



***RESPONSES OF EGGPLANT (*Solanum melongena* L. cv MTe 2)
TOWARDS SALINITY AND DEVELOPMENT OF PLANT GROWTH
ENHANCER IN REDUCING SALT STRESS***

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FS 2019 50



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By

FATEEN KHALIESSA BINTI MOHD ARIFIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

March 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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March 2018

Chairman : Associate Professor Rosimah binti Nulit, PhD
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Salinity is the second most widespread soil problem affecting large areas of world cultivated land and is considered as a serious constraint to increase crop production. Seed germination and seedling growth are both important stages in agronomical aspect of crop establishment and both stages are sensitive towards salt stress. Currently, no study has been done regarding salt tolerance of eggplant (*Solanum melongena* L. var MTe 2). Thus, the present work aimed to compare the effects of different types of salts on seed germination capability, early seedling growth and seed anatomy of the eggplant (*S. melongena* var. MTe 2) and also to develop liquid formulation to enhance germination and growth of salt-stressed MTe 2 seeds. Ten sterilized Mte 2 seeds were treated with 5 ml of 25, 50, 100, 150 and 200 mM of NaCl, KCl, MgCl₂ and MgSO₄ in Petri dish with deionized water as control. After 10 days, the germination performance and early seedling growth were evaluated with their parameters, respectively. The ungerminated seed were transferred to distilled water for percent recovery test and also for histological study. Seeds showed highest tolerance and germination performance in KCl followed by NaCl, MgCl₂ and MgSO₄. The germination percentage, germination rate, relative salt injury rate, seed vigor, salt tolerance, seedling height and biomass were significantly affected by interaction effect between type of salt and concentration of salt. Treatment with 25 mM KCl was found to enhance the germination performance and early seedling growth of MTe 2 seeds. The seeds were unable to germinate even at the lowest concentration of MgCl₂ and MgSO₄. Percent recovery test revealed that while NaCl and KCl imposed an osmotic stress onto MTe 2 seeds, MgCl₂ and MgSO₄ imposed a specific ion toxicity that affects the germination performance. The histological study further revealed that osmotic stress induced a state of dormancy which prevented the protusion of radicle from the testa while specific ion toxicity disrupted cell membrane integrity, interfered with metabolism of germinating seeds and caused a cell death to seed embryo. In this study, potassium chloride (KCl), potassium nitrate (KNO₃) and indole-butyric-acid (IBA) were tested for the effectiveness as a liquid enhancer towards salt stressed MTe 2 seeds. MTe 2 seeds were haloprimered with 200 mM NaCl to induce a salt-stress state to the seeds. The seeds were then dried, surface-sterilized and treated with different

combinations of KCl, KNO₃ and IBA, respectively. After ten days, the germination performance and early seedling growth were evaluated. Results found that the mixture of 25 mM KCl with 6 ppm IBA significantly produced seedlings with the highest seed vigor and seedling length among all other treatments in compared to control. In conclusion, the salt tolerance of eggplant MTe 2 at germination and early growth stages are as follows: NaCl>KCl>MgCl₂>MgSO₄. The mixture of 25 mM KCl and 6 ppm IBA were found to enhance the germination performance and early seedling growth of salt-stressed eggplant MTe 2.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**TINDAK BALAS TERUNG (*Solanum melongena* cv. MTe 2) TERHADAP
KEMASINAN DAN PEMBANGUNAN CECAIR PENGALAK
PERTUMBUHAN UNTUK MENGURANGKAN TEKANAN GARAM**

Oleh

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Kemasinan merupakan masalah kedua utama yang meliputi sebahagian besar tanah di seluruh dunia dan dianggap sebagai kekangan yang serius untuk meningkatkan pengeluaran tanaman. Percambahan biji benih dan pertumbuhan anak benih adalah peringkat penting dalam aspek agronomi untuk penubuhan tanaman dan kedua-dua peringkat tumbesaran tersebut sensitif terhadap tekanan garam. Terkini, tiada lagi kajian telah dilakukan mengenai kadar toleransi terung (*Solanum melongena* cv. MTe 2) terhadap jenis garam yang berbeza. Oleh itu, kajian ini dijalankan bertujuan untuk membandingkan kesan garam yang berbeza terhadap percambahan biji benih, pertumbuhan benih awal dan anatomi biji benih terung (*S. melongena* cv. MTe 2) dan membangunkan formulasi cecair yang boleh meningkatkan percambahan dan pertumbuhan biji benih terung MTe 2 dalam keadaan persekitaran yang ada kemasinan. Sepuluh biji benih MTe 2 diletakkan dalam bekas petri yang telah diletakkan 5 mL 25, 50, 100, 150 dan 200 mM NaCl, KCl, MgCl₂ dan MgSO₄ dan air ternyahion sebagai kawalan. Selepas 10 hari, prestasi percambahan dan pertumbuhan anak benih terung Mte 2 dinilai menggunakan parameter masing-masing. Benih yang tidak bercambah dalam masa 10 hari dalam rawatan dipindahkan ke air suling untuk kajian pemulihan peratus yang selanjutnya dan juga untuk kajian histologi. Benih menunjukkan prestasi toleransi dan percambahan tertinggi di dalam rawatan KCl, diikuti oleh NaCl, MgCl₂ dan MgSO₄. Peratusan percambahan, kadar percambahan, relatif kecederaan garam, kesegaran benih. Toleransi garam, panjang anak benih dan biomas telah terkesan secara signifikan oleh kadar kepekatan dan jenis garam yang berbeza. Rawatan dengan 25 mM KCl pula didapati meningkatkan percambahan dan pertumbuhan benih Mte 2. Tiada percambahan biji benih terung Mte 2 dalam MgCl₂ dan MgSO₄, walaupun pada kepekatan yang terendah. Ujian pemulihan peratus mendedahkan bahawa walaupun NaCl dan KCl mengenakan tegasan osmotik ke atas biji benih terung MTe 2, MgCl₂ dan MgSO₄ mempunyai ketoksikan tertentu yang mempengaruhi prestasi percambahan biji benih tersebut. Kajian histologi seterusnya mendedahkan bahawa tekanan osmosis menyebabkan keadaan dorman pada biji benih dan seterusnya menghalang pengeluaran radikal manakala ketoksikan ion tertentu mengganggu integriti sel membran, mengganggu metabolisme dan menyebabkan kematian sel kepada embrio benih. Dalam kajian ini, KCl,

KNO_3 dan IBA telah diuji untuk keberkesanan sebagai formula cairan. Biji benih terung MTe 2 telah direndam terlebih dahulu dalam 200 mM NaCl untuk memberi tekanan garam kepada biji benih. Kemudian, biji benih dikeringkan, melalui proses sterilasi dan dirawat pula dengan campuran KCl, KNO_3 dan IBA untuk 10 hari. Selepas 10 hari, prestasi percambahan dan pertumbuhan anak benih dinilai. Keputusan mendapati bahawa campuran 25 mM KCl dan 6 ppm IBA secara signifikannya telah menghasilkan benih terpanjang dan terbaik berbanding semua rawatan lain termasuklah rawatan kawalan. Kesimpulannya, toleransi biji benih terung MTe 2 dalam garam yang berbeza adalah seperti berikut: $\text{NaCl} > \text{KCl} > \text{MgCl}_2 > \text{MgSO}_4$. Campuran 25 mM KCl and 6 ppm IBA secara signifikan telah menambah baik percambahan biji benih dan pertumbuhan awal terung MTe 2 yang telah menerima tekanan garam.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

NaCl	sodium chloride
KCl	potassium chloride
MgCl ₂	magnesium chloride
MgSO ₄	magnesium sulphate
CaSO ₄	calcium sulphate
CaCl ₂	calcium chloride
Na ₂ SO ₄	sodium sulphate
NaHCO ₃	sodium bicarbonate
KNO ₃	potassium nitrate
IBA	indole-butyric acid
MTe 2	<i>Malaysia varieti terung 2</i>
mM	milimolar
ppm	part per million
EC	electrical conductivity
dS/m	deciSiemens per meter
EC _e	electrical conductivity of saturated paste extract
mg L ⁻¹	milligram per liter
NaClO	sodium hypochlorite
GP	germination percentage
GR	germination rate
RIR	relative injury rate
ST	salt tolerance of seed
SHR	seedling height reduction

CHAPTER 1

INTRODUCTION

1.1 Background Study

Abiotic stresses are serious issues concerning agricultural production worldwide for the ability of reducing average yields of major crop plants by more than 50%. These stresses have an adverse effects on plants by triggering a series of morphological, physiological, biochemical, and molecular changes in crop plants, hence affecting the growth and reducing the crop productivity. Among these stresses, salinity gives a remarkable impact on farmlands worldwide (FAO & ITPS, 2015). Globally, approximately 25% of the agricultural land is saline and it continues to be a major problem in the arid and semi-arid regions (Karan & Subudhi, 2012).

Seed germination is a critical and decisive phase of a plant life cycle as the survival and continuity of a species depends on the ability of the plant to germinate and establish itself as a seedling in its environment (Deng et al., 2014). A delayed and reduced in seed germination and later seedling emergence may lead to non-uniform stand establishment and reduced yield. (Wojtyła, Lechowska, Kubala & Garnczarska, 2016). Salinity stress is a well-known, major abiotic factor that provokes disorders in seeds and in its entirety affects the germination, seedling development, crop growth and productivity by disrupting homeostasis in water potential, water uptake and ion distribution. Often, secondary stress might develop as the consequence of these primary effects and result in oxidative damage through ion toxicity to the metabolic machinery (Zhang et al., 2014; Chaparzadeh & Hosseinzad-Behboud, 2015).

Eggplant (*Solanum melongena* L.) is an important non-tuberous crop which belongs to the nightshade family of vegetables. It is one of the most common crop grown in India, Pakistan, China, Philippines, Bangladesh and many Asian countries. Besides low in calories and fats, eggplant is also a good source of vitamins and minerals and rich in amide proteins, free reducing sugars and total water soluble sugars. The purple colour of the fruit comes from anthocyanin pigment, an antioxidant that has been found to prevent brain damages and serve as anti-tumoral activities, alongside with other health benefits (Lim, 2013).

1.2 Problem Statement and Justification of Study

Various salts occur worldwide yet, many research only focused on the salinity effects of NaCl on plant. Agricultural soil composition and irrigation water consists combinations of anions and cations of magnesium, potassium, chloride and sulphate. Currently, the effects of various concentrations and types of salt stress on Malaysian variety of eggplant, MTe 2 are not fully understood and no research has been done before. Though previous studies stated that it is moderately sensitive (Demir et al., 2003; Akinci et al, 2004; Shaheen, Naseer, Ashraf & Akram, 2013), the knowledge of salt tolerance of this vegetable plant and how their growth and development are affected by salt stress are outdated and scarce. The response differs depends on the variety, developmental stage, genotype and environment on which the eggplant is being cultivated. Knowledge on the tolerance of eggplant MTe 2 under salt stress may later contribute in minimizing the impact of salinity and thus increase the efficiency of eggplant MTe 2 production in salt-affected soils in Malaysia.

Besides that, the interaction between dormant salt-stressed seed and plant growth regulators on eggplant MTe 2 has never been studied before. The alleviation of salt stress by liquid growth enhancer that is formulated from plant growth regulators may serve as an alternative to reduce loss due to salinity stress on crop production.

1.3 Objectives

Hence the objectives of the present work are as follows:

1. To compare the effects of different concentrations of salts on germination, seedling growth and anatomy of eggplant (*Solanum melongena* L. cv. MTe 2).
2. To develop a liquid growth enhancer to mitigate the effects of salt stress on eggplant seed (*Solanum melongena* L. cv. MTe 2).

REFERENCES

- Abdul-Baki, A. A., & Anderson, J. D. (1973). Vigor determination in soybean seed by multiple criteria. *Crop Science*, 13(6), 630-633.
- Abid, M., Haddad, M., & Ferchichi, A. (2008). Effect of magnesium sulphate on the first stage of development of Luceren. In C. Porqueddu, and M. M. Tavares de Sousa (Eds), *Sustainable Mediterranean Grasslands and Their Multi-functions* (pp. 405-408). Zaragoza: CIHEAM/ FAO/ ENMP/ SPPF.
- Achakzai, A. K. K. (2009). Effect of water stress on imbibition, germination and seedling growth of maize cultivars. *Sarhad Journal of Agriculture*, 25(2), 165-172.
- Adams, E., & Shin, R. (2014). Transport, signalling and homeostasis of potassium and sodium in plants. *Journal of Integrated Plant Biology*, 56, 231-249.
- Akanitapichat, P., Phraibung, K., Nuchklang, R., & Prompitakkul, S. (2010). Antioxidant and hepatoprotective activities of five eggplant varieties. *Food and Chemical Toxicology*, 48, 3017-3021.
- Akinci I. E., Akinci S., Yilmaz, A., & Dikici H. (2004). Response of eggplant varieties (*Solanum melongena*) to salinity in germination and seedling stages. *New Zealand Journal of Crop and Horticultural Science*, 32(2), 193-200.
- Almansouri, M., Kinet, J. M., & Lutts, S. (2001). Effect of salt and osmotic stresses on germination in durum wheat (*Triticum durum* Desf.). *Plant and Soil*, 231, 243-254.
- Al-Fayer, K. A., & Bazaid, S. A. (2014). Improving drought and salinity tolerance in barley by application of salicylic acid and potassium nitrate. *Journal of the Saudi Society of Agricultural Science*, 13, 45-55.
- Al-Hassan, M., Pacurer, A., Lopez-Gresa, M. P., Donat-Torres, M. P., Llinares, J. V., Boscaiu, & Vicente, O. (2016). Effects of salt stress on three ecologically distinct *Plantago* species. *PLoS ONE*, 11(8), 1-21.
- Al-Tardeh, S., & Iraki, N. (2013). Morphological and anatomical responses of two Palestinian tomato (*Solanum lycopersicon* L.) cultivars to salinity during seed germination and early growth stages. *African Journal of Biotechnology*, 12(30), 4788-4797.
- Ameixa, O. M. C. C., Marques, B., Fernandes, V. S., Soares, A. M. V. M., Calado, R., & Lillebo, A. I. (2016). Dimorphic seeds of *Salicornia ramossima* display contrasting germinating responses under different salinities. *Ecological Engineering*, 87, 120-123.
- Asma, A. A. S. (2015). *Impact of salinity on seed germination and antioxidative profiling of tomato leaves (Solanum lycopersicon cv. MT1)*. (Master's thesis, Universiti Putra Malaysia).

- Aydinsakir, K., Ulukapi, K., Kurun, R., & Buyuktas, D. (2013). The effects of different salt source and concentrations on seed germination and seedling growth of pumpkin varieties used as rootstock. *Journal of Food, Agriculture & Environment*, 11(1), 503-510.
- Bahraimi, H., & Razmjoo, J. (2012). Effect of salinity stress (NaCl) on germination and early seedling growth of ten sesame cultivars (*Sesamum indicum* L.). *International Journal of AgriScience*, 2(6), 529-537.
- Banuelos, G. S., Mead, R., & Hoffman, G. J. (1993). Accumulation of selenium in wild mustard irrigated with agricultural effluent. *Agriculture Ecosystem and Environment*, 43, 119-126.
- Bao, G. R., Wu, R. H., Baoburen, Q. Q., Wang, H. L., & Chen, L. (2010). Effect of saline on seed germination of *Onobrychis viciaefolia*. *Journal of Inner Mongolia for Nationalities (Natural Sciences)*, 25(6), 640-642.
- Bekendam, G., & Grob, R. (1979). *Handbook for seedling evaluation*. Zurich, Switzerland: The international Seed Testing Association [ISTA].
- Bewley, J. D., Bradford, K. J., Hilhorst, H. W. M., & Nonogaki, H. (2013). *Seeds Physiology of Development, Germination and Dormancy*, 3rd ed. New York, Heidelberg, Dordrecht London: Springer.
- Brandenburg, W., & Kleier, C. (2011). Effect of MgCl₂ on germination, growth and biomass allocation of the radish cv. "cherry belle". *American Journal of Environmental Science*, 7(2), 132-135.
- Cavallaro, V., Maucieri, C., & Barbera, A. C. (2014). *Lolium multiflorum* Lam. cvs germination under simulated olive mill wastewater salinity and pH stress. *Ecological Engineering*, 71, 113-117.
- Chaparzadeh, N., & Hosseinzad-Behboud, E. (2015). Evidence for enhancement of salinity induced oxidative damage s by salicylic acid in radish (*Raphanus sativus* L.). *Journal of Plant Physiology and Breeding*, 5(1), 23-33.
- Chartzoulakis, K., & Klapaki, G. (2000). Response of two greenhouse pepper hybrids to NaCl salinity during different growth stages. *Scientia Horticulturae*, 86, 247-260.
- Cha-Um, S., Takabe, T., & Kirdmanee, C. (2010). Ion contents, relative electrolyte leakage, proline accumulation, photosynthetic abilities and growth characters of oil palm seedlings in response to salt stress. *Pakistan Journal of Botany*, 42(3), 2191-2020.
- Chehregani, A., Mohsenzade, F., & Ghanad, M. (2011). Male and female gametophyte development in *Cichorium intybus*. *International Journal of Agriculture & Biology*, 13, 603-606.

- Chhun, S., Taketa, S., Tsurumi, S., & Ichii, M. (2013). The effects of auxin on lateral root initiation and root gravitropism in a lateral rootless mutant Lrtl of rice (*Oryza sativa* L.). *Plant Growth Regulator*, 39(2), 161-170.
- Curtain, D., Steppuhn, H., & Selles, F. (1993). Plant responses to sulfate and chloride salinity: Growth and ionic relations. *Soil Science Society of American Journal*, 57, 1304-1310.
- Dantas, B. F., De Sa Ribeiro, L., & Aragao, C. A. (2007). Germination, initial growth and cotyledon protein content of bean cultivars under salinity stress. *Revista Brasileira de Sementes*, 29, 106–110.
- Daunay, M. C. (2008). Eggplant. In J. Prohens, & F. Nuez (Eds), *Vegetables II* (pp. 163). New York: Springer Science+Business Media.
- de Pascual-Teresa, S. (2014). Molecular mechanisms involved in the cardiovascular and neuroprotective effects of anthocyanins. *Archives of Biochemistry and Biophysics*, 559, 68-74.
- Deak, K. I., & Malamy, J. (2006). Osmotic regulation of root system architecture. *Plant Journal*, 43, 17-28.
- Deinlein, U., Stephan, A. B., Horie, T., Luo, W., Xu, G., & Schroeder, J. I. (2014). Plant salt-tolerance mechanisms. *Trends in Plant Science*, 19, 371-379.
- Demidchik, V., Straltsova, D., Medvedev, S. S., Pozhvanov, G. A., Sokolik, A., & Yurin, V. (2013). Stress-induced electrolyte leakage: the role of K⁺ permeable channels and involvement in programmed cell death and metabolic adjustment. *Journal of Experimental Botany*, 65(5), 1259-1270.
- Demir, I., Mavi, K., Ozcoban, M., & Okcu, G. (2003). Effect of salt stress on germination and seedling growth in serially harvested aubergine (*Solanum melongena* L.) seeds during development. *Israel Journal of Plant Sciences*, 51(2), 125-131.
- Deng, Y., Yuan, F., Feng, Z., Ding, T., Song, J., & Wang, B. (2014). Comparative study on seed germination characteristics of two species of Australia saltbrush under salt stress. *Acta Ecologica Sinica*, 34, 337-341.
- Department of Agriculture Peninsular Malaysia. (2012). Vegetables and Cash Crops Statistic. Putrajaya, Malaysia.
- Department of Agriculture Peninsular Malaysia. (2016). Vegetables and Cash Crops Statistic. Putrajaya, Malaysia.
- Dong, T., Dong, T., Tong, J., & Xiao, L. (2012). Nitrate, abscisic acid and gibberellin interactions on the thermo-inhibition of lettuce seed germination. *Plant Growth Regulation*, 66.

- Driessen, P., & Dudal, R. (1991). *Lecture notes on the geography, formation, properties and use of the major soils of the world*. Wageningen Agricultural University, Wageningen, the Netherlands.
- Elhag, A. Z., & Elzain, H. (2012). Sodium chloride tolerance of pineapple (*Ananas comosus* L. Mirrl.) in vitro. *Journal of Applied Agricultural Research*, 4(2), 105-110.
- Elouaer, M. A., & Cherif, H. (2012). Effect of NaCl priming duration and concentration on germination behaviour of Tunisian safflower. *Journal of Stress Physiology and Biochemistry*, 8(3).
- Eskandari, H. (2013). Effects of priming technique on seed germination properties, emergence and field performance of crops: a review. *International Journal of Agronomical Plant Production*, 4, 454-458.
- Fang, J. (2014). Bioavailability of anthocyanins. *Drug Metabolism Reviews*, 46(4), 508-520.
- FAO. (2011). *Current world fertilizer trends and outlook to 2015*. Rome: Food and Agriculture Organization of the United Nations.
- FAO. (2018). <http://www.fao.org/soils-portal/soil-management/management-of-some-problem-soils/salt-affected-soils/en/#c239240> accessed on 16 January 2018.
- FAO & ITPS. (2015). *Status of the World's Soil Resources (SWSR) Main Report*. Rome, Italy: Food and Agriculture Organization of the United Nations and Intergovernmental Technical Panel on Soils.
- Fayez, K. A., & Bazaid, S. A. (2013). Improving drought and salinity tolerance in barley by application of salicylic acid and potassium nitrate. *Journal of the Saudi Society of Agricultural Sciences*, 13, 45-55.
- Fernandez-Garcia, N., Olmos, E., Bardisi, E., De la Garma, J. C., Lopez-Berenguer, C., & Rubio-Asensio, J. S. (2013). Intrinsic water use efficiency controls the adaptation to high salinity in a semi-arid adapted plant, henna (*Lawsonia inermis* L.). *Journal of Plant Physiology*, 171, 64-75.
- Forester, S. C., Choy, Y. Y., Waterhouse, A. L., & Oteiza, P. I. (2014). The anthocyanin metabolites gallic acid, 3-O-methylgallic acid, and 2,4,6-trihydroxybenzaldehyde decrease human colon cancer cell viability by regulating pro-oncogenic signals. *Molecular Carcinogenesis*, 53, 432-439.
- Fu, J., & Wang, S. (2011). Insights into auxin signaling in plant-pathogen interactions. *Frontiers in Plant Science*, 2, 74.
- Fernandez-Garcia, N., Olmos, E., Bardisi, E., la Garma, J. C. D., Lopez-Berenguer, C., & Rubio-Asensio, J. S. (2013). Intrinsic water use efficiency controls the adaptation to high salinity in a semi-arid adapted plant, henna (*Lawsonia inermis* L.). *Journal of Plant Physiology*, 37, 64-75.

- Garcia, R. A., Li, Y., & Rosolem, C. A. (2013). Soil organic matter and physical attributes affected by crop rotation under no-till. *Soil Science Society of American Journal*, 77(5), 1724-1731.
- Gaylord, B., & Egan, T. P. (2008). How salts of sodium, potassium and sulfate affect the germination and early growth of *Atriplex acanthocarpa* (Chenopodiaceae). In M. A. Khan, & D. J. Weber (Eds), *Ecophysiology of high salinity tolerant plants* (pp 1-9). New York, USA: Springer Science + Business Media.
- Ghanad, M. (2015). *Gametophyte development, antioxidant properties and response to salinity of Brassica rapa L. var. parachinensis*. (Doctoral dissertation, Universiti Putra Malaysia).
- Ghodrat, V., Rousta, M. J., & Tadaion, M. S. (2012). Effect of priming with indolebutyric acid (IBA) on germination and growth of wheat under saline conditions. *International Journal of Agriculture and Crop Sciences*, 4(6), 289-292.
- Ghosh, G., Kumar, S., & Saha, K. (2012). Hyperspectral satellite data in mapping salt-affected soil using linear spectral unmixing analysis. *Journal of Indian Society Remote Sense*, 40, 129-136.
- Gomes-Filho, E., Machado Lima, C. R. F., Costa, J. H., da Silva, A. C., da Guia Silva Lima, M., de Lacerda, C. F., & Prisco, J. T. (2008). Cowpea ribonuclease: properties and effect of NaCl-salinity on its activation during seed germination and seedling establishment. *Plant Cell Report*, 27, 147-157.
- Greenway, H., & Munns, R. (1980). Mechanisms of salt tolerance in non-halophytes. *Annual Review of Plant Physiology*, 31, 149-190.
- Grieve, G. M., Grattan, S. R., & Maas, E. V. (2012). Plant salt tolerance. In W. W. Wallander, & K. K. Tanji, (Eds.), *ASCE Manual and Report on Engineering Practice No 71 Agricultural Salinity Assessment and Management (2nd Edition)* (pp. 405-459). Reston: ASCE.
- Grigore, M. N., Toma, C., & Zamfiracha, M. M. (2011). An ecological approach for proposing and describing a new type of soil salinization. In Thomas, R. P. *Proceedings of the Global Forum on Salinization and Climate Change* (pp 21). Rome: Food and Agriculture Organization of the United Nations.
- Guo, J. H., Liu, X. J., Zhang, Y., Shen, J. L., Han, W. X., Zhang, W. F., ... Zhang, F. S. (2010). Significant acidification in major Chinese croplands. *Science*, 327, 1008-1010.
- Hakim, M. A., Abdul, S. J., Hanafi, M. M., Ali, E., Mohd, R. I., Ahmed, S., & Rezaul Karim, S. M. (2014). Effect of salt stress on morpho-physiology, vegetative growth and yield of rice. *Journal of Environmental Biology*, 35, 317-326.
- Hansch, R., & Mendel, R. R. (2009). Physiological functions of mineral micronutrients (Cu, Zn, Mn, Fe, Ni, Mo, B, Cl). *Current Opinion in Plant Physiology*, 12, 259-266.

- Hartmann, H. T., Kester, D. E., Davies, F. T., & Geneve, R. L. (2002). *Plant Propagation: Principles and Practices (7th edition)*. New Jersey: Prentice-Hall.
- Hasanuzzaman, M., Nahar, K., & Fujita, M. (2013). Plant response to salt stress and role of exogenous protectants to mitigate salt-induced damages. In P. Ahmad, M. N. V. Prasad, & M. M. Azooz (Eds), *Ecophysiology and Responses of Plants under Salt Stress* (pp. 25-87). New York, NY: Springer Science+Business Media.
- Hasanuzzaman, M., Nahar, K., Fujita, M., Ahmad, P., Chandna, R., Prasad, M. N. V., & Ozturk, M. (2013). Enhancing plant productivity under salt-stress: Relevance of poly-omics. In P. Ahmad et al. (Eds), *Salt stress in Plants: Signalling, Omics and Adaptations* (pp. 113-156). New York, NY: Springer Science+Business Media.
- Hashim, G. M. (2009). *Salt-affected soils of Malaysia*. In Proceedings of the Workshop on Soil Science in Malaysia towards 2020.
- Hopkins, W. G., & Huner, N. P. A. (2009). *Introduction to Plant Physiology Fourth Edition*. Hoboken, NJ: John Wiley & Sons, Inc.
- Horie, T., Karahara, I., & Katsuhara, M. (2012). Salinity tolerance mechanisms in glycophytes: An overview with the central focus on rice plants. *Rice*, 5, 11.
- Hussain, A., & Khan, A. M. (2004). Effect of growth regulators on stem cutting of *Rosa bourboniana* and *Rosa gruss-an-teplitz*. *International Journal of Agriculture & Biology*, 6(5), 931–932.
- Ibrahim, E. A. (2016). Seed priming to alleviate salinity stress in germinating seeds. *Journal of Plant Physiology*, 192, 38-46.
- IFA. (2012). *Global supply and demand outlook for fertilizer and raw materials*. IFA. (Also available at www.fertilizer.org)
- Imran, M., Mahmood, A., Romheld, V., & Neumann, G. (2013). Nutrient seed priming improves seedling development of maize exposed to low root zone temperatures during early growth. *European Journal of Agronomy*, 49, 141-148.
- Im-Erb, R., & Sukchan, S. (2009). Salt-affected soils in Thailand: Assessment and monitoring of salinization. In: *Advanced in the assessment and monitoring of salinization and status of biosaline agriculture* (pp 19-20) Rome, Italy: Food and Agricultural Organization.
- Im-Erb, R., Neawsuparb, K., Sukchan, S., Srinarong, S., & Sombatpanit, S. (2011). Effect of climate and land use change to increase salinity impact in North-east Thailand. In Thomas, R. P.: *Proceedings of the Global Forum on Salinization and Climate Change* (pp 61-62) Rome, Italy: Food and Agricultural Organization.

- IRRI. (2011). *Stress and disease tolerance: Breeding for salt tolerance in rice*. Retrieved from <http://www.knowledgebank.irri.org/ricebreedingcourse/bodydefault> on 12 October 2016.
- Jabeen, R., & Ahmad, R. (2009). Alleviation of the adverse effects of salt stress by foliar application of sodium antagonistic essential minerals on cotton (*Gossypium hirsutum*). *Pakistan Journal of Botany*, 41(5), 2199-2208.
- Joseph, B., & Jini, D. (2010). Salinity induced programmed cell death in plants: challenges and opportunities for salt-tolerant plants. *Journal of Plant Science*, 5, 376-390.
- Karan, R., & Subudhi, P. K. (2012). Approaches to increasing salt tolerance in crop plants. In P. Ahmad, & M. N. V. Prasad (Eds), *Abiotic Stress Responses in Plants: Metabolism, Productivity and Sustainability*. (pp 63-88). New York, NY: Springer Science+Business.
- Karihaloo, J. L., & Gottlieb, L. D. (1995). Allozyme variation in the eggplant, *Solanum melongena* L. (*Solanaceae*). *Theory of Applied Genetics*, 90, 578-583.
- Khan, M. A., & Weber, D. J. (Eds.). (2008). *Ecophysiology of High Salinity Tolerant Plants*. USA: Springer Science + Business Media.
- Khodarahmpour, Z., Ifar, M., & Motamedi, M. (2012). Effects of NaCl salinity on maize (*Zea mays* L.) at germination and early seedling stage. *African Journal of Biotechnology*, 11, 298-304.
- Knapp, S., Vorontsova, M. S., & Prohens, J. (2013). Wild relatives of the eggplant (*Solanum melongena* L.: Solanaceae): New understanding of species name in a complex group. *PLoS ONE*, 8(2), e57039.
- Koyro, H. W. (2002). Ultrastructural effects of salinity in higher plants. In A. Lauchli, & U. Luttge (Eds), *Salinity: Environment, plants, molecules* (pp 139-157). Kluwer, Amsterdam.
- Lara, T. S., Lira, J. M. S., Rodrigues, A. C., Rakocevic, M., & Alvarenga, A. A. (2014). Potassium nitrate priming affects the activity of nitrate reductase and antioxidant enzymes in tomato germination. *Journal of Agricultural Science*, 6(2), 72-80.
- Lauchli, A., & Grattan, S. R. (2007). Plant growth and development under salinity stress. In M. A. Jenks, P. M. Hasegawa, & Jain, S. M. (Eds.), *Advances in Molecular Breeding Towards Drought and Salt Tolerance Crops* (pp. 1-32). New York: Springer.
- Lee, M. K., & van Iersel, M. W. (2008). Sodium chloride effects on growth, morphology and physiology of chrysanthemum (*Chrysanthemum morifolium*). *HortScience*, 43(6), 1881-1891.

- Lei, B., Ling, Y., Jian-An, W., & Hai-Long, S. (2013). Effects of KNO₃ pretreatment and temperature on seed germination of *Sorbus pohuashanensis*. *Journal of Forestry Research*, 24(2), 309-316.
- Li, J. Y., Jiang, A. L., & Zhang, W. (2007). Salt stress induced programmed cell death in rice root tip cells. *Journal of Integrated Biology*, 49, 481-486.
- Li, T., Hu, Y., Du, X., Tang, H., Shen, C., & Wu, J. (2014). Salicylic acid alleviates the adverse effects of salt stress in *Torreya grandis* cv. Merrillii seedlings by activating photosynthesis and enhancing antioxidant systems. *PLoS ONE*, 9(10), e109492.
- Li, D., Wang, P., Luo, Y., Zhao, M., & Chen, F. (2017). Health benefits of anthocyanins and molecular mechanisms: update form recent decades. *Critical Reviews in Food Science and Nutrition*, 57(8), 1729-1741.
- Li, G., Zhu, Y., Zhang, Y., Lang, J., Chen, Y., & Ling, W. (2013). Estimated daily flavonoid and stilbene intake from fruits, vegetables, and nuts and associations with lipid profiles in Chinese adults. *Journal of the Academy of Nutrition and Dietetics*, 113, 786-794.
- Li, Y. (2008). Effect of salt stress on seed germination and seedling growth of three salinity plants. *Pakistan Journal of Biological Science*, 11, 1268-1272.
- Lim, T. K. (2013). *Fruits Volume 6*. Dordrecht: Springer Science+Business Media.
- Lin, J., Yu, D., Shi, Y., Sheng, H., Li, C., Wang, Y., ... Li, X. (2016). Salt-alkali tolerance during germination and establishment of *Leymus chinensis* in the Songnen Grassland of China. *Ecological Engineering*, 95, 763-769.
- Llanes, A., Bertazza, G., Palacio, G., & Luna, V. (2013). Different sodium salts cause different solute accumulation in the halophyte *Prosopis strombulifera*. *Plant Biology*, 15(1), 118-125.
- Loescher, W., Chan, Z., & Grumet, R. (2011). Options for developing salt-tolerant crops. *Horticultural Science*, 46(8), 1085-1092.
- Macedo, A. F. (2013). Abiotic stress responses in plants: metabolism to productivity. In P. Ahmad, & M. N. V. Prasad (Eds.), *Abiotic stress responses in plants: Metabolism, productivity and sustainability*. New York, USA: Springer Science+Business Media.
- Malvi, U. (2011). Interaction of micronutrients with major nutrients with special reference to potassium. *Karnataka Journal of agricultural Science*, 24(1), 106-109.
- Mantri, N., Patade, V., Penna, S., Ford, R., & Pang, E. (2013). Abiotic stress responses in plants: Present and Future. In P. Ahmad P, & M. N. V. Prasad (Eds). *Abiotic stress response in plants: metabolism, productivity and sustainability* (pp 1-19). New York: Springer Science+Business media.

- Muscolo, A., Sidari, M., Panuccio, M. R., Santonoceto, C., Orsini, F., & De Pascale, S. (2011). Plant responses in saline and semiarid environments: an overview. *The Europea Journal of Plant Science and Biotechnology*, 5, 1-11.
- Maurya, V., & Gothandam, K. M. (2014). Factors influencing the salt stress tolerance in plants: An overview. *Research Journal of Biotechnology*, 9(2), 79-88.
- McKenzie, D., & Orange, P. L. M. (2003). Salinity and sodicity: What's the difference? *The Australian Cottongrower*, 24(28).
- McMahan, C., Kostyal, D., Lhamo, D., & Cornish, K. (2015). Protein influences on guayule and Hevea natural rubber soil and gel. *Journal of Applied Polymers Science*, 132(23), 42051.
- Melling, L., Teng, C. S., & Husni, M. H. A. (2002). Sustainable agriculture development on peat land in Sarawak. In Shamshuddin, J., Hamdan, J. & Samsuri, A. W. (Eds) "Sustainable Land Management", *Malaysian Society of Soil Science*, 20-31.
- Meyer, R. S., Karol, K. G., Little, D. P., Nee, M. H., & Litt, A. (2012). Phylogeographic relationships among Asian eggplants and new perspectives on eggplant domestication. *Molecular Phylogenetic Evolution*, 63(3), 685-701.
- Mian, A. A., Senadheera, P., & Maathuis, F. J. M. (2011). Improving crop salt tolerance: anion and cation transporters as genetic engineering targets. *Plant Stress*, 5(1), 64-72.
- Ministry of Science & Technology. (n.d). *Series of Crop Specific Biology Documents: Biology of Brinjal*, pp 1-27. Government of India.
- Millar, J., & Roots, J. (2012). Changes in Australian agriculture and land use: implications for future food security. *International Journal of Agricultural Sustainability*, 10, 25-39.
- Moocheshi, A. S., Shekoofa, A., Sadeghi, H., & Pessarakli, M. (2014). Drought and salt stress mitigation by seed priming with KNO₃ and urea in various maize hybrids: An experimental approach based on enhancing antioxidant responses. *Journal of Plant Nutrition*, 37, 674-689.
- Munns R. (2002). Comparative physiology of salt and water stress. *Plant Cell Environment*, 25, 239-250.
- Munns, R., & Tester, M. (2008). Mechanisms of salinity tolerance. *Annual Review on Plant Biology*, 59, 651-681.
- Muraki, I., Imamura, F., Manson, J. E., Hu, F. B., Willett, W. C., van Dam, R. M., & Sun, Q. (2013). Fruit consumption and risk of type 2 diabetes: Results from three prospective longitudinal cohort studies. *The BMJ*, 347, f5001.

- Nasri, N., Kaddour, R., Rabhi, M., Plassard, C., & Lachaal, M. (2011). Effect of salinity on germination, phytase activity and phytate content in lettuce seedling. *Acta Physiology Plant*, 33, 935-942.
- Nasri, N., Saidi, I., Kaddour, R., & Lachaal, M. (2015). Effect of salinity on germination, seedling growth and acid phosphatase activity in lettuce. *American Journal of Plant Sciences*, 6, 57-63.
- Nawaz, A., Muhammad, A., Muhammad, A. P., & Afzal, I. (2011). Effect of halopriming on germination and seedling vigour of tomato. *Africa Journal of Agricultural Research*, 6, 3551-3559.
- Neumann, P. M. (2011). Recent advances in understanding the regulation of whole-plant growth inhibition by salinity, drought and colloid stress. In Kader, J. C., & Delseny, M. (Eds), *Advanced Botanical Research Vol 57* (pp. 33-48).
- Nizam, I. (2011). Effects of salinity stress on water uptake, germination and early seedling growth of perennial ryegrass. *African Journal of Biotechnology*, 10(51), 10418-10424.
- Noda, Y., Kneyuki, T., Igarashi, K., Mori, A., & Packer, L. (2000). Antioxidant activity of nasunin, an anthocyanin in eggplant peels. *Toxicology*, 148, 119-123.
- Nonogaki, H., Bassel, G. W., & Bewley, J. D. (2010). Germination, still a mystery. *Plant Science*, 179, 574-581.
- Odjegba, V. J., & Chukwunwike, I. C. (2012). Physiological responses of *Amaranthus hybridus* L. under salinity stress. *Indian Journal of Innovations and Developments*. 1(10), 742-748.
- Orlovsky, N., Japakova, U., Zhang, H., & Volis, S. (2016). Effect of salinity on seed germination, growth and ion content in dimorphic seeds of *Salicornia europaea* L. (*Chenopodiaceae*). *Plant Diversity*, 38, 183-189.
- Othman, Y., Al-Karaki, G., Al-Tawaha, A.R., & Al-Horani, A. (2006). Variation in germination and ion uptake in barley genotypes under salinity conditions. *World Journal of Agricultural Science*, 2, 11-15.
- Panuccio, M. R., Jacobsen, S. E., Akhtar, S. S., & Muscolo, A. (2014). Effect of saline water on seed germination and early seedling growth of the halophyte quinoa. *AoB PLANTS*, 6, plu047.
- Parida, A. K., & Das, A. B. (2005). Salt tolerance and salinity effects on plants: A review. *Ecotoxicology and Environmental Safety*, 60, 324-349.
- Parida, A. K., Das, A. B., & Mitra, B. (2004). Effects of salt on growth, ion accumulation photosynthesis and leaf anatomy of the mangrove, *Bruguiera parviflora*. *Trees-Structure and Function*, 18, 167-174.

- Peerzada, Y. Y., Khalid, U. R. H., Chandna, R., & Ahmad, P. (2012). Role of glutathione reductase in plant abiotic stress. In P. Ahmad, & M.N. V. Prasad (Eds), *Abiotic Stress Responses in Plants: Metabolism, Productivity* (pp 149-158). Springer Science+Business.
- Pessaraki, M., & Szabolcs, I. (2010). Soil salinity and sodicity as particular plant/crop stress factors. In M. Pessaraki (Ed). *Handbook of plant and crop stress. 3rd edition*. Boca Raton: CRC Press.
- Prohens, J., Whitaker, B. D., Plazas, M., Vilanova, S., Hurtado, M., Blasco, M.,... Stommel, J. R. (2013). Genetic diversity in morphological characters and phenolic acids content resulting from an interspecific cross between eggplant, *Solanum melongena*, and its wild ancestor (*S. incanum*). *Annals of Applied Biology*, 162, 242-257.
- Promila, K.m & Kumar, S. (2000). *Vigna radiata* seed germination under salinity. *Biology of Plant*, 43, 423-426.
- Rahdari, P, Tavakoli, S., & Hosseini, S.M. (2012). Studying of salinity stress effect on germination, proline, sugar, protein, lipid and chlorophyll content in purslane (*Portulaca oleracea* L.) leaves. *Journal of Stress Physiology and Biochemistry*, 8(1), 182-193.
- Rahimi, A. (2013). Seed priming improves the germination performance of cumin (*Cuminum syminum* L.) under temperature and water stress. *Industrial Crops and Products*, 42, 454-460.
- Raisi, A., Kalat, S. M. N., & Darban, A. R. S. (2013). The study effects of stratification, temperature and potassium nitrate on seed dormancy breaking *Ferula assa-foetida*. *World Applied Sciences Journal*, 21(3), 379-383.
- Ramzan, A., Hafiz, L. A., Ahmad, T., & Abbasi, N. A. (2010). Effect of priming with potassium nitrate and dehusking on seed germination of gladiolus (*Gladiolus alatus*). *Pakistan Journal of Botany*, 42(1), 247-258.
- Rasheed, R. (2009). *Salinity and extreme temperature effects on sprouting buds of sugarcane (Saccharum officinarum L.); some histological and biochemical studies*. (Doctoral dissertation, University of Agriculture, Faisalabad, Pakistan).
- Ratnakar, A., & Rai, A. (2013). Effect of sodium chloride salinity on seed germination and early seedling growth of *Trigonella foenum-graecum* L. var PEB. *Octa Journal of Environmental Research*, 1(4), 304-309.
- Reddy, P. S., Jogeswar, G., Rasineni, G. K., Maheswari, M., Reddy, A. R., Varshney, R. K., & Kishor, P. B. K. (2015). Proline overaccumulation alleviates salt stress and protects photosynthetic and antioxidant enzyme activities in transgenic sorghum (*Sorghum bicolor* (L.) Moench). *Plant Physiology Biochemical*, 94, 104-113.

- Rhoades, J. D. (1993). Electrical conductivity methods for measuring and mapping soil salinity. In Sparks, D. L. (Ed.) (pp 201-251). *Advances in Agronomy*, 49.
- Römheld, V., & Kirkby, E. A. (2010). Research on potassium in agriculture: needs and prospects. *Plant Soil*, 335, 155–80.
- Rouhi, H. R., Ali, A. S., Farzad, S. Z., Reza, T. A., Mohammad, A. A., & Goudarz, A. (2011). Study of different priming treatments on germination traits of soybean seed lots. *Notulae Scientia Biologicae*, 3, 101–108.
- Ruffino, A. M. C., Rosa, M., Hilal, M., Gonzalez, J. A., & Prado, F. E. (2009). The role of cotyledon metabolism in the establishment of quinoa (*Chenopodium quinoa*) seedlings growing under salinity. *Plant Soil*, 326, 213-224.
- Ruzicka, K., Strader, L. C., & Bailly, A. (2010). *Arabidopsis* PIS encodes the ABCG3 transporter of the auxinic compounds including the auxin precursor indole-3 butyric acid. *Proceedings of the National Academy of Sciences, USA* 107, 10749-10753.
- Sadeghi, H., & Robati, Z. (2015). Response of *Cichorium intybus* L. to eight seed priming methods under osmotic stress conditions. *Biocatalysis and Agricultural Biotechnology*, 4, 443-448.
- Samar, J. M. (2017). *Germination, seedling growth, and anatomical responses of Cucumis sativus cv. MTI2 in different salts and development of germination enhancer*. (Master's thesis, Universiti Putra Malaysia).
- Sarihan, E. O., Ipek, A., Khawar, K. M., Atak, M., & Gurbuz, B. (2005). Role of GA₃ and KNO₃ in improving the frequency of seed germination in *Plantago lanceolata* L. *Pakistan Journal of Botany*, 37(4), 883-887.
- Sarker, B., & Rahim, A. (2013). Yield and quality of mango (*Mangifera indica* L.) as influenced by foliar application of potassium nitrate and urea. *Bangladesh Journal of Agricultural Research*, 38(1), 145-154.
- Sarwar, M. (2012). Effects of potassium fertilization on population build up of rice stem borers (lepidopteron pests) and rice (*Oryza sativa* L.) yield. *Journal of Cereals Oilseeds*, 3, 6-9.
- Sekara, A., Cebula, S., & Kunicki, E. (2007). Cultivated eggplants: origin, breeding objectives and genetic resources, a review. *Folia Horticulturae*, 19(1), 97-114.
- Sauer, M., Robert, S., & Kleine-Vehn, J. (2013). Auxin: simply complicated. *Journal of Experimental Botany*, 64(9), 2565-2577.
- Sevik, H., & Guney, K. (2013). Effects of IAA, IBA, NAA and GA₃ on rooting and morphological features of *Melissa officinalis* L. stem cuttings. *The Scientific World Journal*, 2013, 1-5.

- Sevik, H., & Cetin, M. (2016). Effects of some hormone applications on germination and morphological characters of endangered plant species *Lilium arvinense* L. onion scales. *Bulgarian Chemical Communications*, 48(2), 256-260.
- Shabala, S. (2013). Learning from halophytes: physiological basis and strategies to improve abiotic stress tolerance in crops. *Annals of Botany*, 112, 1209-1221.
- Shabala, S., & Munns, R. (2012). Salinity Stress: Physiological Constraints and Adaptive Mechanisms. In S. Shabala, (Ed.), *Plant Stress Physiology* (pp. 59-93). United Kingdom: CABI.
- Shaheen, S., Naseer, S., Ashraf, M., & Akram, N. A. (2013). Salt stress affects water relations, photosynthesis and oxidative defense mechanisms in *Solanum melongena* L. *Journal of Plant Interactions*, 8(1), 85-96.
- Shamshuddin, J., Azura, A. E., Shazana, M. A. R. S., Fauziah, C. I., Panhwar, Q. A., & Naher, U. A. (2014). Properties and measurement of acid sulfate soils in Southeast Asia for sustainable cultivation of rice, oil palm, and cocoa. *Advance Agronomy*, 124, 91-142.
- Sharma, K. K., Saikia, R., Kotoky, J., Kalita, J. C., & Devi, R. (2011). Antifungal activity of *Solanum melongena* L., *Lawsonia inermis* L. and *Justicia gendarussa* B. against Dermatophytes. *International Journal of PharmaTech Research*, 3(3), 1635-1640.
- Sharma, A. D., Rathore, S. V. S., Srinivasan, K., & Tyagi, R. K. (2014). Comparison of various seed priming methods for seed germination, seedling vigour and fruit yield in okra (*Abelmoschus esculentus* L. Moench). *Scientia Horticulturae*, 165, 75-81.
- Shaul, O. (2002). Magnesium transport and function in plants: the tip of the iceberg. *Biology of Metals*, 15, 309-323.
- Sheldon, A. R., Dalal, R. C., Kirchoff, G., Kopittke, P. M., & Menzies, N. W. (2017). The effect of salinity on plant-available water. *Plant Soil*, 418, 477-491.
- Sosa, L., Llanes, A., Reinoso, H., Reginato, M. & Luna, V. (2005). Osmotic and specific ion effects on the germination of *Prosopis strombulifera*. *Annals of Botany*, 96, 261-267.
- Stefancic, M., Stampar, F., & Osterc, G. (2005). Influence of IAA and IBA on root development and quality of Prunus "GiSelA5" leafy cuttings. *HortScience*, 40(7), 2052-2055.
- Suhana, O., Mohamad, O., Abd Rahman, M., & Zubaid Akbar, M. A. (2016). Genetics of yield and its components in eggplant (*Solanum melongena* L.). *Journal of Tropical Agriculture and Field Science*, 44(1), 137-145.

- Sukifto, M. R. (2016). *Comparative study on seed germination of Malaysian lowland tomato (*Solanum lycopersicon* L. cv MT1) under different types of salinity stress*. (Degree's thesis, Universiti Putra Malaysia).
- Sultana, B., Hussain, Z., Hameed, M., & Mushtaq, M. (2013). Antioxidant activity among different parts of aubergine (*Solanum melongena* L.). *Pakistan Journal of Botany*, 45(4), 1443-1448.
- Szabolcs, I. (1989). *Salt Affected Soils*. Boca Raton, FL: CRC Press.
- United States Department of Agriculture, (1954). Diagnosis and improvement of saline and alkali soils. In *Agriculture Handbook No 60*.
- Tanji, K. K. (1990). (ed). *Agricultural salinity assessment and management*. ASCE manuals and reports on engineering practice No. 71. New York: American Society of Civil Engineering.
- Tao, L., van Staden J. & Cress, W. A. (2000). Salinity induced nuclear and DNA degradation in meristematic of soybean (*Glycine max* L.) roots. *Plant Growth Regulator*, 30, 49-54.
- Teh, C. Y., Maziah, M., Noor, A. S., & Chai, L. H. (2015). In vitro rice shoot apices as simple model to study the effect of NaCl and the potential of exogenous proline and glutathione in mitigating salinity stress. *Plant Growth Regulator*, 75, 771-781.
- Todaro, A., Cimino, F., Rapisarda, P., Catalano, A. E., Barbagallo, R. N. & Spagna, G. (2009). Recovery of anthocyanins from eggplant peel. *Food Chemistry*, 114, 434-439.
- Topacoglu, O., Sevik, H., Guney, K., Unal, C, Akkuzu, E., & Sivacioglu, A. (2016). Effect of rooting hormones on the rooting capabilities of *Ficus benjamina* L. cuttings. *Sumarski List*, 1(2), 39-44.
- Tsegay, B., & Gebreslassie, B. (2014). The effect of salinity (NaCl) on germination and early seedling growth of *Lathyrus sativus* and *Pisum sativum* var. abyssinicum. *African Journal of Plant Science*, 8(5), 225-231.
- USDA. (1954). *Reclamation and improvement of saline and sodic soils*. USDA Handbook 60, Riverside, California.
- Ventura, L., Dona, M., Macovei, A., Carbonera, D., Buttafava, A., Mondoni, A., ... Balestrazzi, A. (2012). Understanding the molecular pathways associated with seed vigor. *Plant Physiology and Biochemistry*, 60, 196-206.
- Vinocur, B., & Altman, A. (2005). Recent advances in engineering plant salt tolerance to abiotic stress: achievements and limitations. *Current Opinion in Biotechnology*, 16, 123-132.

- Vinod. K. K., Krishnan, S. G., Babu, N. N., Nagarajan, M., & Singh, A. K. (2013). Improving salt tolerance in rice: looking beyond the conventional. In P. Ahmad, M. M. Azooz, & M. N. V. Prasad (Eds), *Salt Stress in Plants: Signalling, Omics and Adaptations* (pp. 219-260). New York: Springer Science+Business Media.
- Wahid, A., Farooq, M., Basra, S. M. A., Rasul, E., & Siddique, K. H. M. (2010) Germination of seeds and propagules under salt stress. In Handbook of plant and crop stress. Boca Raton: CRC Press.
- Wang, H., Xu, R. K., Wang, N., & Li, X. H. (2010). Soil acidification of Alfisols as influenced by tea plantation in eastern China. *Pedosphere*, 20, 799-806.
- Wang, M., Zheng, Q., Shen, Q., & Guo, S. (2013). The critical role of potassium in plant stress response. *International Journal of Molecular Science*, 14(4), 7370-7390.
- Wang W., Liu, S., Song, S., & Moller, I. A. (2015). Proteomics of seed development, desiccation tolerance, germination and vigor. *Plant Physiology and Biochemistry*. 86, 1-15.
- Wang, P., Mo, B., Long, Z., Fan, S., & Wang, H. (2016). Factors affecting seed germination and emergence of *Sophora davidii*. *Industrial Crops and Products*, 261-265.
- Weese, T. L., & Bohs, L. (2010). Eggplant origins: out of Africa, into the orient. *Taxon*, 59(1), 3448-54.
- Wojtyla, L., Lechowska, K., Kubala, S., & Garnczarska, M. (2016). Molecular processes induced in primed seeds-increasing the potential to stabilize crop yields under drought conditions. *Journal of Plant Physiology*, 203, 116-126.
- Wong, N. C., & Jaafar, A. M. (1993). *Soil chemical characteristics of vegetable plots at Cameron Highlands*. MARDI Report no. 170.
- Wong, N. C., Lee, B. S., Yuen, P. M., Wan Abdullah, W. Y., & Mohd Ridzuan, S. (2002). Effects of continuous mono-cropping of chrysanthemum flowers under rain-shelter on soil quality. In Shamshuddin, J., Hamdan, J. & Samsuri, A. W., (Eds). "*Sustainable Land Mangement*", Malaysian Society of Soil Science, 236-252.
- Yousuf, B., Gul, K., & Wani, A. A. (2016). Health benefits of anthocyanins and their encapsulation for potential use in food systems: a review. *Critical Reviews in Food Science and Nutrition*, 56(13), 2223-2230.
- Zeng, F., Shabala, L., Zhou, M., Zhang, G., & Shabal, S. (2013). Barley responses to combined waterlogging and salinity stress: separating effects of oxygen deprivation and elemental toxicity. *Frontiers in Plant Science*, 4(313), 1-13.
- Zhang, L., Ma, H., Chen T., Pen, J., Yu, S., & Zhao, X. (2014). Morphological and physiological responses of cotton (*Gossypium hirsutum* L.) plants to salinity. *PLoS ONE*, 9(11), 1-14.

- Zhu, H., & Banuelos, G. (2016). Influence of salinity and boron on germination, seedling growth and transplanting mortality of guayule: a combined growth chamber and greenhouse study. *Industrial Crops and Products*, 92, 236-243.
- Zinck, J. A., & Metternicht, G. (2009). Soil salinity and salinization hazard. In Metternicht G., & Zinck J.A. (Eds). *Remote Sensing of Soil Salinization: Impact on Land Management* (pp. 3-5). Boca Raton, FL: CRC Press Taylor and Francis Group.
- Zorb, C., Senbayram, M., & Peiter, E. (2014). Potassium in agriculture. *Journal of Plant Physiology*, 17, 656-669.

