

UNIVERSITI PUTRA MALAYSIA

PATHOGENICITY STUDY OF EXTRACELLULAR PROTEINS (ECPs) AND CELLULAR MEMBRANE PROTEINS (CMPs) OF Streptococcus agalactiae AND ITS EFFECT AS IMMUNOMODULATOR IN AFRICAN CATFISH (Clariasgariepinus)

NOR ANISKIHA MAT YUNUS

FPV 2016 67

PATHOGENICITY STUDY OF EXTRACELLULAR PROTEINS (ECPs) AND CELLULAR MEMBRANE PROTEINS (CMPs) OF Streptococcus agalactiae AND ITS EFFECT AS IMMUNOMODULATOR IN AFRICAN CATFISH (Clariasgariepinus)

UPM

NOR ANISKIHA MAT YUNUS

A project paper submitted to the Faculty of Veterinary Medicine, Universiti Putra Malaysia In partial fulfilment of the requirement for the DEGREE OF DOCTOR OF VETERINARY MEDICINE Universiti Putra Malaysia Serdang, Selangor DarulEhsan

March 2016

It is hereby certified that I have read this project paper entitled "Pathogenicity Study of Extracellular Proteins (ECPs) and Cellular Membrane Proteins (CMPs) of *Streptococcus agalactiae* and its effect as immunomodulator in African Catfish (*Clariasgariepinus*)", by Nor Aniskiha Mat Yunus and in my opinion it is satisfactory in terms of scope, quality and presentation as partial fulfilment of the requirement for the course VPD 4999-Project

ASSOC. PROFESSOR DR. HASSAN HJ. MOHD DAUD

DVM (UPM), MSc (Stirling, Scotland), PhD (Kingston, England)

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Supervisor)

DR. MOHD FUAD MATORI

DVM (UPM)

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Co-supervisor)

Specially dedicated to my beloved

parents, family and friends

ACKNOWLEDGEMENTS

Alhamdulillah....

First and foremost, my humble thanks to Allah SWT, for giving me the patience, strength and ideas in completing my final year project. A very sinceregratitude and appreciation to my supervisor, Assoc. Prof. Dr Hassan HjMohdDaudfor his support, guidance, and patience in the process of completion of this project. I also would like to express my gratitude to my co-supervisor Dr. MohdFuadMatori for all the help and guidance.

I also would like to thank all the staffs and postgraduates students at the Aquatic Animal Health Unit (AAHU) En.ZainalFitri, En. Azmi, Pn.Raina,CikKumari, CikFairuz,CikNora FatenAfifah, CikNadia, and CikDayah for their help and guidance. Thank you also to the staffs at the Aquatic Lab, FPV, PnLatifah and Dr Diyana.

Special appreciation is also dedicated to my family who have given me the love, care and moral support to the success of this project. Very sincere gratitude to my special friend Humairak, thathad been helping me as well as assisting me to complete my final year project. Last but not least, to all my friends whom have helped me in one way to another.

Thank you.....

iv

CONTENTS

		Page
	TITLE	ì
	CERTIFICATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	CONTENTS	v
	LIST OF TABLES	vii
	LIST OF FIGURES	viii
	LIST OF PLATES	ix
	LIST OF ABBREVIATION	X
	ABSTRAK	xi
	ABSTRACT	xiii
	CHAPTER	
	Chapter 1.0: INTRODUCTION	1
	Chapter 2.0: LITERATURE REVIEW	
	2.1: African catfish (<i>Clariasgariepinus</i>)	4
	2.2: Streptococcus agalactiae	4
	2.3: Clinical signs of S. agalactiae infection	5
	2.4: Pathogenicity of S. agalactiae	6
	2.5: Extracellular proteins (ECPs)	6
	2.6: Cellular Membrane Proteins (CMPs)	6

Chapter 3	3.0: MA	TERIALS	AND	METHOD
Chapter 3	3.0: MA	TERIALS	AND	METHOD

3.1: Experimental fish	8
3.2: Preparation of bacteria inoculum	8
3.2.1: Preparation of Cellular Membrane Proteins (CMPs)	9
3.2.2: Preparation of Extracellular Proteins (ECPs)	9
3.3: Experimental infection trials	9
3.4: Blood sample collection	10
3.5 Immunogenicity testing	10
3.5.1: Agar Gel Precipitation Test (AGPT)	10
3.6: Evaluation	11
3.7: Statistical Analysis	11
Chapter 4.0: RESULTS	12
Chapter 5.0: DISCUSSION	21
Chapter 6.0: CONCLUSION AND RECOMMENDATION	24
Chapter 7.0: REFERENCES	26
Chapter 8.0: APPENDICES	29

LIST OF TABLES

Table 1: Total cumulative mortality and percentage survivability recorded for

 fingerlings *Clariasgariepinus*intraperitoneally injected with Cellular MembraneProteins

 (CMPs) of *S. agalactiae*.

Table 2: Total cumulative mortality and percentage survivability recorded for

 fingerlings
 *Clariasgariepinus*intraperitoneally injected with Extracellular Proteins

 (ECPs) of *S. agalactiae*.

Table 3: SPSS Output 1- The significant between dilution and percentage survivability

 for CMPs and ECPs injected fingerlings.

 Table 4: SPSS Output 2- The significant between bacteria products on mortality of the fingerlings.

LIST OF FIGURES

Figure 1: Graph on relationship between ECPs and CMPs on cumulative mortality of fingerlings.

Figure 2: Percentage Survivability of every dilution of CMPs injected fingerlings

Figure 3: Percentage Survivability of every dilution of ECPs injected fingerlings



LIST OF PLATES

Plate 1:Lethargic fingerlings from ECPs and CMPs group seen at 24 hpi

Plate 2:Inappetance seen at 2nd dpi from ECPs and CMPs group

Plate 3:Erratic swimming behaviour at 3rd dpi from ECPs group

Plate 4:Corneal opacity at 2nd dpi from CMPs group

Plate 5: Exopthalmia seen on 3rd dpi from CMPs group

Plate 6:Caudal and dorsal fin rot at 3rd dpi from ECPs group

Plate 7: AGPT results of CMPs injected fish sera

Plate 8: AGPT results of ECPs injected fish sera

LIST OF ABBREVIATIONS

%	Percent
μl	Micro liter
mL	Milliliter
° C	Degree Celcius
i.p	Intraperitoneal
hpi	Hour post inoculation
dpi	Day post inoculation
SPSS	Statistical Package for the Social Sciences
CFU/ml	Colony forming unit per ml
ECPs	Extracellular Proteins
CMPs	Cellular Membrane Proteins
AGPT	Agar Gel Precipitation Test

ABSTRAK

Abstrakdaripadakertaskerjaprojek yang

dikemukakankepadaFakultiPerubatanVeterinaruntukmemenuhisebahagiandaripadakeper luankursus VPD 4999- ProjekIlmiahTahunAkhirPelajar.

KAJIAN PATOGENISITIPROTEIN EKSTRASELULAR (PES) DAN PROTEIN MEMBRAN SEL (PMS) *Streptococcus agalactiae* DAN KESAN SEBAGAI IMUNOMODULATOR DALAM IKAN KELI AFRIKA (*Clarias gariepinus*)

Oleh

Nor Aniskiha Mat Yunus

2016

Penyelia: Prof. MadyaDr Hassan Hj. MohdDaud Penyeliabersama: DrMohdFuadMatori

 \bigcirc

Protein Membran Sel (PMS) adalah protein permukaan bakteria yang boleh menjadi sumber immunogens manakala Protein Ekstraselular (PES) adalah produk yang dikeluarkan oleh bakteria yang dapat mengaktifkan tindak balas imun hos. Walau bagaimanapun, terdapat kekurangan dalam kajian sebelum ini untuk menilai tindak balas imun ikan keli terhadap PES dan PMS menggunakan AGPT. Kajian ini bertujuan untuk menilai kesan *in vivo*PES dan PMSbakteria *Streptococcus agalactiae* terhadap patogenisiti dan imuniti ikan keli Afrika (*Clarias gariepinus*).

PES dan PMS daripada *S.agalactiae* disuntik secara intraperitoneal (i,p) ke dalam ikan keli Afrika untuk menentukan sama ada bakteria produk dapat menyebabkan patogenisiti dan merangsang tindak balas imun ikan. Bakteria daripada kultur asal diambil menggunakan proses emparan pada 1800xg selama 15 minit untuk memisahkan PES dan PMS daripada larutan. Pencairan bersiri untuk PES dan PMS dilakukan pada 10⁻¹, 10⁻², 10⁻³, 10⁻⁴, dan 10⁻⁵. Morbiditi, mortaliti kumulatif dan peratusan kemandirian direkodkan setiap hari selama 7 hari. Sampel serum daripada ikan bagi setiap PMS kumpulan kawalan pencairanPES dan serta telah diambil melalui venakaudalpedunkel pada hari ke-8 selepas suntikan. Tanda-tanda klinikal seperti tiada selera makan, lesu, berenang tidak menentu, kelegapan kornea dan eksoptalmia diperhatikan dan sera diuji untuk melihat tahap kehadiran antibodi menggunakan Ujian Presipitasi Agar Gel (AGPT).

Hasil kajian menunjukkan bahawa PMSlebih virulen daripada PES berdasarkan kematian, bagaimanapun, secara statistik tidak ada perbezaan yang signifikan di antara ikan disuntik dengan PES dan PMS pada p <0.05. Keputusan juga menunjukkan bahawa tidak terdapat perbezaan yang signifikan di antara pencairan inokula dan peratusan kemandirianuntuk kedua-dua PES dan PMS. Analisis serum melalui AGPT menunjukkan negatif untuk semua PES dan PMS dan ia menunjukkan bahawa tidak ada kompleks antigen-antibodi yang terbentuk bagi setiap pencairan.

6

Kata kunci: *Streptococcus agalactiae*, protein ekstraselular (PES), protein membransel (PMS), ikankeliAfrika, imunomodulasi

ABSTRACT

An abstract of the project paper presented to the Faculty of Veterinary Medicine as a partial fulfilment of requirement for the course VPD 4999- Final Year Project.

PATHOGENICITY STUDY OF EXTRACELLULAR PROTEINS (ECPs) AND

CELLULAR MEMBRANE PROTEINS (CMPs) OF Streptococcus agalactiaeAND

ITS EFFECT AS IMMUNOMODULATOR IN AFRICAN CATFISH

(Clariasgariepinus)

By

Nor Aniskiha Mat Yunus

2016

Supervisor: Assoc. Prof. Dr Hassan Hj. MohdDaud

Co-supervisor: Dr MohdFuadMatori

Cellular Membrane Proteins (CMPs) are bacterial surface proteinsthat could be a source of immunogens while the Extracellular protein (ECPs) are bacterial secretory products that able to activate host'simmune response. However, there is lack of previous studies done to evaluate the immune response of catfish against ECPs and CMPs using AGPT. This study aimed to evaluate the *in vivo* effect of the ECPs and CMPs of *Streptococcus agalactiae* on pathogenicity and immunity of the African catfish (*Clariasgariepinus*) fingerlings.

The ECPs and CMPs of *S.agalactiae* were intraperitoneally (i.p) injected into African catfish fingerlings to determine whether the bacteria products able to cause disease and also stimulate fish's immune response. Bacteria cells from pure culture were harvested by centrifugation at 1800xg for 15 minutes to separate ECPs and CMPs from bacteria suspension. Serial dilutions for ECPs and CMPs were done to give 10⁻¹, 10⁻², 10⁻³, 10⁻⁴, and 10⁻⁵ diluents. The morbidity, cumulative mortality and percentage survivability of the fish were recorded within 7dpi. Pooled serum samples from fingerlings of each dilution of ECPs and CMPs injected fish as well as control group were taken via caudal peduncle vein at 8th dpi. Clinical signs observed include anorexia, lethargy, erratic swimming, corneal opacity and exopthalmia. The sera were tested for antibody production by Agar Gel Precipitation Test (AGPT).

Results showed that CMPs was more virulence than ECPs based on mortality, however, statistically there was no significance difference between the fingerlings injected with ECPs and CMPs at p<0.05. Results also showed that there was no significant difference between the dilutions of the inocula and the percentage survivability of fingerlings for both ECPs and CMPs. Serum analysis via AGPT showed negativity for all ECPs and CMPs indicating that there was no antigen-antibody complex formed for every dilution.

Keywords: *Streptococcus agalactiae*, Extracellular Proteins (ECPs), Cellular Membrane Proteins (CMPs), African catfish, immunomodulation

1.0 INTRODUCTION

According to Department of Fisheries Malaysia, (2013) the total freshwater aquaculture in Malaysia reached about 132, 892.42 metric tonnes and about 38% from total production (50,533.79 metric tonnes) was contributed from freshwater catfish production from all culture system. Freshwater catfish production had the highest production compared to other fish species in Malaysia. Therefore, infectious diseases caused by pathogenic organisms such as *Streptococcus agalactiae* are an important issue that has caused a lot of financial loss in the aquaculture industry and it is increasingly being recognized as a potential constraint on aquaculture production and trade, and cause massive financial loss either through mortality or reduced meat quality, resulting in reduced profit margins (Smith *et al.*, 2003).

Streptococcus agalactiae is a group B streptococcus, it was reported to cause neonatal pneumonia and meningitis in human (Brimil *et al.*, 2006; Johri *et al.*, 2006), mastitis in cows (Brochet*et al.*, 2006; Yildirim *et al.*, 2002) and streptococcal infection in fish (Toranzo *et al.*, 2005).*Streptococcusagalactiae* also reported to cause such erratic swimming, loss appetite, exophthalmia and visceral cavity distension as main clinical signs in infected fish.According to Song *et al.* (2013), the development of vaccines is one of the solutions against the pathogen as a sustainable prevention method to control this emerging disease.Besides outer membrane proteins as a source of immune protective immunogens, recent increasing interests have been paid on extracellular secretory proteins The extracellular secretory proteins easily activate host'simmune response since they are secreted out of cells and are easily contacted with the host (Zhang *et al.*, 2012). Thus, investigation on protective immunogens from extracellular proteins may provide efficient candidates for development of vaccines (Song *et al.*, 2013).

The objectives of this study were:

- To identify the Extracellular Proteins (ECPs) and Cellular Membrane Proteins (CMPs) of *S.agalactiae*
- 2) To determine the clinical signs, mortality and Percentage Survivability in *Clariasgariepinus* fish when injected with ECPs and CMPs of *S.agalactiae*
- 3) To study the pathogenicity and immunity of *Clariasgariepinus* fish towards ECPs and CMPsof *S.agalactiae*.

Justification for this study:

Streptococcosis caused by *S.agalactiae* cause high morbidity and mortality in aquaculture industry primarily in cultured fish and cause great financial loss to the farmer. Therefore, the aim of the study is to evaluate the pathogenicity (clinical signs and mortality) and immune response of *Clariasgariepinus* fish when administered with Extracellular Proteins (ECPs) and Cellular Membrane Proteins (CMPs). Besides the host- agent immune response, the antigenic protein of the bacteria is important and potential for vaccine development. Other than that, there is also lack of study done on pathogenicity and immune response of African catfish fingerlings when challenged with ECPs and CMPs of *S. agalactiae*.

Hypothesis of this study were:

The hypothesis for this study is the Extracellular Proteins (ECPs) and Cellular Membrane Proteins (CMP) of *S. agalactiae* is able to produce pathogenicity (in terms of morbidity and mortality) and produce immunity in African catfish (*Clariasgariepinus*) fingerlings.



7.0 REFERENCES

- Abuseliana, A., Daud, H.M., Abdul Aziz, S., Bejo, S., &Alsaid, M. (2011).
 Pathogenicity of Streptococcus agalactiae Isolated from a Fish Farm in Selangor to Juvenile Red Tilapia (Oreochromis sp.). J. Of Animal And Veterinary Advances, 10(7), 914-919.
- Amrullah, Sukenda, Harris, E., Alimuddin, &Lusiastuti, A. M. (2014).Immunogenicity of the 89 kDa Toxin Protein from Extracellular Products of Streptococcus in Oreochromisniloticus. *Journal Of Fisheries And Aquatic Science*, 9(4), 176-186.
- ArunSudhagar, S. (2016). Diagnostic Tools Used in Fish Disease Diagnosis.
 Aquafind.com. Retrieved 12 March 2016, from http://aquafind.com/articles/FishDiseaseDiagnosis.php
- Avtalion, R. R., Malik, Z., Lefler, E. & Katz, E.(1970). Temperature effect on immune resistance of fish to pathogens.Bamidge Bull. Fish.Cult. Isr. 22, 33–38.
- Batt, C., &Tortorello, M. (2014).*Encyclopedia of food microbiology*. Amsterdam [u.a.]: AP, Academic Press/Elsevier.
- Biller-Takahashi, J., Montassier, H., Takahashi, L., &Urbinati, E. (2014).Proposed method for agglutinating antibody titer analysis and its use as indicator of acquired immunity in pacu, Piaractusmesopotamicus.*Braz. J. Biol.*, 74(1), 238-242.
- Boesen, H., Pedersen, K., Koch, C., & Larsen, J. (1997). Immune response of rainbow trout (Oncorhynchusmykiss) to antigenic preparations from Vibrio anguillarum serogroup O1. Fish & Shellfish Immunology, 7(8), 543-553.
- Evans, J.J., Klesius, P.H., Gilbert, P.M., Shoemaker, C.A., Al Sarawi, M.A., Landsberg, J., Duremdez R., Al Marzouk A. & Al Zenki S. (2002), Characterization of bhemolytic group B *Streptococcus agalactiae* in cultured sea bream, Sparusauratus L., and wild mullet, Liza klunzingeri, in Kuwait. *Journal of Fish Diseases*, 25, 505-513.
- *Fishery Statistics Portal RasmiJabatanPerikanan Malaysia.*(2016). *Dof.gov.my*. Retrieved 12 March 2016, from http://www.dof.gov.my/en/fishery-statistics

- Hossain, M., Haylor, G., &Beveridge, M. (2001).Effect of feeding time and frequency on the growth and feed utilization of African catfish Clariasgariepinus (Burchell 1822) fingerlings.*Aquaculture Research*, 32(12), 999-1004.
- Klesius, P. H., Evans, J.J. & Shoemaker, C.A. (2007). The macrophage chemotactic activity of *Streptococcus agalactiae* and *Streptococcus iniae* Extracellular Products (ECP). Fish Shellfish Immunol., 22: 443-450.
- Kreutz, L., Pavan, T., Alves, A., Correia, A., Barriquel, B., dos Santos, E., &Barcellos, L. (2014). Increased immunoglobulin production in silver catfish (Rhamdiaquelen) exposed to agrichemicals. *Braz. J. Med. Biol. Res.*, 47(6), 499-504.
- Khushiramani, R., Girisha, S.K, &Karunasagar, I. (2007).Cloning and expression of an outer membrane protein ompTS of *Aeromonashydrophila* and study of immunogenicity in fish. Protein Exp Purif.;51(2):303-7.
- Lindahl, G., Stalhammar-Carlemalm, M., &Areschoug, T. (2005).Surface Proteins of Streptococcus agalactiae and Related Proteins in Other Bacterial Pathogens.*Clinical Microbiology Reviews*, 18(1), 102-127.
- Madureira, P., Baptista, M., Vieira, M., Magalhaes, V., Camelo, A., & Oliveira, L. (2007). Streptococcus agalactiae GAPDH Is a Virulence-Associated Immunomodulatory Protein. *The Journal Of Immunology*, *178*(3), 1379-1387.
- Mekuchi, T., Kiyokawa, T., Honda, K., Nakai, T., &Muroga, K. (1995). Vaccination
 Trials in the Japanese Flounder towards Edwardsiellosis. *Fish Pathol.*,30(4), 251-256.
- Navarre, W., &Schneewind, O. (1994).Proteolytic cleavage and cell wall anchoring at the LPXTG motif of surface proteins in Gram-positive bacteria.*Molecular Microbiology*, 14(1), 115-121.

Ouchterlony,Ö.(2009).Antigen-antibodyReactionsinGels.ActaPathologicaMicrobiologicaScandinavica, 32(2), 231-240.

- Pasnik, D., Evans, J., Panangala, V., Klesius, P., Shelby, R., & Shoemaker, C. (2005). Antigenicity of Streptococcus agalactiae extracellular products and vaccine efficacy. *J Fish Diseases*, 28(4), 205-212.
- Pretto-Giordano, L., MÃ¹/₄ller, E., Freitas, J., & Silva, V. (2010).Evaluation on the Pathogenesis of *Streptococcus agalactiae* in Nile Tilapia (*Oreochromisniloticus*).*Brazilian Archives of Biology and Technology*, 53(1), 87-92.
- Rijkers, G. T., Frederix-Wolters, E. M. H. A, &Van Muiswinkel, W. B. (1980).Theimmune system of cyprinid fish.Kinetics and temperature dependence of antibody-producing cells in carp (Cyprinuscarpio).Immunol. 41, 91–97.
- Song, M., J. Xie, X. Peng and H. Li, 2013. Identification of protective immunogens from extracellular secretome of *Edwardsiellatarda*. Fish Shellfish Immunol., 35: 1932-1936.

Zhang, M., H. Wu, X. Li, M. Yang and T. Chen *et al.*, 2012. *Edwardsiellatarda* flagellar protein FlgD: A protective immunogen against edwardsiellosis. Vaccine, 30: 3849-3856.