

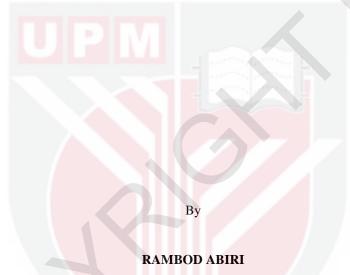
## EMBRYOGENESIS PROTOCOL FOR GENETIC TRANSFORMATION AND FUNCTIONAL ANALYSIS OF JERF1 GENE IN DROUGHT-INDUCED MALAYSIAN RICE CULTIVAR MR219

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FBSB 2018 60



#### EMBRYOGENESIS PROTOCOL FOR GENETIC TRANSFORMATION AND FUNCTIONAL ANALYSIS OF *JERF1* GENE IN DROUGHT-INDUCED MALAYSIAN RICE CULTIVAR MR219



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

September 2018

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#### DEDICATION

"I don't know what your destiny will be, but one thing I know; the only ones among you who will be really happy are those who have sought and have found how to serve." -Albert Schweitzer

This humble work is dedicated to:

My beloved wife, Narges, for your patience, love, friendship and humor, without

which I wouldn't have reached this present stage.

My parents who have supported me through all my various endeavors,

#### Louis and MohammadReza.

My parents in law who have prayed for me, Forouz and Homayoun.

My brother, **<u>Ramin</u>**, and my brother in law, <u>Ali</u> who have encouraged inspired me.

My sisters in law, Jila and Armaghan,

My nephew, Shervin

Thank you for understanding that distance can help one improve his knowledge, even though 6,615 km is a long way from home.

I decided to devote my life to telling the truth, because having survived I owe something to the dead and anyone who does not remember them betrays them, my dearest brothers, <u>Navid</u> and <u>Amir</u>,

who I have lost them, may their souls continue to rest in perfect peace.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

#### EMBRYOGENESIS PROTOCOL FOR GENETIC TRANSFORMATION AND FUNCTIONAL ANALYSIS OF *JERF1* GENE IN DROUGHT-INDUCED MALAYSIAN RICE CULTIVAR MR219

By

#### RAMBOD ABIRI

September 2018

Chair: Noor Azmi Shaharuddin, PhD Faculty: Biotechnology and Biomolecular Sciences

In Malaysia, severe drought stresses have affected residents and agricultural crops, especially rice, in various regions of the country and it is likely to continue and may worsen in the future. To mitigate the problems, new tolerant plant varieties to drought must be developed. The main objective of the current research was to develop a suitable in vitro protocol and elucidate the response of Malaysian rice cultivar MR219 to Jasmonate and Ethylene Response Factor 1 (JERF1) gene in drought condition. The best recipe of callogenesis for MR219 was MS media added with 2 mg L<sup>-1</sup> of 2,4-D and root was the prominent explant. The best priming conditions were seen at 18 hours of hydropriming and 50 mg L<sup>-1</sup> of abscisic acid. For the proliferation phase, the highest efficiency was observed at week four in the MS media supplemented with 2 mg L<sup>-1</sup> of 2,4-D, 2 mg L<sup>-1</sup> of kinetin, 50 mg L<sup>-1</sup> of L-proline, 100 mg L<sup>-1</sup> of casein and 30 mg L<sup>-1</sup> of silicon. MS media with 3 mg  $L^{-1}$  of KIN, 30 mg  $L^{-1}$  of silicon and 2 mg  $L^{-1}$  of NAA was selected as the best media for regeneration. To promote the roots of the regenerated explants, 0.4 mg L<sup>-1</sup> of IBA was found to be the best activator. TRIzol protocol was found to be an appropriate method for isolating high quality RNA from the tomato leaves and fruits. Four hours of NaCl and ABA treatments on the tomato prior to the nucleic acids extractions produced the highest JERF1 expression and led to the successfully isolating the JERF1 gene. The entry and the expression vectors were constructed using Gateway® Technology. Agrobacterium-mediated transformation method was used to transform JERF1 gene to MR219. Transgenicity of the transformed plants was evaluated using polymerase chain reaction, q-PCR and High-Performance Liquid Chromatography. The result of functional study has shown that the average width of the wild type seeds was significantly higher than transgenic seeds. Meanwhile, the seeds ratio of wild type was higher in comparison to transgenic in the second generation. The morpho- and physiological results of the two-week-old rice seedlings had shown that the responses of both wild type and transgenic rice in terms of shoot and root length and plumule fresh and dry weight were significantly different ( $p \le 0.01$ ). The responses of both wild type and transgenic rice in terms of the shoot and root length,

leaf proline, root proline, chlorophyll a and b, total chlorophyll and carotenoid and other pigments were significantly different ( $p \le 0.01$ ) in three-week-old rice. These results have confirmed the significant differences ( $p \le 0.05$ ) between wild type and transgenic plants in terms of the ratio of root to shoot proline in three-week-old rice. The concentrations of some amino acids such as aspartic acid, serine, glycine, proline and cysteine were significantly different between wild type and transgenic plants. Real-time PCR confirmed the over-expression of *OsLTP1*, *OsCDPK13*, *OsP5CS* and *OsSPDS2* by *JERF1* genes in transgenic plants. To conclude, the results of this experiment confirmed the high efficiently of somatic embryogenesis protocol and potential of *JERF1* as a prominent gene under drought stress condition.



Abstrak tesis yang dikemukakan kepada Senate Universiti Putra Malaysia sebagai memeuhi keperluan untak ijazah Doktor Falsafah

#### PROTOCOL EMBRIOGENESIS UNTUK ANALISIS TRANSFORMASI GENETIK DAN FUNGSI GEN *JERF1* DI DALAM PADI MALAYSIA KULTIVAR MR219 TERARUH-KEMARAU

Oleh

#### **RAMBOD ABIRI**

September 2018

Pengerusi: Noor Azmi Shaharuddin, PhD Fakulti: Bioteknologi dan Sains Biomolekul

Di Malaysia, kemarau yang teruk telah menjejaskan kehidupan dan tanaman pertanian, terutamanya padi, di seluruh negara ini dan ja mungkin akan berterusan dan boleh menjadi lebih buruk pada masa akan datang. Untuk mengatasi masalah ini, variati tumbuhan toleran yang rintang kemarau mesti dibangunkan. Objektif utama penyelidikan ini adalah untuk membangunkan protokol *in vitro* dan mengkaji tindak balas kultivar padi MR219 kepada gen Jasmonate dan Ethylene Response Factor 1 (JERF1) dalam keadaan kemarau. Resipi yang terbaik untuk proses kallogenesis untuk MR219 adalah media MS yang ditambah dengan 2 mg L<sup>-1</sup> 2,4-D dengan akar sebagai explan yang paling sesuai. Kondisi proses pemula yang terbaik ialah pada 18 jam pemula-hidro di dalam 50 mg  $L^{-1}$  asid absisik. Untuk fasa proliferasi, keeffisyenan tertinggi didapati pada minggu keempat dalam media MS yang ditambah dengan 2 mg L<sup>-1</sup> 2,4-D, 2 mg L<sup>-1</sup> kinetin, 50 mg L<sup>-1</sup> L-prolin, 100 mg L<sup>-1</sup> kasein dan 30 mg L<sup>-1</sup> silikon. Media MS dengan 3 mg L<sup>-1</sup> KIN, 30 mg L<sup>-1</sup> silikon dan 2 mg L<sup>-1</sup> NAA dipilih sebagai media terbaik untuk proses percambahan. Untuk menggalakkan pertumbuhan akar pada eksplan, 0.4 mg L<sup>-1</sup> IBA didapati sebagai pengaktif terbaik. Kaedah TRIzol merupakan kaedah yang sesuai untuk memencilkan RNA berkualiti tinggi dari daun dan buah tomato. Rawatan NaCl dan ABA pada tomato selama empat jam sebelum pengekstrakan asid nukleik, telah menghasilkan ekspresi JERF1 tertinggi dan menyumbang kepada kejayaan pemencilan gen JERF1. Vektor ungkapan dibina dengan menggunakan teknologi Gateway®. Kaedah transformasi agrobakterium telah digunakan untuk memindahkan gen JERF1 kepada MR219. Transgenisiti pada padi transgenik dinilai menggunakan tindak balas rantaian polimerase, q-PCR dan Kromatografi Cecair Berprestasi Tinggi. Hasil kajian fungsional telah menunjukkan bahawa lebar purata biji padi jenis liar adalah jauh lebih tinggi daripada biji transgenik. Sementara itu, nisbah benih liar lebih tinggi berbanding dengan transgenik pada generasi kedua. Hasil morfologi dan fisiologi dari anak benih padi berusia dua minggu telah menunjukkan perbezaan yang sinifikan (p≤0.01) pada tindakbalas padi liar dan padi transgenik dari segi pucuk dan panjang akar dan plumul berat segar dan kering.

Respons terhadap kedua-dua jenis liar dan padi transgenik dari segi pucuk dan panjang akar, proline daun, proline akar, klorofil a dan b, jumlah klorofil dan karotenoid dan pigmen lain adalah sangat berbeza ( $p\leq0.01$ ) dalam padi yang berusia tiga minggu. Hasil ini telah mengesahkan perbezaan ketara ( $p\leq0.05$ ) antara padi jenis liar dan padi transgenik dari segi nisbah prolin pada akar-pucuk dalam padi berusia tiga minggu. Kepekatan beberapa asid amino seperti asid aspartik, serin, glisin, prolin dan sistin sangat berbeza antara padi liar dan padi transgenik. Analisis qRT-PCR telah mengesahkan pengekspresan lampau gen-gen *OsLTP1, OsCDPK13, OsP5CS* dan *OsSPDS2* oleh gen *JERF1* dalam padi transgenik. Kesimpulannya, hasil eksperimen ini telah berjaya menghasilkan protokol embriogenesis somatik yg berkeeffisyenan tinggi dan mengesahkan potensi *JERF1* sebagai gen yang utama dalam keadaan kemarau.



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This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## TABLE OF CONTENTS

		TABLE OF CONTENTS	
			Page
AB	STRA	АСТ	i
AB	STRA	Κ	iii
AC	KNO	WLEDGEMENT	v
	PRO		vi
		RATION	viii
		TABLES	xiv
		TABLES TFIGURES	
			XV1
LR	ST OF	ABBREVIATIONS	XX
CH 1	IAPTI INT	ER P P P P P P P P P P P P P P P P P P P	1
2	LIT	ERATURE REVIEW	
	2.1	Rice	3
		2.1.1 Rice yield in the world	3
	2.2	Climate changes and drought occurrence in Malaysia	5 6
		<ul><li>2.2.1 Rice production in Malaysia</li><li>2.2.2 Physiological, morphological and agronomical characteristics</li></ul>	0 10
		of rice cultivar MR219	10
	2.3	Plants mechanisms under abiotic stress	10
	2.4		11
	2.5		13
	2.6	APETALA 2/ethylene-response factor, an influential superfamily in plants	13
	2.7	Plant improvement by utilizing genetic engineering strategies	15
		2.7.1 Gene transformation strategies	15
		2.7.2 Plant transformation platforms	17
		2.7.2.1 Stable nuclear expression	17
		2.7.2.2 Transient expression	18
		2.7.2.3 Plastid expression	18
		<ul><li>2.7.2.4 Plant cell-suspension cultures</li><li>2.7.3 Factors influencing the gene transformation in Indica rice hosts</li></ul>	19 19
		2.7.5 Factors influencing the gene transformation in indica fice hosts 2.7.4 Enhancing somatic embryogenesis and plant regeneration	20
	2.8	Tomato a dicotyledons model plant in responses to abiotic stress	20 21
	2.9	Gene expression analysis	22
		Gene recombination simplifies cloning workflow	23
		Functional study of putative rice	23
3	MA	TERIALS AND METHODS	
	3.1	Plant materials and culture conditions	24
	3.2	Callus frequency	24
		3.2.1 Optimization of auxins application on callus frequency	24
		3.2.2 Optimizations of different hydro- and hormonal-priming factors	24

 $\bigcirc$ 

	on explants productions	
	3.2.3 Effect of different explants on callus frequency	26
3.3	Callus growth and proliferation	26
	3.3.1 Effect of various cytokinin concentrations on callus growth	27
	3.3.2 Effect of various adjuvant materials and concentrations on	27
	callus growth	
	3.3.3 Localization of potassium metasilicate in treated samples	27
3.4	Plant regeneration	28
	3.4.1 Effect of various cytokinin concentrations on plant regeneration	28
	3.4.2 Effect of various auxin concentrations on plant regeneration	28
	3.4.3 Effect of various adjuvant material concentrations on plant	28
	regeneration	
3.5	Regenerated plantlets' protection	28
3.6	Statistical analysis of <i>in vitro</i> tissue culture	29
3.7	Optimization of RNA extraction protocols	29
	3.7.1 RNA extraction with modified TRIzol method	29
	3.7.2 Cetyltrimethylammonium bromide (RNA extraction buffer 1;	30
	CTAB)	
	3.7.3 Phenol Chloroform (RNA extraction buffer 2)	30
	3.7.4 RNeasy plant mini kit	30
	3.7.5 NucleoSpin® RNA plant kit	31
3.8	Purification of total RNA using DNase treatment and removal	31
	reagents	
3.9	Isolation of total RNA from leaf of tomato plants by most accurate	31
	protocol	
	3.9.1 Nucleic acid integrity analysis	32
	3.9.2 Statistical analysis of total RNA extraction protocols	32
3.10	JERF1 quality and integrity	32
	3.10.1 Real Time-PCR	32
	3.10.2 Isolation and purification full length of <i>JERF1</i>	33
3.11	Gene transformation procedure	35
	3.11.1 Construction of recombinant plasmids	35
	3.11.2 Identification of recombinant plasmids	35
	3.11.2.1 Plasmid purification	35
	3.11.2.2 Digestion of recombinant plasmids	36
	3.11.3 Expression vector construction	36
	3.11.3.1 Entry clones construction	36
	attB-PCR products producing	36
	BP recombination reaction performing	37
	Competent cell protocol of <i>E. coli</i> (TOP 10)	38
	Transforming competent cells	38
	3.11.3.2 Verification of the recombinant entry vector	38
	3.11.4 Construction of expression clones	39 39
	3.11.4.1 Performing LR recombination reaction	- 39 - 39
	3.11.4.2 Verifying the recombinant expression clones	
	Confirmation recombinant expression clones	39
	by colony PCR and sequencing Purification and sequencing of plasmid	40
	3.11.5 Transferring the construct into <i>Agrobacterium tumefaciens</i>	
	strain	40
	3.11.6 <i>Agrobacterium</i> strain LBA 4404 and selectable markers	40
	5.11.0 Agrobucierium suam LDA 4404 and sciectable markers	40

		4.1.3.3	Effects of various adjuvant material concentrations on	65
			plant regeneration	
	4.1.4	Regener	rated plantlets	68
4.2	Evalu	ation of l	RNA extraction methods in tomato, gene isolation and	70
	expre	ssion ana	lysis of JERF1 and vector construction	
	4.2.1	Compai	rison of different total RNA extraction methods for	70
		tomato	leaves and fruits	
	4.2.2	Total R	NA extraction of tomato leaves and fruits using TRIzol	74
	4.2.3	Gene ex	pression detection of <i>JERF1</i> gene by real-time PCR	76
	4.2.4	Isolatio	n of a full-length coding region for JERF1 gene	79
4.3	Evalu	ation of t	the putative transgenic rice plants	81
	4.3.1	Functio	nal analysis of transgenic rice in T <sub>2</sub> generation	86
		4.3.1.1	Grain characteristics of wild type seeds of MR219 and	86
			transferred seeds	
		4.3.1.2	Effect of drought stress treatment on the physiological	87
			and morphological plant growth parameters in the	
			seedling phase	
		4.3.1.3	Amino acid evaluation of transgenic rice	93
		4.3.1.4	Analysis of various genes over-expression by JERF1	95
			in transgenic plants by real-time PCR	
	4.3.2	In silico	analysis of JERF1 protein	96

# 5 SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH

REFERENCES	102
APPENDICES	128
BIODATA OF STUDENT	140
LIST OF PUBLICATIONS	141

 $\mathbf{G}$ 

## LIST OF TABLES

Table		Page	
2.1	The scientific classification of rice (O. sativa)	3 5	
2.2	Impact of adverse conditions on rice production in Southeast Asia	5	
2.3	Paddy rice area and yield in Malaysia (2015/2016)	6	
2.4	Different paddy rice cultivars released by MARDI in Malaysia	8	
2.5	Agronomic characteristics, physio-chemical properties, pest and diseases tolerant of MR219	10	
2.6	Drought tolerance genes transformed to rice	15	
3.1	Different concentrations of hormonal priming	26	
3.2	KAPA SYBR® qPCR Master Mix Kit	33	
3.3	The sequences of <i>TUB</i> and <i>EF1</i> primers references genes and <i>JERF1</i>	33	
3.4	The sequences of <i>JERF1</i> forward and reverse primers for RT-PCR	34	
3.5	The sequences of forward and reverse adapter primers	37	
3.6	List of primers for <i>JERF1</i> , <i>CamV35S</i> and <i>OsActin</i> used to confirm transgenicity of MR219	43	
3.7	Amino acid concentration of standard for HPLC procedure	46	
3.8	The sequences of OsLTP1, 18S rRNA, OsCDPK13, a-Tubulin, OsP5CS and OsSPDS2 primers references genes and JERF1 primers	47	
4.1	for real- time PCR ANOVA of hydro- priming effects on germination and growth of MR219 rice cultivar	53	
4.2	MR219 fice cultivar Mean comparison of total germination, 50% of germination, mean germination time, germination percentage and germination vigour of	54	
	MR219 under different hydro- priming time condition		
4.3	ANOVA of hormonal-priming effect on germination and growth of	54	
	MR219 rice cultivar	-	
4.4	Mean comparison of total germination, 50% of germination, mean germination time, germination percentage and germination vigour index of MR219 under different hormonal- priming conditions	55	
4.5	The summarised results of tissue culture sections	70	
4.6	ANOVA of purity of concentration of total RNA extracted from	70	
1.0	tomato leaves and fruits using five various protocols	12	
4.7	The average yield of extracted total RNA from tomato leaves and fruits using various extraction protocols	72	
4.8	ANOVA of purity and concentrations of total RNA extracted from tomato leaves and fruits of contaminated samples by SDS and CTAB	73	
4.9	methods ANOVA of purity and concentrations of total RNA extracted from tomato leaves and fruits in different ages phases	75	
4.10	The average of yield and integrity of extracted total RNA from tomato leaves and fruits using TRIzol protocols	76	
4.11	Relative expression patterns and P-value of <i>JERF1</i> , <i>Tubulin</i> and <i>EF1</i> reference genes under various treatments and time in tomato leaves and fruits	78	
4.12	ANOVA results of seeds characters evaluation between wild type seeds and $T_2$ seeds of MR219	86	
4.13	Analysis of variance of the seeds shape variables between wild type	87	

#### seeds and transgenic seeds of MR219

- 4.14 Analysis of variance of the shoot and root length, plumule fresh and dry weight as well as radical fresh and dry weight of wild type and transgenic plants in week two under drought stress condition
- 4.15 Analysis of variance of the shoot and root length, leaf proline, root proline, ratio of root to shoot proline, chlorophyll a and b, total chlorophyll as well as carotenoid and other pigments of wild type and transgenic plants in week three under drought stress conditions
- 4.16 Analysis of amino acid profile of transgenic and wild-type MR219



88

90

 $\int$ 

## LIST OF FIGURES

Figure		Page	
2.1	Global paddy rice area and production (FAO, 2015)	4	
2.2	Paddy rice cultivation fields in Peninsular Malaysia. The eight main	7	
	areas of paddy rice cultivation are presented in yellow boxes (Figure		
2.3	is duplicated from DOA, 2014) Schematic figure of plants signal transduction under stress	11	
2.3	AP2/ERF super family members in different plant species	14	
2.5	Different monocotyledon and dicotyledon plants were used in gene	16	
	transformation by Agrobacterium tumefaciens		
3.1	pDrive cloning vector plasmid map (Qiagen, Germany)	35	
3.2	pDONOR/Zeo vector (Invitrogen, CA, USA) plasmid map	37	
3.3 4.1	Plasmid map of pFAST-G02 vector Effect of different auxins on the seed callus frequency, days to callus	39 48	
4.1	induction and rooty or shooty callus induction percentage. Highest	40	
	callus frequency were observed in the MS media supplemented with		
	2,4-d, whereas other auxins did not induce high percentage callus.		
	Means followed by the same letter are not significantly different		
	based on analysis of variance at the 0.01 level		
4.2	Callogenesis of MR219 in various auxins treatments, a) control	49	
	(MSO), b) 2,4-D, c) NAA, d) picloram, e) dicamba and f) 2,4,5-T		
	after one month. 2,4-D produced calli without shooty or rooty		
	organelle. Other auxins produced calli with some organelles structures		
4.3	Effect of different 2,4-D concentrations on the seed callus frequency,	51	
1.5	days to callus induction and dead seed or no callus induction	51	
4.4	Callogenesis of MR219 in MS media supplemented with 2 mg L <sup>-1</sup> of	52	
	2,4-D, a) embryogenic callus, b) non-embryogenic callus and c) dead		
	callus. Embryogenic callus are nodular and yellowish callus which		
	can proliferate and regenerate in the suitable media		
4.5	Effect of various explants on the callus frequency, days to callus	57	
	induction and dead explants or not callus induction. The callus frequency in root explants were higher than other explants		
4.6	Effect of various cytokinines on the callus growth, a) fresh and, b)	58	
1.0	dry weight. The highest fresh and dry weight were seen in week 4 for	20	
	KIN		
4.7	Callus proliferation steps in, a) week four and, b) week seven in MS	59	
	media supplemented with 2 mg $L^{-1}$ of 2,4-D and 2 mg $L^{-1}$ of KIN		
4.8	Callus fresh and dry weight in week four for different concentrations	60	
	of, a) proline and casein hydrolysate and, b) potassium metasilicate (alliant) in MS models and provide a state $f_{2}$ and $f_{3}$ and $f_{4}$ and $f_{3}$ and $f_{4}$ and		
	(silicon) in MS media supplemented with 2 mg $L^{-1}$ of 2,4-D and 2 mg $L^{-1}$ of KIN. The highest fresh and dry weight calli were observed in		
	the 50 mg $L^{-1}$ L- proline, 100 mg $L^{-1}$ Casein hydroysate and 30 mg L-		
	1 of silicon		
4.9	Scanning electron microscopy image showing the calli in, a) control	61	
	and b) 30 mg L <sup>-1</sup> of silicon (white mass). The white layer of silicon		
	were seen in the calli treated with potassium metasilicate (B)		
4.10	Effect of different cytokinin concentrations on the plant regeneration.	62	
	The highest regeneration percentage were seen in the 3 mg L <sup>-1</sup> of KIN		

- 4.11 Various steps in regeneration of rice calli in medium, a) embryogenesis calli, b) initial green spot, c) lustrous green spot calli with sickle- shaped trichomes, d) regenerated calli, e) shoot induction and f) dead calli in the same medium. The initial step of micropropagation is embryogenesis callus induction by using relevant PGRs. Next, calli should be canalised to the producing green spot and sickle shaped trichomes. Then using suitable cytokinin helps to regenerate calli, producing shoot and root. During this procedure, non- embryogenesis calli is dead due un-specified structure in the callus induction
- 4.12 Effect of different auxins concentrations on the plant regeneration
- 4.13 Effect of different, a) L- proline, casamino acids and b) silicon concentrations on the plant regeneration. The highest regeneration percentage were reported in MS media supplemented with 100 mg L<sup>-1</sup> of L-proline and 30 mg L<sup>-1</sup> of silicon
- Scanning electron microscopy image showing the root, a) control (no 4.14 silicon) and b) 30 mg  $L^{-1}$  of silicon. The microscopy results showed the white silicon spot on the root
- Effect of different IAA, IBA and ABA concentrations (0.1, 0.2, 0.3) 4.15 68 and 0.4 mg L<sup>-1</sup>) on the plant regeneration. The highest regeneration percentage were seen in the MS media added with 0.4 mg L<sup>-1</sup> of IBA
- 4.16 Isolation of total RNA and gDNA extracted from tomato leaves and fruits. The extracted RNA of tomato leaves with A.1) TRIzol, A.2) CTAB 1, A.3) SDS, A.4) RNeasy plant mini kit and A.5) NucleoSpin® RNA plant kit and RNA of tomato fruits with B.1) TRIzol, B.2) CTAB 1, B.3) SDS, B.4) RNeasy plant mini kit and B.5) NucleoSpin<sup>®</sup> RNA plant kit using 1.5% agarose gel and stained with ethidium bromide
- 4.17 Various steps in regeneration of rice calli in medium, a) 74 embryogenesis calli, b) initial green spot, c) lustrous green spot calli with sickle- shaped trichomes, d) regenerated calli, e) shoot induction and f) dead calli in the same medium. The initial step of micropropagation is embryogenesis callus induction by using relevant PGRs. Next, calli should be canalised to the producing green spot and sickle shaped trichomes. Then using suitable cytokinin helps to regenerate calli, producing shoot and root. During this procedure, non- embryogenesis calli is dead due un-specified structure in the callus induction
- 4.18 Total RNA extraction by TRIzol in various phases of tomato. Leaves 75 samples including vegetative growth phase (1-month-old plant) (1), flowering phase (3-month-old plant) (2) and fruiting phase (4-monthold plant) (3) as well as breakers- stage 2 (4), light red- stage 5 (5) and red- stage 6 (5) (6) using 1.5% agarose gel and stained with ethidium bromide
- 4.19 Melting curve of Tubulin (A), EF1 (B) and JERF1 (C) for standard 77 curve dilutions of cDNA template. The melting temperature of 58 °C was obtained
- Relative abundance of JERF1 gene in tomato fruits under (A) ABA 4.20 78 and (B) NaCl treatments
- 4.21 Relative abundance of JERF1 gene in tomato fruits under (A) ABA 78 and (B) NaCl treatments

xvii

63

64 65

67

- 4.22 JERF1 transcript, A) isolated from tomato (Lycopersicon esculentum Mill. cv. MT1) in 58 °C of annealing temperature and B) purified DNA with 1118 bp with use of 1.5% agarose gel and stained with ethidium bromide
- 4.23 JERF1 amplified by site-specific primers flanked to the attb sequences by gradient PCR
- The JERF1 gene existence in the BP recombination reaction 4.24 confirmed by colony PCR
- 4.25 The JERF1 gene existence in the LR recombination reaction 81 confirmed by colony PCR
- 4.26 *JERF1* gene in the *Agrobacterium tumefaciens* colonies using colony 81 PCR
- The gene transformation process of MR219 by JERF1 gene. A) The 4.27 seeds of rice were germinated in the B) petri dishes under different hydro- and hormonal-priming for 9 days. Then the C) root explants of germinated seeds were cultured on the best in vitro media with the effective hormones and adjuvant materials. D) The most embryogenesis calli which induced from roots E) proliferated in the most effective media formulation. Between weeks three and forth which the calli had the highest fresh and dry weight, calli were inoculated in the Agrobacterium tumefaciens containing the plasmid. Then the calli were co-cultured in the selection media containing cefotoxime and herbicide phosphinothricin (BASTA). F) Due to the lack of bar gen in the non- transferred calli, just G) the transferred calli with bar gen which is the prove of JERF1 gen existence can continue to H) regenerate. I) After regeneration of rice in the media the regenerated plants transferred to different media and some abnormality like J) colorless plants were observed then, K) plants in the media were acclimatized. The putative rice then L, M) adjusted with the green house temperature transferred to Yoshida media in the transgenic green house. After adoption to the new condition, the rice plants N, O) transferred into the soil and kept in the P) green house for future functional experiments
- 4.28 Analysis of *bar* gene (1168 bp) in  $T_0$  generation of transgenic rice 4.29
- Analysis of *JERF1* gene (1118 bp) in  $T_0$  generation of transgenic rice 4.30 Analysis of OsActin gene (445 bp) in T<sub>0</sub> generation of transgenic rice
- 83 4.31 Analysis of *CaMV35S* gene (1353 bp) in T<sub>0</sub> generation of transgenic 83 rice.
- 4.32 Analysis of *bar* gene (1168 bp) in  $T_1$  generation of transgenic rice 84 84
- 4.33 Analysis of *JERF1* gene (1118 bp) in T<sub>1</sub> generation of transgenic rice 4.34 Analysis of OsActin gene (445 bp) in T<sub>1</sub> generation of transgenic rice
- 84 Analysis of CaMV35S gene (1353 bp) in  $T_1$  generation of transgenic 4.35 84 rice
- 4.36 Analysis of JERF1 gene (1118 bp) in T<sub>2</sub> generation of transgenic rice 85 seed 85
- Analysis of *bar* gene (1168 bp) in T<sub>2</sub> generation of transgenic rice 4.37
- Analysis of CaMV35S gene (1353 bp) in T<sub>2</sub> generation of transgenic 4.38 85 rice
- 4.39 Analysis of OsActin gene (445 bp) in T<sub>2</sub> generation of transgenic rice 85 4.40 The observation of *gfp* flanked expression to *JERF1* gen in the  $T_2$ 85

(Mature rice seeds of MR219)

80

80

80

82

83

- 4.41 The observation of A) transgenic seeds and B) wild type seeds in  $T_2$ generation. The observation showed that the transgenic seeds were shorter than wild type seeds
- 4.42 Comparisons of (A) plumule fresh and dry weight as well as radical fresh and dry weight and (B) roots and shoots lengths of wild type and transgenic rice under normal and drought stress conditions in week two. The results of plumule fresh and dry weight showed the highest rate in the control treatment of transgenic and wild type plants. The amount of plumule fresh and dry weight was higher in wild type plants than transgenic plants. In the radical fresh and dry weight, the highest and lowest amounts were observed in control treatment of transgenic plants and transgenic plants under drought treatment. The shoot and root length measurement results showed that highest amount of shoot length was seen in wild type plants in treatment condition and wild type plants under drought treatment. Means followed by the same letter are not significantly different based on analysis of variance at the 0.01 level. A) The wild type and B) transgenic rice under drought stress condition and C) the wild type and D) transgenic rice under normal condition
- 4.43 Comparisons of shoot and root length, leaf proline, root proline, ratio of root to shoot proline, chlorophyll a and b, total proline as well as carotenoid and other pigments of wild type and transgenic plants in week three under normal and drought stress conditions. The figure showed the fluctuation trends of all the characteristics for wild and transgenic plants under drought and control condition. Means followed by the same letter are not significantly different based on analysis of variance at the 0.01 level. A) The wild type and B) transgenic rice under normal condition and C) the wild type and D) transgenic rice under drought stress condition
- 4.44 The analysis of amino acids by HPLC chromatograms of wild- type 94 plant
- 4.45 The analysis of amino acids by HPLC chromatograms of transgenic 94 plant
- Analysis of OsLTP1, OsCDPK13, OsP5CS and OsSPDS2 genes 4.46 95 over- expression in wild type and transgenic plants under normal and drought stress conditions
- Analysis of hydrophilicity as well as hydrophobicity for the encoded 4.47 96 protein by JERF1
- 4.48 The amino acids compositions analysis of the protein encoded by 97 JERF1 predicted by ProtScale (http://web.expasy.org/protscale/) in the tool kit of ExPASy
- Secondary structure of JERF1 predicted by PsiPred 97 4.49 98
- Predicted I-TASSER results of normalized B-factor 4.50
- The 3D structure of protein encoded by JERF1 gene predicted by 99 4.51 ITASSER as well as Phyre2 programs

89

87

## LIST OF ABBREVIATIONS

A. tumefaciens	Agrobacterium tumefaciens
aa	Amino acid
AA	Ascorbic acid
ABA	Abscisic Acid
Ala	Alanine
AP2/ERF	APETALA 2/ethylene-responsive
Arg	Arginine
Asp	Aspartic acid
B.C.	Before Christ
BAP	(benzylamino) purine
Вр	Base Pairs
Ca <sup>2+</sup>	Calcium ions
CaMV35s	Cauliflower mosaic virus
cDNA	Complementary DNA
Ch.a	chlorophyll a
Ch.b	chlorophyll b
СТАВ	Hexacetyltrimethyl ammonium bromide
DEPC	Diethyl pyrocarbonate
Dicamba	3,6-dichloro-o-anisic acid
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
dNTPS	deoxynucleotides
DRE/CRT	Dehydration-Responsive Clement/C-Repeat
Ds	Double-stranded
ER	Endoplasmic reticulum
ERF	Ethylene-responsive
ET	Ethylene
EtBr	Ethidium bromide
FAO	Food and Agriculture Organization
G	Gram
g (rcf)	Gravity
G%	The germination percentage
GA3	Gibberellic acid
GFP	Green fluorescent protein
GI	Gibberellins
Glu	Glutamic acid

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	Gly	Jlycine
		Genetic modification
	GVI G	ermination Vigor Index.
	GVI	The germination vigor index
	Н	Hour
	HCl	Hydrochloric acid
	His	Histidine
	HPLC	High performance liquid chromatography
	IAA	Indole-3-acetic acid
	IBA	Abscisic acid
	Ile	Isoleucine
	JA	Jasmonic acid
	JERF1	Jasmonate and Ethylene Response Factor 1
	K <sub>2</sub> SiO <sub>3</sub>	Potassium metasilicate
	Kb	Kilo base-pair
	KIN	Kinetin
	L	Litre
	LB	Luria-bertani
	LB	Lysogeny Broth
	Leu	Leucine
	LiCl	Lithium chloride
	LP	Proline of root
	Lys	Lysine
	M	Molar
	MARDI	Malaysian Agricultural Research and Development Institute
	Met	Methionine
	Mg	Milligram
	mg g <sup>-1</sup>	Milligram per gram
	MGT	Mean germination time
	MGT	Mean Germination Time,
	Min	Minute
	μg	Microgram
	μg μL <sup>-1</sup>	Microgram per microliter
$(\mathbf{G})$	μL	Microliter
	mL	Millilitre
	mM	Millimolar
	MR219	MARDI 219

xxi

	mRNA	Messenger RNA
	MS	Murashige and Skoog
	MSO	MS media free hormone
	NAA	1-Naphthaleneacetic acid
	NaCl	Sodium chloride
	NaCl	Sodium chloride
	NCBI	National Centre for Biotechnology Information
	Ng	Nanogram
	NOA	Naphthoxyacetic acid
	ORF	Open reading frame
	p-CPA	Para-chlorophenoxyacetic acid
	PCR	Polymerase chain reactions
	PEG	polyethylene glycol
	%	Percentage
	PGRs	Plant Growth Regulators
	Phe	Phenylalanine
	Picloram	4-amino-3, 5,6- tricholoropyridinecarboxylic
		acid
	PR	Pathogenesis-related
	Pro	Proline
	PVP	Polyvinylpyrrolidone
	PVPP	Polyvinyl polypyrolidone
	RNA	Ribonucleic acid
	RNase	Ribonuclease
	ROS	Reactive Oxygen Species
	RP	Proline of root
	RP/SP	Proline root to shoot ratio
	RT	Room Temperature
	RT-PCR	Reverse transcriptase polymerase chain reaction
	SA	Salicylic Acid
	SDS	Sodium dodecyl sulphate
	Sec	Second
$(\mathbf{G})$	Ser	Serine
	Spp	Species
	SS	Single-stranded
	T.G	Total Germination
	T50	50 Percentage of Germination,
	150	so reconcise of Germination,

xxii

T50	Time to 50% germination
TAE	Tris acetate EDTA
TBE	Tris borate EDTA
T-DNA	Transferred DNA
TDZ	Thidiazuron
TE	Tris-EDTA
Thr	Threonine
Total chi	Total chlorophyll.
Tyr	Tyrosine
Val	Valine
X-Gal	5-bromo-4-chloro-3-indolyl-β-D-
	galactopyranoside
zeatin	6-(4-hydroxy-3- methyl but-2-enyl amino)- purine)]

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#### **CHAPTER 1**

#### INTRODUCTION

After wheat, rice (*Oryza sativa* L.) is the second most important plant in the world (Al-Amin and Ahmed, 2016). Fluctuation and scarcity of agricultural products lead to spike in prices of rice in recent years (Tadasse *et al.*, 2016; Torero, 2016). In this regard, environmental stresses have been identified as the top reasons for the price spikes by affecting on both food security and livelihoods (De La Fuente *et al*, 2013). Despite the increasing demands for rice products, this plant is exposed to the wide spectrum of environmental stresses that negatively effects on its growth, function and productivity (Bajaj and Mohanty, 2005). Drought and water deficit have been extensively reported as the most hazardous stresses to rice (Wang *et al.*, 2012).

Drought tolerance mechanism is observed in all crops but its extent varies from one species to another species and within species (Jaleel et al., 2009). Tolerance to drought is a complex mechanism, due to interactions between different undesirable factors and various molecular, biochemical and physiological phenomena affecting plant growth as well as development (Razmjoo et al., 2008). Drought defense mechanisms in plants can be classified into four types, including drought -recovery, -escape, -tolerance and avoidance. Different physiological indicators have been applied to evaluate the tolerance of plants under drought conditions such as plant growth regulators contents, amino acid level, water potential, cell membrane stability, leaf and root traits as well as adjustment capabilities. During the years, researches have assessed the molecular and genetic mechanisms of drought resistance plants to obtain the drought-related genes with regard to drought- tolerance and -avoidance. With increasing knowledge of drought resistance mechanisms in model plants, it still a matter of concern to improve drought- resistance and water-saving (Fang and Xiong, 2015; Zheng et al., 2009). A better conceptual understanding of the plant mechanisms could be used to create or select resistance plant varieties, which may obtain a more tolerant plant under drought stress (Jaleel et al., 2009). Under drought stress and water deficit, complex signaling networks have been elaborated in rice. These networks perceives the stress signals and modulates the resistance genes expression (Xiong et al., 2002). Rice reaction to drought differs considerably at diverse organ levels depending upon stress duration and intensity as well as crop growth stage and species (Jaleel et al., 2009).

Genetic modification (GM) is a popular method that researchers have been using to increase the yield of plant products by improving certain traits, including the responses of plants to abiotic and biotic stresses (Ashraf *et al.*, 2008). Success in genetic engineering process depends upon selecting the proper method of transformation to introduce desirable traits into the host genome while concentrating on preserving the individual characteristics of the plant (Abiri *et al.*, 2015). In gene transformation processes, the gene(s) of interest of donor plants, bacteria or viruses are transferred to host plants using various methods (Rivera *et al.*, 2012). In rice, the engineered cultivars were produced through poly ethylene glycol, particle bombardment and electroporation during the 1980s- 1990s (Birch and Franks, 1991). However, the advantages of

Agrobacterium-mediated transformation encouraged researchers to use in the rice genetic transformation (Hiei et al., 1994).

The most important gene family involves in stress responses consists of transcription factors (sequence-specific DNA-binding factors) that play vital roles in influencing or controlling several biological processes. Among the stress- responsive transcription factor-encoding genes, the APETALA 2/ethylene- responsive (AP2/ERF) family genes have been described as the main stress- responsive genes in various physiological networks in rice. *Jasmonate and Ethylene Response Factor 1 (JERF1)* is a tomato protein containing a conserved ERF DNA- binding motif. The *JERF1* overexpression enhanced the tobacco resistance to low temperature, salt concentration and osmotic stress (Zhang *et al.*, 2004). The general objective of the current research was to develop a suitable *in vitro* protocol as well as to elucidate the response of Malaysian rice cultivar MR219 to *JERF1* gene in drought-induced condition. Therefore, the specific objectives of this study were:

- 1. To investigate the effect of different priming factors in germination and establish the most suitable media formulations *in vitro* condition.
- 2. To isolate the drought stress tolerance gene (*JERF1*) from the most suitable explants of Malaysian tomato (*Solanum lycopersicum*) cultivar MT1 and construct the expression vector.
- 3. To transfer the *JERF1* gene to MR219, determine the function of *JERF1* in MR219 rice cultivar and evaluate *JERF1* by computational analysis.

#### REFERENCES

- Abbasi F, Onodera H, Toki S, Tanaka H, Komatsu S (2004) *OsCDPK13*, a calciumdependent protein kinase gene from rice, is induced in response to cold and gibberellin in rice leaf sheath. *Plant Mol Biol* 55:541–552.
- Abiri, R., Zebarjadi, A., Ghobadi, A., Kafashi, A. K., Atabaki, N. (2012). Determination of Advanced Drought Tolerant and Breeder Lines in *Hordeum vulgare* L. under Kermanshsh conditions. *Iranian Journal of field crop science*, 43(1), 175-188.
- Abiri, R., Valdiani, A., Maziah, M., Shaharuddin, N. A., Sahebi, M., Yusof, Z., et al. (2015). A critical review of the concept of transgenic plants: insights into pharmaceutical biotechnology and molecular farming. Current Issues in Molecular Biology, 18, 21-42.
- Abiri, R., Shaharuddin, N. A., Maziah, M., Yusof, Z., Atabaki, N., Sahebi, M., Azizi, P. (2016). Quantitative assessment of indica rice germination to hydropriming, hormonal priming and polyethylene glycol priming. *Chilean journal of* agricultural research, 76, 392-400.
- Abiri, R., Zebarjadi, A., Ghobadi. (2016). Investigation of drought tolerance of barley genotypes during seedling stage using polyethylene glycol. 29(2), 253-265.
- Abiri, R., Maziah, M., N. A., Maziah, M., Yusof, Z., Atabaki, N., et al. (2017a). Enhancing somatic embryogenesis of Malaysian rice cultivar MR219 using adjuvant materials in a high-efficiency protocol. International Journal of Environmental Science and Technology.1-18.
- Abiri, R., Maziah, M., N. A., Maziah, M., Yusof, Z., Atabaki, N., et al. (2017b). Role of ethylene and the APETALA 2/ethylene response factor superfamily in rice under various abiotic and biotic stress conditions. *Environmental and experimental* botany, 134, 33-44.
- Abro, S. A., Qureshi, R., Soomro, F. M., Mirbahar, A. A., & Jakhar, G. (2009). Effects of silicon levels on growth and yield of wheat in silty loam soil. *Pakistan Journal of Botany*, *41*(3), 1385-1390.
- Adams-Phillips, L., Barry, C., & Giovannoni, J. (2004). Signal transduction systems regulating fruit ripening. *Trends in Plant Science*, 9(7), 331-338.
- Agarwal, P. K., Agarwal, P., Reddy, M., & Sopory, S. K. (2006). Role of DREB transcription factors in abiotic and biotic stress tolerance in plants. *Plant Cell Reports*, 25(12), 1263-1274.
- Aggarwal, D., Kumar, A., Sharma, J., & Reddy, M. S. (2012). Factors affecting micropropagation and acclimatization of an elite clone of *Eucalyptus tereticornis* Sm. In Vitro Cellular & Developmental Biology-Plant, 48(5), 521-529.
- Aguilar-Benítez, G., Peña-Valdivia, C. B., Vega, J. R., Castro-Rivera, R., & Ramírez-Tobías, H. M. (2014). Seed Germination and Early Root Growth in Common

Bean and Maize Landraces and Improved Cultivars at Different Water Stress Levels. *International Journal of Applied*, 4(4).

- Ahmad, S., & Hashim, N. (2010). Perubahan iklim mikro di Malaysia. Penerbit Fakulti Sains Sosial dan Kemanusiaan Universiti Kebangsaan Malaysia, Bangi.
- Al-Amin, A. Q., & Ahmed, F. (2016). Food Security Challenge of Climate Change: An Analysis for Policy Selection. *Futures*.
- Alam, M., Tanaka, T., Nakamura, H., Ichikawa, H., Kobayashi, K., Yaeno, T., et al. (2015). Overexpression of a rice heme activator protein gene (OsHAP2E) confers resistance to pathogens, salinity and drought, and increases photosynthesis and tiller number. Plant Biotechnology Journal, 13(1), 85-96.
- Alexander, L., & Grierson, D. (2002). Ethylene biosynthesis and action in tomato: a model for climacteric fruit ripening. *Journal of Experimental Botany*, 53(377), 2039-2055.
- Alker, A. P., Mwapasa, V., & Meshnick, S. R. (2004). Rapid real-time PCR genotyping of mutations associated with sulfadoxine-pyrimethamine resistance in *Plasmodium falciparum. Antimicrobial Agents and Chemotherapy*, 48(8), 2924-2929.
- Allen, M. D., Yamasaki, K., Ohme-Takagi, M., Tateno, M., & Suzuki, M. (1998). A novel mode of DNA recognition by a  $\beta$ -sheet revealed by the solution structure of the GCC-box binding domain in complex with DNA. *The EMBO Journal*, *17*(18), 5484-5496.
- Alonso, J. M., Hirayama, T., Roman, G., Nourizadeh, S., & Ecker, J. R. (1999). EIN2, a bifunctional transducer of ethylene and stress responses in *Arabidopsis*. Science, 284(5423), 2148-2152.
- Arunyanart, S., & Chaitrayagun, M. (2005). Induction of somatic embryogenesis in lotus (*Nelumbo nucifera* Geartn.). *Scientia Horticulturae*, 105(3), 411-420.
- Ashley, J. R. (2004). The Macedonian empire: the era of warfare under Philip II and Alexander the Great, 359-323 BC: McFarland
- Ashraf, M., Athar, H., Harris, P., & Kwon, T. (2008). Some prospective strategies for improving crop salt tolerance. *Advances in Agronomy*, *97*, 45-110.
- Atta, R., Laurens, L., Boucheron-Dubuisson, E., Guivarc'h, A., Carnero, E., Giraudat-Pautot, V., *et al.* (2009). Pluripotency of *Arabidopsis* xylem pericycle underlies shoot regeneration from root and hypocotyl explants grown *in vitro*. *The Plant Journal*, 57(4), 626-644.
- Audi, A., & Muhktar, F. (2009). Effect of pre-sowing hardening treatments using various plant growth substances on cowpea germination and seedling establishment. *Bayero Journal of Pure and Applied Sciences*, 2(2), 44-48.

- Azizi, P., Rafii, M. Y., Mahmood, M., Abdullah, S. N. A., Hanafi, M. M., Latif, M. A., et al. (2017). Evaluation of RNA extraction methods in rice and their application in expression analysis of resistance genes against Magnaporthe oryzae. Biotechnology & Biotechnological Equipment, 31(1), 75-84.
- Azizi, P., Rafii, M. Y., Mahmood, M., Hanafi, M. M., Abdullah, S. N. A., Abiri, R., et al. (2015). Highly efficient protocol for callogenesis, somagenesis and regeneration of Indica rice plants. Comptes Rendus Biologies, 338(7), 463-470.
- Azman, E. A., Jusop, S., Ishak, C. F., & Ismail, R. (2014). Increasing Rice Production Using Different Lime Sources on an Acid Sulphate Soil in Merbok, Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 37(2).
- Bajaj, S., & Mohanty, A. (2005). Recent advances in rice biotechnology—towards genetically superior transgenic rice. *Plant Biotechnology Journal*, 3(3), 275-307.
- Baraldi, P., Bertazza, G., Bregoli, A., Fasolo, F., Rotondi, A., Predieri, S., et al. (1995). Auxins and polyamines in relation to differential in vitro root induction on microcuttings of two pear cultivars. *Journal of Plant Growth Regulation*, 14(1), 49-59.
- Bartlett, J. G., Alves, S. C., Smedley, M., Snape, J. W., & Harwood, W. A. (2008). High-throughput Agrobacterium-mediated barley transformation. *Plant Methods*, 4(1), 22.
- Bartlett, J. M., & Stirling, D. (2003). A short history of the polymerase chain reaction. *PCR Protocols*, 3-6.
- Baskaran, P., & Jayabalan, N. (2009). Psoralen production in hairy roots and adventitious roots cultures of *Psoralea coryfolia*. *Biotechnology Letters*, 31(7), 1073-1077.
- Bates, L., Waldren, R., & Teare, I. (1973). Rapid determination of free proline for water-stress studies. *Plant and soil*, 39(1), 205-207.
- Benchabane, M., Goulet, C., Rivard, D., Faye, L., Gomord, V., & Michaud, D. (2008). Preventing unintended proteolysis in plant protein biofactories. *Plant Biotechnology Journal*, 6(7), 633-648.
- Benjamins, R., & Scheres, B. (2008). Auxin: the looping star in plant development. Annu. Rev. Plant Biol., 59, 443-465.
- Benlioglu, B., Tuna, D., Birsin, M., & Ozgen, A. (2015). Effect of growth regulators on tissue culture parameters in rice (*Oryza sativa* L.). *Ekin J Crop Breed and Gen*, 1-2.
- Bernard, P. S., & Wittwer, C. T. (2002). Real-time PCR technology for cancer diagnostics. *Clinical Chemistry*, 48(8), 1178-1185.

Bhargava, S., & Sawant, K. (2013). Drought stress adaptation: metabolic adjustment

and regulation of gene expression. Plant Breeding, 132(1), 21-32.

- Bhojwani, S. S., & Dantu, P. K. (2013). *Plant tissue culture: an introductory text:* Springer.
- Bhojwani, S. S., & Razdan, M. K. (1986). Plant tissue culture: theory and practice (Vol. 5): Elsevier.
- Birch, R., & Franks, T. (1991). Development and optimisation of microprojectile systems for plant genetic transformation. *Functional Plant Biology*, 18(5), 453-469.
- Birch, R. G. (1997). Plant transformation: problems and strategies for practical application. Annual Review of Plant Biology, 48(1), 297-326.
- Blackman, F. F., & Matthaei, G. L. (1901). On the reaction of leaves to traumatic stimulation. Annals of Botany(3), 533-546.
- Bock, R. (2007). Plastid biotechnology: prospects for herbicide and insect resistance, metabolic engineering and molecular farming. *Current Opinion in Biotechnology*, 18(2), 100-106.
- Box, M. S., Coustham, V., Dean, C., & Mylne, J. S. (2011). Protocol: A simple phenolbased method for 96-well extraction of high quality RNA from Arabidopsis. *Plant Methods*, 7(1), 7.
- Bregitzer, P., & Brown, R. H. (2013). Long-term assessment of transgene behavior in barley: Ds-mediated delivery of bar results in robust, stable, and heritable expression. *In Vitro Cellular & Developmental Biology-Plant*, 49(3), 231-239.
- Buah, J., Tachie-Menson, J., Addae, G., & Asare, P. (2011). Sugarcane juice as an alternative carbon source for *in vitro* culture of plantains and bananas. *American Journal of Food Technology*, 6(8), 685-694.
- Bustin, S. (2002). Quantification of mRNA using real-time reverse transcription PCR (RT-PCR): trends and problems. *Journal of Molecular Endocrinology*, 29(1), 23-39.
- Büttner, M., & Singh, K. B. (1997). Arabidopsis thaliana ethylene-responsive element binding protein (AtEBP), an ethylene-inducible, GCC box DNA- binding protein interacts with an ocs element binding protein. Proceedings of the National Academy of Sciences, 94(11), 5961-5966.
- Cai, S., Jiang, G., Ye, N., Chu, Z., Xu, X., Zhang, J., et al. (2015). A key ABA catabolic gene, OsABA80x3, is involved in drought stress resistance in rice. PloS One, 10(2), e0116646.
- Cerdà, A., & García-Fayos, P. (1997). The influence of slope angle on sediment, water and seed losses on badland landscapes. *Geomorphology*, 18(2), 77-90.

- Chang, S., Puryear, J., & Cairney, J. (1993). A simple and efficient method for isolating RNA from pine trees. *Plant Molecular Biology Reporter*, *11*(2), 113-116.
- Chao, Q., Rothenberg, M., Solano, R., Roman, G., Terzaghi, W., & Ecker, J. R. (1997). Activation of the ethylene gas response pathway in Arabidopsis by the nuclear protein ETHYLENE-INSENSITIVE3 and related proteins. *Cell*, 89(7), 1133-1144.
- Chen, G., Zhu, Y., Wang, H.-Z., Wang, S.-J., & Zhang, R.-Q. (2007). The metabolites of a mangrove endophytic fungus, *Penicillium thomi. Journal of Asian natural* products research, 9(2), 159-164.
- Chen, J., & Ziv, M. (2003). Carbohydrate, metabolic, and osmotic changes in scaled-up liquid cultures of Narcissus leaves. *In Vitro Cellular & Developmental Biology-Plant*, 39(6), 645-650.
- Chen, W., Provart, N. J., Glazebrook, J., Katagiri, F., Chang, H.-S., Eulgem, T., et al. (2002). Expression profile matrix of *Arabidopsis* transcription factor genes suggests their putative functions in response to environmental stresses. *The Plant Cell*, 14(3), 559-574.
- Cheng, J., Zhang, Y., & Li, Q. (2004). Real-time PCR genotyping using displacing probes. *Nucleic Acids Research*, 32(7), e61-e61.
- Chinnusamy, V., Zhu, J., & Zhu, J.-K. (2006). Salt stress signaling and mechanisms of plant salt tolerance *Genetic engineering* (pp. 141-177): Springer.
- Chirgwin, J. M., Przybyla, A. E., MacDonald, R. J., & Rutter, W. J. (1979). Isolation of biologically active ribonucleic acid from sources enriched in ribonuclease. *Biochemistry*, 18(24), 5294-5299.
- Chomczynski, P., & Sacchi, N. (1987). Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Analytical Biochemistry*, *162*(1), 156-159.
- Choudhary, N., Sairam, R., & Tyagi, A. (2005). Expression of  $\Delta^1$ -pyrroline-5carboxylate synthetase gene during drought in rice (*Oryza sativa* L.).
- Chowdhry, C. N., Tyagi, A., Maheshwari, N., & Maheshwari, S. (1993). Effect of Lproline and L-tryptophan on somatic embryogenesis and plantlet regeneration of rice (*Oryza sativa* L. cv. Pusa 169). *Plant Cell, Tissue and Organ Culture, 32*(3), 357-361.
- Christianson, M., & Warnick, D. (1985). Temporal requirement for phytohormone balance in the control of organogenesis in vitro. Developmental Biology, 112(2), 494-497.
- Chugh, A., Eudes, F., & Shim, Y.-S. (2010). Cell-penetrating peptides: Nanocarrier for macromolecule delivery in living cells. *IUBMB Life*, 62(3), 183-193.

- Clermont-Dauphin, C., Suwannang, N., Grünberger, O., Hammecker, C., & Maeght, J.-L. (2010). Yield of rice under water and soil salinity risks in farmers' fields in northeast Thailand. *Field Crops Research*, 118(3), 289-296.
- Dahot, M. U. (2007). Morpho-physiological aspects of micro-propagating banana under different hormonal conditions. *Asian Journal of Plant Sciences*.
- Danquah, A., de Zelicourt, A., Colcombet, J., & Hirt, H. (2014). The role of ABA and MAPK signaling pathways in plant abiotic stress responses. *Biotechnology Advances*, *32*(1), 40-52.
- De Castro, R. D., van Lammeren, A. A., Groot, S. P., Bino, R. J., & Hilhorst, H. W. (2000). Cell division and subsequent radicle protrusion in tomato seeds are inhibited by osmotic stress but DNA synthesis and formation of microtubular cytoskeleton are not. *Plant Physiology*, 122(2), 327-336.
- De Klerk, G.-J., Keppel, M., Ter Brugge, J., & Meekes, H. (1995). Timing of the phases in adventitous root formation in apple microcuttings. *Journal of Experimental Botany*, 46(8), 965-972.
- De Paiva Neto, V. B., & Otoni, W. C. (2003). Carbon sources and their osmotic potential in plant tissue culture: does it matter? *Scientia Horticulturae*, 97(3), 193-202.
- de Silva Gesteira, A., Micheli, F., Ferreira, C. F., & de Mattos Cascardo, J. C. (2003). Isolation and purification of functional total RNA from different organs of cacao tree during its interaction with the pathogen Crinipellis perniciosa. *Biotechniques*, 35(3), 494-501.
- Denecke, J., De Rycke, R., & Botterman, J. (1992). Plant and mammalian sorting signals for protein retention in the endoplasmic reticulum contain a conserved epitope. *The EMBO Journal*, 11(6), 2345.
- Deng, M. Y., Wang, H., Ward, G. B., Beckham, T. R., & McKenna, T. S. (2005). Comparison of six RNA extraction methods for the detection of classical swine fever virus by real-time and conventional reverse transcription–PCR. *Journal of Veterinary Diagnostic Investigation*, 17(6), 574-578.

Din, A. R. J. M., Ahmad, F. I., Wagiran, A., Samad, A. A., Rahmat, Z., & Sarmidi,

- M. R. (2016). Improvement of efficient *in vitro* regeneration potential of mature callus induced from Malaysian upland rice seed (*Oryza sativa* cv. Panderas). *Saudi Journal of Biological Sciences*, 23(1), S69-S77.
- Du, H., Wu, N., Cui, F., You, L., Li, X., & Xiong, L. (2014). A homolog of ETHYLENE OVERPRODUCER, OsETOL1, differentially modulates drought and submergence tolerance in rice. *The Plant Journal*, 78(5), 834-849.
- Edwards, C. E., Ewers, B. E., McClung, C. R., Lou, P., & Weinig, C. (2012). Quantitative variation in water-use efficiency across water regimes and its relationship with circadian, vegetative, reproductive, and leaf gas-exchange traits.

*Molecular Plant*, 5(3), 653-668.

- Egan, J. F., Barlow, K. M., & Mortensen, D. A. (2014). A meta-analysis on the effects of 2, 4-D and dicamba drift on soybean and cotton. *Weed Science*, 62(1), 193-206.
- Erlich Ed, H. (1989). PCR technology: principles and applications for DNA amplification IRL Press at Oxford Univ. Press: Oxford, UK.
- Eyal, Y., Meller, Y., Lev-Yadun, S., & Fluhr, R. (1993). A basic-type PR-1 promoter directs ethylene responsiveness, vascular and abscission zone- specific expression. *The Plant Journal*, 4(2), 225-234.
- Fang, Y., & Xiong, L. (2015). General mechanisms of drought response and their application in drought resistance improvement in plants. *Cellular and Molecular Life Sciences*, 72(4), 673-689.
- FAO. (2014). Statistical Yearbook, Asia and the Pacific Food and Agriculture.
- FAO. (2015). Statistical Pocketbook. Asia and the Pacific Food and Agriculture.
- Farooq, M., Barsa, S. M., & Wahid, A. (2006a). Priming of field-sown rice seed enhances germination, seedling establishment, allometry and yield. *Plant Growth Regulation*, 49(2-3), 285-294.
- Farooq, M., Basra, S., & Hafeez, K. (2006b). Seed invigoration by osmohardening in coarse and fine rice. Seed Science and Technology, 34(1), 181-187.
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., & Basra, S. (2009). Plant drought stress: effects, mechanisms and management *Sustainable agriculture* (pp. 153-188): Springer.
- Feramisco, J., Perona, R., & Lacal, J. (1999). Needle microinjection: a brief history *Microinjection* (pp. 9-15): Springer.
- Finkelstein, R. R., & Lynch, T. J. (2000). The *Arabidopsis* abscisic acid response gene *ABI5* encodes a basic leucine zipper transcription factor. *The Plant Cell*, *12*(4), 599-609.
- Floss, D. M., Falkenburg, D., & Conrad, U. (2007). Production of vaccines and therapeutic antibodies for veterinary applications in transgenic plants: an overview. *Transgenic Research*, 16(3), 315-332.
- Fonseca, C., Planchon, S., Serra, T., Chander, S., Saibo, N. J., Renaut, J., et al. (2015). In vitro culture may be the major contributing factor for transgenic versus nontransgenic proteomic plant differences. Proteomics, 15(1), 124-134.
- Franconi, R., Demurtas, O. C., & Massa, S. (2010). Plant-derived vaccines and other therapeutics produced in contained systems. *Expert Review of Vaccines*, 9(8), 877-892.

- Fujita, M., Fujita, Y., Noutoshi, Y., Takahashi, F., Narusaka, Y., Yamaguchi-Shinozaki, K., *et al.* (2006). Crosstalk between abiotic and biotic stress responses: a current view from the points of convergence in the stress signaling networks. *Current Opinion in Plant Biology*, 9(4), 436-442.
- Fukao, T., Harris, T., & Bailey-Serres, J. (2009). Evolutionary analysis of the Sub1 gene cluster that confers submergence tolerance to domesticated rice. Annals of Botany, 103(2), 143-150.
- Gairola, K., Nautiyal, A., & Dwivedi, A. (2011). Effect of temperatures and germination media on seed germination of *Jatropha curcas* Linn. Advances in Bioresearch, 2(2), 66-71.
- Gao, J. P., Chao, D. Y., & Lin, H. X. (2007). Understanding abiotic stress tolerance mechanisms: recent studies on stress response in rice. *Journal of Integrative Plant Biology*, 49(6), 742-750.
- Gao, F., Wang, Y., Shi, D., Zhang, J., Wang, M., Jing, X., ... & Gritzel, M. (2008). Enhance the optical absorptivity of nanocrystalline TiO2 film with high molar extinction coefficient ruthenium sensitizers for high performance dye-sensitized solar cells. *Journal of the American Chemical Society*, 130(32), 10720-10728.
- Ge, X., Chu, Z., Lin, Y., & Wang, S. (2006). A tissue culture system for different germplasms of indica rice. *Plant Cell Reports*, 25(5), 392-402.
- Gelvin, S. B. (2003). Agrobacterium-mediated plant transformation: the biology behind the "gene-jockeying" tool. *Microbiology and Molecular Biology Reviews*, 67(1), 16-37.
- Gelvin, S. B. (2010). Plant proteins involved in *Agrobacterium*-mediated genetic transformation. *Annual Review of Phytopathology*, 48, 45-68.
- George, E. F., Hall, M. A., & De Klerk, G.-J. (2008). Micropropagation: uses and methods *Plant propagation by tissue culture* (pp. 29-64): Springer.
- Gibson, N. J. (2006). The use of real-time PCR methods in DNA sequence variation analysis. *Clinica Chimica Acta*, 363(1), 32-47.
- Gleba, Y., Klimyuk, V., & Marillonnet, S. (2005). Magnifection—a new platform for expressing recombinant vaccines in plants. *Vaccine*, 23(17), 2042-2048.
- Gordon-Kamm, W. J., Spencer, T. M., Mangano, M. L., Adams, T. R., Daines, R. J., Start, W. G., *et al.* (1990). Transformation of maize cells and regeneration of fertile transgenic plants. *The Plant Cell*, 2(7), 603-618.
- Gu, Y.-Q., Wildermuth, M. C., Chakravarthy, S., Loh, Y.-T., Yang, C., He, X., et al. (2002). Tomato transcription factors *Pti4*, *Pti5*, and *Pti6* activate defense responses when expressed in *Arabidopsis*. *The Plant Cell*, *14*(4), 817-831.

- Gunawardena, A. H., Pearce, D. M., Jackson, M. B., Hawes, C. R., & Evans, D. E. (2001). Characterisation of programmed cell death during aerenchyma formation induced by ethylene or hypoxia in roots of maize (*Zea mays L.*). *Planta*, 212(2), 205-214.
- Haberer, G., & Kieber, J. J. (2002). Cytokinins. New insights into a classic phytohormone. *Plant Physiology*, *128*(2), 354-362.
- Hamidi, R., & Pirasteh-Anosheh, H. (2013). Comparison effect of different seed priming methods on sunflower germination and seedling growth. *Int J Agron Plant Prod*, 4, 1247-1250.
- Haque, M. S., Wada, T., & Hattori, K. (2003). Effects of Sucrose, Mannitol and KH<sub>2</sub>PO<sub>4</sub> on Proliferation of Root Tip Derived Shoots and Subsequent Bulblet Formation in Garlic. *Asian Journal of Plant Sciences*.
- Hare, P., & Cress, W. (1997). Metabolic implications of stress-induced proline accumulation in plants. *Plant Growth Regulation*, 21(2), 79-102.
- Hattori, Y., Nagai, K., Furukawa, S., Song, X.-J., Kawano, R., Sakakibara, H., *et al.* (2009). The ethylene response factors SNORKEL1 and SNORKEL2 allow rice to adapt to deep water. *Nature*, 460(7258), 1026-1030.
- He, C., Wang, L., Liu, J., Liu, X., Li, X., Ma, J., et al. (2013). Evidence for 'silicon'within the cell walls of suspension-cultured rice cells. New Phytologist, 200(3), 700-709.
- Helling, D., Possart, A., Cottier, S., Klahre, U., & Kost, B. (2006). Pollen tube tip growth depends on plasma membrane polarization mediated by tobacco PLC3 activity and endocytic membrane recycling. *The Plant Cell*, 18(12), 3519-3534.
- Hiei, Y., Ohta, S., Komari, T., & Kumashiro, T. (1994). Efficient transformation of rice (*Oryza sativa* L.) mediated by Agrobacterium and sequence analysis of the boundaries of the T-DNA. *The Plant Journal*, 6(2), 271-282.
- Holme, I. B., Krogstrup, P., & Hansen, J. (1997). Embryogenic callus formation, growth and regeneration in callus and suspension cultures of *Miscanthus x ogiformis Honda Giganteus*' as affected by proline. *Plant Cell, Tissue and Organ Culture*, 50(3), 203-210.
- Holsters, M., Waele, D. D., Depicker, A., Messens, E., Montagu, M. v., & Schell, J. (1978). Transfection and transformation of Agrobacterium tumefaciens. Molecular and General Genetics MGG, 163(2), 181-187.
- Hoque, M. E., & Mansfield, J. W. (2004). Effect of genotype and explant age on callus induction and subsequent plant regeneration from root-derived callus of Indica rice genotypes. *Plant Cell, Tissue and Organ Culture,* 78(3), 217-223.
- Horn, M., Woodard, S., & Howard, J. (2004). Plant molecular farming: systems and products. *Plant Cell Reports*, 22(10), 711-720.

- Horton, R. F. (1991). The effect of ethylene and other regulators on coleoptile growth of rice under anoxia. *Plant Science*, *79*(1), 57-62.
- Htwe, N. N., Maziah, M., Ling, H. C., Zaman, F. Q., & Zain, A. M. (2011). Regeneration capacity of cell suspension culture in Malaysian rice genotypes under salinity stress. Asian J Biotechnol, 3, 357-367.
- Huang, Z., Zhang, Z., Zhang, X., Zhang, H., Huang, D., & Huang, R. (2004). Tomato *TERF1* modulates ethylene response and enhances osmotic stress tolerance by activating expression of downstream genes. *Febs Letters*, 573(1-3), 110-116.
- Hussain, Z., Ali, S., Hayat, Z., Zia, M. A., Iqbal, A., & Ali, G. M. (2014). Agrobacterium mediated transformation of DREB1A gene for improved drought tolerance in rice cultivars (Oryza sativa L.). Australian Journal of Crop Science, 8(7), 1114.
- Ikeuchi, M., Sugimoto, K., & Iwase, A. (2013). Plant callus: mechanisms of induction and repression. *The Plant Cell*, 25(9), 3159-3173.
- Ingram, J., Ericksen, P., & Liverman, D. (2012). Food security and global environmental change: Routledge.
- Iwai, H., Masaoka, N., Ishii, T., & Satoh, S. (2002). A pectin glucuronyltransferase gene is essential for intercellular attachment in the plant meristem. *Proceedings of the National Academy of Sciences*, 99(25), 16319-16324.
- Jaakola, L., Pirttilä, A. M., Halonen, M., & Hohtola, A. (2001). Isolation of high quality RNA from bilberry (*Vaccinium myrtillus* L.) fruit. *Molecular Biotechnology*, 19(2), 201-203.
- Jaleel, C. A., Manivannan, P., Wahid, A., Farooq, M., Al-Juburi, H. J., Somasundaram, R., et al. (2009). Drought stress in plants: a review on morphological characteristics and pigments composition. *International Journal of Agriculture* and Biology, 11(1), 100-105.
- Jamil, M., Deog Bae, L., Kwang Yong, J., Ashraf, M., Sheong Chun, L., & Eui Shik, R. (2006). Effect of salt (NaCl) stress on germination and early seedling growth of four vegetables species. *Journal of Central European Agriculture*, 7(2), 273-282.
- Jenesl, B., Moore, H., Caoz, J., & Zhang, W. (2012). Techniques for gene transfer. *Transgenic Plants: Engineering and Utilization.*
- Jia, Y., Zhang, Q.-X., Pan, H.-T., Wang, S.-Q., Liu, Q.-L., & Sun, L.-X. (2014). Callus induction and haploid plant regeneration from baby primrose (*Primula forbesii Franch.*) anther culture. *Scientia Horticulturae*, 176, 273-281.
- Jia, Y., Zhang, Q. X., Pan, H. T., Wang, S. Q., Liu, Q. L., & Sun, L. X. (2014). Callus induction and haploid plant regeneration from baby primrose (*Primula forbesii* Franch.) anther culture. *Scientia Horticulturae*, 176,273-281.

- Jiménez, V. M. (2005). Involvement of plant hormones and plant growth regulators on *in vitro* somatic embryogenesis. *Plant Growth Regulation*, 47(2-3), 91- 110.
- Jin, L. G., Li, H., & Liu, J. Y. (2010). Molecular characterization of three ethylene responsive element binding factor genes from cotton. *Journal of Integrative Plant Biology*, 52(5), 485-495.
- Jofuku, K. D., Omidyar, P. K., Gee, Z., & Okamuro, J. K. (2005). Control of seed mass and seed yield by the floral homeotic gene APETALA2. *Proceedings of the National Academy of Sciences of the United States of America*, 102(8), 3117-3122.
- John, M. E. (1992). An efficient method for isolation of RNA and DNA from plants containing polyphenolics. *Nucleic Acids Research*, 20(9), 2381.
- Johnston, R. J., Poholek, A. C., DiToro, D., Yusuf, I., Eto, D., Barnett, B., *et al.* (2009). Bcl6 and Blimp-1 are reciprocal and antagonistic regulators of T follicular helper cell differentiation. *Science*, 325(5943), 1006-1010.
- Joo, J., Choi, H. J., Lee, Y. H., Lee, S., Lee, C. H., Kim, C. H., *et al.* (2014). Over expression of *BvMTSH*, a fusion gene for maltooligosyltrehalose synthase and maltooligosyltrehalose trehalohydrolase, enhances drought tolerance in transgenic rice. *BMB Reports*, 47(1), 27.
- Kagaya, Y., Ohmiya, K., & Hattori, T. (1999). RAV1, a novel DNA-binding protein, binds to bipartite recognition sequence through two distinct DNA- binding domains uniquely found in higher plants. *Nucleic Acids Research*, 27(2), 470-478.
- Kandasamy, M. K., Gilliland, L. U., McKinney, E. C., & Meagher, R. B. (2001). One plant actin isovariant, ACT7, is induced by auxin and required for normal callus formation. *The Plant Cell*, 13(7), 1541-1554.
- Kareem, I., Ismail, M. R., Puteh, A., Rahim, A. A., Habib, S. H., & Kausar, H. (2013). Potential osmotic and hormonal priming for higher productivity of rice. *Journal* of Food, Agriculture & Environment, 11(2), 737-741.
- Karimi, M., Inzé, D., & Depicker, A. (2002). GATEWAY<sup>™</sup> vectors for Agrobacterium-mediated plant transformation. Trends in Plant Science, 7(5), 193-195.
- Karimi, M., Inzé, D., Van Lijsebettens, M., & Hilson, P. (2013). Gateway vectors for transformation of cereals. *Trends in Plant Science*, 18(1), 1-4.
- Kavitah, G., Taghipour, F., & Huyop, F. (2010). Investigation of factors in optimizing Agrobacterium-mediated gene transfer in Citrullus lanatus cv. Round Dragon. Journal of Biological Sciences, 10(3), 209-216.

Kazan, K. (2015). Diverse roles of jasmonates and ethylene in abiotic stress tolerance.

Trends in Plant Science, 20(4), 219-229.

- Kendrick, M. D., & Chang, C. (2008). Ethylene signaling: new levels of complexity and regulation. *Current Opinion in Plant Biology*, 11(5), 479-485.
- Keyvan, S. (2010). The effects of drought stress on yield, relative water content, proline, soluble carbohydrates and chlorophyll of bread wheat cultivars. *Journal of Animal and Plant Science*, 8(3), 1051-1060.
- Khush, G. (2000). Taxonomy and origin of rice. Aromatic rices, 5-13.
- Kido, N., Yokoyama, R., Yamamoto, T., Furukawa, J., Iwai, H., Satoh, S., *et al.* (2014). The matrix polysaccharide (1; 3, 1; 4)-β-D-glucan is involved in silicondependent strengthening of rice cell wall. *Plant and Cell Physiology*, pcu162.
- Kiefer, E., Heller, W., & Ernst, D. (2012). A simple and efficient protocol for isolation of functional RNA from plant tissues rich in secondary metabolites. *Plant Molecular Biology Reporter*, 18(1), 33-39.
- Kim, H., Lee, K., Hwang, H., Bhatnagar, N., Kim, D.-Y., Yoon, I. S., et al. (2014). Overexpression of PYL5 in rice enhances drought tolerance, inhibits growth, and modulates gene expression. Journal of Experimental Botany, 65(2), 453-464.
- Kim, J. S., Park, H.-M., Chae, S., Lee, T.-H., Hwang, D.-J., Oh, S.-D., et al. (2014). A pepper MSRB2 gene confers drought tolerance in rice through the protection of chloroplast-targeted genes. PloS One, 9(3), e90588.
- Kim, T.-G., Baek, M.-Y., Lee, E.-K., Kwon, T.-H., & Yang, M.-S. (2008). Expression of human growth hormone in transgenic rice cell suspension culture. *Plant Cell Reports*, 27(5), 885-891.
- Kocsy, G., Galiba, G., & Brunold, C. (2001). Role of glutathione in adaptation and signalling during chilling and cold acclimation in plants. *Physiologia Plantarum*, 113(2), 158-164.
- Konaté, S., Kone, M., Kouakou, H., Kouadio, J., & Zouzou, M. (2013). Callus induction and proliferation from cotyledon explants in *Bambara groundnut*. *African Crop Science Journal*, 21(3), 255-263.
- Kovach, M. J., Sweeney, M. T., & McCouch, S. R. (2007). New insights into the history of rice domestication. *Trends in Genetics*, 23(11), 578-587.
- Kovács, Z., Simon-Sarkadi, L., Vashegyi, I., & Kocsy, G. (2012). Different accumulation of free amino acids during short-and long-term osmotic stress in wheat. *The Scientific World Journal*, 2012.
- Lacroix, B., Tzfira, T., Vainstein, A., & Citovsky, V. (2006). A case of promiscuity: *Agrobacterium*'s endless hunt for new partners. *Trends in Genetics*, 22(1), 29-37.

Lambers, H., Atkin, O. K., & Millenaar, F. F. (2002). Respiratory patterns in roots in

relation to their functioning *Plant Roots: The Hidden Half, Third Edition* (pp. 521-552): CRC Press.

- Lau, W., Fischbach, M. A., Osbourn, A., & Sattely, E. S. (2014). Key applications of plant metabolic engineering. *PLoS Biol*, 12(6), e1001879.
- Leng, L. W., & Lai-Keng, C. (2004). Plant regeneration from stem nodal segments of Orthosiphon stamineus Benth., a medicinal plant with diuretic activity. *In Vitro Cellular & Developmental Biology-Plant*, 40(1), 115-118.
- Lenucci, M. S., Cadinu, D., Taurino, M., Piro, G., & Dalessandro, G. (2006). Antioxidant composition in cherry and high-pigment tomato cultivars. *Journal of Agricultural and Food Chemistry*, 54(7), 2606-2613.
- Leyser, O. (2010). The power of auxin in plants. Plant Physiology, 154(2), 501-505.
- Li, C.-H., Wang, G., Zhao, J.-L., Zhang, L.-Q., Ai, L.-F., Han, Y.-F., et al. (2014). The receptor-like kinase SIT1 mediates salt sensitivity by activating MAPK3/6 and regulating ethylene homeostasis in rice. *The Plant Cell*, 26(6), 2538-2553.
- Li, Z., Zhou, H., Peng, Y., Zhang, X., Ma, X., Huang, L., *et al.* (2015). Exogenously applied spermidine improves drought tolerance in creeping bentgrass associated with changes in antioxidant defense, endogenous polyamines and phytohormones. *Plant Growth Regulation*, *76*(1), 71-82.
- Liao, Z., Chen, M., Guo, L., Gong, Y., Tang, F., Sun, X., et al. (2004). Rapid isolation of high-quality total RNA from Taxus and Ginkgo. *Preparative Biochemistry* and Biotechnology, 34(3), 209-214.
- Lichtenthaler, H. K., & Wellburn, A. R. (1983). Determinations of total carotenoids and chlorophylls a and b of leaf extracts in different solvents: Portland Press Limited.
- Lin, Y. J., & Zhang, Q. (2005). Optimising the tissue culture conditions for high efficiency transformation of indica rice. *Plant Cell Reports*, 23(8), 540-547.
- Liu, G., Li, X., Jin, S., Liu, X., Zhu, L., Nie, Y., *et al.* (2014). Overexpression of rice NAC gene *SNAC1* improves drought and salt tolerance by enhancing root development and reducing transpiration rate in transgenic cotton. *PLoS One*, 9(1), e86895.
- Liu, Q., Kasuga, M., Sakuma, Y., Abe, H., Miura, S., Yamaguchi-Shinozaki, K., et al. (1998). Two transcription factors, *DREB1* and *DREB2*, with an *EREBP/AP2* DNA binding domain separate two cellular signal transduction pathways in drought-and low-temperature-responsive gene expression, respectively, in *Arabidopsis. The Plant Cell*, 10(8), 1391-1406.
- Lorenzo, O., Piqueras, R., Sánchez-Serrano, J. J., & Solano, R. (2003). ETHYLENE RESPONSE FACTOR1 integrates signals from ethylene and jasmonate pathways in plant defense. *The Plant Cell*, 15(1), 165-178.

- Løvdal, T., & Lillo, C. (2009). Reference gene selection for quantitative real-time PCR normalization in tomato subjected to nitrogen, cold, and light stress. *Analytical Biochemistry*, 387(2), 238-242.
- Lucas, S., Durmaz, E., Akpinar, B. A., & Budak, H. (2011). The drought response displayed by a DRE-binding protein from Triticum dicoccoides. *Plant Physiology and Biochemistry*, 49(3), 346-351.
- Ma, J. F. (2004). Role of silicon in enhancing the resistance of plants to biotic and abiotic stresses. Soil Science and Plant Nutrition, 50(1), 11-18.
- Ma, Y., Yang, X., Huan, X., Wang, W., Ma, Z., Li, Z., et al. (2016). Rice bulliform phytoliths reveal the process of rice domestication in the Neolithic Lower Yangtze River region. *Quaternary International*.
- Magnani, E., Sjölander, K., & Hake, S. (2004). From endonucleases to transcription factors: evolution of the AP2 DNA binding domain in plants. *The Plant Cell*, *16*(9), 2265-2277.
- Malnoy, M., Reynoird, J., Mourgues, F., Chevreau, E., & Simoneau, P. (2001). A method for isolating total RNA from pear leaves. *Plant Molecular Biology Reporter*, 19(1), 69-69.
- Mannhalter, C., Koizar, D., & Mitterbauer, G. (2000). Evaluation of RNA isolation methods and reference genes for RT-PCR analyses of rare target RNA. *Clinical Chemistry and Laboratory Medicine*, 38(2), 171-177.

MARDI. (2016).

- Marillonnet, S., Thoeringer, C., Kandzia, R., Klimyuk, V., & Gleba, Y. (2005). Systemic Agrobacterium tumefaciens-mediated transfection of viral replicons for efficient transient expression in plants. *Nature Biotechnology*, 23(6), 718-723.
- Marone, M., Mozzetti, S., De Ritis, D., Pierelli, L., & Scambia, G. (2001). Semiquantitative RT-PCR analysis to assess the expression levels of multiple transcripts from the same sample. *Biol Proced Online*, 3(1), 19-25.
- Martin, K. P., Zhang, C.-L., Hembrom, M. E., Slater, A., & Madassery, J. (2008). Adventitious root induction in Ophiorrhiza prostrata: a tool for the production of camptothecin (an anticancer drug) and rapid propagation. *Plant Biotechnology Reports*, 2(2), 163-169.
- Masood, A., Iqbal, N., & Khan, N. A. (2012). Role of ethylene in alleviation of cadmium induced photosynthetic capacity inhibition by sulphur in mustard. *Plant, Cell & Environment, 35*(3), 524-533.
- Matuszewski, L., Persigehl, T., Wall, A., Schwindt, W., Tombach, B., Fobker, M., et al. (2005). Cell Tagging with Clinically Approved Iron Oxides: Feasibility and Effect of Lipofection, Particle Size, and Surface Coating on Labeling Efficiency 1. Radiology, 235(1), 155-161.

- Maya, M. A., & Matsubara, Y.-i. (2013). Influence of arbuscular mycorrhiza on the growth and antioxidative activity in cyclamen under heat stress. *Mycorrhiza*, 23(5), 381-390.
- Meyer, M., de Angelis, M. H., Wurst, W., & Kühn, R. (2010). Gene targeting by homologous recombination in mouse zygotes mediated by zinc-finger nucleases. *Proceedings of the National Academy of Sciences*, 107(34), 15022-15026.
- Mizoi, J., Shinozaki, K., & Yamaguchi-Shinozaki, K. (2012). AP2/ERF family transcription factors in plant abiotic stress responses. *Biochimica et Biophysica* Acta (BBA)-Gene Regulatory Mechanisms, 1819(2), 86-96.
- MO, A. (2010). Comparison of five different RNA isolation methods from equine endometrium for gene transcription analysis. *Kafkas Üniversitesi veteriner fakültesi dergisi*, 16(5).
- Moghaddam, B. E., Mesbah, M., & Yavari, N. (2000). The effect of in planta TIBA and proline treatment on somatic embryogenesis of sugar beet (*Beta vulgaris* L.). *Euphytica*, 112(2), 151-156.
- Moser, C., Gatto, P., Moser, M., Pindo, M., & Velasco, R. (2004). Isolation of functional RNA from small amounts of different grape and apple tissues. *Molecular Biotechnology*, 26(2), 95-99.
- Mostajeran, A., & Rahimi-Eichi, V. (2009). Effects of drought stress on growth and yield of rice (*Oryza sativa* L.) cultivars and accumulation of proline and soluble sugars in sheath and blades of their different ages leaves. *American- Eurasian Journal of Agricultural and Environmental Science*, 5(2), 264-272.
- Munné-Bosch, S., Jubany-Marí, T., & Alegre, L. (2001). Drought-induced senescence is characterized by a loss of antioxidant defences in chloroplasts. *Plant, Cell & Environment*, 24(12), 1319-1327.
- Murashige, T., & Skoog, F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum*, 15(3), 473-497.
- Nakano, T., Suzuki, K., Fujimura, T., & Shinshi, H. (2006). Genome-wide analysis of the ERF gene family in Arabidopsis and rice. *Plant Physiology*, *140*(2), 411-432.
- Nakashima, K., Jan, A., Todaka, D., Maruyama, K., Goto, S., Shinozaki, K., *et al.* (2014). Comparative functional analysis of six drought-responsive promoters in transgenic rice. *Planta*, 239(1), 47-60.
- Narusaka, Y., Narusaka, M., Iwabuchi, M., & Yamasaki, S. (2012). Methods to transfer foreign genes to plants: INTECH Open Access Publisher.

Neely, D. (1979). Tree wounds and wound closure. Journal of Arboriculture (USA).

Ngomuo, M., & Ndakidemi, P. (2013). The effects of auxins and cytokinin on growth

and development of (*Musa sp.*) Var."Yangambi" explants in tissue culture. *American Journal of Plant Sciences*, 4(11), 2174.

- Ning, D., Song, A., Fan, F., Li, Z., & Liang, Y. (2014). Effects of slag-based silicon fertilizer on rice growth and brown-spot resistance. *Plos One*, *9*(7), e102681.
- Off-Farm Employment Participation Amoung Paddy Farmers in the Muda Agricultural Development Authority and Kemasin, 16(2009).
- Nwe, N. H., Mahmood, M., Ho, C. L., Qamaruz Zaman, F., & Md Zain, A. (2011). Regeneration capacity of cell suspension culture in Malaysian rice genotypes under salinity stress. *Asian Journal of Biotechnology*, 3(4), 357-367.
- Obembe, O. O., Popoola, J. O., Leelavathi, S., & Reddy, S. V. (2011). Advances in plant molecular farming. *Biotechnology Advances*, 29(2), 210-222.
- Ohme-Takagi, M., & Shinshi, H. (1995). Ethylene-inducible DNA binding proteins that interact with an ethylene-responsive element. *The Plant Cell*, 7(2), 173-182.
- Pan, R., & Tian, X. (1999). Comparative effect of IBA, BSAA and 5, 6-Cl2-IAA- Me on the rooting of hypocotyl in mung bean. *Plant Growth Regulation*, 27(2), 91-98.
- Pant, B., & Bose, B. (2016). Mitigation of the influence of PEG-6000 imposed water stress on germination of halo primed rice seeds. *International Journal of Agriculture, Environment and Biotechnology*, 9(2), 275-281.
- Pantaleoni, L., Longoni, P., Ferroni, L., Baldisserotto, C., Leelavathi, S., Reddy, V. S., *et al.* (2014). Chloroplast molecular farming: efficient production of a thermostable xylanase by Nicotiana tabacum plants and long-term conservation of the recombinant enzyme. *Protoplasma*, 251(3), 639-648.
- Park, J. S., Gamboni-Robertson, F., He, Q., Svetkauskaite, D., Kim, J. Y., Strassheim, D., ... & Ishizaka, A. (2005). High Mobility Group Box 1 protein (HMGB1) interacts with multiple Toll like receptors. *American Journal of Physiology-Cell Physiology*.
- Park, J. M., Park, C.-J., Lee, S.-B., Ham, B.-K., Shin, R., & Paek, K.-H. (2001). Overexpression of the tobacco *Tsi1* gene encoding an EREBP/AP2–type transcription factor enhances resistance against pathogen attack and osmotic stress in tobacco. *The Plant Cell*, *13*(5), 1035-1046.
- Pawar, B., Prashant, K., Bahurupe, J., Jadhav, A., Anil, K., & Pawar, S. (2015). Proline and glutamine improve *in vitro* callus induction and subsequent shooting in rice. *Rice Science*, 22(6), 283-289.
- Peeters, A. J., Cox, M. C., Benschop, J. J., Vreeburg, R. A., Bou, J., & Voesenek, L. A. (2002). Submergence research using Rumex palustris as a model; looking back and going forward. *Journal of Experimental Botany*, 53(368), 391-398.

- Peng, S., Huang, J., Sheehy, J. E., Laza, R. C., Visperas, R. M., Zhong, X., et al. (2004). Rice yields decline with higher night temperature from global warming. Proceedings of the National academy of Sciences of the United States of America, 101(27), 9971-9975.
- Pérez-Díaz, J., Wu, T.-M., Pérez-Díaz, R., Ruíz-Lara, S., Hong, C.-Y., & Casaretto, J. A. (2014). Organ-and stress-specific expression of the ASR genes in rice. *Plant Cell Reports*, 33(1), 61-73.
- Phongsisay, V., Perera, V. N., & Fry, B. N. (2007). Evaluation of eight RNA isolation methods for transcriptional analysis in *Campylobacter jejuni*. *Journal of Microbiological Methods*, 68(2), 427-429.
- Pirrello, J., Jaimes-Miranda, F., Sanchez-Ballesta, M. T., Tournier, B., Khalil- Ahmad, Q., Regad, F., *et al.* (2006). *Sl-ERF2*, a tomato ethylene response factor involved in ethylene response and seed germination. *Plant and Cell Physiology*, 47(9), 1195-1205.
- Poesen, J., & Savat, J. (1981). Detachment and transportation of loose sediments by raindrop splash: Part II Detachability and transport ability measurements. *Catena*, 8(1), 19-41.
- Qiao, Z.-X., Huang, B., & Liu, J.-Y. (2008). Molecular cloning and functional analysis of an ERF gene from cotton (Gossypium hirsutum). *Biochimica et Biophysica Acta (BBA)-Gene Regulatory Mechanisms*, 1779(2), 122-127.
- Radziah, C. M. C., Nurkhalida, A. S., Zamri, Z., & Ismanizan, I. (2012). Effect of illumination, casein hydrolysate and proline on callus induction of *Oryza sativa* L. CV. MR219. *Malaysian Applied Biology*, 41(1), 37-41.
- Rahman, A., Bannigan, A., Sulaman, W., Pechter, P., Blancaflor, E. B., & Baskin, T. I. (2007). Auxin, actin and growth of the *Arabidopsis thaliana* primary root. *The Plant Journal*, 50(3), 514-528.
- Rai, V. (2002). Role of amino acids in plant responses to stresses. *Biologia Plantarum*, 45(4), 481-487.
- Raineri, D., Bottino, P., Gordon, M., & Nester, E. (1990). *Agrobacterium*–Mediated Transformation of Rice (*Oryza sativa* L.). *Nature Biotechnology*, 8(1), 33-38.
- Rao, A. M., Kumar, I. S., & Kishor, P. K. (2012). Effect of growth regulators and Physiological Gradients on the High frequency plant regeneration from the longterm callus cultures of different germplasms of Rice (Oryza sativa L.). *Journal of Phytology*, 4(2).
- Rachmawati, D., & Anzai, H. (2006). Studies on callus induction, plant regeneration and transformation of Javanica rice cultivars. *Plant Biotechnology*, 23(5), 521-524.

Rashid, U., Ali, S., Ali, G. M., Ayub, N., & Masood, M. S. (2009). Establishment of an

efficient callus induction and plant regeneration system in Pakistani wheat (*Triticum aestivum*) cultivars. *Electronic Journal of Biotechnology*, 12(3), 4-5.

- Razmjoo, K., Heydarizadeh, P., & Sabzalian, M. R. (2008). Effect of salinity and drought stresses on growth parameters and essential oil content of Matricaria chamomile. *Int. J. Agric. Biol*, 10(4), 451-454.
- Ravikumar, G., Manimaran, P., Voleti, S., Subrahmanyam, D., Sundaram, R., Bansal, K., *et al.* (2014). Stress-inducible expression of *AtDREB1A* transcription factor greatly improves drought stress tolerance in transgenic indica rice. *Transgenic Research*, 23(3), 421-439.
- Reddy, V. S., Leelavathi, S., Selvapandiyan, A., Raman, R., Giovanni, F., Shukla, V., *et al.* (2002). Analysis of chloroplast transformed tobacco plants with cry1Ia5 under rice psbA transcriptional elements reveal high level expression of Bt toxin without imposing yield penalty and stable inheritance of transplastome. *Molecular Breeding*, 9(4), 259-269.
- Rivera, A. L., Gómez-Lim, M., Fernández, F., & Loske, A. M. (2012). Physical methods for genetic plant transformation. *Physics of Life Reviews*, 9(3), 308-345.
- Roberts, E., & Ellis, R. (1989). Water and seed survival. Annals of Botany, 63(1), 39-39.
- Rodríguez-Serrano, M., Pazmiño, D., Sparkes, I., Rochetti, A., Hawes, C., Romero-Puertas, M., et al. (2014). 2, 4-Dichlorophenoxyacetic acid promotes Snitrosylation and oxidation of actin affecting cytoskeleton and peroxisomal dynamics. *Journal of Experimental botany*, eru237.
- Roy, I., Mitra, S., Maitra, A., & Mozumdar, S. (2003). Calcium phosphate nanoparticles as novel non-viral vectors for targeted gene delivery. *International Journal of Pharmaceutics*, 250(1), 25-33.
- Rzewuski, G., & Sauter, M. (2008). Ethylene biosynthesis and signaling in rice. *Plant Science*, *175*(1), 32-42.
- Sahebi, M., & Hanafi, M. (2013). Extraction of total RNA from mangrove plants to identify different genes involved in its adaptability to the variety of stresses. *Pakistan Journal of Agriculture Sciences.*
- Sahebi, M., Hanafi, M. M., Abdullah, S. N. A., Nejat, N., Rafii, M. Y., & Azizi, P. (2013). Extraction of total RNA from mangrove plants to identify different genes involved in its adaptability to the variety of stresses. *Pakistan Journal of Agricultural Science*, 50(4), 1-9.
- Sahebi, M., Hanafi, M. M., & Azizi, P. (2016). Application of silicon in plant tissue culture. *In Vitro Cellular & Developmental Biology-Plant*, 1-7.
- Salzman, R., Fujita, T., Zhu-Salzman, K., Hasegawa, P., & Bressan, R. (1999). An improved RNA isolation method for plant tissues containing high levels of phenolic compounds or carbohydrates. *Plant Molecular Biology Reporter*, 17(1),

- Santner, A., & Estelle, M. (2009). Recent advances and emerging trends in plant hormone signalling. *Nature*, 459(7250), 1071-1078.
- Sanusi, W., Jemain, A. A., Zin, W. Z. W., & Zahari, M. (2015). The drought characteristics using the first-order homogeneous Markov chain of monthly rainfall data in peninsular Malaysia. *Water Resources Management*, 29(5), 1523-1539.
- Sasidharan, R., & Voesenek, L. A. (2015). Ethylene-mediated acclimations to flooding stress. *Plant physiology*, 169(1), 3-12.
- Schneiderbauer, A., Sandermann, H., & Ernst, D. (1991). Isolation of functional RNA from plant tissues rich in phenolic compounds. *Analytical Biochemistry*, 197(1), 91-95.
- Seeburg, P. H., Shine, J., Martial, J. A., Ullrich, A., Baxter, J. D., & Goodman, H. M. (1977). Nucleotide sequence of part of the gene for human chorionic somatomammotropin: purification of DNA complementary to predominant mRNA species. *Cell*, 12(1), 157-165.
- Shaaban, A. J., & Low, K. S. (2003). Droughts in Malaysia: A look at its characteristics, impacts, related policies and management strategies. Paper presented at the Water and Drainage 2003 Conference.
- Shaaltiel, Y., Bartfeld, D., Hashmueli, S., Baum, G., Brill-Almon, E., Galili, G., et al. (2007). Production of glucocerebrosidase with terminal mannose glycans for enzyme replacement therapy of Gaucher's disease using a plant cell system. *Plant Biotechnology Journal*, 5(5), 579-590.
- Shao, H.-B., Guo, Q.-J., Chu, L.-Y., Zhao, X.-N., Su, Z.-L., Hu, Y.-C., et al. (2007). Understanding molecular mechanism of higher plant plasticity under abiotic stress. Colloids and Surfaces B: Biointerfaces, 54(1), 37-45.
- Sheng, J., & Citovsky, V. (1996). Agrobacterium-plant cell DNA transport: have virulence proteins, will travel. The Plant Cell, 8(10), 1699.
- Shi, Y., Tian, S., Hou, L., Huang, X., Zhang, X., Guo, H., *et al.* (2012). Ethylene signaling negatively regulates freezing tolerance by repressing expression of CBF and type-A ARR genes in Arabidopsis. *The Plant Cell*, 24(6), 2578-2595.
- Shinozaki, K., & Yamaguchi-Shinozaki, K. (2000). Molecular responses to dehydration and low temperature: differences and cross-talk between two stress signaling pathways. *Current Opinion in Plant Biology*, 3(3), 217-223.
- Shinozaki, K., Yamaguchi-Shinozaki, K., & Seki, M. (2003). Regulatory network of gene expression in the drought and cold stress responses. Current opinion in plant biology, 6(5), 410-417.

- Shinozaki, K., & Yamaguchi-Shinozaki, K. (2007). Gene networks involved in drought stress response and tolerance. *Journal of Experimental Botany*, 58(2), 221-227.
- Shukla, R., Dube, A., & Koshy, E. (2014). Production of high quality embryogenic callus of rice. An International Quarterly J. Life Sciences, 9(3), 1077-1080.
- Singh, K. B., Foley, R. C., & Oñate-Sánchez, L. (2002). Transcription factors in plant defense and stress responses. *Current Opinion in Plant Biology*, 5(5), 430-436.
- Siripornadulsil, S., Traina, S., Verma, D. P. S., & Sayre, R. T. (2002). Molecular mechanisms of proline-mediated tolerance to toxic heavy metals in transgenic microalgae. *The Plant Cell*, 14(11), 2837-2847.
- Sivanesan, I., & Park, S. W. (2014). The role of silicon in plant tissue culture. *Frontiers in Plant Science*, *5*, 571.
- Skoog, F., & Miller, C. (1957). Chemical regulation of growth and organ formation in plant tissues cultured. Paper presented at the Vitro, Symp. Soc. Exp. Biol.
- Solano, R., Stepanova, A., Chao, Q., & Ecker, J. R. (1998). Nuclear events in ethylene signaling: a transcriptional cascade mediated by ETHYLENE- INSENSITIVE3 and ETHYLENE-RESPONSE-FACTOR1. *Genes & Development, 12*(23), 3703-3714.
- Sood, P., Bhattacharya, A., & Sood, A. (2011). Problems and possibilities of monocot transformation. *Biologia Plantarum*, 55(1), 1-15.
- Sridevi, G., Sabapathi, N., Meena, P., Nandakumar, R., Samiyappan, R., Muthukrishnan, S., et al. (2003). Transgenic indica rice variety Pusa Basmati 1 constitutively expressing a rice chitinase gene exhibits enhanced resistance to Rhizoctonia solani. Journal of Plant Biochemistry and Biotechnology, 12(2), 93-101.
- Stefanello, S., Dal Vesco, L. L., Ducroquet, J. P. H., Nodari, R. O., & Guerra, M. P. (2005). Somatic embryogenesis from floral tissues of feijoa (*Feijoa sellowiana* Berg). *Scientia horticulturae*, 105(1), 117-126.
- Stockinger, E. J., Gilmour, S. J., & Thomashow, M. F. (1997). Arabidopsis thaliana CBF1 encodes an AP2 domain-containing transcriptional activator that binds to the C-repeat/DRE, a cis-acting DNA regulatory element that stimulates transcription in response to low temperature and water deficit. Proceedings of the National Academy of Sciences, 94(3), 1035-1040.
- Stoger, E., Sack, M., Fischer, R., & Christou, P. (2002). Plantibodies: applications, advantages and bottlenecks. *Current Opinion in Biotechnology*, 13(2), 161-166.
- Streatfield, S. J. (2007). Approaches to achieve high-level heterologous protein production in plants. *Plant Biotechnology Journal*, 5(1), 2-15.

Streatfield, S. J., & Howard, J. A. (2003). Plant-based vaccines. International Journal

for Parasitology, 33(5), 479-493.

- Su, Y.-H., Liu, Y.-B., & Zhang, X.-S. (2011). Auxin–cytokinin interaction regulates meristem development. *Molecular Plant*, 4(4), 616-625.
- Sugimoto, K., Jiao, Y., & Meyerowitz, E. M. (2010). Arabidopsis regeneration from multiple tissues occurs via a root development pathway. Developmental Cell, 18(3), 463-471.
- Sugiyama, M. (2015). Historical review of research on plant cell dedifferentiation. Journal of Plant Research, 128(3), 349-359.
- Suzuki, Y., Hibino, T., Kawazu, T., Wada, T., Kihara, T., & Koyama, H. (2003). Extraction of total RNA from leaves of Eucalyptus and other woody and herbaceous plants using sodium isoascorbate. *Biotechniques*, *34*(5), 988-990, 992-983.
- Szabados, L., & Savoure, A. (2010). Proline: a multifunctional amino acid. *Trends in Plant Science*, 15(2), 89-97.
- Szweykowska, A. (1974). The role of cytokinin in the control of cell growth and differentiation in culture. tissue culture and plant science, 461-475.
- Tadasse, G., Algieri, B., Kalkuhl, M., & von Braun, J. (2016). Drivers and triggers of international food price spikes and volatility *Food Price Volatility and Its Implications for Food Security and Policy* (pp. 59-82): Springer.
- Talei, D., Valdiani, A., Abdullah, M. P., & Hassan, S. A. (2012). A rapid and effective method for dormancy breakage and germination of King of Bitters (*Andrographis* paniculata Nees.) seeds. Maydica, 57(2), 98-105.
- Talei, D., Valdiani, A., Maziah, M., & Mohsenkhah, M. (2013). Germination Response of MR 219 Rice Variety to Different Exposure Times and Periods of 2450 MHz Microwave Frequency. *The Scientific World Journal*.
- Talei, D., Valdiani, A., Maziah, M., Sagineedu, S. R., & Abiri, R. (2015). Salt stressinduced protein pattern associated with photosynthetic parameters and andrographolide content in Andrographis paniculata Nees. *Bioscience*, *Biotechnology, and Biochemistry*, 79(1), 51-58.
- Tamaki, H., Reguera, M., Abdel-Tawab, Y. M., Takebayashi, Y., Kasahara, H., & Blumwald, E. (2015). Targeting hormone-related pathways to improve grain yield in rice: a chemical approach. *PloS one*, *10*(6), e0131213.
- Terano, R., Mohamed, Z., & Din, N. S. Z. (2016). Determinants of Farmers' Adoption of Clearfield Production System in Malaysia. Agriculture and Agricultural Science Procedia, 9, 103-107.
- Thirugnanasambantham, K., Durairaj, S., Saravanan, S., Karikalan, K., Muralidaran, S., & Islam, V. I. H. (2015). Role of ethylene response transcription factor (ERF)

and its regulation in response to stress encountered by plants. *Plant Molecular Biology Reporter*, 33(3), 347-357.

- Thokozani, B. L., Zulu, D., Sileshi, C. W., Teklehaimanot, Z., Gondwe, D. S., Sarasan, V., & Stevenson, P. (2011). Seed germination and *in vitro* regeneration of the African medicinal and pesticidal plant, *Bobgunnia madagascariensis*. African Journal of Biotechnology, 10(32), 5959-5966.
- Thomashow, M. F. (1999). Plant cold acclimation: freezing tolerance genes and regulatory mechanisms. *Annual Review of Plant Biology*, 50(1), 571-599.
- To, K.-Y. (2000). Identification of differential gene expression by high throughput analysis. *Combinatorial Chemistry & High Throughput Screening*, 3(3), 235-241.
- Torero, M. (2016). Alternative Mechanisms to Reduce Food Price Volatility and Price Spikes: Policy Responses at the Global Level Food Price Volatility and Its Implications for Food Security and Policy (pp. 115-138): Springer.
- Torres, M. A., & Dangl, J. L. (2005). Functions of the respiratory burst oxidase in biotic interactions, abiotic stress and development. *Current Opinion in Plant Biology*, 8(4), 397-403.
- Tournier, B., Sanchez-Ballesta, M. T., Jones, B., Pesquet, E., Regad, F., Latché, A., *et al.* (2003). New members of the tomato ERF family show specific expression pattern and diverse DNA-binding capacity to the GCC box element. *FEBS Letters*, 550(1), 149-154.
- Tremblay, R., Wang, D., Jevnikar, A. M., & Ma, S. (2010). Tobacco, a highly efficient green bioreactor for production of therapeutic proteins. *Biotechnology Advances*, 28(2), 214-221.
- Tyagi, A., Mohanty, A., Bajaj, S., Chaudhury, A., & Maheshwari, S. (1999). Transgenic rice: a valuable monocot system for crop improvement and gene research. *Critical Reviews in Biotechnology*, 19(1), 41-79.
- Tzfira, T., & Citovsky, V. (2006). Agrobacterium-mediated genetic transformation of plants: biology and biotechnology. Current Opinion in Biotechnology, 17(2), 147-154.
- Tzfira, T., Jensen, C. S., Wang, W., Zuker, A., Vinocur, B., Altman, A., et al. (1997). Transgenic Populus tremula: a step-by-step protocol for its Agrobacteriummediated transformation. Plant Molecular Biology Reporter, 15(3), 219-235.
- Valderrama-Cháirez, M. L., Cruz-Hernández, A., & Paredes-López, O. (2002). Isolation of functional RNA from cactus fruit. *Plant Molecular Biology Reporter*, 20(3), 279-286.
- Valvekens, D., Van Montagu, M., & Van Lijsebettens, M. (1988). Agrobacterium tumefaciens-mediated transformation of Arabidopsis thaliana root explants by

using kanamycin selection. *Proceedings of the National Academy of Sciences*, 85(15), 5536-5540

- Vennapusa, A. R., Vemanna, R. S., Reddy, R., Babitha, K. C., Kiranmai, K., Nareshkumar, A., & Sudhakar, C. (2015). An efficient callus induction and regeneration protocol for a drought tolerant rice indica genotype AC39020. *Journal of Plant Sciences*, 3(5), 248-54.
- Verbruggen, N., & Hermans, C. (2008). Proline accumulation in plants: a review. *Amino Acids*, 35(4), 753-759.
- Visarada, K., & Sarma, N. (2002). Qualitative assessment of tissue culture parameters useful in transformation of indica rice. *Current Science*, 82(3), 343-346.
- Voesenek, L., Colmer, T., Pierik, R., Millenaar, F., & Peeters, A. (2006). How plants cope with complete submergence. *New Phytologist*, *170*(2), 213-226.
- Wahid, A., Noreen, A., Basra, S. M., Gelani, S., & Farooq, M. (2008). Priminginduced metabolic changes in sunflower (Helianthus annuus) achenes improve germination and seedling growth. *Bot Stud*, 49, 343-350.
- Wahyuni, S., Sinniah, U. R., Amarthalingam, R., & Yusop, M. K. (2003). Enhancement of seedling establishment in rice by selected growth regulators as seed treatment. *Jurnal Penelitian Pertanian Tanaman Pangan*, 22(1), 51-55.
- Wahyuni, S., Sinniah, U. R., Yusop, M. K., & Amarthalingam, R. (2013). Improvement of seedling establishment of wet seeded rice using GA3 and IBA as seed treatment. *Indonesian Journal of Agricultural Science*, 4(2).
- Walles, B. (1990). Buvat, R. 1989. Ontogeny, Cell Differentiation, and Structure of Vascular Plants. *Nordic Journal of Botany*, 9(5), 498-498.
- Wang, B., Guo, B., Xie, X., Yao, Y., Peng, H., Xie, C., et al. (2012). A novel histidine kinase gene, ZmHK9, mediate drought tolerance through the regulation of stomatal development in Arabidopsis. Gene, 501(2), 171-179.
- Wang, H.-M., Yin, W.-C., Wang, C.-K., & To, K.-Y. (2009). Isolation of functional RNA from different tissues of tomato suitable for developmental profiling by microarray analysis. *Botanical Studies*, 50, 115-125.
- Wang, H., Huang, Z., Chen, Q., Zhang, Z., Zhang, H., Wu, Y., et al. (2004). Ectopic overexpression of tomato JERF3 in tobacco activates downstream gene expression and enhances salt tolerance. Plant Molecular Biology, 55(2), 183-192.
- Wang, W.-S., Pan, Y.-J., Zhao, X.-Q., Dwivedi, D., Zhu, L.-H., Ali, J., et al. (2010). Drought-induced site-specific DNA methylation and its association with drought tolerance in rice (*Oryza sativa* L.). Journal of Experimental Botany, erq391.

Wang, Y., Xue, Y., & Li, J. (2005). Towards molecular breeding and improvement of

rice in China. Trends in Plant Science, 10(12), 610-614.

- Wani, S. H., Sanghera, G. S., & Gosal, S. S. (2011). An efficient and reproducible method for regeneration of whole plants from mature seeds of a high yielding Indica rice (*Oryza sativa* L.) variety PAU 201. *New Biotechnology*, 28(4), 418-422.
- Wassmann, R., Jagadish, S., Peng, S., Sumfleth, K., Hosen, Y., & Sander, B. (2008). Rice production and global climate change: scope for adaptation and mitigation activities. *Ceedin*, 67.
- Wessler, S. R. (2005). Homing into the origin of the AP2 DNA binding domain. *Trends* in *Plant Science*, 10(2), 54-56.
- Whitmore, A. P., & Whalley, W. R. (2009). Physical effects of soil drying on roots and crop growth. *Journal of Experimental Botany*, 60(10), 2845-2857.
- Woo, H.-J., Sohn, S.-I., Shin, K.-S., Kim, J.-K., Kim, B.-G., & Lim, M.-H. (2014). Expression of tobacco tocopherol cyclase in rice regulates antioxidative defense and drought tolerance. *Plant Cell, Tissue and Organ Culture (PCTOC), 119*(2), 257-267.
- Woodhead, M., Taylor, M. A., Davies, H. V., Brennan, R. M., & McNicol, R. J. (1997). Isolation of RNA from blackcurrant (*Ribes nigrum* L.) fruit. *Molecular Biotechnology*, 7(1), 1-4.
- Xia, C.-F., Yin, H., Borlongan, C. V., Chao, L., & Chao, J. (2004). Kallikrein gene transfer protects against ischemic stroke by promoting glial cell migration and inhibiting apoptosis. *Hypertension*, 43(2), 452-459.
- Xie, L., Tan, Z., Zhou, Y., Xu, R., Feng, L., Xing, Y., et al. (2014). Identification and fine mapping of quantitative trait loci for seed vigor in germination and seedling establishment in rice. *Journal of Integrative Plant Biology*, 56(8), 749-759.
- Xiong, L., Schumaker, K. S., & Zhu, J.-K. (2002). Cell signaling during cold, drought, and salt stress. *The Plant Cell*, 14(suppl 1), S165-S183.
- Xiong, L., & You, J. (2012). Use of *OsPP18* Gene in Controlling Rice Drought Resistance: Google Patents.
- Xu, J., Dolan, M. C., Medrano, G., Cramer, C. L., & Weathers, P. J. (2012). Green factory: plants as bioproduction platforms for recombinant proteins. *Biotechnology Advances*, 30(5), 1171-1184.
- Xu, Z.-S., Chen, M., Li, L.-C., & Ma, Y.-Z. (2011). Functions and application of the AP2/ERF transcription factor family in crop improvement. *Journal of Integrative Plant Biology*, 53(7), 570-585.
- Yadav, S. K. (2010). Cold stress tolerance mechanisms in plants. A review. Agronomy for Sustainable Development, 30(3), 515-527.

- Yang, Y., Shah, J., & Klessig, D. F. (1997). Signal perception and transduction in plant defense responses. *Genes & Development*, 11(13), 1621-1639.
- Yaseen, M., Ahmad, T., Sablok, G., Standardi, A., & Hafiz, I. A. (2013). Review: role of carbon sources for *in vitro* plant growth and development. *Molecular Biology Reports*, 40(4), 2837-2849.
- Yinxia, Z., & Te-chato, S. (2012). Callus induction and plantlet regeneration from mature embryos of indica rice (*Oryza Sativa* L.) cultivar Kra Dang Ngah. *Journal of Agricultural Technology*, 8(7), 2423-2433.
- Yoshiba, Y., Kiyosue, T., Nakashima, K., Yamaguchi-Shinozaki, K., & Shinozaki, K. (1997). Regulation of levels of proline as an osmolyte in plants under water stress. *Plant and Cell Physiology*, 38(10), 1095-1102.
- You, J., Zong, W., Hu, H., Li, X., Xiao, J., & Xiong, L. (2014). A STRESS-RESPONSIVE NAC1-regulated protein phosphatase gene rice protein phosphatase18 modulates drought and oxidative stress tolerance through abscisic acid-independent reactive oxygen species scavenging in rice. *Plant Physiology*, 166(4), 2100-2114.
- Yu, S., Zheng, W., Yu, W., Zhang, Y., Jiang, Q., & Zhao, Z. (2009). Formation mechanism of β-phase in PVDF/CNT composite prepared by the sonication method. *Macromolecules*, 42(22), 8870-8874.
- Yusof, F., Hui-Mean, F., Suhaila, J., & Yusof, Z. (2013). Characterisation of drought properties with bivariate copula analysis. *Water Resources Management*, 27(12), 4183-4207.
- Zarei, A., Körbes, A. P., Younessi, P., Montiel, G., Champion, A., & Memelink, J. (2011). Two GCC boxes and AP2/ERF-domain transcription factor ORA59 in jasmonate/ethylene-mediated activation of the *PDF1*. 2 promoter in *Arabidopsis*. *Plant Molecular Biology*, 75(4-5), 321-331.
- Zarembinski, T. I., & Theologis, A. (1997). Expression characteristics of OS-ACS1 and OS-ACS2, two members of the 1-aminocyclopropane-1-carboxylate synthase gene family in rice (Oryza sativa L. cv. Habiganj Aman II) during partial submergence. *Plant Molecular Biology*, *33*(1), 71-77.
- Zažímalová, E., Murphy, A. S., Yang, H., Hoyerová, K., & Hošek, P. (2010). Auxin transporters—why so many?. Cold Spring Harbor perspectives in biology, 2(3), a001552.
- Zeid, I., & Shedeed, Z. (2006). Response of alfalfa to putrescine treatment under drought stress. *Biologia Plantarum*, 50(4), 635-640.
- Zeng, L., Shannon, M., & Grieve, C. (2002). Evaluation of salt tolerance in rice genotypes by multiple agronomic parameters. *Euphytica*, *127*(2), 235-245.

- Zhang, J., & Kirkham, M. B. (1994). Drought-stress-induced changes in activities of superoxide dismutase, catalase, and peroxidase in wheat species. *Plant and Cell Physiology*, 35(5), 785-791.
- Zhang, G., Cui, Y., Ding, X., & Dai, Q. (2013). Stimulation of phenolic metabolism by silicon contributes to rice resistance to sheath blight. *Journal of Plant Nutrition* and Soil Science, 176(1), 118-124.
- Zhang, H., Huang, Z., Xie, B., Chen, Q., Tian, X., Zhang, X., et al. (2004). The ethylene-, jasmonate-, abscisic acid-and NaCl-responsive tomato transcription factor JERF1 modulates expression of GCC box-containing genes and salt tolerance in tobacco. Planta, 220(2), 262-270.
- Zhang, H., Yang, Y., Zhang, Z., Chen, J., Wang, X.-C., & Huang, R. (2008). Expression of the ethylene response factor gene *TSRF1* enhances abscisic acid responses during seedling development in tobacco. *Planta*, 228(5), 777-787.
- Zhang, Z., Li, F., Li, D., Zhang, H., & Huang, R. (2010). Expression of ethylene response factor *JERF1* in rice improves tolerance to drought. *Planta*, 232(3), 765-774.
- Zhao, Y., Ma, Q., Jin, X., Peng, X., Liu, J., Deng, L., et al. (2014). A novel maize homeodomain-leucine zipper (HD-Zip) I gene, Zmhdz10, positively regulates drought and salt tolerance in both rice and arabidopsis. Plant and Cell Physiology, 55(6), 1142-1156.
- Zheng, H., Lin, S., Zhang, Q., Lei, Y., & Zhang, Z. (2009). Functional analysis of 5' untranslated region of a TIR-NBS-encoding gene from triploid white poplar. *Molecular Genetics and Genomics*, 282(4), 381-394.
- Zhou, J., Tang, X., & Martin, G. B. (1997). The *Pto* kinase conferring resistance to tomato bacterial speck disease interacts with proteins that bind a *cis*-element of pathogenesis-related genes. *The EMBO Journal*, 16(11), 3207-3218.
- Ziemienowicz, A., Shim, Y.-S., Matsuoka, A., Eudes, F., & Kovalchuk, I. (2012). A novel method of transgene delivery into triticale plants using the Agrobacterium transferred DNA-derived nano-complex. *Plant Physiology*, 158(4), 1503-1513.
- Zulkarami, B., Razi, I. M., Halimi, M., Mondal, M., Panhwar, Q., & Islam, M. R. (2014). Effectiveness of different phytohormones on grain filling and yield of rice (Oryza sativa L.) under drought stress. *Journal of Food Agricutare and Environment*, 12, 697-700.
- Zuraida, A., Naziah, B., Zamri, Z., & Sreeramanan, S. (2011). Efficient plant regeneration of Malaysian indica rice MR 219 and 232 via somatic embryogenesis system. *Acta Physiologiae Plantarum*, 33(5), 1913-1921.