



EFFECTS OF PROCESSING CONDITIONS ON EXTRACTION OF TOTAL SAPONINS FROM BITTER GOURD (*Momordica charantia* L.) AND ITS ANTIMICROBIAL ACTIVITY AGAINST FOODBORNE PATHOGENS

HANAA ABDELKARIM SALMAN ABOAZRA

FSTM 2016 24



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By

HANAA ABDELKARIM SALMAN ABOAZRA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

November 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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A tropical plant, *Momordica charantia* L., bitter gourd (BG), subcontinent variety, is widely found in Asian countries. The World Health Organization of the United Nations estimates that every year one in three people worldwide get food poisoning leading to food illness or food diseases from consuming contaminated food. Many researches have reported that saponins in BG fruit have biological potency. Therefore, the need to explore a natural source of antimicrobial for food safety and using it as a natural sanitizer was important. The aim of this study is to examine the antimicrobial effect of hot air dried, spray dried, and extrudates of BG crude extracts against selected foodborne pathogens. Optimization of extraction condition using Response Surface Methodology (RSM) was conducted using solvent type, time, and solid to solvent ratio as independent variables while total saponins measured as a dependent response. Different processing methods includes hot air drying at 45°C, spray drying (90, 120, 147.5, 175 and 200°C), and extrusion (80, 100, and 120°C) were applied to produce BG fruit powder. Optimal crude extracts of samples which contain different amount of total saponins were evaluated for antimicrobial activity against eight foodborne pathogens. The extracts that could inhibit the growth of tested pathogens were used in sanitizing test at different exposure times (5, 10, and 15 min). The results of RSM showed that 0.02 g/ml of solid to solvent ratio, time (75 - 120 min), and solvent type (ethanol) was the optimum extraction condition. Hot air dried and extrudate at temperature of 80°C showed a good retention of total saponins with 103.0, 108.8 mg DE/g DM, respectively, compared to initial content of fresh fruit sample 172.7 mg/g DM. BG fruit powder which contain high total saponins had potential bactericidal activity against the tested food pathogens with less than 10 mg/ml in 24 h. Ethanolic extracts of extrudate at temperature of 80°C and hot air dried samples had inhibited the growth of *Klebsiella pneumoniae*, *Staphylococcus epidermidis*, *Bacillus cereus*,

Streptococcus mutans, *Escherichia coli*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa* with a minimum inhibitory concentration (MIC) of 0.625 - 5.625 mg/ml. The sample extracts possessed bactericidal and bacteriostatic activities against foodborne bacteria. Based on time-kill curve, it showed time needed to destroy the bacteria was within 4 h, except for *P. aeruginosa*. In conclusions, different processing method possess different total saponin yields in the dried BG fruit powder could have important bactericidal activity and this study will enhance the attempts of using total saponins compound as a potential natural antibacterial agent in food applications.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KESAN KAEDAH PEMROSESAN PADA PENGEKSTRAKAN JUMLAH SAPONIN DARIPADA PERIA DAN AKTIVITI ANTIMIKROB TERHADAP PATOGEN MAKANAN

Oleh

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Momordica charantia L., buah peria (BG) merupakan tumbuhan tropika yang terdapat secara meluas di negara-negara Asia. Pertubuhan Kesihatan Sedunia Pertubuhan Bangsa-Bangsa Bersatu menganggarkan bahawa setiap tahun masih ada satu dalam tiga orang di seluruh dunia mendapat keracunan makanan dari memakan makanan yang tercemar. Banyak kajian telah melaporkan bahawa saponin dalam buah BG mempunyai potensi biologi. Menyelidik sumber semula jadi antimikrob dari tumbuhan sebagai sanitizer semula jadi adalah perlu. Tujuan kajian ini adalah untuk mengkaji kesan antimikrobial serbuk buah BG yang telah dikeringkan melalui udara panas, semburan kering, dan proses penyemperitan. Pengoptimuman cara pengekstrakan menggunakan Metodologi Response Surface (RSM) telah dijalankan menggunakan jenis pelarut, masa, dan nisbah sampel kepada pelarut sebagai pembolehubah bebas, manakala jumlah saponin adalah respon. Kaedah pemprosesan yang berbeza iaitu pengeringan udara panas pada suhu 45°C, semburan kering (90, 120, 147.5, 175 dan 200°C), dan proses penyemperitan (80, 100, dan 120°C) telah digunakan untuk menghasilkan serbuk buah BG. Ekstrak mentah daripada sampel udara panas, semburan kering dan proses penyemperitan mengandungi kandungan saponin yang berbeza dan kesemua sampel telah dinilai untuk aktiviti antimikrob terhadap lapan patogen makanan. Ekstrak yang boleh menghalang pertumbuhan patogen diuji dengan menggunakan ujian sanitasi pada masa pendedahan yang berlainan (5, 10, dan 15 min). Keputusan RSM menunjukkan bahawa 0.02 g/ml nisbah sampel kepada pelarut, masa (75 - 120 min), dan jenis pelarut (etanol) adalah keadaan optimum pengekstrakan saponin. Sampel daripada pengeringan udara panas dan proses penyemperitan pada suhu 80 °C menunjukkan kandungan saponin 103.0 dan 108.8 mg DE/g DM, masing-masing, berbanding kandungan awal sampel buah BG segar iaitu 172.7 mg/g DM. Serbuk buah

BG yang mengandungi jumlah saponin yang tinggi mempunyai potensi aktiviti bakteria terhadap patogen makanan dengan kurang daripada 10 mg/ml dalam masa 24 jam. Ekstrak etanol daripada serbuk dihasilkan melalui proses penyempitan pada suhu 80°C dan sampel udara panas telah menghalang pertumbuhan *Klebsiella pneumoniae*, *Staphylococcus epidermidis*, *Bacillus cereus*, *Streptococcus mutans*, *Escherichia coli*, *Staphylococcus aureus*, dan *Pseudomonas aeruginosa* dengan kepekatan perencatan minimum (MIC) 0.625 - 5.625 mg/ml. Manakala, masa yang diperlukan untuk memusnahkan bakteria adalah dalam 4 jam, kecuali *P. aeruginosa*. Hasil kajian menunjukkan bahawa ekstrak mentah buah peria memiliki sifat bacteriostatic terhadap bakteria makanan. Pada kesimpulannya, kaedah pemprosesan yang berbeza memberi kesan terhadap jumlah hasil saponin dalam serbuk buah BG dan saponin berpotensi sebagai agen anti-bakteria semulajadi dalam aplikasi makanan.

ACKNOWLEDGEMENTS

Humbles thanks to Almighty Allah. The creator of this universe, who has given knowledge to man to think, finds out and explores the world and his creation. First and foremost, I would like to express my sincere gratitude and appreciation to my supervisory committee, Dr. Rabiha Sulaiman, and Associate Professor Dr. Yaya Rukayadi encouragement and above all, their confidence in me to complete this project. I am very thankful and grateful to Associate Professor Dr. Abdulkarim Sabo Mohammed for comments that greatly improved the discussion section in Chapter 5 and 6. I am also thankful to Professor Dr. Hasanah Mohd. Ghazali and Dr. Rashidah Sukor for their advice. Deep thanks are also extended to Lab assistants from the Engineering Laboratory of Faculty of Food Science and Technology including Encik Amran for his help in my research endeavour. I would like to express my sincere gratitude and appreciation to all Lab assistants from the Biochemistry Laboratory of Faculty of Food Science and Technology, including Encik Azman Asmat for their excellent lab assistance to students. Thanks are also to the staff from the Laboratory of Natural Products, Institute of Bioscience. Special appreciation is also dedicated to my fellow colleagues especially Muhammad Safwan bin Ahamad Bustamam, Nor Husna Yusoff, Lew Kok Fang and Abdel Gani their mutual support during completing this project. Many thanks also to my colleagues at Synthesis and Photochemistry Laboratory in Institute of Bioscience for their help in this study. In addition, thanks are also due to all my colleagues at Food 5 Laboratory especially Mahsa, Shizwan and other post graduate students for their mutual encouragement and exchange of opinions or suggestion in the progress of completing this research. Last but not least, I am very grateful to my family who has always been there for me throughout my study.

I certify that a Thesis Examination Committee has met on 9th November 2016 to conduct the final examination of Hanaa Abdelkarim Salman Aboazra on her thesis entitled "Effects of Processing Conditions on Extraction of Total Saponins from Bitter Gourd (*Momordica Charantia* L.) and its Antimicrobial Activity Against Foodborne Pathogens" in accordance with the Universities and University Colleges Act 1971 and the Constitution of Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

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TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xvi
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Objectives	1
2 LITERATURE REVIEW	3
2.1 Bitter Gourd Plant	3
2.2 Extraction	6
2.3 Saponins	7
2.4 Drying Methods and Extrusion Process	12
2.4.1 Hot Air Drying	12
2.4.2 Spray Drying	13
2.4.3 Extrusion Process	13
2.5 Effect of Drying Methods and Extrusion Process on Plant Phytochemicals	14
2.6 Response Surface Methodology (RSM)	15
2.7 Food Microbial Pathogens	17
2.8 Food Plant's Antimicrobial Agents and its Activity	19
2.9 Food Application of Natural Antimicrobial Compounds	20
3 PROCESS OPTIMIZATION FOR THE EXTRACTION OF TOTAL SAPONINS FROM HOT AIR DRIED BITTER GOURD FRUIT	24
3.1 Introduction	24
3.2 Materials and Methods	24
3.2.1 Materials	24
3.2.2 Sample Preparation	25
3.2.3 Response Surface Methodology (RSM) Experiment	25
3.2.4 Determination of the Response (Total Saponins)	26
3.2.5 Qualitative Evaluation of Saponins	29
3.2.6 Statistical Analysis	31
3.3 Results and Discussion	31
3.3.1 Fitting the Response Surface Model	31
3.3.2 Effect of Independent Variables on Total Saponins	36
3.3.3 The Optimal and Validation of the Predictive Response Model	40

3.3.4	Qualitative Test for Saponins	41
3.4	Conclusion	42
4	EFFECTS OF DRYING METHODS AND EXTRUSION PROCESS ON TOTAL SAPONINS CONTENT AND OTHER PROPERTIES OF BITTER GOURD FRUIT POWDER	43
4.1	Introduction	43
4.2	Experimental	43
4.2.1	Preliminary Study	43
4.2.2	Hot Air Drying Process	44
4.2.3	Extrusion Process	44
4.2.4	Spray Drying Process	46
4.2.5	Powder Analysis	46
4.2.6	Total Saponins in Fresh and Powder Samples	48
4.2.7	Statistical Analysis	49
4.3	Results and Discussion	49
4.3.1	Physicochemical Properties of the BG Powders	49
4.3.2	Effect of Drying and Extrusion Process on the Total Saponins Content	53
4.4	Conclusion	56
5	SCREENING ON ANTIMICROBIAL ACTIVITY OF HOT AIR DRIED, SPRAY DRIED AND EXTRUDATES OF BITTER GOURD FRUIT	57
5.1	Introduction	57
5.2	Experimental	58
5.2.1	Preparation of Bitter Gourd Fruit Extract	58
5.2.2	Microbial Strains Tested	58
5.2.3	Antimicrobial Assays	58
5.3	Results and Discussion	60
5.4	Conclusion	72
6	HOT AIR DRIED AND EXTRUDATES OF BITTER GOURD FRUIT EXTRACTS AS A NATURAL SANITIZER ON EPITHELIAL MICROFLORA IN RAW CHICKEN MEAT	73
6.1	Introduction	73
6.2	Experimental	74
6.2.1	Preparation of Selective Media	74
6.2.2	Extract Preparation	74
6.2.3	Preparation of Bitter Gourd Extract For Treatment	74
6.2.4	Preparation of Chicken Sample and the Treatment	75
6.2.5	Statistical Analysis	75
6.3	Results and Discussion	75
6.4	Conclusion	80
7	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	81
7.1	Summary	81
7.2	Conclusion	81
7.3	Recommendation for Future Research	82

REFERENCES	83
APPENDICES	101
BIODATA OF STUDENT	108
LIST OF PUBLICATIONS	109



LIST OF TABLES

Table		Page
2.1	Biological roles of bitter gour fruit extract	4
2.2	Phytochemical compounds present in bitter gourd fruit ethanol extract	6
2.3	Previous studies on saponins extract with antimicrobial ability	10
2.4	Determination of total saponins in some plants	12
2.5	Antimicrobial activity of bitter gourd fruit extracts	22
3.1	Independent conditions and their levels employed for Box- Behnken design	25
3.2	Box Behnken Design (BBD) model for extraction condition runs	27
3.3	Total saponins in hot air dried bitter gourd fruit powder based on Response Surface Methodology - Box Behnken Design (RSM-BBD)	32
3.4	The regression coefficients and their significance values for the fitted full quadratic model of total saponins	33
3.5	Reducing model of full quadratic model of total saponins	35
3.6	Validation of experimental values under optimized conditions for maximum total saponins	41
3.7	Total saponins in crude (non-defatted) and fraction (defatted) extracts of hot air dried bitter gourd fruit powder	41
3.8	Thin Layer Chromatography (TLC) profile of the extracted saponins fraction and the standard	42
4.1	Preliminary study on bitter gourd fruit varieties (Subcontinent and Chinese phenotype)	50
4.2	Physicochemical properties of the bitter gourd fruit powder	51
4.3	Effect of drying methods and extrusion process on the color of bitter gourd fruit powder	52
4.4	Total saponins in fresh, dried and extrudates of bitter gourd fruit	54

5.1	Antimicrobial assays of hot air dried bitter gourd fruit powder	62
5.2	Disk diffusion assay of spray dried bitter gourd fruit powder	62
5.3	Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of spray dried bitter gourd fruit powde	63
5.4	Antimicrobial assays of extrudates bitter gourd fruit powder	63
5.5	Effective antibacterial parameters of bitter gourd powders	64
5.6	Time-kill data	67
6.1	Yield and total saponins value of ethanolic extract	75
6.2	Effect of hot air dried bitter gourd at 10 mg/ml concentration on TPC, <i>Escherichia coli</i> , <i>Bacillus cereus</i> , and <i>Staphylococcus aureus</i> in chicken meat at different exposure time	77
6.3	Effect of extrudate bitter gourd at 10 mg/ml concentration on TPC, <i>Escherichia coli</i> , <i>Bacillus cereus</i> , and <i>Staphylococcus aureus</i> in chicken meat at different exposure time	77
6.4	Count reduction in chicken meat samples	78

LIST OF FIGURES

Figure		Page
2.1	Bitter gourd varieties	5
2.2	Summary of plant saponins biosynthesis	8
2.3	Structure of the standard diosgenin	11
2.4	Box-Behnken RSM design with three variables	15
2.5	Interaction of some secondary metabolites with cell membranes	19
3.1	Experimental process of bitter gourd for extraction	25
3.2	Normal plot of residual for total saponins yield	32
3.3	Main effects plot for total saponins yield	34
3.4	Scatterplot of total saponins yield	34
3.5	The 3D response surface of full quadratic model for predicted values of total saponins	36
3.6	The 2D response surface of full quadratic model for predicted values of total saponins	37
3.7	Optimization plot for total saponins of RSM-BBD model	38
4.1	A cross-section view of bitter gourd fruit (Subcontinent variety)	42
4.2	Single – screw food extruder and its basic structure	43
5.1	Representative time-kill curve plots for the susceptible bacteria following exposure to bitter gourd extracts	65

LIST OF ABBREVIATIONS

BBD	Box-Behnken Design
BG	Bitter gourd
CFU	Colony forming unit
CLSI	Clinical and Laboratory Standards Institute
D	Diameter
DE	Diosgenin Equivalent
DM	Dry Material
DMSO	Dimethyl sulfoxide
DOE	Design of Experiments
Eq.	Equation
g	Gram
h	Hour
MBC	Minimum Bactericidal Concentration
mg	Milligram
MHA	Mueller Hinton agar
min	Minute
ml	Milliliter
mm	Millimeters
PBS	Phosphate buffered saline
R _f	Retention factor
rpm	Revolutions per minute
RSM	Response Surface Methodology
SEM	Scanning Electron Microscopy
<i>spp.</i>	Species
USDA	US Department of Agriculture
UV	Ultraviolet
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

1.1 Background

Momordica charantia L., bitter gourd (BG) is widely distributed, especially in India, China, Malaysia and other tropical countries. It is often neglected in terms of overall consumption due to its bitter taste. However, BG is among the candidates in recent research for its antidiabetic, anticancer, anti-inflammatory, and anti-triglycerides properties (Harinantenaina *et al.*, 2006; Zulbadli *et al.*, 2011; Joseph and Jini, 2013). Malaysia published a country report on the state of plant genetic resources for food and agriculture (1997-2007) stating that the government encourages the development of value-added processing of 'diversity-rich' products for commercial purposes. Its recommendations include the processing of BG into dietary supplements (Nordin *et al.*, 2007). BG fruit is rich with phytochemical compounds including saponins, which could introduce the fruit to the food industry sector (Reyes-Jáquez *et al.*, 2011). The food market is strongly moving towards products with beneficial nutraceutical values.

BG fruit powders could be used as an ingredient in product formulation and as a source of saponins. Investigation on optimal extraction conditions of total saponins, including factors such as solvent type, extraction time, and solid to solvent ratio of hot air-dried BG powder will provide comprehensive information. For quantification study, this can give an implication of extraction solvent capability in obtaining significant total saponins yields. The finding will be for food and pharmaceutical industries. Additionally, saponins have no noted harmful effect on the kidney or liver at reasonable levels of consumption (Virdi *et al.*, 2003).

Optimized extract that rich in total saponins content from processed bitter gourd fruit may have strong antimicrobial effects against foodborne pathogens, *in vitro*. Use of bitter gourd optimized extract that rich in total saponins content from processed bitter gourd fruit may protect fresh meat from spoilage by reducing its microbial population significantly. This study focus on effect of processing conditions on the extraction of total saponins from BG fruit and determine the antibacterial effect of hot air dried, spray dried, and extrudates of BG powders extracts against *Candida albicans*, *B. cereus*, *E. coli*, *P. aeruginosa*, *S. aureus*, *K. pneumoniae*, *S. mutans* and *S. epidermidis*.

1.2 Objectives

Overall, the research is undertaken with the aim to study effect of processing condition on total saponins yields and examine the antimicrobial effects of dried BG fruit powder. In particular, this study attempts

1. To optimize the extraction conditions for total saponins present in hot air dried BG fruit using response surface methodology (RSM).
2. To determine effect of processing conditions (hot air drying, spray drying, and extrusion process) on physicochemical properties of dried BG fruit powder.
3. To determine the antimicrobial activity of bitter gourd crude extract obtained through RSM model in term of minimum inhibitory concentration (MIC), minimum bactericidal concentration (MBC), and time-kill curve against selected foodborne pathogens.
4. To evaluate antimicrobial activities of bitter gourd crude extract on food application (raw chicken meat) as a natural sanitizer.



REFERENCES

- Abramovitch, R. B. and Martin, G. B. 2004. Strategies used by bacterial pathogens to suppress plant defenses. *Current Opinion in Plant Biology* 7(4): 356-364.
- Acikel, U., Ersan, M. and Acikel, Y. S. 2010. Optimization of critical medium components using response surface methodology for lipase production by *Rhizopus delemar*. *Food and Bioproducts Processing* 88(1): 31-39.
- Adumanya, O. C. U., Obiloma, A. A. and Essien, E. B. 2015. Proximate, vitamins and mineral composition of *Salacia senegalensis* Lam (DC) leaves. *The American Journal of Innovative Research and Applied Sciences* 1(2): 44-52.
- Agrawal, R., Upadhyay, A. and Nayak, P. S. 2013. Drying characteristics of Safed Musli (*Chlorophytum borivilianum*) and its effect on colour and saponin content. *Journal of Pharmacognosy and Phytotherapy* 5(8): 142-147.
- Amid, M., Tan, C. P., Mirhosseini, H., Aziz, N. A. and Ling, T. C. 2011. Optimisation of serine protease extraction from mango peel (*Mangifera indica* Cv. Chokanan). *Food Chemistry* 124(2): 666-671.
- Amira, K., Aminah, A. and Zuhair, A. 2013. Evaluation of bitter melon (*Momordica charantia*) extract administration in the antioxidant and free radical scavenging activities of plasma and liver in male rat. *International Food Research Journal* 20(1): 319-323.
- Anderson, R. A., Conway, H. F., Pfeifer, V. F. and Griffin, E. L. 1969. Roll and extrusion-cooking of grain sorghum grits. *Cereal Science Today* 14(11): 372-376.
- Anilakumar, K. R., Kumar, G. P. and Ilaiyaraja, N. 2015. Nutritional, pharmacological and medicinal properties of *Momordica charantia*. *International Journal of Nutrition and Food Sciences* 4(1): 75-83.
- Anuonye, J. C., Jigam, A. A. and Ndaceko, G. M. 2012. Effects of extrusion-cooking on the nutrient and anti-nutrient composition of pigeon pea and unripe plantain blends. *Journal of Applied Pharmaceutical Science* 2(5): 158-162.
- Anuonye, J. C., Onuh, J. O., Egwim, E. and Adeyemo, S. O. 2010. Nutrient and antinutrient composition of extruded acha/soybean blends. *Journal of Food Processing and Preservation* 34: 680-691.
- AOAC International. Association of Official Analytical Chemists (AOAC). 2006. Official methods of determination the moisture content. The AOAC Official Method 967.03. Washington: AOAC International.

- Applewhite, T. H. 1989. Proceedings of the World Congress on Vegetable Protein Utilization in Human Foods and Animal Feedstuffs. Champaign: The American Oil Chemists Society.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., et al. 2013. Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering* 117: 426-436.
- Babatunde, O. A. and Adewumi, A. O. 2015. Effects of ethanolic extract of garlic, roselle and ginger on quality attributes of chicken patties. *African Journal of Biotechnology* 14(8): 688-694.
- Bafina, P. and Manimehalai, N. 2014. Kokum fruit bar development via response surface methodology (RSM). *Journal of Engineering Research and Applications* 4(2): 223-230.
- Basch, E., Gabardi, S. and Ulbricht, C. 2003. Bitter melon (*Momordica charantia*): a review of efficacy and safety. *American Journal of Health System Pharmacy* 60(4): 356-359.
- Berk, Z. 2009. *Food Process Engineering and Technology*. Burlington: Academic Press.
- Bettaieb, A., Moujahed, N. and Ksouri, R. 2012. Secondary compounds characterization in some *Autochtonous* species from a North-Eastern region of Tunisia. In: Acar, Z., López-Fran cos, A., Porqueddu, C. (Eds). *New Approaches for Grassland Research in A Context of Climate and Socioeconomic Changes*, p. 371-374. Paris: Centre International de Hautes Etudes Agronomiques Méditerranéennes.
- Bhandari, B., Bansal, N., Zhang, M. and Schuck, P. 2013. *Handbook of Food Powders: Processes and Properties*. Cambridge: Woodhead Publishing.
- Bhunia, A. 2007. *Foodborne Microbial Pathogens: Mechanisms and Pathogenesis*. New York: Springer Science and Business Media.
- Bradley, R. 2010. Moisture and total solids analysis. Nielsen, S. (Ed). *Food Analysis*, p. 87-104. Springer Science and Business Media.
- Brennan, C., Brennan, M., Derbyshire, E. and Tiwari, B. K. 2011. Effects of extrusion on the polyphenols, vitamins and antioxidant activity of foods. *Trends in Food Science and Technology* 22(10): 570-575.
- Bukar, A., Uba, A. and Oyeyi, T. 2010. Antimicrobial profile of *Moringa oleifera* Lam. extracts against some food-borne microorganisms. *Bayero Journal of Pure and Applied Sciences* 3(1): 43-48.

- Caliskan, G. and Dirim, S. N. 2013. The effects of the different drying conditions and the amounts of maltodextrin addition during spray drying of sumac extract. *Food and Bioproducts Processing* 91(4): 539-548.
- Carr, B. T., Jaeger, S. R. and MacFie, H. 2010. Statistical design of experiments in the 21st century and implications for consumer product testing. In Jaeger, S. R. and MacFie, H. (Eds). *Consumer-Driven Innovation in Food and Personal Care Products*, p. 427-469. Cambridge: Woodhead Publishing.
- Cervantes-Martínez, C. V., Medina-Torres, L., González-Laredo, R. F., Calderas, F., Sánchez-Olivares, G., Herrera-Valencia, E. E. and Rodríguez-Ramírez, J. 2014. Study of spray drying of the Aloe vera mucilage (*Aloe vera barbadensis* Miller) as a function of its rheological properties. *LWT-Food Science and Technology* 55(2): 426-435.
- Chaturvedi, S., Hemamalini, R. and Khare, S. K. 2012. Effect of processing conditions on saponin content and antioxidant activity of Indian varieties of soybean (*Glycine max* Linn.). *Annals of Phytomedicine* 1(1): 62-68.
- Cheesbrough, M. 2006. *District Laboratory Practice in Tropical Countries*. Norfolk: Cambridge University Press.
- Chen, J., Tian, R., Qiu, M., Lu, L., Zheng, Y. and Zhang, Z. 2008. Trinorcucurbitane and cucurbitane triterpenoids from the roots of *Momordica charantia*. *Phytochemistry* 69: 1043-1048.
- Chen, J., Zhou, Y., Fang, S., Meng, Y., Kang, X., Xu, X. and Zuo, X. 2013. Mathematical modeling of hot air drying kinetics of *Momordica charantia* slices and its color change. *Advance Journal of Food Science and Technology* 5(9): 1214-1219.
- Chen, Y., Xie, M. Y. and Gong, X. F. 2007. Microwave-assisted extraction used for the isolation of total triterpenoid saponins from *Ganoderma atrum*. *Journal of Food Engineering* 81(1): 162-170.
- Cheok, C. Y. 2012. Extraction of total phenolic and total monomeric anthocyanin contents from mangosteen (*Garcinia mangostana* L.) hull. Serdang, Malaysia: Universiti Putra Malaysia, PhD thesis.
- Cheok, C. Y., Salman, H. A. K. and Sulaiman, R. 2014. Extraction and quantification of saponins: a review. *Food Research International* 59: 16-40.
- Chia, Y. Y. and Yap, W. 2011. *In vitro* antimicrobial activity of hexane: petroleum ether extracts from fruits of *Momordica charantia* L. *International Journal of Pharmaceutical and Biological Archive* 2(3): 868-873.

- Clinical and Laboratory Standards Institute (CLSI). 1999. Methods for determining bactericidal activity of antimicrobial agents. Approved standard M26-A. Wayne: National Committee for Clinical and Laboratory Standards.
- Clinical Laboratory Standards Institute (CLSI). 2003. Reference method for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved standard M7-A6. Wayne: National Committee for Clinical and Laboratory Standards.
- Conde, E., Moure, A., Domínguez, H. and Parajó, J. C. 2010. Extraction of natural antioxidants from plant foods. In: Rizvi, S. (Ed). Separation, Extraction and Concentration Processes in the Food, Beverage and Nutraceutical Industries, p. 506–594. Cambridge: Woodhead Publishing.
- Costa, J. G. M., Nascimento, E. M., Campos, A. R. and Rodrigues, F. F. 2010. Antibacterial activity of *Momordica charantia* (Curcubitaceae) extracts and fractions. *Journal of Basic and Clinical Pharmacy* 2(1): 45-51.
- Daboor, S. M., Masood, F. S. S., Al-Azab, M. S. and Nori, E. E. 2015. A review on *Streptococcus mutans* with its diseases dental caries, dental plaque and endocarditis. *Indian Journal of Microbiology Research* 2(2): 76-82.
- Daniel, P., Supe, U. and Roymon, M. G. 2014. A review on phytochemical analysis of *Momordica charantia*. *International Journal of Advances in Pharmacy, Biology and Chemistry* 3(1): 214-220.
- Dillon, V. M. and Board, R. G. 1994. Natural Antimicrobial Systems and Food Preservation. Wallingford: CAB International.
- D'Mello, J. F., Duffus, C. M. and Duffus, J. H. 1991. Toxic Substances in Crop Plants. Cambridge: The Royal Society of Chemistry.
- Dogan, H. and Karwe, M. V. 2003. Physicochemical properties of quinoa extrudates. *Food Science and Technology International* 9(2): 101-114.
- Du, X. W., Wills, R. B. H. and Stuart, D. L. 2004. Changes in neutral and malonyl ginsenosides in American ginseng (*Panax quinquefolium*) during drying, storage and ethanolic extraction. *Food Chemistry* 86(2): 155-159.
- Duangmal, K., Saicheua, B. and Sueprasarn, S. 2008. Colour evaluation of freeze-dried roselle extract as a natural food colorant in a model system of a drink. *LWT-Food Science and Technology* 41(8): 1437-1445.
- El-Said, S. M. and Al-Barak, A.S. 2011. Extraction of insulin like compounds from bitter melon plants. *American Journal of Drug Discovery and Development* 1: 1-7.

- Eskilsson, C. S. and Björklund, E. 2000. Analytical-scale microwave-assisted extraction. *Journal of Chromatography* 902(1): 227-250.
- Fennema, O. R. and Reid, D. S. 2008. Water and ice. In Fennema, O. R., Damodaran, S. and Parkin, K. L. (Eds). *Fennema's Food Chemistry*, p. 17-82. Boca Raton: CRC Press.
- Fernandes, N. P., Lagishetty, C. V., Panda, V. S. and Naik, S. R. 2007. An experimental evaluation of the antidiabetic and antilipidemic properties of a standardized *Momordica charantia* fruit extract. *BioMed Central Complementary and Alternative Medicine* 7(1): 29-36.
- Fluck, H. 1963. Intrinsic and extrinsic factors affecting the production of secondary plant products. In: Swain T. (Ed). *Chemical Plant Taxonomy*, p. 167-186. London: Academic Press.
- Fuchs, H., Bachran, D., Panjideh, H., Schellmann, N., Weng, A., Melzig, M. F. and Bachran, C. 2009. Saponins as tool for improved targeted tumor therapies. *Current Drug Targets* 10(2): 140-151.
- Gertenbach D. 2002. Solid-liquid extraction technologies for manufacturing nutraceuticals. In Shi J. Mazza G. and Maguer ML. (Eds). *Functional Foods: Biochemical and Processing Aspects*, p. 332-365. Boca Raton: CRC Press.
- Ghasemzadeh, A., Jaafar, H. Z. and Rahmat, A. 2015. Phytochemical constituents and biological activities of different extracts of *Strobilanthes crispus* (L.) Bremek leaves grown in different locations of Malaysia. *BioMed Central Complementary and Alternative Medicine* 15(1): 422-432.
- Gil, M. I., Selma, M. V., López-Gálvez, F. and Allende, A. 2009. Fresh-cut product sanitation and wash water disinfection: problems and solutions. *International Journal of Food Microbiology* 134(1): 37-45.
- Gómez-López, V. M. 2012. *Decontamination of Fresh and Minimally Processed Produce*. Oxford: Wiley-Blackwell.
- Grabowski, J. A., Truong, V. D. and Daubert, C. R. 2006. Spray-drying of amylase hydrolyzed sweetpotato puree and physicochemical properties of powder. *Journal of Food Science* 71(5): 209-217.
- Granato, D. and de Araújo Calado, V. M. 2014. The use and importance of design of experiments (DOE) in process modelling in food science and technology. In Granato, D. and Ares, G. (Eds). *Mathematical and Statistical Methods in Food Science and Technology*, p. 1-18. Hoboken: John Wiley and Sons.
- Grover, J. K. and Yadav, S. P. 2004. Pharmacological actions and potential uses of *Momordica charantia*: A review. *Journal of Ethnopharmacology* 93: 123-132.

- Guclu-Ustundag, O. and Mazza, G. 2007. Saponins: properties, applications and processing. *Critical Reviews in Food Science and Nutrition* 47(3), 231-258.
- Guha, M. 2000. Processing and quality of rice-based extruded products. Calcutta, India: Jadavpur University, PhD thesis.
- Gui, Y. and Ryu, G. H. 2013. The effect of extrusion conditions on the acidic polysaccharide, ginsenoside contents and antioxidant properties of extruded Korean red ginseng. *Journal of Ginseng Research* 37(2): 219-226.
- Ha, Y. W., Lim, S. S., Ha, I. J., Na, Y. C., Seo, J. J., Shin, H. and Kim, Y. S. 2007. Preparative isolation of four ginsenosides from Korean red ginseng (steam-treated *Panax ginseng* CA Meyer), by high-speed counter-current chromatography coupled with evaporative light scattering detection. *Journal of Chromatography* 1151(1): 37-44.
- Han, C., Hui, Q. and Wang, Y. 2008. Hypoglycaemic activity of saponin fraction extracted from *Momordica charantia* in PEG/salt aqueous two-phase systems. *Natural Product Research* 22(13): 1112-1119.
- Haralampidis, K., Trojanowska, M. and Osbourn, A. E. 2002. Biosynthesis of triterpenoid saponins in plants. In Dutta, NN. Hammar, F. Haralampidis, D. Karanth, NG. König, A. Krishna, SH. Kunze, G. Nagy, E. Orlich, R. Osbourn, AE. Raghavarao, KS. History and Trends in Bioprocessing and Biotransformation, p. 31-49. Berlin: Springer-Verlag Berlin Heidelberg.
- Harinantenaina, L., Tanaka, M., Takaoka, S., Oda, M., Mogami, O., Uchida, M. and Asakawa, Y. 2006. *Momordica charantia* constituents and antidiabetic screening of the isolated major compounds. *Chemical and Pharmaceutical Bulletin* 54(7): 1017-1021.
- Hecer, C. and Guldaz, M. 2011. Effects of lactic acid, fumaric acid and chlorine dioxide on shelf-life of broiler wings during storage. *African Journal of Microbiology Research* 23: 3880-3883.
- Heng, M. Y., Tan, S. N., Yong, J.W. H. and Ong, E. S. 2013. Emerging green technologies for the chemical standardization of botanicals and herbal preparations. *Trends in Analytical Chemistry* 50: 1-10.
- Hiai, S., Oura, H. and Nakajima, T. 1976. Color reaction of some saponins and saponins with vanillin and sulphuric acid. *Planta Medica* 29(02): 116-122.
- Horax, R., Hettiarachchy, N., Kannan, A. and Chen, P. 2010. Proximate composition and amino acid and mineral contents of *Momordica charantia*. L. pericarp and seeds at different maturity stages. *Food Chemistry* 122: 1111-1115.

- Ibrahim, M. A., Koorbanally, N. A. and Islam, M. 2014. Antioxidative activity and inhibition of key enzymes linked to type-2 diabetes (α -glucosidase and α -amylase) by *Khaya senegalensis*. *Acta Pharmaceutica* 64(3): 311-324.
- Irondi, A. E., Anokam, K. K. and Ndidi, U. S. 2013. Effect of drying methods on the phytochemical composition and antioxidant activities of *Carica papaya* seed. *International Journal of Biosciences* 3(11): 154-163.
- Islam, S., Jalaluddin, M. and Hettiarachchy, N. 2011. Bio-active compounds of bitter melon genotypes (*Momordica charantia* L.) in relation to their physiological functions. *Functional Foods in Heals and Disease* 2: 61-74.
- Iwe, M. O., Van Zuilichem, D. J. and Ngoddy, P. O. 2000. Color of single-screw extruded blends of soy-sweet potato flour—A response surface analysis. *Plant Foods for Human Nutrition* 55(2): 159-168.
- Iwe, M. O., Van Zuilichem, D. J., Stolp, W. and Ngoddy, P. O. 2004. Effect of extrusion cooking of soy-sweet potato mixtures on available lysine content and browning index of extrudates. *Journal of Food Engineering* 62(2): 143-150.
- Jabeen, U. and Khanum, A. 2014. Isolation and characterization of potential food peptide from *Momordica charantia* L. *Arabian Journal of Chemistry* [In Press] <http://dx.doi.org/10.1016/j.arabjc.2014.06.009>.
- Jagessar, R. C., Mohamed, A. and Gomes, G. 2008. An evaluation of the Antibacterial and Antifungal activity of leaf extracts of *Momordica Charantia* against *Candida albicans*, *Staphylococcus aureus* and *Escherichia coli*. *Nature and Science* 6(1): 1-14.
- Joseph, B. and Jini, D. 2013. Antidiabetic effects of *Momordica charantia* (bitter melon) and its medicinal potency. *Asian Pacific Journal of Tropical Disease* 3(2): 93-102.
- Jouda, M. M. 2013. Antibacterial effect of some medicinal plant extracts and their synergistic effect with antibiotic and non-antibiotic drugs. Gaza, Palestine: Islamic University, MSc thesis.
- Kai, T., Jie, Y. and Xin, H. 2014. A technological research on synergetic extraction of *Momordica charantia* L. saponin using microwave-surfactant. *Applied Mechanics and Materials* 563: 384-390.
- Kalaivani, T. 2013. Antimicrobial property of potent medicinal plant *Acacia nilotica* L. wild. *International Journal Pharmacy and Pharmaceutical Sciences* 5(2): 467-470.

- Karimi, E., Jaafar, H. Z. and Ahmad, S. 2011. Phytochemical analysis and antimicrobial activities of methanolic extracts of leaf, stem and root from different varieties of *Labisa pumila* Benth. *Molecules* 16(6): 4438-4450.
- Khan, M. M. H. and Chaudhry, A. S. 2011. A comparative study of low and high quality forages for chemical composition and *in vitro* degradability. *The Journal of Animal and Plant Sciences* 21: 715-723.
- Kim J. 2008. Protective effects of Asian dietary items on cancers - soy and ginseng. *Asian Pacific Journal of Cancer Prevention* 9: 543-548
- Kim, Y. and Wampler, D. J. 2009. Determination of saponin and various chemical compounds in *Camellia sinensis* and genus *Ilex*. Technical Note (SEN-TN-0027). Hamilton: Sensus.
- Kobori, M., Ohnishi-kameyama, M., Akimoto, Y., Yukizaki, C. and Yoshida, M. 2008. α -Eleostearic acid and its dihydroxy derivative are major apoptosis-inducing components of bitter melon. *Journal of Agricultural and Food Chemistry* 56: 10515-10520.
- Kothakota, A., Jindal, N. and Thimmaiah, B. 2013. A study on evaluation and characterization of extruded product by using various by-products. *African Journal of Food Science* 7(12): 485-497.
- Kozačinski, L., Cvrtila Fleck, Ž. Kozačinski, Z., Filipović, I., Mitak, M., Bratulić, M. and Mikuš, T. 2012. Evaluation of shelf life of pre-packed cut poultry meat. *Veterinarski Arhiv* 82: 47-58.
- Kreis, W. and Müller-Uri, F. 2010. Biochemistry of sterols, cardiac glycosides, brassinosteroids, phytoecdysteroids and steroid saponins. In Wink, M. (Ed). *Annual Plant Reviews Volume 40: Biochemistry of Plant Secondary Metabolism*, p. 304-363. Oxford: Wiley-Blackwell.
- Krishnamurthy, P., Tsukamoto, C., Yang, S. H., Lee, J. D. and Chung, G. 2012. An improved method to resolve plant saponins and sugars by TLC. *Chromatographia* 75: 1445-1449.
- Kumar, D. S., Sharathnath, K. V., Yogeswaran, P., Harani, A., Sudhakar, K., Sudha, P. and Banji, D. 2010. A medicinal potency of *Momordica charantia*. *International Journal of Pharmaceutical Sciences Review and Research* 1(2): 95-100.
- Kumar, R., Balaji, S., Uma, T. S. and Sehgal, P. K. 2009. Fruit extracts of *Momordica charantia* potentiate glucose uptake and up-regulate Glut-4, PPAR γ and PI3K. *Journal of Ethnopharmacology* 126(3): 533-537.
- Kwatra, D., Venugopal, A., Standing, D., Ponnurangam, S., Dhar, A., Mitra, A. and Anant, S. 2013. Bitter melon extracts enhance the activity of chemotherapeutic

- agents through the modulation of multiple drug resistance. *Journal of Pharmaceutical Sciences* 102(12): 4444-4454.
- Kwon, J. H., Bélanger, J. M. and Paré, J. J. 2003. Optimization of microwave-assisted extraction (MAP) for ginseng components by response surface methodology. *Journal of Agricultural and Food Chemistry* 51(7): 1807-1810.
- Kyoto Encyclopedia of Genes and Genomes (KEGG). (2015). Retrieved on August 24, 2015 from Genome Website: www.genome.jp/kegg.
- Lawson, J. 2010. *Design and Analysis of Experiments with SAS*. Boca Raton: CRC Press.
- Leelaprakash, G., Rose, J. C., Gowtham, B. M., Javvaji, P. K. and Prasad, S. A. 2011. In vitro antimicrobial and antioxidant activity of *Momordica charantia* leaves. *Pharmacophore* 2(4) 244-252.
- Li Ma, 1998. *Engineering Properties of Foods and Other Biological Materials*. Washington: ASAE.
- Li, W., Lin, Z., Yang, C., Wang, Y. and Qiao, Y. 2015. Study on the chemical constituents of *Momordica charantia* L. leaves and method for their quantitative determination. *Biomedical Research* 26(3): 415-419.
- Lu, Y. L., Liu, Y. H., Liang, W. L., Chyuan, J. H., Cheng, K. T., Liang, H. J. and Hou, W. C. 2011. Antibacterial and cytotoxic activities of different wild bitter gourd cultivars (*Momordica charantia* L. var. *abbreviata* seringe). *Botanical Studies Journal* 52: 427-434.
- Lucas, E. A., Dumancas, G. G., Smith, B. J., Clarke, S. L. and Arjmandi, B. H. 2010. Health benefits of bitter melon (*Momordica charantia*). In Watson, R. R. and Preedy, V. R. (Eds). *Bioactive Foods in Promoting Health: Fruits and Vegetables*, p. 525-549. London: Academic Press.
- Lucera, A., Costa, C., Conte, A., and Del Nobile, M. A. 2012. Food applications of natural antimicrobial compounds. *Frontiers in Microbiology, Antimicrobial, Resistance and Chemotherapy* 3: 1-13.
- Maatalah, M. B., Bouzidi, N. K., Bellahouel, S., Merah, B., Fortas, Z., Soulimani, R., Saidi, S. and Derdour, A. 2012. Antimicrobial activity of the alkaloids and saponin extracts of *Anabasis articulata*. *Journal of Biotechnology and Pharmaceutical Research* 3(3): 54-57.
- Majinda, R. R. T. 2012. Extraction and isolation of saponins. In Sarker, S. D. and Nahar, L. (Eds). *Natural Products Isolation*, p. 415-426. Totowa: Humana Press.

- Makhija, M., Ahuja, D., Nancy, B. C., Gauttan, S., Tiwari, K., Awasthi, A. and Sxena, P. 2011. Evaluation and comparison of antibacterial activities of leaf, seeds, and fruits of extracts of *Momordica charantia*. Research Journal of Pharmaceutical, BioLogical and Chemical Sciences 2: 185-192.
- Makkar, H. P. S., Siddhuraju, P. and Becker, K. 2007. Plant Secondary Metabolites. Totowa: Humana Press.
- Marston, A., Wolfender, J. L. and Hostettmann, K. 2000. Analysis and isolation of saponins from plant material. In Oleszek and Marston (Eds). Saponins in Food, Feedstuffs and Medicinal Plants, p. 1-12. Dordrecht: Kluwer Academic.
- Mead, G. 2004. Poultry Meat Processing and Quality. Cambridge: Woodhead Publishing.
- Mirhosseini, H., Tan, C. P., Hamid, N. S. and Yusof, S. 2008. Effect of Arabic gum, xanthan gum and orange oil on flavor release from diluted orange beverage emulsion. Food Chemistry 107(3): 1161-1172.
- Mishra, P., Mishra, S. and Mahanta, C. L. 2014. Effect of maltodextrin concentration and inlet temperature during spray drying on physicochemical and antioxidant properties of amla (*Emblica officinalis*) juice powder. Food and Bioproducts Processing 92(3): 252-258.
- Mith, H., Dure, R., Delcenserie, V., Zhiri, A., Daube, G. and Clinquart, A. 2014. Antimicrobial activities of commercial essential oils and their components against food-borne pathogens and food spoilage bacteria. Food Science and Nutrition 2(4): 403-416.
- Mohammed, M. 2010. Postharvest handling and quality management of bitter gourds (*Momordica charantia* L.). St. Augustine: The University of the West Indies.
- Montgomery, D. C. 2009. Design and Analysis of Experiments. Hoboken: John Wiley and Sons.
- Moscicki, L. and van Zuilichem, D. J. 2011. Extrusion-cooking and related technique. In Moscicki, L. (Ed). Extrusion-Cooking Techniques: Applications, Theory and Sustainability, p. 1-23. Weinheim: Wiley-VCHA.
- Mugford, S. T. and Osbourn, A. 2013. Saponin synthesis and function. In Bach, T. J. and Rohmer, M. (Eds). Isoprenoid Synthesis in Plants and Microorganisms: New Concepts and Experimental Approaches, p. 405-424. New York: Springer New York.
- Müller, A., Ganzera, M. and Stuppner, H. 2006. Analysis of phenolic glycosides and saponins in *Primula elatior* and *Primula veris* (primula root) by liquid chromatography, evaporative light scattering detection and mass spectrometry. Journal of Chromatography A 1112(1): 218-223.

- Mwambete, K. D. 2009. The *in vitro* antimicrobial activity of fruit and leaf crude extracts of *Momordica charantia*: A Tanzania medicinal plant. *African Health Sciences* 9(1), 34-39.
- Ngan, C. L., Basri, M., Lye, F. F., Masoumi, H. R. F., Tripathy, M., Karjiban, R. A. and Abdul-Malek, E. 2014. Comparison of Box–Behnken and central composite designs in optimization of fullerene loaded palm-based nano-emulsions for cosmeceutical application. *Industrial Crops and Products* 59: 309-317.
- Nordin, M. S., Ariffin, Z., Jajuli, R., Abdullah, W. D. W. and Denis, M. G. 2007. Plant genetic resources for food and agriculture in Malaysia (1997–2007). Report of MOA-ABI and MARDI on the Conservation and Sustainable Utilization of (PGRFA). Malaysia: The Ministry of Agriculture and Agro-based Industries (MOA-ABI), as the National Focal Point, and the Malaysian Agricultural Research and Development Institute (MARDI).
- Oakenfull, D. 1981. Saponins in food - a review. *Food Chemistry* 7(1): 19-40.
- Oenning, G., Juillerat, M. A., Fay, L. and Asp, N. G. 1994. Degradation of oat saponins during heat processing-effect of pH, stainless steel, and iron at different temperatures. *Journal of Agricultural and Food Chemistry* 42(11): 2578-2582.
- Oleszek, W. A. 2000. Saponins. In Naidu, A. S. (Ed). *Natural Food Antimicrobial Systems*, p. 295-324. Boca Raton: CRC Press.
- Oleszek, W. and Bialy, Z. 2006. Chromatographic determination of plant saponins - an update (2002–2005). *Journal Chromatography* 1112: 78–91.
- Oliveira, W. P., Souza, C. R. F., Kurozawa, L. E. and Park, K. J. 2010. Spray drying of food and herbal products. In Woo, M. W., Mujumdar, A. S. and Daud, W. R. W. (Eds). *Spray Drying Technology*, p. 113-156. Singapore: Free e-book Publication [ISBN: 978-981-08-6270-1].
- Oussalah, M., Caillet, S., Saucier, L. and Lacroix, M. 2007. Inhibitory effects of selected plant essential oils on the growth of four pathogenic bacteria: *E. coli* O157: H7, *Salmonella typhimurium*, *Staphylococcus aureus* and *Listeria monocytogenes*. *Food Control* 18(5): 414-420.
- Ozusaglam, M. A. and Karakoca, K. 2013. Antimicrobial and antioxidant activities of *Momordica charantia* from Turkey. *African Journal of Biotechnology* 12(13): 1548-1558.
- Pankey, G. A. and Sabath, L. D. 2004. Clinical relevance of bacteriostatic versus bactericidal mechanisms of action in the treatment of Gram-positive bacterial infections. *Clinical Infectious Diseases* 38(6): 864-870.

- Parhusip, A. J. N. and Sitanggang, A. B. 2011. Antimicrobial Activity of Melinjo Seed and Peel Extract (*Gnetum gnemon*) Against Selected Pathogenic Bacteria. *Microbiology Indonesia* 5(3): 103-112.
- Paris, R. 1963. The distribution of plant glycosides. In Swain, T. (Ed). *Chemical Plant Taxonomy*, p. 337-358. London: Academic Press.
- Pasaribu, T., Astuti, D. A., Wina, E., Sumiati and Setiyono, A. 2014. Saponin content of *sapindus rarak* pericarp affected by particle size and type of solvent, its biological activity on *Eimeria tenella* oocysts. *International Journal of Poultry Science* 13(6): 347-352.
- Passos M. L. and Birchall V. S. 2010. Manipulating physical properties of powders. In Woo, M. W., Mujumdar, A. S. and Daud, W. R. W. (Eds). *Spray Drying Technology*, p. 37-60. Singapore: Free e-book Publication [ISBN: 978-981-08-6270-1].
- Patel, S., Patel, T., Parmar, K., Bhatt, Y., Patel, Y. and Patel, N. M. 2010. Isolation, characterization and antimicrobial activity of charantin from *Momordica charantia* linn. Fruit. *International Journal of Drug Development and Research* 2(3): 629-634.
- Pathare, P. B., Opara, U. L. and Al-Said, F. A. J. 2013. Colour measurement and analysis in fresh and processed foods: a review. *Food and Bioprocess Technology* 6(1): 36-60.
- Petrová, J. 2013. Microbiological quality of fresh chicken breast meat after rosemary essential oil treatment and vacuum packaging. *Scientific Papers Animal Science and Biotechnologies* 46: 140-144.
- Pham, H. N. T., Nguyen, V. T., Vuong, Q. V., Bowyer, M. C. and Scarlett, C. J. 2015. Effect of extraction solvents and drying methods on the physicochemical and antioxidant properties of *Helicteres hirsuta* Lour. Leaves. *Technologies* 3(4): 285-301.
- Phisut, N. 2012. Spray drying technique of fruit juice powder: some factors influencing the properties of product. *International Food Research Journal* 19(4): 1297-1306.
- Premi, M. and Sharma, H. K. 2013. Oil extraction optimization and kinetics from *Moringa oleifera* (PKM 1) seeds. *International Journal of Agriculture and Food Science Technology* 4(4): 371-378.
- Puspanadan, S., Loo, Y. Y., Nillian, E., Kuan, C. H., Goh, S. G., Chang, W. S., Lye, Y. L., Afsah-Hejri, L., Tang, J. Y. H., Rukayadi, Y. and Nakaguchi, Y. 2012. Detection of *Klebsiella pneumoniae* in raw vegetables using most probable number-polymerase chain reaction (MPN-PCR). *International Food Research Journal* 19(4): 1757-1762.

- Rajamoorthi, A., Shrivastava, S., Steele, R., Nerurkar, P., Gonzalez, J. G., Crawford, S., Mark, V. and Ray, R. B. 2013. Bitter melon reduces head and neck squamous cell carcinoma growth by targeting c-Met signaling. *PLoS One* 8(10): e78006.
- Rakholiya, K., Vaghela, P., Rathod, T. and Chanda, S. 2014. Comparative study of hydroalcoholic extracts of *Momordica charantia* L. against foodborne pathogens. *Indian Journal of Pharmaceutical Sciences* 76(2): 148-156.
- Rao, A. V. and Gurfinkel, D. M. 2000. Dietary saponins and human health. In Oleszek, W. and Marston, A. (Eds). *Saponins in Food, Feedstuffs and Medicinal Plants*, p. 1-12. Dordrecht: Kluwer Academic.
- Ren, Y., Chen, Y., Hu, B., Wu, H., Lai, F. and Li, X. 2015. Microwave-assisted extraction and a new determination method for total steroid saponins from *Dioscorea zingiberensis* CH Wright. *Steroids*, 104: 145-152.
- Reyes-Jáquez, D., Vargas-Rodríguez, J., Delgado-Licon, E., Rodríguez-Miranda, J., Araiza-Rosales, E. E., Andrade-González, I. and Medrano-Roldan, H. 2011. Optimization of the extrusion process temperature and moisture content on the functional properties and *In vitro* digestibility of bovine cattle feed made out of waste bean flour. *Journal of Animal Science Advances* 1(2): 100-110.
- Riaz, M. N. 2000. *Extruders in Food Applications*. Lancaster: Technomic.
- Rickert, D. A., Meyer, M. A., Hu, J. and Murphy, P. A. 2004. Effect of extraction pH and temperature on isoflavone and saponin partitioning and profile during soy protein isolate production. *Journal of Food Science* 69(8): 623-631.
- Rodel, W. 2001. Water activity and its measurement in food. In Kress-Rogers, E. and Brimelow, C. J. (Eds). *Instrumentation and Sensors for the Food Industry*, p. 453-474. Cambridge: Woodhead Publishing and CRC Press.
- Romagnì, J. G. 2009. Biosynthesis of chemical signals-de novo synthesis and secondary metabolites. *Chemical Ecology* 1: 393-401.
- Roongruangsri, W. and Bronlund, J. E. 2016. Effect of air-drying temperature on physico-chemical, powder properties and sorption characteristics of pumpkin powders. *International Food Research Journal* 23(3): 962-972.
- Ruiz-Gutiérrez, M. G., Amaya-Guerra, C. A., Quintero-Ramos, A., Pérez-Carrillo, E., Ruiz-Anchondo, T. D. J., Báez-González, J. G. and Meléndez-Pizarro, C. O. 2015. Effect of extrusion cooking on bioactive compounds in encapsulated red cactus pear powder. *Molecules* 20(5): 8875-8892.
- Rukayadi, Y. and Hwang, J. K. 2006. *In vitro* activity of xanthorrhizol against *Streptococcus mutans* biofilms. *Letter in Applied Microbiology* 42: 400-404.

- Rukayadi, Y., Han, S., Yong, D. and Hwang, J. K. 2010. *In vitro* antibacterial activity of panduratin A against enterococci clinical isolates. *Biological and Pharmaceutical Bulletin* 33: 1489–1493.
- Rukayadi, Y., Lee, K., Lee, M., Yong, D. and Hwang, J. K. 2009. Synergistic anticandidal activity of xanthorrhizol in combination with ketoconazole or amphotericin B. *FEMS Yeast Research* 9: 1302-1311.
- Saeed, S. and Tariq, P. 2005. Antibacterial activities of *Mentha piperita*, *Pisum sativum* and *Momordica charantia*. *Pakistan Journal of Botany* 37(4): 997-1001.
- Sagdic, O. 2003. Sensitivity of four pathogenic bacteria to Turkish thyme and oregano hydrosols. *LWT-Food Science and Technology* 36(5): 467-473.
- Santhalakshmy, S., Bosco, S. J. D., Francis, S. and Sabeena, M. 2015. Effect of inlet temperature on physicochemical properties of spray-dried jamun fruit juice powder. *Powder Technology* 274: 37-43.
- Saravolatz, L. D., Pawlak, J., Saravolatz, S. N. and Johnson, L. B. 2013. *In vitro* activity of retapamulin against *Staphylococcus aureus* resistant to various antimicrobial agents. *Antimicrobial Agents and Chemotherapy* 57: 4547-4550.
- Schwalbe, R., Steele-Moore, L. and Goodwin, A. C. 2007. *Antimicrobial Susceptibility Testing Protocols*. Boca Raton: CRC Press.
- Serna-Saldivar, S. O. 2012. *Cereal Grains: Laboratory Reference and Procedures Manual*. Boca Raton: CRC Press.
- Sharma, S., Sharma, M. C., Kohli, D. V. and Chaturvedi, S. C. 2009. Formulation, Evaluation, Wound Healing Studies of Benzene-95% Absolute Ethanol Extract of Leaves. *Journal of Optoelectronics and Biomedical Materials* 1(4): 375-378.
- Sharma, V., Gupta, D. K. and Sharma, J. P. 2011. Selective anti-proliferative effect of ocimum, ginger and bitter gourd on human cancer cells. *Journal of Biotechnology and Biotherapeutics* 2(7): 12-16.
- Shi, J., Xue, S. J., Ma, Y., Li, D., Kakuda, Y. and Lan, Y. 2009. Kinetic study of saponins B stability in navy beans under different processing conditions. *Journal of Food Engineering* 93: 59-65.
- Singh, A., Kuila, A., Yadav, G. and Banerjee, R. 2011. Process optimization for the extraction of polyphenols from okara. *Food Technology and Biotechnology* 49(3): 322-328.
- Singletary, K., Faller, J., Li, J. Y. and Mahungu, S. 2000. Effect of extrusion on isoflavone content and antiproliferative bioactivity of soy/corn mixtures. *Journal of Agricultural and Food Chemistry* 48(8): 3566-3571.

- Snee, L. S., Nerurkar, V. R., Dooley, D. A., Efir, J. T., Shovic, A. C. and Nerurkar, P. V. 2011. Strategies to improve palatability and increase consumption intentions for *Momordica charantia* (bitter melon): A vegetable commonly used for diabetes management. *Nutrition Journal* 10:78.
- Soetan, K. O., Oyekunle, M. A., Aiyelaagbe O. O. and Fafunso, M. A. 2006. Evaluation of the antimicrobial activity of saponins extract of *Sorghum bicolor* L. moench. *African Journal of Biotechnology* 5(23): 2405–2407.
- Sohi Sukhman, K., Sharma, N. S., Hanuman, B., Baljit, S. and Savita, S. 2015. Effect of extrusion variables (moisture content, barrel temperature and screw speed) on the reduction of aflatoxins in maize. *International Research Journal of Biological Sciences* 4(5): 31-37.
- Suhaimi, M. 2013. Review on the application of a tray dryer system for agricultural products. *World Applied Sciences Journal* 22(3): 424-433.
- Supraja, P. and Usha, R. 2013. Antibacterial and phytochemical screening from leaf and fruit extracts of *Momordica Charantia*. *International Journal of Pharma and Bio Sciences* 4(1): 787–793.
- Sylvester, W. S., Son, R., Lew, K. F. and Rukayadi, Y. 2015. Antibacterial activity of Java turmeric (*Curcuma xanthorrhiza* Roxb.) extract against *Klebsiella pneumoniae* isolated from several vegetables. *International Food Research Journal* 22(5): 1770-1776.
- Tam, K. S. 2009. Effects of bitter melon extracts on adipogenesis of 3T3-L1 adipocytes. Pok Fu Lam, Hong Kong: The University of Hong Kong, MSc thesis.
- Tan, S. P. 2016. Extraction and encapsulation of bioactive compounds of bitter melon. Newcastle, Australia: University of Newcastle, PhD thesis.
- Tan, S. P., Parks, S. E., Stathopoulos, C. E. and Roach, P. D. 2014b. Extraction of flavonoids from bitter melon. *Food and Nutrition Sciences* 5(5): 458-465.
- Tan, S. P., Parks, S. E., Stathopoulos, C. E. and Roach, P. D. 2014a. Greenhouse-grown bitter melon: production and quality characteristics. *Journal of the Science of Food and Agriculture* 94(9): 1896-1903.
- Tan, S. P., Stathopoulos, C., Parks, S. and Roach, P. 2014. An optimised aqueous extract of phenolic compounds from bitter melon with high antioxidant capacity. *Antioxidants* 3(4): 814-829.
- Tan, S. P., Tuyen, C. K., Parks, S. E., Stathopoulos, C. E. and Roach, P. D. 2015. Effects of the spray-drying temperatures on the physiochemical properties of an encapsulated bitter melon aqueous extract powder. *Powder Technology* 281: 65-75.

- Tan, S. P., Vuong, Q. V., Stathopoulos, C. E., Parks, S. E. and Roach, P. D. 2014. Optimized aqueous extraction of saponins from bitter melon for production of a saponins-enriched bitter melon powder. *Journal of Food Science* 79(7): 1372-1381.
- Thakur, M., Melzig, M. F., Fuchs, H. and Weng, A. 2011. Chemistry and pharmacology of saponins: special focus on cytotoxic properties. *Botanics: Targets and Therapy* 1: 19-29.
- Traynor, M., Burke, R., Frias, J. M., Gaston, E. and Barry-Ryan, C. 2013. Formation and stability of an oil in water emulsion containing lecithin, xanthan gum and sunflower oil. *International Food Research Journal* 20(5): 2173-2181.
- Tuyen, C. K., Nguyen, M. H. and Roach, P. D. 2010. Effects of spray drying conditions on the physicochemical and antioxidant properties of the Gac (*Momordica cochinchinensis*) fruit aril powder. *Journal of Food Engineering* 98(3): 385-392.
- Ullah, M., Showkat, M., Ahmed, N. U., Islam, S. and Absar, N. 2012. Evaluation of *Momordica charantia* L. fruit extract for analgesic and anti-inflammatory activities using *in vivo* assay. *Research Journal of Medicinal Plant* 6(3): 236-244.
- United States Department of Agriculture (USDA) National Plant Data Center (NRCS). (2011). Flora of North America Expertise Network: *Momordica charantia*. Retrieved on July 10, 2016 from the Integrated Taxonomic Information System on-line database website: www.itis.gov/servlet/Taxonomic Serial No: 22399
- United States Department of Agriculture (USDA). 2015. Quantitative analysis of bacteria in foods as sanitary indicators. Food safety and inspection service laboratories MLG 3.02. Athens: Office of Public Health Science, USDA.
- Utama-ang, N., Chompreeda, P., Haruthaithanasan, V., Lerdvuthisopon, N., Suwonsichon, T., Wood, K. and Watkins, B. A. 2006. Identification of major saponins from jiaogulan extract (*Gynostemma pentaphyllum*). *Kasetsart University Journal* 40: 59-66.
- Vandekinderen, I., Devlieghere, F., Van Camp, J., Denon, Q., Alarcon, S. S., Ragaert, P. and De Meulenaer, B. 2009. Impact of a decontamination step with peroxyacetic acid on the shelf-life, sensory quality and nutrient content of grated carrots packed under equilibrium modified atmosphere and stored at 7°C. *Postharvest Biology and Technology* 54(3): 141-152.
- Virdi, J., Sivakami, S., Shahani, S., Suthar, A. C., Banavalikar, M. M. and Biyani, M. K. 2003. Antihyperglycemic effects of three extracts from *Momordica charantia*. *Journal of Ethnopharmacology* 88(1): 107-111.

- Walker, J. R. L. 1994. Antimicrobial compounds in food plants. In Dillon, V. M. and Board, R. G. (Eds). *Natural Antimicrobial Systems and Food Preservation*, p. 181-204. Wallingford: CAB International.
- Weidner, S., Powalka, A., Karamac, M. and Amarowicz, R. 2012. Extracts of phenolic compounds from seeds of three wild grapevines - comparison of their antioxidant activities and the content of phenolic compounds. *International Journal of Molecular Sciences* 13(3): 3444-3457.
- Wink, M. 2010. Introduction. In Wink, M. (Ed). *Annual Plant Reviews, Functions and Biotechnology of Plant Secondary Metabolites*, p. 1-20. Chichester: Wiley-Blackwell.
- Wink, M. 2015. Modes of action of herbal medicines and plant secondary metabolites. *Medicines* 2(3): 251-286.
- Yeo, Y. L., Chia, Y. Y., Lee, C. H., Sow, H. S. and Yap, W. S. 2014. Effectiveness of Maceration Periods with Different Extraction Solvents on *in-vitro* Antimicrobial Activity from Fruit of *Momordica charantia* L.. *Journal of Applied Pharmaceutical Science* 4(10): 016-023.
- Yousef, A. E. and Carlstrom, C. 2003. *Food microbiology: a laboratory manual*. Hoboken: John Wiley and Sons.
- Youssef, K. M., Mokhtar S. M. 2014. Effect of drying methods on the antioxidant capacity, color and phytochemicals of *Portulaca oleracea* L. leaves. *Journal of Nutrition and Food Sciences* 4(6): 1-6.
- Youssef, T., Guessennd, N. K., Souleymane, M., Abou, Q., Karamoko, Q., Kouadio Fernique, K., Adama, C. and Mireille, D. 2015. Phytochemical screening and *in vitro* evaluation of the antibacterial properties of *Terminalia macroptera* stem bark extracts against selected pathogenic bacteria. *Scholars Research Library, Der Pharmacia Lettre* 7(11): 62-67.
- Yusoff, N. A. H., Noor, N. F. and Rukayadi, Y. 2015. Effects of *Cosmos caudatus* Kunth (*Ulam raja*) extract on microflora in raw chicken meat. *International Journal of Current Microbiology and Applied Sciences* 4(2): 426-435.
- Yusoff, N. A. H., Sanuan, F. M. and Rukayadi, Y. 2015. *Cosmos caudatus* Kunth extract reduced number of microflora in oyster mushroom (*Pleurotus ostreatus*). *International Food Research Journal* 22: 1837-1842.
- Zagory, D. 1999. Effects of post-processing handling and packaging on microbial populations. *Postharvest Biology and Technology* 15: 313-321.
- Zhu, Y., Dong, Y., Qian, X., Cui, F., Guo, Q., Zhou, X. and Xiong, Z. 2012. Effect of superfine grinding on antidiabetic activity of bitter melon powder. *International Journal of Molecular sciences* 13(11): 14203-14218.

Zulbadli, N., Alwi, H. and Hamid, K. H. K. 2011. *Momordica charantia* extraction by using pressurized boiling system and compounds identification through gas chromatography mass spectrometry. International Journal of Engineering and Technology 11(03): 79-84.

