



UNIVERSITI PUTRA MALAYSIA

**GRANULAR FORMULATION OF ENDOPHYTIC FUNGUS,
Hendersonia toruloidea GanoEF1 FOR CONTROLLING *Ganoderma*
DISEASE AND PROMOTING OIL PALM GROWTH**

NUR RASHYEDA RAMLI

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By

NUR RASHYEDA RAMLI

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

GRANULAR FORMULATION OF ENDOPHYTIC FUNGUS, *Hendersonia toruloidea* GanoEF1 FOR CONTROLLING *Ganoderma* DISEASE AND PROMOTING OIL PALM GROWTH

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November 2018

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Oil palm is one of the important crops in Malaysia and plays an important role in the agricultural and economic development of the country. Basal stem rot (BSR) caused by *Ganoderma boninense* is the biggest threat for oil palm production and has been documented to cause a huge damage to the oil palm industry in Malaysia. There is yet an effective control measure for BSR disease. Endophytic fungi have been previously studied and identified as potential biological control agents of many crop diseases. Isolate *Hendersonia toruloidea* GanoEF1 is a novel and promising biological control agents against *G. boninense*. The use of endophytes in form of formulated product is preferred and sought as such preparations offer many advantages during application in the field. Therefore this study was designed with the specific objectives to (i) determine the compatible carriers for viability and quality of *H. toruloidea* GanoEF1 in the preparation of granular formulation, (ii) investigate the effects of granular formulations of *H. toruloidea* GanoEF1 for controlling BSR in oil palm, (iii) determine the biochemical compounds released in oil palm treated with *H. toruloidea* GanoEF1, and (iv) study the effects of the granular formulations developed of on oil palm growth. *H. toruloidea* GanoEF1 isolated from healthy oil palm roots was cultured on potato dextrose agar (PDA) media. A suspension containing 10^8 CFU ml⁻¹ of the conidia cells was prepared as granular formulation by using empty fruit bunch (EFB), rice bran (RB), talc powder, paddy husk (PH) and sawdust (SD) as nutrient supplement mixed with inert ingredient of either kaolin or palm kernel cake powder in the solution containing alginate-pectin as a binder. The best ratio of alginate:pectin for optimum growth of *H. toruloidea* GanoEF1 was 1:3 at temperature 35 °C with the highest number of conidia cell recorded was log 10⁸ CFU g⁻¹ at 30 days after storage. Three best nutrient carrier of EFB, RB and SD that succeeded to sustain the viability of *H. toruloidea* GanoEF1 were further formulated with kaolin (K) or palm kernel cake (PKC) and tested for their viability over 12 months storage and efficacy against *G. boninense*. Amongst them, three

granular formulation of empty fruit bunch-kaolin (EFB-K), empty fruit bunch-palm kernel cake (EFB-PKC) and rice bran-palm kernel cake (RB-PKC) showed highest conidia viability of *H. toruloidea* GanoEF1 more than 10^5 CFU g⁻¹ and recorded more than 50% of percentage inhibition of radial growth (PIRG) values. The effectiveness of *H. toruloidea* GanoEF1 in EFB-PKC, EFB-K and RB-PKC was then evaluated in the glasshouse on their efficacy against *G. boninense*. Disease suppression was highest in the treatment that had the application of *H. toruloidea* GanoEF1 in EFB-PKC with a disease reduction of 65.92% ($P < 0.05$). The percentage of dead seedlings also was significantly lowest in seedlings treated with *H. toruloidea* GanoEF1 in EFB-PKC (26.7%) as compared to the control seedlings (93.3%). The lower percentage of dead seedlings indicates that the lower infection of BSR occurred. Furthermore, the production of POX and PPO were detected in the seedlings pre-inoculated with *H. toruloidea* GanoEF1 and significantly higher than control treatment at the post *G. boninense* challenge inoculation indicating induced resistance is one the mechanism of *H. toruloidea* GanoEF1 to control *Ganoderma* infection. The effect of the formulations on plant growth showed the seedlings treated with *H. toruloidea* GanoEF1 in EFB-PKC gave significantly ($P < 0.05$) highest results on plant height (98.61 cm), girth (38.7 mm), number of frond (11.0), chlorophyll content (60.85 µg/L), root biomass (42.5 g) and leaves biomass (70.1 g) respectively, followed by seedlings treated with granular formulation of EFB-K, RB-PKC and control treatment. The oil palm roots colonized positively to *H. toruloidea* GanoEF1 inoculation with the highest population (between 2.0×10^4 cfu g⁻¹ to 5.3×10^7 cfu g⁻¹) was observed in the treatment of EFB-PKC and EFB-K granular formulations. This phenomenon was supported with the colonization of *H. toruloidea* GanoEF1 within the cortex of the root observed by using transmission electron microscopy (TEM). This study showed the *H. toruloidea* GanoEF1 in granular formulations containing carrier EFB-PKC was the most effective as biological control agent for controlling *Ganoderma* disease and promoting the growth of oil palm seedlings.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**FORMULASI GRANULAR YANG MENGANDUNGI KULAT ENDOFITIK,
Hendersonia toruloidea GanoEF1 UNTUK MENGAWAL PENYAKIT
Ganoderma DAN UNTUK MENGGALAKKAN PERTUMBUHAN POKOK
SAWIT**

Oleh

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Pokok sawit merupakan salah satu tanaman penting di Malaysia dan memainkan peranan penting dalam pembangunan pertanian dan ekonomi negara. Penyakit reput pangkal batang (BSR) yang disebabkan oleh kulat *Ganoderma boninense* adalah ancaman terbesar bagi pengeluaran minyak sawit dan telah didokumentasi menyebabkan kerosakan besar kepada industri sawit di Malaysia. Terdapat beberapa langkah kawalan yang berkesan untuk penyakit BSR. Kulat endofitik telah dikenalpasti sebagai agen kawalan biologi yang berpotensi untuk banyak penyakit. Isolat *Hendersonia toruloidea* GanoEF1 adalah merupakan novel strain yang belum pernah dilaporkan, namun menjanjikan strategi kawalan biologi untuk mengawal *G. boninense*. Penggunaan kulat endofitik dalam pembangunan formulasi juga dipilih kerana banyak kelebihan semasa aplikasi di lapangan. Oleh itu, kajian ini telah direka dengan objektif khusus untuk (i) menentukan substrat nutrien pembawa yang sesuai dengan penghasilan konidia dan kualiti *H. toruloidea* GanoEF1 dalam rumusan formulasi granular, (ii) mengkaji kesan rumusan formulasi granular *H. toruloidea* GanoEF1 untuk mengawal penyakit RPB pada pokok sawit, (iii) menentukan tindak balas biokimia dalam anak sawit yang diinokulatkan dengan *H. toruloidea* GanoEF1, dan (iv) mengkaji kesan-kesan rumusan formulasi granular *H. toruloidea* GanoEF1 pada pertumbuhan vegetatif pokok sawit. Kulat endofitik, *H. toruloidea* GanoEF1 telah dipencilkan daripada akar pokok sawit yang sihat dan dikultur di atas piring agar dektrosa kentang (PDA). Larutan suspensi yang mengandungi 10^8 CFU ml⁻¹ sel-sel konidia telah dirumuskan dalam formulasi granular dengan campuran substrat nutrient pembawa seperti buah tandan buah kosong (EFB), dedak beras (RB), serbuk talkum, sekam padi (PH) dan serbuk habuk kayu (SD) bersama ramuan lengai; serbuk kaolin atau hampas isirung sawit (PKC) dalam larutan alginat-pektin sebagai pengikat. Kajian ini mendapati bahawa nisbah terbaik alginat:pektin untuk pertumbuhan optimum *H. toruloidea*

GanoEF1 adalah 1:3 pada suhu 35 °C dengan jumlah tertinggi sel konidia yang direkodkan adalah 10^8 CFU g^{-1} pada 30 hari selepas penyimpanan. Tiga substrat nutrient pembawa terbaik iaitu EFB, RB dan SD yang berjaya menunjukkan bilangan sel kulat yang tinggi telah dipilih dan diformulasi lagi dengan kaolin (K) atau hampas isirung sawit (PKC) dan ujian kualiti serta aktiviti antagonistik *H. toruloidea* GanoEF1 terhadap *G. boninense* dijalankan setiap bulan selama 12 bulan. Tiga jenis formulasi granular yang mengandungi buah tandan kosong-serbuk kaolin (EFB-K), buah tandan kosong-hampas isirung sawit (EFB-PKC) dan dedak beras-hampas isirung sawit (RB-PKC) berjaya mengekalkan bilangan sel konidia pada 10^5 CFU g^{-1} dan mencatatkan peratusan perencatan pertumbuhan radial (PIRG) terhadap *G. boninense* melebihi 50%. Keberkesanan *H. toruloidea* GanoEF1 dalam formulasi granular EFB-PKC, EFB-K dan RB-PKC telah dipilih bagi menguji keberkesanan setiap satunya dalam mengawal penyakit ke atas anak sawit di tapak semaian. Penindasan penyakit adalah tertinggi dalam rawatan aplikasi *H. toruloidea* GanoEF1 dalam formulasi EFB-PKC dengan pengurangan penyakit sebanyak 65.92% ($P < 0.05$). Peratusan anak sawit mati juga jauh lebih rendah dalam anak sawit yang dirawat dengan *H. toruloidea* GanoEF1 dalam EFB-PKC (26.7%) berbanding dengan kawalan (93.3%). Peratusan anak sawit mati disebabkan oleh jangkitan *G. boninense* yang lebih rendah menunjukkan bahawa jangkitan BSR yang lebih rendah berlaku. Di samping itu, aktiviti POX dan PPO dikesan pada anak sawit yang telah dirawat dengan *H. toruloidea* GanoEF1 dan jauh lebih tinggi daripada rawatan kawalan pada sebelum dan selepas jangkitan *G. boninense*. Kesan formulasi pertumbuhan tumbuhan menunjukkan anak sawit yang dirawat dengan *H. toruloidea* GanoEF1 dalam formulasi granular EFB-PKC telah didapati meningkat secara signifikan ($P < 0.05$) pada ketinggian pokok (98.61 cm), lilitan diameter batang (38.7 mm), jumlah bilangan pelepah (11.0), kandungan klorofil ($60.85 \mu g / L$), serta jumlah biomas akar (42.5 g) dan daun (70.1 g) masing-masing, diikuti oleh anak sawit yang dirawat dengan rumusan EFB-K, RB-PKC dan rawatan kawalan. Kulat *H. toruloidea* GanoEF1 mengkolonisasi akar pokok sawit secara positif dengan populasi tertinggi diperhatikan dalam rawatan formulasi granular EFB-PKC dan EFB-K antara 2.0×10^4 cfu g^{-1} hingga 5.3×10^7 cfu g^{-1} . Fenomena ini disokong dengan pengkolonian *H. toruloidea* GanoEF1 dalam korteks akar yang diamati dengan menggunakan transmisi elektron mikroskop (TEM). Kajian ini menunjukkan *H. toruloidea* GanoEF1 dalam formulasi granular yang mengandungi substrat nutrient pembawa EFB-PKC adalah paling berkesan sebagai agen kawalan biologi untuk mengawal penyakit *Ganoderma* dan meningkatkan pertumbuhan vegetatif pokok sawit.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Oil palm (<i>Elaeis guineensis</i> Jacq.)	4
2.2 Basal stem rot (BSR) disease	5
2.2.1 Symptoms of BSR Infection	7
2.2.2 External Symptoms (Foliar Symptoms)	7
2.2.3 Factors Affecting BSR	9
2.2.4 BSR transmission methods	10
2.3 BSR management practices	11
2.3.1 Fungicides application	11
2.3.2 Physical control: Trench system and mounding	12
2.3.3 Mechanical control: Sanitation	12
2.3.4 Biological control specific for oil palm	13
2.4 Endophytic microorganisms Roles in Agriculture	14
2.4.1 Endophytic fungi	15
2.4.2 Endophytic fungus, <i>Hendersonia toruloidea</i>	17
2.4.3 Endophytic bacteria	17
2.4.4 Endophytic basidiomycete/actinomycetes	18
2.5 Formulation of biological control agent (BCA) product	19
2.5.1 Granular Formulation of biological control agent (BCA) product	20
2.6 Gaps in knowledge	21
3 PREPARATION OF GRANULAR FORMULATION OF <i>Hendersonia toruloidea</i> GanoEF1 AND DETERMINATION OF VIABILITY DURING STORAGE	22
3.1 Introduction	22
3.2 Material and Methods	23

3.2.1	Endophytic fungus, <i>H. toruloidea</i>	23
3.2.2	Preparation of <i>H. toruloidea</i> GanoEF1 inoculant	23
3.2.3	Development of granular formulation of <i>H. toruloidea</i> GanoEF1	23
3.2.3.1	Equipment	23
3.2.3.2	Selection of ratio of selected binder	24
3.2.3.3	Selection of nutrient carrier	25
3.2.4	Effect of long-term storage of formulation on viability <i>H. toruloidea</i> GanoEF1 and efficacy against <i>G. boninense</i>	25
3.2.5	Experimental design and statistical analysis	26
3.3	Results	28
3.3.1	Selection of ratio of selected binder	28
3.3.2	Selection of nutrient carrier	29
3.3.3	Effect of long-term storage of formulation on viability of <i>H. toruloidea</i> GanoEF1 and efficacy against <i>G. boninense</i>	29
3.4	Discussion	31
4	EFFECT OF <i>H. toruloidea</i> GanoEF1 GRANULAR FORMULATION AS AN OIL PALM GROWTH ENHANCER	33
4.1	Introduction	33
4.2	Material and Methods	34
4.2.1	Experimental design	34
4.2.2	Preparation of oil palm seedlings and pre-inoculation with <i>H. toruloidea</i> GanoEF1	34
4.2.3	Data collection on vegetative growth and development	35
4.2.4	Colonisation and population of <i>H. toruloidea</i> GanoEF1 in oil palm roots	35
4.2.5	Transmission Electron Microscopy (TEM)	36
4.2.6	Statistical analysis	36
4.3	Results	36
4.3.1	Effect of different granular formulation of <i>H. toruloidea</i> GanoEF1 as an oil palm growth enhancer	36
4.3.2	Colonisation and population of <i>H. toruloidea</i> GanoEF1 in oil palm roots	40
4.3.3	Transmission Electron Microscopy (TEM)	42
4.4	Discussion	43

5	EVALUATION OF <i>H. toruloidea</i> GanoEF1 GRANULAR FORMULATIONS IN CONTROLLING INFECTION BY <i>G. boninense</i> ON OIL PALM SEEDLINGS	45
	5.1 Introduction	45
	5.2 Material and Methods	46
	5.2.1 Preparation of <i>G. boninense</i> inoculum	46
	5.2.2 Preparation of oil palm seedlings for inoculation	46
	5.2.3 Experimental design	47
	5.2.4 Disease assessment	48
	5.2.4.1 Disease severity of foliar index (DSFI)	48
	5.3 Results	53
	5.4 Discussion	57
6	BIOCHEMICAL RESPONSES INDUCED BY PRE-INOCULATION OF <i>H. toruloidea</i> GanoEF1 IN OIL PALM SEEDLINGS	59
	6.1 Introduction	59
	6.2 Material and Methods	60
	6.2.1 Experimental design	60
	6.2.2 Inoculum Preparation - <i>H. toruloidea</i> GanoEF1 and preparation of seedlings	60
	6.2.3 Sampling for enzyme activity assay	60
	6.2.4 Effect of <i>H. toruloidea</i> GanoEF1 on inducible defense related enzymes	61
	6.2.4.1 Assay of peroxidase (PO) and polyphenol oxidase (PPO)	61
	6.2.4.2 Assay of β -1,3-glucanase	61
	6.2.5 Statistical analysis	61
	6.3 Results	62
	6.3.1 Peroxidase (POX) and polyphenol oxidase (PPO)	62
	6.3.2 β -1,3-glucanase	64
	6.4 Discussion	65
7	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	66
	7.1 Summary and conclusions	66
	7.2 Recommendations for future research	67
	REFERENCES	68
	APPENDICES	
	BIODATA OF STUDENT	

LIST OF TABLES

Table		Page
3.1	Ratio of sodium alginate to pectin in granular formulation of <i>H. toruloidea</i> GanoEF1	24
3.2	Viability of <i>H. toruloidea</i> GanoEF1 conidia cells in granular formulations	30
3.3	Percentage of PIRG value of <i>H. toruloidea</i> GanoEF1 in different formulation and storage at room temperature.	30
4.1	Treatments to determine effect of three granular formulations of <i>H. toruloidea</i> GanoEF1 on oil palm growth	34
4.2	Population of <i>H. toruloidea</i> GanoEF1 from roots of oil palm seedlings aged 3, 6, 9 and 12 months old	41
5.1	Treatments of different granular formulations of <i>H. toruloidea</i> GanoEF1 applied to oil palm seedlings	48
5.2	Signs and symptoms of DSFI scored on a scale of 0-4	49
5.3	Signs and symptoms according to disease severity bole index (DSBI) scored on a scale of 0–4	52
5.4	Signs and symptoms according to disease severity root index (DSRI) scored on a scale of 0–4	52
5.5	Effect of granular formulation of <i>H. toruloidea</i> GanoEF1 on BSR development in oil palm seedlings after eight months of being challenged with <i>G. boninense</i>	54

LIST OF FIGURES

Figure		Page
2.1	Foliar symptoms of <i>Ganoderma</i> infection on mature palm: two or more unopened spear leaves (a), old fronds snapping at the petiole and drooping (b) and collapsed/dead palm (c)	8
2.2	Fruiting bodies of <i>Ganoderma</i> : white mycelium (a), small white button (b) and bracket shaped form (c) at the base of the infected palms	9
3.1	Apparatus used to produce granular formulation of <i>H. toruloidea</i> GanoEF1	24
3.2	Three best granular formulations: rice bran-palm kernel cake (RB-PKC) (a), empty fruit bunch-palm kernel cake (EFB-PKC) (b) and empty fruit bunch-kaolin (EFB-K) (c)	27
3.3	Viability of <i>H. toruloidea</i> GanoEF1 conidia cells in granular formulation with alginate-pectin stored at 30°C, 35°C, and 40°C at 30 days after storage. Note: FA = formulation containing alginate:pectin = 0:1; FB = formulation containing alginate:pectin = 1:3; and FC = formulation containing alginate:pectin = 2:2	28
3.4	Viability of <i>H. toruloidea</i> GanoEF1 conidia cells in granular formulation with different nutrient carriers stored at 30°C, 35°C, and 40°C at 30 days after storage. Note: F-RB = formulation containing RB; F-TP = formulation containing TP; F-EFB = formulation containing EFB; F-PH = formulation containing PH; and F-SD = formulation containing SD.	29
4.1	Effect of <i>H. toruloidea</i> GanoEF1 on plant height of oil palm seedlings. Means with the same alphabet denote a statistically non-significant difference at $P < 0.05$ for each column (n=120)	37
4.2	Effect of <i>H. toruloidea</i> GanoEF1 on girth (top) and frond count (bottom) of oil palm seedlings. Means with the same alphabet denote a statistically non-significant difference at $P < 0.05$ for each column (n=120)	38
4.3	Effect of <i>H. toruloidea</i> GanoEF1 on leaf and root mass of oil palm seedlings. Means with the same alphabet	39

denote a statistically non-significant difference at $P < 0.05$ for each column ($n=120$)

4.4	Effect of <i>H. toruloidea</i> GanoEF1 on leaf and root mass of oil palm seedlings eight months after treatment. Means with the same alphabet denote a statistically non-significant difference at $P < 0.05$ for each column ($n=120$)	40
4.5	A hypha (hy) of <i>Hendersonia</i> GanoEF1 within the cortex of the root (A). The other hypha grows on top of the fibrils material and is partially covered by a dense matrix (B)	42
5.1	Inoculation of oil palm seedlings with <i>G. boninense</i> and treatment with <i>H. toruloidea</i> GanoEF1 granules: (a) a hole was dug up in a large polybag; (b) first application with 5 g of the <i>H. toruloidea</i> GanoEF1 granules on roots and oil palm seedlings were left undisturbed for two weeks; (c) roots of seedlings were placed in contact with RWB colonised with <i>G. boninense</i> PER 71; and (d) a booster application with 50 g of <i>H. toruloidea</i> GanoEF1 granules on roots of oil palm seedlings.	47
5.2	Disease Severity of Foliar Index (DSFI) on infected oil palm seedlings	50
5.3	Progress of disease in oil palm seedlings with different treatments. (Values are mean of 3 replications with vertical bars representing standard error)	53
5.4	Development of BSR in bole and root tissues of oil palm seedlings observed after nine months of being challenged with <i>G. boninense</i> . Means with the same alphabet denote a statistically non-significant difference at $P < 0.05$ for each column ($n = 30$)	55
5.5	Percentage of dead seedlings for different treatments. Means with the same alphabet denote a statistically non-significant difference at $P < 0.05$ for each column ($n = 30$)	56
6.1	Pre and post <i>Ganoderma</i> challenge inoculation sampling intervals for enzyme analysis of oil palm seedlings	60
6.2	Total peroxidase (POX) (top) and polyphenol oxidase (PPO) (bottom) activity in the roots of oil palm seedlings pre-inoculated with pure culture <i>H. toruloidea</i> GanoEF1. Values are mean of 3 replications with	63

vertical bars representing standard error. T1= Seedling untreated, control, T2= Seedling treated with *H. toruloidea* GanoEF1.

- 6.3 Total β -1,3-glucanase activity in the roots of oil palm seedlings pre-inoculated with pure culture *H. toruloidea* GanoEF1 at pre and post *G. boninense* challenge inoculation. Values are mean of 3 replications with vertical bars representing standard error. T1= Seedling untreated, control, T2= Seedling treated with *H. toruloidea* GanoEF1. 64



LIST OF ABBREVIATIONS

BSR	Basal Stem Rot
MA	Malt Agar
MEA	Malt Extract Agar
CMA	Corn Meal Agar
CDA	Czapek's Dox Agar
AMF	Arbuscular Mycorrhiza Fungi
MPOB	Malaysian Palm Oil Board
PGPR	Plant Growth Promoting Rhizobacteria
ISR	Induced Systemic Resistance
PO	Peroxidase
PPO	Polyphenol oxidase
EFB	Empty Fruit Bunch
FFB	Fresh Fruit Bunch
GSM	Ganoderma selective medium
PDA	Potato dextrose agar
PDB	Potato dextrose broth
PIRG	Percentage inhibition of radial growth
CFU/g	Colony forming unit per gram
RCBD	Randomized Completely Block Design
DI	Disease Incidence
AUDPC	Area Under the Disease Progress Curve
DS	Dead Seedlings
DSFI	Disease Severity of Foliar Index
DSBI	Disease Severity of Bole Index
DSRI	Disease Severity of Root Index
RWB	Rubber Wood Blocks
ANOVA	One Way Analysis of Variance

CHAPTER 1

INTRODUCTION

1.1 Background

Oil palm is the world's highest oil crops' producer compared to the other oil crops planted on the same size of land with Malaysia and Indonesia contributes up to 90% of world's palm oil production. High demand for palm oil has led to wider cultivation of oil palm i.e. from 54,000 hectares in 1960 to 5.81 million hectares in 2017, with an annual growth of 10.06%. Besides that, in 2016, it was reported that production of palm oil reached up to 17.32 million tonnes (MPOB, 2017; Nambiappan *et al.*, 2018).

Nowadays, one main constraint faced by the palm oil industry is diseases caused by plant pathogens. The most worrying disease is basal stem rot (BSR) caused by *Ganoderma boninense*. BSR is not new to Malaysia whereby its first attack was not long after oil palm was introduced in the country. The first known attack was on oil palms aged 25 years and above, which were not considered as economically important at that time (Thompson, 1931). Nonetheless, in 1960, as oil palm became a plantation crop, occurrence of BSR started to rise whereby plants, as young as 12–24 months, were attacked (Hoong and Idris, 2010). BSR also infected plants aged 4–5 years (Singh, 1990) and 10 to 15 years in replanted areas (Turner, 1981).

Basal stem rot disease have caused the highest economic loss, estimated about RM 1.5 billion per annum, for both Malaysia and Indonesia compared to other palm oil producing countries such as Africa, Papua New Guinea, and Thailand (Idris *et al.*, 2016; Ommelna *et al.*, 2012;). BSR is very hard to eradicate and numerous ways have been tried to overcome the infection such as applying chemical fungicides, soil mounding, and sanitation by removing infected palms (Mohammed *et al.*, 2014; Idris, 2011). Physical methods such as clean clearing and sanitation involve removal of infected plants from the field. This requires mechanised equipment and a high labour cost. Furthermore, physical methods are more towards preventing spread of disease rather than curing of infected palms. High labour cost and need for periodical observation make physical methods relevant to be applied in severe infected fields, however, it is not sustainable.

On the other hand, manipulation of microorganisms such as fungi, bacteria, and actinomycetes as biological controls has been explored for inhibiting the most severe disease of oil palm (Idris *et al.*, 2010). Biological control is introduced as an alternative to minimise impact on non-targeted organisms and improve agricultural sustainability besides reducing high dependency on mechanisation. Disease caused by *Ganoderma* plays a major role in lowering oil palm yield if no control measures are implemented. Advances in biotechnology have led to a

significant increase in use of microbes as biological control agents against plant pathogens. For instance, biological properties of several antagonistic fungi, such as *Aspergillus* (Shukla & Uniyal, 1989), *Penicillium* (Dharmaputra *et al.*, 1989), and *Trichoderma* (Angel *et al.*, 2016; Sundram, 2013; Nur Ain Izzati & Abdullah, 2008;), and endophytic bacteria (Nurrashyeda *et al.*, 2016; Sapak *et al.*, 2008) have been studied and are proven to have antagonistic effect against *G. boninense*.

The application of microbes as biological agents is a good alternative to control *Ganoderma*, especially in reducing cost of source and maintenance management in the field, compared to chemical fungicides. Endophytic microorganisms, specifically endophytic fungi are therefore a new field of study in biological disease control of *Ganoderma*. The use of endophytic fungi are preferable due to its role as internal colonizers and therefore offering protection from diseases caused by pathogens (Miliute *et al.*, 2015). This was successfully demonstrated with *Hendersonia toruloidea* isolate GanoEF1, *Amphinema byssoides* isolate GanoEF2 and *Phlebia radiata* isolate GanoEF3 which effectively suppressed BSR in oil palm seedlings (Idris *et al.*, 2010).

1.2 Justification

Basal stem rot disease (BSR) is the most destructive disease facing by the oil palm industry. In addition to existing disease management tools, new eco-friendly approaches are being explored to suppress the disease. Studies have proven the potential of endophytic fungi in controlling BSR disease. However, Nurrashyeda's (2018) work used only a pure culture of endophytic fungus, *Hendersonia toruloidea* isolate GanoEF1 in controlling BSR in oil palm seedlings while this study looks at the development of granular formulation of *H. toruloidea* GanoEF1 for application in oil palm plantation. While many biological control products of *Ganoderma* are already in the market, a limited number of these are reported as granular formulation. Liquid and powder formulation are not preferred due to handling difficulties, drift and safety to handlers. The hypothesis of this study is pre-inoculation with granular formulation of *Hendersonia toruloidea* isolate GanoEF1 delays the onset and development of BSR disease in oil palm.

1.3 Research Objectives

- i. To determine carriers that are compatible with viability and quality of endophytic fungus, *H. toruloidea* isolate GanoEF1 in different types of granular formulation.
- ii. To investigate the effects of granular formulations of endophytic fungus, *H. toruloidea* isolate GanoEF1 for controlling BSR in oil palm.
- iii. To determine the biochemical compounds released in oil palm treated with endophytic fungus, *H. toruloidea* isolate GanoEF1.
- iv. To study the effects of granular formulations of endophytic fungus, *H. toruloidea* isolate GanoEF1 on oil palm growth.

REFERENCES

- Abdel-Kader, M.M., El-Mougy, N.S., Aly, M.D.E. and Lashin, S.M. (2012). Long activity of stored formulated bio-agents against some soil-borne plant pathogenic fungi causing root rot of some vegetables. *Journal of Applied Science Research*, 8(4): 1882-1892.
- Abdullah, R. (2005). A decade of oil palm gene manipulation, where are we now?. *Paper presented 9th International Conference on Agricultural Biotechnology*, 6-10th July 2005.
- Abdullah, R. and Idris A.S. (2012). Economic impact of *Ganoderma* incidence on Malaysian oil palm plantation-a case study in Johor. *Oil Palm Industry Economic Journal*, 12(1): 24-30.
- Abdullah, F., Ilias, G. N. M., Nelson, M., Izzati, N. A. M. Z. and Umi Kalsom, Y. (2003). Disease assessment and the efficacy of *Trichoderma* as a biocontrol agent of basal stem rot of oil palms. *Science Putra Research Bulletin*, 11(2): 31-33.
- Alaxandra, A.G. (2014). *Endophytic actinomycetes as a potential agent to control common scab of potatoes*. M.Sc. Thesis. Northern Michigan University.
- Aly, A.H., Debbab, A., Kijer, J. and Prokksch, P. (2010). Fungal endophytes from higher plants: a prolific source of phytochemicals and other bioactive natural products. *Fungal Diversity*, 41(1): 1-6.
- Angel, L. P. L., Sundram, S., Ping, B. T. Y., Yusof, M. T. and Ismail, I. S. (2018). Profiling of anti-fungal activity of *Trichoderma virens* 159c involved in biocontrol assay of *Ganoderma boninense*. *Journal of Oil Palm Research*, 30(1): 83-93.
- Angel, L. P. L., Yusof, M. T., Ismail, I. S., Ping, B. T. Y., Intan Nur Ainni, M. A., Norman, K. and Sundram, S. (2016). An *in vitro* study of the antifungal activity of *Trichoderma virens* 7b and a profile of its non-polar antifungal components released against *Ganoderma boninense*. *Journal of Microbiology*, 54(11): 732-744.
- Anuar, E.N., Nulit, R. and Idris, A.S. (2015). Growth promoting effects of endophytic fungus *Phlebia* GanoEF3 on oil palm (*Elaeis guineensis*) seedlings. *International Journal of Agriculture and Biology* 17(1): 135-141.
- Ariffin, D. and Idris, A.S. (2002). Progress and research on *Ganoderma* basal stem rot of oil palm. *Paper presented at the MPOB Seminar on elevating the national oil palm productivity and recent progress in the management of peat and Ganoderma*. 6-7th May 2002. Bangi Malaysia.

- Ariffin, D., Idris, A.S. and Singh, G. (2000). Status of *Ganoderma* in oil palm. In *Ganoderma Diseases of Perennial Crops*, ed. J. Flood, P.D. Bridge, and M. Holderness, pp. 49-68. Wallingford, UK: CABI Publishing.
- Ariffin, D., Idris, A.S. and Abdul Halim, H. (1989). Significance of the black line within oil palm tissue decay by *Ganoderma boninense*. *Journal of Elaeis* 1:11-16.
- Ariffin, D. and Idris, A.S. (2003). Progress and research on *Ganoderma* Basal Stem Rot of oil palm. In: Mohd Basri, W., Chan, K.W., Mohd Tayeb, D. and Sundram, S. (Eds.). *Proc. Seminar on Elevating National Oil Palm Productivity and Recent Progress in the Management of Peat and Ganoderma*. Malaysian Palm Oil Board, Bangi, Selangor, Malaysia, Pp. 167-205.
- Arnold, A.E and Lutzoni, F. (2007). Diversity and host range of foliar fungal endophytes: 25 are tropical leaves biodiversity hotspots? *Ecology*, 88: 541-549.
- Arnold, A.E., Meji'a L.C., Kylo, Rojas, E.I., Aybard, Z., Robbins, N. and Herre, A. (2003). Fungal endophytes limit pathogen damage in a tropical tree. *PNAS*, 100: 15649-15654.
- Azizah, H. (2003). *Ganoderma* versus mycorrhiza. *Oil Palm Bulletin*, 47: 6-14.
- Badalyan, M.S., Innocenti, G. and Garibyan, G.N. (2004). Interactions between Xylophilic mushrooms and mycoparasitic fungi in dual-culture experiments. *Phytopathologia Mediterranea*, 43: 44-48.
- Bailey, B.A., Bae, H., Strem, M.D., Roberts, D.P., Thomas, S.E., Crozier, J., Samuels, G.J., Choi, I.Y. and Holmes, K.A. (2006). Fungal and plant gene expression during the colonization of cocoa seedlings by endophytic isolates of four *Trichoderma* species. *Planta*, 224: 1449-1464.
- Bailey, B.A., Bae, H., Strem, M.D., Crozier, J., Thomas, S.E., Samuels, G.J., Vinyard, B.T. and Holmes, K.A. (2008). Antibiosis, mycoparasitism, and colonization success for endophytic *Trichoderma* isolates with biological control potential in *Theobroma cacao*. *Biological control* 46(1): 24-35.
- Bakker, P. A. H. M., Raaijmakers, J. M., Bloemberg, G. V., Hofte, M., Lemanceau, P. and Cooke, M. (2007). New perspectives and approaches in plant growth-promoting rhizobacteria research. *European Journal of Plant Pathology* 119: 241-242.
- Basari, S. (2003). *Modification of Alginate Controlled-release Formulations of Alachlor and Diuron with Pectin: Their Release Kinetics and Efficacy*. Unpublished doctoral dissertation, Universiti Putra Malaysia.

- Benjamin, M. and Chee, K.H. (1995). *Basal stem rot of oil palm- a serious problem on inland soils. MAPPS Newsletter*, 19(1): 3
- Bivi, R.M., Siti Noor Farhana, M., Khairulmazmi, A. and Idris, A. (2010). Control of *Ganoderma boninense*: A causal agent of basal stem rot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture & Biology* 12(6): 833-839.
- Breton, F., Hasan, Y., Hariadi, S., Lubis, Z. and De Franqueville, H. (2006). Characterization of parameters for the development of an early screening test for basal stem rot tolerance in oil palm progenies. *Journal of Oil Palm Research*, 24–36.
- Campbell, C.L. and Madden, L.V. (1990). *Introduction to plant disease epidemiology*, pp. 113-121. USA: John Wiley and Sons.
- Card, S., Johnson, L., Teasdale, S. and Caradus, J. (2016). Deciphering endophyte behaviour: the link between endophyte biology and efficacious biological control agents. *FEMS Microbiology Ecology*, 92(8): 1-19.
- Chung, G.F. (2011). Management of *Ganoderma* diseases in oil palm plantations. *Planter*, 87: 325-339.
- Corley, R.H.V. (1976). The genus *Elaeis*. In *Oil Palm Research*, ed. R.H.V. Corley, J.J. Hardon, and B.J. Wood, pp. 3-5. Netherlands: Elsevier Scientific Publisher.
- Dastogeer, K. M. (2018). Influence of fungal endophytes on plant physiology is more pronounced under stress than well-watered conditions: A meta-analysis. *Planta*, 248(6): 1403-1416.
- Dhanya N.N. and Padmavathy, S. (2014). Impact of endophytic microorganisms on plants, environment and humans. *The Scientific World Journal* 2014(2014): 1-11.
- Dharmaputra, O.S., Tjitrosomo, O.H.S. and Abadi, A.I. (1989). Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. *Biotropia*, 3: 41-49.
- Doni, F. , Anizan, I., CheRadziah, C.M.Z., Wan Natasya, W. A., Abidah, A. and Wan Mohtar, W.Y. (2014). Enhanced rice seedling growth by *Trichoderma* sp. FCR1. In *Proceedings of the 13th Symposium of the Malaysian Society of Applied Biology 2014*, ed. M.A. Masni, Y. Shahrizad, O. Ramlan, M.N. Mahanem, S. Abdullah, and A.S. Norrakiah, pp. 123-125. Cherating, Pahang, Malaysia.
- Evans, H.C., Holmes, K.A and Thomas, S.E. (2003). Endophytes and mycoparasites associated with an indigenous forest tree, *Theobroma*

- gileri*, in Ecuador and preliminary assessment of their potential as biocontrol agents of cocoa diseases. *Mycological Progress*, 2: 149-160.
- Faruk, M.I., Rahman, M.L., Mustafa, M.M.H., Rahman, M.M. and Rahman, M.A. (2014). Screening of carrier materials to formulate *Trichoderma harzianum* based bio-fungicide against foot and root rot disease of tomato (*Lycopersicon esculentum* L.). *Bangladesh Journal of Agricultural Research*, 39(2): 197-209.
- Flood, J., Bridge, P.D. and Holderness, M. (2000). *Ganoderma Diseases of Perennial Crops*. CABI Publishing, Wallingford. UK.
- Flood, J., Keenan, L., Wayne, S. and Hasan, Y. (2005). Studies on oil palm trunk as sources of infection in the field. *Mycopathologia*, 159: 101-107.
- Flood, J. and Hasan, Y. (2004). Basal stem rot - taxonomy, biology, epidemiology, economic status and control in South East Asia and Pacific Islands. Proceedings of the Malaysian Palm Oil Board Conference. May 2004. pp. 117-33. Kuala Lumpur: MPOB.
- Gao, F., Dai, C. and Liu, X. (2010). Mechanisms of fungal endophytes in plant protection against pathogens. *African Journal of Microbiology Research*, 4: 1346–1351.
- Garbeva, P., van Overbeek, L.S., van Vuurde, J.W.L. and van Elsas, J.D. (2001). Analysis of endophytic bacterial communities of potato by plating and denaturing gradient gel electrophoresis (DGGE) of 16S rDNA based PCR fragments. *Microbial Ecology*, 41: 369-383.
- Gomez, K.A. and Gomez, A.A. (1984). *Statistical procedures for agricultural research*. John Wiley & Sons.
- Gomez-Vidal, S., Lopez-Liorca, L.V., Jansson, H.B and Salinas, J. (2006). Endophytic colonization of date palm (*Phoenix dactylifera* L.) leaves by entomopathogenic fungi. *Micron*, 37: 624-632.
- Goudjal, Y., Toumatia, O., Sabaou, N., Barakate, M., Mathieu, F. and Zitouni, A. (2013). Endophytic actinomycetes from spontaneous plants of Algerian Sahara: indole-3-acetic acid production and tomato plants growth promoting activity. *World Journal of Microbiology and Biotechnology*, 29: 1821-1829.
- Gray, E.J. and Smith, D.L. (2005). Intracellular and extracellular PGPR: commonalities and distinctions in the plant–bacterium signaling processes. *Soil Biology and Biochemistry*, 37(3): 395-412.
- Hallman, J., Quadt-Halmann, A., Mahafee, W.F. and Kloepper, J.W. (1997). Bacterial endophytes in agricultural crops. *Canadian Journal Of Microbiology*, 43: 895-914.

- Haniff, M.H., Ismail, S. and Idris, A.S. (2005). Gas exchange responses of oil palm to *Ganoderma boninense* infection. *Asian Journal of Plant Sciences*, 4(4): 438-444.
- Harman, G.E., Howell, C.R., Viterbo, A., Chet, I. and Lorito, M. (2004). *Trichoderma* species-Opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2(1): 43-56.
- Hasan, Y. and Turner, P.D. (1994). Research at BAH LIAS research station on basal stem rot of oil palm. In: Holderness, M. (Eds.), Proceedings of the 1st International Workshop on Perennial Crop Diseases caused by *Ganoderma*, 28 November-3 December. UPM, Serdang, Selangor, Malaysia.
- Hasegawa, S., Meguro, A., Shimizu, M., Nishimura, T., Kunoh, H. (2006). Endophytic actinomycetes and their interactions with host plants. *Actinomycetologica*. 20: 72–81.
- Ho, C.T. (1998). Safe and efficient management systems for plantation pests and diseases. *The Planter*, 74 (868): 369-385.
- Hoong, H. W. and Idris, A. S. (2010). Control and management of *Ganoderma* disease of oil palm in Malaysia. In: Second International Seminar on Oil Palm Diseases: Advances in *Ganoderma* Research and Management, Yogyakarta.
- Ho, Y. W. and Khairuddin, H. (1995). Pathogenicity and histopathology of *Ganoderma boninense* on oil palm seedlings. *Journal of Bioscience*, 6: 155-164.
- Ho, Y.W. and Nawawi, A. (1985). *Ganoderma boninense* Pat. from basal stem rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. *Pertanika*, 8: 425-428.
- Howell, C.R. (2003). Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: The history and evolution of current concepts. *Plant Disease*, 87: 4-10.
- Hushiarian, R., Nor Azah, Y. and Dutse, S.W. (2013). Detection and control of *Ganoderma boninense*: strategies and perspectives. *SpringerPlus*, 2(555): 1-12.
- Hyde, K.D. and Soyong, K. (2008). The fungal endophyte dilemma. *Fungal Diversity*, 33: 163-173.
- Leggett, M., Leland, J., Kellar, K. and Epp, B. (2011). Formulation of microbial biocontrol agents—an industrial perspective. *Canadian Journal of Plant Pathology*, 33(2): 101-107.

- Idris, A.S., Ariffin, D., Swinburne, T. and Watt, T.A. (2000). The identity of *Ganoderma* species responsible for basal stem rot (BSR) disease of oil palm in Malaysia-morphological characteristics. *MPOB Information Series, No. 102*. 4pp.
- Idris, A.S. (2009). Basal stem rot in Malaysia- Biology, epidemiology, economic importance, detection and control. *Proc. of the International Conference on Pests and Diseases of Importance to the Oil Palm Industry*. May 2004, Kuala Lumpur.
- Idris, A.S., Arifurrahman & Khushairi, A. (2010). Hexaconale as a preventive treatment for managing *Ganoderma* in oil palm. *MPOB Information Series -533- MPOB TS. No 75*. 4pp.
- Idris, A.S. Ismail, S. and Ariffin, D. (2004). Innovative technique of sanitation for controlling *Ganoderma* at replanting. *MPOB Information Series, No. 213*, 4pp.
- Idris, A. S., Ismail, S., Arrifin, D. and Ahmad, H. (2002). Control of *Ganoderma*-infected palm-development of pressure injection and field applications. *MPOB Information Series, No. 131*. 4pp.
- Idris, A.S. (2011). Biology, detection, control and management of *Ganoderma* in oil palm. In *Further Advances in Oil Palm Research (2000-2010)*, M.W. Basri, Y.M. Choo, and K.W. Chan, pp. 485-521. Bangi, Malaysia. MPOB Publication.
- Idris, A.S., Noor Haida, S and Nurrashyeda, R. (2010). GanoEF1-A fungal biocontrol agent for *Ganoderma* in oil palm. *MPOB Information Series No. 501, MPOB TT No. 444*, June 2010, 4pp.
- Idris, A. S. and Norman, K. (2015). Upstream management practices to reduce *Ganoderma* disease outbreak – The industry’s approaches. *Proc. of the PIPOC 2015 International Palm Oil Congress (Agriculture, Biotechnology and Sustainability)*. MPOB, Malaysia (Oral paper). pp. 9-23. Bangi, Selangor, Malaysia. MPOB.
- Idris, A. S., Nurrashyeda, R., Rusli, M.H., Sundram, S. and Norman, K. (2010). Standard operating procedure (SOP) guidelines for managing *Ganoderma* disease in oil palm. pp. 41. Bangi, Selangor, Malaysia. MPOB.
- Idris, A. S., Sharifah-Muzaimah, S. A., Madihah, A. Z., Norman, K., Kushairi, A., Choo, Y. M. and Wan Ismail, W. H. (2014). EMBIO™ actinoPLUS for biological control of *Ganoderma* disease. *MPOB TT Information Series, 544*. 4pp.
- Ishaq, I., Alias, M.S., Kadir, J. and Kasawani I. (2014). Detection of basal stem rot disease at oil palm plantation using sonic tomography. *Journal of Sustainability Science and Management*, 9(2):52-57.

- Jia, M., Chen, L., Xin, H. L., Zheng, C. J., Rahman, K., Han, T. and Qin, L. P. (2016). A Friendly Relationship between Endophytic Fungi and Medicinal Plants: A Systematic Review. *Frontiers in Microbiology*, 7(906): 1-14.
- Jonathan, R.G., Crystal, A.M., Karen, A.T., Nicola, J.D., Anna, S.B. and Kari, E.D. 2013. Inside root microbiome: Bacterial root endophytes and plant growth promotion. *American Journal of Botany* 100(9): 1738-1750.
- Junaid, J. M., Dar, N. A., Bhat, T. A., Bhat, A. H. and Bhat, M. A. (2013). Commercial biocontrol agents and their mechanism of action in the management of plant pathogens. *International Journal of Modern Plant & Animal Sciences*, 1(2): 39-57.
- Kandan, A., Bashkaran, R. and Samiyappan, R. (2010). Ganoderma- a basal stem rot disease of coconut palm in South Asia and Asia Pacific regions. *Archives of Phytopathology and Plant Protection*, 43: 1445-1449.
- Khairudin, H. (1990). *Basal stem rot of oil palm: incidence, etiology and control*. Master of Agriculture Science Thesis, Universiti Pertanian Malaysia, Selangor, Malaysia.
- Khairudin, H. and Chong, T.C. (2008). An overview of the current status of *Ganoderma* basal stem rot and its management in a large plantation group in Malaysia. *Planter*, 84(988): 469-482.
- Khare, E., Mishra, J. and Arora, N. K. (2018). Multifaceted interactions between endophytes and plant: Developments and Prospects. *Frontiers in Microbiology*, 9: 1-12.
- Kloepper, J. W., Lifshitz, R. and Zablotticz, R. M. (1989). Free-living bacterial inocula for enhancing crop productivity. *Trend Biotechnology*, 7: 39-43.
- Kloepper, J. W. (2003). A review of mechanisms for plant growth promotion by PGPR. In *6th International PGPR workshop (Abstracts and short papers*, ed. Reddy, M. S., Anandaraj, M., Eapen, S. J., Sarma, Y. R., Kloepper, J. W., Indian Institute of Spices Research, Calicut, India.
- Kloepper, J.W. and Ryu, C.M. (2006). Bacterial endophytes as elicitors of induced systemic resistance. In *Microbial Root Endophytes*, pp. 33-52. Springer, Berlin, Heidelberg.
- Kobayashi, D.Y. and Palumbo, J.D. (2000). Bacteria endophytes and their effect on plant and uses in agriculture. In: Bacon, C.W. and Jr. White, J.F. (Eds.), *Microbial Endophytes*, pp:199 -233. Marcel Dekker, Inc. New York.
- Kogel, K.H., Franken, P., and Huckelhoven, R. (2006). Endophyte or parasite- What decide?. *Current Opinion in Plant Biology*, 9: 358-363.

- Kumar, S., Kumar, R. and Hari, O. M. (2013). Shelf-life of *Trichoderma viride* in talc and charcoal based formulations. *Indian Journal of Agricultural Sciences*, 83(5): 566-9.
- Kushairi, A, Loh S. K. , Azman, I., Hishamuddin, E.L., Ong-Abdullah, M., Izuddin, Z. B., Razmah, G., Sundram, S. and Parveez, G. K. (2017). Oil palm economic performance in Malaysia and R&D progress in 2017. *Journal of Oil Palm Research*, 30(2): 163-95.
- Lata, R., Chowdhury, S., Gond, S. K. and White Jr, J. F. (2018). Induction of abiotic stress tolerance in plants by endophytic microbes. *Letters in Applied Microbiology*, 66(4): 268-276.
- Lazarovits, G. and Nowak, J. (1997). Rhizobacteria for improvement of plant growth and establishment. *HortScience* 32(2): 188-192.
- Lehtonen, P.T., Helander, M., Siddiqui, S.A., Lehto, K. and Saikkonen, K. (2006). Endophytic fungus decreases plant virus infections in meadow ryegrass (*Lolium pratense*). *Biology Letters*, 2: 620-623.
- Lim, K. H., Chuah, J. H. and HO, C.H. (1993). Effects of soil heaping on *Ganoderma* infected oil palms. *Proc. of the 1993 PORIM Int'l Palm Oil Congress 'Update and Vision' - Agriculture* (Jalani et al. eds.). PORIM, Bangi, Selangor, Malaysia: 735-738.
- Lisnawita., Hanum, H. and Tantawi, A. R. (2016). Survey of Basal Stem Rot Disease on Oil Palms (*Elaeis guineensis* Jacq.) in Kebun Bukit Kijang, North Sumatera, Indonesia. *International Conference on Agricultural and Biological Sciences (ABS 2016), Earth and Environmental Science*, 41: 1-5.
- Liu, C.H., Zou, W.X., Lu, H. and Tan, R.X. (2001). Antifungal activity of *Artemisia annua* 11 endophyte cultures against phytopathogenic fungi. *Journal of Biotechnology*, 88: 277-282.
- Lo, C. T. and Lin, C.Y. (2002). Screening strains of *Trichoderma* spp for plant growth enhancement in Taiwan. *Plant Pathology Bulletin* 4, (2002): 215-220.
- Lugtenberg, B. J., Caradus, J. R. and Johnson, L. J. (2016). Fungal endophytes for sustainable crop production. *FEMS Microbiology Ecology*, 92(12): 1-17.
- Marciano, R.R., Rute ,T. S., Alan, W.V., Pomella, Cristina S.M., Wellington, L., Araujo, D.R., Dos Santos and Joao, L.A. (2005). Diversity of endophytic fungal community of cacao (*Theobroma cacao*) and biological control of *Crinipellis pernicioso*, causal agent of witches' broom disease. *International Journal of Biological Sciences*, 1: 24-33.

- Marco, G.M. and Stall, R.E. (1983). control of bacterial spot of pepper initiated by strains of *Xanthomonas campestris* pv. *vesicatoria* that differ in sensitivity to copper. *Plant Disease*, 67: 779–781.
- Marella, S. (2014). Bacterial endophytes in sustainable crop production: Applications, recent developments and challenges ahead. *International Journal of Life Sciences Research*, 2(2): 46-56.
- Mazliham, M.S., Pierre, L. and Idris, A.S. (2007). Towards Automatic Recognition and Grading of *Ganoderma* Infection Pattern Using Fuzzy Systems. *International Journal of Medical and Health Sciences*, 1(1): 1-6.
- Mejia, L.C., Rojas, E.I., Maynard, Z., Van Bael, S.A., Arnold, A.E., Hebbbar, P., Samuels, G.J., Robbins, N. and Herre, E.A. (2008). Endophytic fungi as biocontrol agents of *Theobroma cacao* pathogens. *Biological Control*, 46: 4-14.
- Mejia, L.C., Rojas, E.I., Maynard, Z., Arnold, A.E., Kylo, D., Robbins, N. and Herre, E.A. (2003). *Inoculation of beneficial endophytic fungi into Theobroma cacao tissues*. Proceedings of the 14th International Cocoa Research Conference, Accra, Ghana, II. pp: 669–705.
- Miliute, I., Buzaitė, O., Baniulis, D. and Stanys, V. (2015). Bacterial endophytes in agricultural crops and their role in stress tolerance: A review. *Zemdirbyste-Agriculture*, 102(4): 465-478.
- Mohammed, C. L., Rimbawanto, A. and Page, D. E. (2014). Management of basidiomycete root-and stem-rot diseases in oil palm, rubber and tropical hardwood plantation crops. *Forest Pathology*, 44(6): 428-446.
- MPOB. 2003. *Research Progress Report 2003*. MPOB, Bangi, Selangor, Malaysia.
- MPOB. (2017). *Review of the Malaysian oil palm industry 2017*. MPOB, Bangi, Selangor, Malaysia.
- Mudarisova, R. K., Koptyaeva, E. I. and Badykova, L. A. (2017). Solid-phase modification of chitosan, pectin, and arabinogalactan with poorly soluble herbicide chlorsulfuron. *Polymer Science, Series B*, 59(5): 570-576.
- Murphy, D. J. (2014). The future of oil palm as a major global crop: opportunities and challenges. *Journal of Oil Palm Research*, 26(1): 1-24.
- Nahas, E. (1996). Factors determining rock phosphate solubilization by microorganisms isolated from soil. *World Journal of Microbiology and Biotechnology*, 12: 567–572
- Naher, L., Ho, C.LI, Tan, S.G., Yusuf, U.K. and Abdullah, F. (2011). Cloning of transcripts encoding chitinases from *Elaeis guineensis* Jacq. and their

expression profiles in response to fungal infections. *Physiological and Molecular Plant Pathology*, 76: 96–103.

- Naher, L., Tan, S.G., Yusuf, U.K., Ho, C.L. and Siddiquee, S. (2012). Activities of chitinase enzymes in the oil palm (*Elaeis guineensis* Jacq.) in interactions with pathogenic and non-pathogenic fungi. *Plant Omics*, 5(4): 333-336.
- Naher, L., Shafiquzzaman, S., Umi Kalsom, Y. and Mondal M.M.A. (2015). Issues of *Ganoderma* spp. and basal stem rot disease management in oil palm. *American Journal of Agricultural Science*, 2(3): 103-107.
- Nair, D. N. and Padmavathy, S. (2014). Impact of endophytic microorganisms on plants, environment and humans. *The Scientific World Journal*, (2014): 1-11.
- Nambiappan, B., Ismail, A., Hashim, N., Ismail, N., Nazrima, D. S., Nik Abdullah, N. I., Omar, N., Salleh, K.M., Nur Ain, M.H. and Kushairi, A. (2018). Malaysia: 100 years of resilient palm oil economic performance. *Journal of Oil Palm Research*, 30(1): 13-25.
- Nazeeb, M., Barakabah, S.S. and Loong, S.G. (2000). Potential of high density oil palm plantings in diseased environments. *The Planter*, 76: 699-710.
- Nega, A. (2014). Review on concepts in biological control of plant pathogens. *Journal of Biology, Agriculture and Healthcare*, 4(27): 33-54.
- Noraini, M.J., (2007). *Effects of Empty Fruit Bunch Compost and arbuscular mycorrhiza on Nutrient Uptake and Growth of Grain Maize*. Unpublished doctoral dissertation, Universiti Putra, Malaysia,
- Nur Ain Izzati, M.Z. and Abdullah, F. (2008). Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44 (3): 101-107.
- Nurrashyeda, R., Idris A.S., Madihah, A.Z., Maizatul-Suriza, M. and Noor Haida, S. (2018). Biocontrol of Basal Stem Rot (BSR) Disease of Oil Palm Using Endophytic Fungus, *Hendersonia* sp. *International Journal of Pure and Applied Mathematics*, 118(24): 1-22.
- Nurrashyeda, R., Idris, A. S. and Ramle, M. (2011). Viability test of alginate granular formulation of *Hendersonia* GanoEF1 against *Ganoderma boninense* *in vitro*. *Proc. of the Third MPOB-IOPRI International Seminar: Integrated Oil Palm Pests and Disease Management*. pp. 111-115. Bangi, Selangor, Malaysia. MPOB.
- Nur Sabrina, A.A., Sariah, M. and Zaharah, A.R. (2012). Suppression of Basal Stem Rot Disease Progress in Oil Palm (*Elaeis guineensis*) after Copper and Calcium Supplementation. *Pertanika Journal of Tropical Agricultural Science*, 35: 13-24.

- Ommelna, B.G., Jennifer, A.N. and Chong, K.P. (2012). The potential of chitosan in suppressing *Ganoderma boninense* infection in oil palm seedlings. *Journal of Sustainability Science and Management*, 7(2): 186-192.
- Orole, O.O. and Adejumo, T.O. (2009). Activity of fungal endophytes against four maize wilt pathogens. *African Journal of Microbiology Research*, 3(12): 969-973.
- Oses, R., Valenzuela, S. and Freer, J. (2006). Evaluation of fungal endophytes or lignocellulolytic enzyme and wood biodegradation. *International Biodeterioration and Biodegradation*, 57: 129-135.
- Ownley, B.H., Griffin, M.R., Klingeman, W.E. Gwinn, K.D., Moulton, J.K. and Pereira, R.M. (2008). *Beauveria bassiana*: endophytic colonization and plant disease control. *Journal of Invertebrate Pathology*, 98: 267-270.
- Ownley, B.H., Pereira, R.M., Klingeman, W.E., Quigley, N.B. and Leckie, B.M. (2004). *Beauveria bassiana*, a dual purpose biocontrol organism with activity against insect pests and plant pathogens. Emerging Concepts In *Plant Health Management*, pp: 255-269.
- Ownley, B.H., Gwinn, K.D. and Vega, F.E. (2009). Endophytic fungal entomopathogens with activity against plant pathogens: ecology and evolution. *Biocontrol*, 55: 113-128.
- Pandya, U. and Saraf, M. (2010). Application of fungi as a biocontrol agent and their biofertilizer potential in agriculture. *Journal of Advances Research*, 1(1): 90-99.
- Pal, K.K. and Gardener, B.M., (2006). Biological control of plant pathogens. *The Plant Health Instructor*, 2: 1117-1142.
- Paterson, R.R.M. (2007). Internal amplification controls have not been employed in diagnostic fungal PCR, hence potential false negative results. *Journal of Applied Microbiology*, 102: 1-10.
- Paterson, R.R.M., Sariah, M., Zainal Abidin, M.A. and Lima, N. (2008). Prospects for inhibition of lignin degrading enzymes to control *Ganoderma* white rot of oil palm. *Current Enzyme Inhibition*, 4: 172-179.
- Persello-Cartieaux, F., Nussaume, L. and Robaglia, C. (2003). Tales from the underground: molecular plant-rhizobacteria interactions. *Plant Cell Environment*, 26: 189-199.
- Pilotti, C.A. (2005). Stem rots of oil palm caused by *Ganoderma boninense*: pathogen biology and epidemiology. *Mycopathologia*, 159: 129-137.
- Pinruan, U., Rungjindamai, N., Choeyklin, R., Lumyong, S., Hyde, K.D. and Jones, E.B.G. (2010). Occurrence and diversity of basidiomycetous

- endophytes from the oil palm, *Elaeis guineensis* in Thailand. *Fungal Diversity*, 41: 71-88.
- Prapagdee, B., Kuekulvong, C. and Mongkolsuk, S. (2008). Antifungal potential of extracellular metabolite produced by *Streptomyces hygroscopicus* against phytopathogenic fungi. *International Journal of Biological Sciences*, 4: 330-337.
- Ramanathan, A., Vidhasekaran, P. and Samiyappan, R., (2000). Induction of defense mechanisms in greengram leaves and suspension-cultured cells by *Macrophomina phaseolina* and its elicitors. *Journal of Plant Diseases and Protection*, 1: 245-257.
- Ramamoorthy, V., Raguchander, T. and Samiyappan, R. (2002). Induction of defense-related proteins in tomato roots treated with *Pseudomonas fluorescens* Pf1 and *Fusarium oxysporum* f. sp. *lycopersici*. *Plant and Soil*, 239(1): 55-68.
- Ramasamy, S. (1972). Cross-infectivity and decay ability of *Ganoderma* species parasitic to rubber, oil palm and tea. In: *Ganoderma* diseases of perennial crops. Flood, J., Bridge, P.D. and Holderness, M. 2000. CABI publishing: UK.
- Rasmussen, J.B., Hammerschmidt, R. and Zook, M.N. (1991). Systemic induction of salicylic acid accumulation in cucumber after inoculation with *Pseudomonas syringae* pv *syringae*. *Plant Physiology*, 97(4): 1342-1347.
- Ravensberg, W.J. (2010). *The development of microbial pest control products for control of arthropods: a critical evaluation and a roadmap to success*. PhD. Thesis, Wageningen University, Wageningen, NL.
- Rees, R.W., Flood, J., Hasan, Y. and Cooper, R. M. (2009). Basal stem rot infection of oil palm; mode of root and lower stem invasion by *Ganoderma*. *Plant Pathology*, 58: 982-989.
- Rivera-Orduna, F.N., Suarez-Sanchez, R.A., Flores-Bustamante, Z.R., Gracida-Rodriguez, J.N. and Flores-Cotera, L.B. (2011). Diversity of endophytic fungi of *Taxus globosa* (Mexican yew). *Fungal Diversity*, 47: 65-74.
- Rosenani, A.B., Rovica, R., Cheah, P.M. and Lim, C.T. (2016). Growth performance and nutrient uptake of oil palm seedling in prenursery stage as influenced by oil palm waste compost in growing media. *International Journal of Agronomy*, 1-8.
- Roslan, M. M. N. and Haniff, M. H. (2004). The Role Of Leaf Area Index (LAI) In Oil Palm. *Oil Palm Bulletin*, 48: 11-16.
- Rungjindamai, N., Pinruan, U., Choeyklin, R., Hattori, T. and Jones, E.B.G. (2008). Molecular characterization of basidiomycetous endophytes

- isolated from leaves, rachis and petioles of the oil palm, *Elaeis guineensis*, in Thailand. *Fungal Diversity*, 33:139-161.
- Sanderson, F.R. (2005). An insight into dispersal of *Ganoderma boninense* on oil palm. *Mycopathologia* 159: 139-141.
- Sapak, Z., Meon, S. and Zam, A. (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture & Biology*, 10: 127-132.
- Sariah, M. (2003). The potential of biological management of basal stem rot of oil palm: issues, challenges and constraints. *Oil Palm Bulletin*, 47: 1-5.
- Sariah, M., Choo C. W., Zakaria, H. and Norihan, M. S. (2005). Quantification and characterization of *Trichoderma* spp. from different ecosystems. *Mycopathologia*, 159: 113–117.
- SAS. (1995). Guide to the use of PC-SAS Version 6.04 for DOS for Statistical Analysis, SAS Institute, Cary, N.C.
- Schena, L., Nigro, F., Pentimone, I., Ligorio, A. and Ippolito, A. (2003). Control of postharvest rots of sweet cherries and table grapes with endophytic isolates of *Aureobasidium pullulans*. *Journal of Postharvest Biology and Technology*, 30: 209-220.
- Schulz, B., Boyle, C. and Sieber, T. (2006). *Soil biology*, vol. 9. Springer, Berlin, Germany.
- Schulz, B. and Boyle, C., (2005). The endophytic continuum. *Mycological Research*, 109: 661-686.
- Seo, G.S. and Kirk, P.M. (2000). Ganodermataceae: Nomenclature and Classification. In: *Ganoderma diseases of perennial crop*. In *Ganoderma diseases of Perennial Crops*, eds, J. Flood, P.D. Bridge, and M. Holderness, pp. 1-3. CABI Publishing, UK
- Shamala, S. and Idris, A. S. (2009). *Trichoderma* as a biological agent against *Ganoderma* in oil palm. MPOB Information Series, Bangi, Selangor, Malaysia. 4pp.
- Shariffah-Muzaimah, S.A., Idris, A.S., Madihah, A.Z., Dzolkhifli, O., Kamaruzzaman, S. and Cheong, P.C.H. (2015). Isolation of actinomycetes from rhizosphere of oil palm (*Elaeis guineensis* Jacq.) for antagonism against *Ganoderma boninense*. *Journal of Oil Palm Research*, 27(1): 19-29.
- Shariffah-Muzaimah, S. A., Idris, A. S., Madihah, A. Z., Dzolkhifli, O., Kamaruzzaman, S. and Maizatul-Suriza, M. (2018). Characterization of *Streptomyces* spp. isolated from the rhizosphere of oil palm and evaluation of their ability to suppress basal stem rot disease in oil palm

seedlings when applied as powder formulations in a glasshouse trial. *World Journal of Microbiology and Biotechnology*, 34(15): 1-14.

- Shukla, A.N. and Uniyal, K. (1989). Antagonistic interactions of *Ganoderma lucidum* (lyss) karst. against some soil microorganisms. *Journal of Current Science*, 58: 265-267.
- Singh, G. (1991). *Ganoderma*-the scourge of oil palm in coastal areas. *The Planter*, 67: 421-444.
- Singh, G. (1990). *Ganoderma*-the scourge of oil palms in the coastal areas. In *Proceedings of the Ganoderma Workshop*, ed. D. Ariffin, and S. Jalani, pp. 113-131. PORIM, Bangi, Selangor, Malaysia.
- Singh, R. and Dubey A. K. (2015). Endophytic actinomycetes as emerging source for therapeutic compounds. *Indo Global Journal Pharmaceutical Science*, 5(2): 106-116.
- Singh, S.K., Sheeba, R.D. Gupta, S. Rajendra, S.K. Verma, M.A. Siddiqui, A. Mathur and Agarwal, P.K. (2011). Assessment of the role of *Pseudomonas fluorescens* as biocontrol agent against fungal plant pathogens. *Current Botany*, 2(3): 43-46.
- Sieber, T. (2007). Endophytic fungi in forest trees: are they mutualists?. *Fungal Biology Reviews*, 21: 75-89.
- Skidmore, A.M. and Dickinson, C.H. (1976). Colony interaction and hyphal interference between *Septoria nodorum* and phylloplane fungi. *Transaction of British Mycological Society*, 66(1): 57-64.
- Smith, S. E. and Read, D. J. (2008). Arbuscular mycorrhizas. In *Mycorrhizal symbiosis*. Academic press.
- Soepena, H., Purba, R. Y. and Pawirosukarto, S. (2000). A control strategy for basal stem rot (*Ganoderma*) on oil palm. In *Ganoderma Disease of Perennial Crops*, ed. J. Flood, pp. 83-88. CAB International, UK.
- Souza, L.Q.A., Souza, L.D.A., Astolfi Filho, S., Belém Pinheiro, L.M., Sarquis, M.I.M. and Pereira, O.J. (2004). Atividade Antimicrobiana de Fungus Endofíticos Isolados de Plantas Tóxicas da Amazônia: *Palicourea longiflora* (aubl.) rich e *Strychnos cogens* Bentham. *Acta Amazônica*, 34(2): 185-195.
- Sriram, S. and Savitha, M. J. (2011). Enumeration of colony forming units of *Trichoderma* in formulations – precautions to be taken to avoid errors during serial dilution. *Journal of Biological Control*, 25: 64-67.
- Strobel, G.A. (2002). Rainforest endophytes and bioactive products. *Critical Reviews in Biotechnology*, 22(28): 315-333.

- Strobel, G.A. and Long, D.M. (1998). Endophytic microbes embody pharmaceutical potential. *ASM News*, 64: 263-268.
- Stone, J.K., Polishook, J.D., White, J.F. (2004). Endophytic fungi. In *Biodiversity of fungi. Inventory and monitoring methods*, ed. G.M. Muller, G.F. Bills, and M.S. Foster, pp. 241-270. San Diego, USA. Elsevier Academic Press.
- Sturz, A.V., Christie, B.R., Matheson, B.G., Arsenault, W.J. and Buchann, N.A. (1999). Endophytic bacterial communities in the periderm of potato tubers and their potential to improve resistance to soil borne plant pathogens. *Journal of Plant Pathology*, 48: 360-369.
- Sturz, A.V., Christie, B.R. and Nowak, J. (2000). Bacterial endophytes: potential role in developing sustainable systems of crop production. *Critical Reviews in Plant Sciences*, 12: 2404-2421.
- Sundram, S (2014). The effects of *Trichoderma* in surface mulches supplemented with conidial drenches in the disease development of *Ganoderma* basal stem rot in oil palm. *Journal of Oil Palm Research*, 25(3): 314-325.
- Sundram, S. (2013). First report: isolation of endophytic *Trichoderma* from oil palm (*Elaeis guineensis* Jacq.) and their *in vitro* antagonistic assessment on *Ganoderma boninense*. *Journal of Oil Palm Research*, 25(3): 368-372.
- Sundram, S. and Idris, A.S. (2009). *Trichoderma* as a biological control agents against *Ganoderma* in oil palm. *MPOB Information Series No. 463*. pp: 4.
- Sundram, S., Sariah, M., Idris, A.S. and Radziah, O. (2011). Symbiotic interaction of endophytic bacteria with arbuscular mycorrhizal fungi and its antagonistics effect on *Ganoderma boninense*. *The Journal Microbiology* 49: 551-557.
- Suprpta, D. N. (2012). Potential of microbial antagonists as biocontrol agents against plant fungal pathogens. *Journal International Society for Southeast Asian Agriculture Science*, 18: 1-8.
- Suresh Rao, K. S., Pradeep Rao, K. T., Dnyanob Rao, G. S., Agrawal, T. and Kotasthane, A. S. (2016). Root growth stimulation in rice (*Oryza sativa* L.) by seed biopriming with *Trichoderma* sp. *Applied Biological Research*, 18(1): 30-38.
- Susanto, A. (2009). Basal stem rot in Indonesia – Biology, economic importance, epidemiology, detection and control. *Proc. of the International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases*, ed. A. Kushairi, A.S. Idris, and K. Norman, pp. 58-89. Bangi, Selangor, Malaysia. MPOB

- Susanto, A., Sudharto, P.S. and Purba, R.Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159: 153-157.
- Tadych, M. and White, J. F. (2018). Endophytic microbes. In: *Encyclopedia of Microbiology*, ed, M. Schaechter, pp. 431-442. Academic Press, Oxford, UK.
- Ting, A.S.Y., Meon, S., Kadir, J., Radu, S. and Singh, G. (2008). Endophytic microorganisms as potential growth promoters of banana. *BioControl*, 53: 541-553.
- Thomas, S.E., Crozier, J., Aime, M.C., Evans, H.C. and Holmes, K.A. (2008). Molecular characterization of fungal endophytic morphospecies associated with the indigeneous forest tree, *Theobroma gileri*. *Ecuador. Mycological Research*, 112: 852-860.
- Thompson, A. (1931). *Stem-rot of the oil palm in Malaya*. Bulletin Department of Agriculture, Straits Settlements and F.M.S., *Science Series*, 6: 23.
- Tripathi, S., Das, A., Chandra, A. and Varma, A. (2015). Development of carrier-based formulation of root endophyte *Piriformospora indica* and its evaluation on *Phaseolus vulgaris* L. *World Journal of Microbiology and Biotechnology*, 31(2): 337-344.
- Turner, P.D. (1965). The incidence of *Ganoderma* disease of oil palm in Malaysia and its relation to previous crop. *Annals of Applied Biology*, 55: 417-423.
- Turner, P.D. (1981). *Oil Palm Diseases and Disorders*, pp. 88-110. New York Oxford University Press
- Turner, P.D. and Gillbanks, R. (1974). *Oil palm cultivation and management*, Kuala Lumpur: Incorporated Society of Planters. pp. 672.
- Utomo, C., Werner, S., Niepold, F. and Deising, H.B. (2005). Identification of *Ganoderma*, the causal agent of basal stem rot disease in oil palm using a molecular method. *Mycopathologia* 159: 159–170.
- Van Loon, L.C. (1997). Induced resistance in plants and the role of pathogenesis-related proteins. *European Journal of Plant Pathology* 103(9): 753-765.
- Van Loon, L.C. and Bakker, P.A.H.M. (2005). Induced systemic resistance as a mechanism of disease suppression by rhizobacteria. In *PGPR: Biocontrol and biofertilization*, pp. 39-66. Springer, Dordrecht.
- Walker, H.L. and Connick, W.J. (1983). Sodium alginate for production and formulation of mycoherbicides. *Weed Science*, 31(3): 333-338.

- Wang, K., Yan, P.S. and Cao, L.X. (2014). Chitinase from a novel strain of *Serratia marcescens* JPP1 for biocontrol of aflatoxin: Molecular characterization and production optimization using response surface methodology. *BioMed Research International*, 2014: 1-8.
- Yurnaliza, R., Aryantha, I. N. P., Esyanti, R. R. and Susanto, A. (2014). Antagonistic activity assessment of fungal endophytes from oil palm tissues against *Ganoderma boninense* pat. *Plant Pathology Journal*, 13(4): 257-267.
- Zabalbeascoa, P. (2008). The nature of the audiovisual text and its parameters. In *The didactics of Audiovisual Translation, Amsterdam*, ed. , J. Díaz-Cintas, pp. 21-38. John Benjamins.
- Zaiton, S., Sariah, M. and Zainal, A. (2006). Isolation and characterization of microbial endophytes from oil palm roots: implication as biological control agents against *Ganoderma*. *Planter*, 82(966): 587-597.
- Zakria, M., Njoloma, J., Saeki, Y. and Akao, S. (2007). Colonization and nitrogen-fixing ability of *Herbaspirillum* sp. strain B501 gfp1 and assessment of its growth-promoting ability in cultivated rice. *Microbes and Environments*, 22: 197-206.
- Zhang, J., Howell, C.R. and Wheeler, M.H. (1995). Frequency of isolation and the pathogenicity of *Fusarium* species associated with roots of healthy cotton seedlings. *Mycological Research*, 100: 747-752.

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