



UNIVERSITI PUTRA MALAYSIA

**LETHAL DOSE, CLINICAL SIGNS, PATHOLOGICAL CHANGES AND
DISEASE DEVELOPMENT OF *Streptococcus agalactiae* FOLLOWING
INTRAPERITONEAL EXPOSURE TO JAVANESE MEDAKA
(*Oryzias javanicus*, Bleeker 1854)**

SITI SUHAIBA BINTI MASTOR

FS 2018 105



**LETHAL DOSE, CLINICAL SIGNS, PATHOLOGICAL CHANGES AND
DISEASE DEVELOPMENT OF *Streptococcus agalactiae* FOLLOWING
INTRAPERITONEAL EXPOSURE TO JAVANESE MEDAKA
(*Oryzias javanicus*, Bleeker 1854)**

By

SITI SUHAIBA BINTI MASTOR

**This Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree
of Master of Science**

March 2018



© COPYRIGHT UPM

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

LETHAL DOSE, CLINICAL SIGNS, PATHOLOGICAL CHANGES AND DISEASE DEVELOPMENT OF *Streptococcus agalactiae* FOLLOWING INTRAPERITONEAL EXPOSURE TO JAVANESE MEDAKA (*Oryzias javanicus*, Bleeker 1854)

By

SITI SUHAIBA MASTOR

March 2018

Chairman: Mohammad Noor Amal Azmai, PhD

Faculty: Science

This study was conducted to determine the median lethal dose, clinical signs, pathological changes and disease development of *Streptococcus agalactiae* in Javanese medaka (*Oryzias javanicus*, Bleeker 1854) model, following intraperitoneal exposure. Javanese medaka was collected from estuary area of Sungai Pelek, Sepang, Selangor, and brought to the laboratory for quarantine and acclimatization. The fish were then challenged from 10^2 - 10^8 CFU/ml of virulent *S. agalactiae* via intraperitoneal injection. Mortalities and clinical signs were observed until 240 h post infection (hpi), while the dead fish were collected for bacterial isolation and histological analyses. Median lethal dose 50% (LD₅₀) of *S. agalactiae* in Javanese medaka was determined at 5.3×10^2 CFU/ml. Most of the infected fish showing lethargy, erratic swimming pattern, exophthalmia and necrosis at the injection site. The histopathological changes were mainly generalised congestion of the internal organs. *Streptococcus agalactiae* were successfully isolated from the dead fish. In the disease development studies, the number of Javanese medaka mortalities following infection by 10^3 CFU/ml of *S. agalactiae* was directly proportional with concentration of *S. agalactiae* in fish and severity of histopathological findings through 96 hpi. Clinical signs and histopathological assessment also showed that infected fish displayed similar findings compared to several the natural host of *S. agalactiae*. This study concluded that Javanese medaka was

susceptible towards *S. agalactiae* infection and could be a potential alternative test organism for study of streptococcosis in fish.



Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia
sebagai memenuhi keperluan ijazah Master Sains

**DOS KEMATIAN, TANDA KLINIKAL, PERUBAHAN PATOLOGI DAN
PERKEMBANGAN PENYAKIT OLEH *STREPTOCOCCUS AGALACTIAE*
KEATAS MODEL MEDAKA JAWA (*ORYZIAS JAVANICUS*, BLEEKER
1854), MELALUI PENDEDAHAN SECARA SUNTIKAN ANTARA
PERITONEAL**

Oleh

SITI SUHAIBA BINTI MASTOR

Marc 2018

**Pengerusi: Mohammad Noor Amal bin Azmai, PhD
Fakulti: Sains**

Kajian ini telah dilakukan untuk mengkaji dos median kematian, tanda-tanda klinikal, perubahan patologi serta perkembangan penyakit oleh *Streptococcus agalactiae* ke atas medaka Jawa (*Oryzias javanicus*, Bleeker 1854). Medaka Jawa diambil dari Sungai Pelek, Sepang dan dibawa pulang ke makmal untuk penyesuaian kepada suasana makmal. Ikan-ikan tersebut didedahkan kepada beberapa siri kepekatan bakteria *S. agalactiae* daripada 10^2 hingga 10^8 CFU/ml melalui suntikan di ruangan antara peritoneal medaka Jawa. Kematian serta tanda-tanda klinikal direkod sehingga 240 jam selepas suntikan, manakala ikan yang mati diambil untuk pemencilan bakteria dan analisis histologi. Dos kematian yang dikenalpasti ialah 5.3×10^2 CFU/ml. Kebanyakan ikan yang dijangkiti menunjukkan tanda-tanda klinikal seperti keletihan, berenang dengan tidak menentu, eksoftalmia dan nekrosis di kawasan suntikan. Perubahan histopatologi pula menunjukkan lebih kepada kesesakan sel darah merah dalam organ dalaman ikan. *Streptococcus agalactiae* berjaya dibuktikan kehadirannya daripada ikan yang telah mati. Kajian perkembangan penyakit menunjukkan bahawa jumlah kematian selari dengan kepekatan bakteria *S. agalactiae* CFU/g sepanjang 96 hpi. Keputusan kajian menunjukkan bahawa medaka Jawamudah dijangkiti oleh *S. agalactiae* dan berpotensi untuk menjadi organisma alternatif didalam kajian streptococcosis bagi ikan.

ACKNOWLEDGEMENT

My heartiest gratitude goes to my beloved supervisor, Dr. Mohammad Noor Amal Azmai for his patience and support in guiding me throughout my postgraduate study. Working under his guidance has taught me to be a brand new person. I gained a lot of knowledge, skills and experience during my journey and able to make me an independent student in my research project. I want to take this opportunity to also thank my supervisory committee, Prof. Dr. Mohd Zamri Saad and Prof. Dr. Ahmad Ismail for their professional advice and comments.

On the other hand, a big applause goes to the staff of Department of Biology, Faculty of Science, UPM, for all the facilities and helps making me able to complete my research project.

After all, I would like to thank my late father Mastor Mohd Seh, my beloved mother Rahmah Zaini, brothers and sisters for all the prayers, love and support throughout my journey. Last but not least, I would like to thank my friends especially, Fatin Zahidah Abdul Aziz, Munirah Hanapiah, all of Casa Impian's members, and Nurliyana Mohamad, to always be there whenever I need a helping hand in finishing my study.

I certify that a Thesis Examination Committee has met on 12 March 2018 to conduct the final examination of Siti Suhaiba Mastor on her thesis entitled "Lethal dose, clinical signs, pathological changes and disease development of *Streptococcus agalactiae* following intraperitoneal exposure to Javanese medaka (*Oryzias javanicus*, Bleeker 1854)' " in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Syaizwan Zahmir Zulkifli, PhD,

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Ina Salwany binti Md Yassin, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Lee Seong Wei, PhD

Associate Professor
Universiti Malaysia Kelantan
Malaysia
(External Examiner)

NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 June 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of Supervisory Committee were as follows:

Mohammad Noor Amal Azmai, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Mohd Zamri Saad, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Ahmad Ismail, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 June 2018

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism software.

Signature: _____ Date: _____

Name and Matric No: _SITI SUHAIBA BINTI MASTOR (GS44545)

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature : _____
Name of Chairman of
Supervisory Committee : _____

Signature : _____
Name of Member of
Supervisory Committee : _____

Signature : _____
Name of Member of
Supervisory Committee : _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENT	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1	
INTRODUCTION	1
1.1 Introduction	1
1.2 Problem statements	3
1.3 Objectives	3
1.4 Hypothesis	3
2	
LITERATURE REVIEW	5
2.1 Model organism	5
2.1.1 List of model organisms in disease and infectious studies	7
2.1.2 Fish as model organism	9
2.2 Javanese medaka (<i>Oryzias Javanicus</i> , Bleeker 1854)	14
2.2.1 Taxonomy	14
2.2.2 Morphology	15
2.2.3 Biology	16
2.2.4 Reproduction	19
2.2.5 Medaka Life Cycle	20
2.2.6 Ecology	20
2.2.7 Distribution	21
2.3 Streptococcosis	22
2.3.1 <i>Streptococcus agalactiae</i>	23
2.3.2 List of fish infected by <i>Streptococcus</i> sp.	24
2.3.3 Symptoms and clinical signs of streptococcosis in fish	25
2.3.4 Pathological changes of streptococcosis in fish	27
2.3.5 Diagnosis	28
2.3.6 Transmission	30
2.3.7 Control and prevention of streptococcosis	30

3	MATERIALS AND METHODS	32
3.1	Collection of Javanese medaka	32
3.1.1	Sampling location	33
3.1.2	Fish acclimatization	33
3.1.3	Fish quarantine and diseases screening	33
3.2	<i>Streptococcus agalactiae</i>	33
3.2.1	Identification and characterization of <i>S. agalactiae</i>	33
3.2.2	Revirulent of bacteria	34
3.3	Median lethal dose	34
3.3.1	Bacteria culture and preparation	34
3.3.2	Ten-fold serial dilution	34
3.3.3	Intraperitoneal injection	35
3.3.4	Experimental monitoring	35
3.3.5	Lethal dose (LD ₅₀) determination	36
3.3.6	Histopathological assessment	36
3.3.7	Histopathological scoring	36
3.4	Disease development of Javanese medaka	37
3.4.1	Javanese medaka	37
3.4.2	Bacterial concentration	37
3.4.3	Intraperitoneal injection	37
3.4.4	Disease, symptoms, clinical signs and mortality observation	37
3.4.5	Colony Forming Unit/g determination	38
3.4.6	Histological assessment	38
3.4.7	Data analysis	38
4	RESULTS	39
4.1	Clinical signs, symptoms, median lethal dose and histopathological assessment	39
4.1.1	Identification of <i>Streptococcus agalactiae</i>	39
4.2	Disease symptoms and clinical signs of Javanese medaka following <i>S. agalactiae</i> infection	41
4.3	Mortality pattern of Javanese medaka	48
4.4	Median lethal dose of <i>S. agalactiae</i> in Javanese medaka	51
4.5	Histopathological assessments	52
4.5.1	Brain section	53
4.5.2	Kidney section	55
4.5.3	Liver section	55
4.6	Disease development	57
4.6.1	Colony Forming Unit/g of <i>S. agalactiae</i>	57

		57
4.6.2	Mortality pattern of Javanese medaka	57
4.6.3	Relationship between the CFU/g of <i>S. agalactiae</i> and mortality of Javanese medaka	58
4.6.4	Histopathological assessment on disease development	60
5	DISCUSSION	65
5.1	Clinical signs, symptoms, median lethal dose and histopathological assessment	65
5.1.1	Isolation <i>S. agalactiae</i> from infected fish	65
5.1.2	Symptoms and clinical signs	65
5.1.3	Median lethal dose	66
5.1.4	Histopathological assessment	67
5.2	Disease development	68
5.3	Javanese medaka as model organism	69
6	CONCLUSION & RECOMMENDATION	70
6.1	Conclusion	70
6.2	Recommendation	71
	REFERENCES	72
	BIODATA OF STUDENT	89

LIST OF TABLES

Table		Page
2.1	List of model organisms in disease and infectious studies	8
2.2	Number of publications in the Web of Science as of March 2013, regarding on fish model species and several research fields. The searching query was performed by typing '[species name]' in the field topic for the total number of entries or '[species name] AND [research field]' in the field topic for specific research areas.	10
2.3	Number of publications in the Web of Science as of March 2013, regarding on some of the most cultured species worldwide (FAO, 2010) and several research fields. The searching query was performed by typing '[species name]' in the field topic for the total number of entries or '[species name] AND [research field]' in the field 'topic' for specific research areas.	12
2.4	Taxonomy of Javanese medaka	15
2.5	Comparison of several meristic characters of field Javanese medaka (adapted from Yusof et al. (2013))	16
2.6	Localities of Javanese medaka in Peninsular Malaysia	17
2.7	List of fish species infected by streptococcal-bacterial related.	24
4.1	Biochemical test results of <i>S. agalactiae</i> isolated from dead red hybrid tilapia and Javanese medaka.	39
4.2	Symptoms and clinical signs of Javanese medaka infected by <i>S. agalactiae</i> at different bacterial concentration	41
4.3	Mortality pattern of Javanese medaka based on different <i>S. agalactiae</i> concentration up to 240 hpi	49
4.4	Different bacteria treatment with dilution factor and the	51

	number of dead Javanese medaka	
4.5	Histopathological scoring of Javanese medaka challenged with different concentration of <i>S. agalactiae</i> up to 240 hpi.	52
4.6	The CFU/g of <i>S. agalactiae</i> in fish and mortality of Javanese medaka at different hpi.	57
4.7	Histological scoring of selected organs and lesions throughout 96 hpi of <i>S. agalactiae</i> in Javanese medaka	60



LIST OF FIGURES

Figures		Page
2.1	Illustration of <i>Oryzias javanicus</i> (Bleeker, 1854). Above, adult male, and below, adult female. Bar indicates 5mm. (Magtoon & Termvidchakorn, 2009)	15
2.2	Javanese medaka a) female and b) male in laboratory culture (Yusof et al., 2013)	17
2.3	Tilapia naturally infected by <i>S. agalactiae</i> showing (a) exophthalmia and (b) corneal opacity (Zamri-Saad et al., 2010)	26
2.4	Tilapia infected by <i>S. agalactiae</i> showing (a) marked congestion of the viscera and (b) softening of the brain (Zamri-Saad et al., 2010)	26
2.5	Histopathology lesions seen in wild giant Queensland grouper infected with <i>S. agalactiae</i> . (a) Brain showing meningitis. Note thickened meninges with lymphocytic infiltration (arrow) (H&E, -100). (b) Gills with granulomatous inflammation (arrowheads) between adjacent gill filaments (H&E, -100). (c) Eye with prominent granuloma-like lesions (arrows) in the choroid space. Lesions consist of aggregations of macrophages containing coccoid bacteria identified as <i>S. agalactiae</i> (H&E, -40). (d) Heart ventricle with granulomatous inflammation (arrows) of the ventricular myocardium and myositis (H&E, -200) (Bowater et al., 2012)	28
3.1	The red circle indicates the location of Javanese medaka sampling area (Google Maps, 2017)	32
4.1	Positive result of <i>S. agalactiae</i> isolated from the dead fish and cultured on agar with 5% of horse blood. The culture shown β -hemolysis on the blood agar	40
4.2	Javanese medaka showing lethargy and isolated from schooling group in all challenged groups	43
4.3	Infected fish showed erratic swimming (circle) after challenged with 1.8×10^8 CFU/ml of <i>S. agalactiae</i>	44

4.4	The infected fish challenged by 1.8×10^8 CFU/ml of <i>S. agalactiae</i> displaying tissue necrosis at the injection site (circle). This signs was observed in the entire challenged fish group	45
4.5	Exophthalmia (red circle) was observed on the right eye of infected Javanese medaka. However, this clinical signs was only observed in fish infected with 1.8×10^7 CFU/ml of <i>S. agalactiae</i>	45
4.6	(a) control fish brain (arrow) showing the brain was still in contact and had recognizable shape while, (b) infected fish brain (arrow) showing the brain was soften and started to loses its particular brain shape	46
4.7	Javanese medaka shown C-curve body shape clinical signs (arrow) at 1.8×10^7 CFU/ml of <i>S. agalactiae</i> infection	46
4.8	Normal Javanese medaka without any symptoms or clinical signs in negative control group. The fish showed transparent body and easily to recognized any physical changes, if any	47
4.9	(a) Normal brain section of Javanese medaka (Bar = $100\mu\text{m}$) the meninges layer and overall tissue not showing any form of abnormalities or lesions related to streptococcosis (H&E staining). (b) Mild congestion of RBC in the meningeal layer after exposed to 8.4×10^3 CFU/ml of <i>S. agalactiae</i> (Bar = $100\mu\text{m}$)(c) Brain section of infected Javanese medaka by 1.8×10^8 CFU/ml of <i>S. agalactiae</i> . Arrow showing moderate congestion of RBC in the brain meningeal layer (Bar = $100\mu\text{m}$). M= brain meningeal layer. (H&E staining).	54
4.10	(a) Normal kidney section of Javanese medaka (Bar = $100\mu\text{m}$) (H&E staining). (b) Tubular necrosis (arrow) where the nucleus showing an obvious dense indicating karyopyknosis (N) following infection with 1.8×10^5 CFU/ml of <i>S. agalactiae</i> (c) Kidney section of Javanese medaka infected with 10^8 CFU/ml of <i>S. agalactiae</i> showing mild tubular necrosis (N) and mild glomerulus shrinkage (G) (Bar = $100\mu\text{m}$) (H&E staining).	55
4.11	(a) Normal liver section of Javanese medaka (Bar =	56

100µm)(b) Mild RBC congestion was observed at hepatic portal vein (HPV) (arrow) in Javanese medaka infected with 8.4×10^3 CFU/ml of *S. agalactiae* (Bar = 100µm)(c) Liver section of Javanese medaka infected with 1.8×10^8 CFU/ml of *S. agalactiae* showing moderate RBC congestion in the central vein (CV) and necrosis in the hepatocytes (N), and also abundant of visible Kupfer's cells surrounded the hepatocytes (vertical arrow) (Bar = 50µm) (H&E staining).

- 4.12 The relationship between Javanese medaka mortality and CFU/g of *S. agalactiae* until 96 hpi 58
- 4.13 (a) The normal section of negative control of Javanese medaka (Bar = 100µm) (b) The brain section showing mild RBC congestion at the meningeal layer (arrow) at 6 hpi (Bar = 100µm) (c) Moderate RBC congestion at the meningeal layer (arrow) (Bar = 100µm) at 12 hpi (H&E staining). 62
- 4.14 (a) Normal liver section of negative control Javanese medaka (Bar = 100µm) (H&E staining). (b) Liver section of Javanese medaka with moderate focal necrosis (circle) at 36 hpi (Bar = 100µm) (H&E staining). (c) Liver section of Javanese medaka with karyopyknosis event (arrow) at 48 hpi (Bar = 100µm) (H&E staining). 63
- 4.15 (a) The normal kidney section of negative control of Javanese medaka (Bar = 100µm) (b) Kidney section with moderate glomerulus shrinkage (circle) and tubular necrosis (arrow) at 6 hpi (Bar = 100µm) (c) Kidney section with mild glomerulus shrinkage (circle) and intermediate tubular necrosis (arrow) at 36 hpi (Bar = 100µm) (H&E staining). 64

LIST OF ABBREVIATIONS

°C	Degree Celcius
%	Percentage
LD ₅₀	Median lethal dose
mm	Millimeter
mgL ⁻¹	Miligram per liter
ppt	Parts per thousand
°	Degree
N	North
E	East
µm	Micrometer
m	Meter
cm	Centimeter
USD	US Dollar
PCR	Polymerase chain reaction
IHC	Immunohistochemistry
β	Beta
γ	Gamma
GBS	Group B <i>Streptococcus</i>
H&E	Hematoxylin and eosin
hr	Hour
BHIA	brain heart infusion agar
TSA	tryptone soy agar
THBA	Todd-Hewitt broth agar
mg/kg/day	Milligram per kilogram per day
µS/cm	Micro-Siemens per centimeter
NH ₄ ⁺	Ammonia
L	Liter
NO ₂ ⁻	Nitrite
NO ₃ ⁻	Nitrite
CaCO ₃	Calcium carbonate
g	Gram
cc	Cubic centimeter
G	Gauge
CFU/ml	Colony forming unit per milliliter
rpm	Revolutions per minute
MS222	Triacane-metasulphonate
µL	microliter
EtOH	Ethanol
IP	Intraperitoneal
CFU/g	Colony forming unit per gram

CHAPTER 1

INTRODUCTION

1.1 Introduction

Many studies have been conducted in establishing fish as test organism in various research disciplines. Zebrafish, *Danio rerio* (Hamilton, 1822), for example, is excellently playing its role as the model organism in various studies relating to disease, ecotoxicology, neurology, genetic and etc., since it display many interesting criteria as a test organism (Ribas & Piferrer, 2014). Around 320 million of years ago, this particular species was evolving in South Asia with its geographical distribution predominantly in Bangladesh, Nepal and India (Spence et al., 2008). Recently, zebrafish has been introduced around the globe for the purpose of research (Bier & McGinnis, 2008).

However, the introduction of a non-native species in an ecosystem is always likely to cause an ecological problems if the species is able to integrate itself successfully into the ecosystem, resulting in possible detrimental interactions with native species or even on ecosystem functioning (Gozlan et al., 2010). The non-native species which introduced to the environment are also able to give impacts in terms of competition (Gurevitch et al., 1992; Fausch, 1998; Potapov & Lewis, 2004; Simon et al., 2004; Caiola & Sostoa, 2005; McDowall, 2006;), predation (McDowall, 2006; Bampfylde & Lewis, 2007; Yonekura et al., 2007), transmission of a novel disease (Gozlan et al., 2006), hybridization and habitat modification (McDowall, 2006).

Recently, researchers were looking for new native fish species to be established as test organism, in order to reduce the ecological and environmental impacts. Javanese medaka, *Oryzias javanicus* (Bleeker, 1854), is a small fish which massively studied and amenable in many areas of research. It holds many other model organism's characteristics such as short life cycles, and can be found in a diverse groups distributed around Asia (Magtoon & Termvidchakorn, 2009). They live in brackish water, freshwater and also the saltwater. In the same genus with Javanese medaka, Japanese medaka, *Oryzias latipes* (Temminck & Schlegel, 1846) is one of the most established fish model species, where they are widely used in experimental of vertebral biology for many years (Ismail & Yusof, 2011). Based on Koyama et al., (2006), Javanese medaka is commonly found in estuarine waters of southern to eastern Asia.

Many advantageous features are found in medaka as an experimental animal. Throughout the embryonic development, the embryos were totally transparent yet particularly pigment-less mutant, ever since it is an oviparous fish, where the embryonic stage occurs externally (Wakamatsu et al., 2001). Quintet and STIII are the examples of strains which transparent even in the adult stage. Moreover, medaka is able to adapt with low temperature which is its natural tolerance. It is a temperate-zone fish where it is able to survive without thermostatic regulator. The environmental temperature able to control the embryonic development rate, for instance, arresting the embryonic development at 10°C and recommences at 25°C (Kinoshita et al., 2009).

Nevertheless, *Streptococcus agalactiae* is a kind of wide range infectious bacteria infecting humans, terrestrial and aquatic animals (Wongsathein, 2012). Various of disease outbreaks had been reported from this bacteria species, thus, has been responsible as causative agent for neonatal meningitis, pneumonia, sepsis, soft tissue and osteomyelitis infections in humans (Brochet et al., 2006; Johri et al., 2006). Bolaños et al., (2005) mentioned that this particular bacteria species latent to infect pregnant women and elderly people as well as immune-compromised adults which lead to mortality, particularly those with malignancies, diabetes mellitus, liver cirrhosis and a history of previous surgery.

On the other hand, ruminants are also susceptible to the infection by *S. agalactiae*. This pathogen prone to inhabit the mammary glands occasioning in clinical and sub-clinical mastitis in cattle which can utterly affect milk quality and production (Phuektes et al., 2001). Besides, it has also been isolated from several other animals presenting with a disease including frogs, mice, dogs, cats, hamsters, chickens, guinea pigs, horses, emerald monitors, monkeys, camels, bottlenose dolphins and captive saltwater crocodiles (Zappuli et al., 2005; Evans et al., 2006; Bishop et al., 2007).

Fish farmers also faced a huge economic loss due to streptococcosis. In acute *Streptococcus* infection, the fish mortality could be observed between 1 to 7 days, and was able to reach up to 50% of mortality. Recently, *Streptococcus iniae* had just identified in farmed tilapia (*Oreochromis aureus*) in Mexico (Ortega et al., 2017) meanwhile, *S. difficile* (Berridge et al., 2001) were reported to cause diseases in rainbow trout. In Malaysia, Amal et al.,(2013) stated that red tilapia (*Oreochromis* sp.) were mainly suffered by *S. agalactiae* infection as first reported in late 1990's. Lately, reported outbreaks and infections of *S. agalactiae* in tilapia in Malaysia have been widespread (Amal et al., 2013).

1.2 Problem statements

Unintentionally introduction of alien fish species for utilization as test organisms could potentially annihilate local species, thus, threatening the stability of the existing water ecosystem. Moreover, Javanese medaka has never been evaluated as a model organism in bacterial fish diseases study especially for streptococcosis.

Thus, this study was assessed the availability of Javanese medaka as the native model organism, as another alternative for fish model instead of using non-native species for research purposes in this country.

1.3 Objectives

This study was conducted in order to evaluate the capability of Javanese medaka as an alternative test organism for streptococcosis study in fish.

Thus, the objectives of this study are:

- i. to determine the median lethal dose (LD_{50}) of *Streptococcus agalactiae* following intraperitoneal exposure to Javanese medaka *Oryzias javanicus*;
- ii. to determine the clinical signs and pathological changes in Javanese medaka following intraperitoneal exposure to *S. agalactiae*;
- iii. to assess the disease development of *S. agalactiae* following intraperitoneal exposure to Javanese medaka.

1.4 Hypothesis

It is hypothesized that Javanese medaka is suitable as native test organism for study of streptococcosis in fish.

Thus, the null hypotheses (H_0) for this study are:

- i. median lethal dose (LD_{50}) of *S. agalactiae* following intraperitoneal exposure to Javanese medaka cannot be determined;
- ii. clinical signs and pathological changes do not shown by Javanese medaka following intraperitoneal exposure by *S.*

- agalactiae* and are not comparable to the previous studies using different fish species;
- iii. there is no development of streptococcosis observed in Javanese medaka following intraperitoneal exposure by *S. agalactiae*

Thus, the alternative hypotheses (H_a) for this study are:

- i. median lethal dose (LD_{50}) of *S. agalactiae* following intraperitoneal exposure to Javanese medaka can be determined;
- ii. clinical signs and pathological changes shown by Javanese medaka following intraperitoneal exposure by *S. agalactiae* are comparable to the previous studies using different fish species;
- iii. there is development of streptococcosis observed in Javanese medaka following intraperitoneal exposure by *S. agalactiae*

REFERENCES

- Aamri, F. El, Padilla, D., Acosta, F., Caballero, M. J., Roo, J., Bravo, J., ... Real, F. (2010). First report of *Streptococcus iniae* in red porgy (*Pagrus pagrus*, L.). *Journal of Fish Diseases*, 33(11), 901–905. <http://doi.org/10.1111/j.1365-2761.2010.01191.x>
- Abdullah, S., Omar, N., Yusoff, S. M., Obukwho, E. B., Nwunuji, T. P., Hanan, L., & Samad, J. (2013). Clinicopathological features and immunohistochemical detection of antigens in acute experimental *Streptococcus agalactiae* infection in red tilapia (*Oreochromis* spp.). *SpringerPlus*, 2(1), 286. <http://doi.org/10.1186/2193-1801-2-286>
- Abner, S. R., Frazier, C. L., Scheibe, J. S., Krol, R. A., Overstreet, R. M., Walker, W. W., & Hawkins, W. E. (1994). Chronic inflammatory lesions in two small fish species, Medaka (*Oryzias latipes*) and Guppy (*Poecilia reticulata*), used in carcinogenesis bioassays. In *Modulators of Fish Immune Responses: Model for Environmental Toxicology, Biomarkers, and Immunostimulators* (pp. 219–233). USA: SOS Publications.
- Abuseliana, A., Duad, H., Aziz, S. A., Bejo, S. K., & Alsaïd, M. (2010). *Streptococcus agalactiae* the etiological agent of mass mortality on farmed red tilapia (*Oreochromis* sp.). *Journal of Animal and Veterinary Advances*, 9(20), 2640–2646.
- Abuseliana, A. F., Daud, H. M., Abdul Aziz, S., Khairani Bejo, S., & AlSaïd, M. (2011). Pathogenicity of *Streptococcus agalactiae* isolated from a fish in Selangor to Juvenile Red tilapia (*Oreochromis* sp.). *JAVA*, 10(7), 914–919.
- Agnes, W. & Barnes, A. C. (2007). *Streptococcus iniae*: an aquatic pathogen of global veterinary significance and a challenging candidate for reliable vaccination. *Veterinary Microbiology*, 122, 1–15.
- Akhir, M. F., P.C., S., & M.L., H. (2011). Seasonal variation of South China Sea physical characteristics off the east coast of Peninular Malaysia from 2002 – 2010 datasets. *International Journal of Environmental Sciences*, 2(2), 569–575.
- Almendras, F. E., Fuentealba, I. C., Markham, R. F. F., & Speare, D. J. (2000). Pathogenesis of liver lesions caused by experimental infection with *Piscirickettsia salmonis* in juvenile Atlantic salmon, *Salmo salar* L. *Journal of Veterinary Diagnostic Investigation*, 12(6), 552. <http://doi.org/http://dx.doi.org/10.1177/104063870001200610>
- Alsaïd Milud, Daud Hassan Hj Mohd, Mohamed Mustapha Noordin, B. S. K., Mohamed Abdelhadi Yasser, Abuseliana Ali Farag, and H. R. H., Milud Alsaïd, Hassan Hj Mohd Daud, Noordin Mohamed Mustapha, Siti Khairani Bejo, Y., & Mohamed Abdelhadi, Ali Farag Abuseliana, and R. H. H. (2013). Pathological Findings of Experimental *Streptococcus Agalactiae*

Infection in Red Hybrid Tilapia (*Oreochromis* sp.). *International Conference on Chemical, Agricultural and Medical Sciences (CAMS-2013)*, 70–73. <http://doi.org/10.15242/IICBE.C1213075>

- Amal, M. N. A., Saad, M. Z., Zahrah, A. S., & Zulkafli, A. R. (2015). Water quality influences the presence of *Streptococcus agalactiae* in cage cultured red hybrid tilapia, *Oreochromis niloticus* × *Oreochromis mossambicus*. *Aquaculture Research*, 46(2), 313-323.
- Amal, A. M. N., Siti-Zahrah, A., Zulkafli, R., Misri, S., Ramley, A., & Zamri-Saad, M. (2008). The effect of water temperature on the incidence of *Streptococcus agalactiae* infection in cage-cultured tilapia. In *International Seminar on Management Strategies on Animal Health and Production Production Control in Anticipation of Global Warming* (pp. 48–51). Surabaya.
- Amal, M. N. a, Zamri-Saad, M., Iftikhar, a. R., Siti-Zahrah, a., Aziel, S., & Fahmi, S. (2012). An outbreak of *Streptococcus agalactiae* infection in cage-cultured golden pompano, *Trachinotus blochii* (Lac??p??de), in Malaysia. *Journal of Fish Diseases*, 35(11), 849–852. <http://doi.org/10.1111/j.1365-2761.2012.01443.x>
- Ankeny, R. A. (2001). The natural history of *C. elegans* research. *National Review Genetics*, 3, 474–478.
- Ankeny, R. A., & Leonelli, S. (2011). What's so special about model organisms? *Studies in History and Philosophy of Science Part A*, 42(2), 313–323.
- Austin, B., & Austin, D. A. (2007). *Bacterial Fish Pathogens. Diseases of Farmed and Wild Fish* (4th ed.). Chichester: Springer/Prazis Publishing.
- Aziz, F. Z. A., Zulkifli, S. Z., Mohamat-Yusuff, F., Azmai, M. N. A., & Ismail, A. (2017). A Histological Study on Mercury-Induced Gonadal Impairment in Javanese Medaka (*Oryzias javanicus*). *Turkish Journal of Fisheries and Aquatic Sciences*, 17(3), 621-627.
- Bampfyld, C. J., & Lewis, M. A. (2007). Biological control through intraguild predation: case studies in pest control, invasive species and range expansion. *Bulletin of Mathematical Biology*, 69(3), 1031-1066., 69(3), 1031–1066.
- Barato, P., Martins, E. R., Melo-Cristino, J., Iregui, C. A., & Ramirez, M. (2015). Persistence of a single clone of *Streptococcus agalactiae* causing disease in tilapia (*Oreochromis* sp.) cultured in Colombia over 8 years. *Journal of fish diseases*, 38(12), 1083-1087.
- Berridge, B. R., Bercovier, H., & Frelief, P. F. (2001). *Streptococcus agalactiae* and *Streptococcus difficile* 16S–23S intergenic rDNA: genetic homogeneity and species-specific PCR. *Veterinary Microbiology*, 78(2),

165–173.

- Bethesda, M. D. (2010). Model organism for biomedical research. Retrieved April 26, 2010, from <http://www.nih.gov/science/models/>
- Bier, E., & McGinnis, W. (2008). Model organisms in the study of development and disease. *Molecular Basis of Inborn Errors of Development*, 25–48. Retrieved from <http://books.google.com/books?hl=en&lr=&id=wGoj9RtTcVIC&oi=fnd&pg=PA25&dq=Model+Organisms+in+the+Study+of+Development+and+Disease&ots=YDD9Pwoz3k&sig=NJpFt15Zrs4K-BOMSZyoXEeLBGE>
- Bilzer, M., Roggel, F., & Gerbes, A. L. (2006). Role of Kupffer cells in host defense and liver disease. *Liver International*, 26(10), 1175–1186.
- Bishop, E. J., Shilton, C., Benedict, S., Kong, F., Gilbert, G. L., Gal, D., ... Currie, B. J. (2007). Necrotizing fasciitis in captive juvenile *Crocodylus porosus* caused by *Streptococcus agalactiae*: an outbreak and review of the animal and human literature. *Epidemiology and Infection*, 135(8), 1248–1255.
- Bleeker, P. (1854). Faunae ichthyologicae japonicae species novae. *Natuurkundig Tijdschrift Voor Nederlandsch Indië*, 395–426.
- Bolaños, M., Hernández, A., Santana, O. E., Molina, J., & Martín-Sánchez, A. M. (2005). Distribution of *Streptococcus agalactiae* serotypes in samples from non-pregnant adults. *Clinical Microbiology Newsletter*, 27(19), 151–153. *Clinical Microbiology Newsletter*, 27(19), 151–153.
- Bowater, R. O., Forbes-Faulkner, J., Anderson, I. G., Condon, K., Robinson, B., Kong, F., ... Blyde, D. (2012). Natural outbreak of *Streptococcus agalactiae* (GBS) infection in wild giant Queensland grouper, *Epinephelus lanceolatus* (Bloch), and other wild fish in northern Queensland, Australia. *Journal of Fish Diseases*, 35(3), 173–186. <http://doi.org/10.1111/j.1365-2761.2011.01332.x>
- Brochet, M., Couvé, E., Zouine, M., Vallaëys, T., Rusniok, C., Lamy, M. C., & Glaser, P. (2006). Genomic diversity and evolution within the species *Streptococcus agalactiae*. *Microbes and Infection*, 8(5), 1227–1243.
- Bromage, E. S., & Owens, L. (2002). Infection of barramundi *Lates calcarifer* with *Streptococcus iniae*: Effects of different routes of exposure. *Diseases of Aquatic Organisms*, 52(3), 199–205. <http://doi.org/10.3354/dao052199>
- Broussard, G. W., & Ennis, D. G. (2007). *Mycobacterium marinum* produces long-term chronic infections in medaka: a new animal model for studying human tuberculosis. *Comparative Biochemistry and Physiology. Toxicology & Pharmacology: CBP*, 145(1), 45–54. <http://doi.org/10.1016/j.cbpc.2006.07.012>
- Buer, J., & Balling., R. (2003). Mice, microbes and models of infection. *National Review Genetics*, 4, 195–205.

- Buller, N. B. (2004). *Bacteria from Fish and Other Aquatic Animals: A Practical Identification Manual*. Wallingford: CABI Publishing.
- Burian, R. M. (1993). How the choice of experimental organism matters: Epistemological reflections on an aspect of biological practice. *Journal of the History of Biology*, 26(2), 351–367.
- Caiola, N., & Sostoa, A. D. (2005). Possible reasons for the decline of two native toothcarps in the Iberian Peninsula: evidence of competition with the introduced Eastern mosquitofish. *Journal of Applied Ichthyology*, 21(4), 358–363.
- (CCOHS), C. C. for occupational H. and S. (2013). What is a LD50 and LC50? Retrieved November 15, 2017, from <https://www.ccohs.ca/oshanswers/chemicals/ld50.html>
- Chaffin, D. O., Beres, S. B., Yim, H. H., & Rubens, C. E. (2000). The serotype of type Ia and III group B streptococci is determined by the polymerase gene within the polycistronic capsule operon. *Journal of Bacteriology*, 182, 4466–4477.
- Chang, J. P., Johnson, J. D., Sawisky, G. R., Grey, C. L., Mitchell, G., & Booth, M. (2009). Signal transduction in multifunctional neuroendocrine control of gonadotropin secretion and synthesis in teleost-studies on the goldfish model. *General and Comparative Endocrinology*, 161, 42–52.
- Chen, C. Y., Chao, C. B., & Bowser, P. R. (2007). Comparative histopathology of *Streptococcus iniae* and *Streptococcus agalactiae*-infected tilapia. *Bulletin of the European Association of Fish Pathologists*, 27(1), 2–9.
- Clarke, A. E., & Fujimura, J. H. (2014). *The right tools for the job: At work in twentieth-century life sciences*. Princeton University Press.
- Coromines, M., & Valls, M. (2011). (2011) *Organismes Model en Biologia. Societat Catalana de Biologia, Barcelona*. Barcelona: Societat Catalana de Biologia.
- Cresko, W. A., McGuigan, K. L., Philips, P. C., & Postelthwait, J. H. (2007). Studies of threespine stickleback developmental evolution: progress and promise. *Genetica*, 41, 177–184.
- Crnogorac-Jurcevic, T., Brown, J. R., Lehrach, H., & Schalkwyk, L. C. (1997). *Tetraodon fluviatilis*, a new puffer fish model for genome studies. *Genomics*, 41(2), 177–184.
- Dahm, R., & Geisler, R. (2006). Learning from small fry: the zebrafish as a genetic model organism for aquaculture fish species. *Marine Biotechnology*, 8, 329–345.
- D'Amato, M. E., Esterhuysen, M. M., Van Der Waal, B. C., Brink, D., & Volckaert, F. A. (2007). Hybridization and phylogeography of the Mozambique tilapia *Oreochromis mossambicus* in southern Africa

- evidenced by mitochondrial and microsatellite DNA genotyping. *Conservation Genetics*, 8(2), 475-488.
- Darwish, A. M., & Griffin, B. R. (2002). Study shows oxytetracycline controls *Streptococcus* in tilapia. *Global Aquaculture Advocate*, 5, 34-35.
- Darwish, A. M., & Hobbs, M. S. (2005). Laboratory efficacy of amoxicillin for the control of *S. iniae* infection in blue tilapia. *Journal of Aquatic Animal Health*, 17, 197-202.
- Devlin, R. H., & Nagahama, Y. (2002). Sex determination and sex differentiation in fish: an overview of genetic, physiological, and environmental influences. *Aquaculture*, 208(3-4), 191-364.
- Devriese, L. A. (1991). Streptococcal ecovars associated with different animal species: epidemiological significance of serogroups and biotypes. *Journal of Applied Bacteriology*, 71, 478-483.
- Dionne, M. S., Ghorri, N., & Schneider, D. S. (2003). *Drosophila melanogaster* is a genetically tractable model host for *Mycobacterium marinum*. *Infection and Immunity*, 71(6), 3540-3550.
- Duremdez, R., Al-Marzouk, A., Qasem, J. A., Al-Harbi, A., & Gharaball, H. (2004). Isolation of *Streptococcus agalactiae* from cultured silver pomfret, *Pampus argenteus* (Euphrasen). *Journal of Fish Diseases*, 27, 307-310.
- Dushay, M. S., & Eldon, E. D. (1998). *Drosophila* immune responses as models for human immunity. *The American Journal of Human Genetics*, 62(1), 10-14.
- Eldar, A., Bejerano, Y., & Bercovier, H. (1994). *Streptococcus shiloi* and *Streptococcus difficile*, two new streptococcal species causing a meningoecephalitis in fish. *Current Microbiology*, 28, 139-143.
- Eldar, A., Bejerano, Y., Livoff, A., Horovitz, A., & Bercovier, H. (1995). Experimental streptococcal meningo-encephalitis in cultured fish. *Veterinary Microbiology*, 43, 33-40.
- Eldar, A., & Ghittino, C. (1999). *Lactococcus garvieae* and *Streptococcus iniae* infections in rainbow trout *Oncorhynchus mykiss*: Similar, but different diseases. *Diseases of Aquatic Organisms*, 36(3), 227-231. <http://doi.org/10.3354/dao036227>
- Eldar, A., Horovitz, A., & Bercovier, H. (1997). Development of a vaccine against *S. iniae* infection in farmed rainbow trout. *Veterinary Immunology and Immunopathology*, 56, 175-183.
- Endersby, J. A. (2007). *A guinea pig's history of biology*. London: Random Huse.
- Esterhuysen, M. E. D. M. M., Waal, B. C. W. van der, Brink, D., & Volckaert, F. A. M. (2007). Hybridization and phylogeography of the Mozambique

tilapia *Oreochromis mossambicus* in southern Africa evidenced by mitochondrial and microsatellite DNA genotyping. *Conservation Genetics*, 8, 475–488.

- Evans, J. J., Bohnsack, J. F., Klesius, P. H., Whiting, A. A., Garcia, J. C., Shoemaker, C. A., & Takahashi, S. (2008). Phylogenetic relationships among *Streptococcus agalactiae* isolated from piscine, dolphin, bovine and human sources: a dolphin and piscine lineage associated with a fish epidemic in Kuwait is also associated with human neonatal infections in Japan. *Journal of Medical Microbiology*, 57, 1369–1376.
- Evans, J. J., Klesius, P. H., Gilbert, P. M., Shoemaker, C. A., Al Sarawi, M. A., Landsberg, J., ... Al Zenki, S. (2002). Characterization of B-haemolytic group B *Streptococcus agalactiae* in cultured seabream, *Sparus auratus* L., and wild mullet, *Liza klunzingeri* (Day), in Kuwait. *Journal of Fish Diseases*, 25, 505–513.
- Evans, J. J., Klesius, P. H., Pasnik, D. J., & Bohnsack, J. F. (2009). Human *Streptococcus agalactiae* in Nile Tilapia (*Oreochromis niloticus*). *Emerging Infectious Diseases*, 15(5), 774–776.
- Evans, J. J., Pasnik, D. J., & Klesius, P. H. (2010). A commercial rapid optical immunoassay detects *Streptococcus agalactiae* from aquatic cultures and clinical specimens. *Veterinary Microbiology*, 144, 422–428.
- Evans, J. J., Pasnik, D. J., Klesius, P. H., & Al-Ablani, S. (2006). First report of *Streptococcus agalactiae* and *Lactococcus garvieae* from a wild bottlenose dolphin (*Tursiops truncatus*). *Journal of Wildlife Diseases*, 42(3), 561–569.
- Evans, J. J., Shoemaker, C. A., & Klesius, P. H. (2000). Experimental *Streptococcus iniae* infection of hybrid striped bass ž *Morone chrysops* = *Morone saxatilis* / and tilapia ž *Oreochromis niloticus* / by nares inoculation, 197–210.
- Evans, J. J., Shoemaker, C. A., & Klesius, P. H. (2001). Distribution of *Streptococcus iniae* in hybrid striped bass (*Morone chrysops* × *Morone saxatilis*) following nares inoculation. *Aquaculture*, 194, 233–243.
- Facklam, R. (2002). What happened to the streptococci: overview of taxonomic and nomenclature changes. *Clinical Microbiology Review*. *Clinical Microbiology Review*, 15(4), 613–630.
- Fausch, K. D. (1998). Interspecific competition and juvenile Atlantic salmon (*Salmo salar*): on testing effects and evaluating the evidence across scales. *Canadian Journal of Fisheries and Aquatic Sciences*, 55(S1), 218–231.
- Ferguson, H. W. (2006). Systemic pathology of fish. A text and atlas of comparative tissue responses in diseases of teleosts. United Kingdom: Scotian Press.

- Filho C I, Müller E E, Pretto-Giordano L G, & BracarenseAna Paula F. R. L. (2009). Histological findings of experimental *Streptococcus agalactiae* infection in Nile tilapias (*Oreochromis niloticus*). *Brazilian Journal of Veterinary Pathology*, 2(1), 12–15.
- Finkbeiner, W. E., Connolly, A. J., Ursell, P. C., & Davis, R. L. (2009). *Autopsy Pathology: A Manual and Atlas E-Book*. Elsevier Health Sciences.
- Fisheries, F. A. O. (2010). *Aquaculture Department (2009) The state of world fisheries and aquaculture 2010*. Rome.
- Garcia, J. C., Klesius, P. H., Evans, J. J., & Shoemaker, C. A. (2008). Non infectivity of cattle *S. agalactiae* in Nile tilapia (*O. niloticus*) and channel catfish (*Ictalurus punctatus*). *Aquaculture*, 281, 151–154.
- Gozlan, R. E., Britton, J. R., Cowx, I., & Copp, G. H. (2010). Current knowledge on non-native freshwater fish introductions. *Journal of Fish Biology*, 76(4), 751–786. <http://doi.org/10.1111/j.1095-8649.2010.02566.x>
- Gozlan, R. E., Peeler, E. J., Longshaw, M., St-Hilaire, S. & Feist, S. W. (2006). Effect of microbial pathogens on the diversity of aquatic populations, notably in Europe. *Microbes and Infection* 8, 1358–1364
- Grunwald, D. J., & Eisen, J. S. (2002). Headwaters of the zebrafish: emergence of a new model vertebrate. *Nature Reviews Genetics*, 3(9), 717–724.
- Guerrini, A. (2003). *Experimenting with humans and animals: from galen to animal rights*. Baltimore, M. D.: The John Hopkins University Press.
- Gurevitch, J., Morrow, L. L., Wallace, A., & Walsh, J. S. (1992). A meta-analysis of competition in field experiments. *American Naturalist*, 539–572.
- Herbomel, P., Thisse, B., & Thisse, C. (1999). Ontogeny and behaviour of early macrophages in the zebrafish embryo. *Development*, 126, 3735–3745.
- Hernandez, E., Figueroa, J., & Iregui, C. (2009). Streptococcosis on a red tilapia, *Oreochromis* sp., farm: a case study. *Journal of Fish Diseases*, 32(3), 247–252.
- Hetzl, U., König, A., Yildirim, A. Ö., Lämmler, C., & Kipar, A. (2003). Septicaemia in emerald monitors (*Varanus prasinus* Schlegel 1839) caused by *Streptococcus agalactiae* acquired from mice. *Veterinary Microbiology*, 95(4), 283–293.
- Hii, Y. S., Law, A.T., Shazili, N.A.M., Abdul Rashid, M.K., & Mohd Lokman, H. (2006). The Straits of Malacca: Hydrological Parameters, Biochemical Oxygen Demand and Total Suspended Solid. *Journal of Sustainability Science and Management*, 1(1), 1–14.
- Hunn, J. B. (1989). History of Acute Toxicity Tests with Fish, 1863-87.

Investigations in Fish Control.

- Huntingford, F. A., & Ruiz-Gomez, M. L. (2009). Three-spined sticklebacks *Gasterosteus aculeatus* as a model for exploring behavioural biology. *Journal of Fish Biology*, 147, 641–660.
- Ibrahim, N. A., Zaid, M. Y. A., Khaw, H. L., El-Naggar, G. O., & Ponzoni, R. W. (2013). Relative performance of two Nile tilapia (*Oreochromis niloticus* Linnaeus) strains in Egypt: The Abbassa selection line and the Kafr El Sheikh commercial strain. *Aquaculture Research*, 44(3), 508–517. <http://doi.org/10.1111/j.1365-2109.2012.03240.x>
- Inglis, V., Roberts, R. J., & Bromage, N. R. (1993). *Bacterial Diseases of Fish*. Oxford: Blackwell Scientific Publication.
- Inoue, J. G., Miya, M., Tsukamoto, K., & Nishida, M. (2003). Basal actinopterygian relationships: a mitogenomic perspective on the phylogeny of the “ancient fish.” *Molecular Phylogenetics and Evolution*, 26(1), 110–120.
- Inoue, K., & Takei, Y. (2002). Diverse adaptability in *Oryzias* species to high environmental salinity. *Zoological science*, 19(7), 727-734.
- Iregui, C., Barato, P., Rey, A., Vasquez, G., & Verjan, N. (2014). Epidemiology of *Streptococcus agalactiae* and Streptococcosis in Tilapia Fish. *Concept Press Ltd*, 18.
- Ishikawa, Y. (2000). Medakafish as a model system for vertebrate developmental genetics. *BioEssays*, 22(5), 487–495. [http://doi.org/10.1002/\(SICI\)1521-1878\(200005\)22:5<487::AID-BIES11>3.0.CO;2-8](http://doi.org/10.1002/(SICI)1521-1878(200005)22:5<487::AID-BIES11>3.0.CO;2-8)
- Ismail, A., & Yusof, S. (2011). Effect of mercury and cadmium on early life stages of Java medaka (*Oryzias javanicus*): A potential tropical test fish. *Marine Biotechnology*, 63(5–12), 347–349.
- Ismail, A., Yusof, S., & Rahman, F. (2014). Notes on the occurrence of medaka fish in South Johor, Malaysia, 66, 36–40.
- Iwamatsu, T. (1993). The biology of the Medaka. *Saienchituso-Sha*, 5, 41–44.
- Iwamatsu, T., Imski A., Kawamoto, A. & inden, A. (1982). On *Oryzias javanicus* collected at Jakarta, Singapore an West Kalimantan. *Annotationes Zoologici Japonenses*, 55(3).
- Iwamatsu, T., Ohta, T., & Oshima, E. (1988). Medaka ABSTRACT — An SDS-PAGE JEM-IOOB JEM-IOOCX, 373, 353–373.
- Jiménez, A., Tibatá, V., Junca, H., Ariza, F., Verjan, N., & Iregui, C. (2011). Evaluating a nested-PCR assay for detecting *Streptococcus agalactiae* in red tilapia (*Oreochromis* sp.) tissue. *Aquaculture*, 31, 203–206.

- Johnson-sommer, M. (2003). Principles of Disease and Epidemiology Chapter 14 Introduction Pathology , Infection and Disease.
- Johri, A. K., Paoletti, L. C., Glaser, P., Dua, M., Sharma, P. K., Grandi, G., & Rappuoli, R. (2006a). Group B *Streptococcus*: global incidence and vaccine development. *Nature Reviews Microbiology*, 4(12), 932–942.
- Jordan, D. S., & Snyder, J. O. (1906). A review of the Poeciliidae or killifishes of Japan. *Proceedings of the United States National Museum*, 31(1486), 287–290. <http://doi.org/10.5479/si.00963801.31-1486.287>
- Ku. Yaacob.,K.K, Ali A., & Mohd Isa, M. (2007). *Keadaan Laut Perairan Semenanjung Malaysia untuk Panduan Nelayan*.
- Kayansamruaj, P., Pirarat, N., Kondo, H., Hirono, I., & Rodkhum, C. (2015). Genomic comparison between pathogenic *Streptococcus agalactiae* isolated from Nile tilapia in Thailand and fish-derived ST7 strains. *Infection, Genetics and Evolution*, 36, 307–314. <http://doi.org/10.1016/j.meegid.2015.10.009>
- Kinoshita, M., Murata, K., Naruse, K., & Tanaka, M. (2009). *Medaka:biology, management, and experimental protocols*.
- Kiso, K., & Mahyam, M. I. (2003). Distribution and feeding habits of juvenile and young John's snapper *Lutjanus johnii* in the Matang mangrove estuary, west coast of Peninsular Malaysia. *Fisheries Science*, 69(3), 563–568.
- Kitao, T., Aoki, T., & Sakoh, R. (1981). Epizootic caused by β -haemolytic *Streptococcus* species in cultured freshwater fish. *Fish Pathology*, 15, 301–307.
- Klesius, P. H., Shoemaker, C. A., & Evans, J. J. (1999). Efficacy of a killed *S. iniae* vaccine in tilapia (*O. niloticus*). *Bulletin of European Association of Fish Pathology*, 19, 39–41.
- Klesius, P. H., Shoemaker, C. A., & Evans, J. J. (2000). Vaccination: A health management practice for preventing diseases caused by *Streptococcus* in tilapia and other cultured fish. In K. Fitzsimmons & J. C. Filho (Eds.), *Tilapia Aquaculture in the 21st Century, Fifth International Symposium on Tilapia Aquaculture* (pp. 558–564). Rio de Janeiro, Brazil.
- Klesius, P. H., Shoemaker, C. A., & Evans, J. J. (2008). *Streptococcus*: a worldwide fish health problem. In *In Proceeding of 8th International Symposium on Tilapia in Aquaculture ed.* (pp. 83–107). Cairo.
- Kondo, M., Nanda, I., Schmid, M., & Scharl, M. (2009). Sex determination and sex chromosome evolution: insights from medaka. *Sexual Development*, 3, 88–98.
- Kopp, E. B., & Medzhitov, R. (1999). Kopp, E. B., & Medzhitov, R. The Toll-receptor family and control of innate immunity. *Current Opinion in*

Immunology, 11(1), 13–18.

- Koyama, J., Imai, S., Fujii, K., Kawai, S. I., Yap, C. K., & Ismail, A. (2006). Pollution by estrogens in river and estuarine waters around Kuala Lumpur, Malaysia, and their effects on the estuarine Java-medaka, *Oryzias javanicus*. *環境毒性学会誌*, 9(2), 141-147. *環境毒性学会誌*, 9(2), 141-147., 9(2), 141–147.
- Kusuda, R., & Salati, F. (1999). *Enterococcus seriolicida* and *Streptococcus iniae*. In P. T. K. Woo & D. W. Bruno (Eds.), *Fish Diseases and Disorders: Viral, bacterial and fungal infections* (pp. 303–317). London, United Kingdom: CABI Publishing.
- Leonelli, S. (2007). Growing weed, producing knowledge: an epistemic history of *Arabidopsis thaliana*. *Hlstory and Philosophy of the Life Sciences*, 29, 55–87.
- Leonelli, S., & Ankeny, R. a. (2013). What makes a model organism? *Endeavour*, 37(4), 209–212. <http://doi.org/10.1016/j.endeavour.2013.06.001>
- Liew, W. C., Bartfai, R., Lim, Z., Sreenivasan, R., Siegfried, K. R., & Orban, L. (2012). Polygenic sex determination system in zebrafish. *PLoS ONE*, 7(4), e34397. <http://doi.org/10.1371/journal.pone.0034397>
- Lin, B., Chen, S., Cao, Z., Lin, Y., Mo, D., Zhang, H., ... Xu, A. (2007). Acute phase response in zebrafish upon *Aeromonas salmonicida* and *Staphylococcus aureus* infection: Striking similarities and obvious differences with mammals. *Molecular Immunology*, 44(4), 295–301. <http://doi.org/10.1016/j.molimm.2006.03.001>
- Magtoon, W., & Termvidchakorn, A. (2009). A Revised Taxonomic Account of Ricefish *Oryzias* (Beloniformes; Adrianichthyidae), in Thailand , Indonesia and Japan. *The Natural History Journal of Chulalongkorn University*, 9(April), 35–68.
- Mahajan-Miklos, S., Rahme, L. G., & Ausubel, F. M. (2000). Elucidating the molecular mechanisms of bacterial virulence using non-mammalian hosts. *Molecular Microbiology*, 37(5), 981–988. <http://doi.org/10.1046/j.1365-2958.2000.02056.x>
- Manangeeswaran, M., Ireland, D. D., Sykes, J., McWilliams, I., & Verthelyi, D. (2017). Neonatal mouse model to study Zika virus pathogenesis: Host immune response determines ZIKV tropism and outcome of disease. *The Journal of Immunology*, 198(1).
- Mansfield, B. E., Dionne, M. S., Schneider, D. S., & Freitag, N. E. (2003). Exploration of host–pathogen interactions using *Listeria monocytogenes* and *Drosophila melanogaster*. *Cellular Microbiology*, 5(12), 901–911.
- McDowall., R. M. (2006). Crying wolf, crying foul, or crying shame: alien salmonids and a biodiversity crisis in the southern cool-temperate

- galaxioid fishes? *Reviews in Fish Biology and Fisheries*, 16(3), 233–422.
- Mian, G. F., Godoy, D. T., Leal, C. A. G., Yuhara, T. Y., Costa, G. M., & Figueiredo, H. C. P. (2009). Aspects of the natural history and virulence of *S. agalactiae* infection in Nile tilapia. *Veterinary Microbiology*, 139, 180–183.
- Miller, J. D., & Neely, M. N. (2004). Zebrafish as a model host for streptococcal pathogenesis. *Acta Tropica*, 91(1), 53–68. <http://doi.org/10.1016/j.actatropica.2003.10.020>
- Miya, M., Satoh, T. P., & Nishida, M. (2005). The phylogenetic position of toadfishes (order Batrachoidiformes) in the higher ray-finned fish as inferred from partitioned Bayesian analysis of 102 whole mitochondrial genome sequences. *Biological Journal of the Linnean Society*, 85(3), 289–306.
- Mohammad Noor Amal Azmai, & Zamri-Saad, M. (2011). Streptococcosis in Tilapia (*Oreochromis niloticus*): A review. *Pertanika Journal of Tropical Agricultural Science*, 34(2), 195–206.
- Mohammadi, F., Mousavi, S. M., & Rezaie, A. (2012). Histopathological study of parasitic infestation of skin and gill on Oscar (*Astronotus ocellatus*) and discus (*Symphysodon discus*). *International Journal of the Bioflux Society*, 5(2), 88–89.
- Mosi, L., Mutoji, N. K., Basile, F. A., Donnell, R., Jackson, K. L., Spangenberg, T., ... & Small, P. L. (2012). *Mycobacterium ulcerans* causes minimal pathogenesis and colonization in medaka (*Oryzias latipes*): an experimental fish model of disease transmission. *Microbes and infection*, 14(9), 719–729.
- Musa, N., Wei, L. S., Musa, N., Hamdan, R. H., Leong, L. K., Wee, W., ... Abdullah, S. Z. (2009). Streptococcosis in red hybrid tilapia (*Oreochromis niloticus*) commercial farms in Malaysia. *Aquaculture Research*, 40(5), 630–632. <http://doi.org/10.1111/j.1365-2109.2008.02142.x>
- Naruse, K. (1996). Classification and phylogeny of fishes of the genus *Oryzias* and its relatives. *Fish Biol J MEDAKA*, 8, 1–9.
- Naruse, K., Shima, A., Matsuda, M., Sakaizumi, M., Iwamatsu, T., Soeroto, B., & Uwa, H. (1993). Description and phylogeny of rice fish and their relatives belonging to the suborder Adrianichthyoidei in Sulawesi, Indonesia. *Fish Biol J MEDAKA*, 5, 11–15.
- Naruse, K., Tanaka, M., Mita, K., Shima, A., Postlethwait, J., & Mitani, H. (2004). A medaka gene map: the trace of ancestral vertebrate proto-chromosomes revealed by comparative gene mapping. *Genome Research*, 14, 820–828.
- Nelson, J. S. (2006). *Fishes of the world* (4th ed.). Hoboken: John Wiley & Sons. <http://doi.org/http://dx.doi.org/>

- Neritic Zone. (2017). Retrieved July 28, 2017, from <https://www.britannica.com/science/neritic-zone>
- Nguyen, H. T., Kanai, K., & Yoshikoshi, K. (2001). Nguyen, H.T., Kanai, K. and Yoshikoshi, K. Immunohistochemical examination of experimental *Streptococcus iniae* infection in Japanese flounder *Paralichthys olivaceus*. *Fish Pathology*, 36(3), 169–178.
- Nguyen, H. T., Kanai, K., & Yoshikoshi, K. (2002). Ecological investigation of *Streptococcus iniae* in cultured Japanese flounder, *Paralichthys olivaceus* using selective isolation procedures. *Aquaculture*, 205, 7–17.
- Novoa, B., & Figueras, A. (2012). Zebrafish: model for the study of inflammation and the innate immune response to infectious diseases. *Advances in Experimental Medicine and Biology*, 946, 253–275.
- Olivares-Fuster, O., Klesius, P. H., Evans, J. J., & Arias, C. R. (2008). Molecular typing of *Streptococcus agalctiae* isolates from fish. *Journal of Fish Diseases*, 31, 277–283.
- Ortega, C., García, I., Irgang, R., Fajardo, R., Tapia-Cammas, D., Acosta, J., & Avendaño-Herrera, R. (2018). First identification and characterization of *Streptococcus iniae* obtained from tilapia (*Oreochromis aureus*) farmed in Mexico. *Journal of fish diseases*.
- Parenti, L. R. (2008). A phylogenetic analysis and taxonomic revision of ricefishes, *Oryzias* and relatives (Beloniformes, Adrianichthyidae). *Zoological Journal of the Linnean Society*, 154(3), 494–610.
- Parenti, L. R., & Soeroto, B. (2004). *Adrianichthys roseni* and *Oryzias nebulosus*, two new ricefish (Atherinomorpha: Beloniformes: Adrianichthyidae) from Lake Poso, Sulawesi, Indonesia. *Ichthyological Research*, 51, 10–19. <http://doi.org/10.1007/s10228-003-0187-1>
- Patterson, H., Saralahti, A., Parikka, M., Dramsi, S., Trieu-Cuot, P., Poyart, C., ... Rämét, M. (2012). Adult zebrafish model of bacterial meningitis in *Streptococcus agalactiae* infection. *Developmental and Comparative Immunology*, 38(3), 447–455. <http://doi.org/10.1016/j.dci.2012.07.007>
- Pavlidis, M., Theodoridi, A., & Tsalafouta, A. (2015). Neuroendocrine regulation of the stress response in adult zebrafish, *Danio rerio*. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 60, 121-131.
- Pereira, U. P., Mian, G. F., Oliveira, I. C. M., Benchetrit, L. C., Costa, G. M., & Figueiredo, H. C. P. (2010). Genotyping of *Streptococcus agalactiae* strains isolated from fish, human and cattle and their virulence potential in Nile tilapia. *Veterinary Microbiology*, 140, 186–192.
- Persson, E., Berg, S., Trollfors, B., Larsson, P., Ek, E., Backhaus, E., ... Johansson, S. (2004). Serotypes and clinical manifestations of invasive group B streptococcal infections in western Sweden 1998–2001. *Clinical Microbiology and Infection*, 10(9), 791–796.

- Phuektes, P., Mansell, P. D., & Browning, G. F. (2001). Multiplex polymerase chain reaction assay for simultaneous detection of *Staphylococcus aureus* and streptococcal causes of bovine mastitis. 1140-1148. *Journal of Dairy Science*, 84(5), 1140–1148.
- Plumb, J. A. (1999). Health Maintenance and Principal Microbial Disease of Cultured Fishes. In *Iowa State Univeristy Press*. Ames. (p. 328pp).
- Popesku, J. T., Martyniuk, C. J., Mennigen, J., Xiong, H. L., Zhang, D. P., & Xia, X. H. (2008). The goldfish (*Carassius auratus*) as a model for neuroendocrine signaling. *Molecular and Cellular Endocrinology*, 293, 43–56.
- Postlethwait, J. H., Yan, Y.-L., Gates, M. A., Horne, S., Amores, A., Brownlie, A., ... Talbot, W. S. (1998). Vertebrate genome evolution and the zebrafish gene map. *Nature Genetics*, 18, 345–349.
- Potapov, A. B., & Lewis, M. A. (2004). Climate and competition: the effect of moving range boundaries on habitat invasibility. *Bulletin of Mathematical Biology*, 66(55), 975–1008.
- Pourgholam, R., Laluei, F., Saeedi, A. A., Zahedi, A., Safari, R., Taghavi, M. J., ... Pourgholam, H. (2011). Distribution and molecular identification of some causative agents of streptococcosis isolated farmed rainbow trout (*Oncorhynchus mykiss*, Walbaum) in Iran. *Iranian Journal of Fisheries Sciences*, 10(1), 109–122.
- Pretto-giordano, L. G., Müller, E. E., Freitas, J. C. De, & Gomes, V. (2010). Evaluation on the Pathogenesis of *Streptococcus agalactiae* in Nile Tilapia (*Oreochromis niloticus*). *Brazilian Archives of Biology an Technology*, 53(February), 87–92.
- Pulido, E. A., Iregui, C. A., Figueroa, J., & Klesius, P. (2004). Estreptococosis en tilapias (*Oreochromis* spp.) cultivadas en Colombia. *Revista AquaTic*, 20, 97–106.
- Ramakrishnan, M. A. (2016). Determination of 50% endpoint titer using a simple formula. *World Journal of Virology*, 5(2), 85–86. <http://doi.org/10.5501/wjv.v5.i2.85>
- Rasheed, V., Limsuwan, C., & Plumb, J. (1985). Histopathology of bullminnows, Baird & Girard, infected with a non- haemolytic group B Streptococcus sp. *Journal of Fish Diseases*, 8, 65–74.
- Rattanachaiakunsopon, P. Phumkhachorn, P. (2009). Prophylactic effect of *Andrographis paniculata* extracts against *Streptococcus agalactiae* infection in Nile tilapia (*Oreochromis niloticus*). *Journal of Bioscience and Bioengineering*, 107(5), 579–582.
- Ribas, L., & Piferrer, F. (2014). The zebrafish (*Danio rerio*) as a model organism, with emphasis on applications for finfish aquaculture research. *Reviews in Aquaculture*, 209–240. <http://doi.org/10.1111/raq.12041>

- Roberts, T. R. (1998). Systematic observations on tropical Asian medakas or ricefishes of the genus *Oryzias* with descriptions of four new species. *Ichthyological Research*, 45, 213–224.
- Robinson, J. A., & Meyer, F. P. (1966). Streptococcal fish pathogen. *Journal of Bacteriology*, (9), 512.
- Rock, F. L., Hardiman, G., Timans, J. C., Kastelein, R. A., & Bazan, J. F. (1998). A family of human receptors structurally related to *Drosophila* Toll. In *Proceedings of the National Academy of Sciences* (Vol. 95, pp. 588–593).
- Rodkhum, C., Kayansamruaj, P., & Pirarat, N. (2011). Effect of water temperature on susceptibility to *Streptococcus agalactiae* serotype Ia infection in Nile tilapia (*Oreochromis niloticus*). *The Thai Journal of Veterinary Medicine*, 41(3), 309–314.
- Rosenthal, N., & Ashburner, M. (2002). Taking stock of our models: the function and future of stock centres. *National Review Genetics*, 3, 711–717.
- Russo, R., Mitchell, H., & Yanong, R. P. E. (2006). Characterization of *Streptococcus iniae* isolated from ornamental cyprinid fishes and development of challenge models. *Aquaculture*, 256(1–4), 105–110. <http://doi.org/10.1016/j.aquaculture.2006.02.046>
- Salvador, R.; Müller, E.E.; Freitas, J.C.; Leonhadt, J.H.; Pretto-Giordano, L.G. and Dias, J. A. (2005). Isolation and characterization of *Streptococcus* spp. group B in Nile tilapia (*Oreochromis niloticus*) reared in hapas nets and earth nurseries in the northern region of Parana State, Brazil. *Ciência Rural*, 35, 1374–1378.
- Sanders, J. L., Zhou, Y., Moulton, H. M., Moulton, Z. X., McLeod, R., Dubey, J. P., ... & Kent, M. L. (2015). The zebrafish, *Danio rerio*, as a model for *Toxoplasma gondii*: an initial description of infection in fish. *Journal of fish diseases*, 38(7), 675-679.
- Schier, A. F., F., N. S. C., Harvey, M., Malicki, J., SolnicaKrezel, L., & Stainier, D. Y. R. (1996). Mutations affecting the development of the embryonic zebrafish brain. *Development*, 123, 165–178.
- Shiomitsu, K., Kusuda, R., Osuga, H., & Munekiyo, M. (1980). Efficacy of erythromycin (EM) against spontaneous streptococcal infection in cultured yellow tails *Seriola quinqueradiata* was evaluated clinically . The present studi. *Fish Pathology*, 15(1), 17–23.
- Shoemaker, C. A., Klesius, P. H., & Evans, J. J. (2001). Prevalence of *Streptococcus iniae* in tilapia, hybrid striped bass, and channel catfish on commercial fish farms in the United States. *American Journal of Veterinary Research*, 62(2), 174–177.
- Simon, K. S., Townsend, C. R., Biggs, B. J. F., Bowden, W. B., & Frew, R. D.

- (2004). Habitat-specific nitrogen dynamics in New Zealand streams containing native or invasive fish. *Ecosystems*, 7(8), 777–792.
- SIT (2006), The Center for Food Security and Public Health, Iowa State University.
- Siti-Zahrah, A., Padilah, B., Azila, A., Rimatulhana, R., & Shahidan, R. (2008). Multiple streptococcal species infection in cage-cultured red tilapia but showing similar clinical signs. In *Diseases in Asian Aquaculture VI. Fish Health Section, Asian Fisheries Society, Manila, Philippines* (p. 313–320.). Manila, Philippines.
- Slotved, H. C., Kong, F., Lambertsen, L., Sauer, S., & Gilbert, G. L. (2007). Serotype IX, a proposed new *Streptococcus agalactiae* serotype. *Journal of Clinical Microbiology*, 45, 2929–2936.
- Speare, D. J., Ferguson, H. W., Beamish, F. W. M., Yager, J. A., & Yamashiro, S. (1991). Pathology of bacterial gill disease: ultrastructure of branchial lesions. *Journal of Fish Diseases*, 14(1), 1–20. <http://doi.org/http://dx.doi.org/10.1111/j.1365-2761.1991.tb00572.x>
- Spence, R., Gerlach, G., Lawrence, C., & Smith, C. (2008). The behaviour and ecology of the zebrafish, *Danio rerio*. *Biological Reviews*, 83(1), 13–34.
- Suanyuk, N., Kanghear, H., & Khongpradit, R., Supamattaya, K. (2005). *Streptococcus agalactiae* infection in tilapia (*Oreochromis niloticus*). *Songklanakarinn Journal of Science and Technology (Thailand)*.
- Suanyuk, N., Kong, F., Ko, D., Gilbert, G. L., & Supamattaya, K. (2008). Occurrence of rare genotypes of *Streptococcus agalactiae* in cultured red tilapia *Oreochromis* sp. and Nile tilapia *O. niloticus* in Thailand—Relationship to human isolates? *Aquaculture*, 284, 35–40.
- Takeda, H., & Shimada, A. (2010). The art of medaka genetics and genomics: what makes them so unique? In *Annual Review of Genetics* (pp. 217–241). Palo Alto.
- Tan, M. W. (2002a). Cross-species infections and their analysis. *Annual Review of Microbiology*. <http://doi.org/10.1146/annurev.micro.56.012302.161110>
- Tan, S.G. & Yusoff F.M. 2002. Biodiversity in the Straits: What are the opportunities? In: Tropical Marine Environment: Charting Strategies for the Millenium. Yusoff, F.M., Shariff, Ibrahim H.M., Tan S.G. & Tai S.Y. (Eds.) pp 137-154, *Malacca Straits Research and Development Centre (MASDEC)*, University Putra Malaysia
- Taylor, J. S., Braasch, I., Frickey, T., Meyer, A., & Van de Peer, Y. (2003). Genome duplication, a trait shared by 22,000 species of ray-finned fish. *Genome Research*, 13, 382–390.
- Tong, S. L., C.Y., H., & F.Y., P. (2007). Monitoring of Ba, Mn, Cu, and Ni during

- estuarine mixing. *Analytical Science*, 13(1997), 373–378.
- Toranzo, A. E., Magarinos, B., & Romalde, J. L. (2005). A review of the main bacterial fish diseases in mariculture systems. *Aquaculture*, 246, 37–61.
- Villamizar, N., Ribas, L., Vera, L., Piferrer, F. S., & Sanchez V. azquez. (2012). Impact of daily thermocycles on hatching rhythms, larval performance and sex differentiation of zebrafish. *PLoS ONE*, 7(e52153).
- Wakamatsu, Y., Pristiyazhnyuk, S., Kinoshita, M., Tanaka, M., & Ozato, K. (2001). The see-through medaka: a fish model that is transparent throughout life. *Proceedings of the National Academy of Sciences* 98(18), 10046-10050.
- Wallace, R. A., & Selman, K. (1981). Cellular and dynamic aspects of oocyte growth in teleosts. *American Zoologist*, 21(2), 325–343.
- Webster, C. D., & Chhorn, L. (2015). *Minerals. Dietay nutrients, additives, and fish health*. <http://doi.org/10.1002/9781119005568>
- Willett, C. E., Cortes, A., Zuasti, A., & Zapata, A. G. (1999). Early hematopoiesis and developing lymphoid organs in the zebrafish. *Developmental Dynamics*, 214(4), 323–336.
- Willett, C. E., Zapata, A. G., Hopkins, N., & Steiner, L. A. (1997). Expression of ZebrafishragGenes during early development identifies the thymus. *Developmental Biology*, 182(2), 331–341.
- Winn, R. N., Norris, M. B., Brayer, K. J., Torres, C., & Muller, S. L. (2000). Detection of mutations in transgenic fish carrying a bacteriophage λ cll transgene target. In *Proceedings of the National Academy of Sciences* (Vol. 97, pp. 12655–12660).
- Winn, R. N., Norris, M., Muller, S., Torres, C., & Brayer, K. (2001). Bacteriophage λ and plasmid pUR288 transgenic fish models for detecting in vivo mutations. *Marine Biotechnology*, 3, S185–S195.
- Wittbrodt, J., Shima, A., & Scharl, M. (2002). Medaka-A model organism from the far east. *Nature Reviews*, 3, 53–64.
- Wolf, J. C., Dietrich, D. R., Friederich, U., Caunter, J., & Brown, A. R. (2004). Qualitative and quantitative histomorphologic assessment of fathead minnow *Pimephales promelas* gonads as an endpoint for evaluating endocrine-active compounds: a pilot methodology study. *Toxicologic pathology*, 32(5), 600-612.
- Wongsathein, D. (2012). Factors Affecting Experimental *Streptococcus Agalactiae* Infection in Tilapia , *Oreochromis Niloticus*, (September).
- Yamanoue, Y., Miya, M., Inoue, J. G., Matsuura, K., & Nishida, M. (2006). The mitochondrial genome of spotted green pufferfish *Tetraodon nigroviridis*

(Teleostei: Tetraodontiformes) and divergence time estimation among model organisms in fishes. *Genes & genetic systems*, 81(1), 29-39.

Yanong, R. P. E., & Francis-floyd, R. (2013). Streptococcal Infections of Fish. *University of Florida*, 1–5. Retrieved from <http://edis.ifas.ufl.edu>.

Yanong, R. P. E., & Francis-Floyd, R. (2002). *Streptococcal infections of fish. Report from University of Florida. Series from the Department of Fisheries and Aquatic Sciences, Florida Cooperative Extension Service.*

Ye, X., Li, J., Lu, M., Deng, G., Jiang, X., Tian, Y., ... Jian, Q. (2011). Identification and molecular typing of *Streptococcus agalactiae* isolated from pond-cultured tilapia in China. *Fish Sciences*, 77, 623–632.

Yildirim, A. Ö., Lämmle, C., & Weiß, R. (2002). Identification and characterization of *Streptococcus agalactiae* isolated from horses. *Veterinary Microbiology*, 85(1), 31–35.

Yoder, J. A., & Litman, G. W. (2000). Immune-type diversity in the absence of somatic rearrangement. In *Origin and Evolution of the Vertebrate Immune System* (pp. 271–282). Springer Berlin Heidelberg.

Yoder, J. A., Nielsen, M. E., Amemiya, C. T., & Litman, G. W. (2002). Zebrafish as an immunological model system. *Microbes and Infection*, 4(14), 1469–1478.

Yonekura, R., Kohmatsu, Y., & Yuma, M. (2007). Difference in the predation impact enhanced by morphological divergence between introduced fish populations. *Biological Journal of the Linnean Society*, 91(4), 601–610.

Yusof, S., Ismail, A., & Alias, M. S. (2014). Effect of glyphosate-based herbicide on early life stages of Java medaka (*Oryzias javanicus*): A potential tropical test fish. *Marine pollution bulletin*, 85(2), 494-498.

Yusof, S., Ismail, A., & Rahman, F. (2013). Distribution and localities of Java medaka fish. *Journal, Malayan Nature*, 65, 38–46.

Zamri-Saad, M., Amal, M. N. A., & Siti-Zahrah, A. (2010). Pathological changes in red tilapias (*Oreochromis* spp.) naturally infected by *Streptococcus agalactiae*. *Journal of Comparative Pathology*, 143(2), 227–229.

Zappuli, V., Mazzariol, S., Cavicchiolo, L., Petterino, C., Bargelloni, L., & Castagnaro, M. (2005). Fatal necrotizing fasciitis and myotitis in a captive common bottlenose dolphin (*Tursiops truncatus*) associated with *Streptococcus agalactiae*. *Journal of Veterinary Diagnostic Investigation*, 17, 617–622.

BIODATA OF STUDENT

Siti Suhaiba Mastor was born in March 1991 in Pahang, Malaysia. She received her primary education in Sekolah Kebangsaan (LKTP) Kota Gelanggi 2, Jerantut, Pahang and Sekolah Menengah Sains Sultan Haji Ahmad Shah, Kuantan, Pahang for her secondary school.

After receiving her SPM result, she pursued her higher education at Kolej Matrikulasi Kedah, Changlun, Kedah. Siti Suhaiba Mastor received her bachelor degree in Bachelor of Science (Hons) in Biology from Universiti Putra Malaysia in 2014.

