



**UNIVERSITI PUTRA MALAYSIA**

***EFFECTS OF DIFFERENT ROUTES OF VACCINATION AGAINST  
Streptococcus agalactiae IN RED HYBRID TILAPIA FINGERLINGS  
(Oreochromis sp.)***

**AISYAH BT AMINUDDIN**

**FPV 2016 62**

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*Streptococcus agalactiae* IN RED HYBRID TILAPIA FINGERLINGS  
(*Oreochromis* sp.)**

**‘AISYAH BT AMINUDDIN**

**A project paper submitted to the  
Faculty of Veterinary Medicine, Universiti Putra Malaysia**

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## CERTIFICATION

It is hereby certified that we have read this project paper entitled “Effects of different routes of vaccination against *Streptococcus agalactiae* in Red hybrid tilapia fingerlings(*Oreochromis*sp.)”, by ‘AisyahBintiAminuddin and in my opinion, it is satisfactory in terms of scope, quality, and presentation as partial fulfilment of the requirement for the course of VPD 4999 – Project.

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ASSOC. PROF. DR. MD SABRI MOHD YUSOFF,

DVM, MVSc, PhD (UPM)

Associate Professor,

Department of Veterinary Pathology and Microbiology

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Supervisor)

## DEDICATIONS

*To my dear and lovely parents, Mr Aminuddin Ahmad*

*And*

*Mrs.Ruhani Mohamed*

*For making me who I am today*

*To my dearest siblings,*

*Ahmad SyahiqAminuddin*

*'AfifahAminuddin*

*Ahmad SyamimAminuddin*

*Ahmad Syazwan Aziz Aminuddin*

*For being there whenever I needed you*

*“You don't choose your family*

*They are God's gift to you, as you are to them.”*

*-Desmond Tutu*

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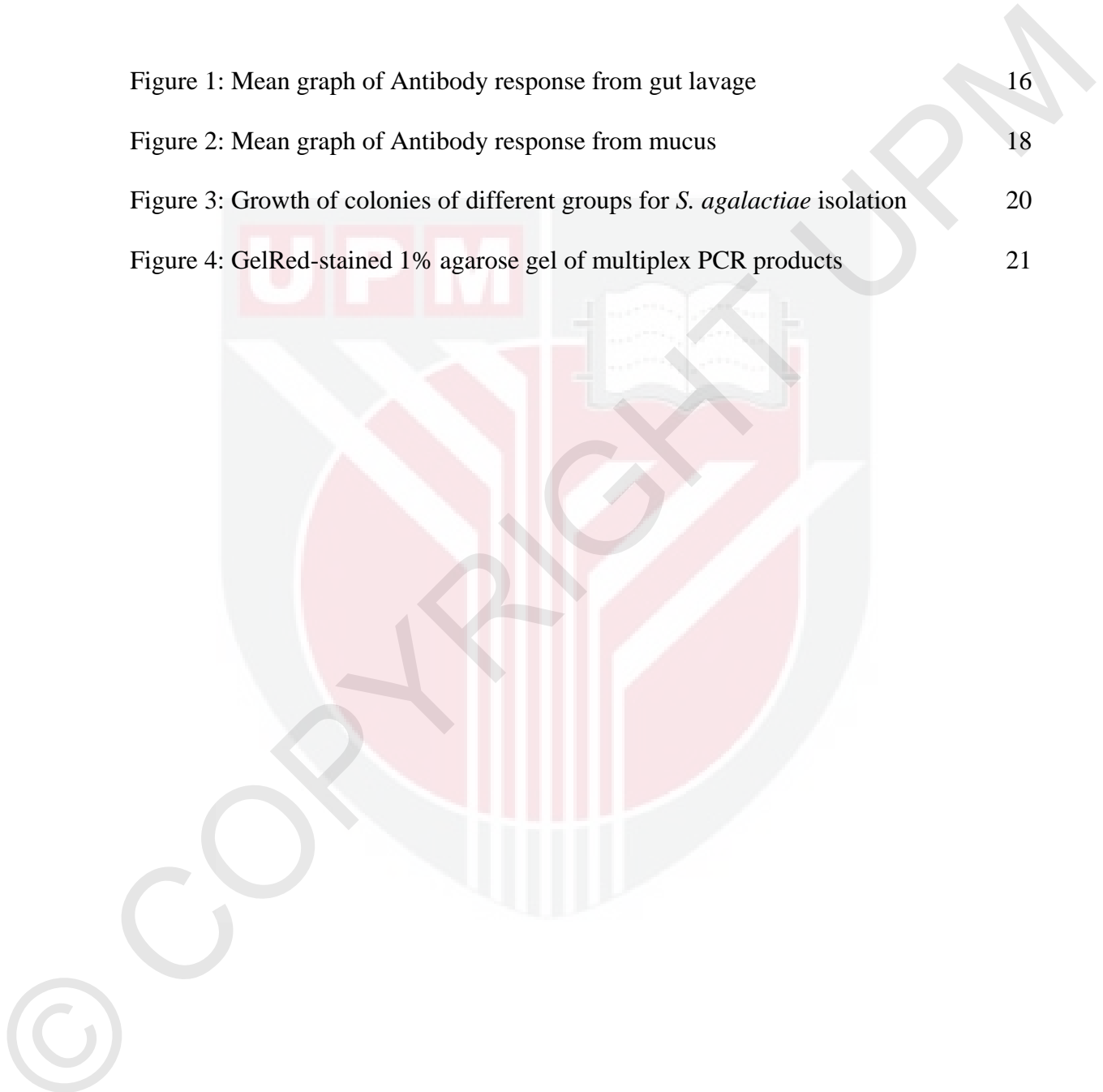
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**LIST OF ABBREVIATION***S. agalactiae**Streptococcus agalactiae*

FKB

Formalin-Killed Bacteria

FCV

Formalin Crude Vaccine

PBS

Phosphate Buffered Saline

BHI

Brain Heart Infusion

ELISA

Enzyme-linked immunosorbent assay

PCR

Polymerase Chain Reaction

CFU/mL

Colony forming unit per mililiter

°C

Degree celcius

rpm

Revolutions per minute

PBST

Phosphate Buffered Saline + Tween-20

BSA

Bovine serum albumin

IgM

Immunoglobulin M

DNA

Deoxyribonucleic acid

UV

Ultraviolet

ANOVA

Analysis of variance

mL

Mililiter

mm

Milimeter

μL

Microliter

&lt;

Less than

Bp

Base pair

## ABSTRAK

Abstrak daripada kertas projek yang dikemukakan kepada Fakulti Perubatan Veterinar untuk memenuhi sebahagian daripada keperluan kursus VPD 4999- Projek

### **KESAN LALUAN VAKSINASI YANG BERBEZA TERHADAP *Streptococcus agalactiae* PADA ANAK TILAPIA HIBRID MERAH (*Oreochromis sp.*)**

Oleh

**'Aisyah Binti Aminuddin**

**2016**

**Penyelia: Prof. Madya Dr. Md Sabri Mohd Yusoff**

Streptokokosis merupakan penyakit yang disebabkan oleh jangkitan *Streptococcus sp.* Penyakit ini merupakan masalah global bagi sector pengeluaran ikan sedunia dan saling berkaitan dengan kerugian yang tinggi dalam sector ekonomi. Kajian ini dilakukan bertujuan untuk mengenalpasti kesan terhadap penghasilan antibody bagi laluan vaksinasi yang berbeza terhadap *Streptococcus agalactiae* dalam anak ikan tilapia merah. Sembilan puluh anak ikan tilapia merah dibahagikan secara rawak kepada tiga kumpulan iaitu kumpulan 1, 2 dan 3 yakni setiap kumpulan mempunyai tiga puluh ekor anak ikan. Dua formulasi bagi formalin vaksin tidak aktif telah dihasilkan iaitu vaksin berasaskan makanan dan semburan. Kumpulan 1 menggunakan vaksin secara semburan selama 3 hari berturut-turut pada minggu pertama dan pada minggu ketiga sebagai dos ransangan jugas elama tiga hari berturut-turut. Bagi kumpulan 2, anak ikan divaksin secara semburan hanya sekali pada minggu pertama dan

diberi vaksin berasaskan makanan pada minggu ketiga sebagai dos penggalak manakala kumpulan 3 kekal sebagai kelompok kawalan tanpa vaksinasi. Kesemua kumpulan dijangkitkan dengan *S. agalactiae*, 109 CFU/mL pada kadar 100 ml melalui intraperitoneum. Setelah dijangkiti, anak ikan berada di bawah pemerhatian bagi sebarang petanda klinikal atau pun kadar kematian. Sampel mucus bagi lima ekor anak ikan daripada setiap kumpulan telah diambil dengan menggunakan steril calitan pada permukaan kulit ikan setiap ming. Cecair lavaj usus juga telah diambil dan kedua-dua sampel diproses menggunakan asaiimunoserap terangkai ensim (ELISA) tak langsung bagi mengenalpasti penghasilan antibody IgM terhadap *S. agalactiae*. Keputusan ELISA menunjukkan penghasilan antibody bagi sampel mucus dan cecair lava jusus bagi anak ikan tilapia yang telah divaksin tidak mempunyai perbezaan bererti walaupun berlainan kaedah vaksinasi telah digunakan.

Kata kunci: *Streptococcus agalactiae*, Tilapia hybrid merah, vaksin semburan, vaksin berasaskan makanan, asaiimunoserap terangkai ensim (ELISA)

**ABSTRACT**

Abstract of the project paper presented to the Faculty of Veterinary Medicines in partial for the course VPD 4999- Final year project.

**EFFECTS OF DIFFERENT ROUTES OF VACCINATION AGAINST  
*Streptococcus agalactiae* IN RED HYBRID TILAPIA FINGERLINGS  
(*Oreochromis sp.*)**

**By**

**'AisyahBintiAminuddin**

**2016**

**Supervisor: Assoc. Prof Dr.SabriMohdYusoff**

Streptococcosis is a disease that develops following infection by *Streptococcus sp.* It is a major problem for fish production worldwide, and it is associated with high economic losses. This study was aimed at investigating the effects of different vaccination route against *Streptococcus agalactiae* in Red hybrid tilapia fingerlings. Ninety fingerlings were randomly divided into three groups, 1, 2 and 3 of 30 each. Two

formalin-killed vaccine formulations were developed, feed based and spray. Group 1 was vaccinated using spray vaccine for 3 consecutive days in the 1<sup>st</sup> week and a booster dose for 3 consecutive days in the 3<sup>rd</sup> week. Group 2 was vaccinated once by using spray vaccine followed by a booster dose using the feed-based vaccine. While Group 3 served as a control group without any vaccination. All groups were challenged with 100 µL of *S. agalactiae* (10<sup>9</sup> CFU/mL) intraperitoneally. Following challenge, the fingerlings were observed for any clinical signs and mortality. Mucus samples of five fish from each group at sampling time were collected by using sterile swab at the surface of the skin. Gut lavage fluid was also collected, and both samples were subjected to indirect enzyme-linked immunosorbent assay (ELISA) to determine the IgM antibody levels against *S. agalactiae*. The results showed that the IgM antibody response in mucus and gut lavage fluids produced by the tilapia immunized with vaccination were not significantly different with each other even though different routes of vaccination were used.

**Keywords:** *Streptococcus agalactiae*, Red hybrid tilapia, spray vaccine, feed-based vaccine, enzyme-linked immunosorbent assay (ELISA)



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## 1.0 INTRODUCTION

### 1.1 Study Background

Streptococcosis can be acute or chronic. Acute streptococcosis usually occurs in the seasons where the water temperature is high resulting in peaks of mortality that last 2 - 3 weeks (Intervet, 2006). The chronic take place when the water temperature is lower and does not cause peaks of mortality. Amal and Zamri (2011) reported that streptococcosis occur by the infection of *Streptococcus* sp. The shape is spherical or ovoid and 0.5 - 2.0  $\mu\text{m}$  in diameter, occur in pairs or chains when grown in liquid media, non-motile, non-spore-forming and appears purple or blue when stained using Gram stain.

*Streptococcus* bacteria are part of the normal flora on animal bodies, but infection and disease also can occur when the bacteria enter through the cuts, abrasions, wounds or when the immune system becomes weakened especially involving the stress. *Streptococcus* spp., which can cause diseases in fish, includes *S. agalactiae* (Suanyuket *et al.*, 2005) *Streptococcus iniae* (Shoemaker *et al.*, 2000) and *Streptococcus difficile* (Berridge *et al.*, 2001). Evans *et al.* (2002) said that Group B *S. agalactiae* is another emerging fish pathogen of freshwater and saltwater fish species throughout the world.

Environmental stress is playing a major role in influencing the fish immunity for examples such as water temperature, high nitrate level and low dissolved oxygen. When the line of immunity in fish is declined, the fish are highly susceptible to streptococcosis. Siti-Zahra *et al.* (2004) and Amalet *et al.* (2008) reported that high



mortality of the fish infection is frequently recorded between April and July that depicts dry season in Malaysia. When fish are infected with streptococcosis, the most common clinical signs shown include anorexia, exophthalmia, ascites and erratic swimming (Evans *et al.*, 2002; Salvador *et al.*, 2005) and meningoencephalitis in fish (Eldar *et al.*, 1995). Mian *et al.* (2009) also said that streptococcosis caused by *S. agalactiae* is a major disease of many fish species, and it is characterized by septicaemia and meningoencephalitis.

Pathogenesis in infected fish involves septicaemia and colonization of several organs such as nares, brain, kidney and intestines (Pasnik *et al.*, 2005). However, other than fish animals such as mice, cats, dogs, hamsters, camels and frogs can also be infected with streptococcosis (Evans *et al.*, 2002).

The vaccine is an antigenic material that stimulates the immune system by developing the adaptive immunity to a pathogen. The main aim of vaccination against an infectious disease is to stimulate host adaptive immune responses to counteract the infection caused by a pathogen. Vaccination is the most effective method to combat disease (Karen and Scott, 2011) and it also an important disease prevention to maintain human and animal health worldwide. Craig *et al.* (2009) reported that vaccines developed for aquaculture have significantly reduced antibiotic use in fish production. Lombart *et al.* (2007) also stated that vaccination is the most effective and cost-effective method of preventing infectious diseases.

## 1.2 Justification

The study in determining the antibody response between spray vaccine and a feed-based vaccine against *S. agalactiae* was never done before. Thus, this study will reveal the information regarding the antibody responses production between these two different methods and at the same time giving the choices of vaccination methods that can be more practical to be practiced in future. Besides, *S. agalactiae* is one of emerging fish pathogen reported in tilapia fish throughout the world including Malaysia.

## 1.3 Objectives

- i. To determine the antibody response in the mucus and gut-lavage in Red hybrid tilapia fingerlings.
- ii. To isolate *S. agalactiae* post challenge in Red hybrid tilapia fingerlings.

## 1.4 Hypothesis

**H<sub>0</sub>:** Presence of antibody responses in mucus and gut-lavage in different route in Red hybrid tilapia fingerlings

**H<sub>A</sub>:** Absence of antibody responses in mucus and gut-lavage in different route in Red hybrid tilapia fingerlings

## REFERENCES

- Abuseliana, A., Daud, H., Aziz, S. A., Bejo, S. K., & Alsaid, M. (2010). *Streptococcus agalactiae* the etiological agent of mass mortality in farmed red tilapia (*Oreochromis* sp.). *J Anim Vet Adv*, 9(20), 2640-2646.
- Alliance, R. U. M. A. (2014). *Responsible Use of Medicines in Agriculture Alliance Guidelines*. 2005.
- Amal A. M. N., SitiZahrah A., Zulkafli R., Misri S., Ramley A. and Zamri-Saad M. (2008). The effect of water temperature on the incidence of *Streptococcus agalactiae* infection in cage cultured tilapia. International seminar on management strategies on animal health and production control in anticipation of global warming, Surabaya, Indonesia.48-51.
- Amal M. N. A., and Zamri-Saad M., (2011) Streptococcosis in Tilapia (*Oreochromis niloticus*): A Review. *Pertanika Journal Tropical Agriculture Science*. 34(2), 195 – 206
- Amal, M. N. A., Zamri-Saad, M., Siti-Zahrah, A., & Zulkafli, A. R. (2014). Control and prevention of streptococcosis in cultured tilapia in Malaysia: a review.
- Amal, M. N., Kutty, B. M., & Abdullah, S. Z. (2008). Streptococcosis in red hybrid tilapia (*Oreochromis niloticus*) commercial farms in Malaysia. *Aquaculture Research*, 40, 630-632.
- Bullock, G.L. (1981). Streptococcal infections of fishes. *Fish Disease Leaflet* (pp. 1-7). United States Department of the Interior, Fish and Wildlife Service.
- Chapman, F. A. (2000). *Culture of hybrid tilapia: A reference profile*. University of Florida Cooperative Extension Service, Institute of Food and Agriculture Sciences, EDIS.
- David J., Pasnik A., Joyce J., Evans A., Phillip H., Klesius B. (2006) Passive immunization of Nile tilapia (*Oreochromis niloticus*) provides significant protection against *Streptococcus agalactiae*. *Fish & Shellfish Immunology*. 21, 365-371
- Diseases of Tilapia - An Introduction*. (2016). *The Fish Site*. Retrieved 8 January 2016, from <http://www.thefishsite.com/articles/139/diseases-of-tilapia-an-introduction/>
- Eldar A., Bejerano Y., Livoff A., Horovitz A., Bercovier H.. (1995). Experimental streptococcal meningo-encephalitis in cultured fish. *Veterinary Microbiology*. 43, 33-40

- Evans J. J., Klesius P.H., Gilbert P.M., Shoemaker C.A., Al Sarawi M.A., Landsberg J., Duremdez R., Al Marzouk A. and Al Zenk S. (2002) Characterization of beta-haemolytic Group B *Streptococcus agalactiae* in cultured sea bream, *Sparus auratus* L and wild mullet, *Liza klunzingeri* in Kuwait. *Journal of Fish Diseases*. 25, 505–513
- Evans, J.J., Pasnik, D.J., Klesius, P.H., & Shoemaker, C.A. (2006a). Identification and epidemiology of *Streptococcus iniae* and *Streptococcus agalactiae* in tilapia, *Oreochromis* spp. *International Symposium on Tilapia in Aquaculture 7* (pp. 25-42). Charles Town, WV, USA, American Tilapia Association
- Gomes, S., & Afonso, A. (2006). Fish vaccination against infections by Streptococcal species and the particular case of Lactococcosis. *Revista Portuguesa de Ciências Veterinárias*, 557, 25.
- Hashim, R., Chong, A. S., Fatan, N. A., Layman, N., & Ali, A. (2002). Production of hybrid red tilapia, *Oreochromis mossambicus* × *O. niloticus*, at varying stocking densities in portable canvas tanks. *Journal of Applied Aquaculture*, 12(3), 1-12.
- Holt, J.G., Krieg, N.R., Sheath, P.A., Staley, J.T., & Williams, S.T. (1994). *Bergey's manual of determination bacteriology* (pp. 532-538). Philadelphia: Williams & Wilkins.
- Intervet aquatic animal health (2006). Streptococcosis in Tilapia (accessed in April 2013).
- J Woods, C., & S Levy, C. (2016). *Streptococcus Group B Infections: Practice Essentials, Background, Pathophysiology*. *Emedicine.medscape.com*. Retrieved 8 January 2016, from <http://emedicine.medscape.com/article/229091-overview#a4>
- Karen P. P., and Scott E. L. (2011). Advances in fish vaccine delivery. *Developmental and Comparative Immunology*. 35, 1256–1262
- Komar, C., Enright, W. J., Grisez, L., & Tan, Z. (2004). Understanding fish vaccination. *Aqua Culture Asia Pacific Magazine*, 27-9.
- Lombard, Pastoret P. P., Moulin A. M., (2007). “A brief history of vaccines and vaccination”. *Reviews.-Off. International Epizootiology*. 26(1):29-48
- Mian G. F., Godoy D. T., Leal C. A. G., Yuhara T. Y., Costa G. M., Figuerido H. C. (2009) Aspects of the natural history and virulence of *S. agalactiae* infection in Nile tilapia. *Veterinary Microbiology*. 136, 180-183.

- Midtløng P. J., Reitan U., Speilberg L. (1996) Efficacy and side effects of injectable furunculosis vaccines in Atlantic salmon (*Salmo salar* L.). *Fish Shellfish Immunology*. 6, 335-350
- Mohamed, L. A., & Soliman, W. S. E. (2013). Development and efficacy of fish vaccine used against some bacterial diseases in farmed Tilapia. *Natural Science*, 11, 120-128.
- Noraini, O., Sabri, M. Y., & Siti-Zahrah, A. (2013). Efficacy of spray administration of formalin-killed *Streptococcus agalactiae* in hybrid Red Tilapia. *Journal of aquatic animal health*, 25(2), 142-148.
- Nur-Nazifah, M., Sabri, M. Y., & Siti-Zahrah, A. (2014). Development and efficacy of feed-based recombinant vaccine encoding the cell wall surface anchor family protein of *Streptococcus agalactiae* against streptococcosis in *Oreochromis* sp. *Fish & shellfish immunology*, 37(1), 193-200.
- Pasnik D. J., Evans J. J., Klesius P. H., (2005). Duration of protective antibodies and correlation with survival in Nile tilapia (*Oreochromis niloticus*) following *Streptococcus agalactiae* vaccination. *Diseases of Aquatic Organisms*. 66, 129-134.
- Salvador R., Muller E. E., Freitas J. C., Leonhardt J. H., Giordano L. G. P., & Dias J. A. (2005). Isolation and characterization of *Streptococcus* spp. group B in Nile Tilapia (*Oreochromis niloticus*) reared in hapas nets and earth nurseries in the northern region of Parana State, Brazil. *Ciencia Rural*. Santa Maria. 35, 1374-1378.
- Shoemaker C. A., Evans J. J. and Klesius P. H. (2000). Density and dose: factors affecting mortality to *Streptococcus iniae* infected tilapia (*Oreochromis niloticus*). *Aquaculture*. 188, 229-235
- Suanyuk N., Kanghear H., Khongpradit R. & Supamattaya K. (2005). *Streptococcus agalactiae* infection in tilapia (*Oreochromis niloticus*). *Songklanakarin Journal Science Technology Aquaculture Science*. 27, 307-319.
- Toranzo, A.E.; Romalde, J.L.; Magariños, B. and Barja, J.L. (2009): The use of veterinary drugs and vaccines in Mediterranean aquaculture. *Options Méditerranéennes*, A / no. 86, 105-176
- Yanong, R. P., & Francis-Floyd, R. (2002). *Streptococcal Infections of Fish* 1.