

# **UNIVERSITI PUTRA MALAYSIA**

# SEQUENCE VARIATION ANALYSIS AND CHARACTERIZATION OF DEFENCE AND STRESS-RELATED GENES IN Coconut Cadang-Cadang Viroid INOCULATED OIL PALM SEEDLINGS

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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

November 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

### SEQUENCE VARIATION ANALYSIS AND CHARACTERIZATION OF DEFENCE AND STRESS-RELATED GENES IN Coconut Cadang-Cadang Viroid INOCULATED OIL PALM SEEDLINGS

By

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December 2018

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Coconut cadang-cadang viroid (CCCVd) is associated with an orange spotting (OS) disorder, which is an emerging problem recently reported affecting oil palms in Malaysia. CCCVd variants were shown to contain low concentrations of several sequence variants of CCCVd in both OS and asymptomatic palms. Knowledge of interaction between CCCVd-infected oil palm is lacking and no study on defence and stress genes upon infection of CCCVd to date. Plant response has become an important approach to understand the host-pathogen interaction. The study was undertaken to study sequence variation of CCCVd oil palm variants from inoculated oil palm seedlings using RT-PCR, cloning and sequencing. Subsequently, detection and characterization of defence and stress genes in CCCVd inoculated oil palm seedlings were investigated. A total of 30 oil palm seedlings comprising ten healthy seedlings, ten seedlings inoculated with plasmid of CCCVd<sub>2460P</sub> and CCCVd<sub>2930P</sub> were used. Total nucleic acid was extracted using modified NETME extraction method. The presence of CCCVd variants were detected by RT-PCR using CCCVd specific primers, cloning and sequencing. For genes study, ten seedlings inoculated with  $CCCVd_{293OP}$  and ten seedlings as control were used to express total of 11 oil palm genes consisting of 4 stress, 4 defence and 3 reference genes. Optimization using gradient PCR and validation of genes were carried out for real-time PCR reaction. The detection of genes were preceded with quantitative real-time PCR using SYBR Green-I. For sequence variation study, RT-PCR analysis showed that all seedlings inoculated with plasmid CCCVd<sub>2460P</sub> and CCCVd<sub>2930P</sub> were detected and generated amplicons at approximately 300 nt on 2% agarose gel electrophoresis. There was no OS symptoms observed for both CCCVd variants. Results of sequencing showed that none of the clones of CCCVd<sub>2460P</sub> were positive with CCCVd, whereas seedlings inoculated with CCCVd<sub>293OP</sub> had 99-100% sequence similarity to CCCVd variant 293 nt (DO097184). Five clones had two to three base substitutions in their sequence compared with the consensus sequence of CCCVd<sub>293OP</sub> which indicate that there were minor sequence variation. However, there are no quasispecies occurs in this study. For genes study, stress and defence genes in CCCVd-inoculated oil palm seedlings were successfully detected and characterized using conventional PCR and real-time PCR.

Real-time PCR was found to be more sensitive than conventional PCR. The validation of genes through DNA sequencing was successful as the sequencing results showed 86-100% sequence similarity to respective genes. The study showed that all the genes involved were significantly different with the control seedlings. Stress and defence genes were significantly different with the time of post-inoculation. In conclusion, there are minor sequence variation present in oil palm seedlings inoculated with CCCVd<sub>2930P</sub> and no quasispecies observed. Defence and stress-related genes were successfully detected and characterized through real-time PCR and conventional PCR in oil palm seedlings inoculated with CCCVd<sub>2930P</sub> plasmid.



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### ANALISIS VARIASI JUJUKAN DAN PENCIRIAN GEN BERKAITAN PERTAHANAN DAN STRES DALAM ANAK-ANAK BENIH KELAPA SAWIT YANG DI INOKULASI Coconut Cadang-Cadang Viroid

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Coconut cadang-cadang viroid (CCCVd) dikaitkan dengan penyakit orange spotting (OS) kelapa sawit, yang mana akhir-akhir ini muncul penyakit OS dan dilaporkan menjejaskan pokok-pokok kelapa sawit di Malaysia. Varian-varian CCCVd menunjukkan bahawa beberapa yarian jujukan CCCVd mempunyai kepekatan yang rendah di dalam OS dan pokok kelapa sawit yang tidak bersimptom. Pengetahuan antara interaksi kelapa sawit yang dijangkiti CCCVd adalah kurang dan tiada kajian sehingga kini mengenai gen pertahanan dan gen stres ketika jangkitan CCCVd. Tindak balas tumbuhan terhadap penyakit telah menjadi pendekatan penting untuk memahami interaksi antara hos-patogen. Kajian ini dilaksanakan untuk mengkaji variasi jujukan CCCVd daripada pokok kelapa sawit yang telah di inokulasi menggunakan ujian RT-PCR, pengklonan dan penjujukan. Selepas itu, pengesanan dan pencirian gen pertahanan dan gen stres dalam anak-anak benih kelapa sawit yang di inokulasi CCCVd di siasat. Sebanyak 30 anak benih kelapa sawit yang terdiri daripada sepuluh anak-anak benih sihat, sepuluh anak benih yang di inokulasi dengan plasmid CCCVd<sub>2460P</sub> dan CCCVd<sub>2930P</sub> telah digunakan. Jumlah asid nukleik di ekstrak menggunakan kaedah pengekstrakan NETME. Kehadiran varian-varian CCCVd di kesan melalui ujian RT-PCR menggunakan primer-primer spesifik CCCVd, pengklonan dan penjujukan. Bagi kajian gen-gen, sepuluh anak benih yang di inokulasi CCCVd<sub>2930P</sub> dan sepuluh anak benih sebagai kawalan digunakan untuk mengekspres sebanyak 11 gen-gen pokok kelapa sawit vang terdiri daripada 4 gen stres, 4 gen pertahanan dan 3 gen rujukan. Pengoptimuman menggunakan gradient PCR dan pengesahan gen-gen dijalankan untuk tindakbalas real-time PCR. Pengesanan gen-gen di dahului dengan kuantitatif real-time PCR menggunakan SYBR Green-I. Bagi kajian variasi jujukan, analisis RT-PCR menunjukkan kesemua anak-anak benih yang di inokulasi dengan plasmid CCCVd<sub>2460P</sub> and CCCVd<sub>293OP</sub> dikesan dan menghasilkan amplikon anggaran 300 nt pada 2% gel agaros elektroforesis. Tiada simptom OS diperhatikan untuk kedua-dua varian CCCVd. Keputusan penjujukan menunjukkan tiada klon CCCVd<sub>2460P</sub> yang positif dengan CCCVd, sedangkan anak-anak benih yang di inokulasi dengan CCCVd<sub>293OP</sub> mempunyai

99-100% persamaan jujukan dengan variann CCCVd 293 nt (DQ097184). Lima klon yang mempunyai dua hingga tiga penggantian dalam jujukannya dibandingkan dengan jujukan konsensus CCCVd<sub>293OP</sub> menunjukkan bahawa terdapat jujukan variasi kecil. Walau bagaimanapun, tiada *quasispecies* berlaku dalam kajian ini. Bagi kajian gen, gen pertahanan dan gen stres dalam anak-anak benih kelapa sawit yang di inokulasi CCCVd telah berjaya di kesan dan dicirikan menggunakan PCR konvensional dan real-time PCR. Real-time PCR didapati lebih sensitif berbanding PCR konvensional. Pengesahan gengen melalui penjujukan DNA telah berjaya kerana keputusan penjujukan menunjukkan 86-100% persamaan jujukan terhadap gen masing-masing. Kajian menunjukkan bahawa kesemua gen-gen yang terlibat berbeza dengan ketara terhadap anak-anak benih kawalan. Gen stress dan gen pertahanan juga menunjukkan perbezaan yang ketara terhadap masa selepas inokulasi. Kesimpulannya, terdapat sedikit variasi jujukan dalam anak-anak pokok yang di inokulasikan dengan CCCVd<sub>2930P</sub> dan tiada quasispecies diperhatikan. Gen stress dan gen pertahanan berjaya di kesan dan dicirikan menggunakan real-time PCR dan konvensional PCR dalam anak-anak pokok kelapa sawit yang di inokulasikan dengan plasmid CCCVd<sub>293OP</sub>.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of Supervisory Committee were as follows:

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# LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
AMV-RT	Avian myeloblastosis virus reverse transcriptase
ASBVd	Avocado sunblotch viroid
bp	Base pair
BLAST	Basic local alignment search tool
С	Central
CA	Chloroform: iso-amyl alcohol mix
CCCVd	Coconut cadang-cadang viroid
CCR	Conserved central region
CTiVd	Coconut tinangaja viroid
CEVd	Citrus exocortis viroid
cDNA	Complementary deoxyribonucleic acid
Cq	Quantification cycle
CRD	Completely randomized design
dATP	Deoxyadenosine triphosphate
dCTP	Deoxycytidine triphosphate
dGTP	Deoxyguanosine triphosphate
dNTP	Mixture of deoxynucleoside-triphosphates in equimolar
	amounts
dTTP	Deoxythymidine triphosphate
ddH2O	Double distilled water
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacetic acid
EG5	Elaeis guineensis genome
EtBr	Ethidium bromide
EtOH	Ethanol
GC	Guanine-cytosine
GOS	Genetic orange spotting'
g	Gram
g	Centrifugal force
ha	Hectare
HCl	Hydrochloric acid
L	Litre
LB	Lysogeny broth
LiCl	Lithium chloride
М	Moles/ liter (Molarity)
М	Marker of DNA ladder
MnT	Million tonnes
MgCl2	Magnesium chloride
mRNA	Messenger RNA
min	Minute
m-	Milli- (10-3)
ml	Milliliter
mM	Millimolar
mg	Milligram
mpi	Month post infection

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μ-	Mic	ro- (10-6)
μl	Mici	roliter
µM	Mic	romolar
NETN	ME Natr	ium Chloride EDTA Tris-HCL Mercaptoethanol
NGS	Nex	t-Generation Sequencing
NCBI	I Nati	onal Center for Biotechnology Information
NaCl	Sodi	ium chloride
Na2F	DTA di-se	adium ethylenediamine tetra acetic acid
NTC	Non	-template control
nt	Nuc	leatides
n.	Nan	$a_{\rm c}$ (10.0)
11- ng	Nan	ogram
	Orar	age spotting
03 nmol	Dia	ige spotting
ршог	PICO	
P	Paul	logenicity
PAG	E Poly	
PCA	Pher	nol: chlorotorm: iso-amyl alconol mix
PCR	Poly	merase chain reaction
PSIV	d Pota	to spindle tuber viroid
qPCR	Real	-time quantitative polymerase chain reaction
RNA	Ribo	onucleic acid
RNAS	sin RNa	ise inhibitor
RT-P	CR Reve	erse transcriptase polymerase chain reaction
rpm	Reve	olutions per minute
rRNA	A Ribo	osomal RNA
SDS	Sodi	ium dodecyl sulfate
SOC	Supe	er optimal broth
S	seco	nd
TBE	Tris	-borate EDTA
ТСН	Terr	ninal conserved hairpin
TCR	Terr	ninal conserved region
Tris	Tris	(hydroxymethyl) aminomethane
Tris-F	HCl Tris	(hydroxymethyl) aminomethane hydrochloride
TuM	V Turr	nin mosaic virus
TL	Terr	nipal left
TR	Terr	ninal right
UV	Ultr	aviolet
	Unit	
ů V	Vari	able
V	Volt	age
vol	volu	ime
	Volu	ume per volume
w/v	Wei	ght per volume
X-gal	5-br	omo-4-chloro-3-Indolyl –D-galactosidase
	0.01	



#### **CHAPTER 1**

#### INTRODUCTION

The oil palm tree (*Elaeis guineensis Jacq.*) originated from West Africa and established in Malaysia in early 1870's and now, oil palm tree has been developed into an agricultural crop (MPOC, 2012a). Oil palm is the most productive oil-bearing crop as Malaysia supplies 10% of the world market oils and fats by utilizing only 4 million hectare of land (Casson, 2003).

The oil palm production is affected by pests and diseases. The most severe disease affects palms in Indonesia and Malaysia is basal stem rot (BSR) disease which is caused by *Ganoderma boninense* (MPOC, 2012b). In addition, threats such as an orange spotting (OS) disorder caused by variants of *Coconut cadang-cadang viroid* (CCCVd) have been reported affecting oil palms in Malaysia (Vadamalai, 2005; Vadamalai *et al.*, 2006).

CCCVd is the causal agent of the lethal cadang-cadang disease of coconut palms (*Cocos nucifera* L.) in the Philippines, which have been estimated to kill over 40 million coconut palms when it was first recognized (Zelazny *et al.*, 1982; Hanold & Randles, 1991; Randles & Rodriguez, 2003). Although OS was recognized as a disorder in the early 19th century in West Africa, but only recently it was associated with variants of CCCVd (Hanold & Randles, 1991; Vadamalai *et al.*, 2006). Since OS is associated with CCCVd, it is crucial to determine the extent of similarity between CCCVd and the CCCVd-like molecules in oil palm with OS (Vadamalai, 2005; Vadamalai *et al.*, 2006, Wu *et al.*, 2013). However, several aspects of its epidemiology in both coconut and oil palm still demand more researches and the principal mode of natural spread of CCCVd is still unknown (Randles & Rodriguez, 2003; Vadamalai *et al.*, 2009).

CCCVd variants associated with OS in oil palm in Malaysia have been reported to propagate in their hosts as populations of closely related sequence variants (quasi-species) (Flores *et al.*, 2005a). Nevertheless, there is lack of study on sequence variation within the host induced by plasmid. The distribution, accumulation and translocation of CCCVd within the oil palm has been studied previously (Thanarajoo, 2014). However, there is still lack of knowledge in the epidemiological aspects and mainly in the host pathogen interaction. In oil palm, several sequence variants of CCCVd reported to be present in low concentration and difficult to detect (Vadamalai, 2005; Mohammadi *et al.*, 2010). Previous study on CCCVd include reverse-transcription polymerase chain reaction (RT-PCR), hybridization assay, polyacrylamide gel electrophoresis (PAGE), ribonuclease protection assay (RPA), reverse transcription loop-mediated isothermal amplification (RT-LAMP), real-time PCR (qPCR) and sequencing (Vadamalai, 2005; Vadamalai *et al.*, 2006; Vadamalai *et al.*, 2009; Cheong, 2012; Joseph, 2012; Wu *et al.*, 2013; Thanarajoo, 2014; Thanarajoo *et al.*, 2014).

Plant response has become a decisive way to gain knowledge of host-pathogen interaction. The interactions of plants and microbial pathogens are among the most

complex phenomena in nature. Relative quantification was used to compare the level of gene expression of a particular gene of interest in a chemically-treated sample relative to the level of gene expression in an untreated sample. The changes are either; up-regulated to strengthen the host defence against pathogen invasion, or down-regulated due to the suppression of the host defence system by the pathogen (Applied Biosytem, 2010). Gene expression studies have been used in virus or viroid-host interaction such as in Turnip mosaic virus (TuMV) (Yang *et al.*, 2007) and *Citrus exocortis viroid* (CEVd) (Vidal *et al.*, 2003). The expression of genes involved in defence and stress responses were widely studied in oil palm infected by *Ganoderma boninense* but no study on genes involved in oil palm infected by CCCVd to date. Therefore, there is an inadequate information and lack of study on host-pathogen interaction of oil palm and CCCVd. Real-time PCR (qPCR) is used due to its highly sensitivity, able to quantify certain transcripts and changes in gene expression as well as produces reliable and rapid quantification results (Pfaffl, 2001).

In view of this, the objectives of this study are:

1. To study sequence variation of CCCVd oil palm variants from inoculated oil palm seedlings using RT-PCR, cloning and sequencing.

2. To detect and characterize defence and stress-related genes in CCCVd inoculated oil palm seedlings.

#### REFERENCES

Agrios, G.N. (2005). Plant Pathology 5th Edition. (pp. 903). *Elsevier Academic Press*. San Diego, California, New York: Academic Press

Applied Biosystem (2010). Introduction to Gene Expression Getting Started Guide.

- Aranda M.A., Fraile, A., Garcia-Arenal F., & Malpica, J.M. (1995). Experimental evaluation of the ribonuclease protection assay method for the assessment of genetic heterogeneity in populations of RNA viruses. *Archives of Virology*, 140, 1373-1383.
- Azni, I. N. A. M., Namasivayam, P., Ling, H. C., Alwee, S. S. R. S., & Manaf, M. A. A. (2014). Differentially expressed transcripts related to height in oil palm. *Journal of Oil Palm Research*, 26(4), 308-316.

Bio-Rad Laboratories (2006). Real-Time PCR Applications Guide.

- Boonham, N., Pérez, L. G., Mendez, M. S., Peralta, E. L., Blockley, A., Walsh, K., Barker, I. & Mumford, R. A. (2004). Development of a real-time RT-PCR assay for the detection of *Potato spindle tuber viroid. Journal of Virological Methods*, 116(2), 139-146.
- Branch, A. D., & Robertson, H. D. (1984). A replication cycle for viroids and other small infectious RNA's. *Science*, 223(4635), 450-455.
- Casson, A. (2003). Oil Palm, Soybeans & Critical Habitat Loss: a review prepared for the WWF Forest Conversion Initiative. *World Wildlife Fund*, Switzerland.
- Chan, P. L., Rose, R. J., Murad, A. M. A., Zainal, Z., Low, E. T. L., Ooi, L. C. L., Oii, S.E., Yahya, S., & Singh, R. (2014). Evaluation of reference genes for quantitative real-time PCR in oil palm elite planting materials propagated by tissue culture. *PloS one*, 9(6), e99774.
- Cheong, L. C. (2012). Incidence of orange spotting and characterization of *Coconut Cadang-Cadang Viroid* variants in Selangor and Sabah oil palm plantations, Malaysia. MSc. Thesis, Universiti Putra Malaysia.
- Chervoneva, I., Li, Y., Schulz, S., Croker, S., Wilson, C., Waldman, S. A., & Hyslop, T. (2010). Selection of optimal reference genes for normalization in quantitative RT-PCR. *BMC bioinformatics*, 11(1), 253.
- Coulter, J. K. & Rosenquist, E. A. (1955). Mineral nutrition of the oil palm: A study of the chemical composition of the frond in relation to chlorosis and yield. *Malayan Agricultural Journal* 38: 214-236.
- Daròs, J. A., Elena, S. F., & Flores, R. (2006). Viroids: an Ariadne's thread into the RNA labyrinth. *EMBO reports*, 7(6), 593-598.

- Demidenko, N. V., Logacheva, M. D., & Penin, A. A. (2011). Selection and validation of reference genes for quantitative real-time PCR in buckwheat (*Fagopyrum esculentum*) based on transcriptome sequence data. *PloS one*, *6*(5), e19434.
- Diener T.O. (2003). Discovering viroids -a personal perspective. *Nature Reviews Microbiology* 1, 75-80.
- Fernandez-Delmond, I., Pierrugues, O., de Wispelaere, M., Guilbaud, L., Gaubert, S., Divéki, Z., Godon, C., Tepfer, M., & Jacquemond, M. (2004). A novel strategy for creating recombinant infectious RNA virus genomes. *Journal of* virological methods, 121(2), 247-257.
- Flores, R., Daròs, J. A., & Hernández, C. (2000). Avsunviroidae family: viroids containing hammerhead ribozymes. Advances in Virus Research, 55, 271-323.
- Flores, R., Hernández, C., Alba, A. E. M. D., Daròs, J. A., & Serio, F. D. (2005a). Viroids and viroid-host interactions. Annual Review of Phytopathology, 43:117-139.
- Flores, R., Randles, J.W., Bar-Joseph, M., Owens, R.A. & Diener, T.O. (2005b). Viroidae. In Virus Taxonomy, Eighth Report of the International Committee on Taxonomy of Viruses, 1145-59. (Eds.) M. A. M. C.M. Fauquet, J. Maniloff, U. Desselberger and A.L. Ball. London: Elsevier/Academic.
- Flores, R., Martelli, G. P., Shaw, M., Marra, R., & Mascia, T. (2009). The phytopathological and epidemiological features of viroids. *Journal of Plant Pathology*, *91*(4, Suplement), 4-19.
- Forde, S. C. M. & Leyritz, M. J. P. (1968). A study of confluent orange spotting of the oil palm in Nigeria. *Journal of the Nigerian Institute for Oil Palm Research* 4: 371-380.
- Fraga, D., Meulia, T., & Fenster, S. (2008). Real-time PCR. Current protocols essential laboratory techniques, (1), 10-3.
- Gadea, J., Mayda, M. E., Conejero, V., & Vera, P. (1996). Characterization of defenserelated genes ectopically expressed in viroid-infected tomato plants. *MPMI-Molecular Plant Microbe Interactions*, 9(5), 409-415.
- Gascon, J. P., & Meunier, J. (1979). Anomalies of genetic origin in the oil palm, *Elaeis Guineensis*. Description and results. *Oleagineux*, *34*(10), 437-445.
- Góra-Sochacka, A. (2004). Viroids: unusual small pathogenic RNAs. *Acta biochimica polonica*-english edition 51(3):587-607.
- Gross, H. J., Liebl, U., Alberty, H., Krupp, G., Domdey, H., Ramm, K., & Sänger, H.
  L. (1981). A severe and a mild *potato spindle tuber viroid* isolate differ in three nucleotide exchanges only. *Bioscience Reports*, 1(3), 235-241.
- Hadidi, A., & Yang, X. (1990). Detection of *pome fruit viroids* by enzymatic cDNA amplification. *Journal of virological Methods*, 30(3), 261-269.

- Hadidi, A., & Candresse, T. (2003). Polymerase chain reaction. Viroids, 115-122. Collingwood, Australia: CSIRO Publishing.
- Hadidi, A., Flores, R., Randles, J., & Semancik, J. (2003). Properties, detection, diseases and their control. In *Viroids*: Collingwood, Australia: CSIRO Publishing.
- Hajeri, S., Ramadugu, C., Manjunath, K., Ng, J., Lee, R., & Vidalakis, G. (2011). *In vivo* generated *Citrus exocortis viroid* progeny variants display a range of phenotypes with altered levels of replication, systemic accumulation and *Virology*, 417(2), 400-409.
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. In *Nucleic acids symposium series*, 41 (41), 95-98. Oxford University Press, London: Information Retrieval Ltd., c1979- c2000.
- Hanold, D. & Randles, J. W. (1991). Detection of *Coconut Cadang-Cadang viroid*-like sequences in oil and coconut palm and other monocotyledons in South-West Pacific. *Annals of Applied Biology* 118: 139-151.
- Hanold, D. (1993). Diagnostic methods applicable to viroids. *Diagnosis of Plant Virus Disease*, 296-313.
- Hanold, D., Semancik, J. S., & Owens, R. A. (2003). Polyacrylamide gel electrophoresis. *Viroids*, 95-102.
- Haseloff, J., Mohamed, N. A., & Symons, R. H. (1982). Viroid RNAs of *cadang cadang* disease of coconuts. *Nature*, 299(5881), 316.
- Heid, C. A., Stevens, J., Livak, K. J., & Williams, P. M. (1996). Real time quantitative PCR. *Genome research*, 6(10), 986-994.
- Ho, C. L., Kwan, Y. Y., Choi, M. C., Tee, S. S., Ng, W. H., Lim, K. A., Lee, Y.P., Ooi, S.E., Lee, W.W., Tee, J.M. & Tan, S. H. (2007). Analysis and functional annotation of expressed sequence tags (ESTs) from multiple tissues of oil palm (*Elaeis guineensis Jacq.*). *BMC genomics*, 8(1), 381.
- Ho, C. L., & Tan, Y. C. (2015). Molecular defense response of oil palm to *Ganoderma* infection. *Phytochemistry*, *114*, 168-177.
- Hodgson, R. A. J., Wall, G. C., & Randles, J. W. (1998). Specific identification of *coconut tinangaja viroid* for differential field diagnosis of viroids in coconut palm. *Phytopathology*, 88(8), 774-781.
- Imperial, J. S., Rodriguez, M. J. B., & Randles, J. W. (1981). Variation in the viroidlike RNA associated with cadang-cadang disease: Evidence for an increase in molecular weight with disease progress. *Journal of General Virology*, 56(1), 77-85.

- Imperial, J. S., & Rodriguez, M. J. (1983). Variation in the *coconut cadang-cadang* viroid: evidence for single-base additions with disease progress. *Philippine* Journal of Crop Science, 8(2), 87-91.
- Imperial, J S., Baustista, R. M. & Randles, J. W. (1985). Transmission of the *coconut cadang-cadang viroid* to six species of palm by inoculation with nucleic acid extracts. *Plant Pathology*, 34: 391–401.
- International Committee on Taxonomy of Viruses (ICTV) (2017). Retrieved 17 August 2017 from (http://www.ictvonline.org)
- Jain, M., Nijhawan, A., Tyagi, A. K., & Khurana, J. P. (2006). Validation of housekeeping genes as internal control for studying gene expression in rice by quantitative real-time PCR. *Biochemical and biophysical research communications*, 345(2), 646-651.
- Joseph, H. (2012). Characterization and pathogenicity of Coconut cadang-cadang viroid variants in oil palm (*Elaeis guineensis Jacq.*) seedlings. PhD Thesis, Universiti Putra Malaysia.
- Kamarudin, N., Seman, I. A., Moslim, R., & Mazmira, M. (2016). Current challenges on pests and diseases of oil palm cultivation. In *Proceedings of the SASS– DOA*, Kota Kinabalu, Sabah
- Kanavedee, R., Vadamalai, G., Lau, W. H., Roslan, N. D., & Sundram, S. (2017). Optimization of total nucleic acid extraction method for detecting *Coconut cadang-cadang viroid* variants in oil palm. *Australasian Plant Pathology*, 46(3), 235-237.
- Lin, C. Y., Wu, M. L., Shen, T. L., Yeh, H. H., & Hung, T. H. (2015). Multiplex detection, distribution, and genetic diversity of *Hop stunt viroid* and *Citrus exocortis viroid* infecting citrus in Taiwan. *Virology journal*, 12(1), 11.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the  $2-\Delta\Delta$ CT method. *Methods*, 25(4), 402-408.
- Luigi, M., & Faggioli, F. (2011). Development of quantitative real-time RT-PCR for the detection and quantification of *Peach latent mosaic viroid*. *European Journal of Plant Pathology*, 130(1), 109-116.
- Mackay, I. M., Arden, K. E., & Nitsche, A. (2002). Real-time PCR in virology. *Nucleic acids research*, 30(6), 1292-1305.
- Mackay, I. M. (2004). Real-time PCR in the microbiology laboratory. *Clinical Microbiology and Infection*, 10(3), 190-212.
- Malaysia palm oil board (MPOB) (2011). Retrieved 19 January 2018 from http://www.palmoilworld.org/about\_malaysian-industry.html

- Malaysian palm oil council (MPOC) (2012a). Retrieved 20 January 2018 from http://www.mpoc.org.my/The\_Oil\_Palm\_Tree.aspx
- Malaysian palm oil council (MPOC) (2012b). Retrieved 20 January 2018 from http://www.mpoc.org.my/Malaysian\_Palm\_Oil\_Industry.aspx
- Maramorosch, K. (1991). The threat of Cadang-cadang disease. *Principes*, 37(4): 187-196.
- Mayo, M. (2003). Base-pairing in RNA virus replication and host plant defence. *Microbiology Today*, 30(1), 12-13.
- Mohamed, N.A., Haseloff, J., Imperial, J.S. and Symons, R.H. (1982). Characterization of the different electrophoretic forms of the *cadang-cadang viroid. Journal of General Virology*, 63(1), 181-188.
- Mohammadi, M. R., Vadamalai, G., & Joseph, H. (2010). An optimized method for extraction and detection of *Coconut cadang-cadang viroid* (CCCVd) from oil palm. *Communications in agricultural and applied biological sciences*, 75(4), 777-781.
- Nagamine, K., Kuzuhara, Y., & Notomi, T. (2002). Isolation of single-stranded DNA from loop-mediated isothermal amplification products. *Biochemical and biophysical research communications*, 290(4), 1195-1198.
- Nie, X. (2005). Reverse transcription loop-mediated isothermal amplification of DNA for detection of Potato virus Y. *Plant Disease*, *89*(6), 605-610.
- Notomi, T., Okayama, H., Masubuchi, H., Yonekawa, T., Watanabe, K., Amino, N., & Hase, T. (2000). Loop-mediated isothermal amplification of DNA. *Nucleic acids research*, 28(12), e63-e63.
- Ooi SE, Choo CN, Ishak Z, Ong-Abdullah M (2012) A candidate auxin-responsive expression marker gene, EgIAA9, for somatic embryogenesis in oil palm (*Elaeis guineensis Jacq.*). *Plant Cell, Tissue and Organ Culture* (*PCTOC*), 110(2), 201-212.
- Owens, R. A., Chen, W., Hu, Y., & Hsu, Y. H. (1995). Suppression of *potato spindle tuber viroid* replication and symptom expression by mutations which stabilize the pathogenicity domain. *Virology*, 208(2), 554-564.
- Papayiannis, L. C. (2014). Diagnostic real-time RT-PCR for the simultaneous detection of *Citrus exocortis viroid* and *Hop stunt viroid*. *Journal of virological methods*, 196, 93-99.
- Peng, J., Fan, Z., & Huang, J. (2012). Rapid Detection of Banana Streak Virus by Loop-mediated Isothermal Amplification Assay in South China. *Journal of Phytopathology*, 160(5), 248-250.
- Pfaffl, M. W. (2001). A new mathematical model for relative quantification in realtime RT–PCR. *Nucleic acids research*, 29(9), e45.

- Puchta, H., Ramm, K., & Sänger, H. L. (1988). The molecular structure of *hop latent viroid* (HLV), a new viroid occurring worldwide in hops. *Nucleic acids research*, 16(10), 4197-4216.
- Randles, J. W., Boccardo, G., Retuerma, M. L., & Rillo, E. P. (1977). Transmission of the RNA species associated with *cadang-cadang* of coconut palm, and the insensitivity of the disease to antibiotics. *Phytopathology*, 67(10), 1211-1216.
- Randles, J.W. (1985). Coconut cadang-cadang viroid. In Subviral Pathogens of Plants and Animals: Viroids and Prions, pp. 39-74. (Eds.) K. Maramorosch, K. & McKelvey, J.J. Elsevier.
- Randles, J. W. (1987). Coconut Cadang-Cadang. (Eds.) Diener, T. O. The Viroids (265-277). New York: Plenum Press.
- Randles, J. W. (1998). CCCVd-related sequences in species other than coconut. (Eds.) Hanold D. & Randles J.W, Report on ACIAR-Funded Research on Viroids and Viruses of Coconut Palm and Other Monocotyledons 1985-1993. 51, 144-152. Canberra: Australia Centre for International Agricultural Research.
- Randles, J. W., Rodriguez, M. J. B., & Imperial, J. S. (1988). Cadang-cadang disease of coconut palm. *Microbiological sciences*, 5(1), 18-22.
- Randles, J. W., & Rodriguez, M. J. B. (2003). *Coconut cadang-cadang viroid. Viroids,* 1st ed. CSIRO Publishing, Victoria, 233-241.
- Rizza, S., Nobile, G., Tessitori, M., Catara, A., & Conte, E. (2009). Real time RT-PCR assay for quantitative detection of *Citrus viroid III* in plant tissues. *Plant Pathology*, *58*(1), 181-185.
- Rodriguez, M. J., & Randles, J. W. (1993). *Coconut cadang-cadang viroid* (CCCVd) mutants associated with severe disease vary in both the pathogenicity domain and the central conserved region. *Nucleic acids research*, 21(11), 2771.
- Sanger, H. L., Klotz, G., Riesner, D., Gross, H. J., & Kleinschmidt, A. K. (1976). Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures. In *Proceedings of the National Academy of Sciences*, 73(11), 3852-3856.
- Sano, T., Candresse, T., Hammond, R. W., Diener, T. O., & Owens, R. A. (1992). Identification of multiple structural domains regulating viroid pathogenicity. In *Proceedings of the National Academy of Sciences*, 89(21), 10104-10108.
- Sarawak Land Consolidation & Rehabilitation Authority (SALCRA) (2012). Pest and Disease Control. Retrieved 26 January 2018 from http://www.salcra.gov.my/en/sustainable-plantation/pest-diseases control.html
- Selvaraja, S., Balasundram, S. K., Vadamalai, G., & Husni, M. H. A. (2012). Spatial variability of orange spotting disease in oil palm. *Journal of Biological Sciences*, 12(4), 232-238.

- Singh, R.P., Ready, K.F.M., & Nie, X. (2003) Viroids: biology. Viroids, (Eds.) Hadidi, A., Flores, R., Randles, J.W., & Semancik, J.S. (pp. 30–48). Collinwood: CSIRO Publishing.
- Sullivan, M., Daniells, E., & Robinson, A. (2012). Pest Datasheet for *Coconut cadang-cadang viroid*, Purdue University, 1–4.
- Tan, Y. C., Wong, M. Y., & Ho, C. L. (2015). Expression profiles of defence related cDNAs in oil palm (*Elaeis guineensis Jacq.*) inoculated with mycorrhizae and *Trichoderma harzianum Rifai* T32. *Plant Physiology and Biochemistry*, 96, 296-300.
- Tan, Y. C., Yeoh, K. A., Wong, M. Y., & Ho, C. L. (2013). Expression profiles of putative defence-related proteins in oil palm (*Elaeis guineensis*) colonized by *Ganoderma boninense*. Journal of plant physiology, 170(16), 1455-1460.
- Tee, S. S. (2009). Transcriptional changes in response to single and combine inoculation of Mycorrhiza and *Ganoderma* in oil palm (*Elaeis guineensis Jacq.*) roots. PhD Thesis, Universiti Putra Malaysia.
- Thanarajoo, S. S., Kong, L. L., Kadir, J., Lau, W. H., & Vadamalai, G. (2014). Detection of *Coconut cadang-cadang viroid* (CCCVd) in oil palm by reverse transcription loop-mediated isothermal amplification (RT-LAMP). *Journal of virological methods*, 202, 19-23.
- Thanarajoo, S.S. (2014). Rapid detection, accumulation and translocation of *Coconut cadang-cadang viroid* variants in oil palm. PhD Thesis, Universiti Putra Malaysia.

ThermoFisher Scientific (2009). qPCR Optimization and Troubeshooting Guide.

- Tindall, K. R., & Kunkel, T. A. (1988). Fidelity of DNA synthesis by the *Thermus* aquaticus DNA polymerase. *Biochemistry*, 27(16), 6008-6013.
- Tomlinson, J. A., Boonham, N., & Dickinson, M. (2010). Development and evaluation of a one-hour DNA extraction and loop-mediated isothermal amplification assay for rapid detection of phytoplasmas. *Plant Pathology*, 59(3), 465-471.
- Tsagris, E. M., Martínez de Alba, Á. E., Gozmanova, M., & Kalantidis, K. (2008). Viroids. *Cellular microbiology*, 10(11), 2168-2179.
- Turner, P. D., & Bull, R. A. (1967). Diseases and Disorders of the Oil Palm in Malaysia. Diseases and Disorders of the Oil Palm in Malaysia.
- Vadamalai, G. (2005). An investigation of orange spotting disorder in oil palm. PhD Thesis, University of Adelaide.
- Vadamalai, G., Hanold, D., Rezaian, M. A. & Randles, J. W. (2006). Variants of Coconut cadang-cadang viroid isolated from an African oil palm (Elaies guineensis Jacq.) in Malaysia. Archives of virology. 151(7). 1447-1456.

- Vadamalai, G., Perera, A. A. F. L. K., Hanold, D., Rezaian, M. A., & Randles, J.W. (2009). Detection of *Coconut cadang-cadang viroid* sequences in oil and coconut palm by ribonuclease protection assay. *Annals of Applied Biology*, 154(1), 117-125.
- Vidal, A. M., Ben-Cheikh, W., Talón, M., & García-Martínez, J. L. (2003). Regulation of gibberellin 20-oxidase gene expression and gibberellin content in citrus by temperature and *citrus exocortis viroid*. *Planta*, 217(3), 442-448.
- Wassenegger, M., Spieker, R. L., Thalmeir, S., Gast, F. U., Riedel, L., & Sänger, H. L. (1996). A single nucleotide substitution converts *potato spindle tuber viroid* (PSTVd) from a noninfectious to an infectious RNA for *Nicotiana tabacum*. *Virology*, 226(2), 191-197.
- Waterston, J. M. (1953). Observations on the influence of some ecological factors on the incidence of oil palm diseases in Nigeria. *Journal of the West African Institute for Oil Palm Research*, 1, 24-59.
- Wu, Y. H., Cheong, L. C., Meon, S., Lau, W. H., Kong, L. L., Joseph, H., & Vadamalai, G. (2013). Characterization of *Coconut cadang-cadang viroid* variants from oil palm affected by orange spotting disease in Malaysia. *Archives of virology*, 158(6), 1407-1410.
- Yang, C., Guo, R., Jie, F., Nettleton, D., Peng, J., Carr, T., Yeakley, J.M., Fan, J.B. & Whitham, S. A. (2007). Spatial analysis of *Arabidopsis thaliana* gene expression in response to Turnip mosaic virus infection. *Molecular plantmicrobe interactions*, 20(4), 358-370.
- Yeap, W. C., Ooi, T. E. K., Namasivayam, P., Kulaveerasingam, H., & Ho, C. L. (2012). EgRBP42 encoding an hnRNP-like RNA-binding protein from *Elaeis guineensis Jacq*. is responsive to abiotic stresses. *Plant cell reports*, 31(10), 1829-1843.
- Yeoh, K. A., Othman, A., Meon, S., Abdullah, F., & Ho, C. L. (2013). Sequence analysis and gene expression of putative oil palm chitinase and chitinase-like proteins in response to colonization of *Ganoderma boninense* and *Trichoderma harzianum*. *Molecular biology reports*, 40(1), 147-158.
- Zelazny, B. (1980). Ecology of cadang-cadang disease of coconut palm in the Philippines. *Phytopathology*, 70(8), 700-703.
- Zelazny, B., Randles, J. W., Boccardo, G., & Imperial, J. S. (1982). The viroid nature of the cadang-cadang disease of coconut palm. *Scientia Filipinas* (Philippines).

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