



***PREPARATION AND CHARACTERISATION OF HYDROGEL FILM AND
NANOFIBER FROM CARBOXYMETHYL-BASED CELLULOSE AND
SAGO PULP FOR CONTROLLED RELEASE APPLICATION***

NAFEESA BINTI MOHD KANAFI

FS 2019 59



**PREPARATION AND CHARACTERISATION OF HYDROGEL FILM AND
NANOFIBER FROM CARBOXYMETHYL-BASED CELLULOSE AND
SAGO PULP FOR CONTROLLED RELEASE APPLICATION**

By

NAFEESA BINTI MOHD KANAFI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirement for the Degree of Master
of Science**

October 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

PREPARATION AND CHARACTERISATION OF HYDROGEL FILM AND NANOFIBER FROM CARBOXYMETHYL-BASED CELLULOSE AND SAGO PULP FOR CONTROLLED RELEASE APPLICATION

By

NAFEESA BINTI MOHD KANAFI

October 2018

Chairman: Norizah Abdul Rahman, PhD
Faculty: Science

Carboxymethyl cellulose (CMC)-based hydrogels show great capability in delivering and occupying small particles like drugs and dyes. CMC-based hydrogel could enhance the capability of a hydrogel and benefit the mankind since it has excellent biocompatibility and biodegradable properties to be use in biomedical application. In this study, a series of CMC and carboxymethyl sago pulp (CMSP) blended with poly(ethylene oxide) (PEO) hydrogel in the form of films and nanofibers were fabricated by using citric acid as a cross-linker. The CMSP used was isolated from sago waste, while CMC was purchased from Fluka Company. CMSP derived from sago waste was studied in the place of CMC because it has similar structure and can help to preserve the environment. For the production of hydrogel nanofibers, the nanofibers were prepared by using electrospinning technique prior cross-linking with citric acid. The electrospinning parameters used were concentration of the polymers blend solution, weight ratio of CMC or CMSP to PEO, applied voltage, tip-to-collector distance and the solution flow rate. The average fiber diameter of the CMSP/PEO nanofibers are from 201 to 300 nm and CMC/PEO from 101 to 200 nm. However, the formation of CMC/PEO nanofibers on the collector was very thin even after several hours of electrospinning, and not able to peel off. Thus, it cannot be further study for fabrication of hydrogel and controlled release. The swelling behaviour of the hydrogels film and nanofibers were optimised based on four parameters; ratio of CMC or CMSP to PEO, percentage of citric acid, temperature and curing time. The results show percentage of swelling and thermal property of CMC/PEO and CMSP/PEO hydrogel was improved compared to CMC and CMSP alone. In controlled release study, methylene blue (MB) was chosen as the model drug due to its

hydrophilic nature. The controlled release results show CMSP/PEO hydrogel nanofibers had the highest percentage of MB loading ($89.20 \pm 0.42\%$) than CMC/PEO and CMSP/PEO hydrogels film. This can be relates with swelling results that show CMSP/PEO hydrogel nanofibers has the highest percentage of swelling ($4366 \pm 975\%$). The MB release study showed that the MB released from CMSP/PEO hydrogel nanofibers was slowly released with pH dependency. The total cumulative percentage release of MB in pH 4.0 (17.04%) and pH 7.34 (19.44%) for CMSP/PEO hydrogel nanofibers are not much different from the CMSP/PEO hydrogel film (pH 4.0 = 14.11% and pH 7.34 = 17.92%), but showed a lower total cumulative percentage release of MB at pH 1.20 (8.91%) and 8.0 (21.21%). The results indicate CMSP/PEO hydrogel nanofibers have a good potential to be used, for example for drug delivery in intestinal area and wound healing.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENYEDIAAN DAN PENCIRIAN HIDROGEL FILEM DAN NANOFIBER
DARIPADA KARBOKSIMETIL BERASASKAN SELULOSA DAN PULPA
SAGU UNTUK APLIKASI KAWALAN PELEPASAN**

Oleh

NAFEESA BINTI MOHD KANAFI

Oktober 2018

Pengerusi: Norizah Abdul Rahman, PhD
Fakulti: Sains

Hidrogel berasaskan CMC menunjukkan keupayaan yang hebat dalam penghantaran dan dipenuhi zarah-zarah kecil seperti dadah dan pewarna. Hidrogel berasaskan CMC boleh meningkatkan keupayaan hidrogel dan memberi faedah kepada manusia memandangkan ia mempunyai ciri-ciri bioserasi dan biodegradasi yang bagus untuk digunakan dalam aplikasi bioperubatan. Dalam kajian ini, satu siri CMC dan karboksimetil pulpa sagu (CMSP) yang dicampurkan dengan poli(etilena oksida) (PEO) dalam bentuk filem-filem hidrogel dan nanofiber direka dengan menggunakan asid sitrik sebagai sambung-silang. CMSP yang digunakan telah diasingkan daripada pulpa sagu, manakala CMC dibeli daripada syarikat Fulka. CMSP yang berasal daripada sisa buangan sagu semulajadi telah dikaji bagi menggantikan tempat CMC kerana ia mempunyai struktur yang sama dan boleh membantu memelihara alam sekitar. Untuk penghasilan nanofiber hidrogel, nanofiber disediakan menggunakan teknik elektrospinning sebelum disambung-silang menggunakan asid sitrik. Parameter-parameter elektrospinning yang digunakan adalah kepekatan cecair campuran polimer, nisbah berat CMC atau CMSP kepada PEO, voltan yang digunakan, jarak dari hujung jarum ke pengumpul dan kadar alir cecair. Purata diameter fiber untuk nanofiber CMSP/PEO adalah daripada 201 hingga 300 nm dan nanofiber CMC/PEO pula adalah daripada 101 hingga 200 nm. Walau bagaimanapun, pembentukan nanofibers CMC/PEO pada pemungutnya sangat nipis walaupun selepas beberapa jam elektrospinning dan sukar untuk dikupas. Jadi, ia tidak boleh diteruskan untuk kajian pelepasan dadah dan pencirian. Tingkah laku bengkak hidrogel filem dan nanofibers dioptimumkan berdasarkan empat parameter, nisbah CMC atau CMSP kepada PEO, peratusan asid sitrik, suhu dan masa pepadatan. Keputusan menunjukkan peratusan pembengkakan dan

sifat terma bagi hidrogel CMC/PEO dan hidrogel CMSP/PEO menunjukkan peningkatan yang baik berbanding CMC dan CMSP sahaja. Dalam kajian pelepasan kawalan, metilena biru (MB) dipilih sebagai model dadah kerana sifat hidrofiliknya. Keputusan pelepasan kawalan menunjukkan nanofiber hidrogel CMSP/PEO mempunyai peratusan tertinggi memuat MB ($89.20 \pm 0.42\%$) berbanding hidrogel CMC/PEO dan hidrogel CMSP/PEO. Ini boleh dikaitkan dengan keputusan pembengkakan yang menunjukkan nanofiber hidrogel CMSP/PEO mempunyai peratusan pembengkakan yang tertinggi ($4366 \pm 975\%$). Kajian pelepasan MB menunjukkan MB yang terlepas daripada nanofiber hidrogel CMSP/PEO adalah merupakan pelepasan yang perlahan dengan kebergantungan kepada pH. Jumlah peratusan kumulatif MB dalam pH 4.0 (17.04%) dan pH 7.34 (19.44%) untuk nanofiber hidrogel CMSP/PEO agak sama dengan hidrogel CMSP/PEO (pH 4.0 = 14.11% and pH 7.34 = 17.92%) tetapi menunjukkan jumlah peratusan kumulatif pelepasan MB yang lebih rendah pada pH 1.20 (8.91%) dan 8.0 (21.21%). Hasil keutusan ini menunjukkan bahawa nanofiber hidrogel CMSP/PEO mempunyai potensi yang baik untuk digunakan dalam penghantaran dadah di kawasan usus dan penyembuhan luka.

ACKNOWLEDGEMENT

Alhamdulillah, all praise and thanks to Allah, I finally completed my master's research. Due to that, I am grateful to all those who supported me in one or another way during this journey of my master.

I would like to express my great appreciation to my supervisor Dr. Norizah Abd. Rahman for her support, guidance, inspiring knowledge, and giving me the opportunity to do my master thesis inside her division. I would also like to express my uncountable thanks to Prof. Dr. Mohd Zobbir Hussein and Prof. Dr. Mansor Ahmad@Ayob for assisting and guiding me during this project.

I would also want to thank Puan Norhashidah binti Talip from Malaysian Nuclear Agency, Bangi for providing me the sago waste and guidance. Furthermore, special thanks to all lecturers, staff, laboratory assistants and postgrad students of Chemistry Department, Faculty of Science, UPM, for their assistance and tolerance.

I wish to express my gratitude to each of my friends, Husna, Aida, Khalilah, Fatimah, Zurmira, Miza, Ain and my labmates for providing me with advice, companion, useful information and helping me throughout my master. I wish to extend my deepest gratitude to my family especially my beloved mother, Roshana binti Othman for supporting, encouraging and keep motivating me throughout this project.

Finally, I cannot forget all the people who I met during these years, thanks for everybody. May Allah reward you with goodness.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Norizah binti Abdul Rahman, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Mansor bin Haji Ahmad@Ayob, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Mohd Zobir bin Hussein, PhD

Professor
Institut Teknologi Maju
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: _____

Signature: _____
Name of Member of
Supervisory
Committee: _____

Signature: _____
Name of Member of
Supervisory
Committee: _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENT	v
APPROVAL	vi
DECLARATION	viii
LIST OF FIGURES	xiii
LIST OF TABLES	xv
LIST ABBREVIATIONS	xvii
CHAPTER	
INTRODUCTION	
1.1 Background of study	1
1.2 Problem statement	3
1.3 Hypothesis and significance of study	4
1.4 Objectives	5
2 LITERATURE REVIEW	
2.1 Carboxymethyl cellulose(CMC)	6
2.2 Carboxymethyl sago pulp (CMSP)	6
2.3 Poly(ethylene) oxide (PEO)	9
2.4 Polymer nanofibers	10
2.5 Electrospinning	11
2.6 Hydrogel	12
2.7 Cross-linking	13
2.8 Controlled release application	15
3 METHODOLOGY	
3.1 Materials and reagents	16
3.2 Isolation of sago pulp from sago waste	16
3.3 Preparation of carboxymethyl sago pulp (CMSP)	17
3.4 Determination of degree of substitution	18
3.4.1 Potentiometric back titration	18
3.5 Preparation of carboxymethyl cellulose (CMC) hydrogel	19
3.5.1 Optimization of CMC/PEO composite hydrogel	19
3.5.2 Determination of gel fraction of optimized CMC/PEO hydrogel	21
3.6 Preparation of carboxymethyl sago pulp (CMSP) hydrogel	21
3.6.1 Optimization of CMSP/PEO compositehydrogel	21
3.7 Polymer nanofibers	22

3.7.1	Optimization electrospinning parameters of CMC/PEO nanofibers	22
3.7.2	Optimization electrospinning parameters of CMSP/PEO nanofibers	23
3.7.3	Optimisation of CMSP/PEO hydrogel nanofibers	24
3.8	Calibration standards	25
3.9	Drug loading and controlled release studies	25
3.10	Characterization	26
3.10.1	Fourier transform infrared (FTIR) spectroscopy	26
3.10.2	Thermogravimetric analysis (TGA) spectroscopy	27
3.10.3	X-ray diffraction (XRD) analysis	27
3.10.4	Ultraviolet-visible (UV-Vis) spectrophotometer	27
3.10.5	Scanning electron microscopy (SEM)	27
4	RESULT AND DISCUSSION	
4.1	Preparation and characterization of carboxymethyl sago pulp (CMSP)	28
4.1.1	Percentage yield of sago pulp and CMSP	28
4.1.2	Spectroscopic analysis of sago pulp, CMSP and CMC	29
4.1.3	Thermal analysis of sago pulp, CMSP and CMC	32
4.2	Optimization of CMC/PEO hydrogel film	34
4.2.1	Swelling studies	34
4.2.2	Gel fraction study	42
4.3	Characterization of CMC/PEO hydrogel	42
4.3.1	Fourier transform infrared (FTIR) spectroscopy	42
4.3.2	Thermogravimetric analysis	45
4.3.3	X-ray powder diffraction spectroscopy	48
4.3.4	Drug loading efficiency	49
4.3.5	Drug release studies	50
4.3.6	Morphology studies	52
4.4	Optimization of CMC/PEO nanofibers	55
4.4.1	Solution parameters	55
4.4.2	Processing parameters	63
4.5	Optimization of CMSP/PEO hydrogel film	74
4.5.1	Swelling studies	74
4.5.2	Gel fraction study	78
4.6	Characterization of CMSP/PEO film	79
4.6.1	Fourier transform infrared (FTIR) spectroscopy	79
4.6.2	Thermogravimetric analysis	81

4.6.3	X-ray powder diffraction spectroscopy	83
4.6.4	Drug loading efficiency	84
4.6.5	Drug release studies	85
4.6.6	Morphology studies	86
4.7	Optimization of CMSP/PEO nanofibers	88
4.7.1	Solution parameters	88
4.7.2	Processing parameters	98
4.7.3	Swelling studies	102
4.7.4	Gel fraction study	104
4.8	Characterization of CMSP/PEO nanofibers	105
4.8.1	Drug loading efficiency	105
4.8.2	Drug release studies	107
4.8.3	Morphology studies	108
5	CONCLUSIONS	110
	REFERENCES	
	APPENDICES	
	BIODATA OF STUDENT	
	LIST OF PUBLICATIONS	

LIST OF FIGURES

Figures	Page
1.1 Sago waste	2
1.2 Sago pulp powder (delignified sago waste)	2
2.1 Etherification of cellulose to carboxymethyl cellulose with sodium monochloroacetic acid in alkaline medium (V Pushpamalar, Langford, Ahmad, & Lim, 2006)	7
2.2 Poly(ethylene oxide) structure	9
2.3 Electrospinning process set up (retrieved from http://substance-en.etsmtl.ca/polymer-fibers-electrospinning/)	12
2.4 Citric acid structure	14
3.1 Sago pulp isolation from sago waste	17
3.2 Dry CMC/PEO hydrogel became swollen after 24 hours immersed in water	20
4.1 a) Sago waste b) Sago pulp c) CMSP d) CMC	29
4.2 FTIR spectra pattern of sago pulp and carboxymethyl sago pulp (CMSP)	30
4.3 IR spectra comparison of CMSP and commercialized CMC	31
4.4 a) Weight loss (%) and b) DTG curve of sago waste, sago pulp, CMSP and CMC	33
4.5 Proposed mechanism of cross-linking reaction between cellulose (CMC) and citric acid	35
4.6 (a) Dry cross-linked CMC/PEO (3:1) film; (b) Swollen CMC/PEO (3:1) hydrogel film	36
4.7 Effects of various parameters; a) ratio of CMC to PEO, b) percentage of citric acid, c) curing temperature, and d) curing time on the percentage of swelling of CMC/PEO hydrogel film	41
4.8 FTIR spectra of non-cross-linked (3:1) (a) and cross-linked (3:1) (b) CMC/PEO films	43
4.9 Effects of different ratio of CMC to PEO	45
4.10 a) Weight loss (%) and b) DTG curve of CMC, PEO, non-cross-linked CMC/PEO and cross-linked CMC/PEO	47
4.11 XRD patterns of PEO, CMC and cross-linked CMC/PEO (3:1) film	49
4.12 Effects of pH value on the release of MB from CMC/PEO hydrogel film	51
4.13 SEM images for surface morphology of a) and b) dry CMC-PEO hydrogel; c) and d) swollen CMC/PEO hydrogel; e) and f) drug-loaded CMC/PEO hydrogel	54
4.14 SEM images of cross-section of a) and b) swollen CMC/PEO hydrogel; c) and d) MB-loaded CMC/PEO hydrogel	54

4.15	Effects of various parameters; a) percentage of citric acid, b) ratio of CMSP to PEO, c) curing temperature, and d) curing time on the percentage of swelling of CMSP/PEO hydrogel film	78
4.16	FTIR spectra of a) non-cross-linked and b) cross-linked (3:1) CMSP/PEO films	80
4.17	a) Weight loss (%) and b) DTG curve of CMSP, PEO, non-cross-linked CMSP/PEO and cross-linked CMSP/PEO	82
4.18	XRD patterns of PEO, CMSP and cross-linked CMSP/PEO film	84
4.19	Effects of pH values on the release of MB from CMSP/PEO hydrogel film	86
4.20	SEM images of surface morphology of a) dry CMSP-PEO hydrogel; b) swollen CMSP/PEO hydrogel; c) MB-loaded CMSP/PEO hydrogel	87
4.21	SEM images of cross-section of (a) swollen CMSP/PEO (3:1) hydrogel; (b) and (c) MB-loaded CMSP/PEO (3:1) hydrogel	88
4.22	Effects of various parameters; a) percentage of citric acid, b) curing temperature on the percentage of swelling of CMSP/PEO hydrogel nanofibers	104
4.23	Proposed interaction of MB with polymer	106
4.24	Methylene blue loading efficiency at different types of hydrogels	107
4.25	Effects of pH value on the release of MB from CMSP/PEO hydrogel nanofibers	108
4.26	SEM images of a) and b) swollen CMSP/PEO hydrogel nanofibers; c) and d) MB-loaded CMSP/PEO hydrogel nanofibers	109

LIST OF TABLES

Tables		Page
3.1	List of chemicals	16
3.2	Optimisation parameters of CMC/PEO hydrogel film	20
3.3	Optimisation parameters of CMSP/PEO hydrogel film	22
3.4	Optimisation parameters of CMC/PEO nanofibers	23
3.5	Optimisation parameters of CMSP/PEO nanofibers	24
3.6	Optimisation parameters of CMSP/PEO hydrogel nanofibers	24
4.1	The absorption bands of pre-treated sago pulp and CMSP and their assignment	30
4.2	The absorption bands of CMSP and commercialized CMC and their assignment	32
4.3	Thermal analysis of sago waste, sago pulp, CMSP and CMC	34
4.4	The absorption bands of non-crosslinked (3:1) and cross-linked (3:1) CMC/PEO films	44
4.5	Thermal analysis of CMC, PEO, non-crosslinked CMC/PEO and cross-linked CMC/PEO films	48
4.6	Percentage of MB loaded into the hydrogel film at different initial concentrations of MB	50
4.7	SEM images and distribution graphs for CMC/PEO nanofibers at different concentrations of CMC/PEO solution	57
4.8	SEM images and distribution graphs of for CMC/PEO nanofibers at different weight ratios of CMC to PEO	60
4.9	SEM images and distribution graphs of CMC/PEO nanofibers when different high voltage been applied	64
4.10	SEM images and distribution graphs of CMC/PEO nanofibers at different tip-to-collector distances	67
4.11	SEM images and distribution graphs of CMC/PEO nanofibers at different flow rate	71
4.12	Comparison of gel fraction between CMC/PEO and CMSP/PEO hydrogel films	78
4.13	The absorption bands of non-cross-linked (3:1) and cross-linked (3:1) CMC/PEO films	80
4.14	Thermal analysis of CMSP, PEO, non-cross-linked CMSP/PEO and cross-linked CMSP/PEO hydrogel films	83
4.15	Percentage of MB loaded into the hydrogel film at different initial concentrations of MB	84
4.16	SEM images and distribution graphs for CMSP/PEO nanofibers at different weight ratios of CMSP to PEO	90
4.17	SEM images and distribution graphs for CMSP/PEO nanofibers at different concentrations of CMSP/PEO	93

	solution	
4.18	SEM images and distribution graphs of CMSP/PEO nanofibers at different flow rates	99
4.19	Comparison of gel fraction between CMC/PEO and CMSP/PEO hydrogel films with CMSP/PEO nanofibers	105
4.20	Percentage of MB loaded into the hydrogel nanofibers at different initial concentrations of MB	106



LIST OF ABBREVIATIONS

CA	Citric acid
CMC	Carboxymethyl cellulose
CMSP	Carboxymethyl sago pulp
CMSS	Carboxymethyl sago starch
CONH	2° amide
CONH ₂	1° amide
DSC	Differential scanning calorimetry
ECM	Extracellular matrix
ECH	Epichlorohydrin
FTIR	Fourier transform infrared
GI	Gastrointestinal
GPC	Gel permeation chromatography
HPMC	Hydroxypropyl methylcellulose
IVD	Intervertebral disc
MB	Methylene blue
NaClO ₂	Sodium chlorite
PAA	Poly(acrylic acid)
PBS	Phosphate-buffered saline
PEO	Poly(ethylene) oxide
PVA	Poly(vinyl alcohol)
Q	Percentage of swelling
SEM	Scanning electron microscopy

SO ₃ H	Sulfonic acid
TGA	Thermogravimetric analysis
US FDA	United State Food and Drug Administration
UV-Vis	Ultraviolet-visible
XRD	X-ray diffraction



CHAPTER 1

INTRODUCTION

1.1 Background of study

In Sarawak, Malaysia, the largest sago-growing areas, sago palms (*Metroxylon sago*) are found in tropical lowland forest and swampy areas in which estimated 54,000 hectares in 2013 by Sarawak agriculture statistics (Bujang & Hassan, 2013; V Pushpamalar et al., 2006). The main product of sago palm, sago starch was exported mainly to Peninsular Malaysia, Japan, Singapore and other countries with total about 48,000 tons in 2013 (Sarawak agriculture statistics, 2013). To isolate sago starch, several mechanical processes involving debarking, rasping, sieving, settling, washing and drying need to go through and at the end of the process, starchy fibrous by-product, sago waste was produced at approximately 7 tons daily from a single processing mill.

Sago waste is light brown in color and still maintains the woody structure as shown in Figure 1.1. It is a lignocellulosic biomass that consist of cellulose, hemicellulose and lignin (Bujang & Hassan, 2013; Veeramachineni et al., 2016). Cellulose is a long and linear polysaccharide polymer which consist of many glucose units that linked to each other by beta 1,4-glycosidic bonds. It has received tremendous attention from researchers nowadays and had been focuses on cellulose derivatives, such as carboxymethylcellulose (Pushpamalar et al., 2006), hydroxypropyl methylcellulose (HPMC) and methylcellulose (Frenot et al., 2006). Since isolation of sago starch requires large amount of water, thus, the residues are mixed with wastewater and easily washed off into nearby streams without proper treatment or deposited in the factory's compound (Bujang & Hassan, 2013). These actions will lead to serious environmental problems in future. To prevent this circumstance, sago waste can become a very economical source of cellulose to the industries due to cheapest, biodegradable and availability of all renewable natural polymers existing in Malaysia. Sago fiber is used to provide bulk for rumen fermentation, sago pith used as animal feed stuff and in livestock industry and sago frond used in pulp and paper industry (Chew et al., 1999). However, the utilization of sago waste should be further explored. Investigating the potential application CMC nanofibers from sago waste drug delivery in this research will not open a new potential application sago waste but also will help to recycle and solve disposal problem of sago waste.



Figure 1.1: Sago waste

Sago pulp is the result of purification of sago waste that is white in color as shown in Figure 1.2. It was reported that 57% w/w sago pulp was successfully isolated from sago waste (V Pushpamalar et al., 2006). The pre-product, sago pulp that consist of cellulose is then been transformed into functionalised cellulose which can be used in various applications such as pharmaceutical excipient and industrial products (V Pushpamalar et al., 2006; Veeramachineni et al., 2016). The purpose of purification of sago waste into sago pulp is to remove the lignin (Veeramachineni et al., 2016).

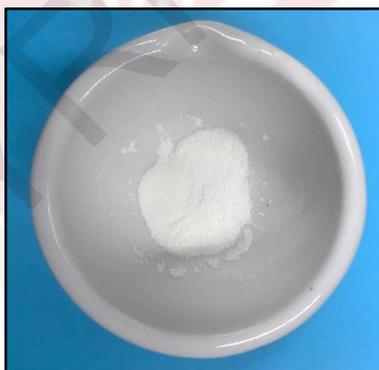


Figure 1.2: Sago pulp powder (delignified sago waste)

Hydrogel is a smart three-dimensional polymeric network structure that capable to hold a lot of water or biological fluids beyond its dry weight without dissolution in water prepared by either chemical or physical cross-linking method (Shen et al., 2016; Tan et al., 2016). Holding capacity of hydrogel is depends on the degree of cross-linking. Hydrophilic groups such as, -OH, -CONH, -CONH₂, and -SO₃H that present in the formulation polymers of hydrogel structure are the reason of the ability of hydrogel to absorb water (Khan & Ranjha, 2014).

The water uptake and release occur when the protonable groups respond to external stimuli such as change in pH, ionic strength, or temperature (Barbucci et al., 2000). The swollen state of hydrogel has a soft, flexible and tissue-like physical properties make it applicable in most biomedical applications. Moreover, hydrogel is sensitive to environment stimuli when there are changes such as pH, temperature, presence of enzyme and glucose (Patel & Mequanint, 2009). To apply hydrogel in drug delivery application, non-toxic and biocompatible materials should be the choices in making the formulation of hydrogel as well as new way to improve the properties of hydrogel in drug delivery application.

Nowadays, widespread of knowledge on the techniques of manufacturing nanomaterial had lead researchers around the world to focus on the preparation of nanomaterial for various applications including biomedical (Haider, Haider, & Kang, 2015a). The focus on nanomaterials specifically nanofibers is because of the special characteristics such as low density, large surface area, high pore volume and tight pore size (Rathinamoorthy et al., 2012). Electrospinning, a highly versatile method is one of the techniques for the fabrication of nanofibers. In biomedical application, electrospun nanofibers have been widely studied for drug and therapeutic agent delivery, wound dressings and tissue engineering using natural or synthetic polymers (Abrigo et al., 2014; Haider, Haider, & Kang, 2015b; Teck, 2017).

1.2 Problem statement

Abundance of sago waste produced daily and cellulose is second major components in sago palm (V Pushpamalar et al., 2006). During isolation of sago starch, sago waste was easily washed off into nearby streams or deposited in the factory's compound (Bujang & Hassan, 2013). These actions can lead to serious environmental problems in future. Sago waste consists of biopolymers such as lignin, cellulose and polysaccharide that can be used for many applications. Cellulose is second major components in sago waste (V Pushpamalar et al., 2006). Apart from that, in the preparation of biopolymer hydrogel, epichlorohydrin was commonly used as cross-linker, however, it can caused carcinogenic by-products and can harm to human. Mostly in electrospinning process, toxic organic solvents are used as the solvent that might leave trace in the nanofibers produced and caused toxicity. In making hydrogel for biomedical application, all carcinogenic by-products and toxic materials need to avoid to ensure only safe materials can enter the human body.

1.3 Hypothesis and significance of study

Natural polymers underwent a re-evaluation as a result of natural biodegradable properties and availability from renewable resources compared to synthetic alternatives. Thus, utilizing the cellulose from sago waste (also known as sago pulp) into valuable product such as carboxymethyl sago pulp (CMSP) could help in preserving the environment as well as benefit the mankind. CMSP also are more cost effective since there are abundance of sago waste available. Moreover, CMSP is a biopolymer that safe to be used for human and suit the requirement in biomedical applications that demanding for non-toxic materials. In this study, CMSP was used to study for drug delivery application in hydrogel form with citric acid as the cross-linker. Citric acid was chosen to replace of commonly used toxic cross-linker such as epichlorohydrin due to its non-toxic property and no toxicity produced even after cross-linking reaction. Recently, the use of nanofibers loaded with drugs for biomedical application has awoken much interest and there were no studies on CMSP-based hydrogel in the form of nanofibers yet. Previous study by Pushpamalar et. al, had shown CMSP hydrogel has good swelling property at different pH media (Vengidesh Pushpamalar et al., 2013). However, to the best of our knowledge, there is no study conducted on the CMSP nanofibers yet. Therefore, CMSP will be electrospun into nanofibers in order to increase the surface area the hydrogel and could improve the delivering and encapsulating of drugs. CMSP will be electrospun into nanofibers by using electrospinning technique. In this study, water was used as a solvent for electrospinning process, which is an ideal solution in biomedical application despite all the toxic organic solvents.

1.4 Objectives

Various studies have been done focusing on economical, safe, and biocompatible materials in building drug delivery system. To accomplish similar aims, a renewable natural polymer from waste (sago pulp) was used in this study in order to open a new potential application sago waste but also will help to recycle and solve disposal problem of sago waste. The objectives of this study are:

- i. To isolate sago pulp and prepare CMSP by using carboxymethylation reaction
- ii. To prepare and optimise cross-linked CMC/PEO and CMSP/PEO films and nanofibers
- iii. To investigate and evaluate the control release of MB for the hydrogel films and nanofibers

REFERENCES

- Abrigo, M., McArthur, S. L., & Kingshott, P. (2014). Electrospun Nanofibers as Dressings for Chronic Wound Care : Advances , Challenges , and Future Prospects, 772-792.
- Adam, P., Sasikanth, K., Nama, S., Suresh, S., & Brahmaiah, B. (2013). Nanofibers - A New Trend In Nano Drug Delivery, 2(2), 118-127.
- Ahmad, B., Abbas, S., Iqbal, Z., Bashir, S., & Ali, J. (2013). Synthesis of Cross Linked PVP Hydrogels and its Use for the Control Release of Anti-Asthmatic Drugs, 14(2), 273-283.
- Akkaya, R., & Ulusoy, U. (2011). Research Article Recep Akkaya* and Ulvi Ulusoy, 39(4), 359-370.
- Alemdar, A., & Sain, M. (2008). Isolation and characterization of nanofibers from agricultural residues - Wheat straw and soy hulls, 99, 1664-1671.
- Arslan, N. (2003). Production of carboxymethyl cellulose from sugar beet pulp cellulose and rheological behaviour of carboxymethyl cellulose, 54, 73-82.
- Azeredo, H. M. C., Kontou-vrettou, C., Moates, G. K., Wellner, N., Cross, K., Pereira, P. H. F., & Waldron, K. W. (2015). Food Hydrocolloids Wheat straw hemicellulose films as affected by citric acid. *Food Hydrocolloids*, 50, 1-6.
- Barbucci, R., Magnani, A., & Consumi, M. (2000). Swelling Behavior of Carboxymethylcellulose Hydrogels in Relation to Cross-Linking , pH , and Charge Density, 7475-7480.
- Bhattarai, N., Gunn, J., & Zhang, M. (2010). Chitosan-based hydrogels for controlled , localized drug delivery ☆. *Advanced Drug Delivery Reviews*, 62(1), 83-99.
- Biswas, A., Kim, S., Selling, G. W., & Cheng, H. N. (2014). Conversion of agricultural residues to carboxymethylcellulose and carboxymethylcellulose acetate. *Industrial Crops & Products*, 60, 259-265.
- Bolio-lópez, G. I., Ross-alcudia, R. E., Veleva, L., Azamar, J. A., Madrigal, G. C., Hernández-villegas, M. M., Bolio-lópez, G. I. (2016). Extraction and Characterization of Cellulose from Agroindustrial Waste of Pineapple (*Ananas comosus* L . Merrill) Crowns, (January).
- Bono, A., Ying, P. H., Yan, F. Y., Muei, C. L., Sarbatly, R., & Krishnaiah, D. (2009). ORIGINAL ARTICLE Synthesis and Characterization of Carboxymethyl Cellulose from Palm Kernel Cake, 3(2073), 5-11.
- Boverhof, D. R., Bramante, C. M., Butala, J. H., Clancy, S. F., Lafranconi, M., West, J., & Gordon, S. C. (2015). Comparative assessment of nanomaterial definitions and safety evaluation considerations. *Regulatory Toxicology and Pharmacology*, 73(1), 137-150.

- Bujang, K. B., & Hassan, M. A. (2013). Recovery of Glucose from Residual Starch of Sago Hampas for Bioethanol Production, 2013.
- Buwalda, S. J., Vermonden, T., & Hennink, W. E. (2017). Hydrogels for Therapeutic Delivery : Current Developments and Future Directions.
- Cassani, D. A. D., Altomare, L., Nardo, D., & Variola, F. (2015). investigation of electrodeposited chitosan : PEO blends. *Journal of Materials Chemistry B: Materials for Biology and Medicine*, 3, 2641–2650.
- Castro, D., Evangelista, R. C., Carbinatto, F. M., Do, A., & Cury, B. S. F. (2013). ScienceDirect Insights into the swelling process and drug release mechanisms from cross-linked pectin / high amylose starch matrices, 9, 3–10.
- Chandrasekaran, A. R., Jia, C. Y., Theng, C. S., Muniandy, T., Muralidharan, S., & Arumugam, S. (2011). Invitro studies and evaluation of metformin marketed tablets-Malaysia, 01(05), 214–217.
- Chang, C., Duan, B., Cai, J., & Zhang, L. (2010). Superabsorbent hydrogels based on cellulose for smart swelling and controllable delivery. *European Polymer Journal*, 46(1), 92–100.
- Chen, W., Li, D., Ei-shanshory, A., El-newehy, M., Ei-hamshary, H. A., Al-deyab, S. S., ... Mo, X. (2015). Colloids and Surfaces B : Biointerfaces Dexamethasone loaded core - shell SF / PEO nanofibers via green electrospinning reduced endothelial cells inflammatory damage. *Colloids and Surfaces B: Biointerfaces*, 126, 561–568.
- Cheng, H. N., & Biswas, A. (2011). Chemical modification of cotton-based natural materials : Products from. *Carbohydrate Polymers*, 84(3), 1004–1010.
- Dahlan, N. A., Ng, S. L., & Pushpamalar, J. (2017). Adsorption of methylene blue onto powdered activated carbon immobilized in a carboxymethyl sago pulp hydrogel, 44271, 1–11.
- Das, N. (2013). PREPARATION METHODS AND PROPERTIES OF HYDROGEL : A REVIEW, 5(3).
- Demirci, S., Celebioglu, A., & Uyar, T. (2014). Polymer Chemistry release properties, 2050–2056.
- El-din, H. M. N., El-naggar, A. W. M., Fadle, F. I. A., Nizam, H. M., El-naggar, A. W. M., Abu-el, F. I., Fadle, F. I. A. (2013). Radiation Synthesis of pH-Sensitive Hydrogels From Carboxymethyl Cellulose / Poly (ethylene Oxide) Blends as Drug Delivery Systems Radiation Synthesis of pH-Sensitive Hydrogels From Carboxymethyl Cellulose / Poly (ethylene Oxide) Blends as Drug Delivery Systems, 4037(March 2016).
- El- Hag Ali, A., Abd El- Rehim, H. A., Kamal, H., & Hegazy, D. E. A. (2008). Synthesis of Carboxymethyl Cellulose Based Drug Carrier Hydrogel Using Ionizing Radiation for Possible Use as Site Specific Delivery System. *Journal of Macromolecular Science, Part A*, 45(8), 628–634.

- Elanthikkal, S., Gopalakrishnanapanicker, U., Varghese, S., & Guthrie, J. T. (2010). Cellulose microfibrils produced from banana plant wastes : Isolation and characterization. *Carbohydrate Polymers*, 80(3), 852–859.
- Elsabee, M. Z., Naguib, H. F., & Morsi, R. E. (2012). Chitosan based nanofibers, review. *Materials Science and Engineering: C*, 32(7), 1711–1726.
- Fekete, T., Borsa, J., Takács, E., & Wojnárovits, L. (2015). Synthesis of cellulose-based superabsorbent hydrogels by high-energy irradiation in the presence of crosslinking agent. *Radiation Physics and Chemistry*, 1–6.
- Fernandes, J. G., Correia, D. M., Botelho, G., Padrão, J., & Dourado, F. (2014). PHB-PEO electrospun fiber membranes containing chlorhexidine for drug delivery applications. *Polymer Testing*, 34, 64–71.
- Foroutan, H., Khodabakhsh, M., & Rabbani, M. (2007). Investigation of synthesis of PVP hydrogel by irradiation. *Iranian Journal of Radiation Research*, 5(3), 131–136.
- Frenot, A., Henriksson, M. W., & Walkenstro, P. (2006). Electrospinning of Cellulose-Based Nanofibers.
- Gainza, G., Villullas, S., Pedraz, J. L., Hernandez, R. M., & Igartua, M. (2015). Advances in drug delivery systems (DDSs) to release growth factors for wound healing and skin regeneration. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 11(6), 1551–1573.
- George, M., & Abraham, T. E. (2007). pH sensitive alginate – guar gum hydrogel for the controlled delivery of protein drugs, 335, 123–129.
- Giani, G., Fedi, S., Barbucci, R., Interuniversitario, C., Medici, S., & Viale, E. (2012). Hybrid Magnetic Hydrogel: A Potential System for Controlled Drug Delivery by Means of Alternating Magnetic Fields, 1157–1169.
- Goyal, R., Macri, L. K., Kaplan, H. M., & Kohn, J. (2015). Nanoparticles and nanofibers for topical drug delivery. *Journal of Controlled Release*.
- Greiner, A., & Wendorff, J. H. (2007). Electrospinning : A Fascinating Method for the Preparation of Ultrathin Fibers *Angewandte*, 5670–5703.
- Guo-en, S. U. N., Hong, T., Chun-ling, Z., Yan-li, D. O. U., & Yi, L. I. (2010). Preparation of Ultrafine Water-soluble Polymers Nanofiber Mats via Electrospinning. *Chemical Research*, 26(2), 318–322.
- Gupta, B., & Agarwal, R. (2014). Antimicrobial and release study of drug loaded PVA / PEO / CMC wound dressings, 1613–1622.
- Gupta, N. V., & Shivakumar, H. G. (2012). Investigation of Swelling Behavior and Mechanical Properties of a pH-Sensitive Superporous Hydrogel Composite, 11(May 2011), 481–493.
- Haider, A., Haider, S., & Kang, I. (2015a). REVIEW A comprehensive review summarizing the effect of electrospinning parameters and potential applications of nanofibers in biomedical and biotechnology. *ARABIAN*

- Haider, A., Haider, S., & Kang, I. (2015b). REVIEW A comprehensive review summarizing the effect of electrospinning parameters and potential applications of nanofibers in biomedical and biotechnology.
- Hashem, M., Sharaf, S., Abd El-Hady, M. M., & Hebeish, a. (2013). Synthesis and characterization of novel carboxymethylcellulose hydrogels and carboxymethylcellulose-hydrogel-ZnO-nanocomposites. *Carbohydrate Polymers*, 95(1), 421-427.
- Hu, W., & Yu, H. (2013). Coelectrospinning of chitosan / alginate fibers by dual-jet system for modulating material surfaces. *Carbohydrate Polymers*, 95(2), 716-727.
- Huang, W. F., Tsui, G. C. P., Tang, C. Y., & Yang, M. (2017). Optimization Strategy for Encapsulation Efficiency and Size of Drug Loaded Silica Xerogel / Polymer Core-Shell Composite Nanoparticles Prepared by Gelation-Emulsion Method, 1-10.
- Huang, Z., Zhang, Y., Kotaki, M., & Ramakrishna, S. (2003). A review on polymer nanofibers by electrospinning and their applications in nanocomposites, 63, 2223-2253.
- Jayakumar, R., Prabakaran, M., Nair, S. V., & Tamura, H. (2010). Novel chitin and chitosan nano fibers in biomedical applications. *Biotechnology Advances*, 28(1), 142-150.
- Kai, D., Liow, S. S., & Loh, X. J. (2014). Biodegradable polymers for electrospinning: Towards biomedical applications. *Materials Science and Engineering: C*, 45, 659-670.
- Khan, S., & Ranjha, N. M. (2014). Effect of degree of cross-linking on swelling and on drug release of low viscous chitosan / poly, 2133-2158.
- Kono, H. (2014). Characterization and properties of carboxymethyl cellulose hydrogels crosslinked by polyethylene glycol. *Carbohydrate Polymers*, 106, 84-93.
- Konwarh, R., Karak, N., & Misra, M. (2013). Electrospun cellulose acetate nano fibers : The present status and gamut of biotechnological applications, 31, 421-437.
- Kumar, S. U., Matai, I., Dubey, P., Bhushan, B., & Sachdev, A. (2014). Supporting information Differentially cross-linkable core-shell nanofibers for tunable delivery of anticancer drugs : Synthesis , characterization and its anticancer efficacy, 1-11.
- Lam, Y. L., Muniyandy, S., Kamaruddin, H., Mansor, A., & Janarthanan, P. (2015). Radiation cross-linked carboxymethyl sago pulp hydrogels loaded with ciprofloxacin: Influence of irradiation on gel fraction, entrapped drug and in vitro release. *Radiation Physics and Chemistry*, 106, 213-222.

- Lee, H. J., Park, Y. H., & Koh, W.-G. (2013). Fabrication of Nanofiber Microarchitectures Localized within Hydrogel Microparticles and Their Application to Protein Delivery and Cell Encapsulation. *Advanced Functional Materials*, 23(5), 591–597.
- Lee, W., & Chen, Y. (2001). Studies on Preparation and Swelling Properties of the N- Isopropylacrylamide / Chitosan Semi-IPN and IPN Hydrogels, (August 2000), 27–29.
- Li, B. D., & Xia, Y. (2004). Electrospinning of Nanofibers : Reinventing the Wheel ?, (14), 1151–1170.
- Li, W., Sun, B., & Wu, P. (2009). Study on hydrogen bonds of carboxymethyl cellulose sodium film with two-dimensional correlation infrared spectroscopy. *Carbohydrate Polymers*, 78(3), 454–461.
- Loh, X. J., Peh, P., Liao, S., Sng, C., & Li, J. (2010). Controlled drug release from biodegradable thermoresponsive physical hydrogel nanofibers. *Journal of Controlled Release*, 143(2), 175–182.
- Lu, P., & Hsieh, Y.-L. (2009). Cellulose nanocrystal-filled poly(acrylic acid) nanocomposite fibrous membranes. *Nanotechnology*, 20(41), 415604.
- Maitra, J., & Shukla, V. K. (2014). Cross-linking in Hydrogels - A Review, 4(2), 25–31.
- Megelski, S., Stephens, J. S., Chase, D. B., & Rabolt, J. F. (2002). Micro- and Nanostructured Surface Morphology on Electrospun Polymer Fibers, 8456–8466.
- Menzel, C., Olsson, E., Plivelic, T. S., Andersson, R., Johansson, C., Kuktaite, R., ... Koch, K. (2013). Molecular structure of citric acid cross-linked starch films. *Carbohydrate Polymers*, 96(1), 270–276.
- Mohsin, M., Farooq, U., Raza, Z. A., Ahsan, M., Afzal, A., & Nazir, A. (2014). Performance enhancement of wool fabric with environmentally-friendly bio-cross-linker. *Journal of Cleaner Production*, 68, 130–134.
- Muniyandy, S., Sathasivam, T., Veeramachineni, A. K., & Janarthanan, P. (2015). Dual Cross-Linked Carboxymethyl Sago Pulp-Gelatine Complex Coacervates for Sustained Drug Delivery, 1088–1105.
- Nista, S. V. G., Bettini, J., & Mei, L. H. I. (2015). Coaxial nanofibers of chitosan-alginate-PEO polycomplex obtained by electrospinning. *Carbohydrate Polymers*, 127, 222–228.
- Nitanan, T., Akkaramongkolporn, P., Rojanarata, T., Ngawhirunpat, T., & Opanasopit, P. (2013). Neomycin-loaded poly (styrene sulfonic acid-co-maleic acid) (PSSA-MA) / polyvinyl alcohol (PVA) ion exchange nanofibers for wound dressing materials. *International Journal of Pharmaceutics*, 448(1), 71–78.
- Olsson, E., Menzel, C., Johansson, C., Andersson, R., Koch, K., & Järnström, L.

- (2013). The effect of pH on hydrolysis, cross-linking and barrier properties of starch barriers containing citric acid. *Carbohydrate Polymers*, 98(2), 1505–1513.
- Patel, A., & Mequanint, K. (2009). Hydrogel Biomaterials.
- Percival, S. L., Mccarty, S., Hunt, J. A., & Woods, E. J. (2014). The effects of pH on wound healing , biofilms , and antimicrobial efficacy.
- Pillay, V., Dott, C., Choonara, Y. E., Tyagi, C., Tomar, L., Kumar, P., Ndesendo, V. M. K. (2013). A Review of the Effect of Processing Variables on the Fabrication of Electrospun Nanofibers for Drug Delivery Applications, 2013.
- Pushpamalar, V., Langford, S. J., Ahmad, M., & Hashim, K. (2013). Preparation of Carboxymethyl Sago Pulp Hydrogel from Sago Waste by Electron Beam Irradiation and Swelling Behavior in Water and Various pH Media, 451–459.
- Pushpamalar, V., Langford, S. J., Ahmad, M., & Lim, Y. Y. (2006). Optimization of reaction conditions for preparing carboxymethyl cellulose from sago waste, 64, 312–318.
- Rachtanapun, P., Luangkamin, S., Tanprasert, K., & Suriyatem, R. (2012). LWT - Food Science and Technology Carboxymethyl cellulose film from durian rind. *YFSTL*, 48(1), 52–58.
- Rangelova, N., Aleksandrov, L., Angelova, T., Georgieva, N., & Müller, R. (2014). Preparation and characterization of SiO₂/CMC/Ag hybrids with antibacterial properties. *Carbohydrate Polymers*, 101, 1166–1175.
- Ranjbar-mohammadi, M., Zamani, M., Prabhakaran, M. P., Bahrami, S. H., & Ramakrishna, S. (2016). Electrospinning of PLGA / gum tragacanth nanofibers containing tetracycline hydrochloride for periodontal regeneration. *Materials Science & Engineering C*, 58, 521–531.
- Rao, S. S., Jeyapal, S. G., & Rajiv, S. (2014). Biodegradable electrospun nanocomposite fibers based on Poly(2-hydroxy ethyl methacrylate) and bamboo cellulose. *Composites Part B: Engineering*, 60, 43–48.
- Rastogi, R. S. J. N. K., & Raj, B. (2010). Moisture Sorption Characteristics of Chitosan / Polyethylene Oxide Blended Films, 266–276.
- Rathinamoorthy, R., Technology, F., & College, P. S. G. (2012). Nanofiber for drug delivery system -, (February), 45–48.
- Raucci, M. G., Demitri, C., Giugliano, D., Benedictis, V. De, Sannino, A., & Ambrosio, L. (2014). Effect of citric acid crosslinking cellulose-based hydrogels on osteogenic differentiation, 2045–2056.
- Reddy, N., & Yang, Y. (2010). Citric acid cross-linking of starch films. *Food Chemistry*, 118(3), 702–711.
- Reza, A. T., & Nicoll, S. B. (2010). Acta Biomaterialia Characterization of novel

photocrosslinked carboxymethylcellulose hydrogels for encapsulation of nucleus pulposus cells. *Acta Biomaterialia*, 6(1), 179–186.

- Rizwan, M., Yahya, R., Hassan, A., Yar, M., Azzahari, A. D., Selvanathan, V., & Sonsudin, F. (2017). pH Sensitive Hydrogels in Drug Delivery : Brief History , Properties , Swelling , and Release Mechanism , Material Selection and Applications.
- Rogina, A. (2014). Applied Surface Science Electrospinning process : Versatile preparation method for biodegradable and natural polymers and biocomposite systems applied in tissue engineering and drug delivery. *Applied Surface Science*, 296, 221–230.
- Sadat, A., Hamid, M., & Hajiesmaeilbaigi, F. (2017). Effect of electrospinning parameters on morphological properties of PVDF nanofibrous scaffolds. *Progress in Biomaterials*, 6(3), 113–123.
- Shalumon, K. T., Binulal, N. S., Selvamurugan, N., Nair, S. V, Menon, D., Furuike, T., Jayakumar, R. (2009). Electrospinning of carboxymethyl chitin / poly (vinyl alcohol) nanofibrous scaffolds for tissue engineering applications. *Carbohydrate Polymers*, 77(4), 863–869.
- Shen, X., Shamshina, J. L., Berton, P., & Rogers, R. D. (2016). Green Chemistry fabrication , properties , and applications, (Mc), 53–75.
- Shet, R., Wong, H., & Dodou, K. (2017). Effect of Drug Loading Method and Drug Physicochemical Properties on the Material and Drug Release Properties of Poly (Ethylene Oxide) Hydrogels for Transdermal Delivery.
- Shi, Y., Wan, A., Shi, Y., Zhang, Y., & Chen, Y. (2014). Experimental and Mathematical Studies on the Drug Release Properties of Aspirin Loaded Chitosan Nanoparticles, 2014.
- Smrke, D. M., Kleinschek, K. S., Maver, T., Kurec, M., & Maver, U. (2015). Electrospun nanofibrous CMC / PEO as a part of an effective pain-relieving wound dressing.
- Stone, S. A., Gosavi, P., Athauda, T. J., & Ozer, R. R. (2013). In situ citric acid crosslinking of alginate / polyvinyl alcohol electrospun nano fi bers. *Materials Letters*, 112, 32–35.
- Sun, X., Wang, H., Jing, Z., & Mohanathas, R. (2013). Hemicellulose-based pH-sensitive and biodegradable hydrogel for controlled drug delivery. *Carbohydrate Polymers*, 92(2), 1357–1366.
- Tan, H. L., Wong, Y. Y., Muniyandy, S., & Hashim, K. (2016). Carboxymethyl sago pulp / carboxymethyl sago starch hydrogel : Effect of polymer mixing ratio and study of controlled drug release, 43652, 1–13.
- Teck, C. (2017). Progress in Polymer Science Nanofiber technology : current status and emerging developments. *Progress in Polymer Science*, 70, 1–17.

- Tezuka, Y., & Tsuchiya, Y. (1996). ^{13}C NMR determination of substituent distribution in carboxymethylcellulose by use of its peresterified derivatives I, *291*, 99–108.
- Thenapakiam, S., Kumar, D. G., Pushpamalar, J., & Saravanan, M. (2013). Aluminium and radiation cross-linked carboxymethyl sago pulp beads for colon targeted delivery. *Carbohydrate Polymers*, *94*(1), 356–363.
- Trivedi, J. H. (2013). Synthesis, Characterization, and Swelling Behavior of Superabsorbent Hydrogel from Sodium Salt of Partially Carboxymethylated Tamarind Kernel Powder-g-PAN, 1992–2003.
- Veeramachineni, A. K., Sathasivam, T., & Muniyandy, S. (2016). applied sciences Optimizing Extraction of Cellulose and Synthesizing Pharmaceutical Grade Carboxymethyl Sago Cellulose from Malaysian Sago Pulp.
- Widsten, P., Dooley, N., Parr, R., Capricho, J., & Suckling, I. (2014). Citric acid crosslinking of paper products for improved high-humidity performance. *Carbohydrate Polymers*, *101*, 998–1004.