



UNIVERSITI PUTRA MALAYSIA

**EFFECTS OF *Bacillus subtilis* UPMB10 INOCULATION ON GROWTH
AND YIELD OF IRRIGATED RICE GROWN IN SEMARAK-KEMASIN
PADDY SOILS AND ITS CLAY MINERALOGY**

NUR FARHANA BINTI CHE HASSAN

FP 2019 40



**EFFECTS OF *Bacillus subtilis* UPMB10 INOCULATION ON GROWTH
AND YIELD OF IRRIGATED RICE GROWN IN SEMARAK-KEMASIN
PADDY SOILS AND ITS CLAY MINERALOGY**

By

NUR FARHANA BINTI CHE HASSAN

**Thesis Submitted to the School of Graduate Studies, Universiti
Putra Malaysia, in Fulfilment of the Requirements for the Degree of
Master of Science**

January 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

EFFECTS OF *Bacillus subtilis* UPMB10 INOCULATION ON GROWTH AND YIELD OF IRRIGATED RICE GROWN IN SEMARAK-KEMASIN PADDY SOILS AND ITS CLAY MINERALOGY

By

NUR FARHANA BINTI CHE HASSAN

January 2017

Chair : Professor Zulkifli Hj. Shamsuddin, PhD
Faculty : Agriculture

Rice is a main diet for half of the global human population and there is an urgent need to increase the rice yield to meet the ever-increasing demand, nationally and globally. Rice cultivation requires more potassium (K) fertilizer compared to nitrogen (N) and phosphorus (P). The challenges are due to the high cost of fertilizer-K and especially due to a majority of the soil K which presents in the insoluble form of mineral rocks and the concentrations of soil solution K are very low. Potassium solubilizing bacteria (KSB) are commonly utilized as plant growth promoters and capable to solubilize different forms of inorganic potassium and improve soil conditions, especially the microbiological and chemical properties, for increased crop production. A sequences of experiments were conducted under laboratory and field conditions with the subsequent objectives; i) to isolate and characterize the KSB from flooded rice, ii) to determine the effects of selected KSB inoculation with different fertilizer-K rates on growth and yield of rice, iii) to determine various forms of soil K concentration as affected by KSB inoculation. Twelve KSB strains were isolated from flooded rice. Four of these strains were selected for their high K solubilization activities namely RS5, RS11, RS13 and ES3, renamed UPMB22, 23, 24 and 25 respectively, along with a reference strain UPMB10, *Bacillus subtilis* originally isolated from oil palm (*Elaeis guineensis* Jacq.) roots. These five strains, especially UPMB10, were able to solubilize K *in-vitro* at the rates between 12.41 to 13.77 mg L⁻¹ in 5 days of incubation and showed beneficial PGPR characteristics (N₂ fixer and P solubilizer). Results from the XRD (X-Ray Diffraction) analysis showed that the soil sampled from Kemasin-Semerak Integrated Agricultural Development Project consisted of a small proportion of 2:1 clay minerals (mica) and behaved as a reservoir for soil-K, critically important to supply the short term plant-K requirements and conserve the soil-K in a long term productive crop ecosystem by reducing K loses. The clay fraction of the soil was dominated by kaolinite

while the other minerals which included quartz, mica, gibbsite and magnetite occurred in minor quantities. Results from the field study indicated that inoculation with *Bacillus subtilis* UPMB10 significantly increased the plant height, tiller numbers and leaf chlorophyll content (SPAD). This significant increase indicates the possible beneficial effects of UPMB10 inoculation on vegetative growth of rice under irrigated system. The maximum grain yield obtained were equivalent to 8.93 and 5.94 t ha⁻¹ with and without UPMB10 inoculation, respectively. Increased rates of K fertilization, complemented with KSB inoculation, increased grain yield production until the 80 kg ha⁻¹ K fertilization rate; equivalent to the grain yield production with the highest rate of uninoculated fertilizer-K (120 kg ha⁻¹). This implies that the usage of UPMB10 at 80 kg ha⁻¹ K fertilization could save 40 kg ha⁻¹ and yet achieve the maximum yield. The total K in soil was observed to significantly decrease after the bacterial inoculation process. The cation exchange capacity (CEC), exchangeable and water soluble K in soil did not show a significant change with UPMB10 application. CEC and water soluble K significantly increased with the application of fertilizer-K (without inoculation) until 80 kg ha⁻¹ while the exchangeable K is significantly decreased. It indicates that 80 kg K ha⁻¹ is sufficient for optimum rice uptake and K balance under irrigated lowland rice system. The study also showed that effective bacterial inoculation could successfully produce rice plants with improved growth and increased yield.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN PENGINOKULASIAN *Bacillus subtilis* UPMB10 TERHADAP
TUMBESARAN DAN HASIL PADI SAWAH YANG DITANAM DI TANAH PADI
SEMARAK-KEMASIN DAN MINERAL LEMPUNGNYA**

Oleh

NUR FARHANA BINTI CHE HASSAN

Januari 2017

Pengerusi : Profesor Zulkifli Hj. Shamsuddin, PhD
Fakulti : Pertanian

Padi adalah makanan ruji untuk separuh populasi dunia dan hasilnya perlu ditingkatkan untuk permintaan tempatan dan juga dunia yang semakin meningkat. Penanaman padi memerlukan lebih potasium (K) berbanding nitrogen (N) dan fosforus (P). Ini disebabkan oleh kos yang tinggi bagi baja K dan juga kerana majoriti, ataupun lebih daripada 90% K dalam tanah wujud dalam bentuk tidak terlarut yakni batuan mineral manakala kepekatan terlarutnya di dalam tanah lazimnya adalah sangat rendah. Bakteria pelarut potasium (KSB) biasanya digunakan sebagai penggalak tumbesaran tumbuhan dan dikenali mampu untuk melarutkan potasium yang tidak organik dalam pelbagai bentuk dan memperbaiki kondisi tanah, terutamanya secara mikrobiologi dan juga kandungan kimia dalam tanah untuk meningkatkan pengeluaran tanaman. Beberapa eksperimen telah dijalankan di makmal dan juga ladang dengan tujuan seperti berikut; i) untuk mengasingkan dan mengkarakterisasi KSB daripada padi sawah, ii) untuk menentukan keberkesanan KSB yang terpilih pada kadar baja K yang berbeza terhadap pertumbuhan dan hasil padi, iii) untuk menentukan kepekatan bentuk K dalam tanah yang dipengaruhi oleh penginokulasian KSB. Dua belas strain KSB diisolasi dari padi sawah. Empat strain terpilih untuk diuji kerana potensi keterlarutan K yang tinggi iaitu RS5, RS11, RS13 dan ES3, dinamakan semula sebagai UPMB22, 23, 24 dan 25, di samping strain rujukan UPMB10, *Bacillus subtilis* yang diisolasikan daripada akar kelapa sawit (*Elaeis guineensis* Jacq.). Kelima-lima strain tersebut terutamanya UPMB10 dapat melarutkan K dalam keadaan *in-vitro* dengan kadar antara 12.41 to 13.77 mg L⁻¹ dalam masa 5 hari inkubasi. Analisis XRD (X-Ray Diffraction) menunjukkan sampel tanah daripada Projek Pembangunan Integrasi Pertanian Kemasin-Semerak terdiri daripada 2:1 lempung mineral (mica) dan bertindak sebagai simpanan untuk K dalam tanah, terutamanya untuk keperluan tumbuhan dalam masa yang singkat. Pecahan lempung tersebut didominasi oleh kaolinite dan

mineral lain termasuklah quartz, mica, gibbsite and magnetite yang terdapat dalam kuantiti yang rendah. Keputusan daripada kajian ladang menunjukkan inokulasi dengan *Bacillus subtilis* UPMB10 secara signifikan meningkatkan tinggi tanaman, bilangan rumpun dan kandungan klorofil daun (SPAD). Peningkatan yang signifikan ini menunjukkan keberkesanan inokulasi UPMB10 pada pertumbuhan padi secara sistem pengairan. Hasil bijirin maksimum yang diperolehi adalah bersamaan dengan 8.93 and 5.94 t ha⁻¹ dengan dan juga tanpa UPMB10 inokulasi. Pembajaan K yang tinggi, dengan penginokulasian KSB, meningkatkan lagi pengeluaran hasil bijirin sehingga pada kadar 80 kg ha⁻¹ pembajaan K; bersamaan dengan kadar yang paling tinggi iaitu (120 kg ha⁻¹) tanpa penginokulasian KSB. Ini menunjukkan aplikasi UPMB10 pada 80 kg ha⁻¹ pembajaan K mampu menjimatkan 40 kg ha⁻¹ baja K dan dapat mencapai hasil yang maksimum. Jumlah kandungan keseluruhan K dalam tanah berkurang secara signifikan selepas diinokulasi dengan bakteria. Kadar pertukaran kation (CEC), pemboleh-tukaran K dan K terlarut dalam tanah tidak menunjukkan perubahan dengan inokulasi UPMB10. Terdapat peningkatan yang signifikan pada CEC dan K terlarut dengan pembajaan K sehingga 80 kg ha⁻¹ (tanpa inokulasi) manakala keboleh-tukaran K menurun secara signifikan. Ini membuktikan yang 80 kg K ha⁻¹ adalah cukup untuk pertumbuhan padi yang optimum dan keseimbangan K di kawasan padi sawah yang dijaga sistem pengairannya. Kajian juga membuktikan bahawa dengan penginokulasian bakteria yang efektif mampu menghasilkan tanaman padi dengan pertumbuhan serta hasil yang lebih tinggi.

ACKNOWLEDGEMENTS

Praise to Allah The Almighty. This dissertation would not be able to be completed without His Mercy and also the guidance, support and help of several individuals, who in one way or another, have contributed or extended their valuable assistance in the preparation and completion of this research.

First and foremost, my utmost gratitude to my supervisor Prof. Dr. Zulkifli Hj. Shamsuddin for his patience and steadfast encouragement as I hurdled through all the obstacles to complete the research. I would have been lost without him. Special thanks to Prof. Dr. Radziah Othman and Dr. Roslan Bin Ismail, their perpetual energy and enthusiasm have motivated many of their advisees including me.

All my lab mates and friends who have made both working in the lab and thesis writing a more enjoyable experience. I warmly thank Suheda, Adilah, Dr. Ali Tan, Kuan, Hun and all other postgraduates for their friendships and companions.

I owe my loving thanks to my husband for his continual support technically and emotionally throughout the entire study. Special gratitude goes to my family and extended family members for their loving support. Thank you all for giving me the strength to plod through good and bad times. The financial support through Graduate Research Fellowship (GRF) by Universiti Putra Malaysia and Long Research Grant Scheme (LRGS) by Ministry of Education are also gratefully acknowledged.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Zulkifli Haji Shamsuddin, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Roslan Bin Ismail, PhD

Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Radziah Binti Othman, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 11 April 2019

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: _____

Signature: _____
Name of Member of
Supervisory
Committee: _____

Signature: _____
Name of Member of
Supervisory
Committee: _____

TABLE OF CONTENT

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xv
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	3
2.1 Rice	3
2.1.1 Growth and Development of Rice	3
2.1.2 Nutritional Requirement	4
2.2 Potassium Fertilizers	6
2.3 Forms of Soils Potassium	7
2.3.1 Soil Solution	7
2.3.2 Exchangeable	7
2.3.3 Non-exchangeable	7
2.3.4 Mineral	8
2.4 Potassium Solubilizing Bacteria (KSB)	8
2.4.1 Mechanism of Solubilization by Potassium Solubilizing Bacteria	9
2.4.2 KSB Effects on Plant Growth and Yield	10
2.5 Clay Mineralogy	12
2.5.1 K-bearing Minerals	12
2.5.2 X-Ray Differentiation	13
3 ISOLATION AND CHARACTERIZATION OF RHIZOSPHERIC AND ENDOPHYTIC BACTERIA FROM RICE ROOTS	14
3.1 Introduction	14
3.2 Materials and Methods	15
3.2.1 Potassium Solubilizing Bacteria Isolation	15
3.2.2 Characterization of Isolates for Potassium	16
3.2.3 Additional Beneficial Characteristics	16
3.3 Results	17
3.3.1 Isolation of Plant Growth-Promoting Rhizobacteria (PGPR) from Rhizospheric and Endophytic Root or Rice	17
3.3.2 Potassium Solubilization Ability by the Isolates	17
3.3.3 Quantitative Test of the isolates for K	19
3.3.4 Nitrogen Fixation and Phosphate	20

	Solubilization Ability Tests	
3.4	Discussion	21
3.5	Conclusion	22
4	CLAY MINERALOGY PROPERTIES OF KEMASIN-SEMERAK RICE SOIL	23
4.1	Introduction	23
4.2	Materials and Methods	24
4.2.1	Soil Sampling	24
4.2.2	Soil Texture Analysis by Pipette Method	24
4.2.3	Soil Chemical Properties	25
4.2.4	Soil Mineralogical Properties – X-Ray Diffraction	26
4.3	Results	27
4.3.1	Soil Texture Analysis	27
4.3.2	Chemical Properties of Soil Sample from Kemasin-Semerak Granary Area	27
4.3.3	Soil Mineralogical Analysis	27
4.4	Discussion	28
4.5	Conclusion	30
5	APPLICATION OF <i>Bacillus subtilis</i> UPMB10 ON SOIL POTASSIUM DISSOLUTION FOR RICE GROWTH AND YIELD UNDER FLOODED CONDITION	31
5.1	Introduction	31
5.2	Materials and Methods	33
5.2.1	Experimental Design	33
5.2.2	Site Characteristics of Kemasin-Semerak	33
5.2.3	Plots Preparation	33
5.2.4	Planting of Rice Seedlings in Plots	33
5.2.5	Application of Chemical Fertilizer	34
5.2.6	Preparation of Bacterial Inoculum	34
5.2.7	Rice Growth and Yield Parameters Measurement	34
5.2.8	Plant Tissue Analysis - Nutrient concentration in leaf and grain of rice	34
5.2.9	Soil Chemical Properties Analysis	35
5.2.10	Statistical Analysis	36
5.3	Results	36
5.3.1	Rice Growth Parameters	36
5.3.2	Rice Yield Parameters	37
5.3.3	Leaf Nutrient Concentration	39
5.3.4	Grain Nutrient Concentration	40
5.3.5	Soil Potassium Chemical Properties	41
5.4	Discussion	42
5.5	Conclusion	45
6	GENERAL DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	46

BIBLIOGRAPHY
APPENDICES
BIODATA OF STUDENT
LIST OF PUBLICATIONS

49
55
63
64



LIST OF TABLES

Table		Page
3.1	Qualitative test of the isolates for K solubilization	18
3.2	Diameter of halo zone formation by KSB strains after 24 h	19
3.3	Ability of 30 bacterial isolates to fix N and solubilize P	21
4.1	Percentage of sand, silt and clay in a soil sample from the Kemasin-Semerak granary area	27
4.2	Chemical properties of soil sample from Kemasin-Semerak granary area	27
4.3	Relative abundance of existed minerals in Kemasin-Semerak soil	28
5.1	The treatments imposed for field experiment	33
5.2	Plant height on 82 and 111 DAT	37
5.3	Tiller numbers on 55, 82 and 111 DAT	37
5.4	Leaf chlorophyll content (SPAD value) on 55 and 82 DAT	37
5.5	Yield components	38
5.6	Effect of PGPR inoculation and fertilizer-K levels on leaf nutrients concentration at harvest	39
5.7	Effect of PGPR inoculation and fertilizer-K levels on rice grain nutrients concentration	40
5.8	Effect of PGPR inoculation and fertilizer-K levels on soil potassium chemical properties and cation exchange capacity	42

LIST OF FIGURES

Figure		Page
2.1	Rice production in Malaysia and its average yield in 2008 – 2012	3
3.1	Halo zone formation of K solubilization ability on Aleksandrov agar medium by KSB isolate	18
3.2	Potassium solubilisation rate at 5 DAI	19
3.3	Positive results of nitrogen fixing ability on N-free agar and phosphate solubilization ability on Pikovskaya agar after 24 hours incubation	20
4.1	XRD pattern of soil sample from Kemasin-Semerak granary area	28
5.1	Estimated grain yield to added K-fertilizer with and without inoculation	38

LIST OF ABBREVIATIONS

N	nitrogen
P	phosphorus
K	potassium
pgpr	plant growth-promoting rhizobacteria
NFB	nitrogen fixing bacteria
PSB	phosphorus fixing bacteria
KSB	potassium solubilizing bacteria
KSM	potassium solubilizing microorganism
mg kg ⁻¹	milligram per kilogram
mg L ⁻¹	milligram per litre
°C	degree Celcius
%	percentage
m	meter
kg	kilogram
g	gram
mg	milligram
t	tonne
ha	hectare
pH	potential different of hydrogen ion
CEC	cation exchange capacity
mL	millilitre
XRD	X-ray Diffraction
AAS	atomic absorption spectrophotometer
AA	autoanalyzer
m ²	meter square
cmolc kg ⁻¹	centimol charge per kilogram
s	second
min	minute
h	hour
TSA	tryptic soy agar
sdH ₂ O	sterile distilled water
rpm	revolution per minute
TC	total carbon
TN	total nitrogen
vol	volume
wt	weight
DAI	day after incubation
DAS	day after sowing
DAT	day after transplanting
cfu	colony forming unit
HCl	hydrochloric acid
HNO ₃	nitric acid
H ₂ SO ₄	sulfuric acid
H ₂ O ₂	hydrogen peroxide
SAS	statistical analysis system
LSD	least significant different
ANOVA	analysis of variance
DOA	Department of Agriculture

NH_4OAC	ammonium acetate
K_2SO_4	potassium sulfate
$\text{Ca}(\text{H}_2\text{PO}_4)_2$	calcium hydrogen phosphate
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	magnesium sulfate heptahydrate
CaCO_3	calcium carbonate
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$	iron chloride hexahydrate
KOH	potassium hydroxide
K_2HPO_4	potassium hydrogen phosphate
NaCl	sodium chloride
CaCl_2	calcium chloride
Fe-EDTA	ferric ethylenediaminetetraacetic acid
H_3BO_3	boric acid
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	manganese sulfate hydrate
NaMoO_4	sodium molybdate
$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	zinc sulfate heptahydrate
$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$	copper sulfate pentahydrate
$\text{Ca}_3(\text{PO}_4)_2$	calcium phosphate
$(\text{NH}_4)_2\text{SO}_4$	ammonium sulfate
KCl	potassium chloride
$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	ferrous sulfate heptahydrate

CHAPTER 1

INTRODUCTION

According to United Nations, the world population is expected to reach 8.9 billion by 2050, with the developing countries of Asia and Africa to absorb the vast majority of the growth (Wood, 2001). The demands placed upon agriculture to supply future food will be one of the greatest challenges due to the rise of world populations. To meet this challenge, greater efforts with focus on the soil-plant-microbe interactions and the agro-ecosystem are needed to enable better perception of the intricate processes which govern the stability of agricultural lands. Soil is a dynamic natural body on the earth's crust. There are several minerals containing essential elements in the soil, most importantly are nitrogen (N), phosphorus (P), and potassium (K). K is the third essential element and it plays a part in the growth, metabolism, and development of plant. Plants with inadequate supply of potassium, will produce unhealthy roots, stunted growth and poor quality seeds (White and Karley, 2010; McAfee 2008).

Half of the world populations consume rice as their staple diet. Its growth is limited by various environmental conditions such as soil and climatic conditions. In Malaysia, there is an urgent need to increase the rice yield to meet the national food requirement. The major challenges for rice cropping are limited arable land, unpredictable water supply and climatic conditions as well as pest and disease problems. Its production requires more fertilizer-K input compared to nitrogen and phosphorus. This is because K is a mobile nutrient and easily leached when not absorbed by plant roots. In rice, K is important especially for the formation of tillers and grains. The K nutrient absorbed by roots can increase the lignification in culms and subsequently enhance the number of tillers and stem strength. The increase in numbers of strong tillers will consequently produce higher yields with improved quality.

Plants uptake K from the soil and its availability is influenced by K dynamics and also the soil K content (Parmar and Sindhu, 2013). Soil K can be classified into four available forms based on the degree of availability to crops, i.e. soil solution, exchangeable, non-exchangeable and mineral K (Darunsontaya et al., 2012). The soil solution K is the crucial source for plant root uptake. This readily available K is usually relatively small and does not provide a good indication of the long term ability of a soil to supply K to plants. Exchangeable-K which is held by the negative charges on soil clay minerals and organic matter exchange sites is in related with water soluble K and collectively known as the available K pool (Meena et al., 2016). Soil-fixed (non-exchangeable) K is an important contribution to plant K supply (Mengel et al., 1993) and is held as fixed ion in the lattice structure of clay minerals.

The use of microorganisms in agriculture is a vital practice to enhance plant nutrient availability. The application of plant growth-promoting rhizobacteria

(PGPR) has grown immensely in the recent years. It is well established for its beneficial effect towards plant growth and development. These bacteria become beneficial when they can fix or solubilize the three essential plant nutrient elements (N, P, K) in soil. This is because some of the elements are in unavailable or bound forms for plant uptake such as being adsorbed by rocks and soil minerals. Nitrogen Fixing Bacteria (NFB), for example, are able to fix the abundantly available atmospheric-N. Some other bacteria can solubilize insoluble P and K in soil and are known as Phosphate Solubilizing Bacteria (PSB) and Potassium Solubilizing Bacteria (KSB), respectively (Tan, 2015). Bacteria will become more beneficial to plant growth when they are also able to produce phytohormones which are undoubtedly vital for its growth and development especially the root system. These bacteria can be grouped as plant growth-promoting rhizobacteria (PGPR) due to these beneficial characteristics.

Plant growth-promoting rhizobacteria also enact a significance function in K mineral cycle and, thus, they can become a green alternative technology to solubilize K minerals for plant uptake. These bacteria which possess potassium solubilizing abilities are termed KSB and they can solubilize the insoluble K into available forms in soil as soil solution K and render them as available K to the plants (Zeng et al., 2012). The major mechanism of mineral K solubilization is due to organic acids synthesized by rhizospheric microorganisms. Organic acids production leads to acidification of the microbial cell and its surrounding environment which promote the solubilization of mineral K.

An imbalanced or surplus application of chemical fertilizers can produce negative environmental impacts and increase the cost of crop production. So, there is an urgent need to apply cost effective and eco-friendly alternatives to effectively improve crop production. Therefore, the application of KSB is viewed a reliable approach in enhancing the agricultural land productivity. This new practice is also asserted to be able to ameliorate the degraded, marginally productive and unproductive agricultural soils (Basak and Biswas, 2012). Currently, minimal information is available on soil potassium dissolution by rhizobacteria and their effects on growth and yield of rice. Hence, the objectives of this study are:

1. To isolate and characterize the KSB from flooded rice.
2. To determine the effects of selected KSB inoculation with different fertilizer-K rates on growth and yield of rice.
3. To determine various forms of soil K concentration as affected by KSB inoculation. There may be a preamble at the beginning of a chapter. The purpose may be to introduce the themes of the main headings.

BIBLIOGRAPHY

- Abdel-Salam, M.A. and Shams, A.S. 2012. Feldspar-K fertilization of potato (*Solanum tuberosum* L.) augmented by biofertilizer. *Journal Agriculture Environmental Science* 12(6):694-699.
- Altamare, C., Norvell, W. A., Bjorkman, T. and Harman, G. E. 1999. Solubilization of phosphates and micronutrients by the plant growth promoting and bacterial fungus *Trichodera harzianum* Rifai, *Applied and Environmental Microbiology* 65: 2926-2933.
- Alves, L., Oliveira, V. L. and Filho, G. N. S. 2010. Utilization of rocks and ectomycorrhizal fungi to promote growth of eucalypt. *Brazilian Journal of Microbiology* 41: 676-684.
- Archana, D. S., Nandish, M. S., Savalagi, V. P. and Alagawadi, A. R. 2012. Screening of potassium solubilizing bacteria (KSB) for plant growth promotional activity. *Bioinfolet* 9(4):627-630.
- Archana, D. S., Nandish, M. S., Savalagi, V. P. and Alagawadi, A. R. 2013. Characterization of potassium solubilizing bacteria (KSB) from rhizosphere soil. *Bioinfolet* 10: 248-257.
- Archana, D. S., Savalgi, V. P. and Alagawadi, A. R. 2008. Effect of potassium solubilizing bacteria on growth and yield of maize. *Soil Biology and Ecology* 28: 9-18.
- Badr, M. A. 2006. Efficiency of K-feldspar combined with organic materials and silicate dissolving bacteria on tomato yield. *Journal of Applied Science Research* 2:1191-1198.
- Badr, M. A., Shafei, A. M. and Sharaf El-Deen, S. H. 2006. The dissolution of K and P-bearing minerals by silicate dissolving bacteria and their effect on sorghum growth. *Research Journal of Agriculture and Biological Sciences* 2(1): 5-11.
- Bagyalakshmi, B., Ponmurugan, P. and Balamurugan, A. 2012. Impact of different temperature, carbon and nitrogen sources on solubilization efficiency of native potassium solubilizing bacteria from tea (*Camellia sinensis*). *Journal of Biological Research* 3(2): 36-42.
- Barre, P., Velde, B., Catel, N. and Abbadie, L. 2007. Soil-plant potassium transfer: impact of plant activity on clay minerals as seen from X-ray diffraction. *Plant and Soil* 292: 137-146.
- Barre, P., Montagnier, C., Chenu, C., Abbadie, L. and Velde, B. 2008. Clay minerals as a soil potassium reservoir: observation and quantification through X-ray diffraction. *Plant and Soil* 302:213-220.
- Basak, B. B and Biswas, D. R. 2009. Influence of potassium solubilizing microorganism (*Bacillus mucilaginosus*) and waste mica on potassium uptake dynamics by sudan grass (*Sorghum vulgare* Pers.) grown under two Alfisols. *Plant and Soil* 317: 235-55.
- Basak, B. B. and Biswas, D. R. 2012. *Modification of waste mica for alternative source of potassium: evaluation of potassium release in soil from waste mica treated with potassium solubilizing bacteria (KSB)*. Lambert Academic Publishing, Germany.
- Baset Mia, M. A., Shamsuddin, Z. H., Wahab, Z. and Marziah, M. 2010. Rhizobacteria as bioenhancer and biofertilizer for growth and yield of banana (*Musa* spp. cv. 'Berangan'). *Scientia Horticulturae* 126: 80-87.

- Bennett, P. C., Choi, W. J. and Rogers, J. R. 1998. Microbial destruction of feldspars. *Mineralogical Magazine* 8(62A):149-150.
- Benton Jones, Jr. 2001. *Laboratory guide for conducting soil tests and plant analysis*. CRC Press LLC, New York.
- Bin, L., Bin, W., Mu, P., Liu, C., Teng, H. H. 2010. Microbial release of potassium from K-bearing minerals by thermophilic fungus *Aspergillus fumigatus*. *Geochimica et Cosmochimica Acta* 72:87-98.
- Darunsontaya, T., Suddhaprikarn, A., Kheoruenromne, I., Prakongkep and Gilkes, R. J. 2012. The forms and availability to plants of soil potassium as related to mineralogy for upland Oxisols and Ultisols from Thailand. *Geoderma* 170: 11-24.
- Davison, J. 1988. Plant beneficial bacteria. *Nature Biotechnology* 6: 282-286.
- Department of Agriculture. 2008. *Panduan mengenali siri-siri tanah utama di Semenanjung Malaysia*. DOA, Malaysia.
- Department of Agriculture. 2014. *Paddy statistics of Malaysia 2013*. DOA, Malaysia.
- Dobereiner J. and Day J. M. 1975. Associative symbiosis in tropical grasses: characterization of microorganisms and dinitrogen fixing sites. pp.518-538. *Proceedings of the 1st International Symposium on Nitrogen Fixation*. Washington State University Press.
- Doberman, A. and Fairhurst, T. 2000. *Rice: nutrient disorders and nutrient management*. Potash and Phosphate Institute, Potash and Phosphate Institute of Canada and International Rice Research Institute.
- Eckert, B., Weber, O. B., Kirchof, G., Halbritter, A., Stoffels, M. and Hartman, A. 2001. *Azospirillum doebereineriae* sp. nov., a nitrogen fixing bacterium associated with the C₄-grass *Miscanthus*. *International Journal of Systematic and Evolutionary Microbiology* 51: 17-26
- Food and Agriculture Organization (FAO) of the United Nations. 2011. Current world fertilizer trends and outlook to 2015. Rome: FAO
- Food and Agriculture Organization (FAO) of the United Nations. 2009. High-level expert forum. Global agriculture towards 2050.
- Gee, G. W. and Bauder, J. W. 1986. Particle-size Analysis. p. 383–411. *In* Methods of soil analysis. Klute, A. (ed). Soil Science Society of America Book Series 5, Madison, WI.
- Glick, B. R. 1995. The enhancement of plant growth by free-living bacteria. *Canadian Journal of Microbiology* 41: 109-117.
- Goldstein, A. H. 1994. Involvement of the quinoprotein glucose dehydrogenase in the solubilization of exogenous mineral phosphates by Gram-negative bacteria. *In Phosphate in microorganisms: cellular and molecular biology*. Torriani-Gorni, A., Yagil, E. and Silver, S. (eds.). ASM Press, Washington.
- Gundala, P. B., Chinthala, P. and Sreenivasulu, B. 2013. A new facultative alkaliphilic, potassium solubilizing, *Bacillus* Sp. SVUNM9 isolated from mica cores of Nellore District, Andhra Pradesh, India. *Research and Reviews: Journal of Microbiology and Biotechnology* 2(1):1–7.
- Han, H. S., Supanjani, and Lee, K.. D. 2006. Effect of co-inoculation with phosphate and potassium solubilizing bacteria on mineral uptake and growth of pepper and cucumber. *Plant Soil and Environment* 52: 130-136.
- Hinsinger, P. 2002. Potassium. *In* Encyclopedia of Soil Science. Lal, R. (ed). Marcel Dekker, Inc., New-York, USA.

- Hinsinger, P., Jaillard, B. and Dufey, E. D. 1992. Rapid weathering of a trioctahedral Mica by the roots of Ryegrass. *Soil Science Society of America Journal* 56:977-982.
- Hinsinger, P., Elsass, F., Jaillard, B. and Robert, M. 1993. Root-induced irreversible transformation of trioctahedral mica in the rhizosphere of rape. *Journal of Soil Science* 44:535-545.
- Hu, X., Chen, J. and Guo, J. 2006. Two phosphate- and potassium-solubilizing bacteria isolated from Tianmu Mountain, Zhejiang, China. *World Journal of Microbiology and Biotechnology* 22:983-990.
- Huang, P. M. 2005. Chemistry of potassium in soils. In *Chemical Processes in Soils*. Tabatabai, M. A. and Sparks, D. L. (eds.). Soil Science Society of America, United States of America.
- International Potash Institute. 1977. Potassium dynamic in the soil. IPI, Switzerland.
- International Rice Research Institute. 2014. Retrieved March 10 2015, from http://www.knowledgebank.irri.org/ericeproduction/0.2._Growth_stages_of_the_rice_plant.htm
- International Rice Research Institute. 2015. Retrieved June 1st 2015, from <http://irri.org/our-impact/increase-food-security>
- Liu, D., Lian, B. and Dong, H. 2012. Isolation of *Paenibacillus* sp. and assessment of its potential for enhancing mineral weathering. *Geomicrobiology Journal* 29(5): 413-421.
- Martin, H. W. and Sparks, D. L. 1985. On the behavior of non-exchangeable potassium in soils. *Communications in Soil Science and Plant Analysis* 16: 133–162.
- McAfee, J. 2008. *Potassium: a key nutrient for plant growth*. Department of Soil and Crop Sciences.
- Meena, V. S., Maurya, B. R. and Verma, J. P. 2014. Does a rhizospheric microorganism enhance K⁺ availability in agricultural soils? *Microbiological Research* 169: 337-347.
- Meena, V. S., Maurya, B. R., Verma, J. P. and Meena, R. S. 2016. *Potassium solubilizing microorganisms for sustainable agriculture*. Springer.
- Mengel, K. 1985. Dynamics and availability of major nutrients in soils. *Advances in Soil Science* 2: 65-131.
- Mengel, K. and Busch, R. 1982. The importance of the potassium buffer power on the critical potassium level in soils. *Soil Science* 133: 27-32.
- Mengel, K., Rahmatullah and Dou, H. 1998. Release of potassium from the silt and sand fractions of loess-derived soils. *Soil Science* 163: 805-813.
- Muralikannan, N. and Anthomiraj, S. 1998. Occurrence of silicate solubilizing bacteria in rice ecosystem. *Madras Agricultural Journal* 85:47-50.
- Niranjan Raj, S. Shetty, H. S. and Reddy, M. S. 2005. Plant growth-promoting rhizobacteria: potential green alternative for plant productivity. In *PGPR: biocontrol and biofertilization*. Siddiqui, Z. A. (ed). Springer, Dordrecht, The Netherlands.
- Officer, S. J., Tillman, R. W., Palmer, A. S. and Whitton, J. S. 2006. Variability of clay mineralogy in two New Zealand steep-land topsoils under pasture. *Geoderma* 132: 427-440.
- Ogaard, A. F. and Krogstad, T. 2005. Release of interlayer potassium in Norwegian grassland soils. *Journal of Plant Nutrition and Soil Science* 168: 80-88.

- Okon, Y. and Labendera-Gonzales, C. A. 1994. Agronomic applications of *Azospirillum*: an evaluation of 20 years worldwide field inoculation. *Soil Biology and Biochemistry* 26: 1591–1601.
- Paramanathan, S. and Noordin, W. D. 1986. Classification of acid sulfate soils of Peninsular Malaysia. *Pertanika* 9: 323-330.
- Park, M., Singvilay, O., Seok, Y., Chung, J., Ahn, K. and Sa, T. 2003. Effect of phosphate solubilizing fungi on P uptake and growth of tobacco in rock phosphate applied soil. *Korean Journal of Soil Science and Fertilizer* 36: 233-238.
- Parmar, P. and Sindhu, S. S. 2013. Potassium solubilization by rhizosphere bacteria: influence of nutritional and environmental conditions. *Journal of Microbiology Research* 3(1): 25-31.
- Pikovskaya, R. I. 1948. Mobilization of phosphorus in soil in connection with vital activity by microbial species. *Microbiologica* 17:362-370.
- Podile, A. R. and Kishore, K. 2007. Plant growth-promoting rhizobacteria. In *Plant associated bacteria*. Gnanamanickam, S. S. (ed). Springer, The Netherlands.
- Prajapati, K. B. and Modi, H. A. 2012. Isolation and characterization of potassium solubilizing bacteria from ceramic industry soil. *CIBTech Journal of Microbiology* 1:8-14.
- Quemener, J. 1986. Important factors in potassium balance sheet. pp. 33-63. *Proceedings of the 13th Congress on Nutrient Balance and Need for Potassium*. International Potash Institute, Bern, Switzerland.
- Ramamoorthy, V., Viswanathan, R., Raguchander, T., Prakasan, V. and Samiyappan, R. 2001. Induction of systemic resistance by plant growth promoting rhizobacteria in crop plants against pests and diseases. *Crop Protection* 20: 1-11.
- Rich, C. I. 1964. Effect of cation size and pH on potassium exchange in Nason soil. *Soil Science* 98: 100-106.
- Rich, C. I. 1972. Potassium in soil minerals. *Proceedings 9th Colloquium of the International Potash Institute (IPI)*. Basel, Switzerland.
- Romheld, V. and Kirkby, E. A. 2010. Research on potassium in agriculture: needs and prospects. *Plant and Soil* 335:155-180.
- Sangeeth, K. P., Bhai, R. S. and Srinivasan, V. 2012. *Paenibacillus glucanolyticus*, a promising potassium solubilizing bacterium isolated from black pepper (*Piper nigrum* L.) rhizosphere. *Journal of Spice and Aromatic Crops* 21(2):118-124.
- Shaaban, E. A., El-Shamma, I. M. S., El Shazly, S., El-Gazzar, A., Abdel-Hak, R. E. 2012. Efficiency of rock-feldspar combined with silicate dissolving bacteria on yield and fruit quality of valencia orange fruits in reclaimed soils. *Journal of Applied Science Research* 8: 4504-4510.
- Shamshuddin, J. Paramanathan, S. and Nik Mokhtar. 1986. Mineralogy and surface charge properties of two acid sulfate soils from Peninsular Malaysia. *Pertanika* 9(2): 167-176.
- Shaviv, A., Mohsin, M., Pratt, P. F. and Mattigod, S. V. 1985. Potassium fixation characteristics of five Southern California soils. *Soil Science Society of America Journal* 49: 1105-1109.
- Sheng, X. F. 2005. Growth promotion and increased potassium uptake of cotton and rape by a potassium releasing strain of *Bacillus edaphicus*. *Soil Biology and Biochemistry* 37:1918-1922.

- Sheng, X. F. and He, L. Y. 2006. Solubilization of potassium-bearing minerals by a wild type strain of *Bacillus edaphicus* and its mutants and increased potassium uptake by wheat. *Canadian Journal of Microbiology* 52:66-72.
- Sheng, X. F. and Huang, W. Y. 2002. Mechanism of potassium release from feldspar affected by the strain NBT of silicate bacterium. *Acta Pedologica Sinica* 39: 863-871.
- Sheng, X. F., Zhao, F., He, H., Qiu, G. and Chen, L. 2008. Isolation, characterization of silicate mineral solubilizing *Bacillus globisporus* Q12 from the surface of weathered feldspar. *Canadian Journal Microbiology* 54:1064-1068.
- Sindhu, S. S., Verma, M.K. and Suman, M. 2009. Molecular genetics of phosphate solubilization in rhizosphere bacteria and its role in plant growth promotion. *In Phosphate solubilizing microbes and crop productivity*. Khan, M.S. and Zaidi, A. (eds). Nova Science Publishers, United States of America.
- Sindhu, S. S, Parmar, P. and Phour, M. 2012. Nutrient cycling: potassium solubilization by microorganisms and improvement of crop growth. *In Geomicrobiology and biogeochemistry: soil biology*. Parmar, N. and Singh, A. (eds.). Springer-Wien, New York, Germany.
- Singh, G., Biswas, D. R. and Marwah, T. S. 2010. Mobilization of potassium from waste mica by plant growth promoting rhizobacteria and its assimilation by maize (*Zea mays*) and wheat (*Triticum aestivum* L.). *Journal of Plant Nutrition* 33: 1236-1251.
- Soil Survey Division Staff. 1993. *Soil survey manual*. Soil Conservation Service. U.S. Department of Agriculture Handbook 18.
- Sparks, D. L. 1987. Potassium dynamics in soils. *Advances in Soil Science* 6: 1-63
- Sparks, D. L. 2000. Bioavailability of soil potassium. *In Handbook of Soil Science*. M. E. Sumner (ed). CRC Press, Boca Raton, FL.
- Sparks, D. L. and Huang, P. M. 1985. Physical chemistry of soil potassium. P. 201-276. *In Potassium in agriculture*. R. D. Munson (ed). American Society of Agronomy, Madison, WI.
- Sperberg, J. I. 1958. The incidence of apatite solubilizing organisms in the rhizosphere and soil. *Australian Journal of Agricultural and Resource Economics* 9:778-781.
- Sparks, D. L. 1987. Potassium dynamics in soils. *Advances in Soil Sciences* 6:1-63.
- Sugumaran, P. and Janarthanam, B., 2007, Solubilization of potassium containing minerals by bacteria and their effect on plant growth. *World Journal of Agricultural Sciences* 3(3): 350-355.
- Supanjani, Han, H. S., Jung, S. J. and Lee, K. D. 2006. Rock phosphate potassium and rock solubilizing bacteria as alternative sustainable fertilizers. *Agronomy for Sustainable Development* 26:233-240.
- Tan, K. Z., Radziah O., Halimi M. S., Khairuddin A. R., Habib S. H. and Shamsuddin Z. H. 2014. Isolation and characterization of rhizobia and plant growth-promoting rhizobacteria and their effects on growth of rice seedlings. *American Journal of Agricultural and Biological Sciences* 9(3): 342-360.
- Tan, K. Z. 2015. Application of rhizobia and plant growth promoting rhizobacteria for increased growth, N₂ fixation and yield of rice. PhD Thesis, Universiti Putra Malaysia.

- Tsadilas, C., Samaras, V. and Stamatiadis, S. 2012. Irrigation and nitrogen fertilization effects on soil chemical properties and cotton yield. *Communications in Soil Science and Plant Analysis* 43: 190-196.
- Uroz, S., Calvaruso, C., Turpault, M-P. and Frey-Klett, P. 2009. Mineral weathering by bacteria: ecology, actors and mechanisms. *Trends in Microbiology* 17:378-387.
- Verma, S. C., Ladha, J. K. and Tripathi, A. K. 2001. Evaluation of plant growth promoting and colonization ability of endophytic diazotrophs from deep water rice. *Journal of Biotechnology* 91: 127-141.
- Vessey, J. K. 2003. Plant growth promoting rhizobacteria as biofertilizers. *Plant and Soil* 255(2): 571-586.
- Wang, J. G., Zhang, F. S., Cao, Y. P. and Zhang, X. L. 2000. Effect of plant types on release of potassium from gneiss. *Nutrient Cycling in Agroecosystems* 56(1): 37-43.
- White, P. J. and Karley, A. J. 2010. Potassium. In Cell biology of metals and nutrients, plant cell monographs. Hell, R. and Mendel, R. R. (eds). Springer, Berlin.
- Wild, A. 1988. Potassium, sodium, calcium, magnesium, sulphur, silicon. In Russell's Soil Conditions and Plant Growth. Alan Wild (ed). Longman Scientific & Technical.
- Wood N. 2001. Nodulation by numbers: the role of ethylene in symbiotic nitrogen fixation. *Trends in Plant Science* 6:501-502.
- Wu, S. C., Cao, Z. H., Li, Z. G., Cheung, K. C. and Wong, M. H. 2005. Effects of biofertilizer containing N- fixer, P and K solubilizers and AM fungi on maize growth: a greenhouse trial. *Geoderma* 125:155-166.
- Xie, J. C. 1998. Present situation and prospects for the world's fertilizer use. *Plant Nutrition and Fertilizer Science* 4:321-330.
- Yoshida, S. 1981. *Fundamentals of rice crop science*. International Rice Research Institute, Philippines.
- Zainudin, P. M. D. H., Elixon, S., Azlan, S., Alias, I., Saad, A., Othman, O., Habibuddin, H., Asfaliza, R., Mohd Najib, M. Y., Siti Norsuha, M. and Amiruddin, M. 2012. MR263 varieti padi baru untuk kawasan sederhana subur. *Buletin Teknologi MARDI* Bil. (1).
- Zakaria, A. A. B. 2009. Growth optimization of potassium solubilizing bacteria isolated from biofertilizer. Thesis, Universiti Malaysia Pahang.
- Zeng, X., Liu, X., Tang, J., Hu, S., Jiang, P. and Li, W. 2012. Characterization and potassium- solubilizing ability of *Bacillus circulans* Z1-3. *Advanced Science Letters* 10: 173-176.
- Zhang, A., Zhao, G., Gao, T., Wang, W., Li, J. and Zhang, S. 2013. Solubilization of insoluble potassium and phosphate by *Paenibacillus kribensis* CX-7: a soil microorganism with biological control potential. *African Journal of Microbiology Research* 7(1):41-47.
- Zhao, F., Sheng, X., Huang, Z. and He, L. 2008. Isolation of mineral potassium-solubilizing bacterial strains from agricultural soils in Shandong Province. *Biodiversity Science* 16: 593-600.

BIODATA OF STUDENT

Nur Farhana Binti Che Hassan was born in Kelantan on 23rd June 1988. She attended Sekolah Kebangsaan Kubur Datu, Kelantan (1995) and Sekolah Kebangsaan Kadok, Kelantan (1996) for her primary school education, and Maahad Muhammadi Perempuan Kota Bharu (2001) for her secondary. She entered Malacca Matriculation College in 2007 and pursued her study and obtained a Bachelor of Agricultural Science (Honours) with major in Crop Production from Universiti Malaysia Sabah (2012). She registered for Master of Science (Soil Science) in 2013 after receiving a scholarship under the MyMaster-MyBrain15 from the Ministry of Education Malaysia (formerly known as Ministry of Higher Education Malaysia). Soon after, she was offered the research assistantship under the Graduate Research Fellowship, UPM. Her research interests are mainly in the area of soil-plant-microbe interactions and soil potassium. She had presented scientific papers in conferences and seminars during her postgraduate study.

LIST OF PUBLICATIONS

Journal paper:

Nur Farhana Che Hassan, Roslan Ismail, Zulkifli H. Shamsuddin and Radziah Othman. 2019. Application of *Bacillus subtilis* UPMB10 on Soil Potassium Dissolution for Rice Growth and Yield under Irrigated Condition. *International Journal of Agriculture and Environmental Research*. (In-press)

Conference/Symposium:

Nur Farhana, C. H. and Zulkifli H. Shamsuddin. 2014. Tolerance of Plant Growth-Promoting Rhizobacterial Isolate (UPMB10) to Acidity and its Effect on Growth of Rice. *In Proceedings of the Soil Science Conference of Malaysia 2014*. (Poster Presenter)

Nur Farhana, C. H. and Zulkifli H. Shamsuddin. 2014. Application of Plant Growth-Promoting on Potassium Uptake, Growth and Yield of Rice. *In LRGs Symposium May 2014*. (Paper Presenter)

Nur Farhana Che Hassan and Zulkifli H. Shamsuddin. 2014. Application of Plant Growth-Promoting Rhizobacteria on Potassium Utilisation, Growth and Yield of Rice. *In National Postgraduate Seminar 2014*. (Poster Presenter)

C. H. N. Farhana, Z. H. Shamsuddin and I. Roslan. 2015. Application of *Bacillus Subtilis* UPMB10 on Potassium Utilization Efficiency for Rice Growth and Yield under Flooded Condition. *In Proceedings of the Soil Science Conference of Malaysia 2015*. (Paper Presenter)

Nur Farhana, C. H., Roslan, I. Radziah, O. and Shamsuddin, Z. H. 2016. Application of Potassium Solubilizing Bacterial Inoculation on Soil Potassium Content, Growth and Yield of Irrigated Rice. *In Proceedings of the Soil Science Conference of Malaysia 2016*. (Paper Presenter)



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2018/2019

TITLE OF THESIS / PROJECT REPORT :

EFFECT OF *Bacillus subtilis* UPMB10 INOCULATION ON GROWTH AND YIELD OF IRRIGATED RICE GROWN IN SEMARAK-KEMASIN PADDY SOILS AND ITS CLAY MINERALOGY

NAME OF STUDENT : NUR FARHANA BINTI CHE HASSAN

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

*Please tick (√)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for:

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

(Signature of Chairman of Supervisory Committee)
Name:

Date :

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]