

UNIVERSITI PUTRA MALAYSIA

POTENTIAL OF NATIVE SAGO STARCH AS PREBIOTIC WITH AN ANTI-OBESITY FUNCTIONALITY

MAYRILYN SOLO THOMPSON LAANG

FSPM 2018 3



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By

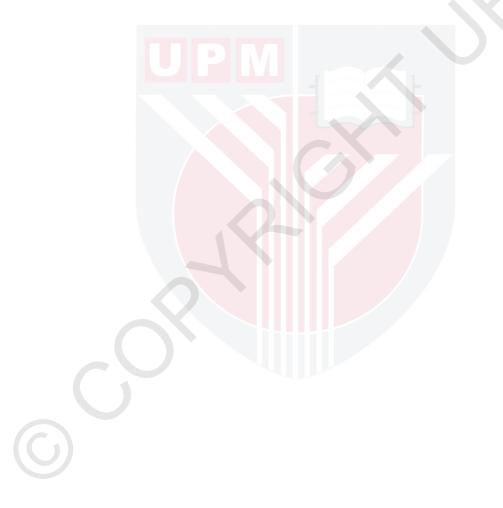
MAYRILYN SOLO THOMPSON LAANG

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

August 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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By

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August 2018

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One hypothesis that has gained attention for the occurrence of obesity is due to the reaction of the microorganisms in our gut. It have been established that the food intake and dietary composition modulates the composition of the gut microbiota. Resistant starch has gained attention due to its benefits to host's health which resembles prebiotics. Sago starch has been reported containing 69% of RS. Sago starch is an important agricultural commodity in Sarawak; however, it is less exploited. This study aimed to determine the potential of sago starch as prebiotics and evaluating the range of low dosage that can confers health benefit in obese management which could be added as functional value to increase its competitiveness with other starches. A 48 male Sprague Dawley rats were fatinduced for 6 weeks prior to RS intervention. Resistant starch intervention was conducted for 8 weeks. Body weight and food intake were observed every week. Faecal samples were collected every 2 weeks interval. At the end of treatment, the rats were sacrificed and gastrointestinal tract were extracted for further analysis. Faecal sample subjected to bacteria profiling using the fluorescent in-situ hybridisation (FISH) technique. Both faecal and caecum sample were subjected to short chain fatty acids analysis using high-performance liquid chromatography. Hepatic lipid content were measured using Folch method. All dosage of sago starch showed a strong correlation of body weight loss, with reduction of food intake. This pattern suggests satiety properties. The increment of dosage RS in treatment showed an increment of *Bifidobacterium* spp. and *Lactobacillus* spp. regardless of phenotype when compared to 0% RS group. This pattern suggests the sago starch having bifidogenic factor. Short chain fatty acids analysis conducted in the faecal and caecum samples showed demonstrated a significant increase of the total SCFA production. Acetate, propionate and butyrate concentration are higher in sago starch group when compared to 0% RS group in both ceacum and faecal sample. Hepatic lipid analysis demonstrated sago starch group (4% SRS, 8% SRS, and 16% SRS) have lower fats accumulation in liver when compared to Hi-maize.

Body fats tissues also showed that RS-enriched diets group have lower fats than the low-fat diet and 0% RS group. The overall results show that sago starch elicits the similar effect as Hi-maize that can bring benefits to health. Sago starch at low dosage has the potential as a prebiotics with anti-obesity functionality with a consistent consumption.



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POTENSI TEPUNG SAGU ASLI SEBAGAI PREBIOTIK YANG MEMPUNYAI FUNSGI ANTI-OBESITI

Oleh

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Satu hipotesis yang mendapat perhatian sebagai punca obesiti adalah reaksi mikroorganisma dalam usus kita. Pengambilan makanan dan komposisi makanan dipercayai boleh megubah komposisi mikrobiota usus. Kanji rintang telah mendapat perhatian kerana manfaatnya kepada kesihatan yang menyerupai prebiotik. Tepung sagu telah dilaporkan mengandungi 69% kanji rintang. Tepung sagu juga adalah komoditi pertanian penting di Sarawak; walau bagaimanapun, ia kurang dieksploitasi. Kajian ini bertujuan untuk menilaikan potensi tepung sagu sebagai prebiotik dan menilai pelbagai dos yang rendah yang dapat memberi manfaat kesihatan dalam pengurusan obes; yang mana laporan ini boleh digunakan untuk menambah fungsi tepung sagu bagi meningkatkan daya saingnya dengan tepung-tepung lain. Sebanyak 48 tikus jantan Sprague Dawley telah dipaksa obes selama 6 minggu. Intervensi kanji rintang dalam makanan dilakukan selama 8 minggu selepas minggu penggemukan. Berat badan dan kadar makanan dicatat setiap minggu. Sampel najis dikumpulkan setiap selang 2 minggu. Pada akhir rawatan, tikus dikorbankan dan saluran usus diekstrak untuk analisis selanjutnya. Sampel najis tertakluk kepada profil bakteria menggunakan teknik hibridisasi insitu fluoresen (FISH). Kedua-dua sampel najis dan usus buntu tertakluk kepada analisa asid lemak rantaian pendek menggunakan kromatografi cecair berprestasi tinggi (HPLC). Kandungan lemak hati dikaji menggunakan kaedah Folch. Semua dos kanji rintang dalam tepung sagu menunjukkan korelasi yang kuat antara penurunan berat badan dengan pengurangan pengambilan makanan. Corak ini menunjukkan sifat kenyang. Peningkatan kanji rintang dalam rawatan menunjukkan kenaikan kiraan Bifidobacterium spp. dan Lactobacillus spp. tidak kira fenotip apabila dibandingkan dengan kumpulan 0% kanji rintang. Corak ini menunjukkan tepung sagu mempunyai faktor bifidogenik. Analisis asid lemak rantaian yang pendek yang dilakukan pada sampel najis dan usus buntu menunjukkan bahawa tepung sagu menunjukkan peningkatan yang ketara dalam

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jumlah pengeluaran SCFA. Kepekatan asetat, propionat dan butirat lebih tinggi berbanding dengan kumpulan 0% kanji rintang dalam usus buntu dan sampel najis. Analisis lipid hepatik menunjukkan kumpulan tepung sagu (4% SRS, 8% SRS dan 16% SRS) mempunyai pengumpulan lemak yang lebih rendah dalam hati berbanding dengan kumpulan *Hi-maize*. Tisu lemak badan juga menunjukkan bahawa kumpulan diet yang diperkaya kanji rintang mempunyai lemak badan yang lebih rendah berbanding diet rendah lemak dan kumpulan 0% kanji rintang. Hasil keseluruhan menunjukkan bahawa tepung sagu mempunyai ciri-ciri yang sama dengan kanji rintang komersial iaitu *Hi-maize* yang mana mampu membawa manfaat kepada kesihatan. Tepung sagu pada dos yang rendah mempunyai potensi sebagai prebiotik yang mempunyai funsgi sebagai anti obesiti dengan penggunaan yang konsisten.



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'Blessed are the merciful for they shall receive mercy." (Matthew 6:7)

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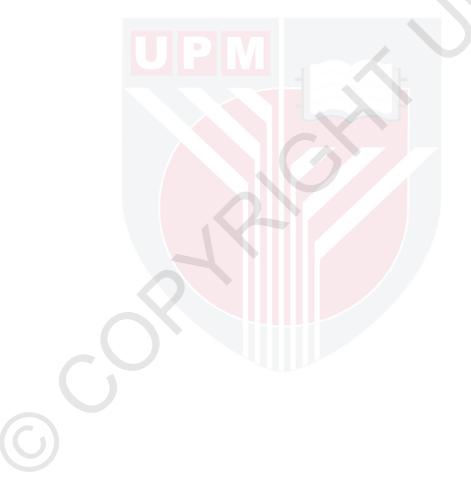
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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

RS	Resistant starch
SCFA	Short chain fatty acid
HPLC	High performance liquid chromatography
m	Meter
kg	Kilogram
μm	Micrometer
g	Gram
DNA	Deoxyribonucleic acid
min	Minute
rpm	Rotation per minute
GOPOD	Glucose oxidase / peroxidase
nm	Nanometer
AMG	Amyloglucosidase
Μ	Molar
mm	Millimetre
mM	mill molar
ANOVA	Analysis of variance
GIP	Gastric inhibitor polypeptide
PYY	Peptide YY
NAFLD	Non-alcoholic fatty liver disease
OP	Obese prone
OR	Obese resistant

CHAPTER 1

INTRODUCTION

The occurrence of obesity is becoming common among adults and children. Worldwide statistic showed 39% of the adult is overweight and obesity in 2014 (World Health Organization, 2017) and estimated to cause 3 to 4 million deaths (Lim *et al.*, 2013). Obesity which predominantly occurs in a developed country has drastically increased in developing country, influenced by modern lifestyles that lack physical activity and unhealthy food choices (Hedley *et al.*, 2004). It has been suggested as a result of increased high simple sugar and high fats diet consumption such as cake, pizza, pastries and etc has been stated as the main culprit of this epidemic (Manz, Amann, Ludwig, Vancanneyt, and Schleifer, 1996). Obesity is a constitutes to the risk factor for type-2 diabetes, liver malfunction, hypertension and cardiovascular diseases (Kyrou and Tsigos, 2009). Realising this, prevention of obesity development is very much essential.

One hypothesis that has gained some attention for the occurrence of obesity is due to the reaction of the microorganisms in our gut. It has been determined that obesity and insulin resistance are associated with low-grade chronic systemic inflammation by the action of bacterial lipopolysaccharide (LPS) (Hotamisligil, 2006). These bacterial LPS is continuously produced in the gut by gram-negative bacteria and transported into intestinal capillaries (Neal *et al.*, 2006). Such LPS is transported from the intestine toward target tissues by a mechanism facilitated by lipoproteins, freshly synthesized from epithelial intestinal cells in response to a high-fat diet (Vreugdenhil *et al.*, 2003) thus triggering the secretion of proinflammatory cytokines (Sweet and Hume, 1996; Wright, Ramos, Tobias, Ulevitch, and Mathison, 1990). These cytokines which are the key inducers to insulin resistance will promote excessive fat accumulation thus leading to development of obesity (Cani, 2007; Cani *et al.*, 2007). Therefore, alteration of the gut microbiota, towards more 'positive and beneficial' environment and prevention of dysbiosis may be the key factors to reduce obesity.

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It is well known that composition of gut microbiota can be modifying by changes in diet. The modulation of gut microbiota is strongly influenced by dietary intake (Conlon and Bird, 2014) as well as genetics and immunology factor (Rodríguez *et al.*, 2015). There are some food ingredient showed positive influence towards the obese gut microbiota in several studies through both *in-vivo* and *in-vitro* experimentation (Hildebrandt *et al.*, 2009; Sarbini, Kolida, Deaville, Gibson, and Rastall, 2014). The imbalance of these composition *i.e* dysbiosis is believed leading to the rising of many diseases. Due to this findings, few studies have been conducted to focus more on gut microbiota potential combating metabolic disease through the fermentation of dietary sources (DiBaise, Frank, and Mathur, 2012). Food ingredient such as inulin and oligofructose has proven its ability positively influence the gut microbiota through several studies (Ramirez-Farias *et al.*, 2008; Wang, X. and Gibson, 1993). The food ingredient that plays important role in positively modulating gut microbiota can also be known as prebiotic. Prebiotic is defined as an indigestible starch fraction that beneficially affects the host by selectively stimulating the growth and/or activity of one or limited number of gut microbiota particularly, *Bifidobacteria* sp. and *Lactobacilli* sp. which subsequently improves the host health (Roberfroid, 2010). Benefits of prebiotics have been proven which include improvement in intestinal permeability and host immunity, reduction of potentially pathogenic bacteria and improved SCFA production (Slavin, 2013).

One of the potential candidates of prebiotics which is gaining attention is resistant starch (RS). The starch is named as resistant starch due to its ability to escape digestion in the small intestine and passes into the colon (Xie, Liu, and Cui, 2006). Consumption of resistant starch is coming to attention due to its ability to confer health benefits to the consumer which resemble prebiotics, and are abundant in many common diets (Zaman and Sarbini, 2015). The beneficial effect of resistant starch has been extensively reviewed and this includes glycaemia response, glucose reduction, fat oxidation, insulin sensitivity and satiety (Belobrajdic, King, Christophersen, and Bird, 2012).

In this study, *Metroxylon sagu* also known as sago is used as tested substrate. Sago starch is one of the most important agricultural commodities in Malaysia, particularly Sarawak where 96% yield are particularly from Sarawak (Uthumporn, Wahidah, and Karim, 2014; Zi-Ni, Rosma, Karim, and Liong, 2015). Previous study reported sago starch containing 69% of resistant starch which was considered high compared to other commercial resistant starch such as Hi-maize®, Novelose, and Fibersym (Zaman, 2015). The amylose content and resistant starch were claimed influencing the digestibility of substrate which contributing factor to confer health benefits to it consumer (Bajury, Rawi, Sazali, Abdullah, and Sarbini, 2017; Falony *et al.*, 2009; Slavin, 2013).

Currently, Malaysia is the largest sago starch exporter with annual extracted starch at a range of 40,000 to 51,000 tonnes per year from 2004 to 2013 (den Besten *et al.*, 2013; Olano-Martin, Mountzouris, Gibson, and Rastall, 2000; Uthumporn *et al.*, 2014). Sago starch extracted per unit area is significantly higher than that of rice, corn, wheat, and cassava which could produce up to 25 tonnes per hectare of starch per year (Silvi, Rumney, Cresci, and Rowland, 1999). However, the consumption of sago only takes up to 3% of other starch resources globally when compared to other dominated starch such as tapioca, potato, and corn which can go up to 300,000 tonnes annually (Zaman, 2015). In order to increase the competitiveness with other sources, a functional value should be added as a marketing strategy and directly contribute the growth of sago-farming. In present study, we are comparing sago starch with the commercial resistant starch type II, Hi-maize. This is due to the similar intrinsic factor of sago starch with Hi-maize (type II RS) which the starches was extracted and occur in its natural granule form (Nugent, 2005). Moreover, Hi-maize has been extensively studied for it beneficial properties towards human's health which has been reviewed for it prebiotics effect (Zaman and Sarbini, 2015).

Appropriate amount of resistant starch is necessary to confer its beneficial physiological effects. Previous studies reported positive impact towards host health such as promotes satiety, increase short chain fatty acids production and reduce body weight (Le Leu, Hu, Brown, Woodman, and Young, 2009; Raben *et al.*, 1994; Zenel and Stewart, 2015). However, high dose of resistant starch was applied in these studies. Generally, food ingredient has low resistant starch (Zi-Ni *et al.*, 2015). Belenguer *et al.* (2006) suggested the daily resistant starch consumption should be approximately 20 g for it to confer health benefits. However, the daily intake of an individual differs for the different region where American, the range of 3 g to 8 g (Demigné *et al.*, 1995); European, approximately 4 g (Dysseler and Hoffem, 1994) and Serbian, approximately 6 g (Byrne, Chambers, Morrison, and Frost, 2015). Hence, in this study, we are evaluating the ability of low dosage of the resistant starch from sago starch which within the range of recommendation daily consumption to confer health benefit particularly to combat obesity or as a weight loss regime.

In addition, obese-prone and obese-resistant phenotype was used in the study to investigate the response of gut microbiota and it fermentation product which could be used as prebiotic potential screening upon the consumption as well as its ability as obese management regime. The diet-induced obese rats mimicking closely to obese human which only some rats developed insulin resistance and dyslipidemia (Belobrajdic *et al.*, 2012; Levin, Hogan, and Sullivan, 1989). The resulted changes can be comparable to the human body rather than using genetic-modified obese rats. This study aims to evaluate the impact of different low dosage of the resistant starch from sago starch that could confer benefits health through *in-vivo* experimentation. The specific objectives of this study as follows:

- 1. To evaluate the anti-obesity Property of Sago Starch using Fat- Induced Rats.
- 2. To investigate the response of gut microbiota and it fermentation product upon the consumption of sago starch for its prebiotic potential.

REFERENCES

- Al-Lahham, S., Peppelenbosch, M., Roelofsen, H., Vonk, R., & Venema, K. (2010). Biological effects of propionic acid in humans; metabolism, potential applications and underlying mechanisms. *Biochimica et Biophysica Acta*, 1801, 1175 - 1183.
- Alvina, M., & Araya, H. (2004). Rapid carbohydrate digestion rate produced lesser short-term satiety in obese preschool children. *European journal of clinical nutrition*, 58(4), 637-642.
- Amin, T., & Mercer, J. (2016). Hunger and satiety mechanisms and their potential exploitation in the regulation of food intake. *Current obesity reports*, 5(1), 106-112.
- Anderson, D. (2005). From Pediatric Nutrition. *Handbook of Pediatric Nutrition*, 53 71.
- Anderson, G., Cho, C. E., Akhavan, T., Mollard, R. C., Luhovyy, B. L., & Finocchiaro, E. T. (2010). Relation between estimates of cornstarch digestibility by the Englyst in vitro method and glycemic response, subjective appetite, and short-term food intake in young men-. American Journal of Clinical Nutrition, 91(4), 932-939.
- Anderson, G., & Woodend, D. (2003). Consumption of sugars and the regulation of short-term satiety and food intake. *American Journal of Clinical Nutrition*, 78(4), 843S-849S.
- Andersson, U., Rosen, L., Wierup, N., Ostman, E., Bjorck, I., & Holm, C. (2010). A low glycaemic diet improves oral glucose tolerance but has no effect on beta-cell function in C57BL/6J mice. *Diabetes, Obesity and Metabolism*, 12, 976 - 982.
- Arora, T., Sharma, R., & Frost, G. (2011). Propionate. Anti-obesity and satiety enhancing factor? *Appetite*, 56, 511 515.
- Arumugam, M., Raes, J., Pelletier, E., Le Paslier, D., Yamada, T., Mende, D. R., . . Batto, J.-M. (2011). Enterotypes of the human gut microbiome. *Nature*, 473(7346), 174-180.
- Avershina, E., Storrø, O., Øien, T., Johnsen, R., Pope, P., & Rudi, K. (2014). Major faecal microbiota shifts in composition and diversity with age in a geographically restricted cohort of mothers and their children. *Federation* of European Microbiological Societies: Microbiology Ecology, 87(1), 280-290.

- Aziz, A., Kenney, L., Goulet, B., & Abdel-Aal El, S. (2009). Dietary starch type affects body weight and glycemic control in freely fed but not energyrestricted obese rats. *Journal of Nutrition*, 139, 1881 - 1889.
- Bajury, D. M., Rawi, M. H., Sazali, I. H., Abdullah, A., & Sarbini, S. R. (2017). Prebiotic evaluation of red seaweed (Kappaphycus alvarezii) using in vitro colon model. *International Journal of Food Sciences and Nutrition*, 1-8.
- Batterham, R. L., Cohen, M. A., Ellis, S. M., Le Roux, C. W., Withers, D. J., Frost, G. S., . . . Bloom, S. R. (2003). Inhibition of food intake in obese subjects by peptide YY3–36. *New England Journal of Medicine*, 349(10), 941-948.
- Batterham, R. L., Cowley, M. A., Small, C. J., Herzog, H., Cohen, M. A., Dakin, C. L., . . Ghatei, M. A. (2002). Gut hormone PYY3-36 physiologically inhibits food intake. *Nature*, 418(6898), 650-654.
- Belenguer, A., Duncan, S. H., Calder, A. G., Holtrop, G., Louis, P., Lobley, G. E., & Flint, H. J. (2006). Two routes of metabolic cross-feeding between Bifidobacterium adolescentis and butyrate-producing anaerobes from the human gut. *Applied and Environmental Microbiology*, 72(5), 3593-3599.
- Belenguer, A., Duncan, S. H., Holtrop, G., Anderson, S. E., Lobley, G. E., & Flint, H. J. (2007). Impact of pH on lactate formation and utilization by human fecal microbial communities. *Applied and Environmental Microbiology*, 73(20), 6526-6533.
- Belobrajdic, D., King, R., Christophersen, C., & Bird, A. (2012). Dietary resistant starch dose-dependently reduces adiposity in obesity-prone and obesity-resistant male rats. *Nutrition and Metabolism*, 9(1), 93.
- Benelam, B. (2009). Satiation, satiety and their effects on eating behaviour. *Nutrition Bulletin*, 34(2), 126-173.
- Bieri, J. G. (1980). Second report of the ad hoc committee on standards for nutritional studies. *The Journal of nutrition*, 110(8), 1726-1726.
- Bindels, L. B., Munoz, R. R. S., Gomes-Neto, J. C., Mutemberezi, V., Martínez, I., Salazar, N., . . . de Los Reyes-Gavilán, C. G. (2017). Resistant starch can improve insulin sensitivity independently of the gut microbiota. *Microbiome*, 5(1), 12.
- Ble-Castillo, J. L., Juárez-Rojop, I. E., Tovilla-Zárate, C. A., García-Vázquez, C., Servin-Cruz, M. Z., Rodríguez-Hernández, A., . . . Díaz-Zagoya, J. C. (2017). Acute Consumption of Resistant Starch Reduces Food Intake but Has No Effect on Appetite Ratings in Healthy Subjects. *Nutrients*, 9(7), 696.

- Blundell, J., De Graaf, C., Hulshof, T., Jebb, S., Livingstone, B., Lluch, A., . . . Van Der Knaap, H. (2010). Appetite control: methodological aspects of the evaluation of foods. *Obesity reviews*, 11(3), 251-270.
- Bodinham, C., Frost, G., & Robertson, M. (2010). Acute ingestion of resistant starch reduces food intake in healthy adults. *British Journal of Nutrition*, 103, 917 922.
- Bolade, M. K., & Bello, S. B. (2006). Selected physicochemical properties of flour from the root of African fan palm (Borassus aethiopum). *International Journal of Food Properties*, 9(4), 701-713.
- Bray, G. A., Paeratakul, S., & Popkin, B. M. (2004). Dietary fat and obesity: a review of animal, clinical and epidemiological studies. *Physiology & behavior*, 83(4), 549-555.
- Byrne, C., Chambers, E., Morrison, D., & Frost, G. (2015). The role of short chain fatty acids in appetite regulation and energy homeostasis. *International journal of obesity (2005), 39*(9), 1331.
- Cani, P. (2007). Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*, 56, 1761-1772.
- Cani, P., Neyrinck, A., Fava, F., Knauf, C., Burcelin, R., Tuohy, K., ... Delzenne, N. (2007). Selective increases of bifidobacteria in gut microflora improve high-fat-diet-induced diabetes in mice through a mechanism associated with endotoxaemia. *Diabetologia*, 50(11), 2374-2383.
- Carlson, T. G., Larsson, K., Dinh-Nguyen, N., & Krog, N. (1979). A study of the amylose-monoglyceride complex by raman spectroscopy. *Starch-Stärke*, 31(7), 222-224.
- Chauhan, M., Henderson, G., & McGuire, W. (2008). Enteral feeding for very low birth weight infants: reducing the risk of necrotising enterocolitis. *Archives of Disease in Childhood: Fetal & Neonatal, 93*, F162 F166.
- Cheng, H., & Lai, M. (2000). Fermentation of resistant rice starch produces propionate reducing serum and hepatic cholesterol in rats. *Journal of Nutrition*, 130, 1991 1995.
- Choi, H., Eo, H., Park, K., Jin, M., Park, E.-J., Kim, S.-H., . . . Kim, S. (2007). A water-soluble extract from Cucurbita moschata shows anti-obesity effects by controlling lipid metabolism in a high fat diet-induced obesity mouse model. *Biochemical and biophysical research communications*, 359(3), 419-425.
- Collado, M. C., Isolauri, E., Laitinen, K., & Salminen, S. (2008). Distinct composition of gut microbiota during pregnancy in overweight and

normal-weight women. *The American journal of clinical nutrition*, 88(4), 894-899.

- Cong, X., Xu, W., Romisher, R., Poveda, S., Forte, S., Starkweather, A., & Henderson, W. A. (2016). Focus: Microbiome: Gut Microbiome and Infant Health: Brain-Gut-Microbiota Axis and Host Genetic Factors. *The Yale Journal of Biology and Medicine*, **89**(3), 299.
- Conlon, M. A., & Bird, A. R. (2014). The impact of diet and lifestyle on gut microbiota and human health. *Nutrients*, 7(1), 17-44.
- Cummings, J., Beatty, E., Kingman, S., Bingham, S., & Englyst, H. (1996). Digestion and physiological properties of resistant starch in the human large bowel. *British Journal of Nutrition*, *75*(5), 733-747.
- Cummings, J., & Englyst, H. (1991). Measurement of starch fermentation in the human large intestine. *Canadian journal of physiology and pharmacology*, 69(1), 121-129.
- Cummings, J., Pomare, E., Branch, W., Naylor, C., & Macfarlane, G. (1987). Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut*, 28, 1221 1227.
- Deepa, G., Singh, V., & Naidu, K. A. (2010). A comparative study on starch digestibility, glycemic index and resistant starch of pigmented ('Njavara'and 'Jyothi') and a non-pigmented ('IR 64') rice varieties. *Journal of food science and technology*, 47(6), 644-649.
- Demigné, C., Morand, C., Levrat, M.-A., Besson, C., Moundras, C., & Rémésy, C. (1995). Effect of propionate on fatty acid and cholesterol synthesis and on acetate metabolism in isolated rat hepatocytes. *British Journal of Nutrition*, 74(2), 209-219.
- den Besten, G., van Eunen, K., Groen, A. K., Venema, K., Reijngoud, D.-J., & Bakker, B. M. (2013). The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *Journal of lipid research*, 54(9), 2325-2340.
- Dethlefsen, L., McFall-Ngai, M., & Relman, D. A. (2007). An ecological and evolutionary perspective on human–microbe mutualism and disease. *Nature*, **449**(7164), 811-818.
- DiBaise, J. K., Frank, D. N., & Mathur, R. (2012). Impact of the gut microbiota on the development of obesity: current concepts. *The American Journal of Gastroenterology Supplements*, 1(1), 22-27.
- Duca, F. A., Sakar, Y., Lepage, P., Devime, F., Langelier, B., Doré, J., & Covasa, M. (2014). Replication of obesity and associated signaling pathways

through transfer of microbiota from obese prone rat. *Diabetes*, DB_131526.

- Duffy, P. H., Lewis, S. M., Mayhugh, M. A., McCracken, A., Thorn, B. T., Reeves, P. G., . . . Feuers, R. J. (2002). Effect of the AIN-93M purified diet and dietary restriction on survival in Sprague-Dawley rats: implications for chronic studies. *The Journal of nutrition*, 132(1), 101-107.
- Duncan, S. H., Belenguer, A., Holtrop, G., Johnstone, A. M., Flint, H. J., & Lobley, G. E. (2007). Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrateproducing bacteria in feces. *Applied and Environmental Microbiology*, 73(4), 1073-1078.
- Dysseler, P., & Hoffem, D. (1994). *Estimation of resistant starch intake in Europe*. Paper presented at the Proceedings of the concluding plenary meeting of EURESTA.
- Eckburg, P. B., Bik, E. M., Bernstein, C. N., Purdom, E., Dethlefsen, L., Sargent, M., . . . Relman, D. A. (2005). Diversity of the human intestinal microbial flora. *Science*, **308**(5728), 1635-1638.
- Englyst, H. N., Kingman, S., & Cummings, J. (1992). Classification and measurement of nutritionally important starch fractions. *European journal of clinical nutrition*, 46, S33-50.
- Falony, G., Lazidou, K., Verschaeren, A., Weckx, S., Maes, D., & De Vuyst, L. (2009). In vitro kinetic analysis of fermentation of prebiotic inulin-type fructans by Bifidobacterium species reveals four different phenotypes. *Applied and Environmental Microbiology*, 75(2), 454-461.
- Fernandes, J., Su, W., Rahat-Rozenbloom, S., Wolever, T., & Comelli, E. (2014). Adiposity, gut microbiota and faecal short chain fatty acids are linked in adult humans. *Nutrition & diabetes*, 4(6), e121.
- Flint, H. J., Duncan, S. H., Scott, K. P., & Louis, P. (2015). Links between diet, gut microbiota composition and gut metabolism. *Proceedings of the Nutrition Society*, **74**(01), 13-22.
- Folch, J., Lees, M., & Sloane-Stanley, G. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, 226(1), 497-509.
- Franks, A. H., Harmsen, H. J., Raangs, G. C., Jansen, G. J., Schut, F., & Welling, G. W. (1998). Variations of bacterial populations in human feces measured by fluorescent in situ hybridization with group-specific 16S

rRNA-targeted oligonucleotide probes. *Applied and Environmental Microbiology*, 64(9), 3336-3345.

- Fukami, H., Tachimoto, H., Kishi, M., Kaga, T., & Tanaka, Y. (2010). Acetic acid bacterial lipids improve cognitive function in dementia model rats. *Journal of agricultural and food chemistry*, 58(7), 4084-4089.
- Fukuda, S., Toh, H., Hase, K., Oshima, K., Nakanishi, Y., Yoshimura, K., . . . Suzuki, T. (2011). Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature*, 469(7331), 543-547.
- Fuller, R. (1991). Probiotics in human medicine. Gut, 32(4), 439.
- Georgieff, M. (2005). Nutrition. Avery's Neonatology: Pathophysiology and Management of the New Born, 380 381.
- Gibson, G., Hutkins, R., Sanders, M. E., Prescott, S. L., Reimer, R. A., Salminen, S. J., . . . Cani, P. D. (2017). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature Reviews Gastroenterology and Hepatology*, 14(8), 491.
- Gibson, G., & Nathalie, D. (2008). Inulin and oligofructose. New Scientific Developments. Avery's Neonatology: Pathophysiology and Management of the New Born, 43, 54 59.
- Gibson, G., & Roberfroid, M. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *The Journal of nutrition*, 125(6), 1401.
- Gibson, G., & Wang, X. (1994). Bifidogenic properties of different types of fructooligosaccharides. *Food Microbiology*, 11(6), 491-498.
- Gibson, G. R., Probert, H. M., Van Loo, J., Rastall, R. A., & Roberfroid, M. B. (2004). Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutrition Research Reviews*, *17*(2), 259-275.
- Gomez, E., Tuohy, K., Gibson, G., Klinder, A., & Costabile, A. (2010). In vitro evaluation of the fermentation properties and potential prebiotic activity of Agave fructans. *Journal of applied microbiology*, *108*(6), 2114-2121.
- Haenen, D., Zhang, J., Souza da Silva, C., Bosch, G., van der Meer, I. M., van Arkel, J., . . . Kemp, B. (2013). A Diet High in Resistant Starch Modulates Microbiota Composition, SCFA Concentrations, and Gene Expression in Pig Intestine–3. *The Journal of nutrition*, 143(3), 274-283.
- Hald, S., Schioldan, A. G., Moore, M. E., Dige, A., Lærke, H. N., Agnholt, J., ... Gregersen, S. (2016). Effects of arabinoxylan and resistant starch on

intestinal microbiota and short-chain fatty acids in subjects with metabolic syndrome: a randomised crossover study. *Public Library of Science (PLoS One)*, 11(7), e0159223.

- Harazaki, T., Inoue, S., Imai, C., Mochizuki, K., & Goda, T. (2014). Resistant starch improves insulin resistance and reduces adipose tissue weight and CD11c expression in rat OLETF adipose tissue. *Nutrition*, *30*(5), 590-595.
- Hedley, A. A., Ogden, C. L., Johnson, C. L., Carroll, M. D., Curtin, L. R., & Flegal, K. M. (2004). Prevalence of overweight and obesity among US children, adolescents, and adults, 1999-2002. *Journal of the American Medical Association*, 291(23), 2847-2850.
- Higgins, J. (2004). Resistant starch: metabolic effects and potential health benefits. *Journal of AOAC International*, 87, 761 - 768.
- Higgins, J., Brand Miller, J., & Denyer, G. (1996). Development of insulin resistance in the rat is dependent on the rate of glucose absorption from the diet. *Journal of Nutrition*, *126*, 596 602.
- Higgins, J., Higbee, D., Donahoo, W., Brown, I., Bell, M., & Bessesen, D. (2004). Resistant starch consumption promotes lipid oxidation. *Nutrition & Metabolism*, 1, 8.
- Higgins, J., Jackman, M., Brown, I., Johnson, G., Steig, A., Wyatt, H., . . . Maclean, P. (2011). Resistant starch and exercise independently attenuate weight regain on a high fat diet in a rat model of obesity. *Nutrition & Metabolism*, 8, 49.
- Hildebrandt, M. A., Hoffmann, C., Sherrill-Mix, S. A., Keilbaugh, S. A., Hamady, M., Chen, Y. Y., . . Wu, G. D. (2009). High-fat diet determines the composition of the murine gut microbiome independently of obesity. *Gastroenterology*, 137(5), 1716-1724. e1712.
- Hirao, K., Kanamori, K., Yoneyama, Y., & Takahashi, S. (2004). Physical and sensory properties of biscuits containing partially replaced sago starch. *Journal of Home Economics of Japan*, 55(9), 715-723.
- Hirao, K., Kondo, T., Kainuma, K., & Takahashi, S. (2018). Starch Properties and Uses as Food for Human Health and Welfare *Sago Palm* (pp. 285-298): Springer.
- Hold, G. L., Schwiertz, A., Aminov, R. I., Blaut, M., & Flint, H. J. (2003). Oligonucleotide probes that detect quantitatively significant groups of butyrate-producing bacteria in human feces. *Applied and Environmental Microbiology*, 69(7), 4320-4324.

- Hosseini, E., Grootaert, C., Verstraete, W., & Van de Wiele, T. (2011). Propionate as a health-promoting microbial metabolite in the human gut. *Nutrition reviews*, 69(5), 245-258.
- Hotamisligil, G. (2006). Inflammation and metabolic disorders. *Nature*, **444**(7121), 860-867.
- Igarashi, M., Kitada, Y., Yoshiyama, H., Takagi, A., Miwa, T., & Koga, Y. (2001). Ammonia as an Accelerator of Tumor Necrosis Factor Alpha-Induced Apoptosis of Gastric Epithelial Cells inHelicobacter pylori Infection. *Infection and immunity*, **69**(2), 816-821.
- Ingredion. (2018). Boost fibre and resistant starch invisibly with HI-MAIZE® resistant starch. Retrieved August, 28, 2018
- Isken, F., Klaus, S., Petzke, K., Loddenkemper, C., Pfeiffer, A., & Weickert, M. (2010). Impairment of fat oxidation under high- vs. low-glycemic index diet occurs before the development of an obese phenotype. *American Journal of Physiology-Endocrinology and Metabolism*, 298, E287 - E295.
- Ivy, J. L. (2004). Regulation of muscle glycogen repletion, muscle protein synthesis and repair following exercise. *Journal of sports science & medicine*, 3(3), 131.
- Jackman, M. R., MacLean, P. S., & Bessesen, D. H. (2010). Energy expenditure in obesity-prone and obesity-resistant rats before and after the introduction of a high-fat diet. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 299(4), R1097-R1105.
- Jaworski, D. M., Namboodiri, A., & Moffett, J. R. (2015). Acetate as a Metabolic and Epigenetic Modifier of Cancer Therapy. *Journal of Cellular Biochemistry*.
- Karim, A., Tie, A., Manan, D., & Zaidul, I. (2008). Starch from the sago (Metroxylon sagu) palm tree—properties, prospects, and challenges as a new industrial source for food and other uses. *Comprehensive Reviews in Food Science and Food Safety*, 7(3), 215-228.
- Keenan, M., Zhou, J., McCutcheon, K., Raggio, A., Bateman, H., Todd, E., ... Hegsted, M. (2006). Effects of resistant starch, a non-digestible fermentable fiber, on reducing body fat. *Obesity (Silver Spring)*, 14, 1523 - 1534.
- Keenan, M. J., Zhou, J., McCutcheon, K., Raggio, A., Bateman, H., Todd, E., ... Hegsted, M. (2006). Effects of resistant starch, a non-digestible fermentable fiber, on reducing body fat. *Obesity (Silver Spring)*, 14, 1523 - 1534.

- Kennedy, A., Martinez, K., Chuang, C.-C., LaPoint, K., & McIntosh, M. (2009). Saturated fatty acid-mediated inflammation and insulin resistance in adipose tissue: mechanisms of action and implications. *The Journal of nutrition*, 139(1), 1-4.
- Kirpitch, A. R., & Maryniuk, M. D. (2011). The 3 R9s of Glycemic Index: Recommendations, Research, and the Real World. *Clinical Diabetes*, 29(4), 155-159.
- Kitcherside, M., Glen, E., & Webster, A. (2000). Fibrecap: an improved method for the rapid analysis of fibre in feeding stuffs. *Animal Feed Science and Technology*, 86(1-2), 125-132.
- Kleessen, B., Stoof, G., Proll, J., Schmiedl, D., Noack, J., & Blaut, M. (1997). Feeding resistant starch affects fecal and cecal microflora and short-chain fatty acids in rats. *Journal of animal science*, 75(9), 2453-2462.
- Korner, J., Inabnet, W., Conwell, I., Taveras, C., Daud, A., Olivero-Rivera, L., ... Bessler, M. (2006). Differential effects of gastric bypass and banding on circulating gut hormone and leptin levels. *Obesity*, 14(9), 1553-1561.
- Kudoh, K., Shimizu, J., Ishiyama, A., WADA, M., TAKITA, T., KANKE, Y., & INNAMI, S. (1999). Secretion and excretion of immunoglobulin A to cecum and feces differ with type of indigestible saccharides. *Journal of nutritional science and vitaminology*, 45(2), 173-181.
- Kumar, A., Sahoo, U., Baisakha, B., Okpani, O. A., Ngangkham, U., Parameswaran, C., . . Sharma, S. (2018). Resistant starch could be decisive in determining the glycemic index of rice cultivars. *Journal of Cereal Science*, 79, 348-353.
- Kyrou, I., & Tsigos, C. (2009). Obesity in the Elderly Diabetic Patient Is weight loss beneficial? No. *Diabetes Care, 32*(suppl 2), S403-S409.
- Langendijk, P. S., Schut, F., Jansen, G. J., Raangs, G. C., Kamphuis, G. R., Wilkinson, M., & Welling, G. W. (1995). Quantitative fluorescence in situ hybridization of Bifidobacterium spp. with genus-specific 16S rRNAtargeted probes and its application in fecal samples. *Applied and Environmental Microbiology*, 61(8), 3069-3075.
- Le Blay, G., Michel, C., Blottiere, H., & Cherbut, C. (2003). Raw potato starch and short-chain fructo-oligosaccharides affect the composition and metabolic activity of rat intestinal microbiota differently depending on the caecocolonic segment involved. *Journal of applied microbiology*, 94(2), 312-320.
- Le Leu, R. K., Hu, Y., Brown, I. L., Woodman, R. J., & Young, G. P. (2009). Synbiotic intervention of Bifidobacterium lactis and resistant starch

protects against colorectal cancer development in rats. Carcinogenesis, 31(2), 246-251.

- Levin, B. E., Hogan, S., & Sullivan, A. (1989). Initiation and perpetuation of obesity and obesity resistance in rats. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 256(3), R766-R771.
- Ley, R. E., Hamady, M., Lozupone, C., Turnbaugh, P. J., Ramey, R. R., Bircher, J. S., . . . Knight, R. (2008). Evolution of mammals and their gut microbes. *Science*, 320(5883), 1647-1651.
- Ley, R. E., Turnbaugh, P. J., Klein, S., & Gordon, J. I. (2006). Microbial ecology: human gut microbes associated with obesity. *Nature*, **444**(7122), 1022-1023.
- Lien, E., Boyle, F., Wrenn, J., Perry, R., Thompson, C., & Borzelleca, J. (2001). Comparison of AIN-76A and AIN-93G diets: a 13-week study in rats. *Food and chemical toxicology*, *39*(4), 385-392.
- Liestianty, D., Rodianawati, I., Patimah, & Mulaidi. (2016). Chemical Composition Of Modified And Fortified Sago Starch (Metroxylonsp) From Northern Maluku. *International Journal of Applied Chemistry*, 12(3), 243-249.
- Lim, S. S., Vos, T., Flaxman, A. D., Danaei, G., Shibuya, K., Adair-Rohani, H., . . Andrews, K. G. (2013). A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *The lancet*, 380(9859), 2224-2260.
- Linden, D. R. (2014). Hydrogen sulfide signaling in the gastrointestinal tract. Antioxidants & redox signaling, 20(5), 818-830.
- Lissauer, T., & Clayden, G. (2007). Neonatal Medicine. Illustrated Text book of Pediatrics, 145 - 168.
- Livesey, G. (1990). Energy values of unavailable carbohydrate and diets: an inquiry and analysis. *American Journal of Clinical Nutrition*, **51**(4), 617-637.
- Lockyer, S., & Nugent, A. (2017). Health effects of resistant starch. *Nutrition Bulletin*.
- Lu, Y., Fan, C., Li, P., Lu, Y., Chang, X., & Qi, K. (2016). Short chain fatty acids prevent high-fat-diet-induced obesity in mice by regulating G proteincoupled receptors and gut microbiota. *Scientific reports*, 6.

- Macfarlane, G., & Macfarlane, S. (2012a). Bacteria, colonic fermentation, and gastrointestinal health. *Journal of AOAC International*, 95, 50 60.
- Macfarlane, G., & Macfarlane, S. (2012b). Bacteria, colonic fermentation, and gastrointestinal health. *Journal of AOAC International*, **95**(1), 50-60.
- Macfarlane, G., Steed, H., & Macfarlane, S. (2008). Bacterial metabolism and health related effects of galacto-oligosaccharides and other prebiotics. *Journal of applied microbiology, 104*, 305 344.
- Maki, K., Pelkman, C., Finocchiaro, E., Kelley, K., Lawless, A., Schild, A., & Rains, T. (2012). Resistant starch from high-amylose maize increases insulin sensitivity in overweight and obese men. *Journal of Nutrition*, 142, 717 723.
- Manz, W., Amann, R., Ludwig, W., Vancanneyt, M., & Schleifer, K.-H. (1996). Application of a suite of 16S rRNA-specific oligonucleotide probes designed to investigate bacteria of the phylum cytophaga-flavobacterbacteroides in the natural environment. *Microbiology*, 142(5), 1097-1106.
- Meyer, D., & Stasse-Wolthuis, M. (2009). The bifidogenic effect of inulin and oligofructose and its consequences for gut health. *European journal of clinical nutrition*, 63(11), 1277-1289.
- Minato, H., & Suto, T. (1978). Technique for fractionation of bacteria in rumen microbial ecosystem. II. Attachment of bacteria isolated from bovine rumen to cellulose powder in vitro and elution of bacteria attached therefrom. *The Journal of General and Applied Microbiology*, **24**(1), 1-16.
- Mookerjea, S., & Sadhu, D. (1955). Metabolism of Acetic Acid in Kidney and Liver Slices of the Hypothyroid Rats *Endocrinology*, 56(5), 507-510.
- Muir, J., Lu, Z., Young, G., Cameron-Smith, D., Collier, G., & O'Dea, K. (1995). Resistant starch in the diet increases breath hydrogen and serum acetate in human subjects. *American Journal of Clinical Nutrition*, *61*, 792 - 799.
- Nayak, B., Berrios, J. D. J., & Tang, J. (2014). Impact of food processing on the glycemic index (GI) of potato products. *Food Research International*, 56, 35-46.
- Neal, M. D., Leaphart, C., Levy, R., Prince, J., Billiar, T. R., Watkins, S., . . . Schreiber, A. (2006). Enterocyte TLR4 mediates phagocytosis and translocation of bacteria across the intestinal barrier. *The Journal of Immunology*, 176(5), 3070-3079.

Nielsen, S. S. (1998). Food analysis (Vol. 86): Springer.

- Nishina, P., & Freedland, R. (1990). Effects of propionate on lipid biosynthesis in isolated rat hepatocytes. *Journal of Nutrition*, *120*, 668 673.
- Nugent, A. P. (2005). Health properties of resistant starch. *Nutrition Bulletin*, **30**(1), 27-54.
- Olano-Martin, E., Mountzouris, K. C., Gibson, G. R., & Rastall, R. A. (2000). In vitro fermentability of dextran, oligodextran and maltodextrin by human gut bacteria. *British Journal of Nutrition*, *83*(3), 247-255.
- Parvez, S., Malik, K., Kang, S., & Kim, H. (2006). Probiotics and their fermented food products are beneficial for health. *Journal of applied microbiology*, 100, 1171 - 1185.
- Patney, N. L., Mehrotra, M. P., Khanna, H. K., & Kumar, A. (1976). Urinary indican excretion in cirrhosis of liver. *The Journal of the Association of Physicians of India*, 24(5), 291-295.
- Peters, S., Pomare, E., & Fisher, C. (1992). Portal and peripheral blood short chain fatty acid concentrations after caecal lactulose instillation at surgery. *Gut*, 33, 1249 1252.
- Puertollano, E., Kolida, S., & Yaqoob, P. (2014). Biological significance of shortchain fatty acid metabolism by the intestinal microbiome. *Current Opinion in Clinical Nutrition & Metabolic Care, 17*(2), 139-144. doi: 10.1097/MCO.0000000000025
- Raben, A., Tagliabue, A., Christensen, N. J., Madsen, J., Holst, J. J., & Astrup, A. (1994). Resistant starch: the effect on postprandial glycemia, hormonal response, and satiety. *The American journal of clinical nutrition*, 60(4), 544-551.
- Raman, M., Ambalam, P., & Doble, M. (2016). Short-Chain Fatty Acids Probiotics and Bioactive Carbohydrates in Colon Cancer Management (pp. 97-115): Springer.
- Ramirez-Farias, C., Slezak, K., Fuller, Z., Duncan, A., Holtrop, G., & Louis, P. (2008). Effect of inulin on the human gut microbiota: stimulation of Bifidobacterium adolescentis and Faecalibacterium prausnitzii. *British Journal of Nutrition*, 101(4), 541-550.
- Reeves, P., Nielsen, F., & Fahey, G. (1993). AIN-93 Purified Diets for Laboratory Rodents: Final Report of the American Institute of Nutrition Ad Hoc Writing Committee on the Reformulation of the AIN-76A Rodent Diet. *Journal of Nutrition*, 123, 1939 - 1951.
- Reeves, P., Nielsen, F. H., & Fahey Jr, G. C. (1993). AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition ad

hoc writing committee on the reformulation of the AIN-76A rodent diet. *Journal of Nutrition*.

- Roberfroid, M. (2007). Prebiotics: the concept revisited. *The Journal of nutrition*, 137(3), 830S-837S.
- Roberfroid, M. (2010). Prebiotic effects: metabolic and health benefits. *British Journal of Nutrition*, **104**, S1-S63.
- Robertson, M., Bickerton, A., Dennis, A., Vidal, H., & Frayn, K. (2005). Insulinsensitizing effects of dietary resistant starch and effects on skeletal muscle and adipose tissue metabolism. *American Journal of Clinical Nutrition*, 82, 559 - 567.
- Robertson, M., Currie, J., Morgan, L., Jewell, D., & Frayn, K. (2003). Prior shortterm consumption of resistant starch enhances postprandial insulin sensitivity in healthy subjects. *Diabetologia*, 46, 659 - 665.
- Rodríguez-Cabezas, M. E., Camuesco, D., Arribas, B., Garrido-Mesa, N., Comalada, M., Bailón, E., . . Pérez-Roca, C. (2010). The combination of fructooligosaccharides and resistant starch shows prebiotic additive effects in rats. *Clinical nutrition*, 29(6), 832-839.
- Rodríguez, J. M., Murphy, K., Stanton, C., Ross, R. P., Kober, O. I., Juge, N., ... Jenmalm, M. C. (2015). The composition of the gut microbiota throughout life, with an emphasis on early life. *Microbial ecology in health and disease*, 26.
- Roediger, W. (1980). Role of anaerobic bacteria in the metabolic welfare of the colonic mucosa in man. *Gut*, 21(9), 793-798.
- Rolfe, R. D. (2000). The role of probiotic cultures in the control of gastrointestinal health. *The Journal of nutrition*, *130*(2), 396S-402S.
- Sa'ad, H., Peppelenbosch, M. P., Roelofsen, H., Vonk, R. J., & Venema, K. (2010). Biological effects of propionic acid in humans; metabolism, potential applications and underlying mechanisms. *Biochimica et Biophysica Acta* (*BBA*)-*Molecular and Cell Biology of Lipids*, 1801(11), 1175-1183.
- Salminen, S., Bouley, C., Boutron, M.-C., Cummings, J., Franck, A., Gibson, G., . . . Rowland, I. (1998). Functional food science and gastrointestinal physiology and function. *British Journal of Nutrition*, 80(S1), S147-S171.
- Sarbini, S., Kolida, S., Deaville, E. R., Gibson, G. R., & Rastall, R. A. (2014). Potential of novel dextran oligosaccharides as prebiotics for obesity management through in vitro experimentation. *British Journal of Nutrition*, 112(8), 1303-1314.

- Sarbini, S., & Rastall, R. (2011). Prebiotics: metabolism, structure, and function. *Functional Food Reviews*, **3**(3), 93-106.
- Sardá, F. A. H., Giuntini, E. B., Gomez, M. L. P., Lui, M. C. Y., Negrini, J. A., Tadini, C. C., . . . Menezes, E. W. (2016). Impact of resistant starch from unripe banana flour on hunger, satiety, and glucose homeostasis in healthy volunteers. *Journal of Functional Foods*, 24, 63-74.
- Scott, K., Martin, J., Campbell, G., Mayer, C., & Flint, H. (2006). Whole-genome transcription profiling reveals genes up-regulated by growth on fucose in the human gut bacterium "Roseburia inulinivorans". *Journal of bacteriology*, 188(12), 4340-4349.
- Shah, N. (2007). Functional cultures and health benefits. *International Dairy* Journal, 17, 1262 - 1277.
- Shen, R.-L., Zhang, W.-L., Dong, J.-L., Ren, G.-X., & Chen, M. (2015). Sorghum resistant starch reduces adiposity in high-fat diet-induced overweight and obese rats via mechanisms involving adipokines and intestinal flora. *Food* and agricultural immunology, 26(1), 120-130.
- Silvi, S., Rumney, C., Cresci, A., & Rowland, I. (1999). Resistant starch modifies gut microflora and microbial metabolism in human flora-associated rats inoculated with faeces from Italian and UK donors. *Journal of applied microbiology*, 86(3), 521-530.
- Slavin, J. (2013). Fiber and prebiotics: mechanisms and health benefits. *Nutrients*, 5(4), 1417-1435.
- So, P.-W., Yu, W.-S., Kuo, Y.-T., Wasserfall, C., Goldstone, A. P., Bell, J. D., & Frost, G. (2007). Impact of resistant starch on body fat patterning and central appetite regulation. *PloS one*, 2(12), e1309.
- Soliman, M. M., Ahmed, M. M., Salah-eldin, A.-e., & Abdel-Aal, A. A.-A. (2011). Butyrate regulates leptin expression through different signaling pathways in adipocytes. *Journal of veterinary science*, *12*(4), 319-323.
- Srikaeo, K., & Sangkhiaw, J. (2014). Effects of amylose and resistant starch on glycaemic index of rice noodles. LWT-Food Science and Technology, 59(2), 1129-1135.
- Sweet, M. J., & Hume, D. A. (1996). Endotoxin signal transduction in macrophages. *Journal of leukocyte biology*, 60(1), 8-26.
- Sybille, T., June, Z., Michael, K., Roy, M., & Maria L, M. (2013). The intestinal microbiota in aged mice is modulated by dietary resistant starch and correlated with improvements in host responses. *FEMS microbiology ecology*, 83(2), 299-309.

- Tojo, R., Suárez, A., Clemente, M. G., de los Reyes-Gavilán, C. G., Margolles, A., Gueimonde, M., & Ruas-Madiedo, P. (2014). Intestinal microbiota in health and disease: role of bifidobacteria in gut homeostasis. World journal of gastroenterology: WJG, 20(41), 15163.
- Topping, D., & Clifton, P. (2001). Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. *Physiol Rev*, 81(Physiological Reviews), 1031 - 1064.
- Topping, D., Fukushima, M., & Bird, A. (2003a). Resistant starch as a prebiotic and synbiotic: state of the art. *Proceedings of the Nutrition Society*, **62**(1), 171-176.
- Topping, D., Fukushima, M., & Bird, A. R. (2003b). Resistant starch as a prebiotic and synbiotic: state of the art. *Proceedings of the Nutrition Society*, 62(01), 171-176.
- Underwood, M., Salzmand, N., Bennett, S., Barman, M., Mills, D., Marcobal, A., .
 . . Sherman, M. (2009). A randomized placebo controlled comparison of 2 prebiotic/probiotic combinations in preterm infants: Impact on weight gain, intestinal microbiota and fecal short chain fatty acids. *Journal of Pediatric Gastroenterology and Nutrition*, 48, 216 225.
- Upadhyaya, B., McCormack, L., Fardin-Kia, A. R., Juenemann, R., Nichenametla, S., Clapper, J., . . . Dey, M. (2016). Impact of dietary resistant starch type 4 on human gut microbiota and immunometabolic functions. *Scientific reports*, *6*, 28797.
- Uthumporn, U., Wahidah, N., & Karim, A. (2014). *Physicochemical properties of* starch from sago (Metroxylon sagu) palm grown in mineral soil at different growth stages. Paper presented at the IOP Conference Series: Materials Science and Engineering.
- Van Kleef, E., Van Trijp, J., Van Den Borne, J., & Zondervan, C. (2012). Successful development of satiety enhancing food products: towards a multidisciplinary agenda of research challenges. *Critical reviews in food science and nutrition*, 52(7), 611-628.
- Van Loo, J., Coussement, P., De Leenheer, L., Hoebregs, H., & Smits, G. (1995). On the presence of inulin and oligofructose as natural ingredients in the western diet. *Critical Reviews in Food Science & Nutrition*, 35(6), 525-552.
- van Munster, I. P., Tangerman, A., & Nagengast, F. M. (1994). Effect of resistant starch on colonic fermentation, bile acid metabolism, and mucosal proliferation. *Digestive diseases and sciences*, *39*(4), 834-842.

- Vandeputte, D., Falony, G., Vieira-Silva, S., Wang, J., Sailer, M., Theis, S., . . . Raes, J. (2017). Prebiotic inulin-type fructans induce specific changes in the human gut microbiota. *Gut*, gutjnl-2016-313271.
- Vreugdenhil, A. C., Rousseau, C. H., Hartung, T., Greve, J. W. M., van't Veer, C., & Buurman, W. A. (2003). Lipopolysaccharide (LPS)-binding protein mediates LPS detoxification by chylomicrons. *The Journal of Immunology*, 170(3), 1399-1405.
- Wahjuningsih, S. B., Marsono, Y., Praseptiangga, D., & Haryanto, B. (2016). Resistant Starch Content and Glycaemic Index of Sago (Metroxylon spp.) Starch and Red Bean (Phaseolus vulgaris) Based Analogue Ric. *Pakistan Journal of Nutrition*, 15(7), 667-672.
- Wang, C., Shoji, H., Sato, H., Nagata, S., Ohtsuka, Y., Shimizu, T., & Yamashiro, Y. (2007). Effects of Oral Administration of bifidobacterium breve on fecal lactic acid and short chain fatty acids in low birth weight infants. *The Journal of Pediatric Gastroenterology and Nutrition*, 44, 252 - 257.
- Wang, X., & Gibson, G. (1993). Effects of the in vitro fermentation of oligofructose and inulin by bacteria growing in the human *Journal of applied microbiology*, 75(4), 373-380.
- Ward, R. E., Benninghoff, A. D., Healy, B. J., Li, M., Vagu, B., & Hintze, K. J. (2017). Consumption of the total Western diet differentially affects the response to green tea in rodent models of chronic disease compared to the AIN93G diet. *Molecular nutrition & food research*, 61(4), 1600720.
- Wolever, T., Fernandes, J., & Rao, A. (1996). Serum acetate:propionate ratio is related to serum cholesterol in men but not women. *Journal of Nutrition*, *126*, 2790 2797.
- Wolever, T., Spadafora, P., & Eshuis, H. (1991). Interaction between colonic acetate and propionate in humans. *American Journal of Clinical Nutrition*, 53(3), 681-687.
- World Health Organization, W. (2017). Obesity and overweight. Retrieved October 2017
- Wright, S. D., Ramos, R. A., Tobias, P. S., Ulevitch, R. J., & Mathison, J. C. (1990). CD14, a receptor for complexes of lipopolysaccharide (LPS) and LPS binding protein. *Science*, 249(4975), 1431-1433.
- Xie, X. S., Liu, Q., & Cui, S. W. (2006). Studies on the granular structure of resistant starches (type 4) from normal, high amylose and waxy corn starch citrates. *Food Research International*, 39(3), 332-341.

- Yamashita, H., Maruta, H., Jozuka, M., Kimura, R., Iwabuchi, H., Yamato, M., ... Kimoto, M. (2009). Effects of acetate on lipid metabolism in muscles and adipose tissues of type 2 diabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. *Bioscience, biotechnology, and biochemistry*, 73(3), 570-576.
- Yan, L., Combs, J., Gerald, F., DeMars, L. C., & Johnson, L. K. (2011). Effects of the physical form of the diet on food intake, growth, and body composition changes in mice. *Journal of the American Association for Laboratory Animal Science*, 50(4), 488-494.
- Yatim, A., & Nor, N. M. (2011). Pink guava (Psidium guajava) puree supplement on high fat diet-induced obese rats: effects on antioxidant enzyme activities, kidney and liver functions. *Sains Malaysiana*, 40(2), 147-154.
- Yun, J. W. (2010). Possible anti-obesity therapeutics from nature–A review. *Phytochemistry*, 71(14-15), 1625-1641.
- Zaman, S. A. (2015). Production Of Resistant Starch Type Iii And Iv And Their In-Vivo Characterisation Using Balb/C Mice Model. (Unplublished Master's Thesis), Universiti Putra Malaysia, Malaysia.
- Zaman, S. A., & Sarbini, S. R. (2015). The potential of resistant starch as a prebiotic. *Critical reviews in biotechnology*(0), 1-7.
- Zenel, A. M., & Stewart, M. L. (2015). High amylose white rice reduces postprandial glycemic response but not appetite in humans. *Nutrients*, 7(7), 5362-5374.
- Zeng, H., Huang, C., Lin, S., Zheng, M., Chen, C., Zheng, B., & Zhang, Y. (2017). Lotus Seed Resistant Starch Regulates Gut Microbiota and Increases Short-Chain Fatty Acids Production and Mineral Absorption in Mice. *Journal of agricultural and food chemistry*, 65(42), 9217-9225.
- Zhou, J., Martin, R., Tulley, R., Raggio, A., Shen, L., Lissy, E., . . . Keenan, M. (2009). Failure to ferment dietary resistant starch in specific mouse models of obesity results in no body fat loss. *Journal of agricultural and food chemistry*, *57*, 8844 8851.
- Zhou, Z., Cao, X., & Zhou, J. Y. (2013). Effect of resistant starch structure on short-chain fatty acids production by human gut microbiota fermentation in vitro. *Starch-Stärke*, 65(5-6), 509-516.
- Zi-Ni, T., Rosma, A., Karim, A., & Liong, M. (2015). Functional Properties of Resistant Starch Type-III from Metroxylon sagu as Affected by Processing Conditions. *Pertanika Journal of Tropical Agricultural Science*, 38(3).