



**UNIVERSITI PUTRA MALAYSIA**

**EFFECTS OF SINGLE AND COMBINED INOCULATION OF PLANT  
GROWTH PROMOTING RHIZOBACTERIA ON GROWTH  
PERFORMANCE OF PADDY (ORYZA SATIVA L.)  
UNDER GLASSHOUSE AND FIELD CONDITIONS**

**MOHAMAD HALID BIN A RAZAK**

**FP 2018 102**



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CONDITIONS**

By

**MOHAMAD HALID BIN A RAZAK**

**Thesis submitted to the School of Graduate Studies,  
Universiti Putra Malaysia, in Fulfillment of the  
Requirements for Degree of Master of Science**

**March 2018**

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## DEDICATION

I dedicate this thesis to my beloved Wife, Mrs. Nursyaza Bt Zulkifli and my parents for their patience, love, understanding and encouragement.

Thank you very much for everything



Abstract of thesis presented to the Senate of Universiti Putra Malaysia,  
in fulfillment of the requirement for the degree of Master of Science

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**March 2018**

**Supervisor : Associate Prof. Halimi Bin Mohd Saud, PhD**  
**Faculty : Agriculture**

Rice (*Oryza sativa* L.) is a staple food in Malaysia grown over 708,148 hectares in various granary areas of Malaysia. In 2016, the average rice production in Malaysia was 2.252 million tons. As the population of Malaysia increases from the current number of 31.2 million, the rice demand also increases. However, Malaysia is only able to achieve 71.2% self-sufficiency level (SSL). Thus, Malaysia has to import rice from other countries, requiring higher productivity in limited granary areas to fulfill domestic demand. A potential solution to this problem is by increasing the usage of plant growth promoting rhizobacteria (PGPR) as a biofertilizer to enhance the productivity and reduced environmental problems caused by application of chemical fertilizers. The objectives of this study were to study the interaction effects of single and combined inoculations of PGPR using *in vitro* assay studies, effects on rice growth and development in glasshouse experiments and effects on rice yield, biomass and nutrient uptake in a field experiment. Determination of panicle breaking strength to minimize panicle grain shattering under field experiments was also conducted in this study. PGPR used in this study were *Bacillus pumilus* GM118 and *Bacillus subtilis* UPMB10 which act as phosphate solubilizing bacteria (PSB) and nitrogen fixing bacteria (NFB), respectively. *B. pumilus* GM118 and *B. subtilis* UPMB10 growth population trends, ability to fix nitrogen and solubilize phosphate on three different selective media and production of indole-3-acetic acid (IAA) were conducted *in vitro* assay. Four treatments of PGPR with different fertilizers treatments (including control) were laid out in a randomized complete block design (RCBD) with four replications

conducted in a glasshouse at Ladang 10, Faculty of Agriculture, Universiti Putra Malaysia. The treatments were as follows: T1: control N0K0; T2: N0K90; T3: N40K30; T4: N40K60; T5: N80K30; T6: N80K60; T7: N120K0 and T8: N120K90 in the present/absent of single and combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10. N and K in the treatments refer to the amount of nitrogen and potassium, respectively. The paddy variety used in glasshouse experiment was MR263 and it was planted using 35 L buckets containing 15 kg of paddy soil. Meanwhile, the field experiment was conducted in Kemasin-Semerak Integrated Agricultural Development Area (IADA), Kelantan. Four treatments were applied in IADA Kemasin-Semerak: T1: control (without bacterial inoculation); T2: single inoculation of *B. pumilus* GM118; T3: single inoculation of *B. subtilis* UPMB10 and T4: combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10. Normal fertilization practices were used in these plots. All data were analyzed statistically using SAS Software Program (Version 9.3) and treatment means were compared using Duncan Multiple Range Test ( $P < 0.05$ ). *In vitro* assay indicated that combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 produced the shortest generation times (14.4 minutes) for cells doubling compared to single inoculations of *B. pumilus* GM118 (36.1 minutes) and *B. subtilis* UPMB10 (24.1 minutes). However, single inoculation of *B. subtilis* UPMB10 is able to produce more IAA (63.8  $\mu\text{g/mL}$ ) compared to combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 (50.1  $\mu\text{g/mL}$ ). Combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 produced the highest grain yield (33%), followed by single inoculations of *B. subtilis* UPMB10 (29%) and *B. pumilus* GM118 (23%) as compared to the control in the glasshouse experiment. *B. pumilus* GM118 showed the highest N contents (15%) while combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 showed the highest K content (16%) compared to control. Detachment of the grain panicle from the pedicle required the highest force for paddy inoculated with single inoculation of *B. subtilis* UPMB10 (11%) and followed by combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 (8%) as compared to the control. In the field experiment, both single and combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 showed significant difference in grain yield and biomass. In grain yield, *B. pumilus* GM118 produced 40%, *B. subtilis* UPMB10 (61%) and combined inoculations (61%) higher compared to the control. For biomass production, *B. pumilus* GM118 produced 33%, *B. subtilis* UPMB10 (42%) and combined inoculations of both (40%) higher compared to the control. IADA Kemasin-Semerak study showed that single inoculation of *B. pumilus* GM118 was able to increase 44% and 126% of plant N and P uptake while *B. subtilis* UPMB10 was able to increase 30% of plant K uptake compared to control, respectively.

This study indicated that co-inoculation having beneficial multi-mechanism of fixing nitrogen, solubilizing phosphate and producing IAA, gave the optimum performance in both glasshouse and field studies. This study proves that inoculation of selected PGPR can enhance rice plant yield, biomass, nutrient uptake and reduce postharvest losses due to grain shattering. The results showed the potential use of combined inoculations of *B. pumilus* GM118 and *B. subtilis* UPMB10 as biofertilizer with multiple beneficial characteristics that can increase rice productivity in the granary areas of Malaysia.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**KESAN INOKULASI TUNGGAL DAN GABUNGAN BAKTERIA  
PENGKALAK PERTUMBUHAN POKOK TERHADAP PRESTASI PADI  
(*Oryza sativa* L.) DALAM KAJIAN RUMAH KACA DAN LAPANGAN**

Oleh

**MOHAMAD HALID BIN A RAZAK**

**Mac 2018**

**Penyelia : Prof. Madya Halimi Bin Mohd Saud, PhD**  
**Fakulti : Pertanian**

Beras (*Oryza sativa* L.) merupakan makanan ruji di Malaysia yang ditanam melebihi 708,148 hektar di kawasan jelapang padi yang berbeza di Malaysia. Secara puratanya, penghasilan padi di Malaysia pada tahun 2016 adalah sebanyak 2.252 juta tan. Bilangan terkini penduduk di Malaysia adalah kira-kira 31.2 juta dan apabila kadar populasi penduduk semakin meningkat, permintaan terhadap beras juga kian meningkat. Walau bagaimanapun, Malaysia hanya mampu mencapai 71.2% kadar keberhasilan padi (SSL). Oleh itu, Malaysia harus mengimport beras dari negara luar dan meningkatkan lagi produktiviti di kawasan jelapang padi yang terhad ini untuk memenuhi kehendak domestik. Potensi untuk menangani masalah ini adalah dengan meningkatkan penggunaan bakteria pengkalak pertumbuhan pokok (PGPR) sebagai biobaja untuk meningkatkan produktiviti serta mengurangkan masalah pencemaran akibat penggunaan baja kimia. Objektif kajian ini adalah untuk mengkaji kesan interaksi di antara inokulasi tunggal dan gabungan inokulasi PGPR dalam kajian *in vitro*, kesannya terhadap pertumbuhan dan perkembangan padi di dalam kajian rumah kaca dan kesannya terhadap hasil padi, biomass dan pengambilan nutrien di kajian lapangan. Penentuan kekuatan pemutusan panikel untuk meminimumkan leraian buah padi di bawah kaedah kawalan dan lapangan juga turut dijalankan. PGPR yang digunakan di dalam kajian ini adalah *Bacillus pumilus* GM118 yang bertindak sebagai bakteria pelarut fosfat (PSB) dan *Bacillus subtilis* UPMB10 yang bertindak sebagai bakteria pengikat nitrogen (NFB). Pertumbuhan populasi, kebolehan untuk mengikat nitrogen dan mengurai fosfat oleh *B. pumilus* GM118 dan *B. subtilis* UPMB10 pada tiga



media terpilih yang berbeza serta penghasilan asid indole-3-asetik (IAA) dijalankan secara *in vitro*. Empat rawatan PGPR dengan rawatan baja yang berbeza (termasuk kawalan) telah dijalankan menggunakan reka bentuk blok lengkap rawak (RCBD) dengan empat replikasi di dalam rumah kaca Ladang 10, Fakulti Pertanian, Universiti Putra Malaysia. Rawatan tersebut adalah seperti berikut: T1: kawalan N0K0; T2: N0K90; T3: N40K30; T4: N40K60; T5: N80K30; T6: N80K60; T7: N120K0 and T8: N120K90 dengan kehadiran atau tanpa kehadiran inokulasi tunggal dan gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10. N dan K di dalam rawatan merujuk kepada jumlah nitrogen dan kalium. Varieti padi yang digunakan adalah MR263 dan ditanam di dalam baldi bersaiz 35 L mengandungi 15 kg tanah sawah. Kajian lapangan pula dijalankan di Kemasin-Semerak Kawasan Pembangunan Bersepadu (IADA), Kelantan. Empat rawatan telah diaplikasikan di IADA Kemasin Semerak (IADA KS) iaitu: T1: Kawalan (tanpa inokulasi bakteria), T2: inokulasi tunggal *B. pumilus* (GM118), T3: inokulasi tunggal *B. subtilis* (UPM B10) dan T4: gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10. Kesemua data yang diperolehi telah dianalisis menggunakan Program Perisian SAS (Versi 9.3), dan purata rawatan telah dibandingkan menggunakan Ujian Pelbagai Duncan (DMRT) ( $P < 0.05$ ). Kajian *in vitro* menunjukkan gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10 menghasilkan sel berganda dua dalam masa yang paling singkat (14.4 minit) untuk sel berganda dua berbanding inokulasi tunggal *B. pumilus*; GM118 (36.1 minit) dan *B. subtilis*; UPMB10 (24.1 minit). Walau bagaimanapun, inokulasi tunggal *B. subtilis* UPMB10 telah menghasilkan lebih banyak IAA (63.8  $\mu\text{g/mL}$ ) berbanding gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10 (50.1  $\mu\text{g/mL}$ ). Gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10 menghasilkan hasil padi yang paling tinggi (33%) dan diikuti oleh inokulasi tunggal *B. subtilis* UPMB10 (29%) dan *B. pumilus* GM118 (23%) berbanding kawalan di dalam kajian rumah kaca. Inokulasi tunggal *B. pumilus* GM118 menunjukkan kandungan N yang tertinggi (15%) sementara gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10 menunjukkan kandungan K yang tertinggi (16%) berbanding kawalan. Penginokulan dengan inokulasi tunggal *B. subtilis* UPMB10 memerlukan daya yang paling tinggi (11%) untuk meleraikan panikel padi daripada tangkai diikuti dengan gabungan inokulasi yang memerlukan 8% lebih daya berbanding kawalan. Inokulasi tunggal dan gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10 menunjukkan perbezaan yang ketara untuk hasil padi dan biojisim di dalam kajian lapangan. Untuk hasil padi, *B. pumilus* GM118 menghasilkan 40%, *B. subtilis* UPMB10 (61%) dan gabungan inokulasi kedua-dua bakteria (61%) lebih tinggi berbanding kawalan. Kajian IADA Kemasin-Semerak menunjukkan inokulasi tunggal *B. pumilus* GM118 mampu meningkatkan pengambilan N sebanyak 44% dan pengambilan P oleh tumbuhan sebanyak 126% sementara *B. subtilis* UPMB10 mampu meningkatkan pengambilan K oleh tumbuhan sebanyak 30% berbanding kawalan.

Oleh yang demikian, kajian menunjukkan gabungan inokulasi mekanisma pelbagai manfaat untuk mengikat nitrogen, melarut fosfat dan menghasilkan IAA dapat memberikan kesan optimum di dalam kajian rumah kaca dan lapangan. Kajian ini membuktikan bahawa inokulasi dengan PGPR terpilih dapat meningkatkan hasil padi, biojisim, pengambilan nutrien dan mengurangkan kerugian pasca-tuai akibat peleraian buah. Hasil kajian menunjukkan potensi penggunaan gabungan inokulasi *B. pumilus* GM118 dan *B. subtilis* UPMB10 sebagai biobaja yang bercirikan pelbagai manfaat untuk meningkatkan produktiviti padi di kawasan jelapang padi di Malaysia.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF GLOSSARY OF ABBREVIATIONS AND SI UNITS

g	Gram
g/L	Gram per liter
H	Hour
L	Liter
mg	Milligram
mg/L	Milligram per liter
mm	Millimeter
min	Minute
M	Molar
$\mu$ M	Micro molar
NA	Nutrient Agar
NB	Nutrient Broth
RPM	Revolution per minutes
UPM	Universiti Putra Malaysia
IADA	Integrated Agricultural Development Area
SOM	Soil Organic Matter
IAA	Indole-3-acetic acid

## CHAPTER 1

### INTRODUCTION

Rice (*Oryza sativa* L.) is the main crop in Asia and it covers half of the arable land used for agriculture in most countries (Cantrell and Hettel, 2004). Rice is the second most widely grown cereal crop and it is a primary food which serves food calories and protein as main part of daily diets for most of the world's population. Based on FAOSTAT (2017) database, world rice production in 2013 was 741.9 million tons and average yield production was 4.51 tons/ha. However, world rice production in 2016 was 740.9 million tons and average yield production was 4.64 tons/ha. The demand for rice is correlated with an increase in population. The Food and Agriculture Organization (FAO) rice market in 2012 stated that the current supply of rice outpaces consumption. However, an increase in cereal crop supply is necessary to overcome the future rice demand in world population. Sheehy and Mitchell (2013) reported that it requires an increase of 50% in rice yield to fulfill the predicted world population of 9.3 billion by 2050.

In Malaysia, rice is the staple food and the third most widely planted crop after oil palm and rubber. Based on FAOSTAT database, area harvested of rice and rice production in Malaysia in 2016 are 708,148 hectare and 2.252 million tons, respectively ([www.fao.org](http://www.fao.org)) and population in Malaysia is about 31.2 million ([www.worldometers.info](http://www.worldometers.info)). However, Malaysia is only able to achieve 71.2% of self-sufficiency level (SSL) in order to produce its own rice to fulfill local rice demand (DOA, 2015). Hence, Malaysia has to import rice from other countries such as Thailand (41%) and Vietnam (32%) to support the rice domestic demand as Malaysia is unable to achieve 100% self-sufficiency level (SSL) of rice production. Besides, to meet the targeted level of SSL, continuous R&D is needed to produce new high yielding varieties and improved conventional method (productivity target needed to fulfill the gap between the actual and the desired production). In conventional method, farmers commonly apply more than the recommended rate of fertilizer to increase rice yield as a short-term solution.

Arshad et al. (2011) stated that Malaysia could sustain 70% SSL in a long-term target and fulfill the agreement on agriculture (AoA) of the World Trade Organization (WTO). They also mentioned that a productivity target based on SSL target was recommended for higher rice production. Therefore, Malaysia needs to propose continues R&D to increase rice production, cultivation area and crop productivity in order to fulfill the demand of a rising population. Hence, one way to overcome this problem is by promoting the use of Plant Growth Promoting Rhizobacteria (PGPR).



PGPR are heterogeneous clusters of microorganisms that have the ability to colonize roots rhizosphere and benefit the crop plants. PGPR are considered as bacteria which originated within rhizosphere or endophytic, free-living or in association with plant roots (Maksimov et al. 2011). Rhizosphere is the zone or area in the soils surrounding plant root and has the highest microbial activity area which have the capability to exchange essential nutrient elements to usable forms by biological stress activator (Muraleedharan et al., 2010). The microbial populations in the rhizosphere are varied compared with the other areas due to root exudates that act as nutrient sources for the microbes (Burdman et al., 2000). According to Lugtenberg et al. (2001), PGPR need to colonize the root surface effectively and compete well with other microbes for nutrient sources and spaces by invading the root in order to give advantage to the plants.

PGPR can act as an essential part in improving plant growth via various modes of mechanisms. The mode of PGPR action which encourages plant growth are biotic and abiotic stress tolerance, enhance nutrient uptake, phytohormones production and siderophore production (Choudhary et al., 2011; García-Fraile et al., 2015). Commonly, a single PGPR inoculation such as *Bacillus* sp., *Pseudomonas* sp. and *Azospirillum* sp. used as inoculants, showed capability to enhance shoot growth, root density and yield. These improvements in plant growth caused by PGPR are due to the potential of fixing and solubilizing mineral fertilizer and production of phytohormones resulting in the increased of nutrients availability and roots permeability (Enebak and Carey, 2000). Combine inoculation of PGPR can also increase plant growth and yield, but compared to single inoculation, combine inoculation provides the plants with more balanced nutrition, and improved the absorption of mineral nutrients (Bashan and Holguin, 1997). Thus, plant growth can be increased by dual inoculation of nitrogen-fixing bacteria and phosphate-solubilizing bacteria which can produce more phytohormones when grown in mixed culture. The combination of microbes that interact synergistically and promotes plant growth is currently an on-going study in many other countries. In Malaysia, similar studies need to be conducted to determine the effectiveness of locally isolated PGPR and their effect on the growth and yield of local rice varieties.

Hence, the present study was focused on the effect of nitrogen fixing bacteria (NFB) and/or phosphate solubilizing bacteria (PSB) strains to improve uptake, growth and yield of rice.

The main objectives of the study are:

- i) To differentiate the response of single and combined inoculations of PGPR bacteria *in vitro* assay studies focusing on population growth and its mechanisms.
- ii) To determine the growth response of inoculated rice plant (*Oryza sativa L.*) with different ratio of chemical fertilizer with single and combined inoculations of PGPR bacteria under glasshouse condition.
- iii) To measure the breaking panicle strength at different applications of chemical fertilizer to minimize panicle grain shattering habit under glasshouse condition.
- iv) To evaluate the growth response of rice plant inoculated with single and combined inoculations of PGPR bacteria under field condition.

## REFERENCES

- Abbas-Zadeh, P., Saleh-Rastin, N., Asadi-Rahmani, H., Khavazi, K., Soltani, A., Shoary-Nejati, A.R. and Miransari, M., 2010. Plant growth-promoting activities of fluorescent pseudomonads, isolated from the Iranian soils. *Acta Physiologiae Plantarum*, 32(2), pp.281-288.
- Addicott, F.T., 1982. Abscission. University of California Press, Berkeley.
- Adesemoye, A.O. and Kloepper, J.W., 2009. Plant-microbes interactions in enhanced fertilizer-use efficiency. *Applied microbiology and biotechnology*, 85(1), pp.1-12.
- Adesemoye, A.O., Torbert, H.A. and Kloepper, J.W., 2009. Plant growth-promoting rhizobacteria allow reduced application rates of chemical fertilizers. *Microbial ecology*, 58(4), pp.921-929.
- Adesemoye, A.O., Torbert, H.A. and Kloepper, J.W., 2010. Increased plant uptake of nitrogen from 15 N-depleted fertilizer using plant growth-promoting rhizobacteria. *Applied soil ecology*, 46(1), pp.54-58.
- Ahmad, M., Zahir, Z.A., Khalid, M., Nazli, F. and Arshad, M., 2013. Efficacy of *Rhizobium* and *Pseudomonas* strains to improve physiology, ionic balance and quality of mung bean under salt-affected conditions on farmer's fields. *Plant Physiology and Biochemistry*, 63, pp.170-176.
- Ahmed, A. and Hasnain, S., 2010. Auxin-producing *Bacillus* sp.: Auxin quantification and effect on the growth of *Solanum tuberosum*. *Pure and Applied Chemistry*, 82(1), pp.313-319.
- Akhtar, M.S. and Memon, M., 2009. Biomass and nutrient uptake by rice and wheat: a three-way interaction of potassium, ammonium and soil type. *Pakistan Journal of Botany*, 41(6), pp.2965-2974.
- Akhtar, N., Arshad, I., Shakir, M.A., Qureshi, M.A., Sehrish, J. and Ali, L., 2013. Co-inoculation with *Rhizobium* and *Bacillus* sp. to improve the phosphorus availability and yield of wheat (*Triticum aestivum* L.). *JAPS, Journal of Animal and Plant Sciences*, 23(1), pp.190-197.
- Alagawadi, A.R. and Gaur, A.C., 1992. Inoculation of *Azospirillum brasilense* and phosphate-solubilizing bacteria on yield of sorghum [*Sorghum bicolor* (L.) Moench] in dry land. *Tropical Agriculture*, 69(4), pp.347-350.
- Aleksandrov, V.G., Blagodyr, R.N. and Ilev, I.P., 1967. Liberation of phosphoric acid from apatite by silicate bacteria. *Mikrobiol Z*, 29, pp.111-114.
- Alizadeh, M.R. and Allameh, A., 2011. Threshing force of paddy as affected by loading manner and grain position on the panicle. *Research in Agricultural Engineering*, 57, pp.8-12.

- Aloni, R., Aloni, E., Langhans, M. and Ullrich, C.I., 2006. Role of cytokinin and auxin in shaping root architecture: regulating vascular differentiation, lateral root initiation, root apical dominance and root gravitropism. *Annals of botany*, 97(5), pp.883-893.
- Amer, G.A. and Utkhede, R.S., 2000. Development of formulations of biological agents for management of root rot of lettuce and cucumber. *Canadian Journal of Microbiology*, 46(9), pp.809-816.
- Amir, H.G., Shamsuddin, Z.H., Halimi, M.S., Ramlan, M.F. and Marziah, M., 2003. N<sub>2</sub> fixation, nutrient accumulation and plant growth promotion by rhizobacteria in association with oil palm seedlings. *Pakistan Journal of Biological Sciences (Pakistan)*.
- Antoun, H., Beauchamp, C.J., Goussard, N., Chabot, R. and Lalande, R., 1998. Potential of *Rhizobium* and *Bradyrhizobium* species as plant growth promoting rhizobacteria on non-legumes: effect on radishes (*Raphanus sativus* L.). *Plant and soil*, 204(1), pp.57-67.
- Apostu, A., Petriman, N., Lulian, T., Mihasan, M., Dunca, S. and Stefan, M., 2010. Isolation and characterization of some rhizobacterial strains with phosphorus solubilizing capabilities. *Analele Stiintifice ale Universitatii" Al. I. Cuza" Din Iasi.(Serie Noua). Sectiunea 2. a. Genetica si Biologie Moleculara*, 11(4).
- Arshad, F.M., Alias, E.F., Noh, K.M. and Tasrif, M., 2011. Food Security: Self-Sufficiency of Rice In Malaysia<sup>1</sup>. *International Journal of Management Studies*, 18(2), pp.83-100.
- Arshad, F.M., Alias, E.F., Noh, K.M. and Tasrif, M., 2011. Food security: Self-sufficiency of rice in Malaysia. *International Journal of Management Studies*, 18(2), pp.83-100.
- Arzanesh, M.H., Alikhani, H.A., Khavazi, K., Rahimian, H.A. and Miransari, M., 2011. Wheat (*Triticum aestivum* L.) growth enhancement by *Azospirillum* sp. under drought stress. *World Journal of Microbiology and Biotechnology*, 27(2), pp.197-205.
- Ashley, M.K., Grant, M. and Grabov, A., 2006. Plant responses to potassium deficiencies: a role for potassium transport proteins. *Journal of Experimental Botany*, 57(2), pp.425-436.
- Ashrafuzzaman, M., Hossen, F.A., Ismail, M.R., Hoque, A., Islam, M.Z., Shahidullah, S.M. and Meon, S., 2009. Efficiency of plant growth-promoting rhizobacteria (PGPR) for the enhancement of rice growth. *African Journal of Biotechnology*, 8(7).

- Aslantaş, R., Çakmakçı, R. and Şahin, F., 2007. Effect of plant growth promoting rhizobacteria on young apple tree growth and fruit yield under orchard conditions. *Scientia Horticulturae*, 111(4), pp.371-377.
- Aung, H.P., Mensah, A.D., Aye, Y.S., Djedidi, S., Oikawa, Y., Yokoyama, T., Suzuki, S. and Bellingrath-Kimura, S.D., 2016. Transfer of radiocesium from rhizosphere soil to four cruciferous vegetables in association with a *Bacillus pumilus* strain and root exudation. *Journal of Environmental Radioactivity*, 164, pp.209-219.
- Ayeh, K.O., Lee, Y., Ambrose, M.J. and Hvoslef-Eide, A.K., 2009. Characterization and structural analysis of wild type and a non-abscission mutant at the development funiculus (Def) locus in *Pisum sativum* L. *BMC Plant Biology*, 9(1), p.76.
- Azziz, G., Bajsa, N., Haghjou, T., Taulé, C., Valverde, Á., Igual, J.M. and Arias, A., 2012. Abundance, diversity and prospecting of culturable phosphate solubilizing bacteria on soils under crop–pasture rotations in a no-tillage regime in Uruguay. *Applied Soil Ecology*, 61, pp.320-326.
- Bagyaraj, D.J., 2013. Mycorrhizal Symbiosis for Sustainable Horticulture. In *Biotechnology in Horticulture: Methods and Application* (pp. 155-177). New India Publishing Agency New Delhi.
- Bahadur, I., Meena V.S., Kumar, S. 2014. Importance and application of Potassic Biofertilizer in Indian agriculture. *International Research Journal of Biological Sciences* 3(12):80-85.
- Bahat-Samet, E., Castro-Sowinski, S. and Okon, Y., 2004. Arabinose content of extracellular polysaccharide plays a role in cell aggregation of *Azospirillum brasilense*. *FEMS Microbiology Letters*, 237(2), pp.195-203.
- Bangerth, F., 2000. Abscission and thinning of young fruit and their regulation by plant hormones and bioregulators. *Plant Growth Regulation*, 31, pp. 43-59.
- Barlog, P and Grzebisz, W. 2004. Effect of timing and nitrogen fertilizer application on winter oilseed rape (*Brassica napus* L.). II. Nitrogen uptake dynamics and fertilizer efficiency. *Journal of Agronomy and Crop Science* 190:314–323.
- Basak, B.B. and Biswas, D.R., 2010. Co-inoculation of potassium solubilizing and nitrogen fixing bacteria on solubilization of waste mica and their effect on growth promotion and nutrient acquisition by a forage crop. *Biology and Fertility of Soils*, 46(6), pp.641-648.

- Bashan, Y. and De-Bashan, L.E., 2010. Chapter two-how the plant growth-promoting bacterium *Azospirillum* promotes plant growth—a critical assessment. *Advances in Agronomy*, 108, pp.77-136.
- Bashan, Y. and Holguin, G., 1997. Short-and medium-term avenues for *Azospirillum* inoculation. *Plant Growth-Promoting Rhizobacteria. Present Status and Future Prospects* (Ogoshi, A., Kobayashi, K., Homma, Y., Kodama, F., Kondo, N. and Akino, S., Eds.), pp.130-149.
- Bashan, Y. and Levanony, H., 1990. Current status of *Azospirillum* inoculation technology: *Azospirillum* as a challenge for agriculture. *Canadian Journal of Microbiology*, 36(9), pp.591-608.
- Bashan, Y., Holguin, G., Bashan, L.E. 2004. *Azospirillum*-plant relationships: agricultural, physiological, molecular and environmental advances (1997-2003). *Canadian Journal of Microbiology* 50:521-577.
- Bashan, Y., Kamnev, A.A. and de-Bashan, L.E., 2013. Tricalcium phosphate is inappropriate as a universal selection factor for isolating and testing phosphate-solubilizing bacteria that enhance plant growth: a proposal for an alternative procedure. *Biology and Fertility of soils*, 49(4), pp.465-479.
- Belimov, A. A., Kojemiakov, P, A. and Chuvarliyeva, C. V., 1995. Interaction between barley and mixed cultures of nitrogen fixing and phosphate-solubilizing bacteria. *Plant Soil* 17: 29–37.
- Belimov, A.A., Postavskaya, S.M., Khamova, O.F., Kozhemyakov, A.P., Kunakova, A.M. and Gruzdeva, E.V., 1994. Effectiveness of barley inoculation with root diazotrophic bacteria and their survival depending on soil-temperature and moisture. *Microbiology*, 63(5), pp.506-510.
- Beneduzi, A., Peres, D., Vargas, L.K., Bodanese-Zanettini, M.H. and Passaglia, L.M.P., 2008. Evaluation of genetic diversity and plant growth promoting activities of nitrogen-fixing bacilli isolated from rice fields in South Brazil. *Applied Soil Ecology*, 39(3), pp.311-320.
- Bennett, P.C., Choi, W.J. and Rogera, J.R., 1998. Microbial destruction of feldspars. *Mineral Management*, 8(62A), pp.149-150.
- Biswas, J.C., Ladha, J.K. and Dazzo, F.B., 2000. Rhizobia inoculation improves nutrient uptake and growth of lowland rice. *Soil Science Society of America Journal*, 64(5), pp. 1644-1650
- Boddey, R.M. and Dobereiner, J., 1988. Nitrogen fixation associated with grasses and cereals: recent results and perspectives for future research. *Plant and soil*, 108(1), pp.53-65.

- Bottini, R., Cassán, F. and Piccoli, P., 2004. Gibberellin production by bacteria and its involvement in plant growth promotion and yield increase. *Applied microbiology and biotechnology*, 65(5), pp.497-503.
- Burd, G.I., Dixon, D.G. and Glick, B.R., 1998. A plant growth-promoting bacterium that decreases nickel toxicity in seedlings. *Applied and Environmental Microbiology*, 64(10), pp.3663-3668.
- Burdman, S., Jurkevitch, E. and Okon, Y., 2000. Recent advances in the use of plant growth promoting rhizobacteria (PGPR) in agriculture. *Microbial interactions in agriculture and forestry (Volume II)*, pp.229-250.
- Burdman, S., Jurkevitch, E., Schwartsburd, B. and Okon, Y., 1999. Involvement of outer-membrane proteins in the aggregation of *Azospirillum brasilense*. *Microbiology*, 145(5), pp.1145-1152.
- Çakmakçı, R., Dönmez, F., Aydın, A. and Şahin, F., 2006. Growth promotion of plants by plant growth-promoting rhizobacteria under greenhouse and two different field soil conditions. *Soil Biology and Biochemistry*, 38(6), pp.1482-1487.
- Çakmakçı, R., Kantar, F. and Sahin, F., 2001. Effect of N<sub>2</sub>-fixing bacterial inoculations on yield of sugar beet and barley. *Journal of Plant Nutrition and Soil Science*, 164(5), pp.527-531.
- Cassan, F., Perrig, D., Sgroy, V., Masciarelli, O., Penna, C. and Luna, V., 2009. *Azospirillum brasilense* Az39 and *Bradyrhizobium japonicum* E109, inoculated singly or in combination, promote seed germination and early seedling growth in corn (*Zea mays* L.) and soybean (*Glycine max* L.). *European Journal of Soil Biology*, 45(1), pp.28-35.
- Cattelan, A.J., Hartel, P.G. and Fuhrmann, J.J., 1999. Screening for plant growth-promoting rhizobacteria to promote early soybean growth. *Soil Science Society of America Journal*, 63(6), pp.1670-1680.
- Chabot, R., Beauchamp, C.J., Kloepper, J.W. and Antoun, H., 1998. Effect of phosphorus on root colonization and growth promotion of maize by bioluminescent mutants of phosphate-solubilizing *Rhizobium leguminosarum biovar phaseoli*. *Soil Biology and Biochemistry*, 30(12), pp.1615-1618.
- Chandramohan, A., Sivasankar, V., Ravichandran, C. and Sakthivel, R., 2013. A probe on the status of microorganisms in the air, soil and solid waste samples of Ariyamangalam dumping site at Tiruchirappalli district, South India. In *Microbiological Research in Agroecosystem Management* (pp. 1-9). Springer India.

- Chaparro, J.M., Sheflin, A.M., Manter, D.K. and Vivanco, J.M., 2012. Manipulating the soil microbiome to increase soil health and plant fertility. *Biology and Fertility of Soils*, 48(5), pp.489-499.
- Chen, J.H., 2006, October. The combined use of chemical and organic fertilizers and/or biofertilizer for crop growth and soil fertility. In *International workshop on sustained management of the soil-rhizosphere system for efficient crop production and fertilizer use* (Vol. 16, p. 20). Land Development Department Bangkok, Thailand.
- Chen, S., Zeng, F.R., Pao, Z.Z. and Zhang, G.P., 2008. Characterization of high-yield performance as affected by genotype and environment in rice. *Journal of Zhejiang University-Science B*, 9(5), pp.363-370.
- Choudhary, D.K., Sharma, K.P. and Gaur, R.K., 2011. Biotechnological perspectives of microbes in agro-ecosystems. *Biotechnology letters*, 33(10), pp.1905-1910.
- Clarkson, D.T. and Hanson, J.B., 1980. The mineral nutrition of higher plants. *Annual review of plant physiology*, 31(1), pp.239-298.
- Cleland, R.E., 2010. Auxin and cell elongation. In *Plant hormones* (pp. 204-220). Springer Netherlands.
- Davies, P.J., 2010. The plant hormones: their nature, occurrence, and functions (pp. 1-15). Springer Netherlands.
- Day, J.M. and Döbereiner, J., 1976. Physiological aspects of N<sub>2</sub>-fixation by a *Spirillum* from *Digitaria* roots. *Soil Biology and Biochemistry*, 8(1), pp.45-50.
- de Freitas, J.R., 2000. Yield and N assimilation of winter wheat (*Triticum aestivum* L., var. Norstar) inoculated with rhizobacteria. *Pedobiologia*, 44(2), pp.97-104.
- de Souza, R., Beneduzi, A., Ambrosini, A., Da Costa, P.B., Meyer, J., Vargas, L.K., Schoenfeld, R. and Passaglia, L.M., 2013. The effect of plant growth-promoting rhizobacteria on the growth of rice (*Oryza sativa* L.) cropped in southern Brazilian fields. *Plant and soil*, 366(1-2), pp.585-603.
- Demetre, L., J.R.M. Zandile, P.S. Nelia, G. Gerda, M.W.M. Eleni, D.D. Yul and P. Whadi-ah, 2011. Food Security in South Africa: A review of national surveys. *Bulletin of the World Health Organization*, 89: 891-899.
- Department of Agriculture (DoA) Peninsular Malaysia, 2014. Paddy Statistics of Malaysia, 2015.



- Department of Agriculture (DoA) Peninsular Malaysia, 2015. Paddy Statistics of Malaysia, 2014.  
[www.doa.gov.my/index/resources/aktiviti\\_sumber/.../perangkaan\\_padi\\_2014.pdf](http://www.doa.gov.my/index/resources/aktiviti_sumber/.../perangkaan_padi_2014.pdf).
- Desai, A. and Archana, G., 2011. Role of siderophores in crop improvement. In *Bacteria in Agrobiolgy: Plant Nutrient Management* (pp. 109-139). Springer Berlin Heidelberg.
- Deubel, A. and Merbach, W., 2005. Influence of microorganisms on phosphorus bioavailability in soils. In *Microorganisms in soils: roles in genesis and functions* (pp. 177-191). Springer Berlin Heidelberg.
- Dhillon, S.S., 1992. Dual inoculation of pretransplant stage *Oryza sativa* L. plants with indigenous vesicular-arbuscular mycorrhizal fungi and fluorescent *Pseudomonas* spp. *Biology and Fertility of soils*, 13(3), pp.147-151.
- Dimkpa, C., Svatoš, A., Merten, D., Büchel, G. and Kothe, E., 2008. Hydroxamate siderophores produced by *Streptomyces acidiscabies* E13 bind nickel and promote growth in cowpea (*Vigna unguiculata* L.) under nickel stress. *Canadian journal of microbiology*, 54(3), pp.163-172.
- Dimkpa, C.O., Merten, D., Svatoš, A., Büchel, G. and Kothe, E., 2009. Siderophores mediate reduced and increased uptake of cadmium by *Streptomyces tendae* F4 and sunflower (*Helianthus annuus*), respectively. *Journal of applied microbiology*, 107(5), pp.1687-1696.
- Dobbelaere, S. and Okon, Y., 2007. The plant growth-promoting effect and plant responses. In *Associative and endophytic nitrogen-fixing bacteria and cyanobacterial associations* (pp. 145-170). Springer Netherlands.
- Dobbelaere, S., Vanderleyden, J. and Okon, Y., 2003. Plant growth-promoting effects of diazotrophs in the rhizosphere. *Critical reviews in plant sciences*, 22(2), pp.107-149.
- Döbereiner, J., 1997. Biological nitrogen fixation in the tropics: social and economic contributions. *Soil Biology and Biochemistry*, 29(5), pp.771-774.
- Döbereiner, J., Reis, V.M., Paula, M.A. and Olivares, F.D., 1993. Endophytic diazotrophs in sugar cane, cereals and tuber plants. In *New horizons in nitrogen fixation* (pp. 671-676). Springer Netherlands.

- Dogan, K., Celik, I., Gok, M. and Coskan, A., 2011. Effect of different soil tillage methods on rhizobial nodulation, biomass and nitrogen content of second crop soybean. *African Journal of Microbiology Research*, 5(20), pp.3186-3194.
- dos Santos, C.L.R., Alves, G.C., de Matos Macedo, A.V., Giori, F.G., Pereira, W., Urquiaga, S. and Reis, V.M., 2017. Contribution of a mixed inoculant containing strains of *Burkholderia* spp. and *Herbaspirillum* ssp. to the growth of three sorghum genotypes under increased nitrogen fertilization levels. *Applied Soil Ecology*, 113, pp.96-106.
- Duarah, I., Deka, M., Saikia, N. and Boruah, H.D., 2011. Phosphate solubilizers enhance NPK fertilizer use efficiency in rice and legume cultivation. *3 Biotech*, 1(4), pp.227-238.
- Dubey, S.K., 1996. Combined effect of *Bradyrhizobium japonicum* and phosphate-solubilizing *Pseudomonas striata* on nodulation, yield attributes and yield of rainfed soybean (*Glycine max*) under different sources of phosphorus in Vertisols. *Indian journal of Agricultural Sciences*, 66(1), pp.28-32.
- Duhoon, S.S., Jain, H.C., Deshmukh, M.R. and Goswami, U., 2001. Integrated nutrient management in kharif sesame (*Sesamum indicum* L.). *Journal of Oilseeds Research*, 18(1), pp.81-84.
- Eiguchi, M. and Sano, Y., 1990. A gene complex responsible for seed shattering and panicle spreading found in common wild rices. *Rice Genetics Newsletter*, 7, pp.105-107.
- Ekin, Z., 2010. Performance of phosphate solubilizing bacteria for improving growth and yield of sunflower (*Helianthus annuus* L.) in the presence of phosphorus fertilizer. *African Journal of Biotechnology*, 9(25), pp.3794-3800.
- El-Fattah, D.A.A., Eweda, W.E., Zayed, M.S. and Hassanein, M.K., 2013. Effect of carrier materials, sterilization method, and storage temperature on survival and biological activities of *Azotobacter chroococcum* inoculant. *Annals of Agricultural Sciences*, 58(2), pp.111-118.
- Elkoca, E., Turan, M. and Donmez, M.F., 2010. Effects of single, dual and triple inoculations with *Bacillus subtilis*, *Bacillus megaterium* and *Rhizobium leguminosarum* bv. Phaseoli on nodulation, nutrient uptake, yield and yield parameters of common bean (*Phaseolus vulgaris* L. cv.'elkoca-05'). *Journal of plant nutrition*, 33(14), pp.2104-2119.
- Emmert, E.A. and Handelsman, J., 1999. Biocontrol of plant disease: a (Gram-) positive perspective. *FEMS Microbiology letters*, 171(1), pp.1-9.

- Enebak, S.A. and Carey, W.A., 2000. Evidence for induced systemic protection to fusiform rust in loblolly pine by plant growth-promoting rhizobacteria. *Plant Disease*, 84(3), pp.306-308.
- Etesami, H. and Alikhani, H.A., 2016. Rhizosphere and endorhiza of oilseed rape (*Brassica napus* L.) plant harbor bacteria with multifaceted beneficial effects. *Biological Control*, 94, pp.11-24.
- Etesami, H., Alikhani, H.A. and Hosseini, H.M., 2015. Indole-3-acetic acid (IAA) production trait, a useful screening to select endophytic and rhizosphere competent bacteria for rice growth promoting agents. *MethodsX*, 2, pp.72-78.
- FAO, 2003. International Rice Commission Newsletters. Dat Van Tran, Food and Agriculture Organization. Rome, Italy, Vol. 52, ISSN 0538-9550.
- FAO, I., IMF, O. and UNCTAD, W., 2011. the World Bank, the WTO, IFPRI and the UN HLTF (2011). *Price Volatility in Food and Agricultural Markets: Policy Responses*. Rome, FAO.
- FAO, U., 2014. FAOstat. Retrieved Feb, 2014.
- FAO, W.F.P., 2012. IFAD. 2012. *The state of food insecurity in the world*, pp.1-63.
- FAOSTAT, 2017. <http://www.fao.org/faostat/en/#data>. Accessed 8 September 2017.
- Fukui, R., Schroth, M.N., Henderson, M., Hancock, J.G. and Firestone, M.K., 1994. Growth patterns and metabolic activity of pseudomonads in sugar beet spermospheres: relationship to pericarp colonization by *Pythium ultimum*. *Phytopathology*, 84(11), pp.1331-1337.
- García-Fraile, P., Menéndez, E. and Rivas, R., 2015. Role of bacterial biofertilizers in agriculture and forestry.
- García-Fraile, P., Menéndez, E. and Rivas, R., 2015. Role of bacterial biofertilizers in agriculture and forestry.
- Ghosh, T.K. and Saha, K.C., 1996. Effects of inoculation of cyanobacteria on nitrogen status and nutrition of rice (*Oryza sativa* L.) in an Entisol amended with chemical and organic sources of nitrogen. *Biology and Fertility of Soils*, 24(1), pp.123-128.
- Glass, A.D., 1989. *Plant mineral nutrition. An introduction to current concepts*. Jones and Bartlett Publishers, Inc..
- Glendinning, J.S., 2000. *Australian soil fertility manual*. CSIRO Publishing.

- Glick, B.R., 1995. The enhancement of plant growth by free-living bacteria. *Canadian Journal of Microbiology*, 41(2), pp.109-117.
- Glick, B.R., 2012. Plant growth-promoting bacteria: mechanisms and applications. *Scientifica*, 2012.
- Glick, B.R., Patten, C.L., Holguin, G. and Penrose, D.M., 1999. Biochemical and genetic mechanisms used by plant growth promoting bacteria. *World Scientific*.
- Glick, B.R., Todorovic, B., Czarny, J., Cheng, Z., Duan, J. and McConkey, B., 2007. Promotion of plant growth by bacterial ACC deaminase. *Critical Reviews in Plant Sciences*, 26(5-6), pp.227-242.
- Goldstein, A.H., 1994. Involvement of the quinoprotein glucose dehydrogenase in the solubilization of exogenous phosphates by gram-negative bacteria. *Phosphate in microorganisms: cellular and molecular biology*. ASM Press, Washington, DC, pp.197-203.
- Gopalakrishnan, S., Srinivas, V., Alekhya, G. and Prakash, B., 2015. Effect of plant growth-promoting *Streptomyces* sp. on growth promotion and grain yield in chickpea (*Cicer arietinum* L). *3 Biotech*, 5(5), pp.799-806.
- Gordon, S.A. and Weber, R.P., 1951. Colorimetric estimation of indoleacetic acid. *Plant Physiology*, 26(1), p.192.
- Gray, E.J. and Smith, D.L., 2005. Intracellular and extracellular PGPR: commonalities and distinctions in the plant–bacterium signaling processes. *Soil Biology and Biochemistry*, 37(3), pp.395-412.
- Guo, J., Tang, S., Ju, X., Ding, Y., Liao, S. and Song, N., 2011. Effects of inoculation of a plant growth promoting rhizobacterium *Burkholderia* sp. D54 on plant growth and metal uptake by a hyperaccumulator *Sedum alfredii* Hance grown on multiple metal contaminated soil. *World Journal of Microbiology and Biotechnology*, 27(12), pp.2835-2844.
- Gupta, A. and Sen, S., 2013. Role of biofertilisers and biopesticides for sustainable agriculture, scholar. google. com.
- Gyaneshwar, P., Parekh, L.J., Archana, G., Poole, P.S., Collins, M.D., Hutson, R.A. and Kumar, G.N., 1999. Involvement of a phosphate starvation inducible glucose dehydrogenase in soil phosphate solubilization by *Enterobacter asburiae*. *FEMS microbiology letters*, 171(2), pp.223-229.

- Hagen, G., 1987. The control of gene expression by auxin. In *Plant hormones and their role in plant growth and development* (pp. 149-163). Springer Netherlands.
- Hameeda, B., Harini, G., Rupela, O.P., Wani, S.P. and Reddy, G., 2008. Growth promotion of maize by phosphate-solubilizing bacteria isolated from composts and macrofauna. *Microbiological Research*, 163(2), pp.234-242.
- Hamilton, M., Hess, R., Ward, T., White, G., Hoskinson, R., Cook, L., USU, B.B. and Siegel, L., 2001. Investigation of Factors Influencing Cesium Mobility and Uptake in Plant/Soil Systems. *Environmental Systems Research Candidates Final Report FY 2000-2001*, p.179.
- Harlan, J.R., 1992. Crops and man (No. Ed. 2). *American Society of Agronomy*.
- Harvey, J.M., 1978. Reduction of losses in fresh market fruits and vegetables. *Annual Review of Phytopathology*, 16(1), pp.321-341.
- Hayat, R., Ali, S., Amara, U., Khalid, R. and Ahmed, I., 2010. Soil beneficial bacteria and their role in plant growth promotion: a review. *Annals of Microbiology*, 60(4), pp.579-598.
- Hecht-Buchholz, C., 1998. The apoplast-habitat of endophytic dinitrogen-fixing bacteria and their significance for the nitrogen nutrition of nonleguminous plants. *Zeitschrift für Pflanzenernährung und Bodenkunde*, 161(5), pp.509-520.
- Hinsinger, P., 2001. Bioavailability of soil inorganic P in the rhizosphere as affected by root-induced chemical changes: a review. *Plant and soil*, 237(2), pp.173-195.
- Hobson, G.E., 1988. Pre-and post-harvest strategies in the production of high quality tomato fruit. *Applied agricultural research (USA)*.
- Hodge, A., 2004. The plastic plant: root responses to heterogeneous supplies of nutrients. *New phytologist*, 162(1), pp.9-24.
- Howard, J.B. and Rees, D.C., 1996. Structural basis of biological nitrogen fixation. *Chemical Reviews*, 96(7), pp.2965-2982.
- <http://www.chemguide.co.uk/physical/equilibria/haber.html>
- [http://www.dairytas.com.au/files/nrm/nutrients/fertsmart\\_info\\_sheet\\_10\\_phosphorus\\_tas\\_fact\\_sheet.pdf](http://www.dairytas.com.au/files/nrm/nutrients/fertsmart_info_sheet_10_phosphorus_tas_fact_sheet.pdf)
- <http://www.galaxypetrochem.in/indole-3-acetic-acid.htm>
- <https://microbewiki.kenyon.edu/index.php/File:Rhizosphere-Bacteria.jpg>

<https://microbewiki.kenyon.edu/index.php/File:Rhizosphere-Bacteria.jpg>

Hu, X., Chen, J. and Guo, J., 2006. Two phosphate-and potassium-solubilizing bacteria isolated from Tianmu Mountain, Zhejiang, China. *World journal of Microbiology and Biotechnology*, 22(9), pp.983-990.

Hutsch, B.W., Augustin, J. and Merbach, W., 2002. Plant rhizodeposition-an important source for carbon turnover in soils. *Journal of Plant Nutrition and Soil Science*, 165(4), p.397.

Iglesias, A., Erda, L. and Rosenzweig, C., 1996. Climate change in Asia: a review of the vulnerability and adaptation of crop production. *Water, Air, and Soil Pollution*, 92(1-2), pp.13-27.

Ilyas, N., Bano, A., Iqbal, S. and Raja, N.I., 2012. Physiological, biochemical and molecular characterization of *Azospirillum* spp. isolated from maize under water stress. *Pakistan Journal of Botany*, 44, pp.71-80.

International Rice Research Institute (IRRI), 2003. Potassium (K). Rice fact sheet. Rice science for a better world, 2003

IRRI, 2004. World Rice Statistics. Retrieved from: [http://www.irri.org/index.php?option=com\\_k2&view=itemlist&layout=category&task=category&id=744&Itemid=100346&lang=en](http://www.irri.org/index.php?option=com_k2&view=itemlist&layout=category&task=category&id=744&Itemid=100346&lang=en).

Jackson, M.B., 1991. Ethylene in root growth and development. *The plant hormone ethylene*, 159181.

Jha, Y., Subramanian, R.B. and Patel, S., 2011. Combination of endophytic and rhizospheric plant growth promoting rhizobacteria in *Oryza sativa* shows higher accumulation of osmoprotectant against saline stress. *Acta physiologiae plantarum*, 33(3), pp.797-802.

Ji, H.S., Chu, S.H., Jiang, W., Cho, Y.I., Hahn, J.H., Eun, M.Y., McCouch, S.R. and Koh, H.J., 2006. Characterization and mapping of a shattering mutant in rice that corresponds to a block of domestication genes. *Genetics*, 173(2), pp.995-1005.

Joe, M.M., Jaleel, C.A., Sivakumar, P.K., Zhao, C.X. and Karthikeyan, B., 2009. Co-aggregation in *Azospirillum brasilense* MTCC-125 with other PGPR strains: Effect of physical and chemical factors and stress endurance ability. *Journal of the Taiwan Institute of Chemical Engineers*, 40(5), pp.491-499.

Katiyar, V. and Goel, R., 2004. Siderophore mediated plant growth promotion at low temperature by mutant of fluorescent pseudomonad\*. *Plant growth regulation*, 42(3), pp.239-244.

- Katsura, K., Maeda, S., Horie, T. and Shiraiwa, T., 2007. Analysis of yield attributes and crop physiological traits of Liangyoupeijiu, a hybrid rice recently bred in China. *Field Crops Research*, 103(3), pp.170-177.
- Kawaguchi, K. and Kyuma, K., 1974. Paddy soils in tropical Asia. *Southeast Asian Studies*, 12(1), pp.3-24.
- Khan, A.L., Waqas, M., Kang, S.M., Al-Harrasi, A., Hussain, J., Al-Rawahi, A., Al-Khiziri, S., Ullah, I., Ali, L., Jung, H.Y. and Lee, I.J., 2014. Bacterial endophyte *Shingomonas* sp. LK11 produces gibberellins and IAA and promotes tomato plant growth. *Journal of Microbiology*, 52(8), pp.689-695.
- Kim, K.Y., Jordan, D. and McDonald, G.A., 1998. Enterobacter agglomerans, phosphate solubilizing bacteria, and microbial activity in soil: effect of carbon sources. *Soil Biology and Biochemistry*, 30(8), pp.995-1003.
- Kloepper, J.W., Gutierrez-Estrada, A. and McInroy, J.A., 2007. Photoperiod regulates elicitation of growth promotion but not induced resistance by plant growth-promoting rhizobacteria. *Canadian Journal of Microbiology*, 53(2), pp.159-167.
- Kloepper, J.W., Lifshitz, R. and Zablotowicz, R.M., 1989. Free-living bacterial inocula for enhancing crop productivity. *Trends in biotechnology*, 7(2), pp.39-44.
- Konishi, S., Izawa, T., Lin, S.Y., Ebana, K., Fukuta, Y., Sasaki, T. and Yano, M., 2006. An SNP caused loss of seed shattering during rice domestication. *Science*, 312(5778), pp.1392-1396.
- Kucey, R.M.N., 1987. Increased phosphorus uptake by wheat and field beans inoculated with a phosphorus-solubilizing *Penicillium bilaji* strain and with vesicular-arbuscular mycorrhizal fungi. *Applied and Environmental Microbiology*, 53(12), pp.2699-2703.
- Kudoyarova, G.R., Melentiev, A.I., Martynenko, E.V., Timergalina, L.N., Arkhipova, T.N., Shendel, G.V., Kuz'mina, L.Y., Dodd, I.C. and Veselov, S.Y., 2014. Cytokinin producing bacteria stimulate amino acid deposition by wheat roots. *Plant Physiology and Biochemistry*, 83, pp.285-291.
- Kumar, A., Maurya, B.R. and Raghuvanshi, R., 2014. Isolation and characterization of PGPR and their effect on growth, yield and nutrient content in wheat (*Triticum aestivum* L.). *Biocatalysis and Agricultural Biotechnology*, 3(4), pp.121-128.

- Kumar, A., Saini, S., Prakash, A. and Johri, B.N., 2009. Influence of cultivation practices on phenotypic and genotypic diversity of antagonistic rhizobacteria isolated from soybean (*Glycine max* L.). In: Abstracts, 1st Asian PGPR Congress for Sustainable Agriculture, Hyderabad. 118.
- Kumar, M., Singh, D.P., Prabha, R., Rai, A.K. and Sharma, L., 2016. Role of Microbial Inoculants in Nutrient Use Efficiency. In *Microbial Inoculants in Sustainable Agricultural Productivity* (pp. 133-142). Springer India.
- Kumar, S., Pandey, P. and Maheshwari, D.K., 2009. Reduction in dose of chemical fertilizers and growth enhancement of sesame (*Sesamum indicum* L.) with application of rhizospheric competent *Pseudomonas aeruginosa* LES4. *European Journal of Soil Biology*, 45(4), pp.334-340.
- Kumar, V. and Narula, N., 1999. Solubilization of inorganic phosphates and growth emergence of wheat as affected by *Azotobacter chroococcum* mutants. *Biology and Fertility of Soils*, 28(3), pp.301-305.
- Ladha, J.K. and Reddy, P.M., 2003. Nitrogen fixation in rice systems: state of knowledge and future prospects. *Plant and Soil*, 252(1), pp.151-167.
- Lambers, H., Mougel, C., Jaillard, B. and Hinsinger, P., 2009. Plant-microbe-soil interactions in the rhizosphere: an evolutionary perspective. *Plant and Soil*, 321(1-2), pp.83-115.
- Lee, K.J., Kamala-Kannan, S., Sub, H.S., Seong, C.K. and Lee, G.W., 2008. Biological control of Phytophthora blight in red pepper (*Capsicum annuum* L.) using *Bacillus subtilis*. *World journal of microbiology and biotechnology*, 24(7), pp.1139-1145.
- Lee, S.W. and Huh, Y.K., 1984. Threshing and cutting forces for Korean rice. *Transactions of the ASAE*, 27(6), pp.1654-1657.
- Lenin, G. and Jayanthi, M., 2012. Indoleacetic acid, gibberellic acid and siderophore production by PGPR isolates from rhizospheric Soils of *Catharanthus roseus*. *International Journal of Pharmaceutical and Biological Archive*, 3, pp.933-8.
- Li, Q., Saleh-Lakha, S. and Glick, B.R., 2005. The effect of native and ACC deaminase-containing *Azospirillum brasilense* Cd1843 on the rooting of carnation cuttings. *Canadian Journal of Microbiology*, 51(6), pp.511-514.



- Lian, B., Fu, P.Q., Mo, D.M. and Liu, C.Q., 2002. A comprehensive review of the mechanism of potassium releasing by silicate bacteria. *Acta Mineralogica Sinica*, 22(2), pp.179-183.
- Lifshitz, R., Kloepper, J.W., Scher, F.M., Tipping, E.M. and Laliberté, M., 1986. Nitrogen-fixing pseudomonads isolated from roots of plants grown in the Canadian High Arctic. *Applied and Environmental Microbiology*, 51(2), pp.251-255.
- Lin, Z., Griffith, M.E., Li, X., Zhu, Z., Tan, L., Fu, Y., Zhang, W., Wang, X., Xie, D. and Sun, C., 2007. Origin of seed shattering in rice (*Oryza sativa* L.). *Planta*, 226(1), pp.11-20.
- Liu, D., Lian, B. and Dong, H., 2012. Isolation of *Paenibacillus* sp. and assessment of its potential for enhancing mineral weathering. *Geomicrobiology Journal*, 29(5), pp.413-421.
- Liu, F., Xing, S., Ma, H., Du, Z. and Ma, B., 2013. Cytokinin-producing, plant growth-promoting rhizobacteria that confer resistance to drought stress in *Platycladus orientalis* container seedlings. *Applied microbiology and biotechnology*, 97(20), pp.9155-9164.
- Liu, S.T., Lee, L.Y., Tai, C.Y., Hung, C.H., Chang, Y.S., Wolfram, J.H., Rogers, R. and Goldstein, A.H., 1992. Cloning of an *Erwinia herbicola* gene necessary for gluconic acid production and enhanced mineral phosphate solubilization in *Escherichia coli* HB101: nucleotide sequence and probable involvement in biosynthesis of the coenzyme pyrroloquinoline quinone. *Journal of Bacteriology*, 174(18), pp.5814-5819.
- Lucas, J.A., García-Cristobal, J., Bonilla, A., Ramos, B. and Gutierrez-Manero, J., 2014. Beneficial rhizobacteria from rice rhizosphere confers high protection against biotic and abiotic stress inducing systemic resistance in rice seedlings. *Plant Physiology and Biochemistry*, 82, pp.44-53.
- Lucy, M., Reed, E. and Glick, B.R., 2004. Applications of free living plant growth-promoting rhizobacteria. *Antonie van Leeuwenhoek*, 86(1), pp.1-25.
- Lugtenberg, B. and Kamilova, F., 2009. Plant-growth-promoting rhizobacteria. *Annual review of microbiology*, 63, pp.541-556.
- Lugtenberg, B.J., Dekkers, L. and Bloemberg, G.V., 2001. Molecular determinants of rhizosphere colonization by *Pseudomonas*. *Annual review of phytopathology*, 39(1), pp.461-490.

- Luo, J.Y., Xie, G.L., Li, B., Luo, Y.C., Zhao, L.H., Wang, X., Liu, B. and Li, W., 2005. Gram-positive bacteria associated with rice in China and their antagonists against the pathogens of sheath blight and bakanae disease in rice. *Rice Science*, 12(3), pp.213-218.
- Lynch, J.M. and Whipps, J.M., 1991. Substrate flow in the rhizosphere. In *The rhizosphere and plant growth* (pp. 15-24). Springer Netherlands.
- Mäder, P., Kaiser, F., Adholeya, A., Singh, R., Uppal, H.S., Sharma, A.K., Srivastava, R., Sahai, V., Aragno, M., Wiemken, A. and Johri, B.N., 2011. Inoculation of root microorganisms for sustainable wheat-rice and wheat-black gram rotations in India. *Soil Biology and Biochemistry*, 43(3), pp.609-619.
- Madhaiyan, M., Poonguzhali, S., Kang, B.G., Lee, Y.J., Chung, J.B. and Sa, T.M., 2010. Effect of co-inoculation of methylotrophic *Methylobacterium oryzae* with *Azospirillum brasilense* and *Burkholderia pyrrocinia* on the growth and nutrient uptake of tomato, red pepper and rice. *Plant and soil*, 328(1-2), pp.71-82.
- Mahmud, Y. and Yahya, B.T., 2001. Crop diversification in Malaysia. *Crop diversification in the Asia-Pacific Region*. Editors: MK Papademetriou and FJ Dent, Food and Agricultural Organization Regional Office, Bangkok, Thailand.
- Maksimov, I.V., Abizgil'Dina, R.R. and Pusenkova, L.I., 2011. Plant growth promoting rhizobacteria as alternative to chemical crop protectors from pathogens (review). *Applied Biochemistry and Microbiology*, 47(4), pp.333-345.
- Marschner, H., 2012. Marschner's mineral nutrition of higher plants, vol. 89.
- Masalha, J., Kosegarten, H., Elmaci, Ö. and Mengel, K., 2000. The central role of microbial activity for iron acquisition in maize and sunflower. *Biology and Fertility of Soils*, 30(5), pp.433-439.
- Mayak, S., Tirosh, T. and Glick, B.R., 2001. Stimulation of the growth of tomato, pepper and mung bean plants by the plant growth-promoting bacterium *Enterobacter cloacae* CAL3. *Biological agriculture & horticulture*, 19(3), pp.261-274.
- McArtney, S.J., Tustin, D.S., Seymour, S., Cashmore, W. and Looney, N.E., 1995. Benzyladenine and carbaryl effects on fruit thinning and the enhancement of return flowering of three apple cultivars. *Journal of Horticultural Science*, 70(2), pp.287-296.
- McManus, M.T., Thompson, D.S., Merriman, C., Lyne, L. and Osborne, D.J., 1998. Transdifferentiation of mature cortical cells to functional abscission cells in bean. *Plant Physiology*, 116(3), pp.891-899.

- Mehdipour Moghadam, M.J., Emtiazi, G. and Salehi, Z., 2012. Enhanced auxin production by *Azospirillum* pure cultures from plant root exudates. *Journal of Agricultural Science and Technology*, 14(5), pp.985-994.
- Mehnaz, S. and Lazarovits, G., 2006. Inoculation effects of *Pseudomonas putida*, *Gluconacetobacter azotocaptans*, and *Azospirillum lipoferum* on corn plant growth under greenhouse conditions. *Microbial Ecology*, 51(3), pp.326-335.
- Meir, S., Philosoph-Hadas, S., Sundaresan, S., Selvaraj, K.V., Burd, S., Ophir, R., Kochanek, B., Reid, M.S., Jiang, C.Z. and Lers, A., 2010. Microarray analysis of the abscission-related transcriptome in the tomato flower abscission zone in response to auxin depletion. *Plant Physiology*, 154(4), pp.1929-1956.
- Melkamu, M., Seyoum, T. and Woldetsadik, K., 2008. Effects of pre-and post harvest treatments on changes in sugar content of tomato. *African Journal of Biotechnology*, 7(8).
- Mia, M.B., Shamsuddin, Z.H. and Mahmood, M., 2012. Effects of rhizobia and plant growth promoting bacteria inoculation on germination and seedling vigor of lowland rice. *African Journal of Biotechnology*, 11(16), pp.3758-3765.
- Miransari, M. and Smith, D.L., 2009. Alleviating salt stress on soybean (*Glycine max* (L.) Merr.)–*Bradyrhizobium japonicum* symbiosis, using signal molecule genistein. *European Journal of Soil Biology*, 45(2), pp.146-152.
- MOA, 2008. Ministry of agriculture and agro base industry of Malaysia. KADA Annual Report, Malaysia Putrajaya.
- MOA, 2011. National Agro-Food Policy 2011-2020. Division of International and Strategic Planning. Putrajaya, Malaysia, ISBN: 978-983-9863-41-3
- Mohapatra, B., Verma, D.K., Sen, A., Panda, B.B. and Asthir, B., 2013. Bio-fertilizer- A Getway to Sustainable Agriculture, *Pop Kheti*. 1(4):97-106.
- Morgan, P.W., 1984. Is ethylene the natural regulator of abscission?. In *Ethylene* (pp. 231-240). Springer Netherlands.
- Muraleedharan, H., Seshadri, S. and Perumal, K., 2010. Biofertilizer (Phosphobacteria), Shri AMM Murugappa Chettiar Research Centre. *Taramani, Chennai-600113*.

- Nadeem, S.M., Naveed, M., Zahir, Z.A. and Asghar, H.N., 2013. Plant–microbe interactions for sustainable agriculture: fundamentals and recent advances. In *Plant Microbe Symbiosis: Fundamentals and Advances* (pp. 51-103). Springer India.
- Nagao, S. and Takahashi, M.E., 1963. Trial Construction of Twelve Linkage Groups in Japanese Rice:(Genetical Studies on Rice Plant, XXVII). *Journal of the Faculty of Agriculture, Hokkaido University= 北海道大學農學部紀要*, 53(1), pp.72-130.
- Nahas, E., 1996. Factors determining rock phosphate solubilization by microorganisms isolated from soil. *World Journal of Microbiology and Biotechnology*, 12(6), pp.567-572.
- Nain, L., Rana, A., Joshi, M., Jadhav, S.D., Kumar, D., Shivay, Y.S., Paul, S. and Prasanna, R., 2010. Evaluation of synergistic effects of bacterial and cyanobacterial strains as biofertilizers for wheat. *Plant and soil*, 331(1-2), pp.217-230.
- Najim, M.M.M., Lee, T.S., Haque, M.A. and Esham, M., 2007. Sustainability of rice production: A Malaysian perspective.
- Najim, M.M.M., Lee, T.S., Haque, M.A. and Esham, M., 2007. Sustainability of rice production: A Malaysian perspective.
- Nautiyal, C.S., Bhadauria, S., Kumar, P., Lal, H., Mondal, R. and Verma, D., 2000. Stress induced phosphate solubilization in bacteria isolated from alkaline soils. *FEMS Microbiology Letters*, 182(2), pp.291-296.
- Nehra, K., Yadav, A.S., Sehrawat, A.R. and Vashishat, R.K., 2007. Characterization of heat resistant mutant strains of *Rhizobium* sp.[Cajanus] for growth, survival and symbiotic properties. *Indian Journal of Microbiology*, 47(4), pp.329-335.
- Nelson, D.R., Bellville, R.J. and Porter, C.A., 1984. Role of nitrogen assimilation in seed development of soybean. *Plant physiology*, 74(1), pp.128-133.
- Nihorimbere, V., Ongena, M., Smargiassi, M. and Thonart, P., 2011. Beneficial effect of the rhizosphere microbial community for plant growth and health. *Biotechnologie, Agronomie, Société et Environnement*, 15(2), p.327.
- Nur Hasnidar, M.K., and Halimi Mohd Saud. 2011. Isolation and screening for nitrogen-fixing and mineral phosphate-solubilizing ability in rhizobia of legume plants. In: Proceedings of The Soil 2011 Conference, Sabah, Malaysia, pp. 343 – 346.

- Nwugo, C.C. and Huerta, A.J., 2008. Effects of silicon nutrition on cadmium uptake, growth and photosynthesis of rice plants exposed to low-level cadmium. *Plant and Soil*, 311(1-2), pp.73-86.
- Oerke, E. C., 2006. Crop losses to pests. *J Agric Sci* 144:31–43
- Oerke, E.C., Dehne, H.W., Schönbeck, F. and Weber, A., 2012. *Crop production and crop protection: estimated losses in major food and cash crops*. Elsevier.
- Okon, Y. and Labandera-Gonzalez, C.A., 1994. Agronomic applications of *Azospirillum*: an evaluation of 20 years worldwide field inoculation. *Soil Biology and Biochemistry*, 26(12), pp.1591-1601.
- Ongena, M.A.R.C. and Thonart, P., 2006. Resistance induced in plants by non-pathogenic microorganisms: elicitation and defense responses. *Floriculture, ornamental and plant biotechnology: advances and topical issues*, pp.447-463.
- Osborne, D.J. and McManus, M.T., 1984. Abscission and the recognition of zone specific target cells. The role of ethylene. In *Ethylene* (pp. 221-230). Springer Netherlands.
- Osborne, D.J., 1989. Abscission. C.R.C. *Critical Rev Plant Sci* 8: 103–129.
- Ozturk, A., Caglar, O. and Sahin, F., 2003. Yield response of wheat and barley to inoculation of plant growth promoting rhizobacteria at various levels of nitrogen fertilization. *Journal of Plant Nutrition and Soil Science*, 166(2), pp.262-266.
- Paddy Statistics of Malaysia 2014., 2015. Department of Agriculture (DOA). [www.doa.gov.my/index/resources/aktiviti\\_sumber/.../perangkaan\\_padi\\_2014.pdf](http://www.doa.gov.my/index/resources/aktiviti_sumber/.../perangkaan_padi_2014.pdf).
- Pal, S.S., 1998. Interactions of an acid tolerant strain of phosphate solubilizing bacteria with a few acid tolerant crops. *Plant and soil*, 198(2), pp.169-177.
- Panhwar, Q.A., Othman, R., Rahman, Z.A., Meon, S. and Ismail, M.R., 2012. Isolation and characterization of phosphate-solubilizing bacteria from aerobic rice. *African Journal of Biotechnology*, 11(11), pp.2711-2719.
- Parmar, N. and Dufresne, J., 2011. Beneficial interactions of plant growth promoting rhizosphere microorganisms. In *Bioaugmentation, biostimulation and biocontrol* (pp. 27-42). Springer Berlin Heidelberg.
- Patterson, S.E., 2001. Cutting loose. Abscission and dehiscence in *Arabidopsis*. *Plant Physiology*, 126(2), pp.494-500.

- Pedraza, R.O., Motok, J., Salazar, S.M., Ragout, A.L., Mentel, M.I., Tortora, M.L., Guerrero-Molina, M.F., Winik, B.C. and Díaz-Ricci, J.C., 2010. Growth-promotion of strawberry plants inoculated with *Azospirillum brasilense*. *World Journal of Microbiology and Biotechnology*, 26(2), pp.265-272.
- Peng, S., Tang, Q. and Zou, Y., 2009. Current status and challenges of rice production in China. *Plant Production Science*, 12(1), pp.3-8.
- Pikovskaya, R.I., 1948. Mobilization of phosphorus in soil in connection with vital activity of some microbial species. *Mikrobiologiya*, 17, p.p 362-370.
- Poonguzhali, S., Madhaiyan, M. and Sa, T., 2008. Isolation and identification of phosphate solubilizing bacteria from chinese cabbage and their effect on growth and phosphorus utilization of plants. *Journal of microbiology and biotechnology*, 18(4), pp.773-777.
- Poorter, H., Niklas, K.J., Reich, P.B., Oleksyn, J., Poot, P. and Mommer, L., 2012. Biomass allocation to leaves, stems and roots: meta-analyses of interspecific variation and environmental control. *New Phytologist*, 193(1), pp.30-50.
- Project, I.R.G.S., 2005. The map-based sequence of the rice genome. *Nature*, 436(7052), pp.793-800.
- Qin, H., Lu, K., Strong, P.J., Xu, Q., Wu, Q., Xu, Z., Xu, J. and Wang, H., 2015. Long-term fertilizer application effects on the soil, root arbuscular mycorrhizal fungi and community composition in rotation agriculture. *Applied Soil Ecology*, 89, pp.35-43.
- Raaijmakers, J.M., Vlami, M. and De Souza, J.T., 2002. Antibiotic production by bacterial biocontrol agents. *Antonie van Leeuwenhoek*, 81(1), pp.537-547.
- Rabalais, N.N., Turner, R.E., Wiseman Jr, W.J. and Dortch, Q., 1998. Consequences of the 1993 Mississippi River flood in the Gulf of Mexico. *River Research and Applications*, 14(2), pp.161-177.
- Radzki, W., Mañero, F.G., Algar, E., García, J.L., García-Villaraco, A. and Solano, B.R., 2013. Bacterial siderophores efficiently provide iron to iron-starved tomato plants in hydroponics culture. *Antonie Van Leeuwenhoek*, 104(3), pp.321-330.
- Rathi, N., Singh, S., Osbone, J. and Babu, S., 2015. Co-aggregation of *Pseudomonas fluorescens* and *Bacillus subtilis* in culture and co-colonization in black gram (*Vigna mungo* L.) roots. *Biocatalysis and Agricultural Biotechnology*, 4(3), pp.304-308.

- Raupach, G.S. and Kloepper, J.W., 2000. Biocontrol of cucumber diseases in the field by plant growth-promoting rhizobacteria with and without methyl bromide fumigation. *Plant Disease*, 84(10), pp.1073-1075.
- Reid, M.S., 1988, March. The role of ethylene in flower senescence. In *IV International Symposium on Postharvest Physiology of Ornamental Plants* 261 (pp. 157-170).
- Revillas, J.J., Rodelas, B., Pozo, C., Martínez-Toledo, M.V. and González-López, J., 2000. Production of B-group vitamins by two *Azotobacter* strains with phenolic compounds as sole carbon source under diazotrophic and adiazotrophic conditions. *Journal of Applied Microbiology*, 89(3), pp.486-493.
- Richardson, A.E., Barea, J.M., McNeill, A.M. and Prigent-Combaret, C., 2009. Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. *Plant and soil*, 321(1-2), pp.305-339.
- Rickard, A.H., Leach, S.A., Buswell, C.M., High, N.J. and Handley, P.S., 2000. Coaggregation between aquatic bacteria is mediated by specific-growth-phase-dependent lectin-saccharide interactions. *Applied and environmental microbiology*, 66(1), pp.431-434.
- Riefler, M., Novak, O., Strnad, M. and Schmölling, T., 2006. Arabidopsis cytokinin receptor mutants reveal functions in shoot growth, leaf senescence, seed size, germination, root development, and cytokinin metabolism. *The Plant Cell*, 18(1), pp.40-54.
- Roberts, J.A., Elliott, K.A. and Gonzalez-Carranza, Z.H., 2002. Abscission, dehiscence, and other cell separation processes. *Annual review of plant biology*, 53(1), pp.131-158.
- Roberts, J.A., Whitelaw, C.A., Gonzalez-Carranza, Z.H. and McManus, M.T., 2000. Cell separation processes in plants—models, mechanisms and manipulation. *Annals of Botany*, 86(2), pp.223-235.
- Rodríguez, H. and Fraga, R., 1999. Phosphate solubilizing bacteria and their role in plant growth promotion. *Biotechnology advances*, 17(4), pp.319-339.
- Roger, Y.S., Adelberg, A.E. and Ingraham, L.J. 1980. General Microbiology. In: *Microbial Growth*. Fourth Edn., Prentice-Hall, Inc., Englewood Cliffs, New Jersey, USA. Pp 270.
- Rogers, P.L., Lee, K.J. and Tribe, D.E., 1979. Kinetics of alcohol production by *Zymomonas mobilis* at high sugar concentrations. *Biotechnology letters*, 1(4), pp.165-170.

- Rose, M.T., Phuong, T.L., Nhan, D.K., Cong, P.T., Hien, N.T. and Kennedy, I.R., 2014. Up to 52% N fertilizer replaced by biofertilizer in lowland rice via farmer participatory research. *Agronomy for sustainable development*, 34(4), pp.857-868.
- Ryu, C., Murphy, J.F., Reddy, M.S. and Kloepper, J.W., 2007. A two-strain mixture of rhizobacteria elicits induction of systemic resistance against *Pseudomonas syringae* and Cucumber mosaic virus coupled to promotion of plant growth on *Arabidopsis thaliana*. *Journal of Microbiology and Biotechnology*, 17(2), p.280.
- Saber, Z., Pirdashti, H., Esmaeili, M., Abbasian, A. and Heidarzadeh, A., 2012. Response of wheat growth parameters to co-inoculation of plant growth promoting rhizobacteria (PGPR) and different levels of inorganic nitrogen and phosphorus. *World Applied Sciences Journal*, 16, pp.213-9.
- Sadasivan, L.A.K.S.H.M.I. and Neyra, C.A., 1985. Flocculation in *Azospirillum brasilense* and *Azospirillum lipoferum*: exopolysaccharides and cyst formation. *Journal of Bacteriology*, 163(2), pp.716-723.
- Şahin, F., Çakmakçı, R. and Kantar, F., 2004. Sugar beet and barley yields in relation to inoculation with N<sub>2</sub>-fixing and phosphate solubilizing bacteria. *Plant and Soil*, 265(1), pp.123-129.
- Sahoo, R.K., Ansari, M.W., Dangar, T.K., Mohanty, S. and Tuteja, N., 2014. Phenotypic and molecular characterisation of efficient nitrogen-fixing *Azotobacter* strains from rice fields for crop improvement. *Protoplasma*, 251(3), pp.511-523.
- Sahoo, R.K., Ansari, M.W., Pradhan, M., Dangar, T.K., Mohanty, S. and Tuteja, N., 2014. Phenotypic and molecular characterization of native *Azospirillum* strains from rice fields to improve crop productivity. *Protoplasma*, 251(4), pp.943-953.
- Sajjad Mirza, M., Ahmad, W., Latif, F., Haurat, J., Bally, R., Normand, P. and Malik, K.A., 2001. Isolation, partial characterization, and the effect of plant growth-promoting bacteria (PGPB) on micro-propagated sugarcane in vitro. *Plant and Soil*, 237(1), pp.47-54.
- Santi, C., Bogusz, D. and Franche, C., 2013. Biological nitrogen fixation in non-legume plants. *Annals of botany*, 111(5), pp.743-767.
- Sarig, S., Blum, A. and Okon, Y., 1988. Improvement of the water status and yield of field-grown grain sorghum (*Sorghum bicolor*) by inoculation with *Azospirillum brasilense*. *The Journal of Agricultural Science*, 110(02), pp.271-277.



- Sasakawa, H. and Yamamoto, Y., 1978. Comparison of the uptake of nitrate and ammonium by rice seedlings influences of light, temperature, oxygen concentration, exogenous sucrose, and metabolic inhibitors. *Plant Physiology*, 62(4), pp.665-669.
- Sattar, M.A. and Gaur, A.C., 1987. Production of auxins and gibberellins by phosphate-dissolving microorganisms. *Zentralblatt für Mikrobiologie*, 142(5), pp.393-395.
- Sayed, R.Z., Reddy, M.S., Kumar, K.V., Yellareddygari, S.K.R., Deshmukh, A.M., Patel, P.R. and Gangurde, N.S., 2012. Potential of plant growth-promoting rhizobacteria for sustainable agriculture. In *Bacteria in agrobiolgy: Plant probiotics* (pp. 287-313). Springer Berlin Heidelberg.
- Schachtman, D.P. and Shin, R., 2007. Nutrient sensing and signaling: NPKS. *Annual Review of Plant Biology*. 58:47–69.
- Schilling, G., Gransee, A., Deuhel, A., Ležoviž, G. and Ruppel, S., 1998. Phosphorus availability, root exudates, and microbial activity in the rhizosphere. *Journal of Plant Nutrition and Soil Science*, 161(4), pp.465-478.
- Schlöter, M., Leubhn, M., Heulin, T. and Hartmann, A., 2000. Ecology and evolution of bacterial microdiversity. *FEMS Microbiology Reviews*, 24(5), pp.647-660.
- Seck, P.A., Diagne, A., Mohanty, S. and Wopereis, M.C., 2012. Crops that feed the world 7: Rice. *Food Security*, 4(1), pp.7-24.
- Seizo, M. 1980. Easy diagnosis of rice cultivation. p 30-31. In: Rice Cultivation for the Million. Japan scientific societies press.
- Sessitsch, A., Howieson, J.G., Perret, X., Antoun, H. and Martinez-Romero, E., 2002. Advances in *Rhizobium* research. *Critical Reviews in Plant Sciences*, 21(4), pp.323-378.
- Sexton, R. and Roberts, J.A., 1982. Cell biology of abscission. *Annual Review of Plant Physiology*, 33(1), pp.133-162.
- Shafi, M., Shah, A., Bakht, J., Shah, M. and Mohammad, W., 2012. Integrated effect of inorganic and organic nitrogen sources on soil fertility and productivity of maize. *Journal of Plant Nutrition*, 35(4), pp.524-537.
- Shaharoon, B., Naveed, M., Arshad, M. and Zahir, Z.A., 2008. Fertilizer-dependent efficiency of *Pseudomonads* for improving growth, yield, and nutrient use efficiency of wheat (*Triticum aestivum* L.). *Applied Microbiology and Biotechnology*, 79(1), pp.147-155.

- Sharma, A.K., 2002. *Biofertilizers for sustainable agriculture* (Vol. 12, pp. 319-324). India.: Agrobios.
- Sharma, J., Namdeo, K.N., Shrivastava, K.B.L., Patel, A.K. and Tiwari, O.P., 2006. Effect of fertility levels, growth regulators and biofertilizers on nutrient contents and uptake of field pea (*Pisum sativum* L.). *CROP RESEARCH-HISAR*-, 32(2), p.192.
- Sharma, P., Sardana, V. and Kandola, S.S., 2011. Response of groundnut (*Arachishypogaea* L.) to Rhizobium Inoculation. *Libyan Agriculture Research Centre Journal International*, 2, pp.101-104.
- Sharpley, A.N., Weld, J.L., Beegle, D.B., Kleinman, P.J., Gburek, W.J., Moore, P.A. and Mullins, G., 2003. Development of phosphorus indices for nutrient management planning strategies in the United States. *Journal of Soil and Water Conservation*, 58(3), pp.137-152.
- Sheehy, J.E. and Mitchell, P.L., 2013. Designing rice for the 21st century: the three laws of maximum yield. *Discuss Paper Series*, 48, p.19.
- Sheng, X.F., Zhao, F., He, L.Y., Qiu, G. and Chen, L., 2008. Isolation and characterization of silicate mineral-solubilizing *Bacillus globisporus* Q12 from the surfaces of weathered feldspar. *Canadian Journal of icrobiology*, 54(12), pp.1064-1068.
- Shinya, O.B.A., KIKUCHI, F. and MARUYAMA, K., 1990. Genetic analysis of semidwarfness and grain shattering of Chinese rice variety "Ai-Jio-Nan-Te". *Japanese Journal of Breeding*, 40(1), pp.13-20.
- Shinya, O.B.A., Noriko, S.U.M.I., Fujimoto, F. and Yasue, T., 1995. Association between grain shattering habit and formation of abscission layer controlled by grain shattering gene sh-2 in rice (*Oryza sativa* L.). *Japanese Journal of Crop Science*, 64(3), pp.607-615.
- Shoda, M., 2000. Bacterial control of plant diseases. *Journal of Bioscience and Bioengineering*, 89(6), pp.515-521.
- Singh, R., Parameswaran, T.N., Prakasa Rao, E.V.S., Puttanna, K., Kalra, A., Srinivas, K.V.N.S., Bagyaraj, D.J. and Divya, S., 2009. Effect of arbuscular mycorrhizal fungi and *Pseudomonas fluorescens* on root-rot and wilt, growth and yield of *Coleus forskohlii*. *Biocontrol Science and Technology*, 19(8), pp.835-841.
- Singh, S. and Kapoor, K.K., 1998. Effects of inoculation of phosphate-solubilizing microorganisms and an arbuscular mycorrhizal fungus on mungbean grown under natural soil conditions. *Mycorrhiza*, 7(5), pp.249-253.

- Sitepu, I.R., Hashidoko, Y., Santoso, E. and Tahara, S., 2007, August. Potent phosphate-solubilizing bacteria isolated from dipterocarps grown in peat swamp forest in Central Kalimantan and their possible utilization for biorehabilitation of degraded peatland. In *Proceedings of the international symposium and workshop on tropical Peatland, Yogyakarta* (pp. 27-29).
- Sivasakthi, S., Kanchana, D., Usharani, G. and Saranraj, P., 2013. Production of plant growth promoting substance by *Pseudomonas fluorescens* and *Bacillus subtilis* isolated from paddy rhizosphere soil of Cuddalore district, Tamil Nadu, India. *International Journal of Microbiological Research*, 4(3), pp.227-233.
- Somasegaran, P. and Hoben, H.J., 1994. Quantifying the growth of rhizobia. In *Handbook for Rhizobia* (pp. 47-57). Springer New York.
- Song, X., Liu, M., Wu, D., Griffiths, B.S., Jiao, J., Li, H. and Hu, F., 2015. Interaction matters: Synergy between vermicompost and PGPR agents improves soil quality, crop quality and crop yield in the field. *Applied Soil Ecology*, 89, pp.25-34.
- Spaepen, S. and Vanderleyden, J., 2011. Auxin and plant-microbe interactions. *Cold Spring Harbor Perspectives in Biology*, 3(4), pp.001438.
- Spaepen, S., Vanderleyden, J. and Remans, R., 2007. Indole-3-acetic acid in microbial and microorganism-plant signaling. *FEMS Microbiology Reviews*, 31(4), pp.425-448.
- Sturz, A.V. and Nowak, J., 2000. Endophytic communities of rhizobacteria and the strategies required to create yield enhancing associations with crops. *Applied Soil Ecology*, 15(2), pp.183-190.
- Subba Rao, N. S. 1999. The rhizosphere and the phyllosphere. In *Soil Microbiology. 4th edition. Science Publishers, Inc., Enfield, New Hampshire*, p. 85.
- Sudhakar, P., Chattopadhyay, G.N., Gangwar, S.K. and Ghosh, J.K., 2000. Effect of foliar application of *Azotobacter*, *Azospirillum* and *Beijerinckia* on leaf yield and quality of mulberry (*Morus alba*). *The Journal of Agricultural Science*, 134(02), pp.227-234.
- Szot, B., Ferrero, A. and Molenda, M., 1998. Binding force and mechanical strength of rice grain. *International agrophysics*, 12, pp.227-230.
- Tabuchi, M., Abiko, T. and Yamaya, T., 2007. Assimilation of ammonium ions and reutilization of nitrogen in rice (*Oryza sativa* L.). *Journal of experimental botany*, 58(9), pp.2319-2327.

- Tahmatsidou, V., O'Sullivan, J., Cassells, A.C., Voyiatzis, D. and Paroussi, G., 2006. Comparison of AMF and PGPR inoculants for the suppression of Verticillium wilt of strawberry (*Fragaria x ananassa* cv. Selva). *Applied Soil Ecology*, 32(3), pp.316-324.
- Tan K.Z., Radziah O., Halim M.S., Khairuddin, A.R., Habib S.H. and Shamsuddin Z.H., 2014. Isolation and characterization of rhizobia and plant growth-promoting rhizobacteria (PGPR) and their effects on growth of rice seedlings. *Americal Journal of Agricultural and Biological Sciences*, 9(3), pp.342-360.
- Tang, W.H., 1994. Yield-increasing bacteria (YIB) and biocontrol of sheath blight of rice. *Improving Plant Productivity with Rhizosphere Bacteria*. Eds. MH Ryder, PM Stephens and GD Bowen, pp.267-273.
- Tanimoto, E., 2005. Regulation of root growth by plant hormones—roles for auxin and gibberellin. *Critical reviews in plant sciences*, 24(4), pp.249-265.
- TATLIDİL, F.F., Kiral, T., GÜNDOĞMUŞ, E., FİDAN, H. and AKTÜRK, D., 2005. The Effect of Crop Losses during Pre-Harvest and Harvest Periods on Production Costs in Tomato Production in the Ayaş and Nallıhan Districts of Ankara Province. *Turkish Journal of Sgriculture and Forestry*, 29(6), pp.499-509.
- Taurian, T., Anzuay, M.S., Angelini, J.G., Tonelli, M.L., Ludueña, L., Pena, D., Ibáñez, F. and Fabra, A., 2010. Phosphate-solubilizing peanut associated bacteria: screening for plant growth-promoting activities. *Plant and Soil*, 329(1-2), pp.421-431.
- Taurian, T., Anzuay, M.S., Ludueña, L.M., Angelini, J.G., Muñoz, V., Valetti, L. and Fabra, A., 2013. Effects of single and co-inoculation with native phosphate solubilising strain *Pantoea* sp J49 and the symbiotic nitrogen fixing bacterium *Bradyrhizobium* sp SEMIA 6144 on peanut (*Arachis hypogaea* L.) growth. *Symbiosis*, 59(2), pp.77-85.
- Taylor, J.E. and Whitelaw, C.A., 2001. Signals in abscission. *New Phytologist*, 151(2), pp.323-340.
- Thurber, C.S., Hepler, P.K. and Caicedo, A.L., 2011. Timing is everything: early degradation of abscission layer is associated with increased seed shattering in US weedy rice. *BMC Plant Biology*, 11(1), p.14.
- Thurber, C.S., Reagon, M., Gross, B.L., Olsen, K.M., Jia, Y. and Caicedo, A.L., 2010. Molecular evolution of shattering loci in US weedy rice. *Molecular Ecology*, 19(16), pp.3271-3284.

- Tilman, D., 1998. The greening of the green revolution. *Nature*, 396(6708), pp.211-212.
- Tiwari, V.N., Lehri, L.K. and Pathak, A.N., 1989. Effect of inoculating crops with phospho-microbes. *Experimental Agriculture*, 25(01), pp.47-50.
- Tsay, Y.F., Ho, C.H., Chen, H.Y. and Lin, S.H., 2011. Integration of nitrogen and potassium signaling. *Annual Review of Plant Biology*, 62, pp.207-226.
- Turan, M., Gulluce, M., Cakmakci, R., Oztas, T. and Sahin, F., 2010. The effect of PGPR strain on wheat yield and quality parameters. In *Proceedings of the 19th World Congress of Soil Science: Soil solutions for a changing world, Brisbane, Australia, 1-6 August 2010. Symposium 2.3. 1 The soil-root interface* (pp. 140-143). International Union of Soil Sciences (IUSS), c/o Institut für Bodenforschung, Universität für Bodenkultur.
- Ueda, N., Kojima, M., Suzuki, K. and Sakakibara, H., 2012. *Agrobacterium tumefaciens* tumor morphology root plastid localization and preferential usage of hydroxylated prenyl donor is important for efficient gall formation. *Plant physiology*, 159(3), pp.1064-1072.
- Ullman, W.J., Kirchman, D.L., Welch, S.A. and Vandevivere, P., 1996. Laboratory evidence for microbially mediated silicate mineral dissolution in nature. *Chemical Geology*, 132(1-4), pp.11-17.
- United Nation, 2004. World Population Prospects 2300. Department of Economic and Social Welfares: Population Division. United Nation, New York.
- Vacheron, J., Desbrosses, G., Bouffaud, M.L., Touraine, B. and Prigent-Combaret, C., 2013. Plant growth-promoting rhizobacteria and root system functioning. *Frontiers in Plant Science*, 356 (4), pp 1-19.
- Valverde, A., Burgos, A., Fiscella, T., Rivas, R., Velazquez, E., Rodríguez-Barrueco, C., Cervantes, E., Chamber, M. and Igual, J.M., 2007. Differential effects of coinoculations with *Pseudomonas jessenii* PS06 (a phosphate-solubilizing bacterium) and *Mesorhizobium ciceri* C-2/2 strains on the growth and seed yield of chickpea under greenhouse and field conditions. In *First International Meeting on Microbial Phosphate Solubilization* (pp. 43-50). Springer Netherlands.
- Van Nguyen, N. and Ferrero, A., 2006. Meeting the challenges of global rice production.

- Vassilev, N., Vassileva, M., Fenice, M. and Federici, F., 2001. Immobilized cell technology applied in solubilization of insoluble inorganic (rock) phosphates and P plant acquisition. *Bioresource Technology*, 79(3), pp.263-271.
- Vejan, P., Abdullah, R., Khadiran, T., Ismail, S. and Nasrulhaq Boyce, A., 2016. Role of plant growth promoting rhizobacteria in agricultural sustainability—a review. *Molecules*, 21(5), p.573.
- Vessey, J.K., 2003. Plant growth promoting rhizobacteria as biofertilizers. *Plant and Soil*, 255(2), pp.571-586.
- Vitousek, P.M., Aber, J.D., Howarth, R.W., Likens, G.E., Matson, P.A., Schindler, D.W., Schlesinger, W.H. and Tilman, D.G., 1997. Human alteration of the global nitrogen cycle: sources and consequences. *Ecological Applications*, 7(3), pp.737-750.
- Wandersman, C. and Delepelaire, P., 2004. Bacterial iron sources: from siderophores to hemophores. *Annual Review of Microbiology*, 58, pp.611-647.
- Wang, Y. and Wu, W.H., 2010. Plant sensing and signaling in response to K<sup>+</sup>-deficiency. *Molecular Plant*, 3(2), pp.280-287.
- Wani, P., Khan, M. and Zaidi, A., 2007. Co-inoculation of nitrogen-fixing and phosphate-solubilizing bacteria to promote growth, yield and nutrient uptake in chickpea. *Acta Agronomica Hungarica*, 55(3), pp.315-323.
- Wasule, D.L., Wadyalkar, S.R. and Buldeo, A.N., 2007. Effect of phosphate solubilizing bacteria on role of *Rhizobium* on nodulation by soybean. In *First international meeting on microbial phosphate solubilization* (pp. 139-142). Springer Netherlands.
- Watanabe, K., Oba, S. and Horiuchi, T., 2003. Allelic test of rice shattering genes sh1 and sh2 in an F<sub>2</sub> population derived from the cross between Momigaredatsu and Dee-geo-woo-gen (*Oryza sativa* L.). *SABRAO JOURNAL OF BREEDING AND GENETICS*, 35, pp.57-64.
- Weerakoon, W.M., Olszyk, D.M. and Moss, D.N., 1999. Effects of nitrogen nutrition on responses of rice seedlings to carbon dioxide. *Agriculture, Ecosystems & Environment*, 72(1), pp.1-8.
- Weller, D.M. and Thomashow, L.S., 1994. Current challenges in introducing beneficial microorganisms into the rhizosphere. *Molecular ecology of rhizosphere microorganisms: Biotechnology and the release of GMOs*, pp.1-18.

- Whitelaw, M.A., 1999. Growth promotion of plants inoculated with phosphate-solubilizing fungi. *Advances in Agronomy*, 69, pp.99-151.
- Whitelaw, M.A., Harden, T.J. and Bender, G.L., 1997. Plant growth promotion of wheat inoculated with *Penicillium radicum* sp. nov. *Soil Research*, 35(2), pp.291-300.
- Wilson, J.W., Wilson, P.W. and Walker, E.S., 1987. Abscission sites in nodal explants of *Impatiens sultani*. *Annals of Botany*, 60(6), pp.693-704.
- Wilson, P.W., Wilson, J.W., Addicott, F.T. and McKenzie, R.H., 1986. Induced abscission sites in internodal explants of *impatiens sultani*: a new system for studying positional control with an appendix: A mathematical model for abscission sites. *Annals of Botany*, 57(4), pp.511-530.
- Wong, W.S., Tan, S.N., Ge, L., Chen, X. and Yong, J.W.H., 2015. The importance of phytohormones and microbes in biofertilizers. In *Bacterial metabolites in sustainable agroecosystem* (pp. 105-158). Springer International Publishing.
- Worldometers, 2018. [www.worldometers.info](http://www.worldometers.info). Accessed 7 April 2018.
- Worlds Food Summit, 1996. World Food Summit Newsletter. 13-17 June 1999. Dr. Jacques Diouf-Director General of FAO. Food Agricultural Organization Corporate Document Repository. Retrieved from: [[http:// www. fao. org/ wfs/ index\\_ en.htm](http://www.fao.org/wfs/index_en.htm)].
- Yanni, Y.G. and Dazzo, F.B., 2010. Enhancement of rice production using endophytic strains of *Rhizobium leguminosarum* bv. *trifolii* in extensive field inoculation trials within the Egypt Nile delta. *Plant and Soil*, 336(1-2), pp.129-142.
- Yanni, Y.G., Rizk, R.Y., Corich, V., Squartini, A., Ninke, K., Philip-Hollingsworth, S., Orgambide, G., De Bruijn, F., Stoltzfus, J., Buckley, D. and Schmidt, T.M., 1997. Natural endophytic association between *Rhizobium leguminosarum* bv. *trifolii* and rice roots and assessment of its potential to promote rice growth. *Plant and Soil*, 194(1-2), pp.99-114.
- Yoneyama, T., Muraoka, T., Kim, T.H., Dacanay, E.V. and Nakanishi, Y., 1997. The natural <sup>15</sup>N abundance of sugarcane and neighbouring plants in Brazil, the Philippines and Miyako (Japan). *Plant and Soil*, 189(2), pp.239-244.
- Yoshida, S., 1981. *Fundamentals of rice crop science*. International Rice Research Institute.

- Young, C.C., 1990. Effects of phosphorus-solubilizing bacteria and vesicular-arbuscular mycorrhizal fungi on the growth of tree species in subtropical-tropical soils. *Soil Science and Plant Nutrition*, 36(2), pp.225-231.
- Young, C.C., Juang, T.C. and Chao, C.C., 1988. Effects of *Rhizobium* and vesicular-arbuscular mycorrhiza inoculations on nodulation, symbiotic nitrogen fixation and soybean yield in subtropical-tropical fields. *Biology and Fertility of Soils*, 6(2), pp.165-169.
- Young, C.C., Juang, T.C. and Guo, H.Y., 1986. The effect of inoculation with vesicular-arbuscular mycorrhizal fungi on soybean yield and mineral phosphorus utilization in subtropical-tropical soils. *Plant and Soil*, 95(2), pp.245-253.
- Yu, X., Liu, X., Zhu, T.H., Liu, G.H. and Mao, C., 2012. Co-inoculation with phosphate-solubilizing and nitrogen-fixing bacteria on solubilization of rock phosphate and their effect on growth promotion and nutrient uptake by walnut. *European Journal of Soil Biology*, 50, pp.112-117.
- Zakry, F.A.A., Shamsuddin, Z.H., Rahim, K.A., Zakaria, Z.Z. and Rahim, A.A., 2012. Inoculation of *Bacillus sphaericus* UPMB-10 to young oil palm and measurement of its uptake of fixed nitrogen using the <sup>15</sup>N isotope dilution technique. *Microbes and Environments*, 27(3), pp.257-262.
- Zee, S.Y., 1979. Abscission layer in rice pedicel. *IRRN*, 4, pp.5-6.
- Zhang, S., Reddy, M.S., Kokalis-Burelle, N., Wells, L.W., Nightengale, S.P. and Kloepper, J.W., 2001. Lack of induced systemic resistance in peanut to late leaf spot disease by plant growth-promoting rhizobacteria and chemical elicitors. *Plant Disease*, 85(8), pp.879-884.
- Zhong, X., Peng, S., Sanico, A.L. and Liu, H., 2003. Quantifying the interactive effect of leaf nitrogen and leaf area on tillering of rice. *Journal of Plant Nutrition*, 26(6), pp.1203-1222.
- Zuo, Y. and Zhang, F., 2011. Soil and crop management strategies to prevent iron deficiency in crops. *Plant and Soil*, 339(1-2), pp.83-95.



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## PUBLICATION

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### Abstract

Mohamad Halid Razak, Halimi Mohd Saud and Mohd Razi Ismail. 2016. Effect of PGPR inoculation on NPK content of rice (*Oryza sativa* L.) variety MR263 growth under glasshouse condition. *Soil science conference of Malaysia, 2015*. Pp. 287-288

Mohamad Halid Razak, Zulkarami Berahim, Mohd Razi Ismail and Halimi Mohd Saud. 2016. *Proceedings 7<sup>th</sup> international agriculture congress 2016*. Pp. 523-526



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