

# IMPROVED PROPAGATION, MAXIMIZING YIELD AND ENHANCING ZERUMBONE PRODUCTION IN *Zingiber zerumbet* (L.) Smith. THROUGH SHADING AND NPK FERTILIZATION

**GOH SUZANNE** 



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## IMPROVED PROPAGATION, MAXIMIZING YIELD AND ENHANCING ZERUMBONE PRODUCTION IN *Zingiber zerumbet* (L.) Smith THROUGH SHADING AND NPK FERTILIZATION

By

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**June 2019** 

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Faculty : Agriculture

Previous studies on Zingiber zerumbet focused mainly on chemical and biological properties of rhizome extract resulting in lack of information on cultivation practices and problems on rhizome dormancy. The determination of shade level and nitrogen (N), phosphorus (P), potassium (K) fertilizer rates are imperative for improvement on growth and rhizome yield of this high medicinal value plant. In addition to having various pharmacological traits, the species are said to be associated with a predominant compound, zerumbone (ZERU), justifying enhancement initiatives on ZERU in the species. The objectives of the present study include improvement in propagation by breaking rhizome dormancy, determination of optimum shade level and maximizing yield through NPK fertilization. The study also aimed at evaluating the inhibition effect on cytotoxic mediators of ZERU in activated microglial cells. At 100 mg/L, 6-Benzylaminopurine (BAP) and ethephon (300 mg/L) performed better in promoting breaking of dormancy in term of highest percentage of bud sprouted. Three shade levels (full sun, 30% and 50% shade levels) and five combinations of NPK fertilizer rates were tested in plant growth and yield performance. Increasing shade level and NPK fertilizer rates showed promoting pattern in plant height, number of leaves, total leaf area per plant, rhizome fresh and dry weight. Field experiment showed that 50% shade with NPK 4 (120 kg N/ha/yr, 140 kg P/ha/yr and 230 kg K/ha/yr) is the best cultural practice with highest rhizome yield (29.71 tonne per hectare) at 10 months after planting (MAP). Production of ZERU as affected by shade level and NPK fertilizer rates was evaluated using High Performance Liquid Chromatography (HPLC) with validated method. The highest concentration of ZERU (40.85% dw/dw) was from plants in pot trial grown under 50% shade and treated with NPK 3 (90 kg N/ha/yr, 105 kg P/ha/yr and 172 kg K/ha/yr). In the field experiment, ZERU decreased with increasing MAP. The highest ZERU content was found in plants harvested at 6 MAP treated with Chicken Manure and NPK 3 (90 kg/ha/yr of N, 105 kg/ha/yr of P and 172 kg/ha/yr of K) at 34.10 % (dw/dw) and 33.04 % (dw/dw) respectively. The rhizome extracts from pot trial and ZERU were used to treat BV2-murine microglial cells with stimulation by lipopolysaccharide (LPS). The results showed selected rhizome extracts and ZERU were significantly reduced nitric oxide (NO) and reactive oxygen species (ROS) level in LPS-stimulated BV2 cells. These results showed that propagation of *Zingiber zerumbet* can be improved by application of BAP at 100 mg/L and ethephon at 300 mg/L while the yield and quality of rhizome was controlled by shading and NPK fertilization. The preliminary study on neuroinflammation showed that ZERU and extracts of rhizome has anti-inflammatory and anti-oxidant properties with significant reduction of NO level and ROS level.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

## PENINGKATAN PEMBIAKAN, MEMAKSIMUMKAN HASIL DAN PENAMBAHBAIKAN PENGELUARAN ZERUMBONE DALAM Zingiber zerumbet (L.) Smith MELALUI RAWATAN TEDUHAN DAN PEMBAJAAN

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Kajian terdahulu ke atas Zingiber zerumbet tertumpu keatas sifat kimia dan biologi ekstrak rizom menyebabkan kurangnya terdapat maklumat mengenai amalan penanaman dan masalah berkaitan dengan kedormanan rizom. Penentuan ke atas tahap naungan dan pembajaan nitrogen (N), phosphorus (P), potassium (K) adalah penting bagi penambahbaikan pertumbuhan dan hasil rizom tumbuhan yang mempunyai nilai perubatan yang tinggi ini. Selain daripada mempunyai beberapa sifat farmakologi, spesies ini dikaitkan dengan satu kompaun yang dominan, zerumbone (ZERU), menjustifikasikan inisiatif penambahbaikan pengeluaran ZERU. Kajian ini telah dijalankan dengan tujuan menambahbaik pembiakan spesies dengan mengatasi keadaan dorman rizom, menentukan paras teduh yang optimum, memaksimumkan hasil melalui pembajaan NPK. Kajian juga bertujuan menilai kesan pengurangan ZERU terhadap mediator sitotoksik dalam sel-sel mikroglial aktif. Kepekatan Benzylaminopurine (BAP) pada 100 mg/L dan ethephon pada 300 mg/L member kesan yang lebih baik bagi pemecahanan kedormanan rizom dengan mempunyai peratus percambahan mata tunas yang paling tinggi. Tiga tahap naungan (penuh matahari, 30% dan 50% tahap naungan) dan lima kombinasi kadar baja NPK diuji dalam pertumbuhan dan prestasi hasil. Tahap naungan dan kadar baja NPK meningkat menunjukkan corak yang menggalakkan dalam ketinggian tumbuhan, jumlah daun, jumlah permukaan daun setiap tumbuhan, berat rizom segar dan rizom kering. Eksperimen lapangan melaporkan 50% naungan dengan NPK 4 (120 kg N/hektar/tahun, 140 kg P/ hektar/tahun and 230 kg K/ hektar/tahun) adalah amalan penanaman terbaik dengan hasil rizom tertinggi (29.71 tan setiap hektar) pada 10 bulan selepas penanaman (BSP). Penghasilan ZERU yang terjejas oleh tahap naungan dan kadar baja NPK dinilai dengan Kromatografi Cecair Prestasi Tinggi dengan kaedah yang disahkan. Kepekatan ZERU tertinggi (40.85% berat kering/berat kering) adalah daripada tanaman dalam percubaan polybags yang ditanam bawah naungan 50% dan dirawat dengan NPK 3 (90 kg N/hektar/tahun, 105 kg P/ hektar/tahun and 172 kg

K/ hektar/tahun). Dalam eksperimen lapangan, ZERU berkurangan dengan peningkatan BSP. Kandungan ZERU tertinggi ditemui di tumbuhan-tumbuhan yang dituai pada 6 BSP yang dirawat dengan tahi ayam dan NPK 3 pada 34.10% (berat kering/berat kering) dan 33.04% (berat kering/berat kering) masing-masing. Ekstrak rhizome dari percubaan polibags dan ZERU digunakan untuk merawat sel-sel mikroglial BV2 dengan rangsangan *lipopolysaccharide* (LPS). Hasilnya menunjukkan ekstrak rizom yang terpilih dan ZERU boleh mengurangkan nitrik oksida (NO) dan tahap spesies oksigen reaktif (ROS) dalam sel BV2 yang dirangsang oleh LPS. Hasil kajian menunjukkan penanaman Zingiber zerumbet boleh ditambahbaikan dengan menggunakan BAP pada 100 mg/L dan ethephon pada 300 mg/L manakala hasil dan kualiti rizom dapat dikawal dengan naungan dan pembajaan NPK. Kajian awal neuroinflammation menunjukkan bahawa ZERU dan ekstrak rizom mempunyai aktiviti anti-keradangan dan anti-oksida dengan berjaya menurunkan tahap NO dan ROS.

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### LIST OF ABBREVIATIONS

μg/mL microgram per mililiter

μM micromolar

μmol/m<sup>2</sup>/s Micromole per meter square per second

ANOVA Analysis of Variance

ATP Adenosine triphosphate

BAP 6-benzylaminopurine

Ca Calcium

Chl a Chlorophyll a

Chl b Chlorophyll b

CM Chicken manure

cm/s centimetre per second

CNB Carbon Nutrient Balance

DCF-DA 2',7'-Dichlorodihydrofluorescein diacetate

DEX Dexamethasone

DMEM Dulbecco's Modified Eagle's Medium

DMRT Duncan Multiple Range Test

DMSO Dimethysulphoxide

DNA Deoxyribonucleic acid

dw Rhizome dry weight

E Transpiration rate

fw Rhizome fresh weight

GCCP Good Cell Culture Practice

G<sub>S</sub> Stomatal conductance

ha hectare

HBSS Hanks' Balanced Salt Solution

HPLC High Performance Liquid Chromatography

K Potassium

kg/ha/yr kilogram per hectare per year

la Leaf area per plant

LOD Limit of Detection

LOQ Limit of Quantification

LPS Lipopolysaccharide

MAP Months after planting

mAU mili Arbitary Unit

Mg Magnesium

mg/mL microgram per mililiter

min minute

mL mililiter

mol/m<sup>2</sup>/s mole per meter square per second

N Nitrogen

ng nanogram

nl Number of leaves per tiller

nm nanometer

NO Nitric Oxide

NPK Nitrogen Phosphate Potassium

nt Number of tiller per plant

P Phosphorus

PA Peak Area

PEP Phosphoenolpyruvate

pH Plant height

P<sub>N</sub> Photosynthesis rate

RNA Ribonucleic acid

ROS Reactive Oxygen Species

RSD Relative Standard of Deviation

SAS Statistical Analysis System

SE Standard of Error

UV Ultraviolet

ZERU Zerumbone

ZZRE Zingiber zerumbet's rhizome extracts

#### **CHAPTER 1**

#### INTRODUCTION

Zingiber zerumbet (L.) Smith or known as 'Lempoyang' in Malaysia, belongs to the family Zingiberaceae. This native rhizomatous herb has been used as food flavoring in culinary preparations and the rhizomes have also been used in traditional medicine. The chemical properties and the biological activities of the rhizome extracts of Lempoyang has been widely studied. Zerumbone (ZERU) has been reported as the predominant compound in the rhizomes of *Z. zerumbet*. Various biological activities of the extracts obtained from the rhizomes were reported including: anti-inflammatory properties (Murakami, Miyamoto, & Ohigashi, 2004), anti-tumour activity (Ramos, Alía, Bravo, & Goya, 2005), preventive towards colon and skin cancer (Tanaka et al., 2001), anti-oxidant activity (Murakami et al., 2002), HIV inhibitory and cytotoxic activities (Dai, Cardellina, Mahon, & Boyd, 1997). Despite the tremendous demand for the rhizomes of *Z. zerumbet*, the agronomic and propagation practices to produce the good quality rhizomes have not been well documented.

Most of the species in the Zingiberaceae family undergo a dormancy period after each growth phase. Overcoming bud dormancy through chemical treatments can improve propagation techniques of *Z. zerumbet* and shorten the dormancy period of rhizomes. Breaking dormancy can also solve the challenges faced during dormancy such as susceptibility towards fungal diseases that further declined success rate in propagation. Application of plant growth regulator such 6-benzylaminopurine (BAP) and 2-chloroethyl phosphoric acid (ethephon) showed success in breaking dormancy in gladiolus corms (Khan, Rahman, & Hossain, 2013) and *Curcuma alismatifolia* (Thohirah, Flora, & Kamalakshi, 2010). Based on the previous literature mentioned, BAP and ethephon were believed to be able to break bud dormancy of *Z. zerumbet*.

The demand for good quality rhizome of Z. zerumbet is increased in Malaysia due to its potential medicinal value. However, the cultural production practices to produce good quality rhizomes are poorly documented and the commercial production of the plant is low. Lempoyang grows under full shade to partial shade areas in the lowland forest (Lim, 2014). Determination of optimum shade level is imperative to improve the growth development in this plant. Optimum light intensity can maximize the growth and yield of plant during growing period (Attridge, 1990). Rhizome yield of Z. officinale and Curcuma longa L. was reduced when growing under high shade level (Hossain et al., 2009; Kratky, Bernabe, Arakaki, White, & Miyasaka, 2013; Sreekala & Jayachandran, 2002). However, response towards shade is varied depending on the natural habitat and physiological changes of the plant. There is lack of scientific literature on the light requirement of Z. zerumbet for optimum growth and yield. In addition, the information on nutrient requirement for Z. zerumbet to produce maximum rhizome yield is lacking. Application of fertilizer is crucial and increasing fertilizer rates significantly increase the rhizome yield of Curcuma longa L. and Zingiber officinale (Akamine et al., 2007; Akhter et al., 2013). The adequate amount of fertilizer to be applied to boost rhizome yield should be known to avoid wastage and soil pollution.

The accumulation of secondary metabolites in plants is greatly affected by the environmental factors such as light intensity, nutrient availability and temperature (Akula & Ravishankar, 2011). Light intensity is the most crucial factor among all the ecological traits. Regulation of plant growth and development by light intensity was attributed by its effect on photosynthesis rate and thus this leads to synthesis of primary and secondary metabolites (Zavala & Ravetta, 2001). High light intensity were resulted in enhancement of total phenolic content in medicinal plants such as *Kaempferia parviflora* (Labrooy, Abdullah, Abdullah, & Stanslas, 2016), *Zingiber officinale* (Ghasemzadeh & Ghasemzadeh, 2011) and *Labisa pumila* (Karimi, Jaafar, Ghasemzadeh, & Ibrahim, 2013). In order to produce and obtain higher phytochemical levels in plants, suitable light intensity and nutrient requirement is necessary.

In the rhizome of *Z. zerumbet*, ZERU was found abundantly and had drawn worldwide attention to study its various effects such as anti-cancer, anti-inflammation and others (Koga, Beltrame, & Pereira, 2016). However, the potential role of ZERU and rhizome extracts to inhibit neurotoxics substances such nitric oxide (NO) and reactive oxygen species (ROS) was not evaluated so far. With the understanding of elevation in excess neurotoxics substances leads to neurodegeneration, the possible therapeutic strategy is to determine ways to suppress the release of neurotoxic substances. Hence, this study aimed to investigate the problem stated above with the general objectives:

- 1. To break dormancy in the rhizome of *Z. zerumbet* through pre-soaked technique using BAP and ethephon
- 2. To determine optimum shade levels and NPK fertilizer for better growth performance and rhizome yield at different harvesting stages
- 3. To enhance the production of zerumbone in rhizome of *Z. zerumbet* under optimal shade levels and NPK fertilizer rates
- 4. To evaluate effect of zerumbone (ZERU) and *Zingiber zerumbet*'s rhizome extracts (ZZRE) to inhibit nitric oxide (NO) and anti-oxidant properties via *in vitro* model of lipopolysaccharide (LPS)-induced brain injury using microglial BV2 cells

#### **REFERENCES**

- Abdelwahab, S. I., Abdul, A. B., Devi, N., Taha, M. M. E., Al-Zubairi, A. S., Mohan, S., & Mariod, A. A. (2010). Regression of cervical intraepithelial neoplasia by zerumbone in female Balb/c mice prenatally exposed to diethylstilboestrol: involvement of mitochondria-regulated apoptosis. *Experimental and Toxicologic Pathology*, 62(5), 461-469.
- Abrams, M. D., & Mostoller, S. A. (1995). Gas exchange, leaf structure and nitrogen in contrasting successional tree species growing in open and understory sites during a drought. *Tree physiology*, 15(6), 361-370.
- Agostini-Costa, T. d. S., Vieira, R. F., Bizzo, H. R., Silveira, D., & Gimenes, M. A. (2012). Secondary metabolites *Chromatography and Its Applications*: InTech.
- Akamine, H., Hossain, M. A., Ishimine, Y., Yogi, K., Hokama, K., Iraha, Y., & Aniya, Y. (2007). Effects of application of N, P and K alone or in combination on growth, yield and curcumin content of turmeric (*Curcuma longa L.*). *Plant production science*, 10(1), 151-154.
- Akhter, S., Noor, S., Islam, M., Masud, M., Talukder, M., & Hossain, M. (2013). Effect of potassium fertilization on the yield and quality of ginger (*Zingiber officinale*) grown on a K deficient terrace soil of level Barind Tract (AEZ 25) in Northern Bangladesh. *Research Findings: e-ifc*(35).
- Aktan, F., Henness, S., Tran, V. H., Duke, C. C., Roufogalis, B. D., & Ammit, A. J. (2006). Gingerol metabolite and a synthetic analogue Capsarol<sup>TM</sup> inhibit macrophage NF-κB-mediated iNOS gene expression and enzyme activity. *Planta Medica*, 72(08), 727-734.
- Akula, R., & Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. *Plant signaling & behavior*, 6(11), 1720-1731.
- Albayrak, S. (2007). Effects of temperature and light intensity on growth of fodder beet (*Beta vulgaris* L. var. crassa Mansf.). *Bangladesh Journal of Botany*, 36(1), 1-12.
- Arimura, C., Finger, F., & Casali, V. (2000). Effect of ANA and BAP on ginger (*Zingiber officinale* Roscoe) sprouting in solid and liquid medium. *Revista Brasileira de Plantas Medicinais*, 2(2), 23-26.
- Asher, C. J., & Lee, M. (1975). Diagnosis and correction of nutritional disorders in ginger.
- Attoe, E., & Osodeke, V. (2009). Effects of NPK on growth and yield of ginger (*Zingiber officinale* Roscoe) in soils of contrasting parent materials of Cross River State. *Electronic Journal of Environmental, Agricultural & Food Chemistry*, 8(12).

- Attridge, T. H. (1990). *Light and plant responses: a study of plant photophysiology and the natural environment*: Cambridge University Press.
- Baby, S., Dan, M., Thaha, A. R., Johnson, A. J., Kurup, R., Balakrishnapillai, P., & Lim, C. K. (2009). High content of zerumbone in volatile oils of *Zingiber zerumbet* from southern India and Malaysia. *Flavour and Fragrance Journal*, 24(6), 301-308.
- Balakrishnan, R., Radzan, R., & Bhattacharyya, S. (1956). Oil of *Zingiber zerumbet* Smith. Part 1. Isolation of various constituents and characterization of the inonocyclic ketone zerumbone. *Perfumery Essential Oil Record*, 47, 274-279.
- Basak, S., Sarma, G. C., & Rangan, L. (2010). Ethnomedical uses of Zingiberaceous plants of Northeast India. *Journal of ethnopharmacology*, 132(1), 286-296.
- Basra, A. (2000). Plant growth regulators in agriculture and horticulture: their role and commercial uses: CRC Press.
- Behura, S. (2001). Effect of nitrogen and potassium on growth parameters and rhizomatic characters of mango-ginger (*Curcuma amada*). *Indian Journal of Agronomy*, 46(4): 474-751.
- Bélanger, J. M., Paré, J. J., & Sigouin, M. (1997). High performance liquid chromatography (HPLC): Principles and applications *Techniques and Instrumentation in Analytical Chemistry* (Vol. 18, pp. 37-59): Elsevier.
- Bennett, R. N., & Wallsgrove, R. M. (1994). Secondary metabolites in plant defence mechanisms. *New phytologist*, 127(4), 617-633.
- Bhuiyan, M. N. I., Chowdhury, J. U., & Begum, J. (2009). Chemical investigation of the leaf and rhizome essential oils of *Zingiber zerumbet* (L.) Smith from Bangladesh. *Bangladesh Journal of Pharmacology*, 4(1), 9-12.
- Blevins, D. G. (1994). Uptake, translocation, and function of essential mineral elements in crop plants. *Physiology and Determination of Crop Yield. Madison:* ASA, CSSA, SSSA, 259-275.
- Bongue Bartelsman, M., & Phillips, D. (1995). Nitrogen stress regulates gene expression of enzymes in the flavonoid biosynthetic pathway of tomato [anthocyane]. *Plant Physiology and Biochemistry (France)*.
- Brochu, S., Olivier, M., & Rivest, S. (1999). Neuronal activity and transcription of proinflammatory cytokines, IκBα, and iNOS in the mouse brain during acute endotoxemia and chronic infection with Trypanosoma brucei brucei. *Journal of neuroscience research*, 57(6), 801-816.
- Brown, P. D., Tokuhisa, J. G., Reichelt, M., & Gershenzon, J. (2003). Variation of glucosinolate accumulation among different organs and developmental stages of *Arabidopsis thaliana*. *Phytochemistry*, 62(3), 471-481.

- Bryant, J., Chapin, F., Reichardt, P., & Clausen, T. (1985). Adaptation to resource availability as a determinant of chemical defense strategies in woody plants *Chemically mediated interactions between plants and other organisms* (pp. 219-237): Springer.
- Bryant, J. P., Chapin III, F. S., & Klein, D. R. (1983). Carbon/nutrient balance of boreal plants in relation to vertebrate herbivory. *Oikos*, 357-368.
- Burkill, I. H. (2015). A dictionary of the economic products of the Malay Peninsula.
- Bustamam, A., Ibrahim, S., Al-Zubairi, A., Met, M., & Syam, M. (2008). Zerumbone: a natural compound with anti-cholinesterase activity. *American Journal of Pharmacology and Toxicology*, 3(3), 209-211.
- Chalker-Scott, L. (1989). The role of phenolic compounds in plant stress responses. Low temperature stress physiology in crops.
- Chalker-Scott, L., & Fuchigami, L. (1989). The role of phenolic compounds in plant stress responses.
- Chane-Ming, J., Vera, R., & Chalchat, J.-C. (2003). Chemical composition of the essential oil from rhizomes, leaves and flowers of *Zingiber zerumbet* Smith from Reunion Island. *Journal of Essential Oil Research*, 15(3), 202-205.
- Chaung, H.-C., Ho, C.-T., & Huang, T.-C. (2008). Anti-hypersensitive and antiinflammatory activities of water extract of *Zingiber zerumbet* (L.) Smith. *Food* and agricultural immunology, 19(2), 117-129.
- Chauser-Volfson, E., & Gutterman, Y. (1998). Content and distribution of anthrone C-glycosides in the South African arid plant species *Aloe mutabilis* growing in direct sunlight and in shade in the Negev Desert of Israel. *Journal of arid environments*, 40(4), 441-451.
- Chen, C.-Y., Peng, W.-H., Tsai, K.-D., & Hsu, S.-L. (2007). Luteolin suppresses inflammation-associated gene expression by blocking NF-κB and AP-1 activation pathway in mouse alveolar macrophages. *Life Sciences*, 81(23-24), 1602-1614.
- Chen, I.-N., Chang, C.-C., Ng, C.-C., Wang, C.-Y., Shyu, Y.-T., & Chang, T.-L. (2008). Antioxidant and antimicrobial activity of Zingiberaceae plants in Taiwan. *Plant foods for human Nutrition*, 63(1), 15-20.
- Chen, X. (2013). Improved Propagation, Early Establishment and Fruit Development of the Miracle Fruit, (Synsepalum Dulcificum Daniell.). Universiti Putra Malaysia.

- Chien, C.-C., Shen, S.-C., Yang, L.-Y., & Chen, Y.-C. (2012). Prostaglandins as Negative Regulators Against Lipopolysaccharide, Lipoteichoic Acid, and Peptidoglycan-Induced Inducible Nitric Oxide Synthase/Nitric Oxide Production Through Reactive Oxygen Species—Dependent Heme Oxygenase 1 Expression in Macrophages. *Shock*, 38(5), 549-558.
- Chirangini, P., & Sharma, G. (2005). In vitro propagation and microrhizome induction in *Zingiber cassumunar* (Roxb.) an antioxidant-rich medicinal plant. *J. Food Agric. Environ*, *3*(1), 139-142.
- Cho, N., Choi, J. H., Yang, H., Jeong, E. J., Lee, K. Y., Kim, Y. C., & Sung, S. H. (2012). Neuroprotective and anti-inflammatory effects of flavonoids isolated from Rhus verniciflua in neuronal HT22 and microglial BV2 cell lines. Food and Chemical Toxicology, 50(6), 1940-1945.
- Chow, Y.-L., Lee, K.-H., Vidyadaran, S., Lajis, N. H., Akhtar, M. N., Israf, D. A., & Syahida, A. (2012). Cardamonin from *Alpinia rafflesiana* inhibits inflammatory responses in IFN-γ/LPS-stimulated BV2 microglia via NF-κB signalling pathway. *International immunopharmacology*, *12*(4), 657-665.
- Chuang, D. Y., Chan, M.-H., Zong, Y., Sheng, W., He, Y., Jiang, J. H., . . . Cui, J. (2013). Magnolia polyphenols attenuate oxidative and inflammatory responses in neurons and microglial cells. *Journal of Neuroinflammation*, 10(1), 786.
- Chung, G., Rowe, R., & Field, R. (1982). Relationship between shoot and roots of cucumber plants under nutritional stress. *Annals of Botany*, 50(6), 859-861.
- Coelho, G. C., Rachwal, M. F., Dedecek, R. A., Curcio, G. R., Nietsche, K., & Schenkel, E. P. (2007). Effect of light intensity on methylxanthine contents of *Ilex paraguariensis* A. St. Hil. *Biochemical systematics and ecology, 35*(2), 75-80.
- Combrinck, M., Williams, J., De Berardinis, M. A., Warden, D., Puopolo, M., Smith, A. D., & Minghetti, L. (2006). Levels of CSF prostaglandin E2, cognitive decline, and survival in Alzheimer's disease. *Journal of Neurology, Neurosurgery & Psychiatry*, 77(1), 85-88.
- Coombs, J., Hall, D. O., & Long, S. (2014). Techniques in bioproductivity and photosynthesis: pergamon international library of science, technology, engineering and social studies: Elsevier.
- Criley, R. (2001). Method of application affects effectiveness of cytokinin in inducing bud break on Heliconia rhizomes. *Horticulture Digest*, 106.
- Croteau, R., Kutchan, T. M., & Lewis, N. G. (2000). Natural products (secondary metabolites). *Biochemistry and molecular biology of plants*, 24, 1250-1319.
- Dai, J.-R., Cardellina, J. H., Mahon, J. B. M., & Boyd, M. R. (1997). Zerumbone, an HIV-inhibitory and cytotoxic sesquiterpene of *Zingiber aromaticum* and *Z. zerumbet. Natural Product Letters, 10*(2), 115-118.

- De Hertogh, A., & Le Nard, M. (1993). Physiological and biochemical aspects of flower bulbs. *The physiology of flower bulbs*. *Amsterdam: Elsevier*, 53-69.
- del Pilar Paz, M., Kuehny, J., McClure, G., Graham, C., & Criley, R. (2004). Effect of rhizome storage duration and temperature on carbohydrate content, respiration, growth, and flowering of ornamental ginger. Paper presented at the IX International Symposium on Flower Bulbs 673.
- Dev, S. (1960). Studies in sesquiterpenes—XVI: Zerumbone, a monocyclic sesquiterpene ketone. *Tetrahedron*, 8(3-4), 171-180.
- Devi, N. B., Singh, P., & Das, A. K. (2014). Ethnomedicinal utilization of Zingiberaceae in the valley districts of Manipur. *J. Environ. Sci. Toxicol. Food Technol*, 8(2), 21-23.
- Dixon, R. A., & Paiva, N. L. (1995). Stress-induced phenylpropanoid metabolism. *The plant cell*, 7(7), 1085.
- Dokic, I., Hartmann, C., Herold-Mende, C., & Régnier-Vigouroux, A. (2012). Glutathione peroxidase 1 activity dictates the sensitivity of glioblastoma cells to oxidative stress. *Glia*, 60(11), 1785-1800.
- Dole, J. M., & Wilkins, H. F. (1999). *Floriculture: principles and species*: Prentice-Hall Inc.
- Emongor, V., & Chweya, J. (1992). Effect of nitrogen and variety on essential-oil yield and composition from chamomile flowers. *Trop Agr*, 69, 290-292.
- Fageria, N. K., & Baligar, V. C. (2003). Fertility management of tropical acid soils for sustainable crop production. *Handbook of soil acidity*, 359-385.
- Fajer, E., Bowers, M., & Bazzaz, F. (1992). The effect of nutrients and enriched CO2 environments on production of carbon-based allelochemicals in Plantago: a test of the carbon/nutrient balance hypothesis. *The American Naturalist*, 140(4), 707-723.
- Fauziah, C., Jamilah, I., & Syed Omar, S. (1997). An evaluation of cation exchange capacity methods for acid tropical soils. *Pertanika journal of tropical agricultural science*, 20, 113-120.
- Fredeen, A. L., Raab, T. K., Rao, I. M., & Terry, N. (1990). Effects of phosphorus nutrition on photosynthesis in *Glycine max* (L.) Merr. *Planta*, *181*(3), 399-405.
- Furutani, S., & Nagao, M. (1986). Influence of Daminozide, Gibberellic acid, and Ethephon on flowering, shoot growth, and yield of ginger. *HortScience*, 21(3), 428-429.
- Gao, H.-M., & Hong, J.-S. (2008). Why neurodegenerative diseases are progressive: uncontrolled inflammation drives disease progression. *Trends in immunology*, 29(8), 357-365.

- Gershenzon, J. (1994). Metabolic costs of terpenoid accumulation in higher plants. *Journal of chemical ecology*, 20(6), 1281-1328.
- Gershenzon, J., Murtagh, G. J., & Croteau, R. (1993). Absence of rapid terpene turnover in several diverse species of terpene-accumulating plants. *Oecologia*, 96(4), 583-592.
- Ghasemzadeh, A., & Ghasemzadeh, N. (2011). Effects of shading on synthesis and accumulation of polyphenolic compounds in ginger (*Zingiber officinale* Roscoe) varieties. *Journal of Medicinal Plants Research*, 5(11), 2435-2441.
- Gonzalez-Scarano, F., & Baltuch, G. (1999). Microglia as mediators of inflammatory and degenerative diseases. *Annual review of neuroscience*, 22(1), 219-240.
- Gowda, J. (1994). Effect of ethrel and benzyladenine on breaking dormancy in gladiolus cormels. Floriculture Technology, Trades and Trends. Oxford and IBH Publishing Co. Pvt. Ltd. Calcutta, 203-204.
- Gregoriou, K., Pontikis, K., & Vemmos, S. (2007). Effects of reduced irradiance on leaf morphology, photosynthetic capacity, and fruit yield in olive (*Olea europaea* L.). *Photosynthetica*, 45(2), 172-181.
- Grusak, M. A., Broadley, M. R., & White, P. J. (2001). Plant Macro- and Micronutrient Minerals. *eLS*, 1-6.
- Guideline, I. H. T. (2005). Validation of analytical procedures: text and methodology Q2 (R1). Paper presented at the International Conference on Harmonization, Geneva, Switzerland.
- Gupta, M. K., & Rajput, S. (2015). Development and Validation of RP-HPLC Method for Quantitation of Itraconazole in Tablets Dosage Form. *International Journal of Pharma Research & Review*, 4(11), 23-29.
- Hampel, H., & Broich, K. (2009). Enrichment of MCI and early Alzheimer's disease treatment trials using neurochemical & imaging candidate biomarkers. *JNHA-The Journal of Nutrition, Health and Aging*, 13(4), 373-375.
- Haque, M. A., Jantan, I., & Harikrishnan, H. (2018). Zerumbone suppresses the activation of inflammatory mediators in LPS-stimulated U937 macrophages through MyD88-dependent NF-κB/MAPK/PI3K-Akt signaling pathways. *International immunopharmacology*, 55, 312-322.
- Hashemi, S., Zulkifli, I., Hair Bejo, M., Farida, A., & Somchit, M. (2008). Acute toxicity study and phytochemical screening of selected herbal aqueous extract in broiler chickens. *Int J pharmacol*, *4*(5), 352-360.
- Hawkesford, M., Horst, W., Kichey, T., Lambers, H., Schjoerring, J., Møller, I. S., & White, P. (2012). Functions of macronutrients *Marschner's Mineral Nutrition of Higher Plants (Third Edition)* (pp. 135-189): Elsevier.

- Hemm, M. R., Rider, S. D., Ogas, J., Murry, D. J., & Chapple, C. (2004). Light induces phenylpropanoid metabolism in Arabidopsis roots. *The Plant Journal*, 38(5), 765-778.
- Hossain, M. A., Akamine, H., Ishimine, Y., Teruya, R., Aniya, Y., & Yamawaki, K. (2009). Effects of relative light intensity on the growth, yield and curcumin content of turmeric (*Curcuma longa* L.) in Okinawa, Japan. *Plant production science*, 12(1), 29-36.
- Hou, J.-l., Li, W.-d., Zheng, Q.-y., Wang, W.-q., Xiao, B., & Xing, D. (2010). Effect of low light intensity on growth and accumulation of secondary metabolites in roots of *Glycyrrhiza uralensis* Fisch. *Biochemical systematics and ecology*, 38(2), 160-168.
- Huang, G.-C., Chien, T.-Y., Chen, L.-G., & Wang, C.-C. (2005). Antitumor effects of zerumbone from *Zingiber zerumbet* in P-388D1 cells in vitro and in vivo. *Planta Medica*, 71(03), 219-224.
- Hussain, S., Sharma, A., Singh, P., & Hore, D. (2006). Effect of varying levels of nitrogen and phosphorus on growth and yield of the medicinal plant, *Alpinia galanga* Wild.
- Ibrahim, H., Khalid, N., & Hussin, K. (2007). Cultivated gingers of peninsular Malaysia: ultilization profiles and micropropagation. *The Gardens' Bulletin Singapore*, 59(1-2), 71-88.
- Islam, A., Asher, C., & Edwards, D. (1978). Germination and early growth of ginger (Zingiber officinale Roscoe), 2: Effects of 2-chloroethyl phosphonic acid or elevated temperature pretreatments. Tropical Agriculture.
- Ivonyi, I., Izsoki, Z., & van der Werf, H. M. (1997). Influence of nitrogen supply and P and K levels of the soil on dry matter and nutrient accumulation of. *Journal of the International Hemp Association*, 4(2).
- Jana, J., Datta, S., Bhaisare, P., & Thapa, A. (2017). Effect of organic, inorganic source of nutrients and Azospirillum on yield and quality of turmeric (*Curcuma longa L.*). *Int. J. Curr. Microbiol. App. Sci, 6*(2), 966-970.
- Jang, D. S., Han, A.-R., Park, G., Jhon, G.-J., & Seo, E.-K. (2004). Flavonoids and aromatic compounds from the rhizomes of *Zingiber zerumbet*. *Archives of pharmacal research*, 27(4), 386-389.
- Jang, D. S., & Seo, E.-K. (2005). Potentially bioactive two new natural sesquiterpenoids from the rhizomes of *Zingiber zerumbet*. *Archives of pharmacal research*, 28(3), 294-296.
- Jeong, K. Y., Pasian, C. C., McMahon, M., & Tay, D. (2009). Growth of six Begonia species under shading. *The Open Horticulture Journal*, 2, 22-28.

- Jones Jr, J. B. (2001). *Laboratory guide for conducting soil tests and plant analysis*: CRC press.
- Jung, H. W., Son, H. Y., Minh, C. V., Kim, Y. H., & Park, Y. K. (2008). Methanol extract of Ficus leaf inhibits the production of nitric oxide and proinflammatory cytokines in LPS- stimulated microglia via the MAPK pathway. *Phytotherapy Research*, 22(8), 1064-1069.
- Jung, H. W., Yoon, C.-H., Park, K. M., Han, H. S., & Park, Y.-K. (2009). Hexane fraction of Zingiberis Rhizoma Crudus extract inhibits the production of nitric oxide and proinflammatory cytokines in LPS-stimulated BV2 microglial cells via the NF-kappaB pathway. Food and Chemical Toxicology, 47(6), 1190-1197.
- Kantar, M., Betts, K., Hulke, B. S., Stupar, R. M., & Wyse, D. (2012). Breaking tuber dormancy in *Helianthus tuberosus* L. and interspecific hybrids of *Helianthus annuus* L.× *Helianthus tuberosus*. *HortScience*, 47(9), 1342-1346.
- Karimi, E., Jaafar, H. Z., Ghasemzadeh, A., & Ibrahim, M. H. (2013). Light intensity effects on production and antioxidant activity of flavonoids and phenolic compounds in leaves, stems and roots of three varieties of *Labisia pumila* Benth. *Australian Journal of Crop Science*, 7(7), 1016.
- Keong, Y. S., Alitheen, N. B., Mustafa, S., Aziz, S. A., Rahman, M. A., & Ali, A. M. (2010). Immunomodulatory effects of zerumbone isolated from roots of *Zingiber zerumbet. Pak J Pharm Sci*, 23(1), 75-82.
- Khan, F., Rahman, M., & Hossain, M. (2013). Effect of benzyladenine and gibberellic acid on dormancy breaking, growth and yield of gladiolus corms over different storage periods.
- Khuankaew, T., Ohyama, T., & Ruamrungsri, S. (2009). Effects of Ethephon Application on Growth and Development of *Curcuma alismatifolia* Gagnep.
- Kim YSJoh, T. (2006). Microglia, major player in the brain inflammation: their roles in the pathogenesis of Parkinson's disease. *Exp Mol Med*, *38*, 333347.
- Kirana, C., McIntosh, G. H., Record, I. R., & Jones, G. P. (2003). Antitumor activity of extract of *Zingiber aromaticum* and its bioactive sesquiterpenoid zerumbone. *Nutrition and cancer*, 45(2), 218-225.
- Kitayama, T. (2011). Attractive reactivity of a natural product, zerumbone. *Bioscience, biotechnology, and biochemistry*, 75(2), 199-207.
- Koga, A. Y., Beltrame, F. L., & Pereira, A. V. (2016). Several aspects of *Zingiber zerumbet*: a review. *Revista Brasileira de Farmacognosia*, 26(3), 385-391.
- Kolářová, P., Bezděčková, L., & Procházková, Z. (2010). Effect of gibberellic acid and ethephon on the germination of European beech dormant and chilled beechnuts. *Journal of Forest Science*, 56(9), 389-396.

- Koo, B.-S., Lee, W.-C., Chung, K.-H., Ko, J.-H., & Kim, C.-H. (2004). A water extract of *Curcuma longa* L.(Zingiberaceae) rescues PC12 cell death caused by pyrogallol or hypoxia/reoxygenation and attenuates hydrogen peroxide induced injury in PC12 cells. *Life sciences*, 75(19), 2363-2375.
- Kowalenko, C. G. (2001). Assessment of Leco CNS-2000 analyzer for simultaneously measuring total carbon, nitrogen, and sulphur in soil. *Communications in Soil Science and Plant Analysis*, 32(13-14), 2065-2078.
- Kratky, B., Bernabe, C., Arakaki, E., White, F., & Miyasaka, S. (2013). Shading Reduces Yields of Edible Ginger Rhizomes Grown in Sub-Irrigated Pots. *UH-CTAHR*, *Rootcroop*, RC-2 May 2013, 1-5.
- Kulpapangkorn, W., & Mai-leang, S. (2012). Effect of plant nutrition on turmeric production. *Procedia Engineering*, 32, 166-171.
- Labrooy, C. D., Abdullah, T. L., Abdullah, N. A. P., & Stanslas, J. (2016). Optimum shade enhances growth and 5, 7-Dimethoxyflavone accumulation in *Kaempferia parviflora* Wall. ex Baker cultivars. *Scientia horticulturae*, 213, 346-353.
- Lako, J., Trenerry, V. C., Wahlqvist, M., Wattanapenpaiboon, N., Sotheeswaran, S., & Premier, R. (2007). Phytochemical flavonols, carotenoids and the antioxidant properties of a wide selection of Fijian fruit, vegetables and other readily available foods. *Food Chemistry*, 101(4), 1727-1741.
- Lang, G. (1987). Endo-, para-and ecodormancy: physiological terminology and classification for dormancy research. *Hortic. Sci.*, 22, 271-277.
- Larsen, K., Ibrahim, H., Khaw, S., & Saw, L. (1999). *Gingers of peninsular Malaysia and Singapore*: Natural History Publications (Borneo).
- Lawlor, D. W. (1993). *Photosynthesis: molecular, physiological and environmental processes*: Longman scientific & technical.
- Lawlor, D. W. (2002). Carbon and nitrogen assimilation in relation to yield: mechanisms are the key to understanding production systems. *Journal of experimental Botany*, 53(370), 773-787.
- Li, P.-F., Dietz, R., & Von Harsdorf, R. (1997). Reactive oxygen species induce apoptosis of vascular smooth muscle cell. *FEBS letters*, 404(2-3), 249-252.
- Lim, T. (2014). Zingiber zerumbet Edible Medicinal and Non Medicinal Plants (pp. 857-876): Springer.
- Lincoln, D., & Couvet, D. (1989). The effect of carbon supply on allocation to allelochemicals and caterpillar consumption of peppermint. *Oecologia*, 78(1), 112-114.

- Liu, B., & Hong, J.-S. (2003). Role of microglia in inflammation-mediated neurodegenerative diseases: mechanisms and strategies for therapeutic intervention. *Journal of Pharmacology and Experimental Therapeutics*, 304(1), 1-7.
- Lujiu, L., Xisheng, G., Jiejun, G., Nan, D., & Lin, Z. (2004). Ginger Response to Potassium in Anhui Province. *Better Crops*, 88(1), 22-24.
- Mahmoud, S. S., & Croteau, R. B. (2002). Strategies for transgenic manipulation of monoterpene biosynthesis in plants. *Trends in plant science*, 7(8), 366-373.
- Makino, A., Sato, T., Nakano, H., & Mae, T. (1997). Leaf photosynthesis, plant growth and nitrogen allocation in rice under different irradiances. *Planta*, 203(3), 390-398.
- Malek, S. N. A., Ibrahim, H., Hong, S. L., Lee, G. S., Chan, K. S., Yusoff, M. M., & Ali, N. A. M. (2005). Essential Oils of *Zingiber ottensii* Valet. and *Zingiber zerumbet* (L.) Sm. from Sabah, Malaysia. *Malaysian Journal of Science*, 24(1), 49-58.
- Marschner, H. (1995). Mineral nutrition of higher plants., 2nd edn (Academic Press: London).
- Masuda, T., Jitoe, A., Kato, S., & Nakatani, N. (1991). Acetylated flavonol glycosides from *Zingiber zerumbet*. *Phytochemistry*, 30(7), 2391-2392.
- McCree, K. J. (1971). The action spectrum, absorptance and quantum yield of photosynthesis in crop plants. *Agricultural Meteorology*, 9, 191-216.
- McGeer, E. G., & McGeer, P. L. (2003). Inflammatory processes in Alzheimer's disease. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 27(5), 741-749.
- Mendis, E., Kim, M.-M., Rajapakse, N., & Kim, S.-K. (2008). Sulfated glucosamine inhibits oxidation of biomolecules in cells via a mechanism involving intracellular free radical scavenging. *European journal of pharmacology*, 579(1-3), 74-85.
- Merrill, J., Koyanagi, Y., Zack, J., Thomas, L., Martin, F., & Chen, I. (1992). Induction of interleukin-1 and tumor necrosis factor alpha in brain cultures by human immunodeficiency virus type 1. *Journal of virology*, 66(4), 2217-2225.
- Modupeola, T., & Olaniyi, J. (2015). Effects of nitrogen (N) fertilizer and plant spacing on the growth and rhizome yield of turmeric (*Curcuma Longa L.*) in Ibadan South-West Nigeria. *Int. J. of Plant Sci. and Eco, 1*(4), 149-154.
- Modupeola, T., Olaniyi, J., Abdul-Rafiu, A., Taylor, O., Fariyike, T., & Oyebamiji, T. (2013). Effect of organic phosphorus fertilizer and plant density on the growth, yield and nutritional value of ginger (*Zingiber officinale*). *Int J Agric Res*, 8, 94-100.

- Mohotti, A. J., & Lawlor, D. W. (2002). Diurnal variation of photosynthesis and photoinhibition in tea: effects of irradiance and nitrogen supply during growth in the field. *Journal of experimental Botany*, 53(367), 313-322.
- More, S. V., Kumar, H., Kim, I. S., Song, S.-Y., & Choi, D.-K. (2013). Cellular and molecular mediators of neuroinflammation in the pathogenesis of Parkinson's disease. *Mediators of inflammation*, 2013.
- Murakami, A., Hayashi, R., Takana, T., Kwon, K. H., Ohigashi, H., & Safitri, R. (2003). Suppression of dextran sodium sulfate-induced colitis in mice by zerumbone, a subtropical ginger sesquiterpene, and nimesulide: separately and in combination. *Biochemical pharmacology*, 66(7), 1253-1261.
- Murakami, A., Miyamoto, M., & Ohigashi, H. (2004). Zerumbone, an antiinflammatory phytochemical, induces expression of proinflammatory cytokine genes in human colon adenocarcinoma cell lines. *Biofactors*, 21(1-4), 95-101.
- Murakami, A., Takahashi, D., Kinoshita, T., Koshimizu, K., Kim, H. W., Yoshihiro, A., . . . Ohigashi, H. (2002). Zerumbone, a Southeast Asian ginger sesquiterpene, markedly suppresses free radical generation, proinflammatory protein production, and cancer cell proliferation accompanied by apoptosis: the α, β-unsaturated carbonyl group is a prerequisite. *Carcinogenesis*, 23(5), 795-802.
- Murakami, A., Takahashi, M., Jiwajinda, S., Koshimizu, K., & Ohigashi, H. (1999). Identification of zerumbone in *Zingiber zerumbet* Smith as a potent inhibitor of 12-O-tetradecanoylphorbol-13-acetate-induced Epstein-Barr virus activation. *Bioscience, biotechnology, and biochemistry, 63*(10), 1811-1812.
- Murakami, A., Tanaka, T., Lee, J. Y., Surh, Y. J., Kim, H. W., Kawabata, K., . . . Ohigashi, H. (2004). Zerumbone, a sesquiterpene in subtropical ginger, suppresses skin tumor initiation and promotion stages in ICR mice. *International journal of cancer*, 110(4), 481-490.
- Nakamura, Y., Yoshida, C., Murakami, A., Ohigashi, H., Osawa, T., & Uchida, K. (2004). Zerumbone, a tropical ginger sesquiterpene, activates phase II drug metabolizing enzymes. *FEBS letters*, *572*(1-3), 245-250.
- Nautiyal, N. (2015). EFFECT OF NITROGEN LEVEL AND PLANT SPACING ON GROWTH AND YIELD OF TURMERIC (Curcuma longa L.). College of Horticulture, Bharsar Campus, VCSG Uttarakhand University of Horticulture and Forestry.
- Nhareet, S. M., & Nur, S. M. (2003). Anti inflammatory property of ethanol and water extracts of *Zingiber zerumbet*. *Indian Journal of Pharmacology*, *35*(3), 181.
- Nwaogu, E., & Ukpabi, U. (2010). Characteristics of Imported Indian Ginger Cultivars in Abia State, Nigeria. *Agricultural Journal*, *5*(1), 31-36.

- Odabas, M. S., Raduğieneuml, J., Camas, N., Janulis, V., & Ivanauskas, L. (2009). The quantitative effects of temperature and light intensity on hyperforin and hypericins accumulation in *Hypericum perforatum* L. *Journal of Medicinal Plants Research*, 3(7), 519-525.
- Okoli, P. S., & Wilson, G. (1986). Response of cassava (Manihot esculenta Crantz) to shade under field conditions. *Field Crops Research*, *14*, 349-359.
- Pandey, A. (1992). Response of turmeric to various levels of nitrogen under terrace condition of mid altitude, Mizoram. *Indian Cocoa*, *Account and Spices Journal*, *16*(1), 14-16.
- Park, J.-H., Park, G. M., & Kim, J.-K. (2015). Zerumbone, sesquiterpene photochemical from ginger, inhibits angiogenesis. *The Korean Journal of Physiology & Pharmacology*, 19(4), 335-340.
- Pawate, S., Shen, Q., Fan, F., & Bhat, N. R. (2004). Redox regulation of glial inflammatory response to lipopolysaccharide and interferonγ. *Journal of neuroscience research*, 77(4), 540-551.
- Paz, M. d. P. (2003). Rhizome manipulation affects growth and development of ornamental gingers.
- Perimal, E. K., Akhtar, M. N., Mohamad, A. S., Khalid, M. H., Ming, O. H., Khalid, S., . . . Israf, D. A. (2011). Zerumbone- Induced Antinociception: Involvement of the l- Arginine- Nitric Oxide- cGMP- PKC- K+ ATP Channel Pathways. *Basic & clinical pharmacology & toxicology*, 108(3), 155-162.
- Phongpaichit, S., Vuddhakul, V., Subhadhirasakul, S., & Wattanapiromsakul, C. (2006). Evaluation of the antimycobacterial activity of extracts from plants used as self-medication by AIDS patients in Thailand. *Pharmaceutical Biology*, 44(1), 71-75.
- Rajendran, L., Ravishankar, G., Venkataraman, L., & Prathiba, K. (1992). Anthocyanin production in callus cultures of *Daucus carota* as influenced by nutrient stress and osmoticum. *Biotechnology letters*, 14(8), 707-712.
- Ralphs, M., Manners, G., & Gardner, D. (1998). Influence of light and photosynthesis on alkaloid concentration in larkspur. *Journal of chemical ecology*, 24(1), 167-182.
- Ramos, S., Alía, M., Bravo, L., & Goya, L. (2005). Comparative effects of food-derived polyphenols on the viability and apoptosis of a human hepatoma cell line (HepG2). *Journal of agricultural and food chemistry*, 53(4), 1271-1280.
- Rao, I. M., Fredeen, A. L., & Terry, N. (1990). Leaf phosphate status, photosynthesis, and carbon partitioning in sugar beet: III. diurnal changes in carbon partitioning and carbon export. *Plant Physiology*, 92(1), 29-36.

- Ravisankar, C., & Muthuswamy, S. (1984). Studies on the endogenous hormonal changes in leaf and rhizome of ginger. *South Indian Hort*, *32*, 347-351.
- Reichardt, P., Chapin, F., Bryant, J., Mattes, B., & Clausen, T. (1991). Carbon/nutrient balance as a predictor of plant defense in Alaskan balsam poplar: potential importance of metabolite turnover. *Oecologia*, 88(3), 401-406.
- Rejdak, K., Eikelenboom, M., Petzold, A., Thompson, E., Stelmasiak, Z., Lazeron, R., . . . Giovannoni, G. (2004). CSF nitric oxide metabolites are associated with activity and progression of multiple sclerosis. *Neurology*, 63(8), 1439-1445.
- Rock, R. B., & Peterson, P. K. (2006). Microglia as a pharmacological target in infectious and inflammatory diseases of the brain. *Journal of Neuroimmune Pharmacology*, *1*(2), 117-126.
- Roy, A., Jana, A., Yatish, K., Freidt, M. B., Fung, Y. K., Martinson, J. A., & Pahan, K. (2008). Reactive oxygen species up-regulate CD11b in microglia via nitric oxide: Implications for neurodegenerative diseases. *Free Radical Biology and Medicine*, 45(5), 686-699.
- Ruslay, S., Abas, F., Shaari, K., Zainal, Z., Sirat, H., Israf, D. A., & Lajis, N. H. (2007). Characterization of the components present in the active fractions of health gingers (*Curcuma xanthorrhiza* and *Zingiber zerumbet*) by HPLC–DAD–ESIMS. *Food Chemistry*, 104(3), 1183-1191.
- Rylski, I., & Spigelman, M. (1986). Effect of shading on plant development, yield and fruit quality of sweet pepper grown under conditions of high temperature and radlation. *Scientia horticulturae*, 29(1-2), 31-35.
- Sabu, M. (2003). Revision of the genus Zingiber in South India. Folia Malaysiana Vol, 4. 1.
- Sanewski, G., Fukai, S., & Giles, J. (1996). Shoot emergence of ginger (*Zingiber officinale* Rosc.) as affected by time of lifting, storage, size, and type of planting pieces. *Tropical Agriculture*.
- Shabir, G. A. (2004). A practical approach to validation of HPLC methods under current good manufacturing practices. *Journal of validation technology*, 10, 210-218.
- Shabir, G. A. (2005). Step-by-step analytical methods validation and protocol in the quality system compliance industry. *Journal of validation technology*, *10*, 314-325.
- Shen, S. C., Wu, M. S., Lin, H. Y., Yang, L. Y., Chen, Y. H., & Chen, Y. C. (2014). Reactive Oxygen Species- Dependent Nitric Oxide Production in Reciprocal Interactions of Glioma and Microglial Cells. *Journal of cellular physiology*, 229(12), 2015-2026.

- Sidahmed, H. M. A., Hashim, N. M., Abdulla, M. A., Ali, H. M., Mohan, S., Abdelwahab, S. I., . . . Vadivelu, J. (2015). Antisecretory, gastroprotective, antioxidant and anti-helicobeter pylori activity of zerumbone from Zingiber Zerumbet (L.) Smith. *PloS one*, *10*(3), e0121060.
- Simon, J., Gleadow, R. M., & Woodrow, I. E. (2010). Allocation of nitrogen to chemical defence and plant functional traits is constrained by soil N. *Tree physiology*, 30(9), 1111-1117.
- Singh, A., & Neopaney, B. (1993). Effect of NPK nulrition and spacing on yield and its attributes in ginger. *Haryana Journal of Horticultural sciences*, 22, 143-143.
- Singh, C. B., Nongalleima, K., Brojendrosingh, S., Ningombam, S., Lokendrajit, N., & Singh, L. (2012). Biological and chemical properties of *Zingiber zerumbet* Smith: a review. *Phytochemistry reviews*, 11(1), 113-125.
- Singh, M., Khan, M. M. A., & Naeem, M. (2016). Effect of nitrogen on growth, nutrient assimilation, essential oil content, yield and quality attributes in Zingiber officinale Rosc. Journal of the Saudi Society of Agricultural Sciences, 15(2), 171-178.
- Sirirugsa, P. (1999). Thai Zingiberaceae: species diversity and their uses. *Pure Appl. Chem*, 70, 1-8.
- Smith, J. A., Das, A., Ray, S. K., & Banik, N. L. (2012). Role of pro-inflammatory cytokines released from microglia in neurodegenerative diseases. *Brain research bulletin*, 87(1), 10-20.
- Smith, S., & Hadley, P. (1989). A comparison of organic and inorganic nitrogen fertilizers: Their nitrate-N and ammonium-N release characteristics and effects on the growth response of lettuce (*Lactuca sativa* L. cv. Fortune). *Plant and soil*, 115(1), 135-144.
- Soeparjono, S. (2016). The effect of media composition and organic fertilizer concentration on the growth and yield of red ginger rhizome (*Zingiber officinale* Rosc.). *Agriculture and Agricultural Science Procedia*, 9, 450-455.
- Spencer, A., Hamill, J. D., & Rhodes, M. J. (1993). In vitro biosynthesis of monoterpenes by Agrobacterium transformed shoot cultures of two Mentha species. *Phytochemistry*, 32(4), 911-919.
- Sreekala, G., & Jayachandran, B. (2002). Influence of shade regimes on the physiological parameters of ginger. *Journal of Spices and Aromatic Crops*, 11(1), 30-34.
- Srikrishnah, S., Peiris, S. E., & Sutharsan, S. (2012). Effect of shade levels on leaf area and biomass production of three varieties of *Dracaena sanderiana* L. in the dry zone of Sri Lanka.

- Srikrishnah, S., & Sutharsan, S. (2015). Effect of different shade levels on growth and tuber yield of turmeric (*Curcuma longa* L.) in the Batticaloa District of Sri Lanka. *Am. Euras. J. Agric. Agric. Environ. Sci*, 15.
- Srivastava, A., Srivastava, S., & Shah, N. (2000). Essential oil composition of *Zingiber zerumbet* (L.) Sm. from India. *Journal of Essential Oil Research*, 12(5), 595-597.
- Stansley, B., Post, J., & Hensley, K. (2012). A comparative review of cell culture systems for the study of microglial biology in Alzheimer's disease. *Journal of neuroinflammation*, 9(1), 115.
- Sulaiman, M. R., Mohamad, T. A. S. T., Mossadeq, W. M. S., Moin, S., Yusof, M., Mokhtar, A. F., . . . Lajis, N. (2010). Antinociceptive activity of the essential oil of *Zingiber zerumbet*. *Planta Medica*, 76(02), 107-112.
- Sung, B., Jhurani, S., Ahn, K. S., Mastuo, Y., Yi, T., Guha, S., . . . Aggarwal, B. B. (2008). Zerumbone down-regulates chemokine receptor CXCR4 expression leading to inhibition of CXCL12-induced invasion of breast and pancreatic tumor cells. *Cancer Research*, 68(21), 8938-8944.
- Swarnathilaka, D., Kottearachchi, N., & Weerakkody, W. (2016). Factors Affecting on Induction of Microrhizomes in Ginger (*Zingiber officinale* Rosc.), Cultivar Local from Sri Lanka. *British Biotechnology Journal*, 12(2), 1.
- Szabolcs, A., Tiszlavicz, L., Kaszaki, J., Pósa, A., Berkó, A., Varga, I. S., . . . Takács, T. (2007). Zerumbone exerts a beneficial effect on inflammatory parameters of cholecystokinin octapeptide-induced experimental pancreatitis but fails to improve histology. *Pancreas*, 35(3), 249-255.
- Taha, M. M. E., Abdul, A. B., Abdullah, R., Ibrahim, T. A. T., Abdelwahab, S. I., & Mohan, S. (2010). Potential chemoprevention of diethylnitrosamine-initiated and 2-acetylaminofluorene-promoted hepatocarcinogenesis by zerumbone from the rhizomes of the subtropical ginger (*Zingiber zerumbet*). *Chemico-Biological Interactions*, 186(3), 295-305.
- Tanaka, T., Shimizu, M., Kohno, H., Yoshitani, S.-i., Tsukio, Y., Murakami, A., . . . Koshimizu, K. (2001). Chemoprevention of azoxymethane-induced rat aberrant crypt foci by dietary zerumbone isolated from *Zingiber zerumbet*. *Life sciences*, 69(16), 1935-1945.
- Thameem Dheen, S., Kaur, C., & Ling, E.-A. (2007). Microglial activation and its implications in the brain diseases. *Current medicinal chemistry*, *14*(11), 1189-1197.
- Thohirah, L., Flora, C., & Kamalakshi, N. (2010). Breaking bud dormancy and different shade levels for production of pot and cut *Cucurma alismatifolia*. *American Journal of Agricultural and Biological Sciences*, 5(3), 285-388.

- Tsai, H.-H., Lee, W.-R., Wang, P.-H., Cheng, K.-T., Chen, Y.-C., & Shen, S.-C. (2013). Propionibacterium acnes-induced iNOS and COX-2 protein expression via ROS-dependent NF-κB and AP-1 activation in macrophages. *Journal of dermatological science*, 69(2), 122-131.
- Varier, N. (1945). *Chemical examination of the rhizomes of Zingiber zerumbet, Smith.*Paper presented at the Proceedings of the Indian Academy of Sciences-Section A.
- Vasant More, S., Kumar, H., Kim, I.-S., Koppulla, S., Kim, B.-W., & Choi, D.-K. (2013). Strategic selection of neuroinflammatory models in Parkinson's disease: evidence from experimental studies. CNS & Neurological Disorders-Drug Targets (Formerly Current Drug Targets-CNS & Neurological Disorders), 12(5), 680-697.
- Wolf, B. (1982). A comprehensive system of leaf analyses and its use for diagnosing crop nutrient status. *Communications in Soil Science and Plant Analysis*, 13(12), 1035-1059.
- Wolfe, J. T., Ross, D., & Cohen, G. M. (1994). A role for metals and free radicals in the induction of apoptosis in thymocytes. *FEBS letters*, 352(1), 58-62.
- Wolff, X. Y., & Coltman, R. R. (1990). Productivity of eight leafy vegetable crops grown under shade in Hawaii. *Journal of the American Society for Horticultural Science*, 115(1), 182-188.
- Xian, M., Ito, K., Nakazato, T., Shimizu, T., Chen, C. K., Yamato, K., . . . Kizaki, M. (2007). Zerumbone, a bioactive sesquiterpene, induces G2/M cell cycle arrest and apoptosis in leukemia cells via a Fas- and mitochondria- mediated pathway. *Cancer science*, 98(1), 118-126.
- Xie, B., & Wang, H. (2006). Effects of light spectrum and photoperiod on contents of flavonoid and terpene in leaves of *Ginkgo biloba L. Journal of Nanjing Forestry University*, 30, 51-54.
- Yeh, W. L., Lu, D. Y., Liou, H. C., & Fu, W. M. (2012). A forward loop between glioma and microglia: Glioma- derived extracellular matrix- activated microglia secrete IL- 18 to enhance the migration of glioma cells. *Journal of cellular physiology*, 227(2), 558-568.
- Yob, N., Jofrry, S. M., Affandi, M., Teh, L., Salleh, M., & Zakaria, Z. (2011). *Zingiber zerumbet* (L.) Smith: a review of its ethnomedicinal, chemical, and pharmacological uses. *Evidence-Based Complementary and Alternative Medicine*, 2011.
- Zakaria, Z., Mohamad, A., Chear, C., Wong, Y., Israf, D., & Sulaiman, M. (2010). Antiinflammatory and antinociceptive activities of *Zingiber zerumbet* methanol extract in experimental model systems. *Medical principles and practice*, 19(4), 287-294.

Zavala, J., & Ravetta, D. (2001). Allocation of photoassimilates to biomass, resin and carbohydrates in *Grindelia chiloensis* as affected by light intensity. *Field Crops Research*, 69(2), 143-149.



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## LIST OF PUBLICATIONS

- Thohirah, L.A., Goh, S., Hassan, S.A. and Stanslas, J. 2016. Pre-soaked technique using BAP and Ethephon in breaking bud dormancy of Lempoyang (*Zingiber zerumbet* (L.) Smith). 26<sup>th</sup> Malaysian Society of Plant Physiology Conference, MSPPC, 9-11<sup>th</sup> August 2016.
- Thohirah, L.A., Goh, S., Hassan, S.A. and Stanslas, J. 2016. Effects of shade and NPK fertilizer on rhizome yield in Lempoyang (*Zingiber zerumbet* (L.) Smith). International Agriculture Congress (IAC) 2016, 4-6<sup>th</sup> October 2016.
- Thohirah, L.A., Goh, S., Stanslas, J. and Hassan, S.A. 2017. Inhibitory effect of Zerumbone and *Zingiber zerumbet* Rhizome extract on nitric oxide production in lipopolysaccharide-stimulated BV2 microglial cells. International Conference on Scientific Frontier in Natural Product Based Drugs, 6-7 July 2017, Singapore.
- Goh, S., Thohirah, L.A., Hassan, S.A. and Stanslas, J. 2018. Breaking dormancy and the effect of shade level and NPK fertilizer rates on the yield of *Zingiber zerumbet* (L.) Smith (Lempoyang). Journal of Agriculture (Published).



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#### TITLE OF THESIS:

# IMPROVED PROPAGATION, MAXIMIZING YIELD AND ENHANCING

# ZERUMBONE PRODUCTION IN ZINGIBER ZERUMBET (L.) Smith

## THROUGH SHADING AND NPK FERTILIZATION

# NAME OF STUDENT: GOH SUZANNE

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