus development in cattle. *Biology of Reproduction*, 81(4): 784-794.
- Foster, J.P., Lamming, G. E. and Peters, A.R. (1980). Short-term relationships between plasma LH, FSH and progesterone concentrations in post-partum dairy cows and the effect of Gn-RH injection. *Journal of Reproduction and Fertility*, 59(2): 321-327.
- Frandsen, R.D., Wilke, W.L. and Fails, A.D. (2009). *Anatomy and physiology of farm animals*. John Wiley and Sons. Singapore. 7th Edition.
- Free, A. H. (1963). Enzymatic determinations of glucose. In *Advances in Clinical Chemistry*, (6): 67-96. Elsevier.
- Freeman, E.W., Purdy, R.H., Coutifaris, C., Rickels, K. and Paul, S.M. (1993). Anxiolytic metabolites of progesterone: correlation with mood and performance measures following oral progesterone administration to healthy female volunteers. *Neuroendocrinology*, 58(4): 478-484.

- French, J.A., Bales, K.L., Baker, A.J. and Dietz, J.M. (2003). Endocrine monitoring of wild dominant and subordinate female *Leontopithecus rosalia*. *International Journal of Primatology*, 24(6), 1281-1300.
- Friggens, N.C., Bjerring, M., Ridder, C., Højsgaard, S. and Larsen, T. (2008). Improved detection of reproductive status in dairy cows using milk progesterone measurements. *Reproduction in Domestic Animals*, 43: 113-121.
- Furr, B.J.A. (1973). Radioimmunoassay of progesterone in peripheral plasma of the domestic fowl in various physiological states and in follicular venous plasma. *Acta Endocrinologica*, 72(1): 89-100.
- Galama, W.T., Graham, L.H. and Savage, A. (2004). Comparison of faecal storage methods for steroid analysis in black rhinoceroses (*Diceros bicornis*). *Zoo Biology* 23: 291-300.
- Galbreath, C.W., Scholljegerdes, E.J., Lardy, G.P., Odde, K.G., Wilson, M.E., Schroeder, J.W. and Vonnahme, K.A. (2008). Effect of feeding flax or linseed meal on progesterone clearance rate in ovariectomized ewes. *Domestic Animal Endocrinology*, 35(2): 164-169.
- Gan, S.D. and Patel, K.R. (2013). Enzyme immunoassay and enzyme-linked immunosorbent assay. *Journal of Investigative Dermatology*, 133(9): e12.
- Gao, W., Stalder, T. and Kirschbaum, C. (2015). Quantitative analysis of estradiol and six other steroid hormones in human saliva using a high throughput liquid chromatography–tandem mass spectrometry assay. *Talanta*, 143: 353-358.
- Garnsworthy, P.C., Sinclair, K.D. and Webb, R. (2008). Integration of physiological mechanisms that influence fertility in dairy cows. *Animal*, 2(8): 1144-1152.
- Garrott, R.A., Monfort, S.L., White, P.J., Mashburn, K.L. and Cook, J.G. (1998). One-sample pregnancy diagnosis in elk using faecal steroid metabolites. *Journal of Wildlife Diseases*, 34(1): 126-131.
- Gascoigne, E., Swain, J., Glover, M., Nabb, L. and McLean, J. (2017). Controlled release progestagen sponges for sheep. *Veterinary Record*, 180(4): 101-102.
- Gholib, G., Agil, M., Supriatna, I., Purwantara, B., Heistermann, M. and Engelhardt, A. (2017). Repeated freeze-thaw cycles but not short-term storage of faecal extracts at ambient temperature influence the stability of steroid metabolite levels in crested macaques. *Jurnal Kedokteran Hewan*, 11(2): 78-85.
- Gholib, G., Heistermann, M., Agil, M., Supriatna, I., Purwantara, B., Nugraha, T. P. and Engelhardt, A. (2018). Comparison of faecal preservation and extraction methods for steroid hormone metabolite analysis in wild crested macaques. *Primates*, 59(3): 281-292.

- Gika, H., Kaklamanos, G., Manesiotis, P. and Theodoridis, G. (2015). Chromatography: High-Performance Liquid Chromatography. In *Encyclopedia of Food and Health* (pp. 93–99). Elsevier Inc., <https://doi.org/10.1016/B978-0-12-384947-2.00159-8>.
- Gillam, E.M., Baba, T., Kim, B.R., Ohmori, S. and Guengerich, F.P. (1993). Expression of modified human cytochrome P450 3A4 in *Escherichia coli* and purification and reconstitution of the enzyme. *Archives of Biochemistry and Biophysics*, 305(1): 123-131.
- Ginther, O.J. (2000). Selection of the dominant follicle in cattle and horses. *Animal Reproduction Science*, 60: 61-79.
- Goding, J.W. (1996). *Monoclonal antibodies: principles and practice*. Elsevier.
- Goldsmith, S.J. (1975). Radioimmunoassay: Review of basic principles. In *Seminars in Nuclear Medicine*, 5 (2):125-152. Elsevier. Graham, J.D. and Clarke, C.L. (1997). Physiological action of progesterone in target tissues. *Endocrine Reviews* 18: 502-519.
- Graham, L., Schwarzenberger, F., Möstl, E., Galama, W. and Savage, A. (2001). A versatile enzyme immunoassay for the determination of progestogens in faeces and serum. *Zoo Biology* 20(3): 227-236.
- Graham, L.H. and Brown, J.L. (1997). Non-invasive assessment of gonadal and adrenocortical function in felid species via faecal steroid analysis. *Zeitschrift fuer Saeugetierkunde (Germany)*.
- Graham, L.H. Goodrowe, K.L., Raeside, J.I. and Liptrap, R.M. (1995). Non- invasive monitoring of ovarian function in several felid species by measurement of faecal estradiol- 17 β and progestins. *Zoo Biology*, 14(3): 223-237.
- Grange, R. D., Thompson, J. P. and Lambert, D. G. (2014). Radioimmunoassay, enzyme and non-enzyme-based immunoassays. *British Journal of Anaesthesia*, 112(2): 213-216.
- Greenspan, N.S. and Cavacini, L.A. (2019). Immunoglobulin function. In *Clinical Immunology, Principle and Practice*. Elsevier Inc (Fifth Edition) (pp. 223-233).
- Guyton, A.C. and Hall, J.E. (1996). *Textbook of medical physiology* (9th edn), 1018-1021, W.B. Saunders Co.
- Hagen, J., Gott, N. and Miller, D. R. (2003). Reliability of saliva hormone tests. *Journal of the American Pharmacists Association*, 43(6): 724-726.
- Hanly, W.C., Artwohl, J.E. and Bennett, B.T. (1995). Review of polyclonal antibody production procedures in mammals and poultry. *Institute of Laboratory Animal Research Journal*, 37: 93-118.

- Hansel, W. and Convey, E. M. (1983). Physiology of the estrous cycle. *Journal of Animal Science*, 57(suppl_2): 404-424.
- Harlow E and Lane D (eds) (1988). *Antibodies: a laboratory manual*, Cold spring Harbor Laboratory, New York.
- Harris, T.R. and Monfort, S.L. (2006). Mating behavior and endocrine profiles of wild black and white Colobus monkeys (*Colobus guereza*): toward an understanding of their life history and mating system *Journal of Primatology*, 68:383–396
- Hattab, S. A., Kadoom, A. K., Palme, R. and Bamberg, E. (2000). Effect of crestar™ on estrus synchronization and the relationship between fecal and plasma concentrations of progestagens in buffalo cows. *Theriogenology*, 54(7): 1007-1017.
- Hauser, B., Deschner, T. and Boesch, C. (2008). Development of a liquid chromatography–tandem mass spectrometry method for the determination of 23 endogenous steroids in small quantities of primate urine. *Journal of Chromatography B*, 862(1-2): 100-112.
- Heistermann, M., Agil, M., Bütke, A. and Hodges, J. K. (1998). Metabolism and excretion of oestradiol-17 β and progesterone in the Sumatran rhinoceros (*Dicerorhinus sumatrensis*). *Animal Reproduction Science*, 53(1-4): 157-172.
- Heistermann, M., Möhle, U., Vervaecke, H., van Elsacker, L. and Keith Hodges, J. (1996). Application of urinary and faecal steroid measurements for monitoring ovarian function and pregnancy in the bonobo (*Pan paniscus*) and evaluation of perineal swelling patterns in relation to endocrine events. *Biology of Reproduction*, 55(4): 844-853.
- Herbison, A. E. (2008). Estrogen positive feedback to gonadotropin-releasing hormone (GnRH) neurons in the rodent: the case for the rostral periventricular area of the third ventricle (RP3V). *Brain Research Reviews*, 57(2): 277-287.
- Hermanson, G.T. (2013). *Bioconjugate techniques*. Academic press.
- Heyman, Y.A., Chavatte-Palmer, P., LeBourhis, D., Camous, S., Vignon, X. and Renard, J. (2002). Frequency and occurrence of late-gestation losses from cattle cloned embryos. *Biology of Reproduction*, 66: 6-13.
- Hidayatik, N., Agil, M., Heistermann, M., Iskandar, E., Yusuf, T. L. and Sajuthi, D. (2018). Assessing female reproductive status of spectral tarsier (*Tarsius tarsier*) using faecal steroid hormone metabolite analysis. *American Journal of Primatology*, 80 (11): e22917.
- Higham, J. P. (2016). Field endocrinology of nonhuman primates: past, present, and future. *Hormones and Behavior*, 84: 145-155.

- Hindle, J. E. and Hodges, J. K. (1990). Metabolism of oestradiol-17 β and progesterone in the white rhinoceros (*Ceratotherium simum simum*). *Journal of Reproduction and Fertility*, 90(2): 571-580.
- Hirata, S. and Mori, Y. (1995). Monitoring reproductive status by faecal progesterone analysis in ruminants. *Journal of Veterinary Medical Science*, 57(5): 845-850.
- Hirschenhauser, K., Kotrschal, K. and Möstl, E. (2005). Synthesis of measuring steroid metabolites in goose feces. *Annals of the New York Academy of Sciences*, 1046(1): 138-153.
- Hodges, J.K., Czekala, N.M. and Lasley, B.L. (1979). Estrogen and luteinizing hormone secretion in diverse primate species from simplified urinary analysis. *Journal of Medical Primatology*, 8:349-364.
- Hogan, L.A., Lisle, A.T., Johnston, S.D., Goad, T. and Robertston, H. (2013). Adult and juvenile sex identification in threatened monomorphic geocrinia frogs using faecal steroid analysis. *Journal of Herpetology*, 47(1): 112-118.
- Hosseini, S., Vázquez-Villegas, P., Rito-Palomares, M. and Martinez-Chapa, S. O. (2018). *Enzyme-linked Immunosorbent Assay (ELISA): From A to Z*. Springer Singapore.
- Howell, M. and Shepherd, M. (2018). *Anaesthesia and Intensive Care Medicine*, 19(10):575-578.
- Hultén, F., Zhang, B. R., Forsberg, M. and Dalin, A. M. (1995). Applying a progesterone assay to faecal samples collected from sows during the oestrous cycle. *Reproduction in Domestic Animals*, 30 (3): 101-105.
- Hunt, K. E. and Wasser, S. K. (2003). Effect of long-term preservation methods on faecal glucocorticoid concentrations of grizzly bear and African elephant. *Physiological and Biochemical Zoology*, 76(6): 918-928.
- Illera, Juan- Carlos, Gema S., Sara C., Maria- Dolores C., Cati G., Leticia Martínez-Fernández, Coralie M. and Miguel C. (2014). "Assessment of ovarian cycles in the African elephant (*Loxodonta africana*) by measurement of salivary progesterone metabolites." *Zoo Biology*, 33(3): 245-249.
- Isobe, N., Akita, M., Nakao, T., Yamashiro, H. and Kubota, H. (2005a). Pregnancy diagnosis based on the faecal progesterone concentration in beef and dairy heifers and beef cows. *Animal Reproduction Science*, 90: 211-218.
- Isobe, N., Nakao, T., Yamashiro, H. and Shimada, M. (2005b). Enzyme immunoassay of progesterone in the faeces from beef cattle to monitor the ovarian cycle. *Animal Reproduction Science*, 87(1-2): 1-10.

- Jamshidi, A.A., Girard, D., Beaudry, F. and Goff, A.K. (2007). Progesterone metabolism in bovine endometrial cells and the effect of metabolites on the responsiveness of the cells to OT-stimulation of PGF₂ α . *Steroids*, 72(13): 843-850.
- Juengel, J.L. and Niswender, G.D. (1999). Molecular regulation of luteal progesterone synthesis in domestic ruminants. *Journal of Reproduction and Fertility*. Supplement, 54, 193-205.
- Kakar, S.S., Rahe, C.H. and Neill, J.D. (1993). Molecular cloning, sequencing, and characterizing the bovine receptor for gonadotropin releasing hormone (GnRH). *Domestic animal endocrinology*, 10(4): 335-342.
- Keevil, B. G. (2013). Novel liquid chromatography tandem mass spectrometry (LC-MS/MS) methods for measuring steroids. *Best practice and research Clinical Endocrinology and Metabolism*, 27(5): 663-674.
- Kesler, D.J. and Garverick, H.A. (1982). Ovarian cysts in dairy cattle: a review. *Journal of Animal Science*, 55(5), 1147-1159.
- Kessler, M.J. (1982). High performance liquid chromatography of steroid metabolites in the pregnenolone and progesterone pathways. *Steroids* 39(1): 21-32.
- Khan, M., Altmann, J., Isani, S. and Yu, J. (2002). A matter of time: evaluating the storage of faecal samples for steroid analysis. *General and Comparative Endocrinology* 128, 57-64.
- Kieslich, K. (1985). Microbial side- chain degradation of sterols. *Journal of Basic Microbiology*, 25(7): 461-474.
- Kim, Y.-S., Zhang, H., Kim, H.-Y. (2000). Profiling neurosteroids in cerebrospinal fluids and plasma by gas chromatography/electron capture negative chemical ionization mass spectrometry. *Analytical Biochemistry*, 277: 187-195.
- Kimmins, S. and MacLaren, L.A. (2001). Oestrous cycle and pregnancy effects on the distribution of oestrogen and progesterone receptors in bovine endometrium. *Placenta*, 22(8): 742-748.
- Krohn, R. I. (2005). The colorimetric detection and quantitation of total protein. *Current protocols in toxicology*, 23(1): A-3I.
- Kleemann, D., Walker, S. and Seamark, R. (1994). Enhanced foetal growth in sheep administered progesterone during the first three days of pregnancy. *Journal of Reproduction and Fertility*, 102: 411-417.
- Koal, T., Schmiederer, D., Pham-Tuan, H., Röhring, C. and Rauh, M. (2012). Standardized LC-MS/MS based steroid hormone profile-analysis. *The Journal of Steroid Biochemistry and Molecular Biology*, 129(3-5): 129-138.

- Köcher, T., Pichler, P., Swart, R. and Mechtler, K. (2011). Quality control in LC-MS/MS. *Proteomics*, 11(6): 1026-1030.
- Koenderman, L., Buurman, W. and Daha, M.R. (2014). The innate immune response. *Immunology Letters*, 162(2): 95-102.
- Koren, L., Mokady, O., Karaskov, T., Klein, J., Koren, G. and Geffen, E. (2002). A novel method using hair for determining hormonal levels in wildlife. *Animal Behaviour*, 63: 403-406.
- Kornmatitsuk, B., Thitaram, C. and Kornmatitsuk, S. (2007). Measurement of Faecal Progesterone Metabolites and its Application for Early Screening of Open Cows Post- insemination. *Reproduction in Domestic Animals*, 42:238-242.
- Kozłowski, C.P., Clawitter, H.L., Thier, T., Fischer, M.T. and Asa, C.S. (2018). Characterization of oestrous cycles and pregnancy in Somali wild asses (*Equus africanus somaliensis*) through faecal hormone analyses. *Zoo Biology*, 37(1): 35-39.
- Kuby, J. (1997). Immunology. 3rd edition. 3rd Ed. WH Freeman.
- Kumar, A., Mehrotra, S., Dangi, S., Singh, G., Singh, L., Mahla, A., Kumar, S. and Nehra, K., (2013). Faecal steroid metabolites assay as a non-invasive monitoring of reproductive status in animals. *Veterinary World*, 6(1): 59-63
- Kupiec, T. (2004). Quality-control analytical methods: High-performance liquid chromatography. *International Journal of Pharmaceutical Compounding*, 8: 223-227.
- Kushnir, M. M., Rockwood, A. L., Roberts, W. L., Yue, B., Bergquist, J. and Meikle, A. W. (2011). Liquid chromatography tandem mass spectrometry for analysis of steroids in clinical laboratories. *Clinical Biochemistry*, 44(1): 77-88.
- Kusina, N.T., Tarwirei, F., Hamudikuwanda, H., Agumba, G. and Mukwena, J. (2000). A comparison of the effects of progesterone sponges and ear implants, PGF₂alpha, and their combination on efficacy of oestrus synchronization and fertility of Mashona goat does. *Theriogenology*, 53(8): 1567-1580.
- Lamming, G.E. and Darwash, A.O. (1998). The use of milk progesterone profiles to characterise components of subfertility in milked dairy cows. *Animal Reproduction Science*, 52(3): 175-190.
- Larson, S., Casson, C. and Wasser, S. (2003). Non-invasive reproductive steroid hormone estimates from faecal samples of captive female sea otters (*Enhydra lutris*). *General and Comparative Endocrinology* 134:18-25.
- Lasley, B.L. and Kirkpatrick, J.F. (1991). Monitoring ovarian function in captive and free-ranging wildlife by means of urinary and faecal steroids. *Journal of Zoo and Wildlife Medicine*, 22(1) :23-31.

- Leenaars, M. and Hendriksen, C.F. (2005). Critical steps in the production of polyclonal and monoclonal antibodies: evaluation and recommendations. *Institute of Laboratory Animal Research Journal*, 46(3), 269-279.
- Leenaars, P.P.A.M., Hendriksen, C.F., De Leeuw, W.A., Carat, F., Delahaut, P., Fischer, R. and Lindblad, E.B. (1999). The production of polyclonal antibodies in laboratory animals. *Atla-Nottingham*, 27: 79-102.
- Lemley, C.O., Butler, S.T., Butler, W.R. and Wilson, M.E. (2008). Insulin Alters Hepatic Progesterone Catabolic Enzymes Cytochrome P450 2C and 3A in Dairy Cows1. *Journal of Dairy Science*, 91(2): 641-645.
- Lemoine, A., Gautier, J.C., Azoulay, D., Kiffel, L., Belloc, C., Guengerich, F.P. and Leroux, J.P. (1993). Major pathway of imipramine metabolism is catalyzed by cytochromes P-450 1A2 and P-450 3A4 in human liver. *Molecular Pharmacology*, 43(5): 827-832.
- Lemus, R. and Karol, M.H. (2008). Conjugation of Haptens in *Methods in Molecular Medicine*. 138:167-182.
- Levy, O., Goriely, S. and Kollmann, T.R. (2013). Immune response to vaccine adjuvants during the first year of life. *Vaccine*, 31(21): 2500-2505.
- Li, Y., Han, C., Zhang, J. and Luo, X. (2000). Establishment of monoclonal antibodies against aflatoxin M1. *Wei sheng yan jiu= Journal of Hygiene Research*, 29(1): 59-60.
- Lipman, N.S., Jackson, L.R., Trudel, L.J. and Weis-Garcia, F. (2005). Monoclonal versus polyclonal antibodies: distinguishing characteristics, applications, and information resources. *Institute of Laboratory Animal Research Journal*, 46 (3): 258-268.
- Llewelyn, M.B., Hawkins, R.E. and Russell, S.J. (1992). Discovery of antibodies. *The British Medical Journal*, 305(6864), 1269-1272.
- Lonergan, P., O'Hara, L. and Forde, N. (2013). Role of dioestrus progesterone on endometrial function and conceptus development in cattle. *Animal Reproduction*, 10 (3): 223-227.
- López-García, M., Romero-González, R. and Frenich, A.G. (2018). Determination of steroid hormones and their metabolite in several types of meat samples by ultra-high performance liquid chromatography—Orbitrap high resolution mass spectrometry. *Journal of Chromatography A*, 1540: 21-30.
- Lozano-Sánchez, J., Borrás-Linares, I., Sass-Kiss, A. and Segura-Carretero, A. (2018). Chromatographic Technique: High-Performance Liquid Chromatography (HPLC). In *Modern Techniques for Food Authentication* (pp. 459-526). Academic Press.

- Lucy, M.C. (2001). Reproductive loss in high-producing dairy cattle: where will it end?. *Journal of Dairy Science*, 84(6): 1277-1293.
- Lynch, J., Khan, M., Altmann, J., Njahira, M. and Rubenstein, N. (2003). Concentrations of four faecal steroids in wild baboons: short-term storage conditions and consequences for data interpretation. *General and Comparative Endocrinology*, 132: 264-271.
- Lynch, J.W., Ziegler, T.E. and Strier, K.B. (2002). Individual and seasonal variation in faecal testosterone and cortisol levels of wild male tufted capuchin monkeys, *Cebus apella nigrinus*. *Hormones and Behavior*, 41: 275-287.
- Macchi, E., Cucuzza, A. S., Badino, P., Odore, R., Re, F., Bevilacqua, L. and Malfatti, A. (2010). Seasonality of reproduction in wild boar (*Sus scrofa*) assessed by faecal and plasmatic steroids. *Theriogenology*, 73(9): 1230-1237.
- Machado, R., Bergamaschi, M.A.C.M., Barbosa, R.T., De Oliveira, C. A. and Binelli, M. (2008). Ovarian function in Nelore (*Bos taurus indicus*) cows after post-ovulation hormonal treatments. *Theriogenology*, 69(7): 798-804.
- Macmillan, K. and Peterson, A. (1993). A new intravaginal progesterone releasing device for cattle (CIDR-B) for oestrous synchronisation, increasing pregnancy rates and the treatment of post-partum anoestrus. *Animal Reproduction Science* 33: 1-25.
- Maggio, E.T. (1980). *Enzyme Immunoassay*. 1st Ed. Crc Press. Boca Ranton FL.
- Magness, R. R., Reynolds, L. P., and Ford, S. P. (1986). Evidence for uterine metabolism of progesterone during early pregnancy in the pig. *Theriogenology*, 25(4):551-558.
- Mann, G.E., Keatinge, R., Hunter, M., Hedley, B.A. and Lamming, G.E. (2005). The use of milk progesterone to monitor reproductive function in beef suckler cows. *Animal Reproduction Science*, 88(3-4): 169-177.
- Masunda, B., Mutisi, C., Hamudikuwanda, H. and Agumbah, J. (1999). The concentration of faecal progestins during the oestrous cycle in Nkone cows and the effect of duration of storage of faecal samples at room temperature on faecal progestin levels. *Tropical Animal Health and Production* 31: 373-381.
- Masunda, B., Mutisi, C., Hamudikuwanda, H. and Agumbah, J. (2002). The Use of Faecal Progestin Measurements to Monitor Reproductive Activity in Mashona Cows in a Smallholder Farming Area of Zimbabwe. *Tropical Animal Health and Production*, 34: 309-318.
- Matissek, R., Wittkowski, R. and Baltes, W. (1992). *High performance liquid chromatography in food control and research*. Behr's Verlag.

- Matysik, S. and Liebisch, G. (2017). Quantification of steroid hormones in human serum by liquid chromatography-high resolution tandem mass spectrometry. *Journal of Chromatography A*, 1526: 112-118.
- McCalley, D.V. (2010). The challenges of the analysis of basic compounds by high performance liquid chromatography: some possible approaches for improved separations. *Journal of Chromatography A*, 1217(6): 858-880.
- McCracken, V.L., Xie, G., Deaver, S.E., Baumgard, L.H., Rhoads, R.P. and Rhoads, M.L. (2015). Hepatic progesterone-metabolizing enzymes cytochrome P450 2C and 3A in lactating cows during thermoneutral and heat stress conditions. *Journal of Dairy Science*, 98 (5): 3152-3157.
- McCrone, I. (2009). Infertility in dairy cows: Ovarian dysfunction and hormonal imbalances. *UK Veterinary Livestock*, 14 (6): 19-25.
- Meisterling, E.M. and Dailey, R.A. (1987). Use of Concentrations of Progesterone and Estradiol-17 β in Milk in Monitoring Postpartum Ovarian Function in Dairy Cows¹. *Journal of Dairy Science*, 70(10): 2154-2161.
- Messmann, S., Bagu, E., Robia, C. and Palme, R. (1999). Measurement of glucocorticoid metabolite concentrations in faeces of domestic livestock. *Journal of Veterinary Medicine Series A* 46: 621-631.
- Mihm, M. and Evans, A.C.O. (2008). Mechanisms for dominant follicle selection in mono-ovulatory species: a comparison of morphological, endocrine and intraovarian events in cows, mares and women. *Reproduction in Domestic Animals*, 43: 48-56.
- Miller, H.M., Foxcroft, G.R., Squires, J. and Aherne, F.X. (1999). The effects of feed intake and body fatness on progesterone metabolism in ovariectomized gilts. *Journal of animal science*, 77(12): 3253-3261.
- Millspough, J.J. and Washburn, B.E. (2003). Within-sample variation of faecal glucocorticoid measurements. *General and Comparative Endocrinology*, 132(1): 21-26.
- Millspough, J.J. and Washburn, B.E. (2004). Use of faecal glucocorticoid metabolite measures in conservation biology research: considerations for application and interpretation. *General and comparative endocrinology*, 138(3): 189-199.
- Millspough, J.J., Washburn, B.E., Milanick, M.A., Beringer, J., Hansen, L.P. and Meyer, T.M. (2002). Non-invasive techniques for stress assessment in white-tailed deer. *Wildlife Society Bulletin*, 899-907.
- Mithileshwari, C., Srivastava, T., Kumar, V., Kumar, A. and Umapathy, G. (2016). Non-invasive assessment of faecal progestagens and pregnancy detection in Himalayan musk deer (*Moschus chrysogaster*). *Theriogenology*, 85(2): 216-223.

- Mizuta, K. (2003). Estudo comparativo dos aspectos comportamentais do estro e dos teores plasmáticos de LH, FSH, progesterona e estradiol que precedem a ovulação em fêmeas Nelore (*Bos taurus indicus*), Angus (*Bos taurus taurus*) e Nelore x Angus (*Bos taurus indicus* x *Bos taurus taurus*) (Doctoral dissertation). Faculdade de Medicina Veterinária Universidade e Aao Paulo, Sao Paulo, Brazil.
- Moal, V., Mathieu, E., Reynier, P., Malthièry, Y. and Gallois, Y. (2007). Low serum testosterone assayed by liquid chromatography-tandem mass spectrometry. Comparison with five immunoassay techniques. *Clinica chimica acta*, 386(1-2): 12-19.
- Moenter, S.M., Brand, R.C. and Karsch, F.J. (1992). Dynamics of gonadotropin-releasing hormone (GnRH) secretion during the GnRH surge: insights into the mechanism of GnRH surge induction. *Endocrinology*, 130 (5): 2978-2984.
- Mohammed, O.B., Green, D.I. and Holt, W.V. (2011). Faecal progesterone metabolites and ovarian activity in cycling and pregnant mountain gazelles (*Gazella gazella*). *Theriogenology*, 75(3): 542-548.
- Mondal, S. and Prakash, B. (2003). Peripheral plasma progesterone concentration in relation to oestrus expression in Sahiwal cows. *Indian Journal of Physiology and Pharmacology*, 47: 111-114.
- Monfort, S.L., Schwartz, C.C. and Wasser, S. K. (1993). Monitoring reproduction in captive moose using urinary and faecal steroid metabolites. *The Journal of Wildlife Management*, 400-407.
- Monfort, S.L., Wasser, S.K., Mashburn, K.L., Burke, M., Brewer, B.A. and Creel, S.R. (1997). Steroid metabolism and validation of non-invasive endocrine monitoring in the African wild dog (*Lycaon pictus*). *Zoo Biology: Published in affiliation with the American Zoo and Aquarium Association*, 16(6):533-548.
- Morrow, C.J. and Monfort, S.L. (1998). Ovarian activity in the scimitar-horned oryx (*Oryx dammah*) determined by faecal steroid analysis. *Animal Reproduction Science*, 53(1-4): 191-207.
- Möstl, E. and R. Palme. (2002). "Hormones as indicators of stress." *Domestic Animal Endocrinology*, 23(1-2): 67-74.
- Möstl, E., Rettenbacher, S., and Palme, R. (2005). Measurement of corticosterone metabolites in birds' droppings: an analytical approach. *Annals of the New York Academy of Sciences*, 1046 (1): 17-34.
- Munro, C.J., Stabenfeldt, G., Cragun, J., Addiego, L., Overstreet, J. and Lasley, B., (1991). Relationship of serum estradiol and progesterone concentrations to the excretion profiles of their major urinary metabolites as measured by enzyme immunoassay and radioimmunoassay. *Clinical Chemistry*, 37: 838-844.
- Mwaanga, E.S. and Janowski, T. (2000). Anoestrus in dairy cows: causes, prevalence and clinical forms. *Reproduction in Domestic Animals*, 35(5): 193-200.

- Navarro, H., Zarco, L., Ducoing, A., Flores, G., and Valencia, J. (1990). Effect of time and temperature of incubation of heparinized caprine blood on concentrations of progesterone detected in plasma. *Theriogenology*, 33(4): 749-755.
- Ncube, H., Duncan, P., Grange, S., Cameron, E. Z., Barnier, F. and Ganswindt, A. (2011). Pattern of faecal 20-oxopregnane and oestrogen concentrations during pregnancy in wild plains zebra mares. *General and Comparative Endocrinology*, 172(3): 358-362.
- Netea, M. G., Schlitzer, A., Placek, K., Joosten, L. A. and Schultze, J. L. (2019). Innate and Adaptive Immune Memory: An Evolutionary Continuum in the Host's Response to Pathogens. *Cell, Host and Microbe*, 25(1): 13-26.
- Newman, M., Pratt, S. M., Curran, D. A. and Stanczyk, F. Z. (2019). Evaluating urinary estrogen and progesterone metabolites using dried filter paper samples and gas chromatography with tandem mass spectrometry (GC-MS/MS). *BMC Chemistry*, 13(1): 20.
- Niswender, G.D. (1981). Mechanisms controlling luteolysis. In *Dynamics of Ovarian Function* (pp. 153-160). Raven Press, New York.
- Niswender, G.D., Juengel, J.L., Silva, P.J., Rollyson, M.K. and McIntush, E.W. (2000). Mechanisms controlling the function and life span of the corpus luteum. *Physiological Reviews*, 80(1): 1-29.
- Norris, R.L., Eaglesham, G.K., Shaw, G.R., Smith, M.J., Chiswell, R.K., Seawright, A.A. and Moore, M.R. (2001). A sensitive and specific assay for glutathione with potential application to glutathione disulphide, using high-performance liquid chromatography-tandem mass spectrometry. *Journal of Chromatography B: Biomedical Sciences and Applications*, 762(1): 17-23.
- Nöthling, J.O. and De Cramer, K.G. (2018). Comparing the values of progesterone in the blood of bitches as measured with a chemiluminescence immunoassay and a radioimmunoassay. *Reproduction in domestic animals*, 53(5): 1136-1141.
- Nugraha, T.P., Heistermann, M., Agil, M., Purwantara, B., Supriatna, I., Gholib, G., van Schaik, C.P. and Weingrill, T., (2017). Validation of a field-friendly extraction and storage method to monitor faecal steroid metabolites in wild orangutans. *Primates*, 58: 285-294.
- Olsen, J.H., Chen, C.L., Boules, M.M., Morris, L.S. and Coville, B.R. (1994). Determination of reproductive cyclicity and pregnancy in Asian elephants (*Elephas maximus*) by rapid radioimmunoassay of serum progesterone. *Journal of Zoo and Wildlife Medicine*, 349-354.
- Oltner, R. and Edqvist, L.E. (1982). Changes in plasma progesterone levels during storage of heparinized whole blood from cow, horse, dog and pig. *Acta Veterinaria Scandinavica*, 23(1): 1-8.

- Orsonneau, J.L., Massoubre, C., Cabanes, M. and Lustenberger, P. (1992). Simple and sensitive determination of urea in serum and urine. *Clinical chemistry*, 38(5), 619-623.
- Otavă, G., Cernescu, H., Mircu, C. and IGNA, V. (2007). Pregnancy diagnosis in cow using progesterone measurements. *Lucrări științifice medicină veterinară*, 95-8.
- Painter, R.H. (1998). IgG. *Encyclopedia of Immunology*. 2nd Ed. Academic Press. Michigan USA.
- Palme, R. (2005). Measuring faecal steroids: guidelines for practical application. *Annals of the New York Academy of Sciences*, 1046: 75-80.
- Palme, R. and Möstl, E. (1997). Measurement of cortisol metabolites in faeces of sheep as a parameter of cortisol concentration in blood. *Zeitschrift fuer Saeugetierkunde (Germany)*.
- Palme, R., Entenfellner, U., Hoi, H. and Möstl, E. (2001). Faecal oestrogens and progesterone metabolites in mares of different breeds during the last trimester of pregnancy. *Reproduction in Domestic Animals*, 36 (5): 273-277.
- Palme, R., Fischer, P., Schildorfer, H. and Ismail, M. (1996). Excretion of infused 14 C-steroid hormones via faeces and urine in domestic livestock. *Animal Reproduction Science* 43: 43-63.
- Palme, R., Rettenbacher, S., Touma, C., El- Bahr, S. M. and Möstl, E. (2005). Stress hormones in mammals and birds: comparative aspects regarding metabolism, excretion, and non-invasive measurement in faecal samples. *Annals of the New York Academy of Sciences*, 1040(1): 162-171.
- Palme, R., Touma, C., Arias, N., Dominchin, M. and Lepschy, M. (2013). Steroid extraction: get the best out of faecal samples. *Wien Tierarztl Monatsschr* 100, 238-246.
- Panda, S., Patra, B. K., Sahu, S. K., Sahoo, N., Mohanty, D. N. and Nahak, A.K. (2017). Faecal Estrogen and Progesterone Concentration in Captive Royal Bengal Tigresses. *International Journal of Livestock Research*, 7(12): 65-73.
- Pappano, D.J., Roberts, E.K. and Beehner, J.C. (2010). Testing extraction and storage parameters for a faecal hormone method. *American Journal of Primatology*, 72(11): 934-941.
- Paris, M.C.J., White, A., Reiss, A., West, M. and Schwarzenberger, F. (2002). Faecal progesterone metabolites and behavioural observations for the non-invasive assessment of oestrous cycles in the common wombat (*Vombatus ursinus*) and the southern hairy-nosed wombat (*Lasiorhinus latifrons*). *Animal Reproduction Science*, 72(3-4): 245-257.

- Parr, R.A., Davis, I.F., Miles, M.A. and Squires, T.J. (1993). Liver blood flow and metabolic clearance rate of progesterone in sheep. *Research in Veterinary Science*, 55(3): 311-316.
- Pastoret, P.P. (1999). Veterinary vaccinology. *Comptes Rendus de l'Académie des Sciences-Series III-Sciences de la Vie* 322, 967-972.
- Patton, M.L., Swaisgood, R.R., Czekala, N.M., White, A.M., Fetter, G.A., Montagne, J.P. and Lance, V. A. (1999). Reproductive cycle length and pregnancy in the southern white rhinoceros (*Ceratotherium simum simum*) as determined by faecal pregnane analysis and observations of mating behavior. *Zoo Biology: Published in affiliation with the American Zoo and Aquarium Association*, 18(2): 111-127.
- Paz, R. C. R., Souza, N. P., Furtado, P. V. and Brown, J. L. (2016). Estradiol and progesterone fecal metabolites analysis in crab-eating-fox (*Cerdocyon thous*). *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 68(3), 636-640.
- Peng, X., Cao, B., Deng, G.Z., Li, C.Y., Ye, L.L. and Yu, H. (2011). Echography characteristics of abnormal ovaries in infertile dairy cows. *Journal of Animal and Veterinary Advances*, 10:1166-1170.
- Penning, T.M., Lee, S.H., Jin, Y., Gutierrez, A. and Blair, I.A. (2010). Liquid chromatography–mass spectrometry (LC–MS) of steroid hormone metabolites and its applications. *The Journal of Steroid Biochemistry and Molecular Biology*, 121(3-5): 546-555.
- Pereira, R.J., Christofoletti, M.D., Blank, M.H. and Duarte, J.M.B. (2018). Uro faecal steroid profiles of captive Blue-fronted parrots (*Amazona aestiva*) with different reproductive outcomes. *General and Comparative Endocrinology*, 260: 1-8.
- Pereira, R.J.G., Polegato, B.F., de Souza, S., Negrao, J.A. and Duarte, J.M.B. (2006). Monitoring ovarian cycles and pregnancy in brown brocket deer (*Mazama gouazoubira*) by measurement of faecal progesterone metabolites. *Theriogenology*, 65(2): 387-399.
- Pérez, R.C.M., Mejía, C.M. and Quintero, L.Z. (1994). Production of antibodies against progesterone from the egg yolk of hens and from rabbit blood serum to be used in radioimmunoassay. *Veterinaria México*, 25(2): 117-125.
- Perry, G.A. (2016). Factors affecting puberty in replacement beef heifers. *Theriogenology*, 86 (1): 373-378.
- Pérsico, J.M.R., Bianchi, C., Tapia, C., Raggio, S. and Marchetti, I. A. (2018). Comparative Quantification of Plasma Progesterone Through Radioimmunoassay and Enzyme-Linked Fluorescent Assay Techniques in Cattle. *Reproduction, Fertility and Development*, 30(1): 184-184.

- Pettitt, B., Wheaton, C. and Waterman, J. (2007). Effects of storage treatment on faecal steroid hormone concentrations of a rodent, the Cape ground squirrel (*Xerus inauris*). *General and Comparative Endocrinology*, 150: 1-11.
- Pier, G.B., Lyczak, J.B., Wetzler, L.M., (2004). Immunology, infection, and immunity. 1st Edition, ASM press. Michigan, USA.
- Posthuma-Trumpie, G. A., van Amerongen, A., Korf, J. and van Berkel, W. J. (2009). Perspectives for on-site monitoring of progesterone. *Trends in Biotechnology*, 27(11): 652-660.
- Proverbio, D., Perego, R., Spada, E., Bagnagatti de Giorgi, G., Belloli, A. and Pravettoni, D. (2013). Comparison of VIDAS and radioimmunoassay methods for measurement of cortisol concentration in bovine serum. *The Scientific World Journal*, 2013.
- Pulido, A., Zarco, L., Galina, C.S., Murcia, C., Flores, G. and Posadas, E. (1991). Progesterone metabolism during storage of blood samples from Gyr cattle: effects of anticoagulant, time and temperature of incubation. *Theriogenology*, 35(5): 965-975.
- Purdy, R.H., Durocher, C.K., Moore Jr, P.H. and Rao, P.N. (1980). Analysis of metabolites of progesterone in bovine liver, kidney, kidney fat, and milk by high performance liquid chromatography. *Journal of Steroid Biochemistry*, 13: 1307-1315.
- Quissell, D.O. (1993). Steroid hormone analysis in human saliva. *Annals of the New York Academy of Sciences*, 694(1): 143-145.
- Rabiee, A., Macmillan, K., Schwarzenberger, F. and Wright, P. (2002). Effects of level of feeding and progesterone dose on plasma and faecal progesterone in ovariectomised cows. *Animal Reproduction Science*, 73: 185-195.
- Rabiee, A.R., Macmillan, K.L. and Schwarzenberger, F. (2001). Excretion rate of progesterone in milk and faeces in lactating dairy cows with two levels of milk yield. *Reproduction Nutrition Development* 41: 309-319.
- Raj, M., Naidu, G., Srinivas, M., Raghunath, M. and Rao, K. (2016). Effect of serum progesterone levels at estrus on conception in graded Murrah buffaloes under field conditions. *Theriogenology Insight: An International Journal of Reproduction in all Animals*, 6(3): 127.
- Rao, C.V. (2005). *Immunology: A Textbook*. Alpha Science Int'l Ltd..
- Rathore, A.S. and Joshi, S. (2018). Process Analysis: *High Performance Liquid Chromatography*. Elsevier Inc.
- Rauh, M. (2009). Steroid measurement with LC-MS/MS in pediatric endocrinology. *Molecular and Cellular Endocrinology*, 301(1-2): 272-281.

- Rauh, M., Gröschl, M., Rascher, W. and Dörr, H. G. (2006). Automated, fast and sensitive quantification of 17 α -hydroxy-progesterone, androstenedione and testosterone by tandem mass spectrometry with on-line extraction. *Steroids*, 71(6): 450-458.
- Read, G.F. (1993). Status report on measurement of salivary estrogens and androgens. *Annals of the New York Academy of Sciences*, 694(1): 146-160.
- Renner, B., Bochschanl, R. and Gerstner, E. (1984). Preparation of 11 α -hemisuccinyl-progesterone for immunological purposes and structural determination of the hapten. *Journal of Steroid Biochemistry*, 20(6): 1247-1251.
- Reuhs, B.L., (2017). High-performance liquid chromatography, *Food Analysis*. Springer, pp. 213-226.
- Ribatti, D. (2016). The discovery of immunoglobulin E. *Immunology letters*, 171, 1-4.
- Roche, J.F. (1996). Control and regulation of folliculogenesis--a symposium in perspective. *Reviews of Reproduction*, 1(1): 19-27.
- Roelofs, J., López-Gatius, F., Hunter, R., Van Eerdenburg, F. and Hanzen, C., (2010). When is a cow in estrus? Clinical and practical aspects. *Theriogenology*, 74: 327-344.
- Rolland, R.M., Hunt, K.E., Kraus, S.D. and Wasser, S.K. (2005). Assessing reproductive status of right whales (*Eubalaena glacialis*) using faecal hormone metabolites. *General and Comparative Endocrinology*, 142: 308-317.
- Safar-Hermann, N., Ismail, M.N., Choi, H.S., Möstl, E. and Bamberg, E. (1987). Pregnancy diagnosis in zoo animals by estrogen determination in feces. *Zoo Biology*, 6(2): 189-193.
- Samad, H.A., Ahmad, N., Rehman, N.U. and Ahmad, I. (2004). Use of milk progesterone assay for monitoring oestrus and early pregnancy in nili-ravi buffaloes. *Pakistan Veterinary Journal*, 24(3): 121-124.
- Sangsrivong, S., Combs, D.K., Sartori, R., Armentano, L.E. and Wiltbank, M.C. (2002). High feed intake increases liver blood flow and metabolism of progesterone and estradiol-17 β in dairy cattle. *Journal of Dairy Science*, 85(11): 2831-2842.
- Sartori, R. and Barros, C.M. (2011). Reproductive cycles in *Bos indicus* cattle. *Animal Reproduction Science*, 124(3-4): 244-250.
- Sartori, R., Haughian, J.M., Shaver, R.D., Rosa, G.J.M. and Wiltbank, M.C. (2004). Comparison of ovarian function and circulating steroids in estrous cycles of Holstein heifers and lactating cows. *Journal of dairy science*, 87(4), 905-920.

- Schally, A.V., Arimura, A., Kastin, A.J., Matsuo, H., Baba, Y., Redding, T.W. and White, W.F. (1971). Gonadotropin-releasing hormone: one polypeptide regulates secretion of luteinizing and follicle-stimulating hormones. *Science*, 173(4001): 1036-1038.
- Schams, D., Schallenberger, E., Menzer, C., Stangl, J., Zottmeier, K., Hoffmann, B. and Karg, H. (1978). Profiles of LH, FSH and progesterone in postpartum dairy cows and their relationship to the commencement of cyclic functions. *Theriogenology*, 10(6): 453-468.
- Schellinger, A.P. and Carr, P.W. (2006). Isocratic and gradient elution chromatography: A comparison in terms of speed, retention reproducibility and quantitation. *Journal of Chromatography A*, 1109(2): 253-266.
- Schoenecker, K.A., Lyda, R.O. and Kirkpatrick J. (2004). Comparison of three Faecal Steroid Metabolites for Pregnancy Detection used with single sampling in Bighorn Sheep (*Ovis Canadensis*). *Journal of Wildlife Diseases*: 40(2):273-28
- Schumacher, M. and Robert, F. (2002). Progesterone: synthesis, metabolism, mechanisms of action, and effects in the nervous system. In *Hormones, brain and behavior* (pp. 683-745). Academic Press.
- Schwarzenberger, F., Francke, R. and Göltenboth, R., (1993). Concentrations of faecal immunoreactive progestagen metabolites during the oestrous cycle and pregnancy in the black rhinoceros (*Diceros bicornis michaeli*). *Journal of Reproduction and Fertility*, 98: 285-291.
- Schwarzenberger, F., Möstl, E., Bamberg, E. and Von Hegel, G. (1992). Monitoring of corpus luteum function by measuring progestagens in faeces of non-pregnant mares (*Equus caballus*) and Przewalski mares (*Equus przewalskii*). *Animal Reproduction Science*, 29(3-4): 263-273.
- Schwarzenberger, F., Möstl, E., Palme, R. and Bamberg, E. (1996a). Faecal steroid analysis for non-invasive monitoring of reproductive status in farm, wild and zoo animals. *Animal Reproduction Science*, 42: 515-526.
- Schwarzenberger, F., Palme, R., Bamberg, E. and Möstl, E. (1997). A review of faecal progesterone metabolite analysis for non-invasive monitoring of reproductive function in mammals. *Zeitschrift fuer Saeugetierkunde (Germany)*.
- Schwarzenberger, F., Rietschel, W., Vahala, J., Holeckova, D., Thomas, P., Maltzan, J., Baumgartner, K., Schaftenaar, W. (2000). Faecal progesterone, estrogen, and androgen metabolites for noninvasive monitoring of reproductive function in the female Indian rhinoceros, *Rhinoceros unicornis*. *General and Comparative Endocrinology*, 119(3): 300-307.
- Schwarzenberger, F., Son, C., Pretting, R. and Arbeiter, K. (1996b). Use of group-specific antibodies to detect faecal progesterone metabolites during the oestrous cycle of cows. *Theriogenology*, 46 (1): 23-32.

- Schwarzenberger, F., Speckbacher, G., and Bamberg E. (1995). Plasma and faecal progesterone evaluations during and after the breeding season of the female vicuna (*Vicuna vicuna*). *Theriogenology*, 43(3): 625-634.
- Schwarzenberger, F., Tomášová, K., Holečková, D., Matern, B. and Möstl, E. (1996c). Measurement of faecal steroids in the black rhinoceros (*Diceros bicornis*) using group- specific enzyme immunoassays for 20- oxo- pregnanes. *Zoo Biology: Published in affiliation with the American Zoo and Aquarium Association*, 15(2): 159-171.
- Schwarzenberger, F., Tomášová, K., Holečková, D., Matern, B. and Möstl, E. (1996). Measurement of faecal steroids in the black rhinoceros (*Diceros bicornis*) using group- specific enzyme immunoassays for 20- oxo- pregnanes. *Zoo Biology*: i, 15(2): 159-171.
- Sell, S. L., Scalzitti, J. M., Thomas, M. L. and Cunningham, K. A. (2000). Influence of ovarian hormones and oestrous cycle on the behavioral response to cocaine in female rats. *Journal of Pharmacology and Experimental Therapeutics*, 293(3): 879-886.
- Shackleton, C. (2010). Clinical steroid mass spectrometry: a 45-year history culminating in HPLC–MS/MS becoming an essential tool for patient diagnosis. *The Journal of Steroid Biochemistry and Molecular Biology*, 121(3-5): 481-490.
- Sheedy, C., MacKenzie, C.R. and Hall, J.C. (2007). Isolation and affinity maturation of hapten-specific antibodies. *Biotechnology Advances*, 25(4): 333-352.
- Shideler, S.E., Savage, A., Ortuno, A.M., Moorman, E.A. and Lasley, B.L. (1994). Monitoring female reproductive function by measurement of faecal estrogen and progesterone metabolites in the white- faced saki (*Pithecia pithecia*). *American Journal of Primatology*, 32(2):95-108.
- Shimada, T., Yamazaki, H., Mimura, M., Inui, Y. and Guengerich, F.P. (1994). Interindividual variations in human liver cytochrome P-450 enzymes involved in the oxidation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. *Journal of Pharmacology and Experimental Therapeutics*, 270(1): 414-423.
- Shirtcliff, E.A., Granger, D.A., Schwartz, E. and Curran, M.J. (2001). Use of salivary biomarkers in bio-behavioral research: cotton-based sample collection methods can interfere with salivary immunoassay results. *Psychoneuroendocrinology*, 26(2): 165-173.

- Shutt, K., Setchell, J.M. and Heistermann, M. (2012). Non-invasive monitoring of physiological stress in the Western lowland gorilla (*Gorilla gorilla gorilla*): validation of a faecal glucocorticoid assay and methods for practical application in the field. *General and Comparative Endocrinology* 179: 167-177.
- Siekman, L. (1979). Determination of steroid hormones by the use of isotope dilution–mass spectrometry: a definitive method in clinical chemistry. In *Hormonal Steroids: Proceedings of the Fifth International Congress on Hormonal Steroids*. pp. 117-123.
- Silvestre, T., Zanetti, E.S., Duarte, J.M., Barriento, F.G., Hirano, Z.M., Souza, J.C., and Passos, F.C. (2017). Ovarian cycle of southern brown howler monkey (*Alouatta guariba clamitans*) through faecal progesterin measurement. *Primates*, 58(1): 131-139.
- Simersky, R., Swaczynova, J., Morris, D.A., Franek, M. and Strnad, M. (2007). Development of an ELISA-based kit for the on-farm determination of progesterone in milk. *Veterinarni Medicina-Praha*, 52(1): 19.
- Singh, A., Chaudhary, S., Agarwal, A. and Verma, A.S. (2014). Antibodies: monoclonal and polyclonal. In *Animal Biotechnology* (265-287). Academic Press.
- Singh, K.V., Kaur, J., Varshney, G.C., Raje, M. and Suri, C.R. (2004). Synthesis and characterization of hapten– protein conjugates for antibody production against small molecules. *Bioconjugate Chemistry*, 15(1): 168-173.
- Sinreih, M., Zukunft, S., Sosič, I., Cesar, J., Gobec, S., Adamski, J. and Rižner, T. L. (2015). Combined Liquid Chromatography–Tandem Mass Spectrometry Analysis of Progesterone Metabolites. *PloS one*, 10(2), e0117984.
- Sist, M.D., Youngblood, M.A. and Williams, J.F. (1987). Using faecal estrone sulfate concentrations to detect pregnancies. *Veterinary Medicine (USA)*.
- Sitruk-Ware, R. (2008). Reprint of pharmacological profile of progestins. *Maturitas*, 61(1-2): 151-157.
- Skenandore, C.S., Pineda, A., Bahr, J.M., Newell-Fugate, A.E. and Cardoso, F.C. (2017). Evaluation of a commercially available radioimmunoassay and enzyme immunoassay for the analysis of progesterone and estradiol and the comparison of two extraction efficiency methods. *Domestic Animal Endocrinology*, 60: 61-66.
- Snyder, L.R., Kirkland, J.J. and Glajch, J.L. (2012). *Practical HPLC method development*. John Wiley and Sons. USA. Pp: 59-99.
- Spencer, T.E., Sandra, O. and Wolf, E. (2008). Genes involved in conceptus–endometrial interactions in ruminants: insights from reductionism and thoughts on holistic approaches. *Reproduction*, 135(2): 165-179.

- Stanley, C.J., Paris, F., Webb, A.E., Heap, R.B., Ellis, S.T., Hamon, M. and Booth, J. M. (1986). Use of a new and rapid milk progesterone assay to monitor reproductive activity in the cow. *The Veterinary Record*, 118(24): 664-667.
- Stevenson, J. S. and Britt, J. H. (1979). Relationships among Luteinizing Hormone, Estradiol, Progesterone, Glucocorticoids, Milk Yield, Body Weight and Postpartum Ovarian Activity in Holstein Cows 1. *Journal of Animal Science*, 48(3): 570-577.
- Stills, H.F., Manning, P., Ringler, D. and Newcomer, C. (2012). Polyclonal antibody production. The laboratory rabbit, guinea pig, hamster and other rodents. Oxford, UK: Elsevier Inc, 259-274.
- Stoops, M.A., Pairan, R.D. and Roth, T.L. (2004). Follicular, endocrine and behavioural dynamics of the Indian rhinoceros (*Rhinoceros unicornis*) oestrous cycle. *Reproduction*, 128(6), 843-856.
- Strier, K.B. and Ziegler, T. (1997). Behavioral and endocrine characteristics of the reproductive cycle in wild marmoset monkeys, *Brachyteles arachnoides*. *American Journal of Primatology*, 42: 299-310.
- Stura, E. A., Arevalo, J. H., Feinstein, A., Heap, R. B., Taussig, M. J. and Wilson, I. A. (1987). Analysis of an anti-progesterone antibody: variable crystal morphology of the Fab'and steroid-Fab'complexes. *Immunology*, 62(4): 511.
- Sunderland, S.J., Crowe, M.A., Boland, M.P., Roche, J.F. and Ireland, JJ. (1994). Selection, dominance and atresia of follicles during the oestrous cycle of heifers. *Journal of Reproduction and Fertility*, 101(3): 547-555.
- Takahashi, T., Hamanaka, S., Imai, K. and Hashizume, K. (2002). Faecal progesterone analysis by time-resolved fluoroimmunoassay (TR-FIA) for monitoring of luteal function in the sika doe (*Cervus nippon centralis*). *Journal of Veterinary Medical Science*, 64(7): 565-569.
- Taylor, R., Kendle, K., Reid, R. and Hung, C. (1987). Chromatography of progesterone and its major metabolites in rat plasma using microbore high-performance liquid chromatography columns with conventional injection and detection systems. *Journal of Chromatography A* 385: 383-392.
- Terio, K.A., Brown, J.L., Moreland, R. and Munson, L. (2002). Comparison of different drying and storage methods on quantifiable concentrations of faecal steroids in the cheetah. *Zoo Biology*, 21: 215-222.
- Thompson, K.V., Mashburn, K.L. and Monfort, S.L. (1998). Characterization of estrous cyclicity in the sable antelope (*Hippotragus niger*) through faecal progestagen monitoring. *General and Comparative Endocrinology*, 112(1): 129-137.

- Tian, W., Wang, L., Lei, H., Sun, Y. and Xiao, Z. (2018). Antibody production and application for immunoassay development of environmental hormones: a review. *Chemical and Biological Technologies in Agriculture*, 5(1): 5.
- Tong, J. C. and Ranganathan, S. (2013). Computer-aided vaccine design. Elsevier Inc. USA.
- Toubi, E., and Vadasz, Z. (2019). Innate immune-responses and their role in driving autoimmunity. *Autoimmunity Reviews*, 18(3) 306-311.
- Touma, C. and Palme, R. (2005). Measuring faecal glucocorticoid metabolites in mammals and birds: the importance of validation. *Annals of the New York Academy of Sciences*, 1046(1): 54-74.
- Trabert, B., Falk, R.T., Stanczyk, F.Z., McGlynn, K.A., Brinton, L.A. and Xu, X. (2015). Reproducibility of an assay to measure serum progesterone metabolites that may be related to breast cancer risk using liquid chromatography-tandem mass spectrometry. *Hormone Molecular Biology and Clinical Investigation*, 23(3): 79-84.
- Tsang, C.P. and Hackett, A. (1979). Metabolism of progesterone in the pregnant sheep near term: identification of 3 β -hydroxy-5 α -pregnan-20-one 3-sulfate as a major metabolite. *Steroids*, 33: 577-588.
- Umaphathy, G., Kumar, V., Kabra, M. and Shivaji, S. (2013). Detection of pregnancy and fertility status in big cats using an enzyme immunoassay based on 5 α -pregnan-3 α -ol-20-one. *General and Comparative Endocrinology*, 180: 33-38.
- Vahdat, F., Seguin, B. E., Whitmore, H. L. and Johnston, S. D. (1984). Role of blood cells in degradation of progesterone in bovine blood. *American Journal of Veterinary Research*, 45(2): 240-243.
- Van de Wiel, D.F.M. and Koops, W. (1986). Development and validation of an enzyme immunoassay for progesterone in bovine milk or blood plasma. *Animal Reproduction Science*, 10(3): 201-213.
- Van der Goot, A.C., Martin, G.B., Millar, R.P., Paris, M.C.J. and Ganswindt, A. (2015). Profiling patterns of faecal 20-oxopregnane concentrations during ovarian cycles in free-ranging southern white rhinoceros (*Ceratotherium simum simum*). *Animal Reproduction Science*, 161: 89-95.
- Van der Lende, T., Schasfoort, R., van der Meer, R. (1992). Monitoring reproduction using immunological techniques. *Animal Reproduction Science*, 28: 179-185.
- Vasconcelos, J.L.M., Sangsritavong, S., Tsai, S.J. and Wiltbank, M.C. (2003). Acute reduction in serum progesterone concentrations after feed intake in dairy cows. *Theriogenology*, 60(5): 795-807.

- Veerkamp, R.F., Beerda, B. and Van der Lende, T. (2003). Effects of genetic selection for milk yield on energy balance, levels of hormones, and metabolites in lactating cattle, and possible links to reduced fertility. *Livestock Production Science*, 83(2-3): 257-275.
- Velloso, A.L., Wasser, S.K., Monfort, S.L. and Dietz, J.M. (1998). Longitudinal faecal steroid excretion in maned wolves (*Chrysocyon brachyurus*). *General and Comparative Endocrinology*, 112(1): 96-107.
- Ventrella, D., Elmi, A., Barone, F., Carnevali, G., Govoni, N. and Bacci, M. (2018). Hair testosterone and cortisol concentrations in pre-and post-rut roe deer bucks: Correlations with blood levels and testicular morphometric parameters. *Animals*, 8(7): 113.
- Visser, S.A., Smulders, C.J., Gladdines, W.W., Irth, H., van der Graaf, P.H. and Danhof, M. (2000). High-performance liquid chromatography of the neuroactive steroids alphaxalone and pregnanolone in plasma using dansyl hydrazine as fluorescent label: application to a pharmacokinetic-pharmacodynamic study in rats. *Journal of Chromatography B: Biomedical Sciences and Applications*, 745: 357-363.
- Vukovic, D., Bozic, A., Relic, R., Stancic, B., Gvozdic, D. and Kucevic, D. (2016). Progesterone concentration in milk and blood serum and reproductive efficiency of cows after Ovsynch treatment. *Turkish Journal of Veterinary and Animal Sciences*, 40: 75-80.
- Walters, D.G., Foster, P.M. and Cottrell, R.C. (1981). High-performance liquid chromatography of progesterone and its metabolites. *Journal of Chromatography A*, 219: 152-155.
- Wang, Y.-H., Liu, S.-Q., Yang, S., Zhang, T.-X., Wei, Y.-T., Zhou, J.-T., Hu, D.-F. and Li L.-H., (2016). Determination of ovarian cyclicity and pregnancy using faecal progesterone in forest musk deer (*Moschus berezovskii*). *Animal Reproduction Science* 170: 1-9.
- Wasser, S.K. and Kathleen E.H. (2005). "Non-invasive measures of reproductive function and disturbance in the barred owl, great horned owl, and northern spotted owl." *Annals of the New York Academy of Sciences*, 1046 (1): 109-137.
- Wasser, S.K., Bevis, K., King, G. and Hanson, E. (1997). Non-invasive physiological measures of disturbance in the northern spotted owl. *Conservation Biology* 11: 1019-1022.
- Wasser, S.K., Kathleen E. H, Janine L. B., Kathy C., Carolyn M. C., Ursula B., Joshua J. M., Shawn, L. and Steven, L.M. (2000). "A generalized faecal glucocorticoid assay for use in a diverse array of nondomestic mammalian and avian species." *General and Comparative Endocrinology*, 120(3): 260-275.

- Wasser, S.K., Monfort, S.L. and Wildt, D.E. (1991). Rapid extraction of faecal steroids for measuring reproductive cyclicity and early pregnancy in free-ranging yellow baboons (*Papio cynocephalus cynocephalus*). *Journal of Reproduction and Fertility*, 92(2): 415-423.
- Wasser, S.K., Monfort, S.L., Southers, J. and Wildt, D.E. (1994). Excretion rates and metabolites of oestradiol and progesterone in baboon (*Papio cynocephalus cynocephalus*) faeces. *Reproduction*, 101(1), 213-220.
- Wasser, S.K., Papageorge, S., Foley, C. and Brown, J.L. (1996). Excretory fate of estradiol and progesterone in the African elephant (*Loxodonta africana*) and patterns of faecal steroid concentrations throughout the oestrous cycle. *General and Comparative Endocrinology*, 102(2): 255-262.
- Wasser, S.K., Risler, L. and Steiner, R.A. (1988). Excreted steroids in primate faeces over the menstrual cycle and pregnancy. *Biology of Reproduction*, 39(4): 862-872.
- Watanabe, T., Nakasaki, H., Kamijou, A., Mitomi, T., Matsuki, H. and Kasuga, H. (1993). The technical problems shoot for the production of anti-low molecular weight hapten antibody. *The Kitasato Archives of Experimental Medicine*, 65: 47-55.
- Weiss, D. J. and Wardrop, K. J. (Eds.). (2011). *Schalm's Veterinary Hematology*. John Wiley & Sons.
- Whitten, P., Brockman, D. and Stavisky, R. (1998). Recent advances in noninvasive techniques to monitor hormone- behaviour interactions. *American Journal of Physical Anthropology*, 107: 1-23.
- Wiseman, B.S., Vincent, D.L., Thomford, P.J., Scheffrahn, N.S., Sargent, G.F. and Kesler, D.J. (1983). Changes in porcine, ovine, bovine and equine blood progesterone concentrations between collection and centrifugation. *Animal Reproduction Science*, 5(3): 157-165.
- Wojtusik, J., Brown, J.L. and Pukazhenti, B.S. (2017). Non-invasive hormonal characterization of the ovarian cycle, pregnancy, and seasonal anoestrus of the female addra gazelle (*Nanger dama ruficollis*). *Theriogenology*, 95: 96-104.
- Xu, Z.Z., Burton, L.J. and Macmillan, K.L. (1997). Reproductive performance of lactating dairy cows following oestrus synchronization regimens with PGF₂ α and progesterone. *Theriogenology*, 47(3): 687-701.
- Yan, L.Y., Robinson, R.S., Shi, Z.D. and Mann, G.E. (2018). Milk progesterone on day 5 following insemination in the dairy cow: associated metabolic variables and reproductive consequences. *South African Journal of Animal Science*, 48(2): 361-368.
- Yellin, T.O. (1972). Estradiol 17 β -hemisuccinate: an improved procedure. *Journal of Lipid Research*, 13(4): 554-555.

- Yimer, N., Rosnina, Y., Wahid, H., Bukar, M.M., Malik, A., Yap, K., Fahmi, M., Ganesamurthi, P., Saharee, A., (2012). Faecal progestin extraction and analysis for non-invasive monitoring of ovarian cycle in beef cows. *Pakistan Veterinary Journal*, 32: 584-588.
- Yimer, N., Rosnina, Y., Wahid, H., Saharee, A.A., Yap, K.C., Ganesamurthi, P., Fahmi, M. and Bukar M.M. (2010). Correlation between plasma progesterone concentrations and faecal Progestins during the oestrus cycle of Kedah Kelantan cows. In *International Seminar on Tropical Animal Production (ISTAP)* :595-598.
- Younglai, E.V., Collins, D.C. and Graham, C.E. (1977). Progesterone metabolism In the Female gorilla. *Journal of Endocrinology*, 75(3): 439-440.
- YÜCEL, F. Ş. and ÇIRAKOĞLU, B. (2000). Production of Monoclonal Antibodies Specific for Progesterone, Estradiol by Simultaneous Injection of Different Steroids. *Turkish Journal of Biology*, 24(4): 697-706.
- Zhang, K. and Fent, K. (2018). Determination of two progestin metabolites (17 α -hydroxypregnanolone and pregnanediol) and different classes of steroids (androgens, estrogens, corticosteroids, progestins) in rivers and wastewaters by high-performance liquid chromatography-tandem mass spectrometry (HPLC-MS/MS). *Science of the Total Environment*, 610:1164-1172.
- Ziegler, T. and Wittwer, D., (2005). Faecal steroid research in the field and laboratory: improved methods for storage, transport, processing, and analysis. *American Journal of Primatology*, 67: 159-174.
- Ziegler, T., Hodges, K., Winkler, P. and Heistermann, M. (2000). Hormonal correlates of reproductive seasonality in wild female Hanuman langurs (*Presbytis entellus*). *American Journal of Primatology*, 51: 119-134.

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LIST OF PUBLICATIONS

Journal Articles

Published:

Innocent Damudu Peter, Abd Wahid Haron, Faez Firdaus Abdullah Jesse, Mokrish Ajat, Mark Hiew Wen Han, Wan Nor Fitri, Muhammad Sanusi Yahaya and Mohammed Saad M. Alamaary. Opportunities and challenges associated with faecal progesterone metabolite analysis. *Veterinary World*, 11(10): 1466-1472. doi: 10.14202/vetworld.2018.1466-1472.

Submitted:

Innocent Damudu Peter, Abd Wahid Haron, Faez Firdaus Abdullah Jesse, Mark Wen Han Hiew and Mokrish Ajat. Concentration of immunoreactive progesterone metabolite in feces of pregnant and non-pregnant cows. Submitted at Animal Reproduction Science.

Innocent Damudu Peter, Abd Wahid Haron, Faez Firdaus Abdullah Jesse, Mark Hiew Wen Han and Mokrish Ajat. Concentrations of immunoreactive progesterone metabolite in feces of pregnant and non-pregnant cows. Submitted at Pakistan Veterinary Journal.

Conference proceedings

Innocent Damudu Peter, Abd Wahid Haron, Faez Firdaus Jesse Abdullah, Mark Hiew Wen Han, Mokrish Moh'd Ajat. Comparison of faecal preservation and extraction methods employed in faecal progestagen analysis in cows. *Proceedings of the 30th Veterinary Association Malaysia Congress, 2018*. pp: 42



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