



***STATISTICAL APPROACH FOR ENZYME PRODUCTION USING
RESPONSE SURFACE METHODOLOGY AND STRUCTURE
PREDICTION OF THERMOSTABLE ORGANIC SOLVENT-TOLERANT
RAND PROTEASE***

RANDA ABDEL KAREEM HUSEIN

FBSB 2018 59



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By

RANDA ABDEL KAREEM HUSEIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirement for the degree of Doctor of Philosophy**

December 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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December 2017

Chairman : Professor Raja Noor Zaliha Raja Abd. Rahman, D.Eng.
Faculty : Biotechnology and Biomolecular Sciences

Serine proteases from the *Bacillus* species extensively applied in the biotechnological application. So far, the broad investigation on proteases gave the basic understanding of their catalytic mechanism and their structure-function. Computational structure analysis and homology modelling can be a key process for the 3D structure reconstruction which facilitates the protein-protein interaction research. Protein crystal is the basic necessity to obtain the 3D structure. The crystallisation process requires ample amount of protein. *Bacillus subtilis* strain Rand could only express the low amount of the protein. The Rand protease has unique characteristics and is the first thermostable and organic solvent tolerant protease that has been reported. Therefore, the structural study is needed to understand the enzyme properties. A statistical approach, response surface methodology (RSM), was performed to optimise the production of extracellular Rand protease in bioreactor stirred tank. Consequently, a face-centred, central composite design (CCD) falling under RSM was employed to enhance the protease activity further. The interactive effect of these parameters resulted in a 1.6-fold increase in protease production. An analysis of the variance showed that the adequacy of the model and verification experiments confirmed its validity. Crystallisation of the purified wild-type protein performed under microgravity conditions in space as well as in the ground. There is no crystal form observed in the ground control but, Rand protease protein successfully crystallised under microgravity conditions. A structural prediction for the Rand protease was built using the Yet Another Scientific Artificial Reality Application (YASARA) structure employing a known 3D structure (subtilisin E-propeptide complex; PDB ID: 1SCJ) as a template, which has the highest sequence similarity (96%) to the Rand protease.

The predicted 3D structure of the Rand protease revealed that the topological organization of the α/β -hydrolase fold consisted of 6 α -helices and 13 β -strands. In silico study of docking, the substrate N-succinyl-alanyl-alanyl-prolyl-phenylalanine-4-nitroanilide in Rand protease resulted in 25 clusters whereby 4 clusters observed to involve the catalytic triad of rand protease that is Asp32, His64, and Ser221. This result is in good agreement with the active site prediction and several experimental studies, which shows the same conserved catalytic triad. Molecular dynamics (MD) simulations were performed in two organic solvents with different $\log P$ values, such as pyridine ($\log P$ 0.71), benzene ($\log P$ 2.0) and pure water, for 10 ns to investigate their effect on the Rand protease structure. In conclusion, the production of Rand protease using a bioreactor through RSM could increase the yield of this enzyme compared to using a shake flask. The structure analysis confirmed the unique characteristics of this enzyme and explained the organic solvent stability of the enzyme. The predicted structure clarified the use of HIC as the first step for purification by highlighting the number of hydrophobic residues on the surface of the protein.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENDEKATAN STATISTIK BAGI PENGELUARAN ENZYME MELALUI
METODOLOGI PENYELESAIAN RESPONSE DAN JANGKAAN
STRUKTUR PROTEASE TERMOSTABIL TAHAN PELARUT ORGANIK**

Oleh

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Serine protease dari *Bacillus* sp. telah digunakan secara meluas dalam aplikasi bioteknologi. Justeru itu, protease telah diselidik secara meluas dari sudut pengetahuan asas mekanisma katalisis dan hubungan fungsi struktur. Analisis struktur berkomputer dan permodelan homologi adalah proses utama untuk pembinaan semula struktur 3 dimensi yang memudahkan kajian hubungan protein. Kristal protein adalah keperluan asas untuk mendapatkan struktur 3 dimensi. Proses penghabluran memerlukan jumlah protein yang mencukupi. *Bacillus subtilis* strain Rand telah hanya dapat mengeluarkan jumlah protein yang rendah. Protease Rand mempunyai ciri-ciri yang unik dan merupakan protease termostabil dan toleran kepada pelarut organik yang pertama telah dilaporkan. Oleh itu, kajian struktur diperlukan untuk memahami sifat-sifat enzim. Satu pendekatan statistik, metodologi permukaan tindak balas (RSM), dilakukan untuk mengoptimumkan penghasilan protease di luar sel dalam bioreaktor. Oleh itu, rekabentuk komposit pusat (CCD) yang berpusat di bawah RSM digunakan untuk meningkatkan lagi aktiviti protease. Kesan interaktif parameter ini menghasilkan peningkatan 1.6 kali ganda dalam penghasilan protease. Analisis varian menunjukkan ketepatan model tersebut dan pengesahan kajian telah menunjukkan ianya berkesan. Enzim yang dituliskan telah dihablurkan di bawah kondisi mikrograviti di angkasa dan juga di bumi. Tiada hablur terberntuk di bumi, tetapi protein Rand protease berjaya dihablurkan di bawah kondisi mikrograviti. Jangkaan struktur Rand protease telah dibuat menggunakan aplikasi YASARA (*Yet Another Scientific Artificial Reality Application*) berdasarkan struktur 3 dimensi (*Subtilisin E-propeptide complex*; PDB ID: 1SCJ) sebagai rujukan yang mempunyai persamaan jujukan terbanyak (96%) dengan Rand protease. Jangkaan struktur 3 dimensi Rand protease mengandungi organisasi topologi α/β -hidrolase lipatan yang mengandungi 6 helik α dan 13 bebenang β . Dalam kajian silico untuk mengendalikan substrat *N-succinyl-alanyl-alanyl-prolyl-*

phenylalanine-4-nitroanilide dalam protease Rand sebanyak 25 kelompok telah dihasilkan di mana 4 kelompok yang diperhatikan melibatkan triad katalitik protease Rand iaitu Asp32, His64, dan Ser221. Hasilnya adalah seiring dengan berikutan ramalan tapak aktif dan beberapa percubaan kajian, menunjukkan triad pemangkin yang sama dipelihara. Simulasi dinamik molekul telah dilakukan di dalam 3 pelarut organik dengan log P berbeza dengan pyridine (log P 0.71), benzene (log P 2.0) dan air selama 10 ns untuk mengkaji kesannya terhadap struktur Rand protease. Kesimpulannya, penghasilan Rand protease menggunakan bioreaktor melalui RSM mampu meningkatkan penghasilan enzim jika dibandingkan dengan menggunakan kelalang kon. Analisis struktur mengesahkan karakter unik enzim ini dan menjelaskan kestabilan enzim di dalam pelarut organik. Jangkaan struktur telah menyokong penggunaan kromatografi hubungan hidrofobik (HIC) sebagai langkah permulaan untuk penulenan enzim ini dengan memberikan bilangan residu yang mempunyai ciri hidrofobik pada permukaan protein.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS AND SYMBOL

Å	Angstrom
ANOVA	Analysis of Variance
Bp	Base pair
BSA	Bovine serum albumin
CaCl ₂ .2H ₂ O	Calcium Chloride
Cm	Centimeter
CCD	Central composite design
Da	Dalton
dH ₂ O	Distilled water
DNA	Deoxyribonucleic acid
DOT	Dissolved oxygen tension
IS	Inoculum size
DTT	Dithiothreitol
EC	Enzyme Commission
EDTA	Ethylene diamine tetra-acetic acid
FID	Free Interface Diffusion
g	Gram
HPLC	High Performance Liquid Chromatography
hr	Hour
IPTG	Isopropyl-β-D-thiogalactopyranoside
kDa	Kilo Dalton
L	Litre
MgSO ₄ .7H ₂ O	Magnesium Sulphate
µg	Microgram
µl	Microlitre
µmol	Micromole
mg	Milligram
ml	Millilitre

min	Minute
M	Molar
MD	Molecular Dynamics
MEGA	Molecular Evolutionary Genetics Analysis
MW	Molecular Weight
PEG	Polyethylene glycol
pH	Negative logarithm of hydrogen ion concentration
ORF	Open reading frame
PDB	Protein Data Bank
PMSF	Phenyl methyl sulphonyl fluoride
pmol	Picomole
PCR	Polymerase chain reaction
RSM	Response Surface Methodology
RNA	Ribonucleic acid
RMSD	Root Mean Square Deviation
rpm	Rotation per minute
sec	Second
sp.	Species
STDV	Standard deviation
Temp	Temperature
TCA	Trichloroacetic acid
TSB	Tryptone Soya Broth
U	Unit of enzyme activity
U/mg	Unit per milligram
v/v	Volume per volume
w/v	Weight per volume



LIST OF SYMBOLS FOR AMINO ACIDS

A	Ala	Alanine
R	Arg	Arginine
N	Asn	Asparagine
D	Asp	Aspartate
C	Cys	Cysteine
E	Glu	Glutamate
Q	Gln	Glutamine
G	Gly	Glycine
H	His	Histidine
I	Ile	Isoleucine
L	Leu	Leucine
K	Lys	Lysine
M	Met	Methionine
F	Phe	Phenylalanine
P	Pro	Proline
S	Ser	Serine
T	Thr	Threonine
W	Trp	Tryptophan
Y	Tyr	Tyrosine
V	Val	Valine

CHAPTER 1

INTRODUCTION

1.1 Background

Currently, enzymes play a significant role, throughout the globe, due to their invaluable applications. Involved in the production of foods, pharmaceuticals and detergents; analysis of clinical samples; alcohol fermentation; and syntheses of organic products; as well as in biotechnology (Malathu *et al.*, 2008; Sanchez and Demain, 2011; Adrio and Demain, 2014). The global market value of enzymes for industrial purposes was approximately \$3.3 billion in 2010 (Souza *et al.*, 2015) and from USD 6.4 to 6.9 billion in 2017 (Ferrer *et al.*, 2016). However, the world enzyme demand is forecasted to grow from average 4.6 percent through 2020 to \$7.2 billion (Freedonia Group 2016).

Enzymes obtained from microbes serve mainly as resources utilised by industries such as those for food, chemicals and their associates in the production of a variety of biotechnological products (Bhunia *et al.*, 2012). The enzyme subtilisin as one of the proteolytic enzymes, given its vast industrial applications, represents the commercial enzyme available in the most massive quantities. It is mainly produced for the manufacture of detergent additives, and in the processing industries for skin and leather. Subtilisin has, therefore, been extensively researched as a promising target for protein engineering from both the basic and applied perspectives (Hiroshi, 1993).

The genus *Bacillus* is one of the main extracellular protease producers, and industries mostly used the species *Bacillus subtilis* in particular for the production of various enzymes. *Bacillus subtilis* is a rod-shaped Gram-positive bacterium, and its natural habit in soil. It may form a hard or harsh, protective sub-terminal endospore that can withstand high temperatures and other extreme environmental conditions without losing viability when in favourable conditions (Pant *et al.*, 2015).

The protein structures provide sufficient details about its functions, the location of functional sites and clues to find the critical residues (Aloy *et al.*, 2001; Reddy *et al.*, 2001; Yao *et al.*, 2003). Moreover, X-ray crystallography increases the understanding of the functions of a protein and can be used to visualise the protein structure at the atomic level (Rupp, 2009). The Rand protease gene has 96% similarity with the subtilisin-propeptide complex gene (PDB ID: 1SCJ). To understand the structural function of Rand protease enzyme, further studies on structure prediction are necessary. *Bacillus subtilis* strain Rand was reported to have produced a thermo-stable and organic solvent protease. Moreover, the enzyme

showed stability in both polar and non-polar organic solvents, which is a unique property among thermostable proteases (Abusham *et al.*, 2009).

1.2 Problem statement

Rand protease is a thermostable and organic solvent tolerant enzyme with broad applications. However, a study on the structure of such an enzyme is limited. Three-dimensional (3D) structure of a protein is needed to understand their properties. In order to solve the structure, a high concentration of protein is required for protein crystallisation. Production of wild-type Rand protease in shake flask is very low and not enough for protein crystallisation.

1.3 Hypothesis

Production of enough and large quantities of the enzyme for crystallisation purposes is a tall order, and protein crystal is the basic necessity of obtaining the 3D structure. Characterization of Rand protease showed that the enzyme is stable at high temperature as well as in the presence of polar and nonpolar organic solvent, which is unique property among other proteases. Nevertheless, conventional optimisation of production medium in shake flask could not increase the amount of enzyme. A further study on enzyme production in bioreactor stirred tank is necessary to improve the yield of protein, enough for crystallisation, study the structure and to understand the enzyme properties.

1.4 Rational of scope of research

In this work, there were four main steps involved. The first step was the production of Rand protease in 7.5 L bioreactor using RSM technique to produce a high quantity of enzyme. Secondly, purification and crystallisation of the enzyme were carried out. Next, isolation of Rand protease gene was conducted. Finally, a further study on protein prediction was done.

1.5 Objective

This study aims to obtain enough amount of Rand protease for crystallisation and to study the structure-function relationship of the enzyme. To achieve this aim, the following work had to be achieved:

- 1) To enhance Rand protease production in the bioreactor by applying response surface methodology (RSM).
- 2) To crystallise Rand protease under gravity and microgravity conditions.
- 3) To isolate Rand protease gene
- 4) To predict 3D structure of Rand protease: a silico study
- 5) To evaluate the effect of organic solvents on overall conformation of Rand protease structure through molecular dynamics (MD) simulations.

REFERENCES

- Abusham, R. A. 2009. Isolation and characterization of thermostable organic solvent tolerant protease from *Bacillus subtilis* isolate Rand. MS. Thesis. Thesis, Universiti Putra Malaysia, Serdang.
- Abusham, R. A., Rahman, R. N. Z. R., Salleh, A. B., and Basri, M. 2009. Optimization of physical factors affecting the production of thermo-stable organic solvent-tolerant protease from a newly isolated halo tolerant *Bacillus subtilis* strain Rand. *Microbial Cell Factories* 8 (1):20.
- Adinarayana, K. and Ellaiah, P. 2002. Response surface optimization of the critical medium components for the production of alkaline protease by a newly isolated *Bacillus* sp. *Journal of Pharmacy and Pharmaceutical Sciences* 5 (3):272-278.
- Adrio, J. L. and Demain, A. L. 2005. Microbial cells and enzymes a century of progress. *Microbial Enzymes and Biotransformations* 17:1-27.
- Adrio, J. L. and Demain, A. L. 2014. Microbial enzymes: Tools for biotechnological processes. *Biomolecules* 4 (1):117-139.
- Agarwal, S. K. 1990. Proteases cathepsins – A view. *Biochemical Education* 18 (2):67-72.
- Agrebi, R., Haddar, A., Hmidet, N., Jellouli, K., Manni, L., and Nasri, M. 2009. BSF1 fibrinolytic enzyme from a marine bacterium *Bacillus subtilis* A26: Purification, biochemical and molecular characterization. *Process Biochemistry* 44 (11):1252-1259.
- Akhavan Sepahy, A. and Jabalameli, L. 2011. Effect of culture conditions on the production of an extracellular protease by *Bacillus* sp. isolated from soil sample of Lavizan Jungle Park. *Enzyme Research* 2011.
- Alberts, B., Bray, D., Lewis, J., Raff, M., Roberts, K., Watson, J. D., and Grimstone, A. 1995. Molecular Biology of the Cell (3rd edn). *Trends in Biochemical Sciences* 20 (5):210-210.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., and Walter, P. 2002. The shape and structure of proteins. In *Molecular Biology of the Cell*. 4th edition. Garland Science, New York.
- Ali, A., Majeed, H., and Abdulhusain, N. 2011. Molecular cloning and expression of *Bacillus stearothermophilus* protease gene in *Escherichia coli*. *Journal of Biology and Life Sciences* 2 (1):26-31.
- Ali, M. S. M., Said, Z. S., Mat, A., Rahman, R. N. Z. R. A., Chor, A. L. T., Basri, M., and Salleh, A. B. 2013. Capillary-seeding crystallization and preliminary

crystallographic Analysis of a solvent-tolerant elastase from *Pseudomonas aeruginosa* strain K. *International Journal of Molecular Sciences* 14 (9):17608-17617.

Almas, S., Hameed, A., and Mohan, P. 2009. Purification and characterization of a novel protease from *Bacillus* strain SAL1. *African Journal of Biotechnology* 8 (15):3603.

Aloy, P., Mas, J., Martí-Renom, M., Querol, E., Avilés, F., and Oliva, B. 2000. Refinement of modelled structures by knowledge-based energy profiles and secondary structure prediction: Application to the human procarboxypeptidase A2. *Journal of Computer-Aided Molecular Design* 14 (1):83-92.

Aloy, P., Querol, E., Aviles, F. X., and Sternberg, M. J. E. 2001. Automated structure-based prediction of functional sites in proteins: applications to assessing the validity of inheriting protein function from homology in genome annotation and to protein docking. *Journal of Molecular Biology* 311 (2):395-408.

Alvarez, M., Zeelen, J. P., Mainfroid, V., Rentier-Delrue, F. o., Martial, J. A., Wyns, L., Wierenga, R. K., and Maes, D. 1998. Triose-phosphate Isomerase (TIM) of the Psychrophilic Bacterium *Vibrio marinus* KINETIC AND STRUCTURAL PROPERTIES. *Journal of Biological Chemistry* 273 (4):2199-2206.

Amitai, G., Shemesh, A., Sitbon, E., Shklar, M., Netanel, D., Venger, I., and Pietrokovski, S. 2004. Network analysis of protein structures identifies functional residues. *Journal of Molecular Biology* 344 (4):1135-1146.

Anbu, P. 2016. Enhanced production and organic solvent stability of a protease from *Brevibacillus laterosporus* strain PAP04. *Brazilian Journal of Medical and Biological Research* 49 (4).

Argos, P., Rossmann, M. G., Grau, U. M., Zuber, H., Frank, G., and Tratschin, J. D. 1979. Thermal stability and protein structure. *Biochemistry* 18 (25):5698-5703.

Asker, M. M., Mahmoud, M. G., El Shebwy, K., and el Aziz, M. S. A. 2013. Purification and characterization of two thermostable protease fractions from *Bacillus megaterium*. *Journal of Genetic Engineering and Biotechnology* 11 (2):103-109.

Baneyx, F. o. 1999. Recombinant protein expression in *Escherichia coli*. *Current Opinion in Biotechnology* 10 (5):411-421.

Barredo, J.-L. 2005. *Microbial enzymes and biotransformations*. Springer.

- Baş, D. and Boyacı, İ. H. 2007. Modeling and optimization I: Usability of response surface methodology. *Journal of Food Engineering* 78 (3):836-845.
- Beg, Q. K. and Gupta, R. 2003. Purification and characterization of an oxidation-stable, thiol-dependent serine alkaline protease from *Bacillus mojavenensis*. *Enzyme and Microbial Technology* 32 (2):294-304.
- Beg, Q. K., Saxena, R., and Gupta, R. 2002. De-repression and subsequent induction of protease synthesis by *Bacillus mojavenensis* under fed-batch operations. *Process Biochemistry* 37 (10):1103-1109.
- Bekler, F. M., Acer, Ö., and Guven, K. 2015. Production and purification of novel thermostable alkaline protease from *Anoxybacillus* sp. KP1. *Cellular and Molecular Biology* 61 (4):113-120.
- Bendtsen, J. D., Nielsen, H., von Heijne, G., and Brunak, S. 2004. Improved prediction of signal peptides: SignalP 3.0. *Journal of Molecular Biology* 340 (4):783-795.
- Benkert, P., Biasini, M., and Schwede, T. 2011. Toward the estimation of the absolute quality of individual protein structure models. *Bioinformatics* 27 (3):343-350.
- Betzl, C., Dauter, Z., Dauter, M., Ingelman, M., Papendorf, G., Wilson, K. S., and Branner, S. 1988. Crystallization and preliminary X-ray diffraction studies of an alkaline protease from *Bacillus lentus*. *Journal of Molecular Biology* 204 (3):803-804.
- Betzl, C., Teplyakov, A., Harutyunyan, E., Saenger, W., and Wilson, K. 1990. Thermitase and proteinase K: a comparison of the refined three-dimensional structures of the native enzymes. *Protein Engineering* 3 (3):161-172.
- Bhunja, B., Basak, B., and Dey, A. 2012. A review on production of serine alkaline protease by *Bacillus* spp. *Journal of Biochemical Technology* 3 (4):448-457.
- Biosciences, A. 2001. Protein purification handbook. *Edition AC* 98.
- Biosciences, A. 2002. Ion Exchange Chromatography, Principles and Methods. *ISBN* 91:970490-3.
- Bjerga, G. E. K., Arsin, H., Larsen, Ø., Puntervoll, P., and Kleivdal, H. T. 2016. A rapid solubility-optimized screening procedure for recombinant subtilisins in *E. coli*. *Journal of Biotechnology* 222:38-46.
- Blom, E.-J., Ridder, A. N., Lulko, A. T., Roerdink, J. B., and Kuipers, O. P. 2011. Time-resolved transcriptomics and bioinformatic analyses reveal intrinsic stress responses during batch culture of *Bacillus subtilis*. *PloS one* 6 (11):e27160.

- Bode, W., Papamokos, E., and Musil, D. 1987. The high-resolution X-ray crystal structure of the complex formed between subtilisin Carlsberg and eglin c, an elastase inhibitor from the leech *Hirudo medicinalis*. *European Journal of Biochemistry* 166 (3):673-692.
- Bohlin, J., Snipen, L., Hardy, S. P., Kristoffersen, A. B., Lagesen, K., Dønsvik, T., Skjerve, E., and Ussery, D. W. 2010. Analysis of intra-genomic GC content homogeneity within prokaryotes. *BMC Genomics* 11:464
- Bolanos-Garcia, V. M. and Chayen, N. E. 2009. New directions in conventional methods of protein crystallization. *Progress in Biophysics and Molecular Biology* 101 (1):3-12.
- Bornot, A., Etchebest, C., and De Brevern, A. G. 2011. Predicting protein flexibility through the prediction of local structures. *Proteins: Structure, Function, and Bioinformatics* 79 (3):839-852.
- Bourgeau, G., Lapointe, H., Peloquin, P., and Mayrand, D. 1992. Cloning, expression, and sequencing of a protease gene (tpr) from *Porphyromonas gingivalis* W83 in *Escherichia coli*. *Infection and Immunity* 60 (8):3186-3192.
- Bowman, G. R. and Pande, V. S. 2009. The roles of entropy and kinetics in structure prediction. *PloS one* 4 (6):e5840.
- Box, G. E. and Wilson, K. 1951. On the experimental attainment of optimum conditions. *Journal of the Royal Statistical Society. Series B (Methodological)* 13 (1):1-45.
- Bradford, M. M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72:248-254.
- Braun, P., Tommassen, J., and Filloux, A. 1996. Role of the propeptide in folding and secretion of elastase of *Pseudomonas aeruginosa*. *Molecular Microbiology* 19 (2):297-306.
- Brown, P., Butts, C. P., Eastoe, J., Fermin, D., Grillo, I., Lee, H.-C., Parker, D., Plana, D., and Richardson, R. M. 2012. Anionic surfactant ionic liquids with 1-butyl-3-methyl-imidazolium cations: characterization and application. *Langmuir* 28 (5):2502-2509.
- Brünger, A. T., Adams, P. D., Clore, G. M., DeLano, W. L., Gros, P., Grosse-Kunstleve, R. W., Jiang, J.-S., Kuszewski, J., Nilges, M., and Pannu, N. S. 1998. Crystallography & NMR system: a new software suite for macromolecular structure determination. *Acta Crystallographica Section D: Biological Crystallography* 54 (5):905-921.

- Bryan, P., Alexander, P., Strausberg, S., Schwarz, F., Lan, W., Gilliland, G., and Gallagher, D. T. 1992. Energetics of folding subtilisin BPN'. *Biochemistry* 31 (21):4937-4945.
- Bryan, P., Wang, L., Hoskins, J., Ruvinov, S., Strausberg, S., Alexander, P., Almog, O., Gilliland, G., and Gallagher, T. 1995. Catalysis of a protein folding reaction: Mechanistic implications of the 2.0. Å structure of the subtilisin-prodomain complex. *Biochemistry* 34 (32):10310-10318.
- Campbell, M. K. 1999. *Biochemistry*, 3rd edition. Harcourt College Pub Hardcover.
- Castillo, B., Bansal, V., Ganesan, A., Halling, P., Secundo, F., Ferrer, A., Griebenow, K., and Barletta, G. 2006. On the activity loss of hydrolases in organic solvents: II. a mechanistic study of subtilisin Carlsberg. *BMC Biotechnology* 6 (1):51.
- Chayen, N. E. 2004. Turning protein crystallisation from an art into a science. *Current Opinion in Structural Biology* 14 (5):577-583.
- Chayen, N. E. and Helliwell, J. R. 2002. Microgravity Protein Crystallization. *Annals of the New York Academy of Sciences* 974 (1):591-597.
- Chayen, N. E., Helliwell, J. R., and Snell, E. H. 2010. 3. Practical methods of crystallization. *Macromolecular Crystallization and Crystal Perfection* 1 (9):24-29.
- Chayen, N. E. and Saridakis, E. 2008. Protein crystallization: from purified protein to diffraction-quality crystal. *Nature Methods* 5 (2):147-153.
- Chayen, N. E., Shaw Stewart, P., Maeder, D., and Blow, D. 1990. An automated system for micro-batch protein crystallization and screening. *Journal of Applied Crystallography* 23 (4):297-302.
- Chayen, N. E., Stewart, P. D. S., and Blow, D. M. 1992. Microbatch crystallization under oil-a new technique allowing many small-volume crystallization trials. *Journal of Crystal Growth* 122 (1):176-180.
- Colovos, C. and Yeates, T. O. 1993. Verification of protein structures: patterns of nonbonded atomic interactions. *Protein Science* 2 (9):1511-1519.
- Cornell, J. A. and Khuri, A. I. 1987. *Response surfaces: designs and analyses*. Marcel Dekker, Inc.
- Cristancho, C. A. M., David, F., Franco-Lara, E., and Seidel-Morgenstern, A. 2013. Discontinuous and continuous purification of single-chain antibody fragments using immobilized metal ion affinity chromatography. *Journal of Biotechnology* 163 (2):233-242.

- DeLucas, L. J. 2001. Protein crystallization – is it rocket science? *Drug Discovery Today* 6 (14):734-744.
- Demain, A. L. and Vaishnav, P. 2009. Production of recombinant proteins by microbes and higher organisms. *Biotechnology Advances* 27 (3):297-306.
- Derewenda, Z. S. 2011. It' s all about crystals. *Acta Crystallographica Section D: Biological Crystallography* 67 (4):243-248.
- Divakar, K., Priya, J. D. A., and Gautam, P. 2010. Purification and characterization of thermostable organic solvent-stable protease from *Aeromonas veronii* PG01. *Journal of Molecular Catalysis B: Enzymatic* 66 (3):311-318.
- Doddapaneni, K. K., Tatineni, R., Potumarthi, R., and Mangamoori, L. N. 2007. Optimization of media constituents through response surface methodology for improved production of alkaline proteases by *Serratia rubidaea*. *Journal of Chemical Technology and Biotechnology* 82 (8):721-729.
- Dong, C., Zhang, C.-Y., Liu, Y.-Y., Zhou, R.-B., Cheng, Q.-D., and Yin, D.-C. 2016. A new design of protein crystallization plates to expand concentration screening space in cross-diffusion microbatch and microbatch methods. *Crystal Growth & Design*.
- Dong, D., Ihara, T., Motoshima, H., and Watanabe, K. 2005. Crystallization and preliminary X-ray crystallographic studies of a psychrophilic subtilisin-like protease Apa1 from Antarctic *Pseudoalteromonas* sp. strain AS-11. *Acta Crystallographica Section F: Structural Biology and Crystallization Communications* 61 (3):308-311.
- Dordick, J. S. 1989. Enzymatic catalysis in monophasic organic solvents. *Enzyme and Microbial Technology* 11 (4):194-211.
- Duckert, P., Brunak, S. r., and Blom, N. 2004. Prediction of proprotein convertase cleavage sites. *Protein Engineering Design and Selection* 17 (1):107-112.
- Ducros, E., Ferrari, M., Pellegrino, M., Raspanti, C., and Bogni, C. 2009. Effect of aeration and agitation on the protease production by *Staphylococcus aureus* mutant RC128 in a stirred tank bioreactor. *Bioprocess and Biosystems Engineering* 32 (1):143-148.
- Edman, P. and Begg, G. 1967. A protein sequenator. *European Journal of Biochemistry* 1 (1):80-91.
- Eisenberg, D. 2003. The discovery of the α -helix and β -sheet, the principal structural features of proteins. *Proceedings of the National Academy of Sciences* 100 (20):11207-11210.
- El-Bastawissy, E., Knaggs, M. H., and Gilbert, I. H. 2001. Molecular dynamics simulations of wild-type and point mutation human prion protein at normal

and elevated temperature. *Journal of Molecular Graphics and Modelling* 20 (2):145-154.

Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution*:783-791.

Fenwick, R. B. and Dyson, H. J. 2016. Classic analysis of biopolymer dynamics is model free. *Biophysical Journal* 110 (1):3-6.

Fernandes, J. o. F. A., McAlpine, M., and Halling, P. J. 2005. Operational stability of subtilisin CLECs in organic solvents in repeated batch and in continuous operation. *Biochemical Engineering Journal* 24 (1):11-15.

Ferrer, M., Martínez-Martínez, M., Bargiela, R., Streit, W. R., Golyshina, O. V., and Golyshin, P. N. 2016. Estimating the success of enzyme bioprospecting through metagenomics: current status and future trends. *Microbial Biotechnology* 9 (1):22-34.

Fiser, A. and Sali, A. 2003. Comparative protein structure modeling. In *Protein structure: Determination, analysis, and application for drug discovery*, ed. D. I. Chasman, pp 167-206 New York: Marcel Dekker, Inc.

Forrester, Wright, J., and Forrester, J. W. 1969. *Urban dynamics*. Vol. 114. mIt press Cambridge.

Fujii, M., Takagi, M., Imanaka, T., and Aiba, S. 1983. Molecular cloning of a thermostable neutral protease gene from *Bacillus stearothermophilus* in a vector plasmid and its expression in *Bacillus stearothermophilus* and *Bacillus subtilis*. *Journal of Bacteriology* 154 (2):831-837.

Gallagher, T., Gilliland, G., Wang, L., and Bryan, P. 1995. The prosegment-subtilisin BPN' complex: crystal structure of a specific 'foldase'. *Structure* 3 (9):907-914.

García-Ruiz, J. M. 2003. Counterdiffusion methods for macromolecular crystallization. *Methods in Enzymology* 368:130-154.

Gaur, R., Tiwari, S., and Singh, S. 2015. Production and characterization of thermotolerant-organic solvent resistant acidic protease by *Pseudomonas aeruginosa* RGSS-09 isolated from dairy sludge. *Asian Journal of Biochemistry* 10 (2):52-66.

Gavira, J. A., Hernandez-Hernandez, M., Gonzalez-Ramirez, L. A., Briggs, R. A., Kole, S. A., and Shaw Stewart, P. D. 2011. Combining counter-diffusion and microseeding to increase the success rate in protein crystallization. *Crystal Growth & Design* 11 (6):2122-2126.

- George, S., Raju, V., Subramanian, T., and Jayaraman, K. 1997. Comparative study of protease production in solid substrate fermentation versus submerged fermentation. *Bioprocess Engineering* 16 (6):381-382.
- Ghorbel, B., Sellami-Kamoun, A., and Nasri, M. 2003. Stability studies of protease from *Bacillus cereus* BG1. *Enzyme and Microbial Technology* 32 (5):513-518.
- Ghosh, A., Chakrabarti, K., and Chattopadhyay, D. 2009. Cloning of feather-degrading minor extracellular protease from *Bacillus cereus* DCUW: dissection of the structural domains. *Microbiology* 155:2049–2057.
- Giberti, F., Salvalaglio, M., and Parrinello, M. 2015. Metadynamics studies of crystal nucleation. *IUCrJ* 2 (2):256-266.
- Gomes, M. T., Teixeira, R. D., Lopes, M. T., Nagem, R. A., and Salas, C. E. 2012. X-ray crystal structure of CMS1MS2: a high proteolytic activity cysteine proteinase from *Carica candamarcensis*. *Amino Acids* 43 (6):2381-2391.
- Gsponer, J., Ferrara, P., and Caflisch, A. 2001. Flexibility of the murine prion protein and its Asp178Asn mutant investigated by molecular dynamics simulations. *Journal of Molecular Graphics and Modelling* 20 (2):169-182.
- Gupta, A. and Khare, S. 2007. Enhanced production and characterization of a solvent stable protease from solvent tolerant *Pseudomonas aeruginosa* PseA. *Enzyme and Microbial Technology* 42 (1):11-16.
- Gupta, A. and Khare, S. 2009. Enzymes from solvent-tolerant microbes: useful biocatalysts for non-aqueous enzymology. *Critical Reviews in Biotechnology* 29 (1):44-54.
- Gupta, R., Beg, Q., and Lorenz, P. 2002. Bacterial alkaline proteases: molecular approaches and industrial applications. *Applied Microbiology and Biotechnology* 59 (1):15-32.
- Gurung, N., Ray, S., Bose, S., and Rai, V. 2013. A broader view: microbial enzymes and their relevance in industries, medicine, and beyond. *BioMed Research International* 2013.
- Hakansson, K., Wang, A. H.-J., and Miller, C. G. 2000. The structure of aspartyl dipeptidase reveals a unique fold with a Ser-His-Glu catalytic triad. *Proceedings of the National Academy of Sciences* 97 (26):14097-14102.
- Hameed, A., Keshavarz, T., and Evans, C. S. 1999. Effect of dissolved oxygen tension and pH on the production of extracellular protease from a new isolate of *Bacillus subtilis* K2, for use in leather processing. *Journal of Chemical Technology and Biotechnology* 74 (1):5-8.

- Hansen, C. L., Classen, S., Berger, J. M., and Stephen, R. 2006. A microfluidic device for kinetic optimization of protein crystallization and in situ structure determination. *Journal of the American Chemical Society* 128 (10):3142-3143.
- Hegyí, H. and Gerstein, M. 1999. The relationship between protein structure and function: a comprehensive survey with application to the yeast genome. *Journal of Molecular Biology* 288 (1):147-164.
- Heidari, H. R. K., Ziaee, A.-A., Amoozegar, M. A., Cheburkin, Y., and Budisa, N. 2008. Molecular cloning and sequence analysis of a novel zinc-metalloprotease gene from the *Salinivibrio* sp. strain AF-2004 and its extracellular expression in *E. coli*. *Gene* 408:196-203
- Hiroshi, T. 1993. Protein engineering on subtilisin. *International Journal of Biochemistry* 25 (3):307-312.
- Hooda, V. 2011. Physicochemical, functional and structural characterization of wheat germin using in silico methods. *Current Research Journal of Biological Sciences* 3 (1):35-41.
- Hurle, D. T. 1993. *Handbook of crystal growth*. Vol. 1. North Holland.
- Ikemura, H., Takagi, H., and Inouye, M. 1987. Requirement of pro-sequence for the production of active subtilisin E in *Escherichia coli*. *Journal of Biological Chemistry* 262 (16):7859-7864.
- Jain, S. C., Shinde, U., Li, Y., Inouye, M., and Berman, H. M. 1998. The crystal structure of an autoprocessed Ser221Cys-subtilisin E-propeptide complex at 2.0 Å resolution. *Journal of Molecular Biology* 284 (1):137-144.
- James, L. C. and Tawfik, D. S. 2003. Conformational diversity and protein evolution – a 60-year-old hypothesis revisited. *Trends in Biochemical Sciences* 28 (7):361-368.
- Jennissen, H. P. 2000. Hydrophobic interaction chromatography: the critical hydrophobicity approach. *International Journal of Biochromatography* 5:131-163.
- Joo, H., Kumar, C., Park, G., Paik, S., and Chang, C. 2003. Oxidant and SDS-stable alkaline protease from *Bacillus clausii* 1-52: Production and some properties. *Journal of Applied Microbiology* 95 (2):267-272.
- Joo, H. S. 2012. Purification and characterization of a novel alkaline protease from *Bacillus horikoshii*. *Journal of Microbiology and Biotechnology* 22 (1):58-68.
- Jørgensen, P. L., Tangney, M., Pedersen, P. E., Hastrup, S., Diderichsen, B., and Jørgensen, S. T. 2000. Cloning and sequencing of an alkaline protease gene

from *Bacillus lentus* and amplification of the gene on the *B. lentus* chromosome by an improved technique. *Applied and Environmental Microbiology* 66 (2):825-827.

Kamal, M., Hoog, J.-O., Kaiser, R., Shafqat, J., Razzaki, T., Zaidi, Z. H., and Jornvall, H. 1995. Isolation, characterization and structure of subtilisin from a thermostable *Bacillus subtilis* isolate. *FEBS Letters* 374 (3):363-366.

Kelley, L. A. and Sternberg, M. J. E. 2009. Protein structure prediction on the Web: a case study using the Phyre server. *Nature Protocols* 4 (3).

Khmelnitsky, Y. L. and Rich, J. O. 1999. Biocatalysis in nonaqueous solvents. *Current Opinion in Chemical Biology* 3 (1):47-53.

Kiesel, B. r. 2008. Book Review: Plasmids: Current Research and Future Trends. By G. Lipps (Ed.): Wiley Online Library.

Kim, S., Quigley, G., Suddath, F., McPherson, A., Sneden, D., Kim, J., Weinzierl, J., and Rich, A. 1973. X-ray crystallographic studies of polymorphic forms of yeast phenylalanine transfer RNA. *Journal of Molecular Biology* 75 (2):421-424.

Kluszens, L. D., Voorhorst, W. G., Siezen, R. J., Schwerdtfeger, R. M., Antranikian, G., van der Oost, J., and de Vos, W. M. 2002. Molecular characterization of fervidolysin, a subtilisin-like serine protease from the thermophilic bacterium *Fervidobacterium pennivorans*. *Extremophiles* 6 (3):185-194.

Krieger, E., Darden, T., Nabuurs, S. B., Finkelstein, A., and Vriend, G. 2004. Making optimal use of empirical energy functions: force-field parameterization in crystal space. *Proteins: Structure, Function, and Bioinformatics* 57 (4):678-683.

Krieger, E., Koraimann, G., and Vriend, G. 2002. Increasing the precision of comparative models with YASARA NOVA-a self-parameterizing force field. *Proteins: Structure, Function, and Bioinformatics* 47 (3):393-402.

Kroes, R. and Reiss, D. 1984. Properties of TGS aqueous solution for crystal growth. *Journal of Crystal Growth* 69 (2-3):414-420.

Kshetri, P., Ningombam, O., and Ningombam, D. 2016. Optimization of Alkaline Protease Production by Alkaliphilic *Bacillus* sp. KW2 in Low Cost Medium using Statistical Approaches. *Applied Microbiology Open Access* 2 (1000117):2.

Kulakova, L., Galkin, A., Kurihara, T., Yoshimura, T., and Esaki, N. 1999. Cold-active serine alkaline protease from the psychrotrophic bacterium *Shewanella* strain Ac10: gene cloning and enzyme purification and characterization. *Applied and Environmental Microbiology* 65 (2):611-617.

- Kumar, P. and Sharma, S. 2015. An overview of purification methods for proteins. *IJAR* 1 (12):450-459.
- Kumar, S., Stecher, G., and Tamura, K. 2016. MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*:54.
- Kumari, S. 2014. Extracellular protease enzyme production using *Micrococcus luteus*-4, *Staphylococcus hyicus*, *Micrococcus luteus*-1, *Pasteurella pneumotrop* and *Micrococcus* sp. isolated from water reservoirs. *International Journal of Current Microbiology and Applied Sciences* 3 (5):772-784.
- Kuzmanic, A. and Zagrovic, B. 2010. Determination of ensemble-average pairwise root mean-square deviation from experimental B-factors. *Biophysical Journal* 98 (5):861-871.
- Kwon, Y. T., Kim, J. O., Moon, S. Y., Yoo, Y. D., and Rho, H. M. 1995. Cloning and characterization of the gene encoding an extracellular alkaline serine protease from *Vibrio metschnikovii* strain RH530. *Gene* 152:59-63.
- Kyte, J. and Doolittle, R. F. 1982. A simple method for displaying the hydropathic character of a protein. *Journal of Molecular Biology* 157 (1):105-132.
- Laane, C., Boeren, S., Vos, K., and Veeger, C. 1987. Rules for optimization of biocatalysis in organic solvents. *Biotechnology and Bioengineering* 30 (1):81-87.
- Laemmli, U. 1970. Most commonly used discontinuous buffer system for SDS electrophoresis. *Nature* 227:680-685.
- Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227 (5259):680-685.
- Lal, R., Batra, A., Aggarwal, M., Wilcox, W., and Trolinger, J. 1991. Growth and study of triglycine sulfate (TGS) crystals in low-g for infrared detector applications.
- Laskar, A., Rodger, E. J., Chatterjee, A., and Mandal, C. 2012. Modeling and structural analysis of PA clan serine proteases. *BMC Research Notes* 5 (1):1.
- Laskowski, R. A., MacArthur, M. W., Moss, D. S., and Thornton, J. M. 1993. PROCHECK: a program to check the stereochemical quality of protein structures. *Journal of Applied Crystallography* 26 (2):283-291.
- LaVallie, E. R. 1995. Production of recombinant proteins in *Escherichia coli*. *Current Protocols in Protein Science*:1-5.

- Leach, A. R. 2001. *Molecular modelling: principles and applications*. Pearson education.
- Li, L. and Ismagilov, R. F. 2010. Protein crystallization using microfluidic technologies based on valves, droplets, and SlipChip. *Biophysics* 39.
- Li, M. and Chang, W. 2009. Protein crystallization. *Photosynthesis Research* 102 (2):223-229.
- Li, S., Yang, X., Yang, S., Zhu, M., and Wang, X. 2012. Technology prospecting on enzymes: application, marketing and engineering. *Computational and Structural Biotechnology Journal* 2 (3):1-11.
- Li, Y., Hu, Z., Jordan, F., and Inouye, M. 1995. Functional analysis of the propeptide of subtilisin E as an intramolecular chaperone for protein folding: refolding and inhibitory abilities of propeptide mutants. *Journal of Biological Chemistry* 270 (42):25127-25132.
- Liu, H. F., Ma, J., Winter, C., and Bayer, R. 2010. Recovery and purification process development for monoclonal antibody production. Paper read at MAbs.
- Liu, Q., Sun, S., Piao, M., and Yang, J. Y. 2013. Purification and Characterization of a Protease Produced by a *Planomicrobium* sp. L-2 from Gut of *Octopus vulgaris*. *Preventive Nutrition and Food Science* 18 (4):273.
- Luft, J. R., Newman, J., and Snell, E. H. 2014. Crystallization screening: the influence of history on current practice. *Acta Crystallographica Section F: Structural Biology Communications* 70 (7):835-853.
- Maciver, B., McHale, R. H., Saul, D. J., and Bergquist, P. L. 1994. Cloning and sequencing of a serine proteinase gene from a thermophilic *Bacillus* species and its expression in *Escherichia coli*. *Applied and Environmental Microbiology* 60 (11):3981.
- Majumdar, S., Goswami, S., Keppen, C., Rai, S. K., and Mukherjee, A. K. 2015. Statistical optimization for improved production of fibrin (Ogen) olytic enzyme by *Bacillus cereus* strain FF01 and assessment of in vitro thrombolytic potential of protease enzyme. *Biocatalysis and Agricultural Biotechnology* 4 (2):191-198.
- Malathu, R., Chowdhury, S., Mishra, M., Das, S., Moharana, P., Mitra, J., Mukhopadhyay, U. K., Thakur, A. R., and Chaudhuri, S. R. 2008. Characterization and wash performance analysis of microbial extracellular enzymes from East Calcutta Wetland in India. *American Journal of Applied Sciences* 5: 1650-1661.
- Malhotra, A. 2009. Tagging for protein expression. *Methods in Enzymology* 463:239-258.

- Markland, F. S. and Smith, E. L. 1971. 16 Subtilisins: Primary structure, chemical and physical properties. *The Enzymes* 3:561-608.
- Marti-Renom, M. A., Stuart, A. C., Fiser, A., Sanchez, R., Melo, F., and Šali, A. 2000. Comparative protein structure modeling of genes and genomes. *Annual Review of Biophysics and Biomolecular Structure* 29 (1):291-325.
- McLachlan, A. D. 1972. A mathematical procedure for superimposing atomic coordinates of proteins. *Acta Crystallographica Section A: Crystal Physics, Diffraction, Theoretical and General Crystallography* 28 (6):656-657.
- McPherson, A. 1999. *Crystallization of biological macromolecules*. Cold Spring Harbor Laboratory Press.
- McPherson, A. 2004. Introduction to protein crystallization. *Methods* 34 (3):254-265.
- McPherson, A. and DeLucas, L. J. 2015. Microgravity protein crystallization. *Npj Microgravity* 1:15010.
- Mian, M. 2014. Optimization of alkaline protease production by *Bacillus licheniformis* MZK05M9 in batch culture using response surface methodology. Thesis, BRAC University, Dhaka, Bangladesh.
- Mótyán, J. A., Tóth, F., and Tózsér, J. 2013. Research applications of proteolytic enzymes in molecular biology. *Biomolecules* 3 (4):923-942.
- Mullis, K. 1990. Recombinant DNA technology and molecular cloning. *Scientific American* 262:56-65.
- Musto, H., Naya, H., Zavala, A., Romero, H., Alvarez-Valin, F., and Bernardi, G. 2004. Correlations between genomic GC levels and optimal growth temperatures in prokaryotes. *FEBS Letters* 573:73-77.
- Nagasawa, K. and Ogamo, A. 1983. Hydrophobic interaction chromatography: Google Patents.
- Newman, J., Burton, D. R., Caria, S., Desbois, S., Gee, C. L., Fazio, V. J., Kvangsakul, M., Marshall, B., Mills, G., and Richter, V. 2013. Crystallization reports are the backbone of Acta Cryst. F, but do they have any spine? *Acta Crystallographica Section F: Structural Biology and Crystallization Communications* 69 (7):712-718.
- Ng, J. D., Gavira, J. A., and García-Ruíz, J. M. 2003. Protein crystallization by capillary counterdiffusion for applied crystallographic structure determination. *Journal of Structural Biology* 142 (1):218-231.
- Nguyen, M. V. C., Zhang, L., Lhomme, S., Mouz, N., Lenormand, J.-L., Lardy, B., and Morel, F. 2012. Recombinant Nox4 cytosolic domain produced by a cell

or cell-free base systems exhibits constitutive diaphorase activity. *Biochemical and Biophysical Research Communications* 419 (3):453-458.

- Ning, L., Yang, P., Wang, Y., Luo, H., Meng, K., Wu, N., Fan, Y., and Yao, B. 2008. Cloning, expression, and characterization of protease-resistant xylanase from *Streptomyces fradiae* var. k11. *Journal of Microbiology and Biotechnology* 18 (3):410-416.
- Nokihara, K. 1998. Procedures leading to primary structure determination of proteins in complex mixtures by gel electrophoresis and modern micro-scale analyses. *Analytica Chimica Acta* 372 (1):21-32.
- Ochi, T., Bolanos-Garcia, V. M., Stojanoff, V., and Moreno, A. 2009. Perspectives on protein crystallisation. *Progress in Biophysics and Molecular Biology* 101 (1):56-63.
- Ogino, H., Watanabe, F., Yamada, M., Nakagawa, S., Hirose, T., Noguchi, A., Yasuda, M., and Ishikawa, H. 1999. Purification and characterization of organic solvent-stable protease from organic solvent-tolerant *Pseudomonas aeruginosa* PST-01. *Journal of Bioscience and Bioengineering* 87 (1):61-68.
- Otálora, F., Gavira, J. A., Ng, J. D., and García-Ruiz, J. M. 2009. Counterdiffusion methods applied to protein crystallization. *Progress in Biophysics and Molecular Biology* 101 (1):26-37.
- Pack, S. P. and Yoo, Y. J. 2004. Protein thermostability: structure-based difference of amino acid between thermophilic and mesophilic proteins. *Journal of Biotechnology* 111 (3):269-277.
- Panda, M. K., Sahu, M. K., and Tayung, K. 2013. Isolation and characterization of a thermophilic *Bacillus* sp. with protease activity isolated from hot spring of Tarabalo, Odisha, India. *Iranian Journal of Microbiology* 5 (2):159.
- Pandey, A., Soccol, C., Rodriguez-Leon, J., and Nigam, P. 2001. Production of enzymes by solid-state fermentation. *Solid-State fermentation in biotechnology. Fundamentals and applications*. Asiatech Publishers, New Delhi.
- Pant, G., Prakash, A., Pavani, J., Bera, S., Deviram, G., Kumar, A., Panchpuri, M., and Prasuna, R. G. 2015. Production, optimization and partial purification of protease from *Bacillus subtilis*. *Journal of Taibah University for Science* 9 (1):50-55.
- Pantoliano, M. W., Whitlow, M., Wood, J. F., Rollence, M. L., Finzel, B. C., Gilliland, G. L., Poulos, T. L., and Bryan, P. N. 1988. The engineering of binding affinity at metal ion binding sites for the stabilization of proteins: subtilisin as a test case. *Biochemistry* 27 (22):8311-8317.

- Park, C.-S. 2012. Effect of tryptic soy broth (TSB) and Luria-Bertani (LB) medium on production of subtilisin CP-1 from *Bacillus* sp. CP-1 and characterization of subtilisin CP-1. *Journal of Life Science* 22 (6):823-827.
- Park, D. H., Lee, H. J., and Lee, E. K. 1997. Crystallization of alkaline protease as a means of purification process. *Korean Journal of Chemical Engineering* 14 (1):64-68.
- Perry, J. J. and Staley, J. 1997. Taxonomy of eubacteria and archaea. In *Microbiology: Dynamics and diversity*, pp 388-413: Fort Worth: Saunders College Publishing.
- Petrey, D. and Honig, B. 2005. Protein structure prediction: Inroads to biology. *Molecular Cell* 20 (6):811-819.
- Petsko, G. A. and Ringe, D. 2004. *Protein structure and function*. New Science Press.
- Potumarthi, R., Ch, S., and Jetty, A. 2007. Alkaline protease production by submerged fermentation in stirred tank reactor using *Bacillus licheniformis* NCIM-2042: effect of aeration and agitation regimes. *Biochemical Engineering Journal* 34 (2):185-192.
- Power, S. D., Adams, R. M., and Wells, J. A. 1986. Secretion and autoprolytic maturation of subtilisin. *Proceedings of the National Academy of Sciences* 83 (10):3096-3100.
- Prakasham, R., Rao, C. S., and Sarma, P. 2006. Green gram husk--an inexpensive substrate for alkaline protease production by *Bacillus* sp. in solid-state fermentation. *Bioresource Technology* 97 (13):1449-1454.
- Prasad, R., Abraham, T. K., and Nair, A. J. 2014. Scale up of production in a bioreactor of a halotolerant protease from moderately halophilic *Bacillus* sp. isolated from soil. *Brazilian Archives of Biology and Technology* 57 (3):448-455.
- Preston, A. 2003. Choosing a cloning vector. *E. coli Plasmid Vectors: Methods and Applications*:19-26.
- Pusey, M. L., Liu, Z.-J., Tempel, W., Praissman, J., Lin, D., Wang, B.-C., Gavira, J. A., and Ng, J. D. 2005. Life in the fast lane for protein crystallization and X-ray crystallography. *Progress in Biophysics and Molecular Biology* 88 (3):359-386.
- Queiroz, J., Tomaz, C., and Cabral, J. 2001. Hydrophobic interaction chromatography of proteins. *Journal of Biotechnology* 87 (2):143-159.
- Rahman, R. N. Z. A., Razak, C. N., Ampon, K., Basri, M., Zin, W. M., Yunus, W., and Salleh, A. B. 1994. Purification and characterization of a heat-stable

alkaline protease from *Bacillus stearothermophilus* F1. *Applied Microbiology and Biotechnology* 40 (6):822-827.

- Rai, S. K. and Mukherjee, A. K. 2010. Statistical optimization of production, purification and industrial application of a laundry detergent and organic solvent-stable subtilisin-like serine protease (Alzwiprase) from *Bacillus subtilis* DM-04. *Biochemical Engineering Journal* 48 (2):173-180.
- Rao, M. B., Tanksale, A. M., Ghatge, M. S., and Deshpande, V. V. 1998. Molecular and biotechnological aspects of microbial proteases. *Microbiology and Molecular Biology Reviews* 62 (3):597-635.
- Rasool, N., Rashid, N., Javed, M. A., and Haider, M. S. 2011. Requirement of pro-peptide in proper folding of subtilisin-like serine protease TK0076. *Pakistan Journal of Botany* 43 (4):2059-2065.
- Rathakrishnan, P. and Nagarajan, P. 2012. Optimizing factors affecting protease production by a *Bacillus cereus* using groundnut shell under solid substrate fermentation. *International Journal of Science and Technology* 1 (2):114-129.
- Rathakrishnan, P. and Nagarajan, P. 2013. Optimization of the production of protease by *bacillus cereus* with response surface methodology using groundnut shell. *International Journal of Pharmaceutical, Chemical and Biological Sciences* 3 (2):200-209.
- Raval, V. H., Pillai, S., Rawal, C. M., and Singh, S. P. 2014. Biochemical and structural characterization of a detergent-stable serine alkaline protease from seawater haloalkaliphilic bacteria. *Process Biochemistry* 49 (6):955-962.
- Rawlings, N. D. and Barrett, A. J. 1999. MEROPS: the peptidase database. *Nucleic Acids Research* 27 (1):325-331.
- Rawlings, N. D., Barrett, A. J., and Bateman, A. 2012. MEROPS: the database of proteolytic enzymes, their substrates and inhibitors. *Nucleic Acids Research* 40 (1):343-350.
- Rawlings, N. D., Morton, F. R., and Barrett, A. J. 2006. MEROPS: the peptidase database. *Nucleic Acids Research* 34 (1):270-272.
- Rayment, I. 1981. A simple method for surveying macromolecular crystallization conditions by microdialysis. *Journal of Applied Crystallography* 14 (2):153-154.
- Rayment, I. 2002. Small-scale batch crystallization of proteins revisited: an underutilized way to grow large protein crystals. *Structure* 10 (2):147-151.
- Reddy, B. V. B., Li, W. W., Shindyalov, I. N., and Bourne, P. E. 2001. Conserved key amino acid positions (CKAAPs) derived from the analysis of common

substructures in proteins. *Proteins: Structure, Function, and Bioinformatics* 42 (2):148-163.

Reddy, L., Wee, Y. J., and Ryu, H. W. 2008. Purification and characterization of an organic solvent and detergent-tolerant novel protease produced by *Bacillus* sp. RKY3. *Journal of Chemical Technology and Biotechnology* 83 (11):1526-1533.

Renge, V., Khedkar, S., and Nandurkar, N. 2012. Enzyme synthesis by fermentation method: A review. *Scientific Reviews and Chemical Communications* 2:585-590.

Rhodes, G. 2014. *Crystallography made crystal clear: a guide for users of macromolecular models*. Academic press.

Rosano, G. n. L. and Ceccarelli, E. A. 2014. Recombinant protein expression in *Escherichia coli*: advances and challenges. *Recombinant protein expression in microbial systems*:7.

Rupp, B. 2009. *Biomolecular crystallography: principles, practice, and application to structural biology*. Garland Science.

Sadava, D. E., Hillis, D. M., Heller, H. C., and Berenbaum, M. 2009. *Life: the science of biology*. Vol. 2. Macmillan.

Saitou, N. and Nei, M. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4 (4):406-425.

Salemme, F. 1972. A free interface diffusion technique for the crystallization of proteins for X-ray crystallography. *Archives of Biochemistry and Biophysics* 151 (2):533-539.

Sambrook, J., Fritsch, E. F., and Maniatus, T. 1989. *Molecular Cloning: A Laboratory Manual*. Second ed. Cold Spring Harbor Laboratory Press: New York.

Sanchez, R. and Sali, A. 1999. Comparative protein structure modeling in genomics. *Journal of Computational Physics* 151 (1):388-401.

Sanchez, S. and Demain, A. L. 2011. Enzymes and bioconversions of industrial, pharmaceutical, and biotechnological significance. *Organic Process Research & Development* 15 (1):224-230.

Saran, S., Isar, J., and Saxena, R. K. 2007. Statistical optimization of conditions for protease production from *Bacillus* sp. and its scale-up in a bioreactor. *Applied Biochemistry and Biotechnology* 141 (2-3):229-239.

- Saraswat, M., Musante, L., Ravidá, A., Shortt, B., Byrne, B., and Holthofer, H. 2013. Preparative purification of recombinant proteins: current status and future trends. *BioMed Research International* 2013.
- Sarath, G., De La Motte, R., and Wagner, F. 1989. Protease assay methods. *Proteolytic enzymes: a practical approach*. IRL Press, Oxford, United Kingdom:25–55.
- Sawant, R. and Nagendran, S. 2014. Protease: an enzyme with multiple industrial applications. *World Journal of Pharmaceutical Sciences* 3:568-579.
- Saxena, R., Sheoran, A., Giri, B., and Davidson, W. S. 2003. Purification strategies for microbial lipases. *Journal of Microbiological Methods* 52 (1):1-18.
- Schumann, W. and Ferreira, L. C. S. 2004. Production of recombinant proteins in *Escherichia coli*. *Genetics and Molecular Biology* 27 (3):442-453.
- Shi, P., Yuan, T., Zhao, J., Huang, H., Luo, H., Meng, K., Wang, Y., and Yao, B. 2011. Genetic and biochemical characterization of a protease-resistant mesophilic β -mannanase from *Streptomyces* sp. S27. *Journal of Industrial Microbiology & Biotechnology* 38 (3):451-458.
- Shinde, U. and Inouye, M. 1995. Folding pathway mediated by an intramolecular chaperone: Characterization of the structural changes in pro-subtilisin E coincident with autoprocessing. *Journal of Molecular Biology* 252 (1):25-30.
- Shinde, U. and Inouye, M. 1996. Propeptide-mediated folding in subtilisin: the intramolecular chaperone concept. In *Subtilisin Enzymes*, pp 147-154: Springer.
- Shinde, U., Liu, J., and Inouye, M. 1997. Protein memory through altered folding mediated by intramolecular chaperones. *Nature* 389 (6650):520-522.
- Siezen, R. J., de Vos, W. M., Leunissen, J. A., and Dijkstra, B. W. 1991. Homology modelling and protein engineering strategy of subtilases, the family of subtilisin-like serine proteinases. *Protein Engineering* 4 (7):719-737.
- Siezen, R. J. and Leunissen, J. A. 1997. Subtilases: the superfamily of subtilisin-like serine proteases. *Protein Science* 6 (3):501-523.
- Singh, R. K., Tiwari, M. K., Singh, R., and Lee, J.-K. 2013. From protein engineering to immobilization: promising strategies for the upgrade of industrial enzymes. *International Journal of Molecular Sciences* 14 (1):1232-1277.
- Singh, S. and Bajaj, B. K. 2016. Bioprocess optimization for production of thermoalkali-stable protease from *Bacillus subtilis* K-1 under solid-state fermentation. *Preparative Biochemistry and Biotechnology* 46 (7):717-724.

- Singh, S. K., Singh, S. K., Tripathi, V. R., Khare, S. K., and Garg, S. K. 2011. Comparative one-factor-at-a-time, response surface (statistical) and bench-scale bioreactor level optimization of thermoalkaline protease production from a psychrotrophic *Pseudomonas putida* SKG-1 isolate. *Microbial Cell Factory* 10 (1):114-127.
- Singh, S. K., Tripathi, V. R., Jain, R. K., Vikram, S., and Garg, S. K. 2010. An antibiotic, heavy metal resistant and halotolerant *Bacillus cereus* SIU1 and its thermoalkaline protease. *Microbial Cell Factories* 9 (1):1.
- Siti, S. D. and Hertadi, R. 2015. Isolation and characterization of organic-solvent stable protease isolated by *Pseudomonas stutzeri* BK AB-12. *Procedia Chemistry* 16:341-348.
- Sloma, A., Rufo, G. A., Theriault, J. K. A., Dwyer, M., Wilson, S. W., and Pero, J. 1991. Cloning and characterization of the gene for an additional extracellular serine protease of *Bacillus subtilis*. *Journal of Bacteriology* 173 (21):6889-6895.
- Smyth, D. R., Mrozkiewicz, M. K., McGrath, W. J., Listwan, P., and Kobe, B. 2003. Crystal structures of fusion proteins with large-affinity tags. *Protein Science* 12 (7):1313-1322.
- Souza, P. M. d., Bittencourt, M. L. d. A., Caprara, C. C., Freitas, M. d., Almeida, R. P. C. d., Silveira, D., Fonseca, Y. M., Ferreira Filho, E. X., Pessoa Junior, A., and Magalhães, P. O. 2015. A biotechnology perspective of fungal proteases. *Brazilian Journal of Microbiology* 46 (2):337-346.
- Spira, W. and Silverman, G. 1979. Effects of glucose, pH, and dissolved-oxygen tension on *Bacillus cereus* growth and permeability factor production in batch culture. *Applied and Environmental Microbiology* 37 (1):109-116.
- Stahl, M. L. and Ferrari, E. 1984. Replacement of the *Bacillus subtilis* subtilisin structural gene with an In vitro-derived deletion mutation. *Journal of Bacteriology* 158 (2):411-418.
- Stevens, R. C. 2000. Design of high-throughput methods of protein production for structural biology. *Structure* 8 (9):177-185.
- Subbian, E., Williamson, D. M., and Shinde, U. 2015. Protein folding mediated by an intramolecular chaperone: Energy landscape for unimolecular pro-subtilisin E maturation. *Advances in Bioscience and Biotechnology* 6 (02):73.
- Subbian, E., Yabuta, Y., and Shinde, U. P. 2005. Folding pathway mediated by an intramolecular chaperone: intrinsically unstructured propeptide modulates stochastic activation of subtilisin. *Journal of Molecular Biology* 347 (2):367-383.

- Subhabrata, S. and Mayura, D. 2006. Industrial and clinical applications excluding diagnostic clinical. *Enzymology* 1:1-25.
- Sullivan, D. C. and Kuntz, I. D. 2004. Distributions in protein conformation space: implications for structure prediction and entropy. *Biophysical Journal* 87 (1):113-120.
- Supangat, S., An, Y. J., Sun, Y., Kwon, S. T., and Cha, S. S. 2010. Purification, crystallization and preliminary crystallographic analysis of a multiple cofactor-dependent DNA ligase from *Sulfophobococcus zilligii*. *Acta Crystallographica Section F: Structural Biology and Crystallization Communications* 66 (12):1583-1585.
- Syed, R., Roja Rani, S., Masoodi, T. A., Shafi, G., and Alharbi, K. 2012. Functional analysis and structure determination of alkaline protease from *Aspergillus flavus*. *Bioinformatics* 8 (4):175-180.
- Tabassum, R., Haseeb, M., and Fazal, S. 2016. Structure prediction of outer membrane protease protein of *Salmonella typhimurium* using computational techniques. *International Journal Bioautomation* 20 (1).
- Takagi, H., Koga, M., Katsurada, S., Yabuta, Y., Shinde, U., Inouye, M., and Nakamori, S. 2001. Functional analysis of the propeptides of subtilisin E and aqualysin I as intramolecular chaperones. *FEBS Letters* 508 (2):210-214.
- Tanaka, S.-i., Saito, K., Chon, H., Matsumura, H., Koga, Y., Takano, K., and Kanaya, S. 2007. Crystal structure of unautoprocessed precursor of subtilisin from a hyperthermophilic archaeon. *Journal of Biological Chemistry* 282.
- Tanaka, S.-i., Takeuchi, Y., Matsumura, H., Koga, Y., Takano, K., and Kanaya, S. 2008. Crystal structure of Tk-subtilisin folded without propeptide: requirement of propeptide for acceleration of folding. *FEBS Letters* 582 (28):3875-3878.
- Tari, C., Genckal, H., and Tokatlı, F. 2006. Optimization of a growth medium using a statistical approach for the production of an alkaline protease from a newly isolated *Bacillus* sp. L21. *Process Biochemistry* 41 (3):659-665.
- Thompson, J. D., Higgins, D. G., and Gibson, T. J. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22 (22):4673-4680.
- Ting, Y. T., Harris, P. W., Batot, G., Brimble, M. A., Baker, E. N., and Young, P. G. 2016. Peptide binding to a bacterial signal peptidase visualized by peptide tethering and carrier-driven crystallization. *International Union of Crystallography* 3 (1).

- Tinoi, J., Rakariyatham, N., and Deming, R. 2005. Simplex optimization of carotenoid production by *Rhodotorula glutinis* using hydrolyzed mung bean waste flour as substrate. *Process Biochemistry* 40 (7):2551-2557.
- Topf, M., Baker, M. L., Marti-Renom, M. A., Chiu, W., and Sali, A. 2006. Refinement of protein structures by iterative comparative modeling and CryoEM density fitting. *Journal of Molecular Biology* 357 (5):1655-1668.
- Tress, M. 2002. Protein tertiary structures: Prediction from amino acid sequences. *eLS*.
- Underkofler, L., Barton, R., and Rennert, S. 1958. Production of microbial enzymes and their applications. *Applied Microbiology* 6 (3):212.
- Valls, C., Pujadas, G., Garcia-Vallve, S., and Mulero, M. 2011. Characterization of the protease activity of detergents laboratory practicals for studying the protease profile and activity of various commercial detergents. *Biochemistry and Molecular Biology Education* 39 (4):280-290.
- Vecerek, B. and Kyslik, P. 1995. Cloning and sequencing of the neutral protease-encoding gene from a thermophilic strain of *Bacillus* sp. *Gene* 158 (1):147-148.
- Velooralappil, N. J., Robinson, B. S., Selvanesan, P., Sasidharan, S., Kizhakkepawothail, N. U., Sreedharan, S., Prakasan, P., Moolakkariyil, S. J., and Sailas, B. 2013. Versatility of microbial proteases. *Advances in Enzyme Research* 1 (3):39-51.
- Verma, A. and Wenzel, W. 2009. A free-energy approach for all-atom protein simulation. *Biophysical Journal* 96 (9):3483-3494.
- Vijayaraghavan, P., Lazarus, S., and Vincent, S. G. P. 2014. De-hairing protease production by an isolated *Bacillus cereus* strain AT under solid-state fermentation using cow dung: Biosynthesis and properties. *Saudi Journal of Biological Sciences* 21 (1):27-34.
- Vishwanatha, K., Rao, A. A., and Singh, S. A. 2010. Acid protease production by solid-state fermentation using *Aspergillus oryzae* MTCC 5341: optimization of process parameters. *Journal of Industrial Microbiology & Biotechnology* 37 (2):129-138.
- Wakayama, N., Yin, D., Harata, K., Kiyoshi, T., Fujiwara, M., and Tanimoto, Y. 2006. Macromolecular crystallization in microgravity generated by a superconducting magnet. *Annals of the New York Academy of Sciences* 1077 (1):184-193.
- Walter, T. S., Mancini, E. J., Kadlec, J., Graham, S. C., Assenberg, R., Ren, J., Sainsbury, S., Owens, R. J., Stuart, D. I., and Grimes, J. M. 2008. Semi-automated microseeding of nanolitre crystallization experiments. *Acta*

- Wang, F., Hao, J., Yang, C., and Sun, M. 2010. Cloning, expression, and Identification of a novel extracellular cold-adapted alkaline protease gene of the marine bacterium strain YS-80-122. *Applied Biochemistry and Biotechnology* 162 (5):1497-1505.
- Wang, S.-L., Yang, C.-H., Liang, T.-W., and Yen, Y.-H. 2008. Optimization of conditions for protease production by *Chryseobacterium taeanense* TKU001. *Bioresource Technology* 99 (9):3700-3707.
- Wang, X., Zhao, H., Liu, G., Cheng, X., and Feng, H. 2016. Improving production of extracellular proteases by random mutagenesis and biochemical characterization of a serine protease in *Bacillus subtilis* S1-4. *Genetics and Molecular Research: GMR* 15 (2).
- Watson, J. D. and Crick, F. H. 1953. Molecular structure of nucleic acids. *Nature* 171 (4356):737-738.
- Watson, S. J. 2009. Molecular dynamics simulations of HIV-1 protease complexed with saquinavir. PhD. Thesis. Thesis, UCL (University College London).
- Welch, A. R., McNally, L. M., Hall, M., and Gibson, W. 1993. Herpesvirus proteinase: site-directed mutagenesis used to study maturational, release, and inactivation cleavage sites of precursor and to identify a possible catalytic site serine and histidine. *Journal of Virology* 67 (12):7360-7372.
- Weng, M., Deng, X., Bao, W., Zhu, L., Wu, J., Cai, Y., Jia, Y., Zheng, Z., and Zou, G. 2015. Improving the activity of the subtilisin nattokinase by site-directed mutagenesis and molecular dynamics simulation. *Biochemical and Biophysical Research Communications* 465 (3):580-586.
- Westhead, D. R., Parish, J. H., and Twyman, R. M. 2002. *Bioinformatics*. BIOS Oxford.
- Whon, T. W., Lee, Y. H., An, D. S., Song, H. K., and Kim, S. G. 2009. A simple technique to convert sitting-drop vapour diffusion into hanging-drop vapour diffusion by solidifying the reservoir solution with agarose. *Journal of Applied Crystallography* 42 (5):975-976.
- Williamson, D. M., Elferich, J., Ramakrishnan, P., Thomas, G., and Shinde, U. 2013. The mechanism by which a propeptide-encoded pH sensor regulates spatiotemporal activation of furin. *Journal of Biological Chemistry* 288 (26):19154-19165.
- Wlodawer, A., Minor, W., Dauter, Z., and Jaskolski, M. 2008. Protein crystallography for non-crystallographers, or how to get the best (but not more) from published macromolecular structures. *Febs Journal* 275 (1):1-21.

- Wong, S. L. and Doi, R. 1986. Determination of the signal peptidase cleavage site in the preprosubtilisin of *Bacillus subtilis*. *Journal of Biological Chemistry* 261 (22):10176.
- Wu, H., Zhang, Z., Hu, S., and Yu, J. 2012. On the molecular mechanism of GC content variation among eubacterial genomes. *Biology Direct* 7 (1):1.
- Wu, J., Bian, Y., Tang, B., Chen, X., Shen, P., and Peng, Z. 2004. Cloning and analysis of WF146 protease, a novel thermophilic subtilisin-like protease with four inserted surface loops. *FEMS Microbiology Letters* 230 (2):251-258.
- Xiang, Z. 2006 Advances in homology protein structure modeling. *Current Protein and Peptide Science* 7 (3):217-227.
- Xu, C. P., Sinha, J., Bae, J. T., Kim, S. W., and Yun, J. W. 2006. Optimization of physical parameters for exo-biopolymer production in submerged mycelial cultures of two entomopathogenic fungi *Paecilomyces japonica* and *Paecilomyces tenuipes*. *Letters in Applied Microbiology* 42 (5):501-506.
- Yabuta, Y., Subbian, E., Takagi, H., Shinde, U., and Inouye, M. 2002. Folding pathway mediated by an intramolecular chaperone: dissecting conformational changes coincident with autoprocessing and the role of Ca²⁺ in subtilisin maturation. *Journal of Biochemistry* 131 (1):31-37.
- Yabuta, Y., Takagi, H., Inouye, M., and Shinde, U. 2001. Folding Pathway Mediated by an Intramolecular Chaperone: Propeptide release modulates activation precision of pro-subtilisin. *Journal of Biological Chemistry* 276 (48):44427-44434.
- Yamagata, H., Masuzawa, T., Nagaoka, Y., Ohnishi, T., and Iwasaki, T. 1994. Cucumisin, a serine protease from melon fruits, shares structural homology with subtilisin and is generated from a large precursor. *Journal of Biological Chemistry* 269 (52):32725-32731.
- Yamagata, Y., Sato, T., Hanzawa, S., and Ichishima, E. 1995. The structure of subtilisin ALP I from alkalophilic *Bacillus* sp. NKS-21. *Current Microbiology* 30 (4):201-209.
- Yang, L., Dordick, J. S., and Garde, S. 2004. Hydration of enzyme in nonaqueous media is consistent with solvent dependence of its activity. *Biophysical Journal* 87 (2):812-821.
- Yao, H., Kristensen, D. M., Mihalek, I., Sowa, M. E., Shaw, C., Kimmel, M., Kavrakli, L., and Lichtarge, O. 2003. An accurate, sensitive, and scalable method to identify functional sites in protein structures. *Journal of Molecular Biology* 326 (1):255-261.

- Yin, L.-J., Lin, H.-H., and Jiang, S.-T. 2010. Bioproperties of potent nattokinase from *Bacillus subtilis* YJ1. *Journal of Agricultural and Food Chemistry* 58 (9):5737-5742.
- Ying, X. G., Shuo, C. S., and Zheng, L. X. 2011. Cloning and heterologous expression of pro-2127, a gene encoding cold-active protease from *Pseudoalteromonas* sp. QI-1. *Advances in Polar Science* 22 (2):124-130.
- Yousef, G. M., Kopolovic, A. D., Elliott, M. B., and Diamandis, E. P. 2003. Genomic overview of serine proteases. *Biochemical and Biophysical Research Communications* 305 (1):28-36.
- Zaghloul, T. I., Hendawy, H. M., Assar, S. E., and Mostafa, M. H. 2002. Enhanced stability of the cloned *Bacillus subtilis* alkaline protease gene in alginate-immobilized *B. subtilis* cells. *Enzyme and Microbial Technology* 30 (862-866).
- Zagrovic, B. and van Gunsteren, W. F. 2007. Computational analysis of the mechanism and thermodynamics of inhibition of phosphodiesterase 5A by synthetic ligands. *Journal of chemical theory and computation* 3 (1):301-311.
- Zaki, M. J. and Bystroff, C. 2008. *Protein structure prediction*. second ed. Humana Press Inc.: New York.
- Zhang, Y. 2008. TASSER server for protein 3D structure prediction. *BMC Bioinformatics* 9 (40).
- Zheng, B., Roach, L. S., and Ismagilov, R. F. 2003. Screening of protein crystallization conditions on a microfluidic chip using nanoliter-size droplets. *Journal of the American Chemical Society* 125 (37):11170-11171.
- Zheng, Z.-l., Zuo, Z.-y., Liu, Z.-g., Tsai, K.-c., Liu, A.-f., and Zou, G.-l. 2005. Construction of a 3D model of nattokinase, a novel fibrinolytic enzyme from *Bacillus natto*: a novel nucleophilic catalytic mechanism for nattokinase. *Journal of Molecular Graphics and Modelling* 23 (4):373-380.
- Zhu, X., Ohta, Y., Jordan, F., and Inouye, M. 1989. Pro-sequence of subtilisin can guide the refolding of denatured subtilisin in an intermolecular process. *Nature* 339:483 - 484.
- Zou, T. 2014. Protein folding and dynamics using multi-scale computational methods. PhD. Thesis. Thesis, Arizona State University.
- Zuckerandl, E. and Pauling, L. 1965. Evolutionary divergence and convergence in proteins. *Evolving genes and proteins* 97:97-166.