

CHARACTERIZATION OF Fusarium Species ASSOCIATED WITH WILT DISEASE OF CUCURBITS AND EFFECTIVENESS OF Trichoderma asperellum AS BIOLOGICAL ENHANCER

ASMA ARIS

FS 2019 48



CHARACTERIZATION OF *Fusarium* Species ASSOCIATED WITH WILT DISEASE OF CUCURBITS AND EFFECTIVENESS OF *Trichoderma asperellum* AS BIOLOGICAL ENHANCER

By

ASMA ARIS

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

August 2019

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

G



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

CHARACTERIZATION OF Fusarium Species ASSOCIATED WITH WILT DISEASE OF CUCURBITS AND EFFECTIVENESS OF Trichoderma asperellum AS BIOLOGICAL ENHANCER

By

ASMA ARIS

August 2019

Chairman: Assoc. Prof. Nur Ain Izzati Mohd Zainudin, PhD Faculty: Science

Fusarium wilt disease caused by Fusarium species is a serious soilborne fungal disease that threatens the production of cucurbits in Malaysia. Despite multiple controls and measures, this economically important pathogenic genus continues intruding the crops. Therefore, this study intends to identify the pathogenicity of *Fusarium* species from five infected hosts namely cucumber, pumpkin, luffa, gourd and rock melon. Infected cucurbits fruit and leaf obtained from the fields were cultured onto pentachloronitrobenzene agar (PCNB). All isolated fungi were purified using a hyphal tip technique and sub-cultured onto potato dextrose agar (PDA) and carnation leaf agar (CLA) for observing morphological characteristics such as macroconidia, microconidia, formation of chlamydospore, colony features and pigmentation. Ninety-four isolates were successfully recovered from 12 locations throughout Peninsular Malaysia. Isolated fungi were also identified molecularly using translation elongation factor 1-alpha (tefla) and beta-tubulin (BT) genes sequences analysis. From the findings, the highest number of identified species were F. incarnatum (52 isolates) followed by F. solani (18 isolates), F. proliferatum (17 isolates), F. oxysporum (6 isolates) and F. longipes (1 isolate). A phylogenetic tree was generated from a combination of $tefl\alpha$ and β -Tub genes sequences. To ascertain the virulence of Fusarium species, all isolates were tested for pathogenicity test by soaking the seeds into $2x10^6$ conidia/mL of conidial suspension for 12 hours. Sterile distilled water served as control. Fusarium solani isolates M1799C and M1800C recorded the highest disease severity of 100% on cucumber, F. solani (C2526P) recorded 93.8% disease severity on pumpkin, F. solani (D2499L) and F. proliferatum (B1777L) recorded 66.75% disease severity on luffa and F. oxysporum (B2547M) recorded 23% disease severity on rock melon. However, no symptoms observed on inoculated gourd plants. In order to eco-friendly control the disease and understand the mechanisms; therefore, Trichoderma asperellum B1902 was

testified as a potential plant biological enhancer in wilt-infected cucumber. Efficacy of *T. asperellum* B1902 was testified by infesting 100 mL/kg inoculum (with concentration $2x10^6$ conidia/mL) onto soil containing infected cucumber plants. After 30 days post-inoculation (dpi), *T. asperellum* B1902-treated plants increased length of stem, area of total leaves and number of leaves. Furthermore, *T. asperellum* B1902 also gave rise to the efficiency of photosystem II (PSII) and enhanced the photosynthetic performance of cucumber plants. The information channeled from this study leads a foundational knowledge for better management in controlling Fusarium wilt infection.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENCIRIAN species Fusarium BERKAIT DENGAN PENYAKIT LAYU PADA CUCURBIT DAN KEBERKESANAN Trichoderma asperellum SEBAGAI PENGGALAK BIOLOGIKAL

Oleh

ASMA ARIS

Ogos 2019

Pengerusi: Prof. Madya Nur Ain Izzati Mohd Zainudin, PhD Fakulti: Sains

Penyakit layu Fusarium disebabkan oleh Fusarium species adalah penyakit kulat bawaan tanah yang serius serta mengancam pengeluaran cucurbit di Malaysia. Walaupun pelbagai kawalan dan langkah dijalankan, genus patogenik yang mempunyai kepentingan ekonomi ini tetap menyerang tanaman cucurbit. Oleh itu, kajian ini bertujuan untuk menentukan kepatogenan spesies Fusarium daripada lima tanaman iaitu timun, labu, petola, labu air dan tembikai wangi. Buah dan daun cucurbit yang dijangkiti diperolehi dari lapangan dikultur di atas agar pentachloronitrobenzene (PCNB). Semua pencilan kulat ditulenkan melalui teknik hujung hifa dan sub-kultur pada media agar kentang dekstrosa (PDA) dan agar daun carnation (CLA) bagi pencirian morfologi seperti makrokonidia, mikrokonidia, pembentukan klamidospora, corak koloni dan pigmentasi. Sembilan puluh empat pencilan berjaya diperoleh daripada 12 lokasi seluruh Semenanjung Malaysia. Pencilan kulat juga dikenalpasti secara molekular menggunakan analisis jujukan gen translation elongation factor 1alpha (tefla) dan beta-tubulin. Daripada penemuan ini, spesis tertinggi dikenalpasti adalah F. incarnatum (52 pencilan), diikuti dengan F. solani (18 pencilan), F. proliferatum (17 pencilan), F. oxysporum (6 pencilan) dan F. longipes (1 pencilan). Filogenetik dendogram telah dihasilkan daripada kombinasi jujukan gen $tefl\alpha$ dan β -Tub. Bagi menentukan kevirulenan spesis Fusarium, kesemua pencilan diuji dengan ujian kepatogenan dengan merendam biji benih ke dalam ampaian konidia dengan kepekatan 2x10⁶ conidia/mL selama 12 jam. Air suling steril digunakan sebagai kawalan. Fusarium solani pencilan M1799C dan M1800C mencatatkan keparahan penyakit tertinggi iaitu 100% pada timun, F. solani (C2526P) mencatat 93.8% keparahan penyakit pada labu, F. solani (D2499L) dan F. proliferatum (B1777L) mencatat 66.75% keparahan penyakit pada petola dan F. oxysporum (B2547M) mencatat 23% keparahan penyakit pada tembikai wangi. Namun, tiada simptom diperhatikan pada labu air yang diinokulasi. Untuk mengawal penyakit ini secara mesraekonomi dan memahami mekanisme tersebut; maka Trichoderma asperellum B1902 telah diuji sebagai penggalak biologikal tumbuhan yang berpotensi terhadap timun layu

terinfeksi. Keberkesanan *T. asperellum* B1902 diuji dengan infestasi 100mL/kg inokulum (pada kepekatan 2x10⁶ conidia/mL) ke dalam tanah berisi pokok timun terinfeksi. Setelah 30 hari selepas inokulasi (hsi), panjang tangkai, luas keseluruhan dan bilangan daun meningkat pada pokok timun dirawat dengan *T. asperellum* B1902. Tambahan lagi, *T. asperellum* B1902 juga meningkatkan kecekapan fotosistem II (PSII) dan menggalakkan prestasi fotosintetik pokok timun. Informasi yang disalurkan daripada kajian ini menyumbang pengetahuan berkenaan pengurusan dalam pengawalan penyakit layu Fusarium.



ACKNOWLEDGEMENT

I would like to express my foremost appreciation to Assoc. Prof. Dr. Nur Ain Izzati Mohd Zainudin as my supervisor. Thank you for all the guidance, knowledge, effort and opportunities you lead throughout the research. I would also like to express my gratitude to Dr. Noor Baity Saidi and Assoc. Prof. Dr. Shamarina Shohaimi as my cosupervisors. I really appreciate all the motivation, contribution on succeeds every experiment and information shared.

Nevertheless, I would like to take this opportunity to thank everyone who lends their contribution and facilitation either directly or indirectly throughout my study. Thank you to all academic staffs, laboratory assistants and supporting staffs Department of Biology, Faculty of Science and Department of Molecular and Cell Biology, Faculty of Biotechnology and Biomolecular, UPM. I would also like to thank my fellow doctoral students, mycology and plant molecular biology laboratory members for their feedback, cooperation and information.

I would like to acknowledge UPM for providing me sponsorships under Fundamental Research Grant Scheme (FRGS) and Ministry of Higher Education (MOHE) for providing me sponsorship under MyPhD scholarship.

Last but not least, my special dedication to my husband Dr. Sopian Abdul Wahab for the love and support. Special thanks to my beloved parents, Aris Tayib and Norasikin Rosdi for constant moral support and patience. Thank you to all proud family members. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Nur Ain Izzati Mohd Zainudin, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Chairman)

Shamarina Shohaimi, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Member)

Noor Baity Saidi, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Science Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, Phd

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 17 October 2019

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including boos, journals, modules, proceedings, popular writing, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature	
Signature	

Date:

Name and Matric No. : <u>Asma Aris, GS44452</u>

Declaration by Members of Supervisory Committee

This is to confirm that:

6

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2013 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Assoc. Prof. Dr. Nur Ain Izzati Mohd Zainudin
Signature: Name of Member of Supervisory Committee:	Assoc. Prof. Dr. Shamarina Shohaimi
Signature: Name of Member of Supervisory Committee:	Dr. Noor Baity Saidi

TABLE OF CONTENTS

ABSTRACT ABSTRAK ACKNOWLF APPROVAL DECLARATT LIST OF TAI LIST OF FIG LIST OF ABI	EDGEM ION BLES FURES BREVIA	ENTS	Page i iii v vi viii xiii xiii xiv xvii
CHAPTER	INT		
1		RODUCTION Packground of study	1
	1.1	Basearch objective	1
	1.2	Research objective	+
2	LIT	ERATURE REVIEW	
-	2.1	Status of Crop Disease in Malaysia	5
		2.1.1 Fusarium wilts in Malaysia	5
	2.2	The Importance of genus <i>Fusarium</i> in ecosystems	6
		2.2.1 Morphological characteristics of <i>Fusarium</i>	7
		species	
		2.2.2 Genetic diversity of <i>Fusarium</i> species	8
		2.2.3 Fusarium wilt disease	10
	2.3	Production and importance of cucurbits	13
		2.3.1 Cucumber (<i>Cucumis sativus</i> Linnaeus)	14
		2.3.2 Pumpkin (<i>Cucurbita pepo</i> Linnaeus)	15
		2.3.3 Luffa (<i>Luffa acutangula</i> L. Roxb.)	16
		2.3.4 Rock melon (<i>cucumis melo</i> Linnaeus)	17
		2.3.5 Gourd (<i>Lagenaria siceraria</i> Molina Standl.)	18
	2.4	The genus Trichoderma	19
		2.4.1 Characterization of <i>Trichoderma</i> species	21
		2.4.2 <i>Inchoderma</i> as a biocontrol agent	22
		2.4.2.1 Wiorphogenesis and development	22
		2.4.2.2 Nycoulopity 2.4.2.3 Symbiosis and endophytism	23 23
		2.4.2.4 Competing for nutrients and space	23
		2.4.2.5 Promoting plant growth	24 24
		2.4.2.6 Enhancing plant defense	24
		mechanisms	24
		2.4.2.7 Antibiosis	25

 \bigcirc

3

MORPHOLOGICAL AND MOLECULAR CHARACTERIZATION OF *FUSARIUM* SPECIES ISOLATED FROM WILT DISEASE OF CUCURBITS

3.1	Introduction 20		
3.2 Materia		als and Methods	27
	3.2.1	Source of cucurbits samples	27
	3.2.2	Fungal isolation, purification and preservation	30
	3.2.3	Microscopic characteristics of <i>Fusarium</i> species	30
	3.2.4	Macroscopic characteristics of <i>Fusarium</i> species	30
	3.2.5	Fungal DNA extraction	30
	3.2.6	Polymerase chain reaction (PCR)	31
	3.2.7	Gel electrophoresis	32
	3.2.8	Nucleotide sequencing and phylogenetic analysis	32
	3.2.9	Sequence deposition	32
3.3	Results	s and Discussion	33
	3.3.1	Morphological characteristics of <i>Fusarium</i>	33
	3.3.2	Molecular identification of <i>Fusarium</i> species associated with wilt of cucurbits in Peninsular Malaysia	42
	3.3.3	Fusarium species diversity	50
3.4	Conclu	ision	53

4

PATHOGENICITY OF *FUSARIUM* SPECIES ISOLATED FROM CUCURBITS IN PENINSULAR MALAYSIA

4.1	Introdu	action	54
4.2	Materi	als and Methods	55
	4.2.1	Conidial suspension and plant preparation	55
	4.2.2	Seed inoculation	57
	4.2.3	Disease assessment	58
4.3	Result	s and Discussion	59
	4.3.1	Pathogenicity of Fusarium isolates on	60
		cucumber	
	4.3.2	Pathogenicity of Fusarium isolates on	62
		pumpkin	
	4.3.3	Pathogenicity of Fusarium isolates on luffa	64
	4.3.4	Pathogenicity of Fusarium isolates on rock	66
		melon	
	4.3.5	Pathogenicity of Fusarium isolates on gourd	67
	4.3.6	Occurrence of Fusarium virulent strains on	69
		cucurbits	
4.4	Conclu	ision	72

5	BIOLOGICAL CHANGES OF FUSARIUM WILT			
	INFECTED Cucumis sativus Linnaeus TREATED			
	WITH Trichoderma asperellum B1902			
	5.1	Introduc	tion	73
	5.2	Material	s and Methods	74
		5.2.1	Fungal strains and inoculum preparation	74
		5.2.2	Plant materials	75
		5.2.3	Fungal inoculation	75
		5.2.4	Gas exchange measurements	76
		5.2.5	Chlorophyll fluorescence measurements	76
	5.3	Results a	and Discussion	76
		5.3.1	Biological changes of infected-cucumber	76
			caused by T. asperellum B1902	
		5.3.2	Photosynthetic performance of wilt infected	79
			cucumber treated with <i>T. asperellum</i> based	
			on chlorophyll fluorescence.	
		5.3.3	Photosynthetic performance of wilt infected	84
			cucumber treated with T. asperellum based	
			on gas exchange measurements	
		5.3.4	Interaction between cucumber plant	86
			treatments and biological parameters	
	5.4	Conclus	ion	87
6	SUM	MARY,	GENERAL CONCLUSION AND	
	REC	OMMEN	DATION FOR FUTURE RESEARCH	
	6.1	Summar	y and General Conclusion	88
	6.2	Recomn	nendation For Future Research	89
REFERENCE	S			90
APPENDICES				111
BIODATA OF	STUD	ENT		162
LIST OF PUB	LICAT	IONS		163

xii

 \mathbf{G}

LIST OF TABLES

Table		Page
3.1	Type of sample from various locations throughout Peninsular Malaysia.	28
3.2	Distribution of <i>Fusarium</i> species isolated from five cucurbits in twelve locations of Peninsular Malaysia	51
4.1	Total isolates tested for pathogenicity on cucurbits according to representative host.	55
4.2	Disease scales for Fusarium wilt assessment.	59
4.3	Disease severity of <i>Fusarium</i> isolates on cucumber in 30 days.	60
4.4	Disease severity of <i>Fusarium</i> isolates on pumpkin in 30 days.	62
4.5	Disease severity of <i>Fusarium</i> isolates on luffa in 30 days.	64
4.6	Disease severity of <i>Fusarium</i> isolates on rock melon in 30 days.	66
4.7	Disease severity of Fusarium isolates on gourd in 30 days	68
4.8	<i>Fusarium</i> virulent strains on cucurbits	70
5.1	Cucumber plant treatments	76
5.2	Plant biological parameters of cucumber plants	78
5.3	Leaf gas exchange measurements of cucumber plants	85
5.4	Cumulative score of cucumber plant treatments	86

 \mathbf{G}

LIST OF FIGURES

Figure		Page
3.1	Distribution of sampling locations throughout Peninsular Malaysia	29
3.2	Micromorphological characteristics of <i>F. oxysporum</i> . A: colony cultures, superscripts a and b indicating colony features (^a) and pigment in media (^b), B: macroconidia, C: microconidia, D: false head microconidia and E: chlamydospore ($-$: 20 µm)	35
3.3	Micromorphological characteristics of <i>F. solani</i> . A: colony cultures, superscripts a and b indicating colony features (^a) and pigment in media (^b), B: macroconidia C: microconidia, D: false head microconidia and E: chlamydospore ($-: 20$ um)	37
3.4	Micromorphological characteristics of <i>F. longipes</i> . A: colony cultures, superscripts a and b indicating colony features (^a) and pigment in media (^b), B: macroconidium and C: macroconidia $(-: 20 \text{ um})$	38
3.5	Micromorphological characteristics of <i>F. incarnatum</i> . A: colony cultures, superscripts a and b indicating colony features (^a) and pigment in media (^b), B: <i>in situ</i> macroconidia, C: macroconidia and D: chained chlamydospores. ($-$: 20 µm).	40
3.6	Micromorphological characteristics of <i>F. proliferatum</i> . A: colony cultures, superscripts a and b indicating colony features (^a) and pigment in media (^b), B: macroconidium, C: microconidia and D: chained microconidia ($-$: 20 µm).	42
3.7	Percentage of the total number of <i>Fusarium</i> species identified by molecular identification using $tefl\alpha$ and β -tubulin sequences analysis.	43
3.8	Banding pattern of <i>tef1a</i> region. A: 22 bands of isolate D2496L to C2518G, B: 20 bands of isolate C2519G to B2540M, C: 22 bands of isolate B2541M to B1781P, D: 20 bands of isolate B1782P to M1801C and E: 10 bands of isolate M1802C to N2216C.	44
3.9	Banding pattern of β -tubulin region. A: 20 bands of isolate D2496L to C2516G, B: 22 bands of isolate C2517G to B2540M, C: 17 bands of isolate B2541M to B2567M, D: 23 bands of isolate B1777L to M1799C and E: 12 bands of isolate M1800C to N2216C.	45
3.10	Phylogenetic tree generated by Maximum Likelihood (ML) analysis of <i>Fusarium</i> isolates of <i>tef1</i> α and β -tubulin gene sequences by using Tamura-Nei model. Alphabets denote the condensed sub-trees. <i>Aspergillus niger</i> CBS513.88 is outgroup.	47
3.11	Expanded sub-tree of cluster A of <i>Fusarium incarnatum</i> . Bootstrap values $\geq 80\%$ are labelled on the branches.	48

 \bigcirc

3.12	Expanded sub-tree of cluster B of <i>Fusarium solani</i> .	49
3 13	Bootstrap values $\geq 80\%$ are labelled on the branches. Expanded sub-tree of cluster C of <i>Fusarium proliferatum</i>	50
5.15	Bootstrap values $\geq 80\%$ are labelled on the branches.	50
4.1	Cucurbits seeds used from Green World Genetics brand. A:	58
	cucumber, B: pumpkin, C: gourd and D: luffa. Cucurbit from	
	Sakata Corporation. E: rock melon.	
4.2	Cucumber roots and plants inoculated with A and G: dH ₂ O	61
	(control), B and H: F. longipes D2504C, C and I: F.	
	oxysporum D2505C, D and J: F. solani M1799C, E and K:	
	<i>F. proliferatum</i> N2215C, F and L: <i>F. incarnatum</i> N2213C	
43	Pumpkin roots and plants inoculated with Fusarium isolates	63
1.5	and non-inoculated plants. A and F: dH ₂ O (control), B and	05
	G: F. oxysporum D2532P. C and H: F. solani C2526P. D	
	and I: F. proliferatum J1793P, E and J: F. incarnatum	
	C2522P on 30 dpi.	
4.4	Luffa roots and plants inoculated with; A and F: dH ₂ O	65
	(control), B and G: F. oxysporum B1783L, C and H: F.	
	solani D2499L, D and I: F. proliferatum B1777L, E and K:	
1.5	F. incarnatum D2496L on 30 dpi	
4.5	Rock melon plants inoculated with A: dH_2O (control), B: F.	6/
	in competition K1706M on 20 dpi	
4.6	Gourd roots inoculated with A and D: dH_2O (control) B and	69
т.0	E: F proliferatum C2518G and C and E: F incarnatum	07
	C2512G on 30 dpi.	
5.1	Trichoderma asperellum B1902 on PDA. A: colony feature	75
	and B: colony pigmentation	
5.2	Cucumber plants after 30 dpi. A: control (distilled water), B:	77
	treated with T. asperellum B1902, C: treated with T.	
	asperellum B1902 and F. solani M1799C and D: treated	
5.0	with F. solani M1799C.	0.0
5.3	Maximum efficiency of photosystem II (Fv/fm) of cucumber	80
	plants treated with <i>I. asperellum</i> B1902. The data are snown	
	as the mean \pm SD of five replicates. The significant differences (P < 0.05) among the treatments are indicated by	
	different superscripts	
5.4	Maximum vield of photosystem II (fv/fo) of cucumber plants	81
	treated with <i>T. asperellum</i> B1902. The data are shown as the	-
	mean \pm SD of five replicates. The significant differences (P	
	< 0.05) among the treatments are indicated by different	
	superscripts.	
5.5	Minimal fluorescence (fo) of cucumber plants treated with <i>T</i> .	82
	asperellum B1902. The data are shown as the mean \pm SD of	
	tive replicates. The significant differences ($P < 0.05$) among	
	the treatments are indicated by different superscripts.	

xv

- 5.6 Performance index (PI) of cucumber plants treated with *T*. *asperellum* B1902. The data are shown as the mean \pm SD of five replicates. The significant differences (P < 0.05) among the treatments are indicated by different superscripts.
- 5.7 Density of reaction centres per PSII antenna chlorophyll (RC/ABS) of cucumber plants treated with *T. asperellum* B1902. The data are shown as the mean \pm SD of five replicates. The significant differences (P < 0.05) among the treatments are indicated by different superscripts.



83

LIST OF ABBREVIATIONS

%	Percentage
⁰ C	Degree Celsius
$\mu g \ cm^{-2} s^{-1}$	Microgram per centimeter square per second
μL	Microliter
ΦPSII	Relative quantum efficiency of PSII
хд	Relative centrifugal/gravitational force
ABA	Abscisic acid
ANOVA	Analysis of variance
BC	Before century
bp	Base pair
CH ₃ CN	Acetonitrile
СНООН	Formic acid
CLA	Carnation leave agar
cm	Centimeter
cm ²	Centimeter square
CRD	Complete randomized design
DNA	Deoxyribonucleic acid
dpi	Day post inoculation
DSI	Disease severity index
Foc	Fusarium oxysporum f. sp. cucumerinum
Fm	Maximum fluorescence
Fv	Variable fluorescence
Fv/fm	Maximum efficiency of photosystem II
Fv/fo	Maximum yield of photosystem II
Fo	Minimal fluorescence
f. sp.	Forma speciales
g	Gram
ha	Hectare
H ₂ O	Water
hr	Hour
JA	Jasmonic acid
KCl	Potassium chloride
kg	Kilogram
LHCs	Light harvesting complexes
nm	Nano meter
Min	Minute
mg	miligram
mL	Milliliter
mm	milimeter
mM	Milimolar
Mt	Million tonne
O_2	Oxygen
L DCD/ID	Probability
PUNB	Pentachloronitrobenzene agar
PDA	Potato dextrose agar
PDB	Potato dextrose broth

6

PFD	Photon flux density
PGPF	Plant growth promoting fungi
PI	Performance index
PSII	Photosystem II
RC/ABS	density of reaction centres per PSII antenna chlorophyll
ROS	Reactive oxygen species
S	Second
SA	Salicylic acid
scm ⁻¹	Second per centimeter
SD	Standard deviation
TBE	Tris-Borate-EDTA buffer
UFLC	Ultra-fast liquid chromatography
WA	Water agar



 \bigcirc

CHAPTER 1

INTRODUCTION

1.1 Background of study

Agriculture provides people with food, where it contributes 7.5% total output of Malaysia economy and making it the fourth highest sector in the country. Malaysia recorded 2.9%, the lowest agriculture growth rate in Southeast Asia together with Thailand and the Philippines (Birthal, Joshi, Roy and Pandey, 2019). As an agricultural-based economy country, it is important to look profoundly into any factors that could enhance productivity. United Nation Food and Agriculture Organization (FAO, 2012), reported that the demand for agricultural products is expecting to be up to 1.1% every year until 2050. On the other hand, knowledge is the key to manage good agriculture. Lack of specialized information could lead to an abandonment of farming and inappropriate agriculture and subsequently affect the biodiversity (Ruxandra and Dacinia, 2010).

The consumerism of crops belong to Cucurbitaceae family is increasing together with the world populations. Among all cucurbits, melon is the most popular crop in the family, followed by cucumber, squash, and pumpkin (Bisognin, 2002). Since the past decades, outnumber of studies revealed the advantages of the crops. This soft-vined plant growing by means of crawling on any surfaces supplies multiple utilizability and essentials. The seed especially has a high source of lipid, and proteins (Mladenovic, Berenji, Ognjanov, Ljubojevic and Cuanovic, 2012) frequently being processed to be used as organic cooking oil (McCreight, Staub, Wehner and Dhillon, 2013). Pharmaceutically, the leave, flower and fruit provide essentials in reducing human health risks such as treatment of hemorrhages in the internal organs, epilepsy, nerve disease, high antioxidants and anti-inflammatory properties. The high contents of β -carotene also can lower the risk of heart attack and cancer (Gabriele, Alberto, Sergio, Fernanda and Marco, 2000; Ismail, Chan, Mariod and Ismail, 2010; Avinash and RavishankarRai, 2013).

In Malaysia, cucumber is the leading crop being produce among any other cucurbits cultivated throughout the country. According to Ministry of Agriculture Malaysia (MOA, 2018), the production of cucumber in 2018 was up to 94, 520 Mt followed by pumpkin 25, 290 Mt, luffa 18, 782 Mt, gourd 5, 413 Mt and rockmelon 3,928.4 Mt. From all these, Peninsular Malaysia produces more hectareage and harvested areas comparing to Sabah and Sarawak. As cucurbits are originally from Western Asia (Deyo and O'Malley, 2008) and essentially a group of tropical plants (Pessarakli, 2016), the cultivation of the crop is compatible in almost every state in Malaysia. The cultivation condition favors to tropical and subtropical regions (Norrizah, Hashim, Siti Fasiha and Yaseer, 2012).

However, several factors such as Phytium crown and root rot, powdery mildew and stem blight (Punja, Tirajoh, Collyer and Ni, 2019; Ishii, Fujiwara and Nishimura,

2018) were found to be contributed and destroying the cucurbit plantations. On top of these factors is a fungal infection caused by *Fusarium* species. Among all plant pathogens, fungi and fungi-like organisms contribute more plant diseases compared to any other group of plant pests (Ellis, Boehm and Mitchell, 2008). *Fusarium oxysporum* was listed as the fifth out of the ten most important plant pathogens in the world (Sharma *et al.*, 2016). Among all the diseases, Fusarium wilt has the highest record of infecting most of the economically important crops. It is one of the oldest described diseases and the most economically important disease of many crops worldwide. It occurs in every continent except Antarctica and new races of the pathogen continue to impact production in many areas around the world. Long-term survival of the pathogen in the soil and the evaluation of new races make the management of Fusarium wilt difficult. The fungi enter through the roots and interfere with the water vessels of the plant. As the infection spreads up into the stems and leaves, it restricts water flow causing the foliage to wilt and yellow (Egel and Martyn, 2007).

Fusarium wilt causes various economic losses. It causes an obvious and direct loss in yield because the plant dies and subsequently brings loss to marketable yields (i.e. fruits that is low quality and cannot be sold because they are too small, misshapen, and low in sugar, cracked and sunburned). About \$253 million loss in Malaysia due to F. oxysporum tropical race 4 (TR4) invasions on banana (Ordonez et al., 2015). Fusarium wilt has also been a vast impact on the agriculture production of fruits and vegetables belong to the Cucurbitaceae family. Indirectly, these impacts increase the agricultural waste disposed into landfills annually by up to 1.2 million tonnes (Agamuthu, Hidzir and Hamid, 2009). Alternatively, several efforts and researchers had trying to overcome this disease invasion. This causative pathogen is a soil-borne fungus and chemical application could not control the disease entirely as it has possible secondary threat on the environment, human and animals (Minuto, Spadaro, Garibaldi and Gullino, 2006). Performing crop rotation, soil solarization, chemical fungicide application and fumigation (Tamietti and Valentino, 2006; Arguelles-Arias, Brans, Joris, and Fickers, 2009; Wang et al., 2013; Ghadikolaei, Cheung and Yung, 2019) which were strictly emphasized previously could not be succeeded as the conidia of the fungus could persistent and survive in the soil for a long period, in critical conditions and also compatible in a wide host range (Cao et al., 2011).

Thus, current information on Fusarium wilt infection is still unable to well manage to control the disease on cucurbits. Although many studies implied various information and alternative on how to reduce Fusarium wilt infection. But still, the infection pervasive persistently in the crop's soil. In fact, most of the studies have not targeted and aimed at the plant defense system as alternative measures to reduce this infection.

The phenomena and behavioral of *Fusarium* species isolated from cucurbits offer a broad significant to many aspects such as virulence, host specificity, toxin production, and functional characteristics. Hence, this study theorized that not all

species isolated from infected cucurbits is the causal pathogen that is responsible for Fusarium wilt infection. Hypothetically, *Fusarium oxysporum*, *F. solani* and *F. proliferatum* are dominantly pathogenic and highly virulent than any other species of the genus that infecting the plant vascular system. The same *Fusarium* species possess different pathogenicity, provided equal environmental condition and nutrient. As a resolution to the Fusarium wilt infection in cucurbits, *Trichoderma* spp. applied as a bio-enhancer onto infected cucumber plants revealed the enhanced photosynthetic performances.

This research study proposes the explanation and discussion on understanding and characterizing the *Fusarium* species. Apart from that, the information on genetic diversity and how the species evolution runs proportionally to time are relatively affected. This study also provides the mechanism and sequential process on how the pathogenesis occurs presenting distinct virulence. Finally, through the advanced technologies on plant physiology, the induction of bio-enhance in cucurbits plants in response to *Fusarium* infection was discovered by analyzing the photosynthetic measurements of cucurbit plants by the application of *T. asperellum*.

Therefore, following with the same mission, this study generally provides updated and additional information related to *Fusarium* species. From the first objective of the study, the findings generally introduced the very basic of all organisms by delivering the picture and visualize how diverse the fungal species associated with cucurbits. The rationale behind this objective is that morphological features are mutually related to a particular function and reactions of the fungus. "*It's one thing to make a picture of what a thing looks like, it's another thing to make a picture of what they are*", quoted by well-known American photographer, Paul Caponigro. Visualize the *Fusarium* characteristics explains us more about the organism. Besides that, molecular identification provides valid and genuine identification of *Fusarium* as it goes directly from DNA, the hereditary materials, instructions, and information of this organism.

The study went deeper into the potential and efficiency of *Fusarium* as a plant pathogen. The determination of *Fusarium* pathogenicity conveys the species virulence. A number of notions reported the pathogenicity of *Fusarium* on cucurbits as a saprophyte, opportunist and parasite. Hence, the pathogenicity test resolves their degree of severity made on the hosts. Over the years, the study on *Trichoderma* as a biocontrol agent progressively being held due to its efficacy to suppress and inhibit the pathogen growth on hosts. This study conveys the application of *T. asperellum* onto infected cucumber plants to enhance the plant growth, defense and induction of phytohormone. This enhancement subsequently affects the photosynthetic efficiency of treated cucumber plants through the analysis of photosystem II efficiency.

1.2 Research objectives

This study was conducted in order to manage a better cultivation of cucurbits; the species diversity and virulence of *Fusarium* species as the pathogen are some of the major factors to be testifying. This study is important in contributing to much further information related to *Fusarium* and understanding plant-pathogen-biocontrol agent interaction. Therefore, the main objectives of this study are:

- i. To identify *Fusarium* species associated with wilt disease of cucurbits based on morphological characteristics and molecular approaches.
- ii. To ascertain the pathogenicity of *Fusarium* species isolated from bottle gourd, cucumber, luffa, pumpkin and rock melon infected samples.
- iii. To examine the biological changes in Fusarium wilt-infected cucumber plants treated with *Trichoderma asperellum*.



REFERENCES

- Abdul-Hasan, F. and Hussein, H.Z. (2016). Genetic diversity of *Fusarium solani* f.sp. *cucurbitae*, the causal root and crown rot of cucurbits (melon) by using molecular markers and control. *American Journal of Plant Sciences*, 7, 2151-2172.
- Achu, B.M., Fokou, E., Tchiégang, C., Fotso, M., and F.M. Tchouanguep. (2005). Nutritive value of some Cucurbitaceae oil seeds from different regions in Cameroon. African Journal of Biotechnology, 4, 1329-1334.
- Agamuthu, P., Hidzir, K.M. and Hamid, F.S. (2009). Drivers of sustainable waste management in Asia. *Waste Management and Research*, 27(7), 625-633.
- Agarwal, M., Kumar, A. Gupta, R. and Upadhyaya, S. (2012). Extraction of polyphenol, flavonoid from *Emblica officinalis*, *Citrus limon*, *Cucumis sativus* and evolution of their antioxidant activity. *Oriental Journal of Chemistry*, 28(2), 993-998.
- Al-Karaawi, H.R. (2017). Insecticidal mycotoxins against cockroach Periplanta americana (Dictyoptera: Blattidae). International Journal of ChemTech Research, 10(2), 778-784.
- Amin, F., Razdan, V.K., Mohiddin, F.A., Bhar, K.A. and Sheikh, P.A. (2010). Effects of volatile metabolites of *Trichoderma* species against seven fungal plant pathogen in vitro. *Journal of Phytopathology*, 2(10), 34-37.
- Anderson, J.P., Badruzsaufari, E., Schenk, P.M., Manners, J.M., Desmond, O.J., Ehlert, C., Maclean, D.J., Ebert, P.R. and Kazan, K. (2004). Antagonistic interaction between abscisic acid and jasmonate-ethylene signaling pathways modulated defense gene expression and disease resistance in *Arabidopsis*. *Plant Cell*, 16, 3460-3479.
- Andrade-Cetto, A., Becerra-Jimenez, J., Cardenas-Vazque, R. (2008). Alfaglucosidase-inhibiting activity of some Mexican plants used in the treatment of type 2 diabetes. *Journal of Ethnopharmacology*, 116, 27–32.
- Aoki, T., O'Donnell, K., Homma, Y. and Lattanzi, A.R. (2003). Sudden-death syndrome of soybean is caused by two morphologically and phylogenetically distinct species within the *Fusarium solani* species complex – *F. virguliforme* in North America and *F. tucumaniae* in South America. *Mycologia*, 95, 660–684.
- Arguelles-Arias, A., Brans, A., Joris, B., Fickers, P. (2009). Bacillus amyloliquefaciens GA1 as a source of potent antibiotics and other secondary metabolites for biocontrol of plant pathogens. Microbiological Cell Factories 8, 1–12.
- Armengol, J., Jose, C., Moya, M.J., Sales, R., Vicent, A. and Garcia-Jimenez, J. (2000). *Fusarium solani* f.sp. *cucurbitae* race 1, a potential pathogen of grafted watermelon production in Spain. *EPPO Buletin*, 30(2), 179-183.
- Avinash, T.S. and RavishankarRai, V. (2013). Identification of diverse fungi related with selected cucurbitaceae vegetables. *Journal of Agricultural Technology*. 9(7), 1837-1848.
- Bae, H., Sicher, R.C., Kim, S.H., and Strem, M.D. (2009). The benefiacial endophyte *Trichoderma hamatum* isolate DIS219b promotes growth and delays the onset of the drought response in *Theobroma cacao*. *Journal of Experimental Botany*, 60(11), 3279-3295.
- Bae, S.J., Mohanta, T.K., Chung, J.Y., Ryu, M., Park, G., Shim, S., Hong, S.B., Seo, H., Bae, D.W., Bae, I., Kim, J.J. Bae, H. (2016). *Trichoderma*

metabolites as biological control agents against *Phytophthora* pathogens. *Biological Control.* 92, 128-138.

- Balasubramanian, G., Udayasoorian, C. and Prabu, P.C. (2007). Effects of shortterm exposure of simulated acid rain on the growth of Acacia nilotica. *Journal of Tropical Forestry and Sciences*, 19, 198-206.
- Bhimani, M.D., Golakiya, B.B. and Akbari, L.F. (2018). Evaluation os different fungicides against fenugreek wilt (*Fusarium oxysporum* Schlecht.). *International Journal of Chemical Studies*, 6(2), 29-34.
- Bilai, V. I. (1970). Experimental morphogenesis in the fungi of the genus Fusarium and their taxonomy. *Annales Academiae Scientiarum Fennica Ser. A, IV, Biologica, 168,* 7-18.
- Birthal, P.S., Joshi, P.K., Roy, D. and Pandey, G. (2019). *Transformation and sources growth in Southest Asian Agriculture*. International Food Policy and Research Institute. India: New Delhi (pp. 4)
- Bisognin, D.A. (2002). Origin and evolution of cultivated cucurbits. *Cienca Rural*, 32(5), 715-723.
- Bokhari, N.A. and Perveen, K. (2012). Antagonistic action of *Trichoderma* harzianum and *Trichoderma viride* against *Fusarium solani* causing root rot of tomato. African Journal of Mircobiology Research, 6(44), 7193-7197.
- Booth, C. (1971). *The genus Fusarium*. Commonwealth Mycological Institute. Kew, Surrey, England. (pp. 237).
- Boughalleb, N. and El-Mahjoub, M. (2007). Frequency of *Fusarium oxysporum* f. sp. *niveum* and *Fusarium solani* f. sp. *cucurbitae* from watermelon seeds and their effect on disease incidence. *Research Journal of Parasitology*, 2, 32-38.
- Bowman, T., Garcia, R., Turkson, J. and Jove, R. (2000). STATs in oncogenesis. Oncogene, 19, 2474-2488.
- Brotman, Y., Briff, E., Viterbo, A. and Chet, I. (2008). Role of swollenin, an expansin-like protein from *Trichoderma* in plant root colonization. *Plant Physiology*, 147, 779-789.
- Brunner, K., Susanne, Z., Rosalia, C., Woo, S.L., Loritto, M. ubice, C.P. and Mach, R.L. (2005). Improvement of the fungal biocontrol agent *Trichoderma atroviride* to enhance both antagonism and induction of plants systemic disease resistance. *Applied and Environmental Microbiology*, 71(7), 3959-3965.
- Burgess, L. W., Liddell, C. M. and Summerell, B. A. (1988). *Laboratory manual for Fusarium research* (2nd ed.) Sydney, Australia: University of Sydney (pp. 3)
- Burgess, L.W., Summerell, B.A., Backhouse, D., Benyon, F. and Levic, J. (2006). Biodiversity and population studies in *Fusarium*. *Sydowia*, 48(1), 1-11.
- Byrne, K.P. and Wolfe, K.H. (2005). The Yeast Gene Order Browser: combining curated homology and syntenic context reveals gene fate in polyploid species. *Genome Research*, 15(10), 1456-1461.
- Cao, Y., Zhang, Z., Ling, N., Yuan, Y., Zheng, X., Shen, B. and Shen, Q. (2011). Bacillus subtilis SQR 9 can control Fusarium wilt in cucumber by colonizing plant roots. Biology and Fertility of Soils, 47(5), 495-506.
- Carreras-Villasenor, N., Sanchez-Arreguin, J.A. and Herrera-Estrella, A.H. (2012). *Trichoderma*: sensing the environment for survival and dispersal. *Microbiology*, 158, 3-16.

- Casas-Flores, S., Rios-Memberg, M., Bibbins, M., Ponce-Noyola, P. and Herrera-Estrella, A. (2004). BLR-1 and BLR-2, key regulatory elements of photoconidiation and mycelial growth in *Trichoderma atroviride*. *Microbiology*, 150, 3561-3569.
- Chaverri, P., Gazis, R. and Samuels, G.J. (2011). *Trichoderma amazonicum*, a new endophytic species on *Hevea brasiliensi* and *H. Guianensis* from the Amazon basin. *Mycologia*, 103, 139-151.
- Chehri, K. (2011). Occurrence of *Fusarium* species associated with economically important agriculture crops in Iran. *African Journal of Microbiology Research*, 5(24), 4043-4048.
- Chehri, K., Salleh, B. and Zakaria, L. (2015). Morphological and phylogenetic analysis of *Fusarium solani* species complex in Malaysia. *Fungal Microbiology*, 69, 457-471.
- Chen, J.G. (2005). Effects of sugar-removed pumpkin zymptic powders in preventing and treating the increase of blood glucose in alloxan-induced diabetic mice. China *Journal of Clinical and Rehabilitee*, 9, 94-95.
- Chen, Z., Chen, X., Yan, H., Li, W., Li, Y., Cai, R. (2015). The Lipoxygenase Gene Family in Poplar: Identification, Classification, and Expression in Response to MeJA Treatment. *PLOS One*, 10(4), e0125526.
- Chen, Z.D., Huang. R.K., Wen, J.L. and Yuan, G.Q. (2015). Development of pathogenicity and AFLP to characterize *Fusarium oxysporumm* f. sp. *momordicae* isolates from bitter gourd in China. *Journal of Phytopathology*, 163(3), 202-211.
- Choi, I.Y., Kim, J.H., Lee, H.W., Park, J.H. and Shin, H.D. (2015). First report of Fusarium wilt of zucchini caused by *Fusarium oxysporum*, in Korea. *Journal of Microbiology*, 43(2), 174-178.
- Chouaki, T., Lavarde, V. Lachaud, L., Raccurt, C.P. and Hennequin, C. (2002). Invasive infections due to *Trichoderma* species: report of 2 cases, finding of In Vitro susceptibility testing and review of literature. *Clinical Infectious Diseases*, *35*(11), 1360-1367.
- Christenhusz, M.J.M. and Byng, J.W. (2016). The number of known plants species in the world and its annual increase. *Phytotaxa*, 261(3), 201-217.
- Cohen, R., Orgil, G., Burger, Y., Saar, U., Elkabetz, M., Tadmor, Y., Edelstein, M., Belausov, E., Maymon, M., Freeman, S. and Yarden, O. (2014). Differences in the responses of melon accessions to Fusarium root and stem rot and their colonization by *Fusarium oxysporum* f.sp. radicescucumerinum. Plant Pathology, 64(3), 655-663.
- Contrares-Cornejo, H.A., Macias-Rodriguez, L., Cortes-Penagos, C. and Bucio-Lopez, J. (2009). *Trichoderma virens*, a plant beneficial fungus, enhances biomass production and promotes lateral root growth through an auxindependent mechanism in Arabidopsis. *Plant Physiology*, *149*, 1579-1592.
- Cui, J., Wang, Y., Han, J. and Cai, B. (2016). Analyses of the community compositions of root rot pathogenic fungi in the soybean rhizosphere soil. *Chilean Journal of Agriculture Research*, 76(2), 179-187.
- Cumagun, C.J.R., Aguirre, J.A., Relevante, C.A. and Balatero, C.H. (2010). Pathogenicity and aggressiveness of *Fusarium oxysporum* Schl. in bottle gourd and bitter gourd. *Plant Protection Science*, 46(2), 51-58.
- Cummings, N.J., Ambrose, A., Braithwaite, M., Bissett, J., Roslan, H.A., Abdullah, J. and Hill, R.A. (2016). Diversity of root-endophytic *Trichoderma* from Malaysian Borneo. *Mycological Progress*, 15(50), 1-14.

- Czembor, E., Stepien, L. and Wasiewicz, A. (2015). Effect of environmental factors on *Fusarium* species and associated mycotoxins in maize grain grown in Poland. *Plos ONE*, *10*(7), e0133644.
- Daniel, H.C.F., Wilfredo, F.F., Francisco, C.R., Gabriel, G.M. and Epifanio, C.D.A. (2014). Antibiosis In Vitro of Trichoderma strains metabolic extract on mycelial growth and reproductive capacity of Fusarium oxysporum isolated from pepper plants (Capsicum annuum L.). British Biotechnology Journal, 4(4), 387-399.
- Darvishnia, M. (2013). Morphological and phylogenetic studies of *Fusarium* species in Iran. *Journal of Novel Applied Science*, 2(4), 1134-1142.
- Daryaei, A., Jones, E.E., Ghazalibglar, H., Glare, T.R. and Falloon, R.E. (2016). Effects of temperature, light and incubation period on production, germination and bioactivity of *Tricoderma atroviride*. *Journal of Applied Microbiology*, 120(4), 999-1009.
- De Medeiros, E., Gusatavo, P.D., Santos, L.A.R.D. and Lima, J.R.D.S. (2017). Soil organic carbon, microbial biomass and enzyme activities responses to natural regeneration in a tropical dry region in Northeast Brazil. *Catena*, *151*, 137-146.
- De wit, P.J.G.M. (2007). How plants recognize pathogens and defend themselves. Cell Molecular and Life Science, 64, 2726-2732.
- Dean, R., Van Kan, J.A., Pretorius, Z.A., Hammond-Kosack, K.E., Di Pietro, A., Spanu, P.D., Dickman, M., Kahmann, R., Decker-Walters, D.S., Staub, J.E., Chung, S.M., Nakata, E., Quemada, H.D. (2012). The top 10 fungal pathogens in molecular plant pathology. *Molecular Plant Pathology*, 13(4), 414-430.
- Decker-Walters, D.S., Wilkins-Ellert, M., Chung, S.M., Staub, J.E. (2004). Discovery and genetic assessment of wild bottle gourd [*Lagenaraia siceraria*(Mol.) Standley; *Cucurbitaceae*] from Zimbabwe. *Economic Botany*, 58, 501-508.
- Dennis, C. and Webster, J. (1971). Antagonistic properties of species-groups of *Trichoderma*: II. Production of volatile antibiotics. *Transaction of the British Mycological Society*, *57*(1), 41-48.
- Deyo, A. and O'Malley, B. (2008). Cucurbitaceae. Food for thoughts: The science, cultural and politics of food spring, 2008. Retrieved from https://www.scribd.com/document/314853193/Cucurbitaceae
- Dhoro Milcah. (2010). Identification and differentiation of *Fusarium* species using selected molecular methods (Master thesis, University of Zimbabwe). Retrieved

 $\frac{\text{http://ir.uz.ac.zw/bitstream/handle/10646/827/Dhoro thesis.pdf; jsessionid}{=12A5C915104DC731443340CEF009D491?sequence=1}$

- Djovonic, S., Pozo, M.J., Dangott, L.J., Howell, C.R. and kenerley, C.M. (2006). Sm1, a proteinaceous elicitors secreted by the biocontrol *Trichoderma virens* induces plant defense responses and systemic resistance. *Molecular Plant-Microbe Interaction*, 19, 838-853.
- Doehlemann, G., Okman, B., Zhu, W. and Amir, S. (2017). Plant pathogenic fungi. *Microbial Spectrum*, 5(1), 703-72.
- Doni, F., Isahak, A., Radziah, C., Zain, C.M., Arifin, S.M., Nurashiqin, W.,Mohamad Wan Mohtar, W. and Yusoff, W. (2014). Formulation of *Trichoderma* sp. SL2 inoculants using different carriers for soil treatment in rice seedling growth. *Springer Plus*, 3, 1-5.

- Drizhinina, I.S., Kopchinskiy, A.G., Kubicek, C.P. (2006). The first 100 *Trichoderma* species characterized by molecular data. *Mycoscience*, 47, 55-64.
- Du Plessis, I.L., Drizhinina, I.S., Atanasova, L., Yarden, O. and Jacobs, K. (2018). The diversity of *Trichoderma* species from soil in South Africa, with five new additions. *Mycologia*, 110(3), 559-583.
- Dubey, S.C., Kumari, P. and Singh, V. (2014). Phylogenetic relationship between different race representative populations of *Fusarium oxysporum* f. sp. *ciceris* in respect of translation elongation factor-1 α , β -tubulin and internal transcribed spacer region genes. *Archives of Microbiology*, 196(6), 445-452.
- Dwivedi, N. and Dwivedi, S.K. (2016). Histopathological observations in guava root during wilting caused by *Fusarium* species: a scanning electron microscopy study. *International Journal of Fruit Science*, *16*(3), 335-340.
- Egel, D.S. and Martyn, R.D. (2007). Fusarium wilt of watermelon and other cucurbits. The Plant Health Instructor. Retrieved from https://www.apsnet.org/edcenter/disandpath/fungalasco/pdlessons/Pages/F usariumWatermelon.aspx
- Ehiagbonare, J.E. and Onyebi, H.I. (2009). Seed storage evaluation imbibition capacity assessment and seed pre-sowing treatment studies on *Newbouldia laevis* (P. Beauv) ex bureau. *Scientific Research and Essays*, *4*, 453-456.
- Einax, E. and Voigt, K. (2003). Oligonucleotide primers for the universal amplification of beta-tubulin genes facilitate phylogenetic analysis in the regnum fungi. *Organisms Diversity and Evolution*, *3*, 185-194.
- Ellis, S.D., Boehm, M.J. and Mitchell, T.K. (2008). Fungal and fungal-like diseases of plants Fact Sheets, Agriculture and Natural Resources. *The Ohio State University*, (pp. 1-4).
- Ellis, J. and Foster, G.D. (2012). The top 10 fungal pathogens in molecular plant pathology. *Molecular Plant Pathology*, 13(4), 414-430.
- El-Sharkawy, H.H.A., Rashad, Y.M. and Ibrahim, S.A. (2018). Biocontorol of stem rust disease of wheat using arbuscular mycorhizal fungi and *Trichoderma* spp. *Physiological and Molecular Plant Pathology*, *103*, 84-91.
- Erickson, D.L., Smith, B.D., Clarke, A.C., Sandweiss, D.H. and Tuross, N. (2005). An Asian origin for a 10, 000-year-old domesticated plant in the Americas. *PNAS*, 10(51), 18315–18320.
- Escandell, J.M., Kaler, P., Recio, M.C., Sasazuki, T., Shirasawa, S., Augenlicht, L. (2008). Activated KRas protects colon cancer cells from cucurbitacininduced apoptosis. *Biochemical Pharmacology*, 76, 198-207.
- Eziashi, E.I., Omamor, I.B. and Odigie, E.E. (2007). Antagonism of Trichoderma viride and effects of extracted water soluble compounds from Trichoderma species and benlate solution on Ceratocystis paradoxa. *African Journal of Biotechnology*, 6, 388-392.
- Eziashi, E.I., Omamor, I.B., Odigie, E.E. (2010). Antagonism of Trichoderma viride and effects of extracted water soluble compounds from Trichoderma species and benlate solution on Ceratocystis paradoxa. *African Journal of Biotechnology*, 6(4), 388-392.
- FAO, Food and Agriculture Organization of United Nation. (2012). World Agriculture Towards 2030/2050: the 2012 Revision (pp. 3). ESA Working Paper 12-03, ROME.

- FAOSTAT, Food and Agriculture Organization of the United Nations (2009). *Feeding the world.* Retrieved from http://www.fao.org/3/i3107e/i3107e03.pdf
- Fernande, M.R. and Chen, Y. (2005). Pathogenicity of *Fusarium* species on different plant parts of spring wheat under controlled conditions. *Plant Disease*, 89(2), 164-169.
- Ferriol, M., Pico, B. and Nuez, F. (2003). Genetic diversity of germplasm collection of *Cucurbita pepo* using SRAP and AFLP markers. *Theoretical and Applied Genetic*, 107, 271-282.
- Fisher, N. L., L. W. Burgess, T. A. Toussoun, and P. E. Nelson. (1982). Carnation leaves as a substrate and for preserving *Fusarium* species. *Phytopathology*. 72:151-153.
- Freeman, B.C. and Beattie, G.A. (2008). An overview of plant defenses against pathogens and herbivores. *Plant Defense*, 1-12
- Gabriele, S., Alberto, P., Sergio, G., Fernanda, F. and Marco, M.C. (2000). Emerging potentials for an antioxidant therapy as a new approach to the treatment of systemic sclerosis. *Toxicology*, 155, 1-15.
- Gams, W. and Bissett, J. (2002). *Morphology and identification of Trichoderma: Trichoderma and Gliocladium- basic biology, taxonomy and genetics.* Taylor and Francis Ltd. (pp. 3-31).
- Geiser, D.M. (2003). Practical molecular taxonomy of fungi. In: Lange, L. and Tkacz, J. (Ed). Advance in fungal biotechnology for industry, medicine and agriculture. Kluwer Academic Publishers, Dordrecht, The Netherlands (pp. 474).
- Geiser, D. M., Aoki, T., Bacon, C. W., Baker, S. E., Bhattacharyya, M. K., Brandt, M. E., Brown, D. W., Burgess, L. W., Chulze, S., Coleman, J. J., Correll, J. C., Covert, S. F., Crous, P. W., Cuomo, C. A., De Hoog, G. S., Di Pietro, A., Elmer, W. H., Epstein, L., Frandsen, R. J. N., Freeman, S., Gagkaeva, T., Glenn, A. E., Gordon, T. R., Gregory, N. F., HammondKosack, K. E., Hanson, L. E., del Mar Jímenez-Gasco, M., Kang, S., Kistler, H. C., Kuldau, G. A., Leslie, J. F., Logrieco, A., Lu, G., Lysøe, E., Ma, L.-J., McCormick, S. P., Migheli, Q., Moretti, A., Munaut, F., O'Donnell, K., Pfenning, L., Ploetz, R. C., Proctor, R. H., Rehner, S. A., Robert, V. A. R. G., Rooney, A. P., bin Salleh, B., Scandiani, M. M., Scauflaire, J., Short, D. P. G., Steenkamp, E., Suga, H., Summerell, B. A., Sutton, D. A., Thrane, U., Trail, F., Van Diepeningen, A., VanEtten, H. D., Viljoen, A., Waalwijk, C., Ward, T. J., Wingfield, M. J., Xu, J.-R., Yang, X.-B., Yli-Mattila, T., and Zhang, N. (2013). One fungus, one name: Defining the genus Fusarium in a scientifically robust way that preserves longstanding use. *Phytopathology*, 103,400-408.
- Geiser, D.M., Ivey, M.L., Hakiza, G., Juba, J.H. and Miller, S.A. (2005). *Gibberella xylarioides* (anamorph: *Fusarium xylarioides*), a causative agent of coffee wilt disease in Africa, is a previously unrecognized member of the *G. fujikuroi* species complex. *Mycologia*, 97(1), 191-201.
- Geiser, D.M., Jimenez-Gasco, M.M., Kang, S., Makalowska, I., Veeraraghavan, N., ard, T.J., Zhang, N., Kuldau, G.A. and O'Donnell, K. (2004).
 FUSARIUM-ID v. 1.0; A DNA seuence database for identifying *Fusarium. European Journal of Plant Pathology*, 110, 473-479.

- Ghadikolaei, M.A., Cheung, C.S., and Yung, K.F. (2019). Study of combustion, performance and emissions of a diesel engine fueled with in blended and fumingation modes. *Fuel*, 235(1), 288-300
- Ghasemzadeh, A., Hawa, Z.E.J., Asmat, R., Puteri Edaroyati, M.W. and Mohd Ridzwan, A.H. (2010). Effect of different light intensities on total phenolics and flavonoids synthesis and antioxidant activities in young ginger varieties (*Zingiber officinale Roscoe*). International Journal of Molecular Sciences, 11(10), 3885-3897.
- Gonzalo, M.J., Oliver, M., Garcia-Mas, J., Monfort, A., Dolcet-Sanjuan, R., Katzir, N., Arus, P. and Monforte, A.J. (2005). Simple-sequence repeat markers used in merging linkage maps of melon (*Cucumis melo L.*). *Theoretical Applied Genetic*, 110, 802-811.
- Gordon, W.L. (1944). The occurrence of *Fusarium* species in Canada. I. Species of *Fusarium* isolated from farm samples of cereal seed in Manitoba. *Canadian Journal of Plant Pathology*, 22, 282-286.
- Graham, L.E., Graham, J.M. and Wilcox, L.W. (2006). *Plant biology: upper saddle river*. Michigan, USA: Pearson Education, Incorporation. (pp. 200-202).
- Groenewald, S. (2006). *Biology, pathogenicity and diversity of Fusarium* oxysporum f. sp. cubense. Master Thesis, University of Pretoria, Pretoria.
- Hafizi, R., Salleh, B and Latiffah, Z. (2013). Morphological and molecular characterization of *Fusarium solani* and *F. oxysporum* associated with crown disease of oil palm. *Brazilian Journal of Microbiology*, 44(3), 959-968.
- Hajar, I.I., Chan, K.W., Mariod, A.A. and Maznah, I. (2010). Phenolic content and antioxidant activity of cantaloupe (*Cucumis melo*) methanolic extracts. *Food Chemistry*, 119, 643-647.
- Harman, G.E., Howell, C.R., Viterbo, A., Chet, I. and Lorito, M. (2004a). *Trichoderma* species opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2, 43-56.
- Harman, G.E., Petzoldt, R., Comis, A. and Chen, J. (2004b). Interactions between *Trichoderma harianum* strain T22 and maize inbred line Mo17 and effects of this interaction on diseases caused by *Pythium ultmum* and *Colletotrichum graminicola*. *Phytopathology*, 94(2), 147-153.
- Hermosa, R., Viterbo, A., Chet, I. and Enrique, M. (2012). Plant-beneficial effect of *Trichoderma* and of its genes. *Microbiology*, 158, 17-25.
- Hernandez-Onate, M.A., Esquivel-Naranjo, E.U., Mendoza-Mendoza, A., Stewart, A. and Herrera-Estrella, A.H. (2012). An injury-response mechanism conserved across kingdoms determines entry of the fungus *Trichoderma atroviride* into development. *Proceeding of National Academy of Science* USA, 109, 14918-14923.
- Heydari, A. and Pessarakli, M. (2010). A review on biological control of fungus plant pathogens using microbial antagonists. *Journal of Biological Sciences*, 10(4), 273-290.
- Hillis, D.M. and Bull, J.J. (1993). An empirical test of boostrapping as a method for assessing confidence in phylogenetic analysis. *Systemic Biology*, 42(2), 182-192.
- Hofstede, R.J.M. and De Riuter, W.P.J. (2014). *Disease resistance in cucmber plants*. US. Patent. No 8,8955,812. Washington, D.C., USA Patent and Trademark Office.

- Hoitink, H.A.J., Maden, L.V. and Dorrance, A.E. (2006). Systemic resistance induced by *Trichoderma* spp: interactions between the hosts, the pathogen, the biocontrol agent and soil organic matter quality. *Phytopathology*, *96*, 196-189.
- Hu, J.L., Lin, X.G., Wang, J.H., Shen, W.S., Wu, S., Peng, S.P. and Mao, T.T. (2010). Arbuscular mycorrhizal fungal inoculation enhances suppression of cucumber Fusarium wilt in greenhouse soils. *Pedosphere*, 20(5), 586-593.
- Huang, S., Li, R., Zhang, Z., Li, L., Gu, X, Fan, W., Lucas, W.J. et al., (2009). The genome of cucumber, *Cucumis sativus* L. *Nature Genetics*, 41(12), 1275-1283.
- Huang, Z.Q., Lu, X.H., Sun, M.H., Guo, R.J., Diepeningen, A.D. and Li., S.D. (2019). Transcriptome analysis of virulence differentiated *Fusarium* oxysporum f.sp. cucumerinum isolates during cucumber colonization reveals pathogenicity profiles. *BMC Genominc*, 20, 1-17.
- Hussain, M.Z., Rahman, M.A., Nurul Islam, M., Latif, M.A. and Bashar, M.A. (2012). Morphological and molecular identification of *Fusarium* oxysporum Sch. isolated from guava wilt in Bangladesh. Bangladesh Journal of Botany, 41(1), 49-54.
- Ingle, A.P. (2017). Diversity and identity of *Fusarium* species occurring on fruits, vegetables and food grains. *Nusantara Bioscience*, 1(1), 44-51.
- Islam, M.T., Yashidoko, Y., Deora, A., Ito, T. and Tahara, S. (2005). Suppression of damping-off-disease in host plants by the rhizoplane bacterium *Lysobacter* sp. strain SB-K88 is linked to plant colonization and antibiosis against soilborne peronosporomycetes. *Applied Environmental Microbiology*, 71, 3786-3796.
- Islam, S., Akanda, A.M., Prova, A., Islam, M.T. and Hossain, M.M. (2015). Isolation and identification of plant growth promoting rhizobacteria from cucumber rhizosphere and their effect on plant growth promotion and disease suppression. *Frontiers in Microbiology*, 2(6), 1-12.
- Ishii, H., Fujiwara, M. and Nishimura, K. (2018). Systemic resistance inducer acibezolar-S-methyl (ASM) and its microencapsulated formulations: their long-lasting control efficacy against cucumber disease and mitigation of phototoxicity. *Pest Management Science*, *75*(3), 801-808.
- Ismail, H.I., Chan, K.W., Mariod, A.A. and Ismail, M. (2010). Phenolic content and antioxidant activity of cantaloupe (*Cucumis melo*) methanolic extracts. *Food Chemistry*, 119, 643-647.
- Jacobs, A., Wyk, P.S.V., Marasas, W.F.O., Wingfield, B.D., Wingfield, M.J. and Coutinho, T.A. (2010). *Fusarium ananatum* sp. nov. in the *Gibberella fujikuroi* species complex from pineapples with fruit rot in South Africa. *Fungal Biology*, 114(7), 515-527.
- Jamiolkowsa, A., Wagner, A. and Sawicki, K. (2011). Fungi colonizing root of zucchini (*Cucurbita pepo* L. var. giromontina) plants and pathogenicity of *Fusarium* spp. to zucchini seedlings. Acta Agrobotanica, 64(1), 73-78.
- Jeyarajan, R. and Nakkeeran, S. (2000). Exploitation of microorganisms and viruses as biocontrol agents for crop disease management. In: Upadhyay R.K., Mukerji K.G., Chamola B.P. (eds) Biocontrol Potential and its Exploitation in Sustainable Agriculture. Springer, Boston, MA. (pp. 95-116).

- Joffe, A. Z. (1974). A modern system of *Fusarium* taxonomy. *Mycopathologia et Mycologia Applicata*, 53, 201-228.
- Kalaji, H.M., Gonvindjee, Bosa, K., Koscielniak, J. and Zuk-Golaszewska, K. (2011). Effect of salt stress on photosystem II efficiency and CO₂ assimilation of two Syrian barley landraces. *Environmental and Experimental Botany*, 73, 64-72.
- Kalaji, H.M., Schansker, G., Brestic, M., Bussoti, F., Calatayud, A., Ferroni, L., Golstev, V., Guidi, L., Jajoo, A., Li, P., Losciale, P., Mishra, V.K., Misra, A.N., Nebauer, S.G. and Pancaldi, S. (2017). Frequently asked questions about chlorophyll fluorescence, the sequel. *Photosynthesis Research*, 132(1), 13-66.
- Kamarubahrin, A.F., Haris, A., Mohd Daud, S.N., Zulkefli, Z., Ahmad, N., Muhamed, N.A. and Abdul Shukor, S. (2018). The potential of pumpkin (*Cucurbita moschata* duschene) as commercial crop in Malaysia. *Pertanika Journal of Scholarly Research Reviews*, 4(3), 1-10.
- Karaca, G. and Kahveni, E. (2009). First report of Fusarium oxysporum f.sp. radices-cucumerinum on cucumbers in Turkey. New Disease Reports. 20, 9.
- Karlsson, I., Edel-Hermann, V., Gautheron, N., Durling, M.B., olseth, A.K., Steinberg, C., Persson, P. and Friberg, H. (2015). Genus-specific primers for study of *Fusarium* communities in field samples. *Applied Enviironmental Microbiology*, 82(2), 491-501.
- Kerenyi, Z., Moretti, A., Waalwijk, C., Olah, B. and Hornok, L. (2004). Mating type sequences in asexually reproducing Fusarium species. *Applied and Environmental Microbiology*, 70(8), 4419-4423.
- Khairani, A.N. (2018). Agriculture: addressing food security in Malaysia. The Edge Malaysia <u>https://www.theedgemarkets.com/article/agriculture-addressing-food-security-malaysia</u>.
- Khaledi, N., Taheri, P., Falahati, R.M. (2016). Identification, virulence factors characterization, pathogenicity and aggressiveness analysis of Fusarium spp., causing wheat head blight in Iran. *European Journal of Plant Pathology*, 147(4), 897-918.
- Knierim, D., Tsai, W.S., Maiss, E. and Kenyon, L. (2014). Molecular diversity of poleroviruses infecting cucurbit crops in four countries reveals the presence of members of six distinct species. *Archives of Virology*, 159(6), 1459-1465.
- Kopchinsiy, A., Komon, M., Kubicek, C.P. and Drizhinina, I.S. (2005). TrichoBLAST: a multilocus database for *Trichoderma* and *Hypocrea* identifications. *Mycological Research*, 109(6), 658-660.
- Kredics, L., Chen, L., edves, O., Buchner, R., Hatvani, L., Allaga, H., Nagy, V.D., haled, J.M., Alharbi, N.S. and Vagvolgyi, C. (2018). Molecular tools for monitoring *Trichoderma* in agricultural environments. *Frontiers Microbiology*, 9, 1599
- Krempels, D. (2011). Laboratory: environment and development, the effects of environment on seed germination. Retrieved from

http://www.bio.miami.edu/dana/161/evolution/161S11_germination.pdf

Kreye, C., Bouman, B.A.M., Faronilo, J.E. and Llorca, L. (2009). Causes for soil sickness affecting early plant growth in aerobic rice. *Field Crops Research*, 114(2), 182-187.

- Krober, W., Plath, I., Heklau, H. and Bruelheide, H. (2015). Relating stomatal conductance to leaf functional traits. *Journal of Visualized Experiments*, 104, 1-7.
- Kubicek, C.P., Herrera-Estrella, A., Seidl-Seiboth, V., Martinbez, D.A. and Druzhinina, I.S. (2011). Comparative genome sequence analysis underscores mycoparasitism as the ancestral life style of *Trichoderma*. *Genome Biology*, 12, R40
- Kucuk, C. and Kivanc, M. (2004). In Vitro antifungal activity of strains of Trichoderma harzianum. Turkish Journal of Biology, 28, 111-115.
- Kumar, P., Kamle, M., Misra, A.K., O'Donovan, A., Pagano, M. and Modi, D.R. (2016). Identification and characterization of *Fusarium mangiferae* as pathogen of mango malformation in India. *Brazilian Archives of Biology* and Technology, 59, 1-9.
- Kumar, A. (2013). *Trichoderma*: a biological weapon for managing plant diseases and promoting sustainability. *International Journal of Agricultural Science and Veterinary Medical*, 1(3), 106-121.
- Laraba, I., Boureghda, H., Abdallah, N., Bouaicha, O., Obanor, F., Morretti, A., Geiser, D.M., Kim, H.S., McCormick, S.P., Proctor, R.H., Kelly, A.C., Ward, T.J. and O'Donnell, K. (2017). Population genetic structure and mycotoxin potential of wheat crown rot and head blight pathogen *Fusarium culmorum* in Algeria. *Fungal Genetics and Biology*, 103, 34-41.
- Latiffah, Z., Mohd Zariman, M. and baharuddin, S. (2007). Diversity of *Fusarium* species in cultivated soils in Penang. *Malaysian Journal of Microbiology*, 3(1), 27-30.
- Lawrence, E.S. (1980). Contributions from biochemistry and plant physiology. In: Madison, W.I., Murphy, L.S., Doll, E. and Welch, F. (Ed). *Moving up the yield curve: advances and obstacles.* (pp. 25-43)
- Lee, S., Yap, M., Behringer, G., Hung, R. and Bennett J.W. (2016). Volatile organic compounds emitted by *Trichoderma* species mediate plant growth. *Fungal Biology and Biotechnology*, 3(1), 1-14.
- Leslie, J.F. and Summerell, B.A. (Ed) (2006). *The Fusarium laboratory manual*. USA: Blackwell Publishing (pp. 2-20).
- Li, B.J., Liu, Y., Shi, Y.X. and Xie, X.W. (2010). First report of crown rot of grafted cucucmber caused by *Fusarium solani* in China. *Plant Disease*, 94(11), 1377.
- LICOR. (1989). Li-1600 steady state porometer. Retrieved from https://www.licor.com/documents/auoxn0ewmmka5r5inwcf
- Lin, X. and Heitman, J. (2005). Chlamydospore formation during hyphal growth in *Cryptococcus neoformans. Eukaryotic Cell, 4,* 1746-1754.
- Lindenthal, M., Treiner, U., Dehne, H.M. Oerke, E.C. (2004). Effect of downy mildew development on transpiration of cucumber leaves visualized by digital infrared thermography. *The American Phytopathological Society*, 95(3), 233-240.
- Liu, H., Song, C. and Ram, A. (2017). Advances in applied biotechnology: Proceedings of the 3rd International Conference on Applied Biotechnology (ICAB2016), November 25-27, 2016, Tianjin, China. Springer Nature Singapore Pte Ltd. (Pp. 589).
- Loukou, A.L. (2007). Macronutrient composition of tree cucurbit species cultivated for seed consumption in Côte d'Ivore. *African Journal of Biotechnology*, *6*, 529-533.

- Mace, M.E.; Bell, A.A.; Beckman, C.H. (1981). *Fungal Wilt Diseases of Plants* Academic Press: New York, USA. (pp. 640.)
- MacHardy, W.E. & Beckman, C.H. (1981) Vascular wilt Fusaria: Infections and pathogenesis. In Nelson, P.E., Toussoun, T.A. and Cook, R.J. (Ed). *Fusarium: Disease, biology and taxonomy.* The Pennsylvania State University Press, University Park (pp. 365-366)
- Mahmood, T., Mehnaz, S., Fleischmann, F., Ali, R., Hashmi, Z.H. and Iqbal, Z. (2014). Soil sterilization effects on root growth and formation of rizosheaths in wheat seedlings. *Pedobiologia*, 57, 123-130.
- MOA. Malaysia Department of Agriculture (2018). Vegetables and cash crops statistic. Retrieved from http://www.doa.gov.my/index/resources/aktiviti_sumber/sumber_awam/m aklumat_pertanian/perangkaan_tanaman/perangkaan_sayur_tnmn_ladang_ 2017.pdf
- Manavalan, L.P. and Nguyen, H.T. (2017). Drought tolerance in crops: physiology to genomics. Plant Stress Physiology. (2nd Ed). CAB International. Boston, USA. (Pp. 3)
- Manshor, N., Rosli, H., Ismail, N.A., Salleh, B. and Zakaria, L. (2012). Diversity of *Fusarium* species from highland areas in Malaysia. *Tropical Life Sciences Research*, 23(2), 1-5.
- Maryani, N., Lombard, L., Poerba, Y.S., Subandiyah, S., Crous, P.W. and Kema, G.H.J. (2019). Phylogeny and genetic diversity of the banana Fusarium wilt pathogen *Fusarium oxysporum* f.sp. *cubense* in the Indonesian centre of origin. *Studies in Mycology*, 92, 155-194.
- Masratul Hawa, M., Salleh, B. and Latiffah, Z. (2010). Characterization and intraspecific variation of *Fusarium semitectum* (Berkeley and Ravenal) associated with red-fleshed dragon fruit (*Hylocereus polyrhizus* [Weber] Britton and Rose) in Malaysia. *African journal of Biotechnology*, 9(3), 273-284.
- Matsumoto, Y., Ogawara, T., Miyagi, M., Watanabe, N. and Kuboyama, T. (2011). Response of wild *Cucumis* species to inoculation with *Fusarium* oxysporum f.sp. melonis Race 1, 2y. Journal Japan Horticulture Science, 80(4), 414-419.
- Matuo, T. (1972). Taxonomic studies of phytopathogenic fusaria in Japan. *Review* of *Plant Protection Research*, *5*, 34-45.
- Maxwell, K. and Johnson, G.N. (2000). Chlorophyll fluorescence- a practical guide. *Journal of Experimental Botany*, 51(345), 659-668.
- McCallum, H., Barlow, N. and Hone, J. (2001). How should pathogen transmission be modeled? *Trends ecological Evolutionary*, *16*(6), 295-300
- McCreight, J.D., Staub, J.E., Wehner, T.C. and Dhillon, N.P.S. (2013). Gene Global: Familiar and exotic cucurbits have Asian origins. *HortScience*, 48(9), 1078-1089.
- Meena, M., Swapnil, P., Zehra, A., Dubey, M.K. and Upadhyay, R.S. (2017). Antagonistic assessment of *Trichoderma* spp. by producing volatile and non-volatile compounds against different fungal pathogens. *Archives of Phytopathology and Plant Protection*, 50(13-14), 629-648.
- Mehl, H.L. and Epstein, L. (2007). Identification of *Fusarium solani* f.sp. *cucurbitae* race 1 and race 2 with PCR and production of disease-free pumpkin seeds. *Plant Disease*, 91(10), 1288-1292.

- Messiaen, C. M., and R. Cassini. (1968). Fusarium Research. IV. The systematics of Fusarium. *Annnals of Epiphytas*, 19, 387-454.
- Michael, Nee. (1990). The domestication of cucurbita (Cucurbitaceae). *Economic Botany*, 44(3), 56-68.
- Michielse, C.B. and Rep, M. (2009). Pathogen profile update: Fusarium oxysporum. Molecular Plant Pathology, 10, 311–324.
- Minuto, A., Spadaro, D., Garibaldi, A. and Gullino, M.L. (2006). Control of soilbrone pathogens of tomato using a commercial formulation of *Streptomyces griseoviridis* and solarization. *Crop protection*, 25, 467-475.
- Mishra, R.K., Pandey, B.K., Pandey, A., Mohd Zeeshan, P.N. (2012). Specific detection of *Fusarium oxysporum* f. sp. *psidii* (Fop) causing wilt disease of *Psidium guajava* L. in India. In: National Symposium on Microbes in Health and Agriculture, JNU, New Delhi (pp.87).
- Mladenovic, E., Berenji, J., Ognjanov, V., Ljubojevic, M. and Cuanovic, J. (2012). Genetic variability of bottle gourd *Lagenaria siceraria* (Mol.) Standley and its morphological characterization by multivariate analysis. *Archive of Biological Science*, 64(2), 573-583.
- Mohd Zainudin, N.A.I., Farah Aqila, H., Nur Azizah, K. and Nur Syuhada, Z. (2017). Characterization and pathogenicity of *Fusarium proliferatum* and *Fusarium verticillioides*, causal agents of Fusarium ear rot of corn. *Turkish Journal of Biology*, 41, 220-230.
- Molina, A.B., Fabregar, E., Sinohin, V.G., Yi, G. and Viljoen, A. (2009). Recent occurrence of *Fusarium oxysporum* f.sp. *cubense* tropical race 4 in Asia. *Acta Horticulture*, 828, 109-116.
- Mousavizadeh, S.J., Mashayekhi, K., Garmakhany, A.D., Enteshamnia, A. and Jafari, S.M. (2010). Evaluation of some physical properties of cucumber (*Cucumis sativus* L.). Journal of Agricultural Science and Technology, 29(4), 107-114.
- Mucharroma, H. and Kuc, J. (1991). Oxalates and phosphate induce systemic resistance against diseases caused by fungi, bacteria and viruses in cucmumber. *Crop Protection*, 10(4), 265-270.
- Mukherjee, M., Mukherjee, P.K., Horwitz, B.A., Zachow, C., Berg, G. and Zeilinger, S. (2012). *Trichoderma*-plant-pathogen interactions: advances in genetic of biological control. *Indian Journal of Microbiology*, *52*, 522-529.
- Mukherjee, P.K., Neema, K.N., Maity, N., and Sarkar, B.K. (2013). Phytochemical and therapeutic potential of cucumber. *Fitoterapia*, 84, 227-236.
- Murchie, E.H. and Lawson, T. (2013). Chlorophyll fluorescence analysis: a guide to good practice and understanding some new applications. *Journal of Experimental Botany*, 64(13), 3983-3998.
- Mwaniki, P.K., Abang, M.M., Wagara, I.N., Wolukau, J.N. and Schroers, H.J. (2011). Morphology, pathogenicity and molecular identification of *Fusarium* spp. from wilting eggplants in Tanzania. *African Crop Science Conference Proceedings*, 10, 217-221.
- Naher, L., Hazreen, N., Siti Aminah, M.Z. and Siddiquee, S. (2017). Isolation of antagonistic *Trichoderma* spp. against selected phytopathogenic fungi from the field soils in Kelantan. *Malaysian Journal of Microbiology*, 13(2), 73-78.
- Najihah, A., Nur Ain Izzati, M.Z., Yong, S.Y.C. and Nik Mohd Iham, M.N. (2017). Characterization of *Fusarium proliferatum* and *Fusarium verticillioides*

based on species-specific gene and microsatellites analysis. *Sains Malaysiana*, 46(12), 2425-2432.

- Nalim, F.A., Samuels, G.J., Wijesundera, R.L. and Geiser, D.M. (2011). New species from the *Fusarium solani* species complex derived from perithecia and soil in the Old World tropics. *Mycologia*, 103(6), 1302-1330.
- Nawrocka, J. and Malolepsza, U. (2013). Diversity in plant systemic resistance induced by *Trichoderma*. *Biological Control*, 67(2), 149-156.
- Nawrocka, J., Malolepsza, U., Szymczak, K. and Szczech, M. (2018). Involvement of metabolic components, volatile compounds, PR proteins and mechanical strengthening in multilayer protection of cucumber plants against *Rhizoctonia solani* activated by *Trichoderma atroviride* TRS25. *Protoplasma*, 225(1), 359-373.
- Nelson, P. E., Toussoun, T. A. and Marasas, W. F. 0. (1983). Fusarium species: an illustrated manual for identification. Pennsylvania State University Press, University Park. (pp. 5).
- Nelson, P.E., Dignani, M.C. and Anaissie, E.J. (1994). Taxonomy, biology and clinical aspects of *Fusarium* species. *Clinical Microbiology Reviews*, 7(4), 479-504.
- Ng, L.C., Ngadin, A., Azhari, A. and Zahari, N.A. (2015). Potential of *Trichoderma* spp. as biological control agents against Bakanae pathogen (*Fusarium fujikuroi*) in rice. *Asian Journal of Plant Pathology*, 9(2), 46-58.
- Nimchuk, Z., Eulgem, T. and Holt, B.F. (2003). Recognition and response in the plant immune system. *Annual Review of Genetics*, 37(1), 579-609.
- Nirmaladevi, D. and Srivinas, C. (2012). Cultural, morphological and pathogenicity variation in *Fusarium oxysporum* f.sp. *lycopersici* causing wilt of tomato. *Journal of Life Science*, 2(1), 1-16.
- Nor Azliza, I., Masratul Hawa, M., Nik Mohd Izham, M.N. and Latiffah, Z. (2017). Funmonisin B1-producing *Fusarium* species from agricultural crops in Malaysia. *Crop Protection*, 98, 70-75.
- Norrizah, J.S., Hashim, S.N., Siti Fasiha, F. and Yaseer, S.M. (2012). β-carotene and antioxidant analysis of three different rock melon (*Cucumis melo* L.) cultivars. *Journal of Applied Science*, 12(17), 1846-1852.
- Nur Ain Izzati, M.Z., Siti Nordahliawate, M.S., Nor Azliza, I. and Salleh, B. (2009). Distribution and diversity of *Fusarium* species associated with grasses in ten states throughout Peninsular Malaysia. *Biotropia*, 16(2), 55-64.
- O'Donnell, K., Gueidan, C., Sink, S., Johnston, P.R., Crous, P.W., Glenn, A., Riley, R., Zitomer, N.C., Colyer, P., Waalwijk, C., Lee, T.V., Moretti, A., Kang, S., Kim, H.S., Geiser, D.M., Juba, J.H., Baayen, R.P., Cromey, M.G., Bithell, S., Sutton, D.A., Skovgaard, K., Ploetsz, R., Corby, K.H., Elliott, M., Daviz, M. and Sarver, B.A. (2009). A two-locus DNA sequence database for typing plant and human pathogens within the *Fusarium oxysporum* species complex. *Fungal Genetic Biology*, 46(12), 936-948.
- O'Donnell, K. and Cigelnik, E. (1997). Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus *Fusarium* are nonorthologous. *Molecular Phylogenetics and Evolution*, 7(1), 103-116.

- O'Donnell, K., Cigelnik, E. and Nirenberg, H.I. (1998). Molecular systematics and phylogeography of the *Gibberella fujikuroi* species complex. *Mycologia*, 90, 465-493.
- O'Donnell, K., Kistler, H.C., Cigelnik, E. and Ploetz, R.C. (1998). Multiple evolutionary origins of the fungus causing Panama disease of banana: Concordant evidence from nuclear and mitochondrial gene genealogies. *Proceedings of the National Academy of Sciences of the United States of America*, 95, 2044–2049.
- Obst, A., Guntber, B., Baek, R., Lepschy, J. and Tisehner, H. (2002). Weather conditions conducive to *Gibberella zeae* and *Fusarium graminearum* head blight in wheat. *Journal of Applied Genetics*, 43, 185-192.
- Ohara, T. and Tsuge, T. (2004). FoSTUA, encoding a basic helix-loop-helix protein, differentially regulates development of three kinds of asexual spores, macroconidia, microconidia and chlamydospores, in the fungal plant pathogen *Fusarium oxysporum*. *Eukaryotic Cell*, *3*(6), 1412-1422.
- Ordonez, N., Seidl, M.F., Waaljik, C., Drenth, A., Kilian, A., Thomma, B.P.H.J, Ploetz, R.C. and Kema, G.H.J. (2015). Worse come to worst: Bananas and Panama disease-when plant and pathogen clones meet. *PLOS Pathology*, *11*(11), e1005197.
- Oumouloud, A., El-Otmani, M., Chikh-Rouhou, H., Claver, A.G., Torres, R.G., Perl-Treves, R. and Alvarez, J.M. (2013). Breeding melon for resistance to *Fusarium* wilt: recent development. *Euphytica*, 192(2), 155-169.
- Paksoy, M. and Aydin, C. (2004). Some physical properties of edible squash (*Cucurbita pepo* L.) seeds. *Journal of Food Engineering*, 65(2), 225-231.
- Papagianni, M. (2014). Characterization of fungal morphology using digital image analysis techniques. Journal of Microbial and Biochemical Technology, 6(4), 189-194.
- Papavizas, G.C. (1985). *Trichoderma* and *Gliocladium*: biology, ecology and potential for biocontrol. *Annual Review of Phytopathology*, 23, 23-54.
- Paris, H.S. (1989). Historical records, origins and development of the edible cultivar groups of *Cucurbita pepo* (Cucurbitaceae). *Economic Botany*, 43, 423-443.
- Patil, J., Goel, S.R. and Yadav, S. (2018). Effect of Meloidogyne incognita and Fusarium oysporum f. sp. cucumerinum on cucumber grown under protected cultivation. Journal of Entomology and Zoology Studies, 6(1), 1004-1007.
- Pavlovic, S., Ristic, D., Vucurovic, I., Stevanovic, M., Stojanovic, S., Kuzmanovic, S., Starovic, M. (2016). Morphology, pathogenicity and molecular identification of *Fusarium* spp. associated with anise seeds in Serbia. *Notulae Botanicae Horti Agrobiotanici*, 44(2), 411-417.
- Perez-Hernandez, A., Porcel-Rodriguez, E. and Gomez-Vazquez, J. (2017). Survival of *Fusarium solani* f.sp. *cucurbitae* and fungicide application, soil solarization and biosolarization for control of crown and foot rot of zucchini squash. *Plant Disease*, 101, 1507-1514.
- Pessarakli, M. (2016). *Handbook of cucurbits: growth, cultural practice and physiology* (Pp. 5). CRC Press, Taylor & Francis Group: USA
- Pimple, B.P., Kadam, P.V. and Patil, M.J. (2011) antidiabetic and antihyperlipidemic activity of *Luffa acutangula* fruit extracts in streptozotocin induced Niddm rats. *Asian Journal of Pharmaceutical and Clinical Research*, 4(2), 156-163.

- Poornima, S. (2011). Evaluation of disease control and plant growth promotion potential of biocontrol agents on *Pisum sativum* and comparison of their activity with popular chemical control agent-carbendazim. *Journal of Toxicological Environmental Health and Sciences*, *3*, 127-138.
- Pouzoulet, J., Pivovaroff, A.L., Santiago, L.S. and Rolshausen, P.E. (2014). Can vessel dimension explain tolerance toward fungal vascular wilt diseases in woody plants? Lessons from Dutch elm disease and esca disease in grapevine. *Frontiers in Plant Science*, 5, 1-11.
- Pradeep, F.S. and Pradeep, B.V. (2013). Optimization of pigment and biomass production from *Fusarium moniliforme* under submerged fermentation condotions. *Internation Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 526-532.
- Pu, X., Xie, B., Li, P., Mao, Z., Ling, J., Shen, H., Zhang, J., Huang, N. and Lin, B. (2014). Analysis of defense-related mechanism in cucumber seedlings in relation to root colonization by non-pathogenic *Fusarium oxysporum* CS-20. *Microbiology*, 355, 142-151.
- Punja, Z.K., Tirajoh, A., Collyer, D and Ni, L. (2019). Efficacy of *Bacillus subtilis* strain QTS713 (Rhapsody) against four maor diseases if greenhouse cucumbers. *Crop Protection*, 124, 104845.
- Qualtoha, T.F., Lopes, F.A.C., Steindorff, A.S. Brandao, R.S., Jesuino. R.S.A. and Ulhoa, C.J. (2013). Mycoparasitism studies of *Trichoderma* species against three phytopathogenic fungi: evaluation of antagonism and hydrolytic enzyme production. *Biotechnology Letters*, 35(9), 1461-1468.
- Ramezami, M., Karimi Abdolmaleki, M., Shabani, S. and Dehestani, A. (2017). The role of potassium phosphite in chlorophyll fluorescence and photosynthetic parameter of downy mildew-challenged cucumber (*Cucumis sativus*) plants. Archives of Phytopathology and Plant Protection, 50(17-18), 927-940.
- Raillo, A. (1935). Diagnostic estimation of morphological and cultural characters of species in the genus *Fusarium*. *Phytopathology*, 7, 1-100.
- Raihanah, R., Mohd, Y.R., Siti Izera, I., Martini, M.Y., Gous, M. and Magaji, U. (2018). Breeding for anthracnose disease in resistance in Chilli: progress and prospects. *International Journal of Molecular Science*, 19(10), 3122-3143.
- Rashid, U., Rehman, H.A., Hussain, I., Ibrahim, M. and Haider, M.A. (2011). Muskmelon (*Cucumis melo*) seed oil: a potential non-food oil source for biodiesel production. *Energy*, 3(9), 5632-5639.
- Raupach, G.S., Liu, L., Murphy, J.F., Tuzun, S. and Kloepper, J.W. (1996). Induced resistance in cucumber mosaic virus using plant growth promoting rhizobacteria. *Plant Disease*, 80, 891-894.
- Raza, W., Ling, N., Zhang, R., Huang, Q., Xu, Y. and Shen, Q. (2017). Success evaluation of the biological control of Fusarium wilts of cucumber, banana and tomato since 2000 and future research strategies. *Critical Reviews in Biotechnology*, 37(2), 202-212.
- Redda, E.T., Ma, J., Mei, J, Li, M., Wu, B. and Jiang, X. (2018). Antagonistic potential of different isolates of *Trichoderma* against *Fusarium* oxysporum, Rhizoctonia solani and Botrytis cinerea. European Journal of Experimental Biology, 8(2), 1-8.
- Reddy, M.S., Ilao, R.I., Faylon, P.S., Dar, W.D. and Batchelor, W.D. (2014). Recent advances in biofertilizers and biofungicides (PGPR) for

sustainable agriculture. Cambridge Scholars Publishing: Newcastle, UK. (pp. 397).

- Rex, J.H., Pfaller, M.A., Walsh, T.J., Chaturavedi, V., Espinel-Ingroff, A., Ghannoum, M.A., Gosey, L.L., Odds, F.C., Rinaldi, M.G., Sheehan, D.J. and Warnock, D.W. (2001). Antifungal susceptibility testing: practical aspects and current challenges. *Clinical Microbial Reviews*, 14(4), 643-658.
- Roberts, D.P., Lakshman, D.K., McKenna, L.F., Emche, S.E. and Bauchan, G. (2016). Seed treatment with ethanol extract of *Serratia marcescens* is compatible with Trichoderma isolates for control of damping-off of cucumber caused by *Phythium ultimum*. *Plant Disease*. 100(7): 1278-1287.
- Robertson, A.E. (2010). First report of cucurbit chlorotic yellows virus infecting cucurbits in Taiwan. *Plant Disease*, 94(9), 1168
- Robertson, J.I., Russell, R.M., Preisler, H.K. and Savin, N.E. (2007). *Bioassays* with Arthropods 2nd Edition. Boca Raton: CRC Press. (pp. 199).
- Rogayah, S., Muhammad Hanim, H., Roslinda, A.R., Chein-Yeong, W. and Janna, O.A. (2018. Malaysian *Carica papaya L. var Eksotika: current research* strategies fronting challenges. *Frontier Plant Science*, 9, 1308.
- Roslianah, F.S. and Sariah, M. (2006). Characterization of Malaysian isolates of *Fusarium* from tomato and pathogenicity testing. *Research Journal of Microbiology*, 1(3), 266-272.
- Rouphael, Y., Rivera, C.M., Cardarelli, M., Fanasca, S. and Colla, G. (2006). Leaf area estimation from linear measurements in zucchini plants of different ages. *Journal of Horticultural Science and Biotechnology*, 81(2), 238-241.
- Rusli, M.H., Idris, A.S. and Cooper, R.M. (2016). Evaluation of Malaysian soils for potential suppressiveness of Fusarium wilt of oil palm caused by *Fusarium oxysporum* f.sp. *elaeidis. Journal of Micobial and Biochemical Technology*, 8(6), 459-464.
- Ruxandria, M.P. and Dacinia, C.P. (2010). Organic agriculture as component of sustainable development. Romania's case. *International Journal of Bioflux Society*. 2(2): 121-132. Schmoll, M., Esquivel-Naranjo, E.U. and Herrera-Estrella, A. (2010). *Trichoderma* in the light of day: physiology and development. *Fungal Genetic Biology*, 47(11), 909-911.
- Saeed, I., Chen, X., Bachir, D.G., Chen, L. and Hu, Y.G. (2017). Association of mapping for photosynthesis and yield traits under two moisture conditions and their drought indices in winter bread wheat (*Triticum aestivum* L.) using SSR markers. Australian Journal of Crop Science, 11(3), 248-267.
- Salami, A. O., Bankole, F.A. and Adepoju, P.O. (2018). Biocontrol potentials of *Trichoderma harzianum* and Glomus facultative on *Fusarium oxysporum* causing Fusarium wilt disease of tomato (*Lycopersicum esculentum*). *Journal of Mycology*, 1(3), 1-11
- Sallenave-Namont, C., Pouchus, Y.F., Robiou du, P.T. Lassus, P. and Verbist, J.F. (2000). Toxigenic saprophytic fungi in marine shellfish farming areas. *Mycopathologia*, 149(1), 21-25.
- Samuels, G.J., Ismaeil, A., Bon, M.C., De Respinis, S. and Petrini, O. (2010). *Trichoderma asperellum sensu lato* consists of two cryptic species. *Mycologia*, 51, 71-88.

- Samuels, G.J., Ismaeil, A., Mulaw, T.B., Szakacs, G., Druzhinina, I.S., Kubicek, C.P. and Jaklitsch, W.M. (2012). The *Longibrachiatum* clade of *Trichoderma*: a revision with new species. *Fungal Diversity*, 55, 77-108.
- Saravanakumar, K, Yu, C., Dou, K., Wang, M., Li, Y. and Chen, J. (2016). Synergistic effect of *Trichoderma*-derived antifungal metabolites and cell wall degrading enzymes on enhanced biocontrol of *Fusarium oxysporum* f. sp cucumerinum. Biological Control, 94, 37-46.
- Savory, E.A., Granke, L.L., Quesada-Ocampo, L.M., Varbanova, M, Hausbeck, M.K. and Day, B. (2011). The cucurbit downy mildew pathogen *Pseudoperonospora cubensis. Molecular Plant Pathology*, 12(3), 217-226.
- Schmoll, M., Seibel, C., Tisch, D., Dorrer, M. and Kubicek, C.P. (2010). A novel class of peptide pheromone precursors in ascomycetous fungi. *Molecular Microbiology*, 77(6), 1483-1501.
- Schoonhoven, A.V. and Pastor-Corales, M.A. (1994). *Standard system for the evaluation of bean germplasm*. International Center of Tropical Agriculture (CIAT), California. (pp. 39).
- Schuster, A. and Schmoll, M. (2010). Biology and biotechnology of *Trichoderma*. *Applied Microbiology and Biotechnology*, 87(3), 787-799.
- Sebastian, P., Schaefer, H., Telford, I.R.H. and Renner, S.S. (2010). Cucumber (*Cucumis sativus*) and melon (*C. melo*) have numerous wild relatives in Asia and Australia, and the sister species of melon is from Australia. *Proceedings of the National Academy of Sciences*, 107, 14269–14273.
- Segarra, G., Casanova, E., Aviles, M. and Trillas, I. (2010). *Trichoderma* asperellum T34 controls Fusarium wilt disease in tomato plants in soilless culture through competition for iron. *Microbe Ecology*, 59, 141-149.
- Seo, Y. and Kim, Y.H. (2017). Potential reasons for prevalence of Fusarium wilt in oriental melon in Korea. *Plant Pathology Journal*, 33(3), 249-23.
- Shah, J.J., Thanki, Y.J. and Kothari, I.L. (1980). Skeletal fibrous net in fruits of Luffa cylindrica M. Roem, and Luffa acutangula Roxb. In: M. Nagaraj and C.P. Malik (eds.). Current trends in botanical research. Kalyani Publishers, New Delhi. (pp. 61-72)
- Shah, S., Nasreen, S. and Sheikh, P.A. (2012). Cultural and morphological characteristics of *Trichoderma* spp. associated with green mold disease of *Pleurotus* spp. in Kashmir. *Research Journal of Microbiology*, 7, 139-144.
- Shahidi, F., and Naczk, M. (2004). *Phenolics in food and nutraceuticals*. Boca Raton, FL: CRC Press.
- Shahraki, M., Heydari, A. and Hassanzadeh, N. (2009). Investigation of antibiotic, siderophore and volatile metabolites production by *Bacillus* and *Pseudomonas* bacteria. *Iran Journal of Biology*, 22, 71-85.
- Sharifah Siti Maryam. (2018). Characterization of Trichoderma species isolated from soil and efficacy of Trichoderma asperellum as biocontrol agent of Fusarium wilt disease of banana, Master Thesis, Universiti Putra Malaysia.
- Sharma, M., Anindita, S., Raju, G., Gaurav, A., Avijit, T., NAgavardhini, A, Suresh, P. and Rajeev, K.V. (2016). Genome wide transcriptome profiling of *Fusarium oxysporum* f. sp. *ciceris* conidial germination reveals new insights into infection-related genes. *Scientific Reports*, 6, 37353.
- Sharma, P., Vignesh, K.P., Ramesh, R., Saravanan, K., Deep, S., Sharma, M., Mahesh, S. and Dinesh, S. (2011). Biocontrol genes from *Trichoderma* species-a review. *African Journal of Biotechnology*, 10(86), 19898-19907.

- Sharma, P.K., Gothalwa, R. and Tiwari, R.K.S. (2013). Isolation of cold tolerant antifungal strains of *Trichoderma* sp. from northern hilly zones of Chhattisgarh. *International Journal of Plant Protection*, 6(2), 236-240.
- Shukla, N., Awasthi, R.P., Rawat, L. and Kumar, J. (2012). Biochemical and physiological responses of rice (*Oryza sativa* L.) as influenced by *Trichoderma harzianum* under drought stress. *Plant Physiology and Biochemistry*, 54, 78-88.
- Singh, S., Kumar, R., Yadav, S., Kumar, R., Kumari, P. and Singh, R.K. (2018). Effect of bio-control agents on soil borne pathogens: A review. *Journal of Pharmacognosy and Phytochemistry*, 7(3), 406-411.
- Sidmore, A.M. and Dickinson, C.H. (1976). Colony interaction and hyphal interference between *Septoria nodorum* and Phylloplane fungi. *Transaction of the British Mycological Society*, 66(1), 57-64.
- Snyder, W. C., and H. N. Hansen. (1940). The species concept in *Fusarium*. *American Journal of Botany*, 27, 64-67.
- Soesanto, L., Utami, D.S. and Rahayuniati, R.F. (2011). Morphological characteristics of four *Trichoderma* isolates and two endophytic *Fusarium* isolates. Canadian *Journal on Scientific and Industrial Research*, 2(8), 294-306.
- Sohrab, S.S., Mandal, B., Ali, A., and Varma, A. (2010). Chlorotic curly stunt: a severe *Begomovirus* disease of bottle gourd in Northern India. *Indian Journal Virology*, 21(1), 56-63.
- Steinkellner, S., Mammerler, R. and Vierheilig, H. (2005). Microconidia germination of the tomato pathogen *Fusarium oxysporum* in the presence of root exudates. *Journal of Plant Interaction*, 1(1), 23-30.
- Stepansky, A., Kovalski, I. and Perl-Treves, R. (1999). Intraspecific classification of melons (*Cucumis melo L.*) in view of their phenotypic and molecular variation. *Plant Systematic Evolution*, 217, 313-333.
- Stepien, L., Koczy, G. and Waskiewicz, A. (2013). Diversity of *Fusarium* species and mycotoxins contaminating pineapple. *Journal of Applied Genetics*, 54(3), 367-380.
- Stevenson, D., Fred, J.E., Wang, L. and Jane, J.L. (2007). Oil and tocopherol content and composition of pumpkin seed oil in 12 cultivars. *Journal of Agriculture and Food Chemistry*, 55(10), 4005-4013.
- Suhaida, S. and Nur Ain Izzati, M.Z. (2013). The efficacy of *Trichoderma harzianum* T73s as biocontrol agent of Fusarium ear rot diasease of maize. *International Journal of Agriculture and Biology*, *15*(6), 1175-1180.
- Sukanya, R. and Jayalakshmi, S.K. (2017). Response inoculation technique to seed and seedling infection by *M. phaseolina* in sorghum. *Advances in Plants and Agriculture Research*, 6(1), 1-2.
- Sultana, N. and Ghaffar, A. (2010). Effect of fungicides, microbial antagonists and oilcakes in the control of *Fusarium solani*, the cause of seed rot, seedling and root infection of bottle gourd, bitter gourd and cucumber. *Pakistan Journal of Botany*, 42(2), 2921-2934.
- Summerell, B.A. Salleh, B. and Leslie, J.F. (2003). A utilitarian approach to *Fusarium* identification. *Plant Disease*, 87(2), 117-128.
- Sun, Y., Frankenberg, C., Jung, m. Joiner, J., Guanter, L., Kohler, P. and Magney, T. (2018). Overview of solar-induced chlorophyll fluorescence (SIF) from the orbiting carbon observatory-2: retrieval, cross-mission comparison and

global monitoring for GPP. Remote Sensing of Environment, 209, 808-823.

- Sun, F., Sun, S., Zhu, L., Duan, Z. and Zhu, Z. (2019). Confirmation of *Fusarium* oxysporum as a causal agent of mung bean wilt in China. Crop Protection, 117, 77-85.
- Sun, Y., Li, Y., Wang, M., Wang, C., Ling, N., Mur, L.A.J. Shen, Q. and Guo, S. (2018). Redox imbalance contributed differently to membrane damage of cucumber leaves under water stress and *Fusarium* infection. *Plant Science*, 274, 171-180.
- Taghdi, Y., Hermosa, R., Dominguez, S., Rubio, M.B., Essalmani, H., Nicolas, C. and Monte, E. (2015). Effectiveness of composts and *Trichoderma* strains for control of Fusarium wilt of tomato. *Phytopathologia Mediterranea*, 54(2), 232-240.
- Tamietti, G. and Valentino, D. (2006). Soil solarization as an ecological method for the control of Fusarium wilt of melon in Italy. *Crop Protection*, 25, 389– 397.
- Taylor, J.W., acobson, D.J., roken, S., Kasuga, T., Geiser, D.M., Hibbett, S. and Fisher, M.C. (2000). Phylogenetic species recognition and species concepts in fungi. *Fungal Genetics and Biology*, 31, 21-32.
- Trivedi, M.K., Branton, A., Trivedi, D., Nayak, G., Gangwar, M. and Jana, S. (2015). Evaluation of vegetative growth parameters in biofield treated bottle gourd (*Lagenaria siceraria*) and okra (*Abelmoschus esculentus*). *International Journal of Nutrition and Food Sciences*, 4(6), 688-694.
- United State Department of Agriculture. (2013). *Cucurbita pepo L. field pumpkin*. United State: Author.
- Vargas, W.A., Mandawe, J.C. and Kenerly, C.M. (2009). Plant-derived sucrose is the key element in the symbiotic association between *Trichoderma virens* and maize plant. *Plant Physiology*, 151, 792-808.
- Verma, A. and Jaiswal, S. (2015). Bottle gourd (*Lagenaria siceraria*) juice poisoning. *World Journal of Emergency Medicine*, 6(4), 308-309.
- Verma, N.P., Kaur, I., Masih, H., Singh, A.K. and Singla, A. (2017). Efficacy of *Trichoderma* in controlling Fusarium wilt in tomato (Solanum lycopersicum L.). Research in Environment and Life Science, 10(7), 635-639.
- Viterbo, A. and Chet, I. (2006). TasHyd1, a new hydrophobin gene from the biocontrol agent *Trichoderma asperellum*, is involved in plant root colonization. *Molecular Plant Pathology*, 7, 249-258.
- Viterbo, A., Inbar, J. and Hadar, Y. (2007). Plant disease biocontrol and induced resistance via fungal mycoparasites: The mycota IV, environmental and microbial relationships. 2nd Edition. Springer-Verlag. Berlin.
- Vitti, A., Elisa, P., Nali, C., Lovelli, S., Sofo, A., Valerio, M., Scopa, A. and Nuzacci, M. (2016). *Trichoderma harzianum* T22 induces systemic resistance in tomato infected by cucumber mosaic virus. *Front Plant Science*, 7, 1520
- Walker, L., LeVine, H. and Jucker, M. (2006). Koch's postulates and infectious proteins. Acta Neuropathology, 112, 1-4.
- Walsh, T.J., Groll, A., Hiemenz, J., Fleming, R., Roilides, E. and Anaissie, E. (2004). Infections due to emerging and uncommon medically important fungal pathogens. *Clinical Microbiology Infection*, 10(1), 48-66.

- Walsh, J., Laurence, M., Liew, E., Sangalang, A., Summerell, B. and Petrovic, T. (2010). *Fusarium*: two endophytic novel species from tropical grasses of northern Australia. *Fungal Diversity*, 44(1), 149-159.
- Wang, B.B., Yuan, J., Zhang, J., Shen, Z.Z., Zhang, M.X., Li, R., Ruan, Y.Z., Shen, Q.R., (2013). Effects of novel bioorganic fertilizer produced by *Bacillus amyloliquefaciens* W19 on antagonism of Fusarium wilt of banana. *Biology and Fertility of Soils*, 49, 435–446.
- Wang, M., Sun, Y., Sun, G., Liu, X., Zhai, L., Shen, Q. and Guo, S. (2015). Water balance altered in cucumber plants infected with *Fusarium oxysporum* f. sp. cucumerinum. Scientific Reports, 5(1), 1-7.
- Wang, H. and Ng, T.B. (2002). Luffangulin, a novel ribosome inactivating peptide from ridge gourd (*Luffa acutangula*) seeds. *Life Science*, 70, 899-906.
- Wollenweber, H.W. and Reinking, O.A. (1925). Aliquot Fusaria tropicola nova vel revisa. *Phythopathology*, 15, 155-169.
- Yan, K. Han, G., Ren, C., Zhao, S., Wu, X. and Bian, T. (2018). Fusarium solani infection depressed photosystem performance by inducing foliage wilting in apple seedlings. Frontiers Plant Science, 9(479), 1-10.
- Yang, P. (2014). The antibiosis of *Trichoderma asperellum* resistance plant pathogenic fungi. *Advanced Materials Research*, 1073(1076), 1067-1070.
- Yang, X., Lihua, C., Yong, X. and Shen, Q. (2011). Formulations can affect rhizosphere colonization and biocontrol efficiency of *Trichoderma* harzianum SQR-T037 against Fusarium wilt of cucumber. *Biology and Fertility of Soils*, 47(3), 239-248.
- Yang, X., Wang, X., Wei, M., Hikosaka, H. and Goto, E. (2009). Changes in growth and photosynthetic capacity of cucumber seedlings in response to nitrate stress. *Brazillian Journal of Plant Physiology*, 21(4), 309-317.
- Yasmeen, R. and Shaheed Siddiqui, Z. (2017). Physiological response of crop plants against *Trichoderma harzianum* in saline environment. *Acta Botanica Croatica*, 76(2), 1154-162.
- Ye, S.F., Yu, J.Q., Peng, Y.H., Zheng, J.H. and Zhou, L.Y. (2004). Incidence of Fusarium wilt in *Cucumis sativus* L. is promoted by cinnamic acid, an autotoxin in root exudates. *Plant and Soil*, 263(1), 143-150.
- Yedidia, I., Srivastva, W.A., Kapulnik, Y. and Chet, I. (2001). Effects of *Trichoderma harzianum* on microelement concentrations and increase growth of cucumber plants. *Plant Soil*, 235, 235-242.
- Yoder, J.A., Glenn, B.D., Benoit, J.B. and Zettler, L.W. (2008). The giant Madagascar hissing-cockroach (*Gromphadorhina portentosa*) as a source of antagonistic moulds: concerns arising from its used in a public setting. *Mycoses*, 51(2), 95-98.
- Zehra, A., Meena, M., Dubey, M.K., Amir, M. and Upadhyay, R.S. (2017). Synergistic effects of plant defense elicitors and *Trichoderma harzianum* on enhanced induction of antioxidant defense system in tomato against Fusarium wilt disease. *Botanical Studies*, 58(1), 44-58.
- Zhang, K., Su, Y.Y. and Cai.L. (2013). An optimized protocol of single spore isolation for fungi. *Cryptogamie Mycologie*, *34*, 349-356.
- Zhang, S., Raza, W., Yang, X., Hu, J., Huang, Q., Xu, Y., Liu, X., Ran, W. and Shen, Q. (2008). Control of Fusarium wilt disease of cucumber plants with the application of bioorganic fertilizer. *Biology and Fertility of Soils*, 44, 1073-1080.

- Zhang, C., Zheng, H., Wu, X., X.H., Han, K., Peng, J., Lu, Y., Lin, L., Xu, P., Wu, X., Li, G., Chen, J. and Yan, F. (2018). Genome-wide identification of new reference genes for RT-qPCR normalization in CGMMV-infected *Lagenaria siceraria*. *Peer Journal*, 6, 1-24.
- Zhao, Z., Liu, H., Luo, Y., Zhou, S., An, L., Wang, C., Jin, Q., Zhou, M. and Xu, J.R. (2014). Molecular evolution and functional divergence of tubulin superfamily in the fungal tree of life. *Scientific Reports*, 4(1), 1-13.
- Zhou, B., Yan, J., Shuo, Z. Xian, L. and Gao, Z. (2014). Phylogeny and pathogenicity of *Fusarium* spp. isolated from greenhouse melon soil in Liaoning Province. *Saudi Journal of Biological Science*, 21(4), 374-379.
- Zhu, W.Y., Huang, L., Chen, L., Yang, J.T., Wu, J.N., Qu, M.L., Yao, D.Q., Guo, C.L., Lian, H.L., He, H.L., Pan, J.S. and Cai, R. (2016). A high-density linkage map for cucumber (*Cucumis sativus* L.): Based on specific length amplified fragment (SLAF) sequencing and QTL analysis of fruit traits in cucumber. *Plant Science*. 7(437), 1-11.