



UNIVERSITI PUTRA MALAYSIA

***THE EFFECT OF UPMB10 AND PSEUDOMONAS SPP. ON PADDY
GROWTH IN DIFFERENT DENSITY***

ABDULLAH BIN SUWARDI

FP 2014 2

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**FACULTY OF AGRICULTURE
UNIVERSITI PUTRA MALAYSIA
SERDANG, SELANGOR DARUL EHSAN**

2013/2014

THE EFFECT OF UPMB10 AND *PSEUDOMONAS* SPP. ON PADDY GROWTH
IN DIFFERENT DENSITY

BY

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A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia, in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science

Faculty of Agriculture

Universiti Putra Malaysia

2013/2014

ENDORSEMENT

This project report entitled “**The effect of UPMB10 and *Pseudomonas* spp. on paddy growth in different density**” is prepared by Abdullah bin Suwardi and submitted to the Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of degree of Bachelor of Agricultural Science.

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Dedicated to My Parents

Hj. Suwardi bin Bugi & Hjh. Masniah binti Katile

Support-Encouragement-Constant Love

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ACKNOWLEDGMENT

Assalamualaikum W.B.T.,

In the name of Allah S.W.T., The Most Gracious, The Most Merciful. Thanksgiving to the Illahi for His grace I can completed this final year projects was successfully.

My sincere appreciation and gratitude to my supervisor, Assoc. Prof. Dr. Mohd Halimi Mohd Saud for his guidance, advice and help throughout this project.

Also thanks to the lecturers and staffs Department of Agriculture Technology, Department of Crop Science and Department of Land Management for their commitment to provide instructing. Also not be forgotten to my friends who are helping and providing moral support to me

Lastly, special thanks to my parent, Hj Suwardi and Hjh Masniah who is my soul and passion to complete this project.

Thank you. Wassalam.

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ABSTRACT

Oryza sativa or rice plant is a type of cereal crops and one of the most popular crops in worldwide. Malaysia with a population of 28 million people is not able to supply sufficient amount of rice and had to import it. This is due to the less area of cultivation. In production systems, farmers are using chemical fertilizer as the method to increase the plant growth. In long time effect, this method can affect to soil fertility, environment and to human beings. One of the cultivation techniques in increasing rice plant growth and yield are by implying several ways in maximising use of land and in the same time, it will contribute to sustainable agriculture. The objectives for this study are to determine the effect of the planting density and plant growth promoting rhizobacteria (PGPR) to increase rice plant growth. The project was carried out in a glasshouse using polybag with different seedling numbers (1 seedling per hill, 2 seedlings per hill and 3 seedlings per hill) and 5 ml (1×10^8 CFU/ml) different PGPR per plant (UPMB10, *Pseudomonas* spp. and UPMB10 mix with *Pseudomonas* spp.) will be inoculated into the soil three days and 31 days after transplanting (DAT). Each treatment with five replications will be arranged in Randomized Complete Block Design (RCBD) and data collected analysis with Analysis of Variance (ANOVA). The effect of the planting density and PGPR can be seen through a number of tiller, plant height, dry weight, root morphology and contain of nutrient after 45 days.

ABSTRAK

Oryza sativa atau tanaman padi adalah sejenis tanaman bijirin dan salah satu tanaman yang paling popular di seluruh pelusuk dunia. Malaysia dengan jumlah penduduk 28 juta orang tidak dapat membekalkan jumlah beras yang mencukupi dan terpaksa mengimportnya. Ini adalah kerana kawasan penanaman adalah kurang. Dalam sistem pengeluaran, petani menggunakan baja kimia sebagai kaedah untuk meningkatkan kadar pertumbuhan tanaman. Hakikatnya pada masa panjang, kaedah ini boleh memberi kesan kepada kesuburan tanah, alam sekitar dan manusia. Salah satu teknik penanaman dalam meningkatkan pertumbuhan tanaman padi dan hasil adalah dengan melaksanakan beberapa cara dalam memaksimumkan penggunaan tanah dan dalam masa yang sama, ia akan menyumbang kepada pertanian lestari. Objektif bagi kajian ini adalah untuk menentukan kesan kepadatan penanaman dan rizobakteria penggalak pertumbuhan tanaman (PGPR) untuk meningkatkan pertumbuhan pokok padi. Projek ini telah dijalankan di dalam rumah kaca menggunakan pasu dengan bilangan anak benih yang berlainan (1 anak benih setiap lubang, 2 anak benih setiap lubang dan 3 anak pokok setiap lubang) dan 5 ml (1×10^8 CFU/ml) PGPR berbeza setiap tumbuhan (UPMB10, *Pseudomonas* spp. dan UPMB10 bercampur dengan *Pseudomonas* spp.) akan disuntik ke dalam tanah selepas tiga hari dan 31 hari selepas menanam (DAT). Setiap rawatan dengan lima replikasi akan disusun dalam Rekabentuk Rawak Berblok (RCBD) dan data analisis Analisis Varians (ANOVA). Kesan kepadatan tanaman dan PGPR dapat dilihat melalui bilangan sulur, ketinggian pokok, berat kering, morfologi akar dan kandungan nutrisi selepas 45 hari.

CHAPTER 1

INTRODUCTION

Rice (*Oryza sativa*) is one of the food groups to be a very high demand from all over the world as well as corn and wheat. In order to enhance production to satisfy consumers' needs, many transformations had been made such as development of variety, double cropping systems and high fertilizer rate usage, especially chemical fertilizers. The use of chemical fertilizers at the maximum rate admitted supplying sufficient nutrients to crops with a fast pace. However, the continuing provision to be felt in a long time on the quality of soil and the environment.

Malaysian did not have a large of crop acreage for planting of rice for fulfil local consumption and had to import rice from neighbouring countries such as Thailand and Myanmar because suitability land for rice planting either from soil type or fertility is less.

To overcome the shortage of the crop land and excessive use of chemical fertilizer there are two suggestions to increase the density of rice plants per hole and use of plant growth-promoting rhizobacteria (PGPR).

An amount of land suitable for rice cultivation is less and the usual rice cultivation system practiced by farmers in Malaysia is one seedling per hole does not guarantee the high production. This project will focus on a system where more than one seedlings hole. This is intended to maximize the usage of seedlings per hectare, simultaneously improve the quality and yield of rice.

Besides that, a plant growth-promoting rhizobacteria (PGPR) is one of biological alternatives to increase the rate of crop growth without given negative impact. Growth of a crop failure can be caused by lack of nutrients and nutrient formed are not readily used. Bacteria used in this project are nitrogen fixation bacteria (NFC) and phosphate solubilizing bacteria (PSB). These bacteria performance to provide nutrients in a suitable form and can be used by plants.

Thus, this project will be conducted to achieve this objective:

- a. To observe the effect of difference bacteria to paddy growth
- b. To determine the effect of difference density (number seedling per hill) to paddy growth
- c. To determine the interaction between different bacteria and density to paddy growth

REFERENCES

- Alias, I. (2002). MR 219, a new high-yielding rice variety with yields of more than 10 mt/ha. MARDI, Malaysia. FFTC: *An international information center for small scale farmers in the Asian and Pacific region*. 12-18.
- Amir, H.G., Z.H. Shamsuddin, M.S. Halimi, M.F. Ramlan and M. Marziah. (2003). Potential use of rhizobacteria for sustainable oil palm seedling production. *Malaysian Journal of Soil Science*, 3, 39-50.
- Agriculture Department of Statistic. (2012). Paddy statistics of Malaysia. www.doa.gov.my.
- Baset Mia, M. B., Shamsuddin, Z. H., & Mahmood, M. (2012). Effects of rhizobia and plant growth promoting bacteria inoculation on germination and seedling vigor of lowland rice. *African Journal of Biotechnology*, 11(16), 3758-3765.
- Biswas, J.C., J.K. Ladha and F.B. Dazzo. (2002). Rhizobia inoculation improves nutrient uptake and growth of lowland rice. *Soil Science Soc. Am. J.*, 164:1644-1650.
- Choudhury, A.T.M.A. and Y.M. Khanif, (2001). Evaluation of the Effects of Nitrogen and Magnesium Fertilization on Rice Yield and Fertilizer Nitrogen Efficiency using ¹⁵N Tracer Technique. *Journal Plant Nutrition, Soil*, 108: 281-285. 24: 855-871.

Choudhury, A.T.M.A. and I.R. Kennedy, (2005). Nitrogen Fertilizer Losses from Rice Soils and Control of Environmental Pollution Problems. *Communications in Soil Science and Plant Analysis*, 36: 1625-1639

De Freitas, J. R., Banerjee, M. R., & . (1997). Phosphate-solubilizing rhizobacteria enhance the growth and yield but not phosphorous uptake of canola (*Brassica napus* L.) *Biology and Fertility of Soils*, 24, 358-364.

De Salamone, I. E. G., Hynes, R. K., & Nelson, L. M. (2001). Cytokinin production by plant growth promoting rhizobacteria and selected mutants. *Canada Journal Microbial.*, 47, 404-411.

Desrochers, A., S.M. Landhausser and V.J. Lieffers. (2002). Coarse and fine root respiration in aspen (*Populus tremuloides*). *Tree Physiol.* 22:725–732.

Development of Agriculture Organization Board, Pahang (LKPP).(2013).Varities of paddy. <http://www.lkppadi.com.my/>.

Evans L.T., Wardlaw I.F., Williams C.N. (1964). Environmental control of growth. *In: Barnard C(Ed) Grasses and Grassland, Macimillan and Co. Ltd. London* .102-105.

FAO (Food and Agriculture Organization). (2012). Top Production of Rice. www.fao.org/.

Faruque, G.M., (1996). Effect of fertilizer nitrogen and seedling(s) per hill on the growth and yield of long grain rice. *M.S. Thesis, Department of Agronomy, IPISA, Joydevpur, Gajipur, Bangladesh, 143, 67-73.*

Gyaneshwar, P., Kumar, G.N., Parekh, L.J., & Poole, P.S. (2002). Role of soil microorganisms in improving P nutrition of plants. *Plant and Soil, 245, 83-93.*

Heulin, T., Guckert, A. & Balandreau, J. (1987). Stimulation of root exudation of rice seedlings by azospirillum strains: carbon budget under gnotobiotic conditions. *Biology Fertility Soil, 4, 9-14.*

Integrated taxonomic information system. (2013). *oryza sativa. L.*
<http://www.itis.gov/>.

Kim, K.Y., Jordan, D., & Krishnasn, H.B. (1998). Expression of genes from *Rahnella aquatilis* that are necessary for mineral phosphate solubilizing in *Escherichia coli*. *FEMS Microbiology Letters, 159, 121-127*

Mahamud J.A, M.M. Haque and M. Hasanuzzaman.(2013). Growth, dry matter production and yield performance of transplanted Aman rice varieties influenced by seedling densities per hill. *International journal of sustainable agriculture, 5(1), 16-24*

Maziah, M., A.R. Zuraida, M.S. Halimi, Shamsuddin, Z.H. and Sreeramanan. (2009). Effectiveness of PGPR inoculation in the presence of nitrogen and carbon

sources on growth of *in vitro* banana plantlets. *Advances in Environmental Biology*, 3(2): 129-143.

Mia MAB, Shamsuddin ZH, Zakaria W, Marziah M (2007). Associative nitrogen fixation by *Azospirillum* and *Bacillus* spp. in bananas. Infomusa, *African Journal of Biotechnology*, 16 : 11-15.

Mia MAB, Shamsuddin ZH, , Marziah M. (2012). Effects of rhizobia and plant growth promoting bacteria inoculation on germination and seedling vigor of lowland rice. *African Journal of Biotechnology*, 11 (16), 3758-3765.

Miah, M.N.H., S. Talukder, M.A.R. Sarkar and T.H. Ansari, (2004). Effect of number of seedling per hill and urea supergranules on growth and yield of the rice cv. *BINA Dhan 4*. *J. Biol. Sci.*, 4(2): 122-129.

Mirza Hasanuzzaman, M.L. Rahman, T.S. Roy, J.U. Ahmed and A.S.M. Zobaer. (2009). Plant characteristic, yield components, and yield of late transplanted aman rice as affected by plant spacing and number of seedling per hill. *Advance in Biological Research*, 3(5-6), 201-207.

Nautiyal, C. S., Bhadauria, S., Kumar, P., Lal, H., Mondal, R., & Verma, D. (2000). Stress-induced phosphate solubilization in bacteria isolated from alkaline soils. *FEMS Microbiology Letters*, 182, 291-296.

Pregitzer, K.S., M.J. Laskowski, A.J. Burton, V.C. Lessard and D.R. Zak. (1998).

Variation in sugar maple root respiration with root diameter and soil depth. *Tree Physiol.* 18:665–670.

Ramezanpour, M.R. (2010). Genetic Diversity and Efficiency of Indole Acetic Acid

Production by the Isolates of *Pseudomonads fluorescent* from Rhizosphere of Rice (*Oryza sativa L.*). *American- Eurasian Journal Agriculture & Environment Science.*, 7(1): 103-109.

Rangarajan, S., L.M Saleena and S. Nair. (2002). Diversity of *Pseudomonas* spp.

isolated from rice rhizosphere populations grown along a salinity gradient. *Microbial Ecology*, 43, 280-9.

Rodriguez, H., R. Fraga, T. Gonzalez, and Y. Bashan. (2009). Genetics of phosphate

solubilization and its potential applications for improving plant-growth-promoting bacteria. *Plant Soil* 287, 15-21.

Rovira, A.D. (1969). Interaction between plants roots and soil microorganisms. *Ann.*

Rev. Microbial., 19, 241-266.

Sahrawat, K.L. (2000). Macro and Micronutrients Removed by Upland and Lowland

Rice Cultivars in West Africa. *Commun. Soil Science Plant Analysis.*, 31: 717-723.

Sanico A.L., S. Peng, R.C. Laza, & R.M. Visperas. (2002). Effect of seedling age and seedling number per hill on snail damage in irrigated rice. *Crop Protection*, 21, 137–143.

Silvia G.A and Jorge A.T. (2010). Tillering regulation by endogenous and environmental factors and its agricultural management. *The Americas Journal of Plant Science and Biotechnology*. 35-48.

Singh, H.P. (1990). Effect of nursery seedling density and seedlings per hill on late transplanted rice. *Intl. Rice Res. Newsl.*, 15(4): 20.

Stevenson and Cole, M. A. (1999). *Cycles of soil :Carbon nitrogen, phosphorus, sulfur, micronutrients*, 2nd Edition. New York:Wiley.

Vessey J.K. (2003). Plant growth promoting rhizobacteria as biofertilizers. *Plant and Soil*, 255: 571-586.

Volder, A., D.R. Smart, A.J. Bloom and D.M. Eissenstat. (2005). Rapid decline in nitrate uptake and respiration with age in fine lateral roots of grape: implications for root efficiency and competitive effectiveness. *New Phytol.* 165:493–501

Volkmar & Breme. (1998). Effect of seed inoculation with a strain of *Pseudomonas fluorescens* on root growth and activity of wheat in well watered and drought-stressed glass-fronted rhizotrons. *Canadian Journal of Plant Sciences*, 78, 545-551.

Zakry, A.A., A.B. Khairuddin, S. Zulkifli and A.B.Rahim. (2012). Contribution of nitrogen from biofertilizer inoculum to young oil palm under field condition. *International Symposium on Managing Soils for Food Security and Climate Change Adaptation and Mitigation Vienna, Austria.*

Zhao, Wu, C.S., Meng, C.F., Lu, X.N., Teng, C.Q. (2002). The agronomic availability and economic effect of applying phosphate rock to acid paddy soil. *Phosphate Comp. Fertil.*, 17(3):67–69.