



UNIVERSITI PUTRA MALAYSIA

***EVALUATION OF ALVEOLAR BONE MICROMORPHOLOGICAL
CHANGES IN EXPERIMENTALLY INDUCED PERIODONTAL
DISEASE RAT MODEL BY USING MICRO-COMPUTED
TOMOGRAPHY (MICRO-CT)***

CHEW XUAN-YEE

FPV 2018 16

**EVALUATION OF ALVEOLAR BONE MICROMORPHOLOGICAL
CHANGES IN EXPERIMENTALLY INDUCED PERIODONTAL
DISEASE RAT MODEL BY USING MICRO-COMPUTED
TOMOGRAPHY (MICRO-CT)**

CHEW XUAN-YEE

A project paper submitted to the

Faculty of Veterinary Medicine, Universiti Putra Malaysia

In partial fulfillment of the requirement for the

DEGREE OF DOCTOR OF VETERINARY MEDICINE

Universiti Putra Malaysia

Serdang, Selangor Darul Ehsan.

MARCH 2018

It is hereby certified that we have read this project paper entitled “Evaluation of Alveolar Bone Micromorphological Changes in Experimentally Induced Periodontal Disease Rat Model by Using Micro-Computed Tomography (Micro-CT)”, by Chew Xuan-Yee and in our opinion it is satisfactory in terms of scope, quality, and presentation as partial fulfillment of the requirement for the course VPD 4999–Final Year Project.

DR. CHEN HUI CHENG

DVM (UPM), MVM (UPM), DVSc (GUELPH)

Associate Professor

Department of Companion Animal Medicine and Surgery

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Supervisor)

DR. LAU SENG FONG

DVM (UPM), PhD (UTRECHT)

Senior Lecturer,

Department of Veterinary Clinical Studies,

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Co-Supervisor)

DR. ROZANALIZA RADZI

DVM (UPM), PhD (YAMAGUCHI)

Senior Lecturer,

Department of Veterinary Clinical Studies,

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Co-Supervisor)

DEDICATIONS

FAMILY

To Giek King and Wee Chin for their unconditional love
And Mianzi, Mianxin and Yan-Yu for their continuous support

LECTURERS

To my supervisor and co-supervisor for all their guidance, advices and assistance

FRIENDS

To my beloved classmates and rotamates of batch 2013/2018

ACKNOWLEDGEMENTS

First and foremost I would like to express my deepest gratitude to my caring, supportive and patient supervisors, Assoc Prof Dr Chen Hui Cheng, Dr Lau Seng Fong and Dr Rozanaliza Radzi. Their dedication and guidance throughout the succession of the study are priceless, not to mention their willingness in sharing their immense knowledge and kind advice, helping me on different aspects of the project.

Besides, zillion thanks also goes to my supportive classmates of DVM 2018, study groupmates and rotationmates. Their kindness showed up in every nook and corner, driving us going an extra mile, making this learning process lively and meaningful. Wishing all of you success in future endeavours.

Last but not least, I would like to thank my beloved family for loving and supporting me in every possible way throughout the journey.

CONTENTS

	PAGE NUMBER
TITLE	i
CERTIFICATION	ii
DEDICATIONS	iv
ACKNOWLEDGEMENT	v
LIST OF FIGURES	viii
LIST OF ABBREVIATIONS	ix
ABSTRAK	x
ABSTRACT	xii
1.0 INTRODUCTION	1
2.0 LITERATURE REVIEW	
2.1 Advantages of Micro-Computed Tomography (Micro- CT) in Bone Micromorphological Evaluation	4
2.2 Sprague-Dawley Rats as Experimental Model	6
2.3 Methods to Induce Periodontal Disease in Animal Models	8
2.4 Area of Interest	10
2.5 Bone Parameters	12
3.0 MATERIALS AND METHODS	

3.1 Animal Model and Housing	15
3.2 Experimental Protocol	15
3.2.1 Anaesthesia and Patient Preparation	16
3.2.2 Placement of Ligature	17
3.2.3 Intra gingival Injection of <i>Prophyromonas gingivalis</i> Lipopolysaccharide (Pg-LPS)	18
3.3 Micro-CT Evaluation	20
3.4 Statistical Analysis	21
4.0 RESULTS	22
5.0 DISCUSSION	28
6.0 CONCLUSION	35
7.0 RECOMMENDATIONS	35
8.0 REFERENCES	36
9.0 APPENDICES	41

LIST OF FIGURES

		PAGE NUMBER
Figure 1	Percent Bone Volume, BV/TV changes	22
Figure 2	Trabecular Thickness, TbTh changes	23
Figure 3	Bone Mineral Density, BMD changes	25
Figure 4	Total Porosity, PoTot changes	26

LIST OF ABBREVIATIONS

LPS	Lipopolysaccharide
<i>Pg</i>-LPS	<i>Prophyromonas gingivalis</i> Lipopolysaccharide
Micro-CT	Micro Computed Tomography
BV/TV	Percent Bone Volume
TbTh	Trabecular Thickness
BMD	Bone Mineral Density
PoTot	Total Porosity
<i>Pg/Fn</i>	<i>Prophyromonas gingivalis/Fusobacterium nucleatum</i>
VOI	Volume of Interest
ROI	Region of Interest
p-Akt	Protein Kinase B
TLR4	Toll-like Receptor 4
NF-κB	Nuclear Factor-kappa B
IL-1β	Interleukin1β
IL-6	Interleukin 6
IL-8	Interleukin 8
RANK	Receptor Activator of NF-κB Ligand
BALB/c	An Albino, Laboratory-bred Strain of House Mouse
ANOVA	Analysis of Variance

ABSTRAK

Abstrak daripada kertas projek yang dikemukakan kepada Fakulti Perubatan Veterinar untuk memenuhi sebahagian daripada keperluan kursus VPD 4999–Projek Ilmiah Tahun Akhir

PENILAIAN PERUBAHAN MIKROMORFOLOGI TULANG ALVEOLAR PADA MODEL TIKUS YANG DIJANGKITI PENYAKIT PERIODONTAL SECARA EKSPERIMEN DENGAN PENGGUNAAN TOMOGRAFI TERKOMPUTERAN MIKRO (MICRO-CT)

oleh

Chew Xuan-Yee

2018

Penyelia: Prof. Madya Dr. Chen Hui Cheng

Penyelia bersama: Dr. Lau Seng Fong, Dr. Rozanaliza Radzi

Periodontitis didorong oleh disbiosis bakteria periodontopatogenik yang boleh mengakibatkan kehilangan tulang alveolar. Tujuan kajian ini adalah untuk menilaiperubahan mikromorfologi tulang alveolar yang disebabkan oleh ligatur, suntikan LPS dan gabungan suntikan LPS dalam model tikus. Data imbasan mikro-CT dari 72 hemimaxilla tikus dianalisis menggunakan Skyscan 1076. Kumpulan rawatan terdiri daripada Kawalan, Ligatur, Ligatur-lipopolisakarida (Ligature-LPS) dan *Porphyromonas gingivalis*-lipopolisakarida (*Pg*-LPS). Setiap rawatan terdiri daripada 7, 14 dan 30 hari. Pada hari 7, kumpulan Ligatur and Ligatur-LPS menunjukkan penurunan ketara ($p < 0.05$) dalam jumlah tulang

peratus (BV / TV), ketebalan trabekular (TbTh), dan kepadatan mineral tulang (BMD), manakala jumlah porositi (PoTot) meningkat, menunjukkan penyerapan tulang. Perubahan ini secara beransur-ansur berkurang, dan pada hari ke 30, mereka tidak signifikan ($p > 0.05$) berbanding dengan kumpulan Kawalan. Ini mungkin menunjukkan kejadian pembentukan semula tulang. Kumpulan *Pg*-LPS tidak menunjukkan perubahan ketara dalam semua parameter. Keputusan ini menunjukkan bahawa Ligatur and Ligatur-LPS menyebabkan penyerapan tulang lebih banyak daripada *Pg*-LPS dalam tikus.

Kata kunci: Penyakit periodontal, model Ligatur, model Ligatur-lipopolisakarida (Ligature-LPS), *Porphyromonas gingivalis*-lipopolisakarida(*Pg*-LPS), parameter tulang.

ABSTRACT

An abstract of the project paper presented to the Faculty of Veterinary Medicine in partial fulfillment of the course VPD 4999 Final Year Project.

EVALUATION OF ALVEOLAR BONE MICROMORPHOLOGICAL CHANGES IN EXPERIMENTALLY INDUCED PERIODONTAL DISEASE RAT MODEL BY USING MICRO-COMPUTED TOMOGRAPHY (MICRO-CT)

by

Chew Xuan-Yee

2018

Supervisor: Assoc. Prof. Dr. Chen Hui Cheng

Co-supervisors: Dr. Lau Seng Fong, Dr. Rozanaliza Radzi

Periodontitis is driven by dysbiotic of periodontopathogenic bacteria that may lead to alveolar bone loss. The purpose of this study was to evaluate the alveolar bone micromorphological changes induced by ligature, injection of LPS and combination of ligature-LPS injection in the rat model. Micro-CT scanned data from 72 hemimaxilla of previously induced rats were analysed using Skyscan 1076. Treatment groups consisted of Control, Ligature, Ligature-lipopolysaccharide (Ligature-LPS) and *Porphyromonas gingivalis*-lipopolysaccharide (*Pg*-LPS). Each treatment comprised of day 7, 14 and 30. At day 7, both Ligature and Ligature-LPS groups showed significant decrease ($p < 0.05$) in percent bone volume (BV/TV), trabecular thickness (TbTh), and bone

mineral density (BMD), but increased in total porosity (PoTot), indicating bone resorption. These changes gradually reduced, and by day 30, they were insignificant ($p > 0.05$) when compared to the Control group. This likely indicates the occurrence of bone remodeling. The *Pg*-LPS group showed no significant change in all parameters. These results showed that Ligature and Ligature-LPS caused more bone resorption than *Pg*-LPS in rats.

Keywords: Periodontal disease, Ligature model, Ligature-lipopolysaccharide (Ligature-LPS) model, *Porphyromonas gingivalis*-lipopolysaccharide (*Pg*-LPS), bone parameters

1.0 INTRODUCTION

Periodontitis is defined as a chronic inflammatory disease affecting tooth supporting tissues, initiated by a synergistic and dysbiotic microbial community bacteria as the primary etiologic factor and culminating in the destruction of the dental attachment apparatus (Hajishengallis & Lamont, 2012). Host defense, genetic and environmental factors are responsible for different periodontal conditions that can be observed in humans (Libermanl et al., 2011). While gingivitis does not affect the underlying supporting structures of the teeth, periodontitis results in loss of connective tissue and bone support and can progress to bone destruction, tooth mobility and finally, tooth loss (Martins et al., 2016).

Various animal species were used in experimental study models in periodontology, for instance, rats, hamsters, rabbits, ferrets, dogs, pigs and primates. All of these animals were used in order to mimic human periodontal diseases in an attempt to reveal pathogenetic mechanisms of periodontal disease and find cure (Balci Yuce, 2016). Rodents, with rats in particular, are important models for experimental periodontal research because rats are easy to handle and inexpensive (Oz & Puleo et al., 2011). One of the most successful study approaches, germ-free or gnotobiotic rats such as Sprague-Dawley rats, made the study of dental plaque and bacterial biofilm in periodontal research possible.

Balci Yuce (2016) depicted that Ligation model and Lipopolysaccharide (LPS) injection model are few periodontal disease induction method that has been used in previous study. Ligation model with silk suture material is the most common way of inducing periodontal disease, however, other retentive device are acceptable to be used as well. The ligatures are thought to facilitate local accumulation of bacteria and thereby enhance bacteria-mediated inflammation and bone loss (Graves et al., 2008). Meanwhile for LPS, it has been suggested that LPS can penetrate gingival connective tissue and induce a local inflammatory response that leads to periodontal bone resorption (Cheng et al., 2010). However, to induce the same lesion, this method may need a longer time as it is associated with inducing chronic periodontal disease (Bostanci & Belibasakis, 2012). On the other hand, combination of Ligature and LPS has not been described clearly in previous studies.

Therefore, this study aims to evaluate the alveolar bone micromorphological changes induced by ligature, injection of LPS and combination of ligature-LPS injection in the Sprague Dawley rat model. The hypotheses were:

Ho: There is no significant difference in Percent Bone Volume (BV/TV), Trabecular Thickness (TbTh), Total Porosity (PoTot) and Bone Mineral Density (BMD) among Ligature, Ligature-LPS and Pg-LPS injection in periodontal disease induction.

Ha: There is significant difference in Percent Bone Volume (BV/TV), Trabecular Thickness (TbTh), Total Porosity (PoTot) and Bone Mineral Density (BMD) among Ligature, Ligature-LPS and Pg-LPS injection in periodontal disease induction.



8.0 REFERENCES

- Abu-Amer, Y. (2013). NF- κ B signaling and bone resorption. *Osteoporosis International*, 24(9), 2377–2386. <https://doi.org/10.1007/s00198-013-2313-x>
- Baiker, M., Snoeks, T. J. A., Kaijzel, E. L., Que, I., Dijkstra, J., Lelieveldt, B. P. F., & Löwik, C. W. G. M. (2012). Automated bone volume and thickness measurements in small animal whole-body MicroCT data. *Molecular Imaging and Biology*, 14(4), 420–430. <https://doi.org/10.1007/s11307-011-0522-2>
- BALCI YUCE, H. (2017). Periodontoloji Alanında Hayvan Çalışmaları: Deneysel Periodontal ve Periimplant Hastalığın İndüksiyonu. *Cumhuriyet Dental Journal*, 20(1), 62–62. <https://doi.org/10.7126/cumudj.307312>
- Bostanci, N., & Belibasakis, G. N. (2012). Porphyromonas gingivalis: An invasive and evasive opportunistic oral pathogen. *FEMS Microbiology Letters*, 333(1), 1–9. <https://doi.org/10.1111/j.1574-6968.2012.02579.x>
- Bouxsein, M. L., Boyd, S. K., Christiansen, B. A., Guldberg, R. E., Jepsen, K. J., & Müller, R. (2010). Guidelines for assessment of bone microstructure in rodents using micro-computed tomography. *Journal of Bone and Mineral Research*, 25(7), 1468–1486. <https://doi.org/10.1002/jbmr.141>
- Bruker. (2017). Analysis of bone by micro-CT General information, 1–41. Retrieved from http://umanitoba.ca/faculties/medicine/units/cacs/sam/media/MN001_Bone_microCT_analysis_general.pdf
- Bruker-microCT. (2012). Morphometric parameters measured by Skyscan™ CT - analyser software . *Reference Manual*, 1–49. Retrieved from <http://bruker-microct.com/next/CTAn03.pdf>
- Charles, J. F., & Aliprantis, A. O. (2014). Osteoclasts: More than “bone eaters.” *Trends in Molecular Medicine*, 20(8), 449–459. <https://doi.org/10.1016/j.molmed.2014.06.001>
- Cheng, W. C., Huang, R. Y., Chiang, C. Y., Chen, J. K., Liu, C. H., Chu, C. L., & Fu, E. (2010). Ameliorative effect of quercetin on the destruction caused by experimental periodontitis in rats. *Journal of Periodontal Research*, 45(6), 788–795. <https://doi.org/10.1111/j.1600-0765.2010.01301.x>
- Cirelli, J. A., Park, C. H., MacKool, K., Taba, M., Lustig, K. H., Burstein, H., & Giannobile, W. V. (2009). AAV2/1-TNFR:Fc gene delivery prevents periodontal disease progression. *Gene Therapy*, 16(3), 426–436.

<https://doi.org/10.1038/gt.2008.174>

- Clark, D. P., & Badea, C. T. (2014). Micro-CT of rodents: State-of-the-art and future perspectives. *Physica Medica*, *30*(6), 619–634. <https://doi.org/10.1016/j.ejmp.2014.05.011>
- Cnudde, V., Cnudde, J. P., Dupuis, C., & Jacobs, P. J. S. (2004). X-ray micro-CT used for the localization of water repellents and consolidants inside natural building stones. *Materials Characterization*, *53*(2–4), 259–271. <https://doi.org/10.1016/j.matchar.2004.08.011>
- Cnudde, V., & Jacobs, P. J. S. (2004). Monitoring of weathering and conservation of building material through non-destructive X-ray computed microtomography. *Environmental Geology*, *46*(3–4), 477–485.
- Coates, M. E. (1973). Gnotobiotic animals in nutrition research. *The Proceedings of the Nutrition Society*, *32*(2), 53–58. <https://doi.org/10.1079/PNS19730015>
- Donos, N., Park, J. C., Vajgel, A., de Carvalho Farias, B., & Dereka, X. (2018). Description of the periodontal pocket in preclinical models: limitations and considerations. *Periodontology 2000*, *76*(1), 16–34. <https://doi.org/10.1111/prd.12155>
- Dumitrescu, A. L., El-Aleem, S. A., Morales-Aza, B., & Donaldson, L. F. (2004). A model of periodontitis in the rat: Effect of lipopolysaccharide on bone resorption, osteoclast activity, and local peptidergic innervation. *Journal of Clinical Periodontology*, *31*(8), 596–603. <https://doi.org/10.1111/j.1600-051X.2004.00528.x>
- Explanation and Recommendations for Reporting of CT Parameters Current as of October 31, 2013 Recommended Set of Cortical Parameters: Total Area (TA), Bone Area (BA), Medullary Area (MA), Cortical Thickness (Ct.Th), Polar Moment of Inertia (p. (2013).
- Feldkamp, L. A., Goldstein, S. A., Parfitt, A. M., Jesion, G., Kleerekoper, M. (1989). The Direct Examination of Three-Dimensional Bone Architecture In Vitro by Computed Tomography. *Journal of Bone and Mineral Research*, *4*(1)
- Graves, D. T., Fine, D., Teng, Y. T. A., Van Dyke, T. E., & Hajishengallis, G. (2008). The use of rodent models to investigate host-bacteria interactions related to periodontal diseases. *Journal of Clinical Periodontology*, *35*(2), 89–105. <https://doi.org/10.1111/j.1600-051X.2007.01172.x>
- Gunter, C. (2002). Human biology by proxy. *Nature*, *420*(6915), 509. <https://doi.org/10.1038/420509a>

- Hajishengallis, G., & Lamont, R. J. (2012). Beyond the red complex and into more complexity: The polymicrobial synergy and dysbiosis (PSD) model of periodontal disease etiology. *Molecular Oral Microbiology*, 27(6), 409–419. <https://doi.org/10.1111/j.2041-1014.2012.00663.x>
- Hana, H. M., Radzi, R., Lau, S. F., Noordin, M. M., & Chen, H. C. (2017). Induction of periodontal disease via retentive ligature, lipopolysaccharide injection and their combination in a rat model. Proc. 29th VAM Congress Malaysia. Universiti Putra Malaysia, Serdang.
- Hwang, S. Y., & Putney, J. W. (2011). Calcium signaling in osteoclasts. *Biochimica et Biophysica Acta - Molecular Cell Research*, 1813(5), 979–983. <https://doi.org/10.1016/j.bbamcr.2010.11.002>
- Karatas, O. H., & Toy, E. (2014). Three-dimensional imaging techniques: A literature review. *European Journal of Dentistry*, 8(1), 132–140. <https://doi.org/10.4103/1305-7456.126269>
- Kesavalu, L., Sathishkumar, S., Bakthavatchalu, V., Matthews, C., Dawson, D., Steffen, M., & Ebersole, J. L. (2007). Rat model of polymicrobial infection, immunity, and alveolar bone resorption in periodontal disease. *Infection and Immunity*, 75(4), 1704–1712. <https://doi.org/10.1128/IAI.00733-06>
- Liberman, D. N., Pilau, R. M., Orlandini, L. F., Gaio, E. J., & Rösing, C. K. (2010). Comparison of two methods for alveolar bone loss measurement in an experimental periodontal disease model in rats. *Brazilian Oral Research*, 25(1), 80–84. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/21271178>
- Lu, H., Xu, M., Wang, F., Liu, S., Gu, J., Lin, S., & Zhao, L. (2016). Chronic stress accelerates ligature-induced periodontitis by suppressing glucocorticoid receptor signaling. *Experimental and Molecular Medicine*, 48(3), e223-10. <https://doi.org/10.1038/emm.2015.127>
- Martins, C. S., Leitão, R. F. C., Costa, D. V. S., Melo, I. M., Santos, G. S., Lima, V., Baldim, V., Wong, D. V. T., Bonfim, L. E., Melo, C. B., de Oliveira, M. G., Brito, G. A. C. (2016). Topical HPMC/S-nitrosoglutathione solution decreases inflammation and bone resorption in experimental periodontal disease in rats. *PLoS ONE*, 11(4), 1–19. <https://doi.org/10.1371/journal.pone.0153716>
- Matsuda, Y., Kato, T., Takahashi, N., Nakajima, M., Arimatsu, K., Minagawa, T., Sato, K., Ohno, H., Yamazaki, K. (2016). Ligature-induced periodontitis in mice induces elevated levels of circulating interleukin-6 but shows only weak effects on adipose and liver tissues. *Journal of Periodontal Research*, 51(5), 639–646. <https://doi.org/10.1111/jre.12344>

- Oz, H. S., & Puleo, D. A. (2011). Animal models for periodontal disease. *Journal of Biomedicine and Biotechnology*, 2011. <https://doi.org/10.1155/2011/754857>
- Park, C. H., Abramson, Z. R., Taba, M., Jin, Q., Chang, J., Kreider, J. M., Giannobile, W. V. (2007). Three-Dimensional Micro-Computed Tomographic Imaging of Alveolar Bone in Experimental Bone Loss or Repair. *Journal of Periodontology*, 78(2), 273–281. <https://doi.org/10.1902/jop.2007.060252>
- Particelli, F., Mecozzi, L., Beraudi, A., Montesi, M., Baruffaldi, F., & Viceconti, M. (2012). A comparison between micro-CT and histology for the evaluation of cortical bone: Effect of polymethylmethacrylate embedding on structural parameters. *Journal of Microscopy*, 245(3), 302–310. <https://doi.org/10.1111/j.1365-2818.2011.03573.x>
- Paz ICL, A., Ldg, B., & Bruno, L. (2006). Bone mineral density: review. *Brazilian Journal of Poultry Science Revista Brasileira*, 8(2), 69–73.
- Prabhakar, A. R., Chakraborty, A., Nadig, B., & Yavagal, C. (2017). Finite element stress analysis of restored primary teeth : A comparative evaluation between stainless steel crowns and preformed zirconia crowns, 68–75. <https://doi.org/10.4103/ijohs.ijohs>
- Putnins, E. E. (2014). *Studies on Periodontal Disease*. <https://doi.org/10.1007/978-1-4614-9557-4>
- Schindeler, A., Mills, R. J., Bobyn, J. D., & Little, D. G. (2017). Preclinical Models for Orthopedic Research and Bone Tissue Engineering. *Journal of Orthopaedic Research*, (July 2017), 1–9. <https://doi.org/10.1002/jor.23824>
- Struillou, X., Boutigny, H., Soueidan, A., & Layrolle, P. (2010). Experimental animal models in periodontology: a review. *The Open Dentistry Journal*, 4, 37–47. <https://doi.org/10.2174/1874210601004010037>
- Truyers, I. (2015). The use of micro - computed tomography in the diagnosis of dental and oral disease in rabbits ., (7), 2014–2015. <https://doi.org/10.1186/s12917>
- Turnbaugh, P. J., Ridaura, V. K., Faith, J. J., Rey, F. E., Knight, R., & Gordon, J. I. (2009). The Effect of Diet on the Human Gut Microbiome: A Metagenomic Analysis in Humanized Gnotobiotic Mice. *Science Translational Medicine*, 1(6), 6ra14–6ra14. <https://doi.org/10.1126/scitranslmed.3000322>
- Vignery, A., Baron, R. (1980). Dynamic histomorphometry of alveolar bone

remodeling in the adult rat. *General Histology and Cytology*. 196(2), 191-200. <http://onlinelibrary.wiley.com/doi/10.1002/ar.1091960210/full>



COPYRIGHT UPM