

DEVELOPMENT OF SANDWICH AND LABEL-FREE ELECTROCHEMICAL IMMUNOSENSOR FOR DETECTION OF HER2 BREAST CANCER BIOMARKER

ZUR MIRA AZIZAH @ NOR HAIZA BINTI LAH

FS 2019 72



DEVELOPMENT OF SANDWICH AND LABEL-FREE ELECTROCHEMICAL IMMUNOSENSOR FOR DETECTION OF HER2 BREAST CANCER BIOMARKER

By

ZUR MIRA AZIZAH @ NOR HAIZA BINTI LAH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

August 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

DEVELOPMENT OF SANDWICH AND LABEL-FREE ELECTROCHEMICAL IMMUNOSENSOR FOR DETECTION OF HER2 BREAST CANCER BIOMARKER



An electrochemical immunosensor for determination of HER2 biomarker was developed using two different types of immunoreaction, which are label-free and sandwich assay. The fabrication of label-free immunosensor involved several steps; (1) pre-treatment of screen printed carbon electrode (SPCE) (2) the binding of primary anti-HER2 antibody to pre-treated SPCE via EDC/NHS (3) bovine serum albumin (BSA) blocking (4) the binding of HER2 antigen (5) electroanalytical performance of the label-free immunosensor. While, in sandwich immunosensor, lead sulphide quantum dots coated with secondary antibodies (Ab2-PbS QDs) were used as a label and the fabrication involved additional several steps which were; (1) synthesis and bioconjugation of PbS QDs with secondary onto-HER2 antibody (2) the sandwich of Ab2-PbS QDs onto label-free immunosensor (3) acid dissolution of Pb²⁺ from label and detection via square wave voltammetry (SWV). The electrochemical properties of the developed immunosensors were analysed by cyclic voltammetry (CV) and electron impedance spectroscopy (EIS) using Fe(CN)6^{-3/-4} as a redox probe. Under optimal condition, label-free and sandwich immunosensors were assessed by using differential pulse voltammetry (DPV) and SWV, respectively. The analytical performances revealed that both immunosensor give satisfactory sensitivity, selectivity and reproducibility towards HER2 cancer biomarker. For a linear range of 0.04-2.0 ng/ml of label-free immunosensor, the limit of detection obtained was 0.01 ng/ml while for sandwich immunosensor, the limit of detection obtained was 1.1 ng/ml for a linear range of 2.0-10 ng/ml Furthermore, excellent

i

recovery also obtained in human serum sample for both label-free and sandwich immunosensor. It was discovered that the recovery percentage of HER2 antigen in sandwich immunosensor was 102.06 % with RSD percentage of 7.51 %, while for label-free immunosensor, the percentage recovery is 97.10 % with RSD percentage of 10.89 %.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

PEMBANGUNAN IMMUNOSENSOR ELEKTROKIMIA SANDWIC DAN TANPA LABEL BAGI PENGESANAN BIO PENANDA BARAH PAYUDARA HER2

Oleh

 ZUR MIRA AZIZAH @ NOR HAIZA BINTI LAH

 Ogos 2018

 Pengerusi
 : Shahrul Ainliah Alang Ahmad, PhD

 Fakulti
 : Sains

Immunosensor elektrokimia untuk penentuan HER2 biomarker telah dibangunkan menggunakan dua jenis imunoreaksi yang berbeza, yang bebas label dan sandwic assay. Pembuatan imunosensor bebas label melibatkan beberapa langkah; (1) pra-rawatan elektrod karbon bercetak skrin (SPCE) (2) proses immobilisasi antibodi anti-HER2 primer untuk SPCE yang telah diaktifkan melalui EDC / NHS (3) penyebaran serum albumin bovine (BSA) HER2 antigen (5) prestasi elektroanalisis imunosensor bebas label. Sementara itu, dalam imunosensor sandwic, titik kuantum yang dilapisi dengan antibodi sekunder (Ab2-PbS QDs) telah digunakan sebagai label dan fabrikasi melibatkan beberapa langkah tambahan; (1) sintesis dan biokonjugasi PbS QDs dengan antibodi ke-HER2 sekunder (2) sandwic Ab2-PbS QDs ke immunosensor bebas larutan label (3) pembubaran asid Pb²⁺ dari label dan pengesanan melalui voltammetry gelombang persegi (SWV). Sifat-sifat elektrokimia imunosensor telah dianalisis oleh voltammetry kitaran (CV) dan spektroskopi impedans elektron (EIS) menggunakan Fe (CN)6-3 / -4 sebagai penyelidikan redoks. Di dalam keadaan optimum, imunosensor bebas label dan sandwic dinilai dengan menggunakan voltmeter nadi (DPV) dan SWV, masing-masing. Keputusan analitikal mendedahkan bahawa kedua-dua imunosensor memberikan sensitiviti, selektiviti dan keboleh ulangan yang memuaskan terhadap biopenanda HER2. Untuk julat linear 0.04-2.0 ng/ml immunosensor bebas label, had pengesanan yang diperoleh adalah 0.01 ng/ml manakala untuk immunosensor sandwic, had pengesanan yang diperoleh adalah 1.1 ng / ml untuk julat linear 2.0-10 ng / ml Selain itu, pemulihan yang sangat baik juga diperolehi dalam sampel serum manusia untuk kedua-dua bebas label dan imunosensor sandwic. Hasil penemuan

iii

menunujukkan peratusan pemulihan antigen HER2 dalam imunosensor sandwic adalah 102.06% dengan peratusan RSD sebanyak 7.51%, manakala bagi imunosensor bebas label, pemulihan peratusan adalah 97.10% dengan peratusan RSD sebanyak 10.89%.



ACKNOWLEDGEMENT

First and foremost I would like to thank Allah the almighty for giving the strength to finish up this thesis. Without His love and blessing, this thesis would not be possible.

I would like to express my gratitude to my main supervisor, Dr. Shahrul Ainliah Alang Ahmad for her guidance and advice through this research project. Without her knowledge and expertise, this research project would not be possible.

I am highly indebted to my co-supervisor, Dr. Mazliana Ahmad Kamarudin on her help during my process in learning on synthesis of quantum dots. I also would like to extend my gratitude to my other co-supervisors, Dr. Jaafar Abdullah and Dr. Janet Lim Hong Ngee for their advice for my research project.

Special appreciations also go to my lab mates in lab 103 for their useful guide in conducting experiments.

Finally, I would like to express my gratitude to my parents, Puan Jamaliah and Encik Lah for their prayer and encouragement. Without their support, I would not be able to finish up this thesis.

Thank you.

ZUR MIRA AZIZAH @ NOR HAIZA BINTI LAH

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Shahrul Ainliah Alang Ahmad, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Chairman)

Mazliana Ahmad Kamarudin, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Member)

Jaafar Abdullah, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Member)

Janet Lim Hong Ngee, PhD Senior Lecturer Faculty of Science Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fullyowned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
 - there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No.:	

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee: _	
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

ABSTR ABSTR ACKNO APPRO DECLA LIST O LIST O LIST O	ACT AK OWLEDG OVAL RATION F TABLE F FIGUR F ABBR	SEMENT I ES RES EVIATIONS	Page I iii v vi viii xiii xiv xvii
СНАРТ	ER		
1		DDUCTION	4
	1.1	Background	1
	1.2	Lead sulphide quantum dots (PhS ODs)	2 1
	1.5	Problem statement and research motivation	4 5
	1.4	Objectives	7
	1.0	Objectives	'
2	LITER	RATURE REVIEW	
	2.1	Immunosensors	8
	2.2	Pre-treatment of Electrode	10
	2.3	Immobilization of antibodies onto electrode	14
	2.4	Conjugation of quantum dots with antibodies	19
		2.4.1 Adsorption	19
		2.4.2 Covalent binding	20
	2.5	Quantum dots-based sensor	21
		2.5.1 Electrochemical immunosensor	22
		2.5.2 Electrochemiluminescent immunosensor	23
		2.5.3 Fluorescence immunosensor	25
	2.6	Acid dissolution for determination of quantum dots	28
	2.7	Summary	31
3	МАТЕ	FRIALS AND METHODS	
J	3.1	Materials and reagents	32
	3.2	Instrumentations	32
	0.2	3.2.1 Electrochemical analysis	32
		3.2.1.1 Cyclic Voltammetry	32
		3.2.1.2 Electron Impedance Spectroscopy	34
		3.2.1.3 Differential Pulse Voltammetry	35
		3.2.1.4 Square Wave Voltammetry	36
		3.2.2 UV-visible spectroscopy (UV-Vis)	36
		3.2.3 Fourier transform infrared (FTIR)	37
		3.2.4 Transmission electron microscopy (TEM)	37
		3.2.5 X-ray photoelectron spectroscopy (XPS)	38
		3.2.6 Zeta potential	39
		3.2.7 X-ray diffraction (XRD)	39

 $\widehat{}$

	3.3	Methodology	40
		3.3.1 Synthesis and bioconjugation of lead sulfide quantum dots (PbS QDs) with antiHER2	40
		3.3.2 Fabrication of label-free immunosensor (SPCE/Ab1/HER2)	41
		3.3.3 Fabrication of the sandwich immunosensor (SPCE/Ab1/HER2/Ab2-Pbs QDs)	42
		3.3.4 Electroanalytical performances	43
		3.3.4.1 Label-free immunosensor	43
		3.3.4.2 Sandwich immunosensor	43
		3.3.5 Sensitivity, selectivity, repeatability and	44
		reproducibility studies	b.
		3.3.6 Serum sample analysis	45
4	RES	SULTS AND DISCUSSION	
	4.1	Synthesis and bioconjugation of PbS QDs with	46
		anti-HER2 antibody	
		4.1.1 Transmission Electron Microscopy	46
		4.1.2 UV-Visible	47
		4.1.3 Zeta Potential	47
		4.1.4 XRD Analysis	48
	4.2	Fabrication of label-free immunosensor	50
		4.2.1 Optimization of anodic pre-treatment of screen printed carbon electrode (SPCE)	50
		4.2.2 Optimization of experimental parameters	51
		4.2.3 Fourier Transform Infrared (FTIR) Analysis	54
	4.3	Fabrication of sandwiched immunosensor	55
		4.3.1 Optimization of bioconjugation of PbS QDs (Ab2-PbS QDs) onto SPCE	55
		4.3.2 Optimization of acid dissolution of Pb ²⁺ ions	56
		4.3.3 Control Study	59
	4.4	Characterization of label-free immunosensor	60
		(SPCE/AD1/HER2) and sandwiched	
		1111111110Sells01 (SPCE/AD1/HERZ/AD2-PDS QDS)	60
		4.4.1 A-ray photoelectron spectroscopy analysis	62
		4.4.2 Electrochemical analysis	63
		4.4.2.1 Gyclic voltarimetry analysis 4.4.2.2 Electron impedance spectroscopy	64
	45	Flectroanalytical performances	66
	ч .5	4 5 1 Label-free immunosensor	66
		4.5.2 Sandwich immunosensor	68
_			
5		ICLUSION AND RECOMMENDATIONS FOR	
	51	Conclusion	72
	5.2	Recommendation for future research	73
	2		
REFERE	NCES	5	74

6

APPENDICES BIODATA OF STUDENT LIST OF PUBLICATIONS

84 88 89



LIST OF TABLES

Table		Page
 2.1 The summary of pre-tree 2.2 The summary on strate electrodes for conjugat immunosensor applies 	eatment of electrodes egies used in different type of ion of different type of biomolecules in	13 18
2.3 The summary of application of ap	ation of quantum dots in different	27
 4.1 Zeta potential of PbS G 4.2 Comparison of HER2 r methods 	QDs and Ab2-PbS QDs esults for the recently proposed	48 69

 \bigcirc

LIST OF FIGURES

Figure		Page	
1.1	The progression of normal cancer into malignant cancer	1	
1.2	Comparison between (a) labelled and (b) label-free detection method	4	
2.1	The depiction of integrated unit that form a biosensor	8	
2.2	Sandwich-type electrochemical immunosensor for detection of HBsAg based on palladium nanoparticles/δ-manganese dioxide/halloysite nanotube		
2.3	In situ generation of an aryl diazonium cation followed by electrochemical reduction of the generated aryl diazonium cation to effect carbon-carbon coupling		
2.4	Schematic display of the different steps involved in the construction of an electrochemical immunosensor for PYY involving the grafting of 4-ABA diazonium salt onto a rGO-modified GCE and the covalent immobilization of anti-PYY		
2.5	Schematic representation of the preparation of the (A) DA- Fe3O4-FC-Ab2 nanoparticles and (B) immunosensor		
2.6	Schematic representation for preparing the complex of antibody (IgG) and ProteinA/PEG–QD	20	
2.7	Illustration of the DNA assay developed on the biosensor surface	23	
2.8	Fabrication process and the detection mechanisms of the ECL immunosensor		
2.9	Simultaneous detection of two proteins using PbS and CdS QDs labels and a mercury film modified electrode		
2.10	Schematic representation of the electrochemical stripping detection of DNA hybridization based on PbS nanoparticles labelled oligonucleotides DNA probes		
3.1	Cyclic voltammogram excitation signal	33	
3.2	Voltammogram of a Single electron oxidation-reduction	34	
3.3	The depiction of XPS instrumentation	38	
3.4	(a) Synthesis of PbS QDs (b) bioconjugation of PbS QDs with secondary antiHER2 antibody (Ab2)	41	
3.5	The development of label-free immunosensor	42	

 \bigcirc

- 3.6 The depiction of development of sandwich immunosensor 43 for HER2 detection based on PbS QDs (a) the fabrication of sandwich immunosensor and (b) acid released procedure
- 4.1 TEM images and size distribution of (a) PbS QDs (b) Ab2- 46 PbS QDs
- 4.2 Absorption spectra of PbS QDs and Ab2-PbS QDs 47
- 4.3 The mechanism of conjugation between PbS QDs and 48 HER2 antibody
- 4.4 XRD analysis of (a) galena (b) Ab2-PbS QDs and (c) PbS 49 QDs
- 4.5 Representative result of optimization of SPCE in 0.1 M KCl 51 with 10 mM potassium ferricyanide as redox probe (a) Concentration of H2SO4 (b) Potential applied (c) Pretreatment time (CV Range: -0.8 V -0.8 V, 100 mV/s)
- 4.6 Example of differential pulse voltammetry peak position 52 (peak position: 0.02 V)
- 4.7 Representative result of the optimization of (a) type of 53 electrolyte (b) pH of electrolyte (c) Ab1 incubation time and (d) HER2 antigen incubation time in 10 mM PBS buffer of pH 5.5 (DPV: -0.3 V to 0.3 V)
- 4.8 FTIR spectra of (a) bare SPCE, (b) activated SPCE and (c) 54 SPCE/Ab1/HER2
- 4.9 Representative result of the optimization of (a) conjugation 56 time (b) concentration of Ab2 and (c) incubation time on SPCE of Ab2-PbS QDs conjugated in 10 mM PBS buffer of pH 5.5 (DPV: -0.3 V 0.3 V)
- 4.10 The reference peak for Pb²⁺ ions released in the electrolyte 57 solution
- 4.11 Effect of dissolution time, deposition time and deposition 58 potential of acid released procedure (SWV: -1.0 V to -0.4 V)
- 4.12 (a) The comparison of sample with and without 60 bioconjugation of Ab2-PbS QDs and PbS QDs (b) The comparison of sample with and without formation of mercury film on SPCE (c) The comparison of sample with and without HER2 antigen on sandwich immunosensor
- 4.13 Narrow scans (a) C1s of SPCE/Ab1/HER2 (b) C1s of 62 SPCE/Ab1/HER2/Ab2-PbS QDs (C) O1s of SPCE/Ab1/HER2 (d) O1s of SPCE/Ab1/HER2/Ab2-PbS SPCE/Ab1/HER2 QDs (e) N1s of (f) N1s of SPCE/Ab1/HER2/Ab2-PbS QDs S2p (g) of SPCE/Ab1/HER2/Ab2-PbS SPCE/Ab1/HER2 (h) S2p of

QDs (i) Pb4f of SPCE/Ab1/HER2 (j) Pb4f of SPCE/Ab1/HER2/Ab2-PbS QDs

- 4.14 The cyclic voltammograms of bare electrode, activated 64 electrode, SPCE/Ab1/HER2 and SPCE/Ab1/HER2/Ab2-PbS QDs in 10 mM PBS buffer of pH 5.5 with 10 mM potassium ferricyanide as redox probe. (CV Range: -0.8 V-0.8 V, 100 mV/s)
- 4.15 Electron impedance spectroscopy of bare electrode, 65 activated electrode, modified electrode and electrode labelled-QDs in 10 mM PBS buffer of pH 5.5 with 10 mM potassium ferricyanide as redox probe. (0.15 V at different electrode)
- 4.16 The illustration of the Randles equivalent circuit 65
- 4.17 Calibration curve. Relationship between peak current and 66 concentration of HER2 biomarker (DPV: -0.3 V to 0.3V, LOD = 26.78 ng/ml)
- 4.18 (a) The reproducibility of the method in 5 samples of 68 sandwich immunosensors (RSD % = 4.68 %) (b) The stability of sandwich immunoassay up to 6 cycles (c) The interference study of 4 samples (1) 100 ng/ml HER2 (2) 100 ng/ml HER2 + 0.001 g/ml glucose (3) 100 ng/ml + 0.001 g/ml BSA (4) 100 ng/ml + 0.001 g/ml AA (d) Comparison of human serum dilution factor to control
- 4.19 (a) SWV of sample solution taken at different concentration 69 of HER2 (A): 0 (B): 1 (C): 2 (D): 4 (E): 8 ng/ml (F): 10 ng/ml (G): 40 ng/ml (H): 100 ng/ml and (b) Calibration curve. Relationship between peak current and concentration of HER2 biomarker (SWV: -1.0 V to -0.4 V, LOD = 13.04 ng/ml)
- 4.20 (a) The reproducibility of the method in 5 samples of sandwich immunosensors (RSD % = 4.68 %) (b) The stability of sandwich immunoassay up to 6 cycles (c) The interference study of 4 samples (1) 100 ng/ml HER2 (2) 100 ng/ml HER2 + 0.001 g/ml glucose (3) 100 ng/ml + 0.001 g/ml BSA (4) 100 ng/ml + 0.001 g/ml AA
 A1.1 XPS wide scan of label-free immunosensor 84
- A2.1 XPS wide scan of sandwich immunosensor 85
- A3.1 The image of PbS QDs at 50 nm resolution 86
- A4.1 The image of Ab2-PbS QDs at 50 nm resolution 87

LIST OF ABBREVIATIONS

	Ab1	Primary Antibody
	Ab2	Secondary Antibody
	AFP	Alpha Fero Protein
	Anti-CEA	Anti-Carcinoembryonic
	Anti-cTnT	Anti-Cardiac Troponin T
	Anti HER2	Anti-Human Epidermal Growth Factor Receptor 2
	APTES	(3-Aminopropyl)triethoxysilane
	AuNPs	Gold nanoparticles
	BSA	Bovine serum albumin
	CDI	Carbonylimidazole
	C-Dot	Carbon dot
	CDSe/CdZns	Cadmium Selenide/Cadmium Zinc
	CDTe	Cadmium telluride
	CEA	Carcinoembryonic Antigen
	CLB	Clenbuterol
	CNT	Carbon Nanotube
	CV	Cyclic Voltammetry
	DNA	Deoxyribonucleic acid
	DPV	Differential Pulse Voltammetry
	DTG	Dithioglycerol
	ECL	Electrochemiluminescence
	EDC	1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride
	EIS	Electrochemical Impedence Spectroscopy
	ELISA	Enzyme-Linked Immunosorbent Assay
	EV	Extracellular Vesicles
	FISH test	Fluorescence In Situ Hybridization Test

	FRET	Fluorescence Resonance Energy Transfer
	FTIR	Fourier Transform Infrared Spectroscopy
	GCE	Glassy carbon electrode
	GS	Graphene Sheet
	LOD	Limit of Detection
	NHS	N-Hydroxysuccinimide
	HER2	Human Epidermal Growth Factor Receptor 2
	HNT	Halloysite Nanotube
	HRP	Horse Radish Peroxidase
	IgG	Immunoglobulin G
	ІНС	Immunohistochemistry
	IR	Infrared
	ІТО	Indium Tin Oxide
	ксі	Potassium chloride
	mAb	Monoclonal Antibody
	МВ	Magnetic Beads
	MCGCE	Magnetic Controlled Glassy Carbon Electrode
	MES	2-(N-morpholino)ethanesulfonic Acid
	MNP	Magnetic Nanoparticles
	MSN	Mesoporous Silica Nanoparticles
	OA	Okadaic Acid
	PbS QDs	Lead Sulphide Quantum Dots
	PDA	Polydopamine
	PSA	Prostate Specific Antigen
	QDs	Quantum Dots
	rGO	Reduced Graphene Oxide
	SPCE	Screen Printed Carbon Electrode
	SPE	Screen Printed Electrode
	ТЕМ	Transmission Electron Microscopy

xviii

TGLThioglycerolTPSATotal Prostate Specific AntigenUV-VISUltraviolet VisibleXPSX-Ray Photoelectron SpectroscopyXRDX-Ray Diffraction



CHAPTER 1

INTRODUCTION

1.1 Background

Breast cancer is a tumour that can either be malignant or benign in which latter does not invade other tissues. However a malignant tumour can invade other parts of body through lymph system, hence causing a cancer.



Figure 1.1 : The progression of normal cells into malignant cancer (Australia, 2017)

Figure 1.1 shows the cancer progress when there are changes in cells. These cells produce signals to control how much and how often the cells divide. However, the faulty in this cell can cause the cells to multiply abnormally and cause it to form tumour. This tumour becomes a cancer once it invades the lymphatic system and blood vessels.

There are three current tests that have been used extensively for detection of breast cancer. They are screening, diagnostic and monitoring tests. These tests depend on categories of breast cancer related issues. Screening tests such as mammogram is used to identify breast cancer at early stage before the cancer cell is further developed so that the treatment is easier. While, diagnostic tests such as biopsy are tested in order to identify the symptoms of breast cancer, which further can identify types of treatment needed. Furthermore, the test can also determine if the cancer cells have spread or not. Lastly, monitoring tests are used to determine how well the treatment is and can be used to check any recurrence.(Breastcancer.org, 2017)

Among the available test, screening test is significant to obtain information of breast cancer cell before the symptoms arise because early detection of cancer can increase the chances of successful treatment. There are different kinds of screening test available including the physical exam and history, laboratory tests, imaging procedures and genetic tests. From the test mentioned, laboratory tests are sensitive and more reliable detection which involves the use of blood marker formed from the response of body towards the tumours. These biomarkers can be found in blood, urine, or body tissue of a cancer patient.

In this project, a simple and disposable of label-free and label immunosensor integrating the lead sulfide quantum dots (PbS QDs) as a reader for detection of HER2 cancer biomarker was constructed. PbS QDs was synthesized with thioglycerol (TGL) and dithioglycerol (DTG) as a capping agent to produce PbS QDs with hydroxyl surfaces. The as-synthesized PbS QDs was conjugated with antiHER2 antibody by using carbonylimidazole (CDI) as a linker. Screen printed carbon electrode (SPCE) was used as the platform for the immunosensor development. In order to improve the electrode interface, the transducer surface was functionalized. The surface of SPCE was electrochemically pre-treated to form carboxyl functional group which then activated with EDC/NHS linker for immobilization of primary anti-HER2 (Ab1). The quantification of HER2 biomarker was performed through acid dissolution of the bioconjugated PbS QDs. The released cations were transferred to an electrochemical cell and measured using square wave voltammetry (SWV).

1.2 Immunosensor

Immunosensor is a biosensor with affinity-ligand based in which the immunological reaction is coupled with transducer (Azam *et al.*, 2014). The key to the development of an immunosensor is the non-covalent specific binding of antibodies to antigens. There are numerous analytical technique that took advantage of this specific interaction in many industries such as clinical diagnostic and analysis, food control safety, environmental monitoring, forensic analysis, drug screening and etc. In this regard, immunosensor emerges as a significant, continuously growing scientific industry (Kokkinos *et al.*, 2016).

There are many types of technique used for detection in immunosensor, for instance, electrochemical, optical, microgravimetric and thermometric. All type of detection can either be labelled or non-labelled. The non-labelled usually detect the physical changes during the immunology reaction while the



labelled sensor used the signal generating label which is more sensitive and versatile detection mode (Luppa *et al.*, 2001)

Recently, electrochemical immunosensor appear as a more promising alternative for immunosensing. This is due to the fact that it is possible for miniaturize modern microelectronics which allows building of microelectrodes that useful for multiplexing. Furthermore, it is well suited for detection of very small volumes of samples ranging from microliters to nanoliters. In addition, electrochemical approach is favourable due to low cost and large scale production of electronic devices for high-throughput analysis. Apart from that, it is suitable for complex and coloured sample because of the scarcity of electrochemical interference in real samples (Ricci *et al.*, 2012).

The detection mechanism of an electrochemical immunosensor employs antibodies as capture and detection means to produce electrical charges for the quantitative analysis of target molecules. One of the main issues in developing immunosensor system is to achieve high sensitivity of detection. Numerous current nanotechnologies allowed the development of innovative electrochemical biosensors with high sensitivity. This is achieved by employing various nanomaterials which able to facilitate the electron transfer and carry signal tracers in combination with surface modification and bioconjugation techniques (Cho *et al.*, 2018).

There are many different formats that can be used to build an electrochemical immunosensor. In this case, two approaches can be used which are label-free and labelled formats. For the label-free format (Figure 1.2 (b)), it able to sense directly the protein bind to the antibody which allow detecting proteins as they bind in real-time. (Mehrabani *et al.*, 2014). While for labelled format (Figure 1.2 (a)) or most commonly known as sandwich format, it used the immobilized antigens and labelled antibodies. Sandwich format generally adopted to prevents all the problems related to antibody immobilization such as loss of affinity and orientation of the immobilized protein.



Figure 1.2 : Comparison between (a) labelled and (b) label-free detection method (Mehrabani *et al.*, 2014)

1.3 Lead Sulfide Quantum Dots (PbS QDs)

Lead sulphide is an inorganic compound with a chemical formula of PbS. The naturally occurring ore of this compound is known as galena. It is one of the earliest semiconductors found due to the fact that it is easily crystallized in sodium chloride. PbS most commonly used in infrared detector, photo optic application and to enhance heat conduction and regulating friction coefficient in friction industry. It has a molecular weight of 239.90 g/mol with a lattice constant of 5.936 Å. While for the electrical properties, it has a small band gap which is 0.37 eV (Azo Materials, 2018).

Lead sulphide quantum dots (PbS QDs) have attracted much attention in recent years due to the excellent properties compare to the bulk PbS. It is most used in single junction and tandem photovoltaics (PV) applications due to a band gap that can be size-tuned from 0.41 to 1.6 eV(Rekemeyer *et al.*, 2017). Furthermore, PbS QDs with optical transitions in the near infrared region are now used in third-generation photovoltaic cells, photodetectors, and light-emitting diodes (Litvin *et al.*, 2017). The availability for synthetic route for creating size tuned PbS QDs and high stability as compared to PbSe QDs making it more favourable for these applications (Márquez *et al.*, 2017).



Although PbS QDs mostly used in electronics, energy conversion and storage (Pal *et al.*, 2017), several research have generated enormous application of this material into biosensing technology. Therefore, attempts have been made in recent years for the inclusion of PbS QDs in biosensor. For instance, an electrochemical biosensor based on PbS QDs for detection of carcinoembryonic antigen tumor marker (Wang *et al.*, 2008), Escherichia Coli O157:H7 Detection (Wang *et al.*, 2015), DNA hybridization (Zhu *et al.*, 2004b)and simultaneous detection of Rabbit immunoglobulin G antigen (IgG) and carcinoembryonic antigen (CEA) (Qian *et al.*, 2011) was developed.

In thid regard, the present study explores the application of PbS QDs for detection of breast cancer biomarker using HER2 antigen.

The toxicity of lead is the main concern for the PbS QDs to be employed for clinical study purposes. The toxicity of lead is very much dependent upon the route of exposure whether, dose, chemical structure, solubility, particle size, matrix incorporation and other physiological and physicochemical factors (Dieter et al., 1993). In this regard, several researchers have been investigated the effect of lead concentration towards biological organism. For instance, Dieter and co-workers (1993) have initiated a study on lead bioavailability in F344 rats that have been fed with lead acetate, lead oxide, lead sulphide, or lead ore concentrate from Skagway Alaska. From the study, there is no mortality and symptoms of lead toxicity observed (Dieter et al., 1993). Apart from that it was found that PbS compound have no effect on red blood cells and is highly insoluble and stable at the pH of bloods hence almost non-toxic to human (Bischoff et al., 1928). According to world health organization (WHO), there is also limited evidence of lead compounds carcinogenicity. However, there is sufficient evidence of carcinogenic of lead compounds towards experimental animal. The evaluation of lead compounds overall bring to the probability of carcinogenic to human (Group 2A) (World Health Organization (WHO), 2018). Overall, the toxicity of PbS QDs is negligible attributed to low bio-availability which consequently leads to low risk.

1.4 Problem Statement And Research Motivation

Enzyme-Linked Immunosorbent Assay or ELISA is one of the current clinical diagnostic tools for early detection of breast cancer. It is often used in biomedical research for quantification of specific antigen or antibodies in a given sample. It is highly sensitive and most of ELISA kits are portable and ready to use. However, ELISA has few limitations observed. Firstly, over a long period of time, the enzyme-mediated colour most likely to change, hence result in a false result. Secondly, the non-specific binding of antigen and antibody could occur lead to false-positive result (Gan and Patel, 2013).

Substantial studies on immunosensors have been carried out based on piezoelectric transducer. The working principle of the piezoelectric immunosensor is based on formation of antigen-antibody complexes that increases the mass as compared with antigen or antibody alone which detected through a device such as quartz crystal balance or a microcantilever (Moina and Ybarra, 2012b). Due to its simplicity, low cost and real time response, it gains significant attention in the field of bioanalytical assays and characterization as one of the competitive tools (Montoya *et al.*, 2009). However, it possesses some drawbacks that set as a challenge for clinical analysis such as complex instrumentation and long incubation time with samples.

As an alternative, optical immunosensors have been widely used as a mean of immunosensing. Typically, biological element is immobilized on the surface of transducer where it will generate optical signal when interacted with target analyte, such as fluorescence.

Fluorescence technique for detection of breast cancer is one of the most widely used immunosensor where the conjugate antibody is labelled with fluorescent probe. The antigen antibody complexes formed cause optical changes towards surrounding. However, most fluorescence immunoassay requires a complex instrumentation. Furthermore, fluorescence tends to have photobleaching effect (Moina and Ybarra, 2012a).

To overcome these issues, electrochemical technique was used as a method for breast cancer biomarker detection. This technique have several advantages over other methods, such as, miniaturised, easy to use and portable (Hammond *et al.*, 2016). Apart from that, electrochemical immunosensor is a rapid, simple and cost effective method (Tian *et al.*, 2016). Electrochemical detection involves the conversion of the event of antigenantibody interaction into electrical signal. This technique able to display high specificity and selectivity of antibody-antigen complexes by monitoring the current response. In this regard, this technique was employed for this study as a method on detection of HER2 breast cancer biomarker based on label-free and sandwich immunosensor.

Recently, quantum dots (QDs) attract much attention due to their superior properties. For instance, effective suppression of non-specific adsorption of biomolecules at the surface and less aggregation is the most significant criteria. Furthermore, the colloid stability can be improved through functionalization of the surface of QDs (Drbohlavova *et al.*, 2009). Due to this pursuit, QDs was employed as a label for the fabrication of sandwich immunosensor. In this regard, PbS QDs was synthesized and bioconjugated with secondary anti HER2. The synthesis process involved the use of lead(II) acetate and sodium sulphide as the precursors with two additional ligand which are thioglycerol (TGL) and dithioglycerol (DTG). Previous reported procedure have shown that the PbS QDs were more stable for several months upon usage of TGL/DTG instead of using either one of them (Zhao *et al.*, 2005). Apart from that, the hydroxyl tail groups available are easily tunable for the bioconjugation process through carbonylimidazole (CDI) linker.

The utilization of PbS QDs as a label sensing for detection of HER2 electrochemical sandwich immunosensor has not yet been explored. Hence, the suitability and stability of the PbS QDs conjugation is still unknown. In order for the anti-HER2 conjugated PbS QDs effectively immobilized onto the

surface of modified screen printed carbon electrode, the optimum condition of layer formation was studied.

1.5 Objectives

The aim of the research is to develop label-free and label electrochemical immunosensors for the detection of cancer biomarkers. In sandwich-type (label) immunosensor, conjugated PbS QDs was used as a label. In order to achieve this overall goal, the specific technical objectives are:

- 1. To synthesize and bioconjugate PbS QDs with antibody
- 2. To develop label-free and sandwich immunosensor
- 3. To analyse the performance of label-free and sandwich immunosensor towards different concentration of HER2

REFERENCES

- AL-KHAFAJI, Q., et al. 2012. An electrochemical immunoassay for HER2 detection. *Electroanalysis*, 24, 735-742.
- AUSTRALIA, C. C. 2017. What is cancer? [Online]. Available: <u>http://www.cancer.org.au/about-cancer/what-is-cancer/</u> [Accessed 16 June 2017].
- AZAM, M. S., et al. 2014. Advancements and application of immunosensors in the analysis of food contaminants. *Nusant. Biosci*, 6, 186-195.
- AZO MATERIALS. 2018. Lead (II) Sulfide (PbS) Semiconductors [Online]. Available: https://www.azom.com/article.aspx?ArticleID=8440 [Accessed 24 Sept 2018].
- BHUCKORY, S., et al. 2016. Direct conjugation of antibodies to the ZnS shell of quantum dots for FRET immunoassays with low picomolar detection limits. *Chemical Communications*, 52, 14423-14425.
- BISCHOFF, F., et al. 1928. Studies on the toxicity of various lead compounds given intravenously. Journal of Pharmacology and Experimental Therapeutics, 34, 85-109.
- BOLAND, S., et al. 2008. Designing stable redox-active surfaces: chemical attachment of an osmium complex to glassy carbon electrodes prefunctionalized by electrochemical reduction of an in situ-generated aryldiazonium cation. *Langmuir*, 24, 6351-6358.
- BRAIEK, M., et al. 2012. An electrochemical immunosensor for detection of Staphylococcus aureus bacteria based on immobilization of antibodies on self-assembled monolayers-functionalized gold electrode. *Biosensors*, 2, 417-426.
- BREASTCANCER.ORG. 2017. *HER2 Status* [Online]. Available: <u>www.breastcancer.org/symptoms/diagnosis/her2</u> [Accessed 19 June 2017].
- BUI, M.-P. N., *et al.* 2012. Electrochemical determination of cadmium and lead on pristine single-walled carbon nanotube electrodes. *Analytical Sciences*, 28, 699-704.
- CANEVARI, T. C., *et al.* 2016. High performance electrochemical sensors for dopamine and epinephrine using nanocrystalline carbon quantum dots obtained under controlled chronoamperometric conditions. *Electrochimica Acta*, 209, 464-470.

- CHEMISTRY LIBRETEXTS. 2018. *Cyclic Voltammetry* [Online]. Available: https://chem.libretexts.org/Textbook_Maps/Analytical_Chemistry/Supp lemental_Modules_(Analytical_Chemistry)/Instrumental_Analysis/Cycli c_Voltammetry.
- CHEN, J., *et al.* 2016. Direct water-phase synthesis of lead sulfide quantum dots encapsulated by β-lactoglobulin for in vivo second near infrared window imaging with reduced toxicity. *Chemical Communications*, 52, 4025-4028.
- CHEN, S., et al. 2008. Antibody microarrays for protein and glycan detection. Clinical Proteomics: From Diagnosis to Therapy, 101-111.
- CHO, I.-H., et al. 2018. Current Technologies of Electrochemical Immunosensors: Perspective on Signal Amplification. Sensors, 18, 207.
- CUI, W., *et al.* 2017. A new and facile strategy for determination of lead and cadmium using silver electrodes manufactured from digital versatile discs. *Chemical Research in Chinese Universities*, 33, 799-803.
- DAHAGHIN, Z., et al. 2018. Simultaneous determination of lead (II) and cadmium (II) at a glassy carbon electrode modified with GO@ Fe3O4@ benzothiazole-2-carboxaldehyde using square wave anodic stripping voltammetry. Journal of Molecular Liquids, 249, 1125-1132.
- DEER, C., et al. 2005. Synthesis of Mn2+-doped PbS quantum dots and their spectroscopic properties. American laboratory, 37, 17-20.
- DIETER, M. P., et al. 1993. Comparison of lead bioavailability in F344 rats fed lead acetate, lead oxide, lead sulfide, or lead ore concentrate from Skagway, Alaska. Journal of Toxicology and Environmental Health, Part A Current Issues, 39, 79-93.
- DIFFERENCEBETWEEN.COM. 2012. Difference Between Wavelength and Wavenumber [Online]. Available: https://www.differencebetween.com/difference-between-wavelengthand-vs-wavenumber/ [Accessed 24 Sept 2018].
- DOLDÁN, X., et al. 2016. Electrochemical Sandwich Immunosensor for Determination of Exosomes Based on Surface Marker-Mediated Signal Amplification. *Analytical Chemistry*, 88, 10466-10473.
- DRBOHLAVOVA, J., et al. 2009. Quantum dots—characterization, preparation and usage in biological systems. International journal of molecular sciences, 10, 656-673.
- EL MHAMMEDI, M. A., et al. 2010. Square wave voltammetry for analytical determination of cadmium in natural water using Ca10 (PO4) 6 (OH) 2-modified platinum electrode. American Journal of Analytical Chemistry, 1, 150.

- EMAMI, M., *et al.* 2014. An electrochemical immunosensor for detection of a breast cancer biomarker based on antiHER2–iron oxide nanoparticle bioconjugates. *Analyst*, 139, 2858-2866.
- FAN, F., et al. 2013. Determination of lead by square wave anodic stripping voltammetry using an electrochemical sensor. Analytical Sciences, 29, 571-577.
- FANJUL-BOLADO, P., *et al.* 2008. Electrochemical characterization of screen-printed and conventional carbon paste electrodes. *Electrochimica Acta*, 53, 3635-3642.
- FELIX, F. S., *et al.* 2005. Determination of Cd, Pb and Ni by square wave stripping voltammetry in particulate matter collected in workplace atmosphere of some Brazilian industrial foundries. *Journal of the Brazilian Chemical Society*, 16, 801-807.
- FLORES-ACOSTA, M., et al. 2003. Excitonic absorption of spherical PbS nanoparticles in zeolite A. *Solid state communications*, 128, 407-411.
- FOUBERT, A., et al. 2016. Bioconjugation of quantum dots: Review & impact on future application. *TrAC Trends in Analytical Chemistry*, 83, 31-48.
- FRASCO, M. F., et al. 2009. Semiconductor quantum dots in chemical sensors and biosensors. Sensors, 9, 7266-7286.
- GAMRY INSTRUMENTS. 2018a. Basics of Electrochemical Impedance Spectroscopy [Online]. Available: https://www.gamry.com/applicationnotes/EIS/basics-of-electrochemical-impedance-spectroscopy/ [Accessed 1 May 2017].
- GAMRY INSTRUMENTS. 2018b. Differential Pulse Voltammetry Purpose [Online]. Available: https://www.gamry.com/Framework%20Help/HTML5%20-%20Tripane%20-%20Audience%20A/Content/PV/Experimental Techniques/Differential

%20Pulse%20Voltammetry/Purpose.htm [Accessed 1 May 2017].

GAMRY INSTRUMENTS. 2018c. Square-wave Voltammetry Purpose [Online]. Available: https://www.gamry.com/Framework%20Help/HTML5%20-%20Tripape%20%20Audience%20A/Content/P\//Experimental_Techn

%20Tripane%20%20Audience%20A/Content/PV/Experimental_Techn iques/Square-wave%20Voltammetry/Purpose.htm [Accessed 23 Sept 2018].

GAN, S. D., et al. 2013. Enzyme immunoassay and enzyme-linked immunosorbent assay. Journal of Investigative Dermatology, 133, 1-3.

- GEOCHEMICAL INSTRUMENTATION AND ANALYSIS. 2018. X-ray Powder Diffraction (XRD) [Online]. Available: https://serc.carleton.edu/research_education/geochemsheets/techniqu es/XRD.html [Accessed 23 Sept 2018].
- GOLDMAN, E. R., et al. 2002. Avidin: a natural bridge for quantum dotantibody conjugates. Journal of the American Chemical Society, 124, 6378-6382.
- GUERRERO, S., *et al.* 2015a. Electrochemical immunosensor for sensitive determination of the anorexigen peptide YY at grafted reduced graphene oxide electrode platforms. *Analyst*, 140, 7527-7533.
- GUERRERO, S., et al. 2015b. Electrochemical immunosensor for sensitive determination of the anorexigen peptide YY at grafted reduced graphene oxide electrode platforms. *Analyst*, 140, 7527-7533.
- HAMMOND, J. L., et al. 2016. Electrochemical biosensors and nanobiosensors. Essays in biochemistry, 60, 69-80.
- HAN, H.-S., et al. 2015. Quantum dot/antibody conjugates for in vivo cytometric imaging in mice. Proceedings of the National Academy of Sciences, 112, 1350-1355.
- HANSEN, J. A., et al. 2006. Quantum-dot/aptamer-based ultrasensitive multianalyte electrochemical biosensor. *Journal of the American Chemical Society*, 128, 2228-2229.
- HNAIEN, M., et al. 2008. Immobilization of specific antibody on SAM functionalized gold electrode for rabies virus detection by electrochemical impedance spectroscopy. *Biochemical Engineering Journal*, 39, 443-449.
- HONG, H., et al. 2012. Electroluminescence Biosensor for DNA Determination Based on CdTe Quantum Dots Double-Tagging Fe_3O_4@ Au Core/Shell Magnetic Nanoparticles [J]. Chinese Journal of Analytical Chemistry, 6, 006.
- JIN, T., et al. 2010. Antibody–ProteinA conjugated quantum dots for multiplexed imaging of surface receptors in living cells. *Molecular BioSystems*, 6, 2325-2331.
- JUNG, Y., et al. 2008. Recent advances in immobilization methods of antibodies on solid supports. *Analyst*, 133, 697-701.
- KERMAN, K., *et al.* 2007. Quantum dot-based immunosensor for the detection of prostate-specific antigen using fluorescence microscopy. *Talanta*, 71, 1494-1499.

- KOKKINOS, C., *et al.* 2011. Disposable Nafion-modified micro-fabricated bismuth-film sensors for voltammetric stripping analysis of trace metals in the presence of surfactants. *Talanta*, 84, 696-701.
- KOKKINOS, C., et al. 2016. Electrochemical immunosensors: Critical survey of different architectures and transduction strategies. *TrAC Trends in Analytical Chemistry*, 79, 88-105.
- KOKKINOS, C., et al. 2015. Quantum dot-based electrochemical DNA biosensor using a screen-printed graphite surface with embedded bismuth precursor. *Electrochemistry Communications*, 60, 47-51.
- KOTAGIRI, N., et al. 2014. Antibody quantum dot conjugates developed via copper-free click chemistry for rapid analysis of biological samples using a microfluidic microsphere array system. *Bioconjugate chemistry*, 25, 1272-1281.
- LI, H., *et al.* 2011. Electrochemical immunosensors for cancer biomarker with signal amplification based on ferrocene functionalized iron oxide nanoparticles. *Biosensors and Bioelectronics*, 26, 3590-3595.
- LI, Y., et al. 2016a. An ultrasensitive sandwich-type electrochemical immunosensor based on δ-MnO 2 and palladium nanoparticles covered natural halloysite nanotubes for the detection of hepatitis B surface antigen. New Journal of Chemistry, 40, 558-563.
- LI, Y., et al. 2016b. A sandwich-type electrochemical immunosensor based on the biotin-streptavidin-biotin structure for immunoglobulin G. Scientific reports, 6.
- LITVIN, A. P., et al. 2017. Photoluminescence of lead sulfide quantum dots of different sizes in a nanoporous silicate glass matrix. *The Journal of Physical Chemistry C*, 121, 8645-8652.
- LIU, Y., et al. 2013. A quantum dots-based electrochemical assay towards the sensitive detection of tumor cells. *Electrochemistry Communications*, 33, 59-62.
- LOS ALAMOS NATIONAL LABORATORY. 2011. XPS works [Online]. Available:https://www.lanl.gov/orgs/nmt/nmtdo/AQarchive/04summer/ XPS.html [Accessed 1 May 2017].
- LU, G. W., *et al.* 2010. Emulsions and microemulsions for topical and transdermal drug delivery. *Handbook of Non-Invasive Drug Delivery Systems.* Elsevier.
- LUPPA, P. B., et al. 2001. Immunosensors—principles and applications to clinical chemistry. *Clinica Chimica Acta*, 314, 1-26.

- MARQUES, R. C., et al. 2014. Electrochemical immunosensor for the analysis of the breast cancer biomarker HER2 ECD. *Talanta*, 129, 594-599.
- MÁRQUEZ, A. M., et al. 2017. Structural and electronic properties of lead sulfide quantum dots from screened hybrid density functional calculations including spin–orbit coupling effects. *Theoretical Chemistry Accounts*, 136, 58.
- MARTINS, G. V., et al. 2017. Based Sensing Device for Electrochemical Detection of Oxidative Stress Biomarker 8-Hydroxy-2'deoxyguanosine (8-OHdG) in Point-of-Care. Scientific Reports, 7, 14558.
- MAZUMDER, S., et al. 2009. Review: biofunctionalized quantum dots in biology and medicine. Journal of Nanomaterials, 2009, 38.
- MEHRABANI, S., et al. 2014. Hybrid integrated label-free chemical and biological sensors. Sensors, 14, 5890-5928.
- MICHIGAN STATE UNIVERSITY. Visible and Ultraviolet Spectroscopy [Online]. Available:https://www2.chemistry.msu.edu/faculty/reusch/virttxtjml/spe ctrpy/uv-vis/spectrum.htm [Accessed 1 May 2017].
- MOINA, C., et al. 2012a. Fundamentals and applications of immunosensors. Advances in immunoassay technology, 65-80.
- MOINA, C., et al. 2012b. Fundamentals and applications of immunosensors. Advances in immunoassay technology. InTech.
- MONTALBETTI, C. A., et al. 2005. Amide bond formation and peptide coupling. *Tetrahedron*, 61, 10827-10852.
- MONTOYA, A., et al. 2009. Fundamentals of piezoelectric immunosensors. Piezoelectric transducers and applications. Springer.
- MOREIRA, F. T., *et al.* 2016. Screen-printed electrode produced by printedcircuit board technology. Application to cancer biomarker detection by means of plastic antibody as sensing material. *Sensors and Actuators B: Chemical*, 223, 927-935.
- MORGAN, C. L., et al. 1996. Immunosensors: technology and opportunities in laboratory medicine. *Clinical chemistry*, 42, 193-209.
- NEJO, A. O., et al. 2012. Facile synthesis of organically capped PbS nanoparticles. Journal of Alloys and Compounds, 537, 19-23.
- NIU, X., et al. 2017. Simple anodization of home-made screen-printed carbon electrodes makes significant activity enhancement for hydrogen

evolution: the synergistic effect of surface functional groups, defect sites, and hydrophilicity. *Electrochimica Acta*, 235, 64-71.

- NOOR AINI, B., et al. 2016. Development of formaldehyde biosensor for determination of formalin in fish samples; malabar red snapper (Lutjanus malabaricus) and longtail tuna (Thunnus tonggol). *Biosensors*, 6, 32.
- PAL, C., et al. 2017. Charge transport in lead sulfide quantum dots/phthalocyanines hybrid nanocomposites. *Organic Electronics*, 44, 132-143.
- PAN, Y., et al. 2017. A Novel Quantum Dot Fluorescence Immunosensor Based on Magnetic Beads and Portable Flow Cytometry for Detection of Okadaic Acid. *Procedia Technology*, 27, 214-216.
- PANDIT, B., et al. 2017. Large scale flexible solid state symmetric supercapacitor through inexpensive solution processed V2O5 complex surface architecture. *Electrochimica Acta*, 242, 382-389.
- PATEL, J. D., et al. 2012. Room temperature synthesis of aminocaproic acidcapped lead sulphide nanoparticles. *Materials Sciences and Applications*, 3, 125.
- PATHAK, Y., et al. 2012. Antibody-mediated Drug Delivery Systems: Concepts, Technology, and Applications, John Wiley & Sons.
- PATRIS, S., et al. 2014. Nanoimmunoassay onto a screen printed electrode for HER2 breast cancer biomarker determination. *Talanta*, 130, 164-170.
- PAULINO, A. T., et al. 2008. Square wave voltammetry in the determination of Ni2+ and Al3+ in biological sample. *Analytical Sciences*, 24, 1443-1447.
- PEI, X., et al. 2013. Sandwich-type immunosensors and immunoassays exploiting nanostructure labels: A review. *Analytica chimica acta*, 758, 1-18.
- PHYSICAL ELECTRONICS. 2018. XPS / ESCA [Online]. Available: https://www.phi.com/surface-analysis-techniques/xps-esca.html [Accessed 1 May 2017`].
- QIAN, J., et al. 2011. Simultaneous detection of dual proteins using quantum dots coated silica nanoparticles as labels. *Biosensors and Bioelectronics*, 28, 314-319.
- REKEMEYER, P. H., et al. 2017. Minority Carrier Transport in Lead Sulfide Quantum Dot Photovoltaics. *Nano letters*, 17, 6221-6227.

- REVERBERI, R., et al. 2007. Factors affecting the antigen-antibody reaction. Blood transfusion, 5, 227.
- RICCI, F., et al. 2012. A review of experimental aspects of electrochemical immunosensors. *Electrochimica Acta*, 84, 74-83.
- SALIH, F. E., *et al.* 2017. Electrochemical detection of lead (II) at bismuth/poly (1, 8-diaminonaphthalene) modified carbon paste electrode. *Arabian Journal of Chemistry*, 10, 596-603.
- SOCIETY FOR ENDOCRINOLOGY. *Peptide* YY [Online]. Available: <u>www.yourhormones.info/hormones/peptide_yy.aspx</u> [Accessed 16 June 2017].
- SOUNDERYA, N., *et al.* Upconversion nanoparticles for imaging cells. 13th international conference on biomedical engineering, 2009. Springer, 1741-1744.
- TIAN, L., et al. 2016. Ultrasensitive sandwich-type electrochemical immunosensor based on trimetallic nanocomposite signal amplification strategy for the ultrasensitive detection of CEA. Scientific Reports, 6.
- VO, N., et al. 2015. Conjugation of E. coli O157: H7 antibody to CdSe/ZnS quantum dots. Journal of Nanomaterials, 2015, 8.
- WAGNER, M. K., *et al.* 2010. Use of quantum dots in the development of assays for cancer biomarkers. *Analytical and bioanalytical chemistry*, 397, 3213-3224.
- WANG, J., et al. 1996. Electrochemical activation of screen-printed carbon strips. *Analyst*, 121, 345-350.
- WANG, S., *et al.* 2008. Electrochemical immunoassay of carcinoembryonic antigen based on a lead sulfide nanoparticle label. *Nanotechnology*, 19, 435501.
- WANG, Y., et al. 2015. Electrochemical immunosensor using nanoparticlebased signal enhancement for Escherichia coli O157: H7 detection. *IEEE Sensors Journal*, 15, 4692-4699.
- WARWICK. 2010. Transmission Electron Microscopy (TEM) [Online]. Available: https://warwick.ac.uk/fac/sci/physics/current/postgraduate/regs/mpags warwick/ex5/techniques/structural/tem/ [Accessed 25 Sept 2018].
- WEI, H., et al. 2007. Enhanced electrochemical performance at screenprinted carbon electrodes by a new pretreating procedure. *Analytica chimica acta*, 588, 297-303.

- WEI, Y., *et al.* 2016. Sandwich-type electrochemical immunosensor for the detection of AFP based on Pd octahedral and APTES-M-CeO 2-GS as signal labels. *Biosensors and Bioelectronics*, 79, 482-487.
- WORLD HEALTH ORGANIZATION (WHO). 2018. Available: https://monographs.iarc.fr/agents-classified-by-the-iarc/ [Accessed 24 Sept 2018].
- WU, D., et al. 2016. Label-free Electrochemiluminescent Immunosensor for Detection of Prostate Specific Antigen based on Aminated Graphene Quantum Dots and Carboxyl Graphene Quantum Dots. Scientific reports, 6.
- WU, Q. S., et al. 2006. Synthesis and optical properties of the semiconductor lead sulfide nanobelts. Bulletin of the Korean Chemical Society, 27, 377-380.
- WU, Y., et al. 2013. Highly specific and ultrasensitive graphene-enhanced electrochemical detection of low-abundance tumor cells using silica nanoparticles coated with antibody-conjugated quantum dots. *Analytical chemistry*, 85, 3166-3173.
- XUAN, X., et al. 2016. A fully integrated and miniaturized heavy-metaldetection sensor based on micro-patterned reduced graphene oxide. *Scientific reports*, 6, 33125.
- YAN, P., et al. 2014. Ultrasensitive detection of clenbuterol by quantum dots based electrochemiluminescent immunosensor using gold nanoparticles as substrate and electron transport accelerator. Sensors and Actuators B: Chemical, 191, 508-515.
- YANG, M., et al. 2011. Sensitive electrochemical immunosensor for the detection of cancer biomarker using quantum dot functionalized graphene sheets as labels. Sensors and Actuators B: Chemical, 155, 357-360.
- ZHANG, P., *et al.* 2014. Simple and sensitive detection of HBsAg by using a quantum dots nanobeads based dot-blot immunoassay. *Theranostics*, 4, 307.
- ZHAO, X., et al. 2005. Synthesis and optical properties of thiol-stabilized PbS nanocrystals. *Langmuir*, 21, 1086-1090.
- ZHOU, X., *et al.* 2014. Facile synthesis and electrochemical properties of two dimensional layered MoS2/graphene composite for reversible lithium storage. *Journal of Power Sources*, 251, 264-268.
- ZHU, N., *et al.* 2004a. Lead sulfide nanoparticle as oligonucleotides labels for electrochemical stripping detection of DNA hybridization. *Electroanalysis*, 16, 577-582.

ZHU, N., *et al.* 2004b. Lead sulfide nanoparticle as oligonucleotides labels for electrochemical stripping detection of DNA hybridization. *Electroanalysis: An International Journal Devoted to Fundamental and Practical Aspects of Electroanalysis,* 16, 577-582.

