

XYLANASE PRODUCTION BY Penicillium oxalicum T3.3 USING RICE STRAW AS SUBSTRATE FOR BIOBLEACHING OF BAMBOO PULPS

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FBSB 2017 47



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By NUR IDAYU BINTI ZAHARI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Master of Science

May 2017

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Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in the fulfilment of the requirement for the degree of Master of Science

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Chairman: Associate Professor Umi Kalsom bt. Md Shah, PhDFaculty: Biotechnology and Biomolecular Sciences

Recently, xylanase has raised high attention in pulp and paper industry due to their bleach-boosting properties, which assist bleaching process. Xylanase can improve the performance of pulp. It is a hydrolytic enzyme that can degrade the linear polysaccharide beta-1,4-xylan into xylose. Thus, the aims of this research are to select potential xylanase-producing fungi with enhance xylanase production and to evaluate the performance of the xylanase in assisting the biobleaching process of bamboo pulps. Four fungi species namely *Penicillium oxalicum* T3.3, *Aspergillus niger* ATCC 6275, Colletotrichum gliosporoides and Pycnoporus sanguineus were screened for xylanase production based on the observation of clear zone formation on Malt Extract Agar (MEA) containing xylan. P. oxalicum T3.3 and A. niger ATCC 6275 showed larger clear zone formation on the agar plate. These fungi were grown in shake flask by submerged fermentation containing rice straw as substrate. P. oxalicum T3.3 showed highest xylanase activity (65.89 U/mL) with lowest carboxymethyl cellulase (CMCase) (1.88 U/mL) and filter paperase (FPase) activity (0.16 U/mL) after 4 days fermentation at 30°C. The cultural conditions for xylanase production by P. oxalicum T3.3 was investigated under various concentration of rice straw, initial pH, temperature, agitation speed, nitrogen sources and additional of surfactants. The best cultural conditions was determined by one-factor-at-a-time method. By optimizing rice straw concentration at 1% (w/v), initial pH at 6, temperature 35°C, agitation speed at 150 rpm, using yeast extract as the nitrogen source, and addition of Tween 80 as surfactant, the highest xylanase production of 79.12 U/mL was obtained. That is 20% improvement compared to that before optimization. The xylanase was found more stable at pH range of 4.0 to 7.0 and temperature from 45°C to 55°C. The enzyme retained 72% and 51% of its activity after incubation for 2 h and 4 h, respectively at 50°C and pH 6. At -80°C, xylanase was still above its half life for 2 weeks. Biobleaching of bamboo pulps with xylanase was most effective at an enzyme dose of 5 U/g oven dried pulp at pH 7 and incubated in water bath at 50° C. Biobleaching process increased paper brightness by

7.51 points at optimum conditions. Results of chemical composition of biobleached bamboo pulps revealed that lignin content reduced by 25% with only slightly reduction of holocellulose and α -cellulose content by 0.5% and 12% compared to chemically bleached bamboo pulp. Thus, cellulase-poor xylanase produced from *P. oxalicum* T3.3 grown on rice straw has high potential for biobleaching application in pulp and paper industry in terms of technical and biological performance and economical aspects.



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Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGHASILAN XYLANASE OLEH *Penicillium oxalicum* T3.3 MENGGUNAKAN JERAMI PADI SEBAGAI SUBSTRAT UNTUK AGEN BIOPELUNTURAN PULPA BULUH

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Baru-baru ini, xilanase menarik perhatian industri pulpa dan kertas disebabkan oleh ciriciri meransang pelunturan, yang mana boleh membantu proses meluntur. Xilanase boleh meningkatkan prestasi pulpa. Ianya adalah enzim hidrolitik yang mana boleh menguraikan polisakarida linear beta-1,4-xylan kepada xylosa. Oleh itu, tujuan kajian ini adalah untuk memilih kulat berpotensi sebagai penghasil xilanase, menentukan keadaan kultur terbaik yang meningkatkan penghasilan xilanase dan untuk menilai prestasi xilanase dalam membantu biopelunturan pulpa buluh. Empat spesis kulat iaitu Penicillium oxalicum T3.3, Aspergillus niger ATCC 6275, Colletotrichum gliosporoides dan Pycnoporus sanguineus telah disaring untuk penghasilan xilanase berdasarkan pemerhatian pembentukan zon jelas di atas Agar Ekstrak Malt (MEA) yang mengandungi xylan. P. oxalicum T3.3 and A. niger ATCC 6275 menunjukkan pembentukan zon jelas lebih besar di atas agar. Kulat ini dihidupkan dalam kultur cecair mengandungi jerami padi sebagai substrat. P. oxalicum T3.3 menunjukkan aktiviti xilanase paling tinggi (65.89 U/mL) dengan paling rendah carboxymethyl selulase with (CMCase) (1.88 U/mL) dan filter paperase (FPase) activity (0.16 U/mL) selepas 4 hari fermentasi pada 30°C. Keadaan kultur penghasilan xilanase oleh P. oxalicum T3.3 dikaji di bawah pelbagai kepekatan jerami padi, pH awal, suhu, kelajuan goncangan, sumber nitrogen dan penambahan surfaktan. Penentuan keadaan kultur dilakukan dengan kaedah satu faktor pada satu masa. Dengan mengoptimumkan kepekatan substrat pada 1% dan pH awal pada 6, suhu 35°C, kelajuan gonjangan pada 150 rpm, ekstrak yis sebagai sumber nitrogen, dan penambahan Tween 80 sebagai surfaktan, xilanase yang tinggi dihasilkan sebanyak 79.12 U/mL. Oleh itu, 20% penambahan berbanding sebelum pengoptimuman. Xilanase dicirikan dan didapati lebih stabil pada julat pH 4.0 hingga 7.0 dan suhu dari 45°C hingga 55°C. Enzim boleh mengekalkan 72% dan 51% aktivitinya selepas 2 dan 4 jam inkubasi pada 50°C dan pH 6. Pada suhu -80°C, xilanase masih di atas separuh hayat untuk 2 minggu. Biopelunturan pulpa buluh dengan xylanase adalah paling berkesan pada dos enzim 5 U/g ketuhar kering pulpa, pada pH 7 and inkubasi dalam air pada 50°C.

Di bawah keadaan yang optimum, proses biopelunturan meningkatkan kecerahan kertas sebanyak 7.51 mata. Keputusan komposisi kimia pulpa buluh biopelunturan menunjukkan kandungan lignin berkurang sebanyak 25% dengan hanya sedikit pengurangan kandungan holoselulosa dan alpha-selulosa sebanyak 0.5% dan 12% berbanding pulpa pelunturan. Oleh itu xilanase rendah selulase dihasilkan oleh *P. oxalicum* T3.3 ditumbuhkankan atas jerami padi mempunyai potensi besar dalam aplikasi biopelunturan dalam perusahaan pulpa dan kertas dari segi teknikal dan prestasi biologikal serta aspek ekonomi.



ACKNOWLEDGEMENTS

Foremost, I am grateful and would like to express my sincere gratitude to my supervisor, Assoc. Prof. Dr. Umi Kalsom binti Md Shah for her invaluable guidance, continuous encouragement and constant support of my Master research. I really appreciate her guidance from the initial until to the final level that enabled me to develop an understanding of this research thoroughly. Without her advice and assistance, it would be a lot tough to complete. I also sincerely thank for her time spending proofreading and correcting.

I would like to express my gratitude to my supervisory committe members, Dr. Ainun Zuriyati Mohamed @ Asa'ari and Assoc. Prof. Dr. Rosfarizan Mohamad, for their help and guidance in completing the study.

My sincere thanks to my fellow labmates, Husna and Fairuzana for stimulating fruitful discussion, support me during difficult condition and for the fun we had together. I also thank all master students, lecturers and members of staff of Biotechnology Lab 1.6, and Biotechnology 1, who always help, assist and guide in order to ensure the research went smoothly and successfully.

I acknowledge my sincere indebtedness and gratitude to my parents and sibling for their love, dream and sacrifice throughout my life. I am really thankful for their patience, and understanding that were inevitable to make this work possible. I cannot find appropriate words to express my appreciation for their devotion, support and faith in during the journey to achieve my dreams.

Last but not least, i would like to thank everyone whom contributes to my research directly or indirectly. Their comments and suggestions are valuable in the completion of this study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

	BSA	Bovine Serum Albumin
	CAZY	Carbohydrate-Active Enzyme
	CE	Carbohydrate Esterase
	CMC	Carboxymethycellulose
	CMCase	Carboxymethycellulase
	CTMP	Chemi-Thermo-Mechanical Pulping
	DNS	Dinitrosalicylic Acid
	E.C.	European Commission
	FOA	Food and Agriculture Organization
	FPase	Filter Paperase
	GH	Glycoside Hydrolysis
	ISO	International Organization for Standardization
	LSD	Least Significant difference
	MADA	Muda Agriculture Development Authority
	MARDI	Malaysian Agricultural Research and Development
		Institute
	MEA	Malt extract Agar
	NAG	N-acetylglucosamine
	ORF	Open Reading Frame
	PDA	Potato Dextrose Agar
	PGW	Pressure Groundwood Pulping
	pI	Isoelectric Point
	RSM	Response Surface Methodology
	SAS	Statistical Analysis Software
	SD	Standard Deviation
	TAPPI	Technical Association of the Pulp and Paper Industry
	TIM-barrel	Triosephosphate Isomerase-barrel
(\mathbf{O})	TMP	Thermo-mechanical Pulping
	UPM	Universiti Putra Malaysia
	USDA	United States Department of Agriculture

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LIST OF SYMBOLS

%	Percentage
:	Ratio
<	Less than
>	More than
°C	Degree celcius
µg/mL	Microgram/mililiter
μL	Microliter
C ₆ H ₅ O ₇ Na ₃ .2H ₂ O	Sodium citrate
cm	Centimeter
CO ₂	Carbon dioxide
CoCl ₂	Cobalt chloride
FeSO ₄ .7H ₂ 0	Ferrous sulphate heptahydrate
g	Gram
g	Gravity
h	Hour
H ₂ O ₂	Hydrogen peroxide
HCL	Hydrochloric acid
IU/g	Unit/gram
IU/mL	Unit/mililiter
kDa	Kilodalton
L	Liter
М	Molar
Mg/mL	Miligram/mililiter
mL	Mililiter
mM	Milimolar
mM	Milimolar
MnSO ₄ .H ₂ O	Zinc sulphate heptahydrate
Ν	Normality
Na ₂ HPO ₄ .7H ₂ O	Dibasic sodium phosphate
NaCl	Sodium chloride

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NaClO ₂	Sodium chlorite
NaOH	Sodium hydroxide
Nm	Nanometer
Rpm	Revolution per minute
U/o.d.	Unit/oven dry
v/v	Volume/volume
w/v	Weight/volume
β	Beta

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CHAPTER 1

INTRODUCTION

Nowadays the applications of xylanase have been diversified from textile to the pulp and paper industry (Dedhia et al., 2014). In the pulp and paper industry, large amounts of chlorine-based chemicals was used during bleaching process which are toxic, mutagenic and persistent. The chlorinated organic compound and absorble organic halides are produced when chlorine reacted with lignin, thus contributed as the major sources of environmental pollution (Yeasmin et al., 2011). Owing to this problem, public awareness has increased which forced the government to put stringent regulation concerning pulp and paper effluents (Dedhia et al., 2014). Hence, an environmental friendly technology is a need in the paper industry. In 1986, biobleaching which is basic idea of enzymeaided bleaching was published. The usage of xylanase to treat pulp prior to bleach enhances the bleaching process by degrading the xylan present in the wood pulp and ease the release of lignin (Sharma et al., 2014). According to Boruah et al., (2015), pretreatment by using xylanase from *P.meleagrinum* can reduced lignin content, small amount of cellulose and increased the brightness of bamboo pulp. There are several conditions to ensure the successful of biobleaching, which is xylanase enzyme must be able to withstand high temperature and alkaline conditions. Besides, xylanase is preferably contain low or free-cellulase because high cellulase resulted in the lost of cellulose, degrade the quality of pulp and increase the cost of effluent treatment (Bhakyaraj, 2014).

Xylanase is an extracellular enzyme which is the main xylanolytic enzyme in degradation of β -1, 4 backbone of xylan in hemicellulose component of plant cell wall (Hatanaka, 2012). Degradation of xylan produced small fragments such as mono-, diand trisaccharides of β -D-xylopyranosyl that are used by microorganism as main carbon source (Kanimozhi & Nagalakshmi, 2014). Different types of miroorganisms can produced xylanase enzyme such as bacteria, actinomyces and fungi except the mammals (Chandra et al., 2012). However, the microbial xylanase was preferred in xylanase production compared to other producers, because they meet industrial application (Goswami & Rawat, 2015). This is because, microbial sources are easy to handle for large production, has short generation time, are structurally stable, easy genetic manipulation and their availability (Sindhu et al., 2011). Due to high production level and extracellular secretion, filamentous fungi usually were used in industrial application (Selvarajan & Veena, 2017). There are many fungi producing xylanase such as Aspergillus, Trichoderma, and Penicillium (Burlacu et al., 2016). Thus, in this study Penicillium oxalicum T3.3, Aspergillus niger ATCC 6275, Colletotrichum gliosporoides, and Pcynoporus sanguineus were evaluated for xylanase production for application in biobleaching of bamboo pulp.

Due to inducible characteristic, xylanase requires inducer for maximum production of xylanase activity. In submerged fermentation, xylanase was induced by various sources of xylan (Pandey et al., 2014). Thus, when fungi are grown in medium containing xylan as carbon source, the production of xylanase also increased. The sources of xylan can be rice straw, beechwood xylan, and sugarcane bagasse. The level of xylanase production is different depend on the sources of xylanase (Motta et al., 2013). In Malaysia, 2.7 million tonnes of rice was produced and the ratio to rice straw is 1:1 (FOA 2015; MADA 2010). Besides the rice straw, Malaysia also produced a lot of oil palm trunk. The trunk's rich cellulose and hemicellulose content are promising feed stock for fermenting sugars (Ang et al., 2013). The rice straw were used as the source of the xylan in xylanase production in order to reduce the environment pollution and production cost.

From our finding, there are few research on the best cultural conditions for xylanase production by *Penicillium oxalicum* using submerged fermentation. Thus a study on the effect of cultural conditions such as concentration of rice straw, initial pH, temperature, agitation speed, nitrogen sources and additional of surfactants are important consideration to determine the best cultural condition for xylanase production. The most important aspects to reduce the production cost are optimization of media and process condition. The conventional methods of optimization included changing one parameter and fixing the others parameter at certain level (Dedhia et al., 2014). Optimization of *one-factor-at-a-time* techniques were used in submerged fermentation to produce high xylanase activity.

Xylanase is widespread in the market, but most widely available xylanases cannot withstand with the harsh conditions during industrial operation. In biobleaching technology, although xylanase from bacteria have higher values in this application compared to fungi, but they have low level enzyme activity. Thus, fungi are more preferable as producer when the crude enzyme were applied in application due to high level enzyme production (Selvarajan & Veena, 2017). Therefore, the aim of this research is to identify the potential of cellulase poor xylanase production by selected fungi using as rice straw, as an economical substrate to reduce the production cost and to evaluate the effectiveness of crude xylanase in assisting biobleaching process of pulp.

1.1 Objectives

The specific objectives of this study were as follows:

- 1. To select the potential fungal isolates from laboratory culture collection for xylanase production.
- 2. To optimize the cultural conditions for xylanase production by the selected fungus.
- 3. To evaluate the effectiveness of crude xylanase in assisting biobleaching process.

REFERENCES

- Abdel-Sater, M. A., & El-Said, A. H. M. (2001). Xylan-decomposing fungi and xylanolytic activity in agricultural and industrial wastes. *International Biodeterioration and Biodegradation*, 47(1), 15–21. doi:10.1016/S0964-8305(00)00113-X
- Adesina, F. C., & Onilude, A. A. (2013). Isolation, identification and screening of xylanase and glucanase-producing microfungi from degrading wood in Nigeria. *African Journal of Agricultural Research*, 8(34), 4414–4421. doi:10.5897/AJAR2013.6993
- Ahuja, S. K., Ferreira, G. M., & Moreira, A. R. (2004). Utilization of enzymes for environmental applications. *Critical Reviews in Biotechnology*, 24(2-3), 125– 154. doi:10.1080/07388550490493726
- Akpinar, O., Ak, O., Kavas, A., Bakir, U., & Yilmaz, L. (2007). Enzymatic production of xylooligosaccharides from cotton stalks. *Journal of Agricultural and Food Chemistry*, 55(14), 5544–5551. doi:10.1021/jf063580d
- Amid, M., Manap, M. Y., Hussin, M., & Mustafa, S. (2015). A novel aqueous two phase system composed of surfactant and xylitol for the purification of lipase from pumpkin (Cucurbita moschata) seeds and recycling of phase components. *Molecules*, 20(6), 11184–11201. doi:10.3390/molecules200611184
- Anthony, T., Raj, K. C., Rajendra, A., Gunasekaran, P. (2003). High molecular weight cellulase-free xylanases from alkali-tolerant *Aspergillus fumigatus* AR1. *Enzyme and Microbial Technology*, 32, 647-654
- Anusha, M., Sushma, T., & Sarkar, S. (2013). Isolation, identification and screening of xylanase producing fungi from apple (Pyrus malus L.). *Annals of Plant Sciences*. Retrieved from http://ebioscholar.com/ojs/index.php/ap/article/view/329
- Anwar, Z., Gulfraz, M., & Irshad, M. (2014). Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy: A brief review. *Journal of Radiation Research and Applied Sciences*, 7(2), 163–173. doi:10.1016/j.jrras.2014.02.003
- Ashori, A. (2004). Development of high quality printing paper using kenaf (*Hibiscus cannabinus*) fibers. PhD Thesis.
- Ateş, S., Deniz, İ., Kirci, H., Atik, C., & Okan, O. T. (2015). Comparison of pulping and bleaching behaviors of some agricultural residues. *Turkish Journal of Agriculture and Forestry*, 39, 144–153. doi:10.3906/tar-1403-41
- Azmy, H. M., & Abd. Razak, O. (2000). Availability of non-wood resources. Part 1.
 Bamboo as an alternative to rubberwood. In Ahmad Shakri, M. S., Ho, K. S.,
 & Mohd. Dahlan, J. (ed.), *Proceedings of the National Seminar on Alternatives*

to Rubberwood (pp. 55-61). 26 September 2000, Forest Research Institute of Malaysia, Kepong, Malaysia.

- Bajaj, B. K., Sharma, M., & Sharma, S. (2011). Alkalistable endo-β-1,4-xylanase production from a newly isolated alkalitolerant *Penicillium* sp. SS1 using agroresidues. *3 Biotech*. doi:10.1007/s13205-011-0009-5
- Bakri, Y., Al-Jazairi, M., & Al-Kayat, G. (2008). Xylanase production by a newly isolated Aspergillus niger SS7 in submerged culture. Polish Journal of Microbiology / Polskie Towarzystwo Mikrobiologów = The Polish Society of Microbiologists, 57(3), 249–51. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/19004246
- Bakri, Y., Masson, M., & Thonart, P. (2010). Isolation and identification of two new fungal strains for xylanase production. *Applied Biochemistry and Biotechnology*, 162(6), 1626–34. doi:10.1007/s12010-010-8944-x
- Bansal, N., Raman, S., Janvega, C., & Soni, S. K. (2012). Production of xylanasecellulase complex *Bacillus subtilis* NS7 for the biodegradation of agrowaste residues, *Lignocellulose*, 1(3), 196-209.
- Basar, B. (2011). Improvement of xylanase production by recombinant Escherichia coli DH5a using fed-batch fermentation. Master Thesis.
- Batalha, L. A. R., Colodette, J. L., Gomide, J. L., Barbosa, L. C. A., Maltha, C. R. A., & Gomes, F. J. B. (2012). Dissolving pulp production from bamboo. *BioResources*, 7(1), 640–651.
- Bhakyaraj, R. (2014). Isolation, Production and Characterization of Xylanase from Bacillus sp. isolated form soil samples. *International Journal of Advanced Multidisciplinary Research*, 1(1), 41–51.
- Biermann, C. J. (1996). *Pulping fundamentals. Handbook of Pulping and Papermaking*. San Diego:Academic Press.
- Boruah, P., Dowarah, P., Hazarika, R., Yadav, A., Barkakati, P., & Goswami, T. (2015). Xylanase from Penicillium meleagrinum var. viridiflavum - a potential source for bamboo pulp bleaching. *Journal of Cleaner Production*, *116*, 259–267. doi:10.1016/j.jclepro.2015.12.024
- Chandra, S., Sharma, R., Jasuja, N. D., Saxena, R., & Rana, S. (2012). Xylanase assay of fungal isolates from semi-arid Soil of India. *Indian Journal of Fundamental* and Applied Life Sciences, 2(4), 7–12.
- Cho, Y. J., Hwang, H. J., Kim, S. W., Song, C. H., & Yun, J. W. (2002). Effect of carbon source and aeration rate on broth rheology and fungal morphology during red pigment production by *Paecilomyces sinclairii* in a batch bioreactor. *Journal* of Biotechnology, 95(1), 13–23. doi:10.1016/S0168-1656(01)00445-X

- Collins, T., Gerday, C., & Feller, G. (2005). Xylanases, xylanase families and extremophilic xylanases. *FEMS Microbiology Reviews*, 29(1), 3–23. doi:10.1016/j.femsre.2004.06.005
- Cui, F., & Zhao, L. (2012). Optimization of xylanase production from *Penicillium* sp.WX-Z1 by a two-step statistical strategy: Plackett-Burman and Box-Behnken experimental design. *International Journal of Molecular Sciences*, 13(8), 10630–10646. doi:10.3390/ijms130810630
- Dedhia, B. S., Vetal, M. D., Rathod, V. K., & Levente, C. (2014). Xylanase and laccase aided bio-bleaching of wheat straw pulp. *Canadian Journal of Chemical Engineering*, 92(1), 131–138. doi:10.1002/cjce.21798
- Doherty, W. O. S., Mousavioun, P., & Fellows, C. M. (2011). Value-adding to cellulosic ethanol: Lignin polymers. *Industrial Crops and Products*, 33(2), 259–276. doi:10.1016/j.indcrop.2010.10.022
- Dong, W. K., Tae, S. K., Young, K. J., & Jae, K. L. (1992). Adsoption kinetic and behaviors of celluloase components on microcrystalline cellulose. *Journal of Fermentation and Bioengineering*, 73, 461-469
- Farrell, R. L., Hata, K., & Wall, M. B. (1997). Solving pitch problems in pulp and paper processes by the use of enzymes or fungi. In *Advances in Biochemical Engineering Biotechnology* (Vol. 57, pp. 197–212). doi:10.1007/BFb0102075
- Food and Agriculture Organization (FAO) (2014). Available from: http://faostat.fao.org. (accessed October 1, 2014).
- Food and Agriculture Organization (FAO) (2015). Available from:<u>http://faostat.fao.org</u>. (accessed February 21, 2016).
- Gullichsen, J. (2000). *Fiber line operations*. In: Gullichsen J, & Fogelholm, C. J. (ed.), Chemical pulping –papermaking science and technology (p A19 (Book 6A)). Helsinki: Fapet Oy.
- Haki, G. D., & Rakshit, S. K. (2003). Developments in industrially important thermostable enzymes: A review. *Bioresource Technology*, 89(1), 17–34. doi:10.1016/S0960-8524(03)00033-6
- Hatanaka, K. (2012). Incorporation of fluorous glycosides to cell membrane and saccharide chain elongation by cellular enzymes. *Top Current Chemistry*, 308, 291–306. doi:10.1007/128
- Helle, S. S., Duff, S. J. B., & Cooper, D. G. (1993). Effect of surfactants on cellulose hydrolysis. *Biotechnology and Bioengineering*, 42(1993), 611–617.
- Hori, C. (2012). Effects of xylan on production of cellulolytic enzymes by the basidiomycete Phanerochaete chrysosporium. <u>http://www.worthington-biochem.com/introbiochem/inhibitors.html</u>, 08 July 2017, 11: 14 am.

- Iqbal, H. M. N., Ahmed, I., Zia, M. A., & Irfan, M. (2011). Purification and characterization of the kinetic parameters of cellulase produced from wheat straw by Trichoderma viride under SSF and its detergent compatibility. *Advances in Bioscience and Biotechnology*, 2(0), 149–156. doi:10.4236/abb.2011.23024
- Irfan, M., Nadeem, M., Syed, Q. A., & Baig, S. (2010). Submerged cultivation of *Aspergillus niger* on pretreated sugarcane bagasse. *World Journal of Agricultural Sciences*, 6(4), 466-472.
- Irshad, M., Asgher, M., Sheikh, M. A., & Nawaz, H. (2011). Purification and characterization of laccase produced by *Schyzophylum commune* IBL-06. in solid state culture of banana stalks. *BioResources*, 6(3), 2861–2873.
- Jeya, M., Nguyen, N. P. T., Moon, H. J., Kim, S. H., & Lee, J. K. (2010). Conversion of woody biomass into fermentable sugars by cellulase from *Agaricus arvensis*. *Bioresource Technology*, 101(22), 8742–8749. doi:10.1016/j.biortech.2010.06.055
- Kanimozhi, K., & Nagalakshmi, P. K. (2014). Original research article xylanase production from *Aspergillus niger* by solid state fermentation using agricultural waste as substrate. *International Journal of Current Microbiology and Applied Sciences*, 3(3), 437–446.
- Karim, A., Nawaz, M. A., Aman, A., & Ul Qader, S. A. (2014). Hyper production of cellulose degrading endo (1,4) β-d-glucanase from *Bacillus licheniformis* KIBGE-IB2. *Journal of Radiation Research and Applied Sciences*, 8(2), 160–165. doi:10.1016/j.jrras.2014.06.004
- Knob, A., & Carmona, E. C. (2008). Xylanase production by *Penicillium sclerotiorum* and its characterization. *World Applied Sciences Journal*, 4(2), 277–283. doi:10.1080/08905430903320958
- Knob, A., Beitel, S. M., Fortkamp, D., Terrasan, C. R. F., & Almeida, A. F. De. (2013). Production, purification, and characterization of a major *Penicillium* glabrum xylanase using brewer's spent grain as substrate. *BioMed Research International*, 2013. doi:10.1155/2013/728735
- Koponen, R. (1991). Enzyme systems prove their potential. *Pulp and paper International*, 33, 81–83.
- Lembaga Kemajuan Pertanian Muda (MADA). (2010). Laporan Tahunan, Alor Setar, Kedah.
- Li, Y., Cui, F., Liu, Z., Xu, Y., & Zhao, H. (2007). Improvement of xylanase production by *Penicillium oxalicum* ZH-30 using response surface methodology. *Enzyme* and Microbial Technology, 40(5), 1381–1388. doi:10.1016/j.enzmictec.2006.10.015

- Lian, H. L., You, J. X., Huang, Y. N., & Li, Z. Z. (2012). Effect of refining on delignification with a laccase /xylanase treatment. *BioResources*, 7(4), 5268– 5278.
- Liu, J., Yuan, X., Zeng, G., Shi, J., & Chen, S. (2006). Effect of biosurfactant on cellulase and xylanase production by *Trichoderma viride* in solid substrate fermentation. *Process Biochemistry*, 41, 2347-2351. doi:10.1016/j.procbio.2006.05.014
- Longui, E. L., Brémaud, I., Da Silva, F. G., Lombardi, D. R., & Alves, E. S. (2012). Relationship among extractives, lignin and holocellulose contents with performance index of seven wood species used for bows of string instruments. *IAWA Journal*, 33(2), 141-149.
- Meryandini, A., Hendarwin, T., Sapurudin, D., & Lestari, Y. (2006). Characterization of Xylanase *Streptomyces* spp. SKK1-8. *Hayati: Journal of Biosciences*, 13(4), 151–155.
- Motta, F., Andrade, C., & Santana, M. (2013). A Review of xylanase production by the fermentation of xylan: classification, characterization and applications. *Intech, Chapter 10*, 251–275. doi:10.5772/53544
- Nagar, S., Jain, R. K., Thakur, V. V., & Gupta, V. K. (2013). Biobleaching application of cellulase poor and alkali stable xylanase from *Bacillus pumilus* SV-85S. 3 *Biotech*, 3(4), 277–285. doi:10.1007/s13205-012-0096-y
- Nair, S. G., Sindhu, R., & Shashidhar, S. (2010). Enzymatic bleaching of kraft pulp by xylanase from *Aspergillus sydowii* SBS 45. *Indian Journal of Microbiology*, 50(3), 332–338. doi:10.1007/s12088-010-0049-2
- Nakamura, S., Wakabayashu, K., Nakai, R., Aono, R., & Horikoshi, K. (1993). Purification and some properties of an alkaline xylanase from alkaliphilic *Bacillus* sp. strain 41M-1. *Applied of Environment Microbiology*, 59(7), 2311-2316
- Noraida Abd Rahim, S., Sulaiman, A., Halim Ku Hamid, K., Aini Edama, N., & Samsu Baharuddin, A. (2015). Effect of agitation speed for enzymatic hydrolysis of tapioca slurry using encapsulated enzymes in an enzyme bioreactor. *International Journal of Chemical Engineering and Applications*, 6(1), 38–41. doi:10.7763/IJCEA.2015.V6.447
- Ó'Fágáin, C. (2003). Enzyme stabilization Recent experimental progress. *Enzyme and Microbial Technology*, 33(2-3), 137–149. doi:10.1016/S0141-0229(03)00160-1
- Opasols, a O., & Adewoye, S. O. (2010). Assessment of degradability potential of *Penicillium oxalicum* on crude oil. *Advances in Applied Science Reserach*, 1(1), 182–188.

- Pal, A., & Khanum, F. (2011). Identification and optimization of critical medium components using statistical experimental designs for enhanced production of xylanase from *Aspergillus flavus* DFR-6. *Food Technology and Biotechnology*, 49(2), 228–236.
- Palma, M. B., Ferreira Milagres, A. M., Prata, A. M. R., & de Mancilha, I. M. (1996). Influence of aeration and agitation rate on the xylanase activity from *Penicillium janthinellum. Process Biochemistry*, 31(2), 141–145. doi:10.1016/0032-9592(95)00042-9
- Pandey, S., Shahid, M., Srivastava, M., Singh, A., Sharma, A., Kumar, V., & Srivastava, Y. K. (2014). Effect of Various Physiological Parameters and Different Carbon Sources on Cellulase and Xylanase Induction by Different Strains of *Trichoderma* Species. *Enzyme Engineering*, 3(1), 1–5. doi:10.4172/2329-6674.1000120
- Pierson, Y., Bobbink, F., & Yan, N. (2013). Alcohol mediated liquefaction of lignocellulosic materials: A mini review. *Chemical Engineering & Process Techniques*, 1(2), 1014–1019.
- PiresDo Nascimento, R., Marques, S., Alves, L., Girio, F., Amaral-Collaco, M. T., Rodrigues Sacramento, D., Pinto da Silva Bon, E., & Rodrigues Coelho, R. R. (2003). A novel strain of *Streptomyces malaysiensis* isolated from Brazilian soil produces high endo-β-1,4- xylanase titres. *World Journal of Microbiology and Biotechnology*, 19(9), 879–881. doi:10.1023/B:WIBI.0000007287.03281.dd
- Purwanto, L. A., Ibrahim, D., & Sudrajat, H. (2009). Effect of agitation speed on morphological changes in Aspergillus niger hyphae during production of tannase. World Journal of Chemistry, 4(1), 34–38.
- Ramakrishnan, J., & Narayanan, M. (2013). Studies on xylanase producing thermophilic Streptomyces sp from compost soil. International Journal of PharmTech Research, 5(3), 1386–1392.
- Rennie, E. a., & Scheller, H. V. (2014). Xylan biosynthesis. Current Opinion in Biotechnology, 26, 100–107. doi:10.1016/j.copbio.2013.11.013
- Ribeiro, L. F. C., Ribeiro, L. F., Jorge, J. A., & Polizeli, M. L. T. M. (2014). Screening of filamentous fungi for xylanases and cellulases not inhibited by xylose and glucose. *British Biotechnology Journal*, 4(1), 30–39.
- Salihu, A., & Alam, Z. (2012). Production and applications of microbial lipases: A review. *Scientific Research and Essays*, 7(30), 2667–2677. doi:10.5897/SRE11.2023
- Samanta, a. K., Kolte, A. P., Senani, S., Sridhar, M., & Jayapal, N. (2011). A simple and efficient diffusion technique for assay of endo β-1,4-xylanase activity. *Brazilian Journal of Microbiology*, 42(4), 1349–1353. doi:10.1590/S1517-83822011000400016

- Sarkar, N., & Aikat, K. (2012). Cellulase and xylanase production from rice straw by a locally isolated fungus Aspergillus fumigatus NITDGPKA3 under solid state fermentation – statistical optimization by response surface methodology. Journal of Technology Innovations in Renewable Energy, 1, 54–62. doi:10.6000/1929-6002.2012.01.01.7
- Sarkar, N., & Aikat, K. (2014). Aspergillus fumigatus NITDGKA3 provides for increased cellulase production. International Journal of Chemical Engineering, 2014,9. doi: 10.1155/2014/959845
- Sathiyavathi, M., & Parvatham, R. (2013). Industrial application of xylanase in the crude enzyme extract from *Trichoderma* sp.MS 2010. *Asian Journal of Pharmaceutical and Clinical Research*, 6(SUPPL. 2), 90–94.
- Schär-zammaretti, P., Dillmann, M., Amico, D., Affolter, M., Ubbink, J., Scha, P., & Amico, N. D. (2005). Influence of fermentation medium composition on physicochemical surface properties of *Lactobacillus acidophilus*. *Applied and Enviromental Microbiology*, 71(12), 8165–8173. doi:10.1128/AEM.71.12.8165
- Scurlocka, J. M. O., Dayron, D. C., & Hamesb, B. (2000). Bamoboo: an overlooked biomass resource?. *Biomass and Energy*, 19, 229-224.
- Sekyere, D. (1994). Potential of bamboo (Bambusa Vulgaris) as a source of raw material for pulp and paper in Ghana. *Ghana Journal of Forestry, 1.*
- Sepahy, A. A., Ghazi, S., & Sepahy, M. A. (2011). Cost-Effective Production and Optimization of alkaline xylanase by indigenous *Bacillus mojavensis* AG137 fermented on agricultural waste. *Enzyme Research*, 2011, 9. doi:10.4061/2011/593624
- Sharma, A., Thakur, V. V., Shrivastava, A., Jain, R. K., Mathur, R. M., Gupta, R., & Kuhad, R. C. (2014). Xylanase and laccase based enzymatic kraft pulp bleaching reduces adsorbable organic halogen (AOX) in bleach effluents: A pilot scale study. *Bioresource Technology*, 169, 96–102. doi:10.1016/j.biortech.2014.06.066
- Sindhu, R., Kuttiraja, M., Binod, P., Janu, K. U., Sukumaran, R. K., & Pandey, A. (2011). Dilute acid pretreatment and enzymatic saccharification of sugarcane tops for bioethanol production. *Bioresource Technology*, 102(23), 10915– 10921. doi:10.1016/j.biortech.2011.09.066
- Sindhu, R., Kuttiraja, M., Binod, P., Janu, K. U., Sukumaran, R. K., & Pandey, A. (2011). Dilute acid pretreatment and enzymatic saccharification of sugarcane tops for bioethanol production. *Bioresource Technology*, 102(23), 10915– 10921. doi:10.1016/j.biortech.2011.09.066
- Singh, P., & Gill, P. K. (2006). Production of inulinases: Recent advances. Food Technology and Biotechnology, 44(2), 151–162.

- Sixta, H. (2006). *Chemical Pulping*. In H. Sixta (ed.), Handbook of Pulp (pp. 1–20). KGaA, Weinheim: Wiley-Vch Verlag GmbH & Co.
- Smook, G. A. (1992). Overview of pulping methodology. Handbook for Pulp & Paper Technologists. Vancouver: Angus Wilde Publications.
- Sohpal, V. K., Dey, A., & Singh, A. (2010). Investigate of process parameters on xylanase enzyme activity in Melanocarpus Albomyces batch culture. *Proceedings of the World*
- Sorek, N., Yeats, T. H., Szemenyei, H., Youngs, H., & Somerville, C. R. (2014). The implications of lignocellulosic biomass chemical composition for the production of advanced biofuels. *BioScience*, 64(3), 192–201. doi:10.1093/biosci/bit037
- Sridevi, B., & Charya, M. A. S. (2011). Isolation, identification and screening of potential cellulase-free xylanase producing fungi, 10(22), 4624–4630. doi:10.5897/AJB10.2108
- Taneja, K., Gupta, S., & Kuhad, R. C. (2002). Properties and application of a partially purified alkaline xylanase from an alkalophilic fungus Aspergillus nidulans KK-99. Bioresource Technology, 85(1), 39–42.
- TAPPI Test Method T203-om-99 (1996). Technical Association of the Pulp and Paper Industry, Tappi Press, Atlanta, GA
- TAPPI Test Method T204 cm-97 (1996). Technical Association of the Pulp and Paper Industry, Tappi Press, Atlanta, GA
- TAPPI Test Method T222-om-02 (1996). Technical Association of the Pulp and Paper Industry, Tappi Press, Atlanta, GA
- TAPPI Test Method T452-om-98 (1996). Technical Association of the Pulp and Paper Industry, Tappi Press, Atlanta, GA.
- Techapun, C., Poosaran, N., Watanabe, M., & Sasaki, K. (2003). Thermostable and alkaline- tolerant microbial cellulase-free xylanases produced from agricultural wastes and the properties required for use in pulp bleaching bioprocesses: a review. *Process Biochemistry*, 38(9), 1327–1340. doi:10.1016/S0032-9592(02)00331-X
- Thygesen, A., Oddershede, J., Lilholt, H., Thomsen, A. B., & Ståhl, K. (2005). On the determination of crystallinity and cellulose content in plant fibres. *Cellulose*, 12(6), 563–576. doi:10.1007/s10570-005-9001-8
- Tiwari, P., Jain, R., Kumar, K., Panik, R., & Sahu, P. K. (2011). An evaluation of antimicrobial activities of root extract of *Calendula officinalis*(LINN.). *Newsletter*, 892, 886–892.
- United States Department of Agriculture (USDA) (2014). Available from: http://worldriceproduction.com (accessed June 24, 2015).

- Wahab, R., Mustafa, M. T., Rahman, S., Salam, M. A., Sulaiman, O., Sudin, M., & Mohd Sukhairi, M. R. (2012). Relationship between physical, anatomical and strength properties of 3-year-old cultivated tropical bamboo *Gigantochloa scortechinii*. *Journal of Agricultural and Biological Science*, 7(10), 782–791. Retrieved from http://www.arpnjournals.com/jabs/research_papers/rp_2012/jabs_1012_464.p df
- Wahab, R., Mustafa, M. T., Salam, M. A., Sudin, M., Samsi, H. W., & Rasat, M. S. M. (2013). Chemical composition of four cultivated tropical bamboo in genus *Gigantochloa. Journal of Agricultural Science*, 5(8), 66–75. doi:10.5539/jas.v5n8p66
- Wise, L., Murphy, M., & Daddieco, A. (2017). Chlorine holocennulose, its fractionation and bearing on summative wood analysis and on studies on the hemicelluloses. *Paper Trade Journal* 122(2), 11-19.
- Wong, K. K., Tan, L. U., & Saddler, J. N. (1988). Multiplicity of beta-1,4-xylanase in microorganisms: functions and applications. *Microbiological Reviews*, 52(3), 305–317.
- Yeasmin, S., Kim, C. H., Park, H. J., Sheikh, M. I., Lee, J. Y., Kim, J. W., Back, K. K., Kim, S. H. (2011). Cell surface display of cellulase activity-free xylanase enzyme on saccharomyces cerevisiae EBY100. *Applied Biochemistry and Biotechnology*, 164(3), 294–304. doi:10.1007/s12010-010-9135-5
- Ziebell, A., Gracom, K., Katahira, R., Chen, F., Pu, Y., Ragauskas, A., Dixon, R. A., & Davis, M. (2010). Increase in 4-coumaryl alcohol units during lignification in alfalfa (Medicago sativa) alters the extractability and molecular weight of lignin. *Journal of Biological Chemistry*, 285(50), 38961–38968. doi:10.1074/jbc.M110.137315