

#### **UNIVERSITI PUTRA MALAYSIA**

### IMPROVEMENT IN ORGANOGENESIS AND THE DEVELOPMENT OF A TRANSFORMATION PROCEDURE FOR CUCUMBER AND MUSKMELON

A. K. M. MOHI UDDIN

**FSMB 1998 20** 



## IMPROVEMENT IN ORGANOGENESIS AND THE DEVELOPMENT OF A TRANSFORMATION PROCEDURE FOR CUCUMBER AND MUSKMELON

By

A. K. M. MOHI UDDIN

Dissertation Submitted in Fulfilment of the Requirements for the Degree of Doctor of Philosophy in the Faculty of Food Science and Biotechnology, Universiti Putra Malaysia.

December 1998



# Specially Dedicated Jo My

Father (Alhajj Md. Lutful Hoque)

Mother (Tahera Hoque)

Father-in-law (Md. Salimullah)

Mother-in-law (Masuma Salim)

and

Wife (Nazneen Salim)



#### **ACKNOWLEDGEMENTS**

First and foremost, my deepest thanks to **ALLAH** for He has guided me through and provided me wisdom, strength and comfort to complete the degree successfully.

I would like to extend my sincere gratitude to my supervisors, Dr. Suhaimi Napis, Dr. K. Harikrishna, Dr. Zaliha Christine Abdullah, Dr. M. Kamal Uddin Chowdhury and Dr. Tan Siang Hee for all their guidance, help, tutelage and invaluable advice during my Ph. D. project and the preparation as well as completion of this thesis. Their criticisms and suggestions have been most constructive and are highly appreciated. Their patience, trust and enthusiasm have left depth of feeling that could not be expressed in words. I express my sincere thanks to Professor Marziah Mahmood for her advice too.

I am profoundly indebted to my parents, father and mother-in-law, brothers, sisters and wife for their sacrifices and encouragement to do higher study in the field of Plant Biotechnology and would like to express my sincere thanks and deepest gratitude to them for their invaluable love and prayers throughout the years to complete my study. I dedicated this work to my parents, father and mother-in-law, with all my love. To my wife, whose love, help, understanding, and encouragement has been the biggest motivation in completing this degree, I dedicated this work to her, with all my love, too. My daughter, Ramisa also deserves appreciation for her patience and sacrifice. A special thanks to Professor Md. Sharif, Ex. Chairman and Head of Department of Botany, Jagannath University College, Bangladesh, Assistant Professor Dr. Md. Imdadul Hoque, Department of Botany, University of Dhaka, Bangladesh and Mrs. Nurjahan Begum, Lecturer, Department of Botany, Jagannath University College, Bangladesh for their valuable advice and help.



Accordingly, I would like to say thank to the Ministry of Science, Technology, and the Environment, Malaysia for financial support (Graduate Assistantship) (IRPA Grants, 50304 and 51267) which gave me the opportunity to pursue my Ph. D. degree in Malaysia. I also express my sincere thanks to the Government of The People's Republic of Bangladesh for providing me one way plane fare. Indeed, I wish to thank the Rubber Research Institute of Malaysia (RRIM) for providing *Agrobacterium* strains.

I would also like to thank my friends, entire staff of Faculty of Food Science and Biotechnology and Graduate School, Universiti Putra Malaysia and staffs of Tissue Culture and Genetic laboratory, Biotechnology Department for their friendship, invaluable help, and encouragement throughout my endeavour here.

I hope any who are not mentioned by name will recognise my gratitude for their kindness, advice, and moral support on completion of my degree at Universiti Putra Malaysia. To all others who have contributed as well as assisted me in providing different inputs in one way or another to the successful completion of my study throughout my student life, they are conferred my appreciation.



#### TABLE OF CONTENTS

ACKNOWLE	DGEMENTS
LIST OF TAE	BLES
LIST OF FIG	URES
	ATES
	_
	BREVIATION
ABSTRACT-	
ABSTRAK	
CHARTER	
CHAPTER	
1	GENERAL INTRODUCTION
	Plant Genetic Manipulation
	Study on Cucumber and Muskmelon
	Justification of the Study
	Objectives of the Study
II	LITERATURE REVIEW
	Study of Tissue Culture on Cucumber and Muskmelon
	Organogenesis
	Somatic Embryogenesis
	Effect of Hormone on Organogenesis
	Effect of Hormone on Somatic Embryogenesis
	Effect of Carbohydrates on Shoot Regeneration
	Physical State of the Media
	Effect of Physical Environment
	Somaclonal Variation in Cucumber
	Somaclonal Variation in Muskmelon
	Study on Ethylene Inhibitors AgNO <sub>3</sub> and CoCl <sub>2</sub>
	AgNO <sub>3</sub> and CoCl <sub>2</sub> Action on Cucumber
	AgNO <sub>3</sub> and CoCl <sub>2</sub> Action on Muskmelon
	Study on Infection of Oncogenic Agrobacterium
	Virulence Consideration
	Crown Gall and Hairy Root Induction
	Study on Plant Genetic Transformation Physical Methods for Plant Gene Transfer
	Agrobacterium-mediated Transformation
	Biology of Agrobacterium
	Recombinant Vector Plasmids
	Genes Required for Transformation Study
	Intron-containing Reporter Gene
	Identification of Transgenic Plants
	Genetic Selection



	Conformation of Transformation and Stability of DNA	32
III	OPTIMIZATION OF RAPID SHOOT ORGANOGENESIS IN CUCUMBER AND MUSKMELON	34
		J-7
	Preface	34
	Materials and Methods	35
	Seed Surface Sterilisation	35
	Seed Germination	35
	Explant Preparation	36
	ExperimentsCulture Conditions and Shoot Primordia Induction	36
		37 38
	Shoot Primordia ElongationStudies on Phenotypical Normal and Abnormal Shoots	
	Root Induction	39 40
	Dark Incubation	40
	Statistical Analysis	40
	Acclimatisation of Regenerants	41
	Results (Part I)	42
	Callus Induction	42
	Effect of BAP on Cucumber Shoot Primordia Induction	43
	Effect of BAP and 2,4-D on Cucumber Shoot	
	Primordia Induction	46
	Effect of BAP and NAA on Cucumber Shoot	50
	Primordia Induction	50
	Shoot Primordia Elongation	55
	Effect of Hormone on Phenotypically Normal Shoot Production	59
	Effect of Hormone on Phenotypically Abnormal Shoot	
	ProductionEffect of Auxin (IBA) on Root Induction	64
	Effect of Dark Incubation on Root Induction	67
		68
	Results (Part II)	74
	Effect of BAP on Muskmelon Shoot Primordia	74
	Shoot Primordia Elongation	76
	Effect of BAP and 2,4-D or NAA on Muskmelon	, 0
	Shoot Primordia Induction	80
	Shoot Primordia Elongation	85
	Phenotypical Normal Muskmelon Shoot Production Phenotypically Abnormal Shoot Production of	86
	Muskmelon	88
	Effect of Auxin on Muskmelon Root Induction	88
	Effect of Dark Incubation on Root Induction	90
	Acclimatisation of Cucumber and Muskmelon	
	Regenerants	92
	Discussion	96



IV	IMPROVEMENT OF CUCUMBER ORGANOGENESIS BY SILVER NITRATE (AGNO <sub>3</sub> ) AND COBALT CHLORIDE (COCI <sub>2</sub> )
	Preface 1
	Materials and Methods1
	Source of Explants1
	Preparation of Shoot Primordia Induction Medium 1
	Experiments1
	Results (Part I)1
	Effect of AgNO <sub>3</sub> on Cucumber Callus Induction1
	Effect of AgNO <sub>3</sub> with SGM on Cucumber Shoot Regeneration1
	Effect of AgNO <sub>3</sub> in SPI Medium on Cucumber
	Shoot Regeneration1
	Effect of AgNO <sub>3</sub> in SGM and SPI Medium on
	Cucumber Shoot Regeneration1 Shoot Primordia Elongation1
	Shoot Primordia Elongation 1 Effect of AgNO <sub>3</sub> on the Production of Normal Shoots-
	Root Induction 1
	Effect of AgNO <sub>3</sub> on Muskmelon Shoot Regeneration
	Discussion 1
	Results (Part II) 1
	Effect of CoCl <sub>2</sub> on Cucumber Callus Induction1
	Effect of CoCl₂ in SGM on Shoot Regeneration1
	Effect of CoCl₂ in SPI Medium on Cucumber Shoot
	Regeneration1
	Effect of CoCl₂ in SGM and SPI Medium on
	Cucumber Shoot Regeneration1
	Shoot Primordia Elongation
	Effect of CoCl <sub>2</sub> on Production of Morphologically
	Normal Shoots 1
	Root Induction 1
	Effect of CoCl <sub>2</sub> on Muskmelon Shoot Regeneration 1 Discussion
V	IDENTIFICATION OF SUSCEPTIBILITY OF DIFFERENT  AGROBACTERIUM STRAINS
	Park as
	Preface
	Materials and Methods
	Germination of Seedlings 1
	Medium for <i>Agrobacterium</i> Culture
	Experiments 1
	Statistical Analysis 1
	Results 1
	Crown-gall Induction in Cucumber and Muskmelon
	Plants



	Hairy Root Induction in Cucumber and Muskmelon
	Plants
	Effect of Dark Incubation on Crown Gall and Hairy
	Root Induction Effect of Light Intensity on Crown Gall and Hairy Root
	Induction Effect of Acetosyringone on Crown Gall
	InductionEffect of Acetosyringone to Hairy Root
	Production
	Discussion
/I	STUDIES ON GENETIC TRANSFORMATION MEDIATED BY AGROBACTERIUM TUMEFACIENS—
	Preface
	Materials and Methods
	Plant Materials
	Agrobacterium Strain and Vector Plasmid
	Agrobacterium Culture
	Feeder Layer Preparation
	Determination of Minimal Inhibitory Concentration
	(MIC) of Kanamycin
	Preparation of Regeneration Medium
	Experiments
	Statistical Analysis
	Results
	Role of Pre-culture Medium
	Effect of Co-cultivation Period on Cucumber and
	Muskmelon Shoot Regeneration
	Role of Feeder Layer on Cucumber and Muskmelon
	Shoot Regeneration
	Role of Acetosyringone on Cucumber and
	Muskmelon Shoot Regeneration
	Discussion
/II	ANALYSIS OF TRANSFORMANTS
	Preface
	Materials and Methods
	Analysis of GUS Enzyme Activity
	GUS Assay
	Components of GUS Extraction Buffer
	Genetic Analysis
	DNA Isolation from Plants and Agrobacterium
	Primer Selection
	Polymerase Chain Reaction and Gel
	Electrophoresis
	Southern Blotting
	Results
	Confirmation of Transformation Through GUS
	· · · · · · · · · · · · · · · · · ·



	Expression	198
	Effect of Feeder Layer on GUS Expression	198
	Effect of Acetosyringone on GUS Expression	198
	Confirmation of Transformation Through Genetic	130
		004
	Analysis	200
	PCR Analysis	200
	Southern Blot Analysis	200
	Discussion	202
VIII	GENERAL DISCUSSION AND CONCLUSION———	20
	Experimental Approach	001
	Overview on Cucumber and Muskmelon	20
		001
	Organogenesis	205
	Overview on the Improvement of Cucumber Shoot	
	Organogenesis by AgNO <sub>3</sub> and CoCl <sub>2</sub>	207
	Overview on Agrobacterium Susceptibility to	
	Cucumber and Muskmelon	207
	Overview on Cucumber and Muskmelon	201
		004
	Transformation and Confirmation Study	208
	Conclusion	210
	Strategies for Further Study	21
REFFERENCES		213
APPENDIX		227
A	Murashige and Skoog (MS) Medium	228
В	Tobacco Suspension Culture Medium and B5 Vitamins	229
С	LB and YMB Medium	230
Ď	Additional Tables	231
b	Additional Tables	23
VITA		242
	List of Publication in Refereed Journals	243
	List of Publication in Proceedings	243



#### LIST OF TABLES

of BAP on Multiple Shoot Primordia Induction from Place and Hypocotyls of SS, SC, SL, TL and TG cultiber	ivars of
nt Numbers of Shoots Induced from Explants of S and TG Cultivars at Different BAP Levels	
of BAP and 2,4-D on Shoot Primordia Induction its of SS, SC, SL, TL and TG Cultivars	
on on Numbers of Shoots Induced from Explants on the Cultivars at Different BAP and 2,4-D Combination	
ice of BAP and NAA on Shoot Primordia Inductionts of SS, SC, SL, TL and TG Cultivars	
s of BAP and NAA on Number of Shoot Primordia In explants of Cucumber Cultivars	
arison of Percentage of Elongated Shoots Derivents of Five Cucumber Cultivars on Elongation Medium	
s of BAP and Combinations of BAP with 2,4-D or N typically Normal Shoot Induction from Cucumber Exp	
s of IBA on Root Induction from Regenerated Shonber Cultivars, Namely SS, SC, SL, TL and TG	
ce of Dark Incubation on Root Induction from Non- s of Five Cucumber Cultivars SS, SC, SL, TL and TG-	
of BAP on Shoot Primordia Induction from Dats of Muskmelon	
on on Numbers of Shoot Primordia Induced from Dats of Muskmelon by BAP	
ce of BAP on Elongation of 10, 15, and 20-day-old dia, Derived from Explants of Muskmelon	
of BAP on Height of 10, 15, and 20-day-old Elos, Derived from Different Explants of Muskmelon	
nce of Combinations of BAP-2,4-D and BAP-Nanelon Shoot Primordia Induction	
ons on Number of Shoots Induced from Musl ts by Hormone Combinations of BAP+2,4-D NAA	and
of BAP on Elongation of 15-day-old Shoot Pried from Medium Containing BAP Either with 2,4-D or	
ice of BAP Added to Elongation Medium on Pheno Il Shoot Production from Muskmelon Explants ed on SPI Medium Containing BAP	Which



19 Effect of BAP on Production of Phenotypical Derived from Explants of Muskmelon on SPI Med BAP either with 2,4-D or NAA	lium Containing
20 Effect of NAA on Root Induction from Differe Muskmelon cv. Birdie	
21 Effect of Dark Incubation on Root Induction fro Shoots of Muskmelon	om Non-rooted
22 Variation on Rates of Cucumber Plantlets Acclimat Environment and Survived in Soil	
23 Variation on Rates of Muskmelon Plantlets A Ambient Environment and Survived in Soil	
Effect of AgNO <sub>3</sub> with SGM on Shoot Primordia Different Explants of SS	
.2 Influence of AgNO <sub>3</sub> with SGM on Shoot Primordia Different Explants of TG	
.3 Effect of AgNO <sub>3</sub> in SPI Medium on Shoot Reg Different Explants of SS	
.4 Influence of AgNO <sub>3</sub> in SPI Medium on Shoot Reg Explants of TG	
.5 Effect of AgNO <sub>3</sub> in SGM and SPI Medium on Shoo from Different explants of SS	
.6 Influence of AgNO <sub>3</sub> in the SGM and SPI Med Regeneration from Different explants of TG	
.7 Shoot Primordia Elongation Ability of Cucumber Different Explants of SS and TG Cultivars on MSO-	
.8 Influence of AgNO <sub>3</sub> on Production of Morpholo Cucumber Shoots	
.9 Rotting Ability of Cucumber Shoots Derived Explants of SS and TG Cultivars at 0.5 mg/l IBA	
10 Shoot Regeneration of Muskmelon cv. Birdie at D Concentration Levels	
11 Effect of CoCl <sub>2</sub> in SGM on Shoot Primordia Different Explants of SS	Induction from
12 Influence of CoCl <sub>2</sub> in SGM on Shoot Primordia Different Explants of TG	
13 Effect of CoCl <sub>2</sub> in SPI Medium on Shoot Primordia Different Explants of SS	
14 Influence of CoCl₂ in SPI Medium on Shoot Prim from Different Explants of TG	
15 Effect of CoCl₂ in Both SGM and SPI Medium on S Induction from Different Explants of SS	



4.16	Primordia Induction from Different Explants of TG
4.17	Variation on Cucumber Shoot Elongation Ability of Different Explants of SS and TG on MSO Medium
4.18	Effect of CoCl <sub>2</sub> on Production of Morphologically Normal Cucumber Shoots Derived from Different Explants of SS and TG Cultivars
4.19	Percentage of Rooting in Shoots Derived from Different Explants of SS and TG on Medium Containing IBA at 0.5 mg/l
4.20	Effect of CoCl <sub>2</sub> on shoot primordia induction from different explants of muskmelon cv. Birdie
5.1	Strain-dependent Variation in Numbers of Hairy Root Induction on Cucumber cv. SS and muskmelon cv. Birdie (BD)
6.1	Effect of Co-cultivation Period on Shoot Regeneration of Cucumber and Muskmelon
6.2	Effect of feeder layer Incubation on Cucumber and Muskmelon Shoot Regeneration
6.3	Effect of feeder layer on the Rates of Survival of Cucumber and Muskmelon Regenerants in Selection Medium
6.4	Role of Acetosyringone (20, 40, 100 µM) on Shoot Regeneration of Cucumber and Muskmelon Explants Cultured on MS+cefotaxime for 2-3 days
6.5	Effect of acetosyringone (20, 40, 100 μM) on Shoot Regeneration of Cucumber and Muskmelon Explants Cultured on MS+Cefotaxime+Kanamycin
6.6	Effect of Acetosyringone on the Rates of Cucumber and Muskmelon Survival Regenerants in Selection Medium
7.1	Effect of Feeder Layer Incubation on Variation of Numbers of Shoots Showed GUS Expression
7.2	Effect of Acetosyringone on Variation of Numbers of Shoots



#### Additional Table

1	Analysis of Variance for the Effect of Treatment (BAP Concentration) on Shoot Primordia Induction (Variable) from Different Cultivars of Cucumber	231
2	Analysis of Variance for the Effect of Treatment (BAP+2,4-D Concentration) on Shoot Primordia Induction (Variable) From Different Cultivars of Cucumber	231
3	Analysis of Variance for the Effect of Treatment (BAP+NAA Concentration) on Shoot Primordia Induction (Variable) From Different Cultivars of Cucumber	231
4	Analysis of Variance for the Effect of Treatment (MSO) on Shoot Primordia Elongation (Variable) of Cucumber	232
5	Analysis of Variance for the Effect of Treatment (Hormone) on Induction of Cucumber Phenotypically Normal Shoots (Variable)	232
6	Analysis of Variance for the Effect of Treatment (Hormone) on Root Induction (Variable) of Cucumber Regenerated Shoots	232
7	Analysis of Variance for the Effect of Treatment (Dark Incubation) on Root Induction (Variable) of Cucumber Regenerated Shoots	233
8	Analysis of Variance for the Effect of Treatment (hormone Concentration or combination) on Muskmelon Shoot Primordia Induction (Variable)	233
9	Analysis of Variance for the Effect of Treatment hormone Concentration or combination) on Elongation (Variable) of 10, 15 and 20-day-old Muskmelon Shoot Primordia	233
10	Analysis of Variance for the Effect of Treatment (hormone Concentration or combination) on Induction of Phenotypically Normal Muskmelon Shoots (Variable)	234
11	Analysis of Variance for the Effect of Treatment (hormone Concentration or combination) on Induction of Phenotypically Normal Muskmelon Shoots (Variable)	234
12	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ], Added to Seed Germination Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. SS	234
13	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ], Added to Seed Germination Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. TG	235
14	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ], Added to Regeneration Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. SS	235
15	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ], Added to Regeneration Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. TG	235
16	Analysis of Variance for the Effect of Treatment (Silver Nitrate	



	Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. SS	2
17	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ], Added to Seed Germination Medium and Regeneration Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. TG	2
18	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ]) on Shoot Primordia Elongation (Variable) of Cucumber cvs. SS and TG	2
19	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ]) on Induction of Phenotypically Normal Shoots (Variable) from Cucumber cvs. SS and TG	2
20	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ]) on Root Induction (Variable) from Cucumber Shoots cvs. SS and TG	2
21	Analysis of Variance for the Effect of Treatment (Silver Nitrate [AgNO <sub>3</sub> ]) on Shoot Primordia Induction (Variable) from Muskmelon cv BD	2
22	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ], Added to Seed Germination Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. SS	2
23	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ], Added to Seed Germination Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. TG	2
24	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ], Added to Regeneration Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. SS	2
25	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ], Added to Regeneration Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. TG	2
26	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ], Added to Seed Germination Medium and Regeneration Medium) on Shoot Primordia Induction Variable) of Cucumber cv. SS	2
27	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ], Added to Seed Germination Medium and Regeneration Medium) on Shoot Primordia Induction (Variable) of Cucumber cv. TG	2
28	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ]) on Shoot Primordia Elongation (Variable) of Cucumber cvs. SS and TG	2
29	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ]) on Induction of Phenotypically Normal Shoots (Variable) from Cucumber cvs. SS and TG	2
30	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ]) on Root Induction (Variable) from Cucumber Shoots cvs. SS and TG	2



31	Analysis of Variance for the Effect of Treatment (Cobalt Chloride [CoCl <sub>2</sub> ]) on Shoot Primordia Induction (Variable) from Muskmelon cv BD	240
32	Analysis of Variance for the Effect of Co-cultivation Period on Shoot Regeneration from Explants Cultured on Medium Containing Cf Either with or without Km (Variable)	240
33	Analysis of Variance for the Effect of Feeder Layer Incubation on Shoot Regeneration from Explants of Cucumber and Muskmelon Cultured on Medium Containing Cf Either with or without Km (Variable)	241
34	Analysis of Variance for the Effect of Acetosyringone on Shoot Regeneration from Explants of Cucumber and Muskmelon Cocultured for 10 and 20 min (Variable) and subsequently cultured on MS+Cf	241
35	Analysis of Variance for the Effect of Acetosyringone on Shoot Regeneration from Explants of Cucumber and Muskmelon Cocultured for 10 and 20 min (Variable) and subsequently cultured on MS+Cf+Km	0.44
		241



#### **LIST OF FIGURES**

Figure		Page
3.1	Effect of BAP on Shoot Regeneration from Proximal Cotyledon and Hypocotyl of Cucumber Cultivars	48
3.2	Effect of BAP on Number of Shoots Induced Per Proximal Cotyledon and Hypocotyl of Cucumber Cultivars	48
3.3	Effect of BAP and 2,4-D on Shoot Regeneration of Cucumber	52
3.4	Effect of BAP and 2,4-D on Numbers of Shoots Induced From Explants of Various Cucumber Cultivars	52
3.5	Effect of BAP and NAA on Induction of Shoot Primordia from Explants of Cucumber Cultivars	57
3.6	Effect of BAP and NAA on Number of Shoots Induced from Cucumber Explants	57
3.7	Effect of BAP or in Combinations with 2,4-D or NAA on Elongation of Shoots Obtained from Cucumber Explants	61
3.8	Effect of BAP or in Combinations with 2,4-D or NAA on Height of Elongated Shoots Obtained from Cucumber Explants	61
3.9	Effect of BAP or in Combinations with 2,4-D or NAA on Induction of Phenotypically Normal Shoots Obtained from Cucumber Explants	65
3.10	Effect of BAP or in Combinations with 2,4-D or NAA on Height of Phenotypical Normal Shoots Induced from Explants of Cucumber-	65
3.11	Effect of BAP or in Combinations with 2,4-D or NAA on Induction of Phenotypically Abnormal Shoots Obtained from Cucumber Explants	69
3.12	Effect of BAP or BAP+2,4-D or BAP+NAA on Height of Phenotypical Normal Shoots Obtained from Cucumber Explants	69
3.13	Effect of IBA on Root Induction from Cucumber Shoots	72
3.14	Effect of IBA on Average Number of Roots Induced from Cucumber Shoots	72
3.15	Effect of Dark Incubation on Root Induction from Non-rooted Cucumber Shoots	73
3.16	Effect of Dark Incubation on Induction of Average Number of Roots per Shoot	73
3.17	Effect of BAP on Shoot Regeneration from Muskmelon Explants	78
3.18	Effect of BAP on Average Number of Shoots Induced per Explant of Muskmelon	78



4.1	Increase of Shoot Regeneration from Proximal Cotyledon and Hypocotyl of SS and TG Cultivars by AgNO <sub>3</sub>
4.2	Enhancement of Number of Shoots Induced from Proximal Cotyledon and Hypocotyl of SS and TG Cultivars
4.3	Enhancement of Shoot Regeneration from Explants of SS and TG Cultivars
4.4	Enhancement of Number of Shoots Induced from Explants of SS and TG Cultivars
4.5	Effect of AgNO <sub>3</sub> on Shoot Regeneration from Explants of SS and TG Cultivars
4.6	Highest Average Numbers of Shoots Induced from Explants of SS and TG Cultivars
4.7	Increase of Shoot Regeneration from Proximal Cotyledon and Hypocotyl of SS and TG Cultivars by CoCl <sub>2</sub>
4.8	Enhancement of Number of Shoots Induced from Proximal Cotyledon and Hypocotyl of SS and TG Cultivars
4.9	Enhancement of Shoot Regeneration from Explants of SS and TG Cultivars
4.10 :	Enhancement of Number of Shoots Induced from Explants of SS and TG Cultivars
4.11	Effect of CoCl <sub>2</sub> on Shoot Regeneration from Explants of SS and TG Cultivars
4.12	Effect of CoCl₂ on Average Numbers of Shoots Induced from Explants of SS and TG Cultivars
5.1	Strain-dependent Variation in Crown Gall Induction on Cucumber, Explants cv. SS
5.2	Strain-dependent Variation in Crown Gall Induction on Muskmelon Explants cv. BD
5.3	Variation in hairy root Induction Percentage on Cucumber cv. SS Inoculated with <i>A. rhizogenesis</i>
5.4	Variation in Hairy Root Induction Percentage on Muskmelon Explants cv. BD Inoculated with <i>A. rhizogenesis</i>
5.5	Variation in Hairy Root Induction Percentage under Dark Incubation on Muskmelon Explants Inoculated with A. rhizogenesis
5.6	Effect of Dark Incubation on Induction of Average Numbers of Hairy Roots from Muskmelon Explants Inculated with A rhizogenesis
5.7	Effect of Acetosyringone on Crown Gall Induction from Stem Explants of Cucumber
5.8	Effect of Acetosyringone on Crown Gall Production from Petiole



5.9	Muskmelon 1
5.10	Effect of Acetosyringone on Crown Gall Induction from Petiole Explants of Muskmelon
5.11	Effect of Acetosyringone on Hairy Root Induction from Stem Explants of Cucumber, cv. SS
5.12	Effect of Acetosyringone on Hairy Root Induction from Petiole Explants of Cucumber, cv. SS
5.13	Effect of Acetosyringone on Hairy Root Induction from Stem Explants of Muskmelon1
5.14	Effect of Acetosyringone on Hairy Root Induction from Petiole Explants of Muskmelon
5.15	Variation on Number of Hairy Roots Induced from Cucumber Stem Inoculated by <i>A. rhizogenes</i> Grown with Acetosyringone
5.16	Variation on Number of Hairy Roots Induced from Cucumber Petiole Explants Inoculated by <i>A. rhizogenes</i> Strains Grown with Acetosyringone
5.17	Variation on Numbers of Hairy Roots Induced from Muskmelon Stem Inoculated by <i>A. rhizogenes</i> Strains Grown with Acetosyringone
5.18	Variation on Numbers of Hairy Roots Induced from Muskmelon Petioles Inoculated by A. mizogenes Grown with Acetosyringone
6.1	A Schematic Diagram of T-DNA Region of a Binary Vector Plasmid p35SGUSINT Used in the Transformation Study of Cucumber and Muskmelon



#### LIST OF PLATES

Plate		Page
3.1	Shoot Primordia Induction in Cucumber	47
3.2	Induction of Cucumber Shoots	58
3.3	Elongation of Cucumber Shoot Primordia	62
3.4	Phenotypical Abnormalities in Cucumber	66
3.5	Root Induction in Cucumber Shoots	70
3.6	Shoot Primordia Induction in muskmelon	77
3.7	Shoot Primordia Elongation	83
3.8	Induction of Phenotypical Normal and Abnormal Shoots	89
3.9	Root Induction	94
3.10	Acclimatisation of Cucumber and Muskmelon Plantlets	95
5.1	Callus Initiated from Wound Site of Cucumber Stem (a) and Crown Gall Formed in Muskmelon Petiole (b) After Infection with Agrobacterium tumefaciens Strain LBA 4404	153
5.2	Hairy Root Initiated from Wound Site of Cucumber Stem (a) and Muskmelon Stem (b) 14 Days After Infection with Agrobacterium rhizogenes Strain 8196	157
5.3	Callus Initiated from Wound Site of Muskmelon Stem at 39.3 μmol m <sup>-2</sup> s <sup>-1</sup> (a) Subsequently Formed Hairy Roots at 23.6 μmol m <sup>-2</sup> s <sup>-1</sup> (b) 14 Days After Infection with <i>A. rhizogenes</i> strain A 105	162
5.4	(i) Hairy Roots Initiated from Inoculated site of Cucumber Stem (a) Muskmelon Stem (b) Petiole (c) 21 Days After Infection with A. Rhizogenes Strain 8196 with Acetosyringone. (ii) Hairy Roots Elongated on Hormone Free MS Medium (a) and Elongation Stopped on MS Medium with 2,4-D (b).	170
6.1	The Muskmelon Regenerated Shoots Died on Medium Containing Kanamycin	188
6.2	The Transformed cucumber shoots survived on Rooting medium (MS+kanamycin)	188
7.1	The Leaves of Putative Transformed Shoots Derived from	



	Staining (Left). Control Leaves were Unresponsive (Right)	201
7.2	PCR Amplification of the GUS Gene from Genomic DNA Isolated from Transgenic Cucumber (lane 3) and Muskmelon (lane 6) Plants. No Bands Obtained from Negative Control Plants (lane 4 and 5). Lane 2 is the Positive Vector Control and Lane 1 is 1kb Ladder	201
7.3	Southern Blot Hybridisation of <i>Hind</i> III Restricted DNA from Transgenic Cucumber (lane 2), Muskmelon (lane 5) and Untransformed Control Plants (lane 3, 6). Lanes 1 and 4 are the Positive Control (vector DNA)	201



#### **LIST OF ABBREVIATION**

ANOVA analysis of variance

BAP 6-benzylaminopurine

Bp base pair

CaMV cauliflower mosaic virus

CAT chloramphenicol acetyl transferase

Cf Cefotaxime

CPA p-chlorophenoxyacetic acid

cv cultivar

cvs cultivars

d day

DMRT duncan multiple range test

2,4-D 2,4-dichlorophenoxyacetic acid

g gram

GUS  $\beta$ -glucuronidase

h hour

IAA indole acetic acid

IBA indole-3-butyric acid

2iP 6- $(\gamma-\gamma$ -dimethylallylamino) purine

Kg Kilogram

Km kanamycin

L liter

LB left border

LB medium Luria-Bertani medium

M molar

Min minute

ml millilitre



MS Murashige and Skoog

MUG 4-methyl umbelliferyl glucuronide

NAA naphthalene acetic acid

NOS nopaline syntheses

nptll neomycin phosphotransferase II

OD optimal density

PCR polymerase chain reaction

P<sup>H</sup> hydrogen ion concentration

RB right border

Rf rifampicin

Ri root-inducing

SAS statistical analysis system

s second

2,4,5-T 2,4,5-trichlorophenoxyacetic acid

TDZ thidiazuron

Ti tumour-inducing

v/v volume/volume

w/v weight/volume

*X-gluc* 5-bromo-4-chloro-3-indolyglucuronide

 $\boldsymbol{\mu} \qquad \quad \text{micro}$ 



Abstract of dissertation submitted to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy.

IMPROVEMENT IN ORGANOGENESIS AND THE DEVELOPMENT OF A TRANSFORMATION PROCEDURE FOR CUCUMBER AND MUSKMELON

By

A.K.M. MOHI UDDIN

December 1998

Chairman: Dr. Suhaimi Napis

Faculty: Food Science and Biotechnology

A comprehensive study was carried out to optimise and improve a direct shoot organogenesis system to facilitate the transfer of reporter genes (GUSINT and nptll) into cucumber (Cucumis sativus L.) and muskmelon (C. melo L.) plants. The studies carried out were: (i) a comparative study on direct and indirect shoot regeneration (ii) the improvement of a direct shoot regeneration system by using an ethylene action inhibitor, silver nitrate and an ethylene biosynthesis inhibitor, cobalt chloride (iii) identification of suitable strains of Agrobacterium tumefaciens and A. rhizogenes that are able to infect cucumber and muskmelon plants (iv) development of an intron containing gene transformation system for both cucurbit species through A. tumefaciens and (v) analysis of transformants.

For the comparative study of direct and indirect shoot regeneration, four different types of explants from five elite commercial cucumber cultivars namely Spring Swallow, Suyo Cross, Suyo Long, Tasty Glory, Tasty Green and one muskmelon cultivar called Birdie were used. Explants from both species were cultured onto Murashige and Skoog medium containing different concentrations of 6-benzylaminopurine alone and in combination with either 2,4-dichlorophenoxyacetic acid or naphthalene acetic acid. 6benzylaminopurine alone at 1.0 or 2.0 mg/l significantly (p<0.05) induced shoot primordia

from the largest number of proximal cotyledon and hypocotyl explants of all cucumber cultivars and muskmelon explants tested. However, the addition of either 2,4-dichlorophenoxyacetic acid or naphthalene acetic acid to 6-benzylaminopurine, resulted in a reduction of the shoot primordia induction rate.

Higher number of shoots induced from explants as well as high numbers of morphological normal shoots were obtained when explants were cultured on medium containing 6-benzylaminopurine alone. Specific concentrations of indole-butyric acid and naphthalene acetic acid significantly (p<0.05) contributed to root initiation from the largest number of cucumber and muskmelon shoots, respectively. Dark treatment was sufficient to significantly induce root formation from the non-rooting cucumber and muskmelon shoots.

The addition of either silver nitrate or cobalt chloride to the seed germination medium or shoot primordia induction medium caused a significant (p<0.05) enhancement of shoot regeneration rate from cucumber cv. SS and TG explants compared to the control. The regeneration rate was further enhanced when these two ethylene inhibitors were added to both SGM and SPI media. Furthermore, the number of shoots induced from explants of both Spring Swallow and Tasty Green cultivars was also enhanced upon the same treatment. However, muskmelon shoot induction and regeneration were reduced when the same treatment was employed.

In the *Agrobacterium*-mediated transformation experiments inoculation of cucumber cultivar Spring Swallow and muskmelon cultivar Birdie explants with *A. tumefaciens* and *A. rhizogenes* wild type strains revealed the different degrees of virulence of both bacteria. It was found that the virulence of both *Agrobacterium* species was enhanced when acetosyringone was added to the culture of inoculum

