



***GROWTH PERFORMANCE AND PHYTOCHEMICALS OF  
Vernonia amygdalina Delile AS AFFECTED BY LIGHT INTENSITY,  
GROWING MEDIA, HARVEST TIME AND STORAGE DURATION***

**NURSUHAILI A. BAKAR**

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By

**NURSUHAILI A. BAKAR**

**Thesis Submitted to the School of Graduate Studies,  
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Degree of Master of Science**

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## DEDICATION

*Specially dedicated to my beloved late father, Mr. A. Bakar Sulong, my mother, Mrs. Lijah Yusof and my siblings, Mrs. Nurhima A. Bakar, Mrs. Norhanisah A. Bakar, Mr. Mohd Amin A. Bakar and Mr. Mohd Aimran A. Bakar, for their endless love, sacrifices, support, understandings, motivation, advice and encouragement.*



Abstract of thesis presented to the senate of universiti putra malaysia in fulfillment of the requirement for the degree of Master of Science

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OF *Vernonia amygdalina* Delile AS AFFECTED BY LIGHT INTENSITY,  
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**January 2019**

**Chair : Martini Mohammad Yusoff, PhD**  
**Faculty : Agriculture**

*Vernonia amygdalina* Del., locally known as pokok bismillah, is a potential herb with high medicinal value for the cure of many types of human ailments. It is found grown in a variety of habitats, causing wide variation in the growth performance and phytochemical content. In this regard, a field experiment and two laboratory experiments were conducted at Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia. The objectives of the study were to evaluate the effects of light intensity and suitable growing media on growth, physiology and biomass yield of *V. amygdalina*. The treatments for the field experiment comprised of three levels of light intensity (30%, 50% and 100% that equivalent to open field) and four types of growing media (soil, cocopeat, empty fruit bunch (EFB) and burnt paddy husk (BPH)). Seedlings were raised in a nursery and transferred to the field at 4 weeks after propagated. Plants were harvested at 18 weeks after transplanted (WAT) to the field. Results indicated that *V. amygdalina* grown under 50% light intensity in EFB growing medium had the highest biomass yield (490 kg ha<sup>-1</sup>) with a greater plant height (182 cm), number of branches (3 branches), stem diameter (22 mm), root surface area (3631 cm) and photosynthesis rate (22.6 μmol CO<sub>2</sub> m<sup>-2</sup>s<sup>-1</sup>) among the treatments imposed.

Experiment 2 was carried out to determine the suitable harvest time for optimum phytochemical contents in *V. amygdalina* leaves. Leaves samples from plants grown at the optimum requirement of the light intensity and growing media on *V. amygdalina* (50% light intensity, EFB) obtained from Experiment 1 were used in this study. The experiment comprised of six treatments of different harvest time (3, 6, 9, 12, 15 and 18 WAT). Results revealed that total phenolic content obtained were 91 and 84 mg GAE/g DW in leaf samples of plants harvested at 9 and 18 WAT, respectively. Similar pattern was found on total flavonoid content where it showed an increment from 3 to 9 WAT, then decreased sharply at week

12 but rising again until 18 WAT. Antioxidant activity (DPPH radical scavenging activity) was high from plants harvested at 9, 15 and 18 WAT. As harvest time increased the total chlorophyll content and plant nutrients content tended to increase. Heavy metals content were below the permissible limits of World Health Organization (WHO).

Experiment 3 was carried out to quantify the phytochemical contents of dried *V. amygdalina* leaves at different storage durations. The experiment consisted of seven treatments of different storage durations (0, 3, 6, 9, 12, 15 and 18 weeks). The results showed the 12-week storage duration of dried leaves of *V. amygdalina* achieved high stability of phytochemicals of total phenolic and flavonoid contents. There were decreases of 44% and 58% of total phenolic and flavonoid contents for 18-week storage duration, respectively. The nutrient content also decreased until 18 weeks of storage. However, *V. amygdalina* could be as a great source of nitrogen, potassium and magnesium since the contents were above the recommended daily allowance (RDA) set by WHO. Therefore, *V. amygdalina* is suitable to be harvested at 18 WAT in order to get the optimum phytochemicals content with minimum heavy metal contamination as well as high in biomass yield. Heavy metal contents and microbial contamination were at tolerable levels as they were below the permissible limits prescribed by WHO. Thus, *V. amygdalina* dried leaves were considered safe to be consumed until 18 weeks of storage.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Sarjana Sains

**PRESTASI PERTUMBUHAN DAN KANDUNGAN FITOKIMIA BAGI *Vernonia  
amygdalina* Delile YANG DIPENGARUHI OLEH INTENSITI CAHAYA,  
MEDIA TANAMAN, MASA TUAIAN DAN TEMPOH PENYIMPANAN**

Oleh

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**Fakulti : Pertanian**

*Vernonia amygdalina* Del., juga dikenali sebagai pokok bismillah, ialah sejenis herba yang berpotensi dengan nilai perubatan yang tinggi untuk merawat pelbagai penyakit manusia. Ianya ditemui hidup dalam pelbagai habitat yang menyebabkan variasi yang luas dalam pertumbuhan dan kandungan fitokimianya. Justeru, satu eksperimen lapangan dan dua eksperimen di makmal telah dijalankan di Jabatan Sains Tanaman, Fakulti Pertanian, Universiti Putra Malaysia. Eksperimen pertama dilaksanakan bagi menilai kesan intensiti cahaya dan kesesuaian media tanaman ke atas pertumbuhan, fisiologi, dan hasil biomas *V. amygdalina*. Rawatan untuk eksperimen di lapangan adalah terdiri daripada tiga tahap intensiti cahaya (30%, 50% dan 100% (bersamaan dengan lapangan terbuka)) dan empat jenis media tanaman (tanah, habuk kelapa, tandan buah kosong (EFB) dan sekam padi bakar (BPH). Anak pokok dibesarkan di nurseri dan dipindahkan ke lapangan 4 minggu selepas disemai. Pokok-pokok dituai pada minggu ke 18 selepas ditanam di lapangan (MST). Keputusan kajian menunjukkan *V. amygdalina* yang ditanam di bawah 50% intensiti cahaya dan menggunakan media tanaman EFB menunjukkan hasil biomas tertinggi (490 kg hektar<sup>-1</sup>) dengan ketinggian pokok yang lebih tinggi (182 cm), bilangan cabang (3 cabang), diameter batang (22 mm), luas permukaan akar (3631 cm) dan kadar fotosintesis (22.6  $\mu\text{mol m}^{-2}\text{s}^{-1}$ ).

Eksperimen 2 dijalankan bagi menentukan masa tuai yang sesuai bagi mendapatkan kandungan fitokimia yang optimum dalam daun *V. amygdalina*. Sampel daun daripada pokok yang ditanam dengan keperluan optimum intensiti cahaya dan media tanaman ke atas *V. amygdalina* (50% intensiti cahaya, EFB) diperolehi daripada daripada Eksperimen 1 telah digunakan dalam kajian ini. Eksperimen ini terdiri daripada enam rawatan masa tuai (3, 6, 9, 12, 15 dan 18 MST). Keputusan kajian mendapati kandungan jumlah kandungan fenolik yang diperolehi ialah 91 dan 84 mg GAE/g berat kering dalam sampel daun

daripada pokok yang dituai pada 9 dan 18 MST. Pola yang sama didapati ke atas jumlah kandungan flavonoid di mana ia menunjukkan peningkatan bermula pada tempoh 3 hingga 9 MST, kemudian turun mendadak pada 12 MST dan meningkat semula sehingga 18 MST. Aktiviti antioksidasi adalah tinggi pada pokok yang dituai pada 9, 15 dan 18 MST. Semakin meningkat masa tuaian, kandungan jumlah klorofil dan nutrien yang diambil oleh pokok juga meningkat. Kandungan logam berat berada di bawah tahap yang dibenarkan oleh Pertubuhan Kesihatan Sedunia (WHO).

Eksperimen 3 dilaksanakan bagi menentukan kuantiti kandungan fitokimia dalam daun kering *V. amygdalina* pada tempoh penyimpanan yang berbeza. Eksperimen ini mengandungi tujuh rawatan tempoh penyimpanan (0, 3, 6, 9, 12, 15 dan 18 minggu). Keputusan kajian menunjukkan tempoh penyimpanan selama 12 minggu bagi daun kering *V. amygdalina* mencapai kestabilan fitokimia yang tinggi bagi kandungan jumlah fenolik dan flavonoid. Terdapat penurunan sebanyak 44% dan 58% bagi kandungan jumlah fenolik dan flavonoid sehingga tempoh penyimpanan selama 18 minggu. Kandungan nutrien juga menurun sehingga minggu ke 18 penyimpanan. Walau bagaimanapun, *V. amygdalina* boleh dijadikan sebagai sumber yang terbaik bagi nitrogen, kalium dan magnesium memandangkan kandungannya adalah melebihi peruntukan harian yang ditetapkan oleh WHO. Justeru itu, *V. amygdalina* adalah sesuai dituai pada minggu ke 18 MST untuk mendapatkan kandungan fitokimia yang optimum dengan kontaminasi logam berat yang minimum sebagaimana hasil biomas. Kandungan logam berat dan kontaminasi mikrob berada di bawah paras maksima yang dibenarkan oleh WHO. Oleh itu, daun kering *V. amygdalina* adalah selamat untuk digunakan sehingga tempoh penyimpanan selama 18 minggu.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

|                   |  |
|-------------------|--|
| cm                | Centimetre   |
| °C                | Degree celsius   |
| $\pi$             | Pi   |
| $\beta$           | Beta   |
| g                 | Gram   |
| h                 | Hour   |
| $\mu$             | Micro  |
| kg                | Kilogram   |
| L                 | Litre  |
| MJ                | Megajoule  |
| m                 | Metre  |
| min               | Minute   |
| mg                | Miligram   |
| mL                | Millilitre   |
| mm                | Millimetre   |
| nm                | Nanometer  |
| r                 | Radius   |
| s                 | Second   |
| v/v               | Volume per volume  |
| w                 | Weight   |
| %                 | Percentage   |
| ABTS              | 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid) |
| Al                | Aluminium  |
| AlCl <sub>3</sub> | Aluminium chloride                                       |
| AOAC              | Association of Official Agricultural Chemists            |
| BPH               | Burnt paddy husk   |
| CO <sub>2</sub>   | Carbon dioxide   |
| Ca                | Calcium  |
| CFU               | Colony forming units                                     |
| Chl               | Chlorophyll  |
| Cu                | Copper   |
| DPPH              | 1, 1-diphenyl-2-picrylhydrazyl                           |
| DW                | Dry weight   |
| EFB               | Empty fruit bunch  |
| EPP               | Entry point project                                      |
| ETP               | The economic transformation programme                    |
| FAO               | Food and Agricultural Organization                       |
| Fe                | Iron   |
| Fi                | The fraction of incident PAR                             |
| FRAP              | Ferric reducing antioxidant power                        |

|                               |                                     |
|-------------------------------|-------------------------------------|
| GAE                           | Gallic acid equivalent              |
| GNI                           | Gross national income               |
| Gs                            | Stomatal conductance                |
| HCl                           | Hydrochloric acid                   |
| HNO <sub>3</sub>              | Nitric acid                         |
| H <sub>2</sub> O <sub>2</sub> | Hydrogen peroxide                   |
| K                             | Potassium                           |
| LAI                           | Leaf area index                     |
| LDPE                          | Low-density polyethylene            |
| LSD                           | Least significant difference        |
| Mg                            | Magnesium                           |
| NKEA                          | National Key Economic Area          |
| M                             | Molar                               |
| MPL                           | Maximum permissible limits          |
| N                             | North                               |
| N                             | Nitrogen                            |
| NA                            | Nutrient agar                       |
| NaNO <sub>2</sub>             | Sodium nitrite                      |
| NaOH                          | Sodium hydroxide                    |
| P                             | Phosphorus                          |
| PAR                           | Photosynthetically active radiation |
| Pb                            | Lead                                |
| PDA                           | Potato dextrose agar                |
| Pn rate                       | Net photosynthesis rate             |
| PSII                          | Photosystem II                      |
| QE                            | Quercetin                           |
| RDA                           | Recommended daily allowance         |
| ROS                           | Reactive oxygen species             |
| RuBP                          | Ribulose-1,5-bisphosphate           |
| TFC                           | Total flavonoid content             |
| TPC                           | Total phenolic content              |
| S                             | South                               |
| SE                            | Standard error                      |
| SLA                           | Specific leaf area                  |
| T                             | Transpiration rate                  |
| UV                            | Ultraviolet                         |
| WAT                           | Weeks after transplanted            |
| WHO                           | World Health Organization           |
| Zn                            | Zinc                                |

## CHAPTER 1

### INTRODUCTION

Malaysia has been ranked 4<sup>th</sup> in Asia as the most biodiverse country due to the richness of the nation's various plant species, many of which have high medicinal values (Mohammad Azmin et al., 2016). A herbal plant is a plant species which has high medicinal value commonly utilized in herbal medicine. In herbal medicine, almost all plant parts such as leaves, barks, roots, seeds, flowers and fruits are valuable for therapeutic purposes (Craig, 1999). Due to the potential of many species of herbs, the government under the Agriculture National Key Economic Area (NKEA) in The Economic Transformation Programme (ETP) has included the herbal industry as one of the Entry Point Project (EPPs) marked as a new source of economic growth and sustainability in Malaysia. The initial phase of this EPP was targeted at commercializing five types of herbs, namely Kacip Fatimah, Tongkat Ali, Hempedu Bumi, Misai Kucing and Dukung Anak (Ministry of Agriculture and Agro-based Industry, 2011). This initiative supports the country to become a potential hub for high-value herbal products in order to generate high returns for gross national income (GNI) by year 2020 (Malaysia Investment Development Authority, 2016).

One of the potential herbal plants is *Vernonia amygdalina* from the Family Asteraceae, native to Africa and widely distributed in Asia and commonly found growing wild in Malaysia and Singapore (Wong et al., 2013). It is known as "Bitter Leaf" or "Daun Bismillah" which contains phytochemical compounds that are useful for pharmacological purposes (Ramachendrin, 2015). The leaves are green in colour with bitter taste and have their own characteristic aroma (Udochukwu et al., 2015). This herbal plant has been confirmed to show anti-cancer, anti-malarial, antibacterial, antiparasitic, antihelmitic, antithrombotic and antidiabetic properties (Audu et al., 2012). Traditionally, the leaves of *V. amygdalina* are consumed as vegetable or liquid extract (tonics) to cure fever, hiccups, anemia, kidney and stomach disorders and to relieve toothache and gingivitis as it acts as antimicrobial agent (Imaga & Bamigbetan, 2013).

*Vernonia amygdalina* is commonly consumed in Cameroon which has a high market demand for its products. The price of dried leaves has been sold at USD 7.5 by Afriproduct per kg in 2010. Besides, there were also health products that have been commercialized, namely Diabetes 5 and EdoTide Plus (Yeap et al., 2010). According to Atangwho et al. (2013), this herb was only recently found to grow wild in Malaysia and it is traditionally used in the control of hypertension and diabetes mellitus, but no commercial product has been developed. However, many studies on its nutritional properties, pharmacological, medicinal and antioxidant value have been reported in recent years (Kadiri & Olawoye, 2016).

Since the consumption of herbal products has been on the increase nowadays, this means that the supply of raw materials of this plant needs to be increased and be easily obtained by entrepreneurs and manufacturers. Production of high yielding *V. amygdalina* with higher phytochemicals content requires optimum agronomic as well as effective postharvest handling practices. Both of these factors are crucial since it can influence the growth, yield of herbal plants and quality of medicinal products (Fonseca et al., 2006).

*Vernonia amygdalina* has been found to grow in nature close to rivers and forest margins, and is classified as a drought tolerant plant although adequate water supply is more suitable for its growth (Oyeyemi et al., 2017). Presently, information on the commercial cultivation of this plant is rather scarce and the herbal plant has not been well studied. In consideration of abundance of phytochemicals in this herb for pharmacological purposes, determination of suitable crop agronomy practices for better plant growth and yield of phytochemical content is therefore important. The agronomic practices are related to crop management such as choice of growing media in relation to abiotic factors such as light, water supply, temperature and others.

Light is one of the abiotic factors that has long been known to influence not only plant performance, but also the production of both primary and secondary metabolites (Hemm et al., 2004; Liu et al., 2002). Photosynthetically active radiation (PAR) in quantum flux density in active wavebands of 400-700 nm are the determinants of plant growth and crop yield (Biscoe & Gallanger, 1977; Williams et al., 1965). Normal plant growth needs optimal light irradiance because excessively high and low irradiances would result in photo-inhibition and light deficiency, respectively (Nur Faezah et al., 2015). In addition, the incident quantum flux intercepted by a plant canopy determines the photosynthetic rate, hence its biomass yield (Hippis et al., 1983).

Growing media is one of the planting components needed to cultivate *V. amygdalina* as this herb has yet to be grown commercially by the farmers. Even though this herb is said to thrive easily in any types of soil, better growth on humus-rich soil has been reported by Oyeyemi et al. (2017). Recently, organic matter is gaining interest as a growing media for sustainable agricultural practices in order to reuse the agricultural waste, ensure better environmental safety issues and promote soil conservation (Yadav et al., 2013). Agricultural organic wastes have been reported as a great source of natural organic fertilizer with their biological properties in plant physiology, such as improving plant yield and ensuring sufficient nutrient uptake by the roots (Zandonadi et al., 2013). Growing media, apart from enhancing soil biological, physical and chemical properties also aids in increasing the production of phytochemicals in the plants. Soils amended with agricultural wastes as a source of organic nutrients have the ability to improve the nutritional value and phytochemical content in crops (Theunissen et al., 2010).

Other than the requirement of optimum light and growing media, harvesting time also influences the phytochemical compounds in herbs (Raya et al., 2015). The production of phytochemicals in medicinal plants was found to vary at different harvesting times since it is also influenced by hormonal and enzyme balance, carbohydrates, nutrients and water content in the plants (Brasileiro et al., 2015).

Plants are good sources of natural phytochemicals of antioxidants and secondary metabolites (Ghasemzadeh et al., 2010). There are an abundance of bioactive compounds in *V. amygdalina* such as sesquiterpene lactones, phenol, flavonoids, alkaloids, tannins, cyanogenic glycosides, saponins, steroid, phytate, anthraquinone and oxalate which have been reported (Udochukwu et al., 2015; Yeap et al., 2010). It is well known that the antioxidant activity of medicinal plants and herbs are contributed more by the phenolic and flavonoid compounds (Ferreira et al., 2007; Rodrigo et al., 2006).

In addition, *V. amygdalina* is prompt to immediate physiological and physical deterioration when harvested (Ejike & Ndukwu, 2017). Thus, appropriate postharvest handling during storage of dried herbs as well as storage duration are the most fundamental factors in maintaining plant metabolic activity instead of using synthetic antioxidants to preserve the medicinal plants before processing the plant raw materials into herbal products (Mediani et al., 2014). However, the environmental factors such as light, temperature and also microbial contamination could affect the capacity of dried materials to maintain their quality and shelf life during storage (Masand et al., 2014).

As *V. amygdalina* is a new potential herb in pharmacology, understanding its underlying process in agronomic practices in the manipulation of light intensity and growing media is desirable for developing package technology for the commercial cultivation of this herb, including the suitable preference of plant harvesting time and storage duration in order to obtain desirable quality of phytochemical compounds. The specific objectives of the study on *V. amygdalina* were as follows:

1. To evaluate the effects of optimum light intensity and suitable growing media on physiological parameters, growth and yield of *V. amygdalina*.
2. To determine the suitable harvesting time for optimum phytochemical content of *V. amygdalina*.
3. To quantify the phytochemical content of *V. amygdalina* at different storage duration.

## REFERENCES

- Abdulkadir, A. R., Jahan, M. S., & Zawawi, D. D. (2015). Effect of chlorophyll content and maturity on total phenolic, total flavonoid contents and antioxidant activity of *Moringa oleifera* leaf (Miracle tree). *Journal of Chemical and Pharmaceutical Research*, 7(5): 1147-1152.
- Abad, M., Noguera, P., Puchades, R., Maquieira, A., & Noguera, V. (2002). Physico-chemical and chemical properties of some coconut dusts for use as a peat substitute for containerized ornamental plants. *Journal of Bioresource Technology*, 82: 241-245.
- Abdulrahman, D. K., Othman, R., & Mohd Saud, H. (2016). Effects of empty fruit bunch biochar and nitrogen-fixing bacteria on soil properties and growth of sweet corn. *Malaysia Journal of Soil Science*, 20: 177-194.
- Abdulrazzaq, H., Jol, H., Husni, A., & Abu-Bakar, R. (2015). Biochar from empty fruit bunches, wood, and rice husks: effects on soil physical properties and growth of sweet corn on acidic soil. *Journal of Agricultural Science*, 7(1): 192-200.
- Abu Bakar, R., Darus, S. Z., Kulaseharan, S., & Jamaluddin, N. (2011). Effects of ten year application of empty fruit bunches in an oil palm plantation on soil chemical properties. *Nutrient Cycling in Agroecosystem*, 89: 341–349.
- Abu-Darwish, M. (2009). Essential oils yield and heavy metals content of some aromatic medicinal plants. *Advances in Environmental Biology*, 3(3): 296–301.
- Adamson, H. Y., Chow, W. S., Anderson, J. M., Vesk, M., & Sutherland, M. W. (1991). Photosynthetic acclimation of *Tradescantia albiflora* to growth irradiance: morphological, ultrastructural and growth responses. *Journal of Plant Physiology*, 82: 353-359.
- Adaramoye, O. (2008). Lipid-lowering effects of methanolic extract of *Vernonia amygdalina* leaves in rats fed on high cholesterol diet. *Journal of Vascular Health and Risk Management*, 4(1): 235-241.
- Adelusi, A., & Aileme, J. (2006). Effects of light and nutrient stress on some growth parameters of cowpea (*Vigna radiata* (L.) Walp). *Research Journal of Botany*, 1(2): 95–103.
- Ademola, I., & Eloff, J. (2011). Anthelmintic activity of acetone extract and fractions of *Vernonia amygdalina* against *Haemonchus contortus* eggs and larvae. *Tropical Animal Health and Production*, 43(2): 521-527.
- Adesanoye, O. A., & Farombi, E. O. (2014). *In Vitro* antioxidant properties of methanolic leaf extract of *Vernonia amygdalina* Del. *Nigerian Journal Physiology Science*, 12: 91-101.

- Adiukwu, P., Kayanja, F., Nambatya, G., Rugera, S., Ezeonwumelu, J., Tanayen, J., Murokore, B., Twikirize, O., Twinomujuni, S., Byamugisha, D., & Imanirampa, L. (2013). Antipyretic and antinociceptive properties of the aqueous extract and saponin from an edible vegetable: *Vernonia amygdalina* leaf. *African Journal of Food, Agriculture, Nutrition and Development*, 13(2): 7588-7589.
- Affendy, H., Aminuddin, M., Azmy, M., Amizi, M., Assis, K., & Tamer, A. (2011). Effect of organic fertilizers application to the growth of *Orthosiphon stamineus* Benth. intercropped with *Hevea brasiliensis* Willd. and *Durio zibethinus* Murr. *International Journal of Agricultural Research*, 6(2): 180–187.
- Affendy, H., Aminuddin, M., Arifin, A., Mandy, M., Julius K., & Tamer, A. T. (2010). Effects of light intensity on *Orthosiphon stamineus* benth. seedlings treated with different organic fertilizers. *International Journal of Agricultural Resources*, 5: 201-207.
- Agbankpe, A. J., Bankole, S. H., Dougnon, T. J., Yehouenou, B., Hounmanou, Y. M. G., & Baba-Moussa, L. S. (2015). Comparison on nutritional values of *Vernonia amygdalina*, *Creteva adansonii* and *Sesamum radiatum*: Three main vegetables used in traditional medicine for the treatment of bacterial diarrhoe in Southern Benin (West Africa). *Journal of Food and Public Health*, 5(4): 144-149.
- Ahl, H. A. H. S., & Ali S. S. (2016). Growth, herb and essential oil of *Marrubium vulgare* as affected by phenological stages and planting dates. *Journal of Chemical and Pharmaceutical Research*, 8(5): 863–872.
- Ahmad, F., Zaidi, M., Sulaiman, N., & Abdul Majid, F. (2015). Issues and challenges in the development of the herbal industry in Malaysia. *Persidangan Kebangsaan Ekonomi Malaysia Ke-10 (PERKEM 10)*, 10(1): 227–238.
- Aimin, L., Shenghua, L., Xianjin, W., Min, H., Ronghui, G., Lin, W., & Anna, H. (2015). Influence of light intensity on the yield and quality of *Houttuynia cordata*. *Journal of Plant Production Science*, 18(4): 522-528.
- Ajayi-Oladosu, R. N., Dienye, H. E., & Agha, I. (2017). *Vernonia amygdalina* (Bitter leaf) extracts as preservative for catfish (*Clarias gariepinus*). *International Journal of Nutritional Science and Food Technology*, 3(1): 102–108.
- Akani, N. P., Jumbo, B., & Nwankwo, C.E. (2017). Effect of drying temperatures on mineral composition and bacterial populations of *Vernonia amygdalina* (Bitter leaf). *Research Journal of Food Science and Quality Control*, 3(2): 39-49.
- Akinyele, I.O., & Shokunbi, O. S. (2015). Comparative analysis of dry ashing and wet digestion methods for the determination of trace and heavy metals in food samples. *Food Chemistry*, 173: 682-684.

- Alabri, T. H., Al Musalami, A. H., Hossain, M. A., Weli, A. M., & Al-Riyami, Q. (2014). Comparative study of phytochemical screening, antioxidant and antimicrobial capacities of fresh and dry leaves crude plant extracts of *Datura metel* L. *Journal of King Saud Univ-Sci*, 26(3): 237-243.
- Alan, W. M. (2007). Coir dust, a viable alternative to peat moss. Available from: [http://flrec.ifas.ufl.edu/Hort/Environmental/Media\\_Nutrition/COIR%20potential.htm](http://flrec.ifas.ufl.edu/Hort/Environmental/Media_Nutrition/COIR%20potential.htm). [Accessed on 10 March 2019].
- Ali, A., Chong, C. H., Mah, S. H., Abdullah, L. C., Choong, T. S. Y., & Chua, B. L. (2018). Impact of storage conditions on the stability of predominant phenolic constituents and antioxidant activity of dried piper betle extracts. *Journal of Molecules*, 23(2): 1–15.
- Alifar, N., Ghehsareh, A. M., & Honarjoo, N. (2010). The effect of growth media on cucumber yield and its uptake of some nutrient elements in soilless culture. *Journal of Science and Technology of Greenhouse Culture*, 1(1):19-25.
- Aliyu, A. B., Musa, A. M., Abdullahi, M. S., Ibrahim, H., & Oyewale, A. O. (2011). Phytochemical screening and antibacterial activities of *Vernonia ambigua*, *Vernonia blumeoides* and *Vernonia oocephala* (Asteraceae). *Acta Poloniae Pharmaceutica-Drug Research*, 68(1): 67-73.
- Alwakeel, S. S. (2008). Microbial and heavy metals contamination of herbal medicines. *Research Journal of Microbiology*, 3(12): 683-691.
- Aktas, M., & Ates, A. (1998). Causes of Nutritional Disorders in Plants. Ostim-Ankara: Nurol Matbaacilik A.S.
- Ambo-rape, R., Lajus, D. L., & Schreider, M. J. (2011). Heavy metal impact on growth and leaf asymmetry of seagrass, *Halophila ovalis*. *Journal of Environmental Chemistry and Ecotoxicology*, 8(96): 149-159.
- Arbona, V., Manzi, M., Ollas, C., & de Gómez-Cadenas, A. (2013). Metabolomics as a tool to investigate abiotic stress tolerance in plants. *International Journal of Molecular Sciences*, 14: 4885–4911.
- Aresta, R. B., & Fukai, S. (1984). Effects of solar radiation on growth of cassava (*Manihot esculenta* crantz.). II. Fibrous root length. *Field Crops Research*, 9: 361–371.
- Arnold, J. B., Frensch, J., & Taylor, A. R. (2006). Influence of inorganic nitrogen and pH on the elongation of maize seminal roots. *Annals of Botany*, 97: 867-873.
- Asante, D. B., Effah-Yeboah, E., Barnes, P., Abban, H. A., Ameyaw, E. O., Boampong, J. N., & Dadzie, J. B. (2016). Antidiabetic effect of young and old ethanolic leaf extracts of *Vernonia amygdalina* : a comparative study. *Journal of Diabetes Research*, 1–13.
- Asiah, A., Mohd Razi, I., Mohd Khanif, Y., Marziah, M., & Shaharuddin, M.



- (2004). Physical and chemical properties of coconut coir dust and oil palm empty fruit bunch and the growth of hybrid heat tolerant cauliflower plant. *Journal of Tropical Agricultural Science*, 27(2): 121–133.
- Atanassova, M., Georgieva, S., & Ivancheva, K. (2011). Total phenolic and total flavonoid contents, antioxidant capacity and biological contaminants in medicinal herbs. *Journal of the University of Chemical Technology and Metallurgy*, 46(1): 81–88.
- Atangwho, I. J., Egbung, G. E., Ahmad, M., Yam, M. F., & Asmawi, M. Z. (2013). Antioxidant versus anti-diabetic properties of leaves from *Vernonia amygdalina* Del. growing in Malaysia. *Journal of Food Chemistry*, 141(4): 3428–3434.
- Audu, B. S., Omerinde, J. O., Gosomji, I. J., & Wazhi, P. E. (2017). Histopathological changes in the gill and liver of *Clarius gariepinus* exposed to acute concentrations of *Vernonia amygdalina*. *Annual Research International*, 14(1): 2576-2587.
- Audu, S. A., Alemika, E. T., Abdurraheem, R. O., Abdulkareem, S. S., Abdurraheem, R. B., & Ilyas, M. (2012). A study review of documented phytochemistry of *Vernonia amygdalina* (Family Asteraceae) as the basis for pharmacologic activity of plant extract. *Journal of Natural Sciences Research*, 2(7): 1–9.
- Awang, Y., Shaharom, S. S., Mohamad, R., & Selamat, A. (2009). Chemical and physical characteristics of cocopeat-based media mixtures and their effects on the growth and development of *Celosia cristata*. *American Journal of Agricultural and Biological Sciences*, 4(1): 63-71.
- Awang, Y., & Ismail, M. (1997). The growth and flowering of some annual ornamentals on coconut dust. *International Science Horticulture*, 450: 31-38.
- Bagchi, G. D., Dwivedi, P. D., Haider, F., Singh, S., Srivastava, S., & Chattopadhyay, S. (2003). Seasonal variation in vasicine content in *Adhatoda* species grown under north Indian plain conditions. *Journal of Medicinal and Aromatic Plant Sciences*, 25: 37-40.
- Bala Sambo, A., Jamiu Oyewole, O., Innocent Jonah, G., & Ponnak Ezekial, W. (2017). Histopathological changes in the gill and livers of *Clarias gariepinus* exposed to acute concentrations of *Vernonia amygdalina*. *Animal Research International*, 14(1): 2576–2587.
- Balsubramanian, P., & Chinnamuthu, C. R. (2017). Charred rice husk to improve the soil moisture retention and nutrient management under rainfed groundnut (*Arachis hypogaea* L.). *Journal of Agroecology and Natural Resources Management*, 4(9): 325-330.
- Barrett, G. E., Alexander, P. D., Robinson, J. S., & Bragg, N. C. (2016). Achieving environmentally sustainable growing media for soilless plant cultivation systems – A review. *Journal of Science Horticulture*, 212(9): 220–234.

- Bekhradi, F., Delshad, M., Kashi, A. A., Babalar, M., & Ikhani, S. (2014). Effect of plant density in some basil cultivars on yield and radiation use efficiency. *Journal of Biodiversity and Environmental Sciences*, 5(1): 91-96.
- Bergquist, S., Gertsson, U., Nordmark, L. Y., & Olsson, M. E. (2007). Effects of shade nettings, sowing time and storage on baby spinach flavonoids. *Journal of the Science of Food and Agriculture*, 87: 2464-2471.
- Beuchat, L. R., & Ryu, J. H. (1997). Produce handling and processing practices. *Emerging Infectious Diseases*, 3: 459-465.
- Biscoe, P. V., & Gallagher, J. N. (1977). Weather, dry matter production and yield. *The Conference on Environmental Effects on Crop Physiology*. Long Ashton Research Station, University of Bristol, London: Academic Press.
- Bittebiere, A. K., Renaud, N., Clement, B., & Mony, C. (2012). Morphological response to competition for light in the clonal *Trifolium repens* (fabaceae). *American Journal of Botany*, 99(4): 646-654.
- Bongers, F., Pons, T. L., & Rijkers, T. (2000). The effect of tree height and light availability on photosynthetic leaf traits of four neotropical species differing in shade tolerance. *Journal of Functional Ecology*, 14(1): 77-86.
- Bote, A. D., & Struik, P. (2011). Effects of shade on growth, production and quality of coffee (*Coffea arabica*) in Ethiopia. *Journal of Horticulture and Forestry*, 3(11): 336-341.
- Bowman, D. M. J. S., & Kirkpatrick, J. B. (1986). Establishment, suppression and growth of *Eucalyptus delegatensis* in multiaged forests. I. The effects of fire on mortality and seedling establishment. *Australian Journal of Botany*, 34:63-72.
- Boyd, R. S. (1998). Hyperaccumulation as a plant defence strategy. In Brooks RR, ed, *Plants That Hyperaccumulate Heavy Metals*. CAB International, Wallingford, UK.
- Brahmi, F., Mechri, B., Dhibi, M., & Hammami, M. (2015). Effect of growth stage and solvent extract on the antioxidant potential of olive leaves. *Journal of Plant Sciences*, 3(31): 1-7.
- Brasileiro, B. G., Leite, J. P. V., Casali, V. W. D., Pizziolo, V. R., & Coelho, O. G. L. (2015). The influence of planting and harvesting times on the total phenolic content and antioxidant activity of *Talinum triangulare* (Jacq.) Willd. *Journal Agronomy*, 37(2): 249.
- Bray, R. H., & Kurtz, L. T. (1945). Determination of total organic and available forms of phosphate in soils. *Soil Science*, 59: 39-45.
- Buajan, S., Jinfu, L. I., Zhongsheng, H., Xueping, F., & Muhammad, A. (2017).

The effect of light on micro-environment and specific leaf area within the gap, subtropical forest, China. *Journal of Botany*, 49(1): 273–282.

- Bugno, A., Buzzo, A. A., Nakamura, C. T., Pereira, T. C., Matos, D., & Pinto, T. J. A. (2005). Evaluation of microbial contamination in plant drugs. *Brazilian Journal of Pharmaceutical Sciences*, 41(4): 491-497.
- Campbell, G. S., & Norman, J. M. (1989). Plant canopies: their growth, form and function. Cambridge: Cambridge University Press.
- Casierra-Posada, F., & Avila-Leon, O. (2015). Shade tolerance of Marigold plants (*Calendula officinalis*). *UDCA Magazine of Current and Scientific Disclosure*, 18(1): 119–126.
- Cetinkaya, O., Sumer, A., Sungur, A., Adiloglu, S., & Akbulak, C. (2010). Fe, Cu, Zn, Mn availability and terrestrial distribution. Plant Nutrition and Fertilizer Congress, Izmir, Turkey, p. 695.
- Chanli, H. U. (2012). *Factors affecting phytochemical composition and antioxidant activity of ontario vegetable crops*. Master Thesis. The University of Guelph, Ontario, Canada.
- Chaves, T. P., Santana, C. P., & Veras, G. (2013). “Seasonal variation in the production of secondary metabolites and antimicrobial activity of two plant species used in Brazilian traditional medicine”. *African Journal of Biotechnology*, 12(8): 47-53.
- Chen, G., Wang, S., Huang, X., Hong, J., Du, L., Zhang, L. & Ye, L. (2015). Environmental factors affecting growth and development of Banlangen (*Radix Isatidis*) in China. *Journal of Plant Science*, 9(11), 421–426.
- Chiew, L. K., & Zaharah, A. R. (2002). The effects oil palm empty fruit bunches on oil palm nutrition and yield, and soil chemical properties. *Journal of Oil Palm Research*, 14(2): 1-9.
- Chouaieb, H., Ayadi, I., Zouari, S., Fakhfakh, N., Zaidi, S., & Zouari, N. (2012). Effect of phenological stage and geographical location on antioxidant activities of tunisian horehound: *Marrubium vulgare* L. (Lamiaceae). *Journal of Biologically Active Products from Nature*, 2(4): 232–238.
- Chua, I. Y. P., King, P. J. H., Ong, K. H., Sarbini, S. R., & Yiu, P. H. (2015). Influence of light intensity and temperature on antioxidant activity in *Premna serratifolia* L. *Journal of Soil Science and Plant Nutrition*, 15(3): 605-614.
- Claussen, J. W. (1996). Acclimation abilities of three tropical rainforest seedlings to an increase in light intensity. *Journal of Forest ecology and Management*, 80: 245-255.
- Clement, K., & Sankat, V. M. (1996). Shelf life of the green herb ‘shado beni’ (*Eryngium foetidum* L.) stored under refrigerated conditions. *Postharvest Biology and Technology*, 7(1–2): 109-118.

- Close, D. C., & McArthur, C. (2002). Rethinking the role of many plant phenolics- protections from photodamage not herbivores? *Journal of Oikos*, 99: 166–172.
- Cockshull, K. E., Graves, C. J., & Cave, C. R. J. (1992). The influence of shading on yield of glasshouse tomatoes. *Journal of Horticultural Science*, 67: 11-24.
- Coley, P. D., Bryant, J. P., & Chapin, F. S. (1985). Resource availability and plant antiherbivore defense. *Journal of Science*, 230: 895–899.
- Coombs, J., Hind, G., & Leegood, R. C. (1986). Analytical techniques. In *Techniques in bioproductivity and photosynthesis* 2nd ed. (Coombs, J., Hall, D. O., Long, D. O. and Scurlock, J. M. O., ed.) Pp. 223–240.
- Cornelius, E. (1999). Light interception and utilisation. 1st ed. *Plants in Action*. Pp. 1–6.
- Craig, W. J. (1999). Health-promoting properties of common herbs. *The American Journal of Clinical Nutrition*, 70(3): 491-499.
- Cutter, A. N. (2002). Microbial control by packaging: A review. *Critical Reviews In Food Science and Nutrition*, 42(2): 151-161.
- Dabrowski, P., Pawluskiewicz, B., Kalaji, H. M., & Baczewska, A. H. (2013). The effect of light availability on leaf area index, biomass production and plant species composition of park grasslands in Warsaw. *Plant soil Environment*, 59(12): 543-548.
- Damascos, M. A., Ronquim, C. C., & e Prado, C. H. B. A. (2005). Gas exchange and plant growth after defoliation on *Leandra lacunosa*, a cerrado woody species with continuous leaf production. *Brazilian Archives of Biology and Technology*, 48(6): 967-974.
- David, I. (2015). Entomotoxicant potential of bitter leaf, *Vernonia amygdalina* powder in the control of cowpea bruchid, *Callosobruchus maculatus* (Coleoptera: Chrysomelidae) infesting stored cowpea seeds. *Octa Journal of Environmental Research*, 3(3): 226–234.
- de Carvalho Goncalves, J. F., de Sousa Barreto, D. C., dos Santos, Jr, U. M., Fernandes, A. V., Barbosa Sampaio, P. D. T., & Buckeridge, M. S. (2005). Growth, photosynthesis and stress indicators in young rosewood plants (*Aniba rosaeodora* Ducke) under different light intensities. *Brazilian Journal of Plant Physiology*, 17: 325-334.
- de Freitas Aroujo, M. G., & Bauab, T. M. (2012). Microbial quality of medicinal plant materials. *INTECH*. Pp. 67-81.
- del-Toro-Sanchez, C. L., Gutierrez-Lomeli, M., Lugo-Cervantes, E., Zurita, F., Robles-Garcia, M. A., Ruiz-Cruz, S., Aguilar, J. A., Rio, J. A. M. D., & Guerrero-Medina, P. J. (2015). Storage effect on phenols and on the antioxidant activity of extracts from *Anemopsis californica* and inhibition

of elastase enzyme. *Journal of Chemicals*, 1-8.

- Demir, Z., & Gulser, C. (2015). Effects of rice husk compost application on soil quality parameters in greenhouse conditions. *Eurasian Journal of Soil Science*, 4(3): 185-190.
- Demmig-Adams, B., & Adams, W. W. (1992). Carotenoid composition in sun and shade leaves of plants with different life forms. *Plant Cell Environment*, 15: 411-419.
- Deveci, M., & Uzun, E. (2011). Determination of phenolic compounds and chlorophyll content of Spinach (*Spinacia oleracea* L.) at different growth stages. *Asian Journal of Chemistry*, 23(8): 3739-3743.
- Dias, J., Pimenta, J., Medri, M., Torres Boeger, M., & de Freitas, C. (2007). Physiological aspects of sun and shade leaves of *Lithraea molleoides* (Vell.) Engl. (Anacardiaceae). *Brazilian Archives of Biology and Technology*, 50(1): 91–99.
- Dinelli, E., & Lomboni, A. (1996). Metal distribution in plants growing on copper mine spoils in Northern Apennines, Italy: the evaluation of seasonal variation. *Applied Geochemistry*, 11: 375-85.
- Dixon, R.A., & Paiva, N. L. (1995). Stress-induced phenyl propanoid metabolism. *Plant Cell*, 7: 1085–1097.
- Doymaz, L. (2005). Dry behavior of green beans. *Journal of Food Engineering*. 69: 161-165.
- Dragana Jakovljevic, Z., Milan Stankovic, S., & Marina Topuzovic, D. (2013). Seasonal variability of *Chelidonium majus* L. secondary metabolites content and antioxidant activity. *EXCLI Journal*, 12: 260–268.
- Du, L., Liu, H., Yan, M., Li, J., & Li, J. (2017). Individual plasticity of the shade response of the invasive *Solidago canadensis* in China. *PLoS ONE*, 12(1): 1–11.
- Duda, S. C., Marghitas, L. A., Dezmirean, D., Duda, M., Margaoan, R., & Bobis, O. (2015). Changes in major bioactive compounds with antioxidant activity of *Agastache foeniculum*, *Lavandula angustifolia*, *Melissa officinalis* and *Nepeta cataria*: Effect of Harvest Time and Plant Species. *Industrial Crops and Products*, 77: 499-507.
- Egharevba, C., Osayemwenre, E., Imieje, V., Ahomafor, J., Akunyuli, C., & Udu-cosi, A. A. (2014). Significance of bitter leaf (*Vernonia amagdalina*) in tropical diseases and beyond : a review. *Malaria Chemotherapy Control and Elimination*, 3(1): 1–10.
- Ejike, C. E., & Ndukwu, M. C. (2017). Pre-harvest and post-harvest factors affecting bioactive compounds from *Vernonia amygdalina* (Del.). *Research Journal of Medicinal Plants*, 11(2): 32-40.

- Elizabeth, A. O. (2013). Shade conditions prevents anthraquinone production in leaves of *Vernonia amygdalina* plants. *International Journal of Science and Nature*, 4(4): 713-715.
- Erasto, P., Grierson, D. S., & Afolayan, A. J. (2007). Antioxidant constituents in *Vernonia amygdalina* leaves. *Pharmaceutical Biology*, 45(3), 195–199.
- Erasto, P., Grierson, D. S., & Afolayan, A. J. (2006). Bioactive sesquiterpene lactones from the leaves of *Vernonia amygdalina*. *Journal of Ethnopharmacology*, 106(1): 117-120.
- Eruygur, N., Turgut, K., Yur, S. & Ozek, T. (2018). The effect of the plant age and growth period on the nutritional substance, chlorophyll and steviol glycoside rates in *Stevia (Stevia rebaudiana Bertonii)* leaves. *Communications in Soil Science and Plant Analysis*, 49(1): 1-12.
- Erwan, Ismail, M. R., Sariah, M., Saud, H. M., Habib, S. H., Kausar, H., & Naher, L. (2013). Effect of oil palm frond compost amended coconut coir dust soilless growing media on growth and yield of cauliflower. *International Journal of Agriculture Biology*, 15: 731–736.
- Eyong, E. U., Agiang, M. A., Atangwho, I. J., Iwara, I. A., Odey, M. O., & Ebong, P. E. (2011). Phytochemicals and micronutrients composition of root and stem bark extracts of *Vernonia amygdalina* Del. *Journal of Medicine and Medical Science*, 2(6): 900-903.
- Ezuruike, U. F., & Prieto, J. M. (2014). The use of plants in the traditional management of diabetes in Nigeria: Pharmacological and toxicological considerations. *Journal of Ethnopharmacology*, 155(2): 857–924.
- Fageria, N. K. (2012). Role of soil organic matter in maintaining sustainability of cropping systems. *Communications in Soil Science and Plant Analysis*, 43(16): 2063-2113.
- Fazlil Ilahi, W. F., & Ahmad, D. (2017). A study on the physical and hydraulic characteristics of cocopeat perlite mixture as a growing media in containerized plant production. *Jurnal Sains Malaysiana*, 46(6): 975–980.
- Fawzy, Z. F., Behairy, A. G., & Shehata, S. A. (2005). Effect of potassium fertilizer on growth and yield of sweet pepper plants (*Capsicum annuum* L.) *Egypt Journal of Agriculture Research*, 2(2): 599-610.
- Fedoruk, A. (2011). *Development of a quantitative microbial risk assessment model for foodborne pathogens in herbs and spices*. Master Thesis. University of Guelph, Ontario, Canada.
- Feng, Y., & van Kleunen, M. (2014). Responses to shading of naturalized and non-naturalized exotic woody species. *Annals of Botany*, 114: 981-988.
- Fernandes, V., de Almeida, L. B., da S. Feijo, E. V. R., da C. Silva, D., de Oliveira, R. A., Mielke, M. S., & do B. Costa, L. C. (2013). Light intensity

on growth, leaf micromorphology and essential oil production of *Ocimum gratissimum*. *Brazilian Journal of Pharmacognosy*, 23(3): 419-424.

- Ferreira, I. C. F. R., Baptista, P., Vilas-Boas, M., & Barros, L. (2007). Free-radical scavenging capacity and reducing power of wild edible mushrooms from northeast Portugal: individual cap and stipe activity. *Journal of Food Chemistry*, 100: 1511-1516.
- Fini, A., Ferrini, F., Frangi, P., Amoroso, G., & Giordano, C. (2010). Growth, leaf gas exchange and leaf anatomy of three ornamental shrubs grown under different light intensities. *European Journal of Horticulture Science*, 75(3): 111-117.
- Fonseca, J. M., Rushing, J. W., Rajapakse, N. C., Thomas, R. L., & Riley, M. B. (2006). Potential implications of medicinal plant production in controlled environments: The case of feverfew (*Tanacetum parthenium*). *Journal of Horticulture Science*, 41(3): 531–535.
- Garriga, M., Retamales, J. B., Romero-Bravo, S., Caligari, P. D. S. & Lobos, G. A. (2014). Chlorophyll, anthocyanin, and gas exchange changes assessed by spectroradiometry in *Fragaria chiloensis* under salt stress. *Journal of Integrative Plant Biology*, 56(5): 505-515.
- Gaur, A., & Adholeya, A. (2004). Prospects of Arbuscular Mycorrhizal Fungi in Phytoremediation of Heavy Metal Contaminated Soils. *Current Science*, 86(4): 528–534.
- Gezahan, A. M. (2016). *Integrated nutrient management for maize-soybean cropping system*. PhD Thesis. Universiti Putra Malaysia, Serdang, Malaysia.
- Ghasemzadeh, A., Jaafar, H. Z. E., Rahmat, A., Wahab, P. E. M., & Halim, M. R. A. (2010). Effect of different light intensities on total phenolics and flavonoids synthesis and anti-oxidant activities in young ginger varieties (*Zingiber officinale* Roscoe). *International Journal of Molecular Sciences*, 11(10): 3885–3897.
- Ghehsareh-Mohammadi, A., & Shirani, M. (2012). Evaluation of chemical properties of Date-palm waste as culture media and its effect on number and yield of Tomato. *Annual Research and Review in Biology*, 5(1): 18–24.
- Gilbert, G. A., Knight, J. D., Vance, C. P., & Allan, D. L. (2000). Proteoid root development of phosphorus deficient lupin is mimicked by auxin and phosphonate. *Annals of Botany*, 85: 921–928.
- Gomez, S., Guenni, O., & Bravo de Guenni, L. (2012). Growth, leaf photosynthesis and canopy light use efficiency under differing irradiance and soil N supplies in the forage grass *Brachiara decumbens* Stapf. *Grass and Forage Science*, 68: 395-407.
- Gratani, L. (2014). Plant phenotypic plasticity in response to environmental

- factors. *Advances in Botany*, 1(1): 1-17.
- Gulcin, İ. (2006). Antioxidant activity of caffeic acid (3,4-Dihydroxycinnamic Acid). *Toxicology*, 217 (2): 213–220.
- Haefele, S. M., Knoblauch, C., Gummert, M., Konboon, Y., & Koyama, S. (2009). Black carbon (biochar) in rice-based systems: characteristics and opportunities. Springer Netherlands, 445-463.
- Haferkamp, M. (1988). Environmental factors affecting plant productivity. In *Fort Keogh Research Symposium*, Pp. 132.
- Halt, M. (1998). Molds and mycotoxins in herb tea and medicinal plants. *European Journal of Epidemiology*, 14: 269-274.
- Hamdan, A.B., Mohd Tayeb, D., & Ahmad Tarmizi, M. (2006). Effects of empty fruit bunch application in oil palm on a bris soil. *Oil Palm Bulletin*, 52(5): 48-58.
- Hanum, C., Ginting, J., Rauf, A., & Habibi Nasution, A. R. (2017). Growth and quision N, P, K of oil palm seedling on plant media made from solid decanter and oil palm empty fruit bunches. *International Journal of Science and Research Methodology*, 5(3): 9-22.
- Hassan, A., Mohamad, M., Azmy, M., Ayob, M. A., Kamu, A., Tabet, A., & Tamer. (2011). Effect of organic fertilizers application to the growth of *Orthosiphon stamineus Benth.* intercropped with *Hevea Brasiliensis Willd.* and *Durio zibethinus Murr.* *International Journal of Agricultural Research*, 6 (2): 180-187.
- Hebert, Y., Guingo, E., & Loudet, O. (2001). The response of root/shoot partitioning and root morphology to light reduction in maize genotypes. *Journal of Crop Science*, 41: 363-371.
- Hemm, M.R., Rider, S.D., Ogas, J., Murry, D.J. & Chapple, C. (2004). Light induces phenylpropanoid metabolism in Arabidopsis roots. *The Plant Journal*, 38: 765-778.
- Hipps, L. E., Aspar, G., & Kanemasu, E. T. (1983). Assessing the interception of photosynthetically active radiation in winter wheat. *Agricultural Meteorology*, 28: 253-259.
- Hortensteiner, S., & Krautler, B. (2011). Chlorophyll breakdown in higher plants. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1807(8): 977–988.
- Huang, C. J., Wei, G., Jie, Y. C., Xu, J. J., Anjum, S. A., & Tanveer, M. (2016). Effect of shade on plant traits, gas exchange and chlorophyll content in four Ramie cultivars. *Photosynthetica*, 54(3): 390–395.
- Hue, S. M., Boyce, A. N., & Somasundram, C. (2011). Influence of growth stage and variety on the pigment levels in *Ipomoea batatas* (Sweet potato) leaves. *African Journal of Agricultural Research*, 6(10): 2379–2385.



- Ibrahim, N. A. (2017). *Extraction of antioxidant compounds from Vernonia amygdalina* (Asteraceae). Degree thesis. Universiti Putra Malaysia, Serdang, Selangor, Malaysia.
- Ibrahim, M. H., Jaafar, H. Z. E., Karimi, E., & Ghasemzadeh, A. (2013). Impact of organic and inorganic fertilizers application on the phytochemical and antioxidant activity of Kacip Fatimah (*Labisia pumila* Benth). *Molecules*, 18: 10973-10988.
- Ibrahim, M. H., & Jaafar, H. Z. E. (2012). Reduced photoinhibition under low irradiance enhanced Kacip fatimah (*Labisia pumila* Benth) secondary metabolics, phenyl alaninelyase and antioxidant activity. *International Journal of Molecular Sciences*, 13: 5290-5306.
- Ibrahim, M. H., & Jaafar, H. Z. E. (2011). Photosynthetic capacity, phytochemical efficiency and chlorophyll content of three varieties of *Labisia pumila* Benth. exposed to open field and greenhouse growing conditions. *Acta Physiologia Plant*, 33: 2179-2185.
- Idris, O. A., Wintola, O. A., & Afolayan, A. J. (2017). Phytochemical and antioxidant activities of *Rumex crispus* L. in treatment of gastrointestinal helminths in Eastern Cape Province, South Africa. *Asian Pasific Journal of Tropical Biomedicine*, 7(12): 1071-1078.
- Ijeh, I. I., & Ejike, C. E. C. C. (2011). Current perspectives on the medicinal potentials of *Vernonia amygdalina* Del. *Journal of Medicinal Plants Research*, 5(7): 1051–1061.
- Imaga, N. O. A., & Bamigbetan, D. O. (2013). In vivo biochemical assessment of aqueous extracts of *Vernonia amygdalina* (Bitter leaf). *International Journal of Nutrition and Metabolism*, 5(2): 22-27.
- Inugraha, Maghfoer, M. D., & Widaryanto, E. (2014). Response of stevia (*Stevia rebaudiana* Bertoni M) to nitrogen and potassium fertilization. *IOSR Journal of Agriculture and Veterinary Science*, 7(10): 47-55.
- Isayenkov, S. V. (2014). Plant vacuoles: Physiological roles and mechanisms of vacuolar sorting and vesicular trafficking. *Cytology and Genetics*, 48(2): 127-137.
- Izyani, R., & Mohamed Zabawi, A. G. (2010). Cocopeat: potential growing media for Kacip Fatimah. Proceedings national coconut conference 2009: opportunities for a sunrise industry: Damai Laut, Perak (Malaysia), 28-30 Jul 2009. Pp. 201-204.
- Iwu, M. (2013). *Handbook of African Medicinal Plants, Second Edition*. Hoboken: Taylor and Francis.
- Jaafar, H. Z. E., Ibrahim, M. H., & Philip, E. (2009). Leaf gas exchange properties of three varieties of *Labisia pumila* Benth. under greenhouse conditions. *Journal of Tropical Plant Physiology*, 3: 16-24.

- Jain, A., Poling, M. D., Karthikeyan, A. S., Blakeslee, J. J., Peer, W. A., Titapiwatanakun, B., Murphy, A. S., & Raghothama, K. G. (2007). Differential effects of sucrose and auxin on localized phosphate deficiency-induced modulation of different traits of root system architecture in Arabidopsis. *Plant Physiology*, 144: 232–247.
- Jaishankar, M., Tseten, T., Anbalagan, N., Mathew, B.B., & Beeregowda, K. N. (2014). Toxicity, mechanism, and health effects of some heavy metals. *Journal of Interdisciplinary Toxicology*, 7(2): 60-72.
- James, S. A., & Bell, D. T. (2000). Influence of light availability on leaf structure and growth of two *Eucalyptus globulus* ssp. *globulus* provenances. *Tree Physiology*, 20: 1007-1018.
- Jensen, P. J., Hangarter, R. P., & Estelle, M. (1998). Auxin transport is required for hypocotyl elongation in light-grown but not dark-grown Arabidopsis. *Plant Physiology*, 116: 455-462.
- Jimenez-Zamora, A., Delgado-Andrade, C., Rufian-Henares, J. A. (2016). Antioxidant capacity, total phenols and color profile during the storage of selected plants used for infusion. *Food Chemistry*, 199: 339-346.
- Joanne, W., Linda, S., & Chris, W. (2014). Food-borne and waterborne diseases. *Prescott's Microbiology*, 9th ed. McGraw Hill Education. Pp. 915-924.
- Jonathan, S. G., Onile, O. G., Asemoloye, M. D. Z., & Omotayo, O. O. (2018). Effect of storage time on nutrient, biodegrading fungi and aflatoxin contents in Bitter leaf (*Vernonia amygdalina*) and JUTE (*Corchorus olitorius*). *Journal of Microbial and Biochemical Technology*, 10(2): 49-55.
- Jonathan, S. G., Ajayi, I., & Omitade, Y. (2011). Nutritional compositions, fungi and aflatoxins detection in stored 'gbodo' fermented (*Dioscorea rotundata*) and 'elubo ogede' fermented (*Musa parasidiaca*) from south western Nigeria. *African Journal of Food Sciences*, 5: 105-110.
- Judd, L., Jackson, B., & Fonteno, W. (2015). Advancements in root growth measurement technologies and observation capabilities for container-grown plants. *Journal of Plants*, 4(3): 369–392.
- Kacar, B., & Inal, A. (2008). *Plant Analysis*. Ankara: Nobel Publication Distribution.
- Kadiri, O., & Olawoye, B. (2016). *Vernonia amygdalina*: An underutilized vegetable with nutraceutical potentials- a review. *Turkish Journal of Agriculture- Food Science and Technology*, 4(9): 763-768.
- Kaewseejan, N., & Siriamornpun, S. (2015). Bioactive components and properties of ethanolic extract and its fractions from *Gynura procumbens* leaves. *Industrial Crops and Products*, 74: 271-278.
- Kambizi, L., & Afolayan, A. (2001). An ethnobotanical study of plants used for

the treatment of diabetes in the warangal district. *Journal of Ethnopharmacology*, 77(1): 5-9.

Kancheva, R., Borisova, D., & Georgiev, G. (2014). Chlorophyll assessment and stress detection from vegetation optical properties. *Ecological Engineering and Environment Protection*, 1: 34-43.

Kasote, D. M., Katyare, S. S., Hedge, M. V., & Bae, H. (2015). Significance of antioxidant potential of plants and its relevance to therapeutic applications. *International Journal of Biological Sciences*, 11(8): 982-991.

Kavitha, B., Jothimani P., & Rajannan, G. (2013). Empty fruit bunch- A potential organic manure for agriculture. *International Journal of Science, Environment and Technology*, 2(5): 930-937.

Kayode, J. (2004). Eco-physiological studies on *Vernonia amygdalina* in Ekiti State, Nigeria. *Pakistan Journal of Scientific and Industrial Research*, 47: 227-230.

Kiczorowska, B., Klebaniuk, R., Bakowski, M., & Al-Yasiry, M. H. (2015). Culinary herbs- the nutritive value and content of minerals. *Journal of Elementology*, 20(3): 599-608.

Kenneth, C. (1989). The herb, spice and medicinal plant digest, 7(3): 1-5.

Khadka, R. B., Marasini, M., Rawal, R., Gautam, D. M., & Acedo Jr, A. L. (2017). Effects of variety and postharvest handling practices on microbial population at different stages of the value chain of fresh tomato (*Solanum lycopersicum*) in western Terai of Nepal. *BioMed Research international*, Pp. 6.

Khalafalla, M. M., Abdellatef, E., Daffalla, H. M., Nassrallah, A. A., Aboul-Enein, K. M., Lightfoot, D. A., & El-Shemy, H. A. (2009). Antileukemia activity from root cultures of *Vernonia amygdalina*. *Journal of Medicinal Plants Research*, 3(8): 556-562.

Kirigia, D., Winkelmann, T., Kasili, R., & Mibus, H. (2018). Development stage, storage temperature and storage duration influence phytonutrient content in cowpea (*Vigna unguiculata* L. Walp.). *Heliyon*, 4(6): 1-24.

Kitao, M., Lei, T.T., Koike, T., Tobita, H., & Maruyama, Y. (2000). Susceptibility to photoinhibition of three deciduous broadleaf tree species with different successional traits raised under various light regimes. *Plant Cell Environment*, 23: 81-89.

Kopjar, M., Orsolcic, M., & Pilizota, V. (2014). Anthocyanins, phenols, and antioxidant activity of sour cherry puree extracts and their stability during storage. *International Journal of Food Properties*, 17(6): 1393-1405.

Korir, R. K. C. (2017). *Microbial and heavy metal contaminations in selected herbal medicinal products sold in Nairobi, Kenya*. Thesis. University of

Nairobi, Kenya.

- Koyama, K., Ikeda, H., Poudel, P.R. & Goto-Yamamoto, N. (2012). Light quality affects flavonoid biosynthesis in young berries of Cabernet sauvignon grape. *Phytochemistry*, 78: 54-64.
- Krishnaiah, D., Sarbatly, R., & Bono, A. (2007). Phytochemical antioxidants for health and medicine: A move towards nature. *Biotechnology and Molecular Biology Reviews*, 1: 97-104.
- Kuczynska, P., Jemiola-Rzeminska, M., & Strzalk, K. (2015). Photosynthetic pigments in diatoms. *Marine Drugs*, 13(9): 5847–5881.
- Kumar, D. S. (2013). Efficient method of storage of dry herbs. *Journal for Drugs and Medicines*, 5(2): 1-3.
- Kumar, R., Sharma, S., & Pathania, V. (2013). Effect of shading and plant density on growth, yield and oil composition of clary sage (*Salvia sclarea* L.) in north western Himalaya. *Journal of Essential Oil Research*, 25(1): 23–32.
- Kumar, R. N., Chakraborty, S., & Kumar, J. I. N. (2012). Influence of light and developmental stages on active principles of *Andrographis paniculata* (Burm.f.) Wall. ex Nees. *Indian Journal of Sciences Research*, 3(1): 91–95.
- Kupper, H., Mijovilovich, A., Meyer-Klaucke, W., & Kroneck, P. M. H. (2004). Tissue- and age-dependent differences in the complexation of cadmium and zinc in the cadmium/zinc hyperaccumulator *Thlaspi caerulescens* (Ganges ecotype) revealed by x-ray absorption spectroscopy. *Journal of Plant Physiology*, 134(2): 748–757.
- Lambers, H., Chapin, F. S., & Pons, T. L. (1998). Plant physiological ecology. New York (NY): Springer.
- Larson, R.A. (1998). The antioxidants of higher plants. *Journal of Phytochemistry*. 27: 969–978.
- Lavres Junior, J., Deus Gomesdos Santos Junior, J., & Monteiro, F. A. (2010). Nitrate reductase activity and spad readings in leaf tissues of guinea grass submitted to nitrogen and potassium rates. *Brazilian Journal of Soil Sciences*, 34: 801-809.
- Lawlor D. W. (2002). Carbon and nitrogen assimilation in relation to yield: mechanisms are the key to understanding production systems, *Journal of Experimental Botany*, 53: 773-787.
- Law-Ogbomo, K., & Enobakhare, D. (2007). The use of leaf powders of *Ocimum gratissimum* and *Vernonia amygdalina* for the management of *Sitophilus oryzae* (Lin.) in stored rice. *Journal of Entomology*, 4(3): 253–257.
- Lee, C. H., Kwon, O. K., & Kim, Y. J. (2005). Rooting characteristics of stem tip

cuttings in *Ardisia pusilla* as influenced by cutting stage, rooting medium, temperature, and plant growth regulator pretreatment. *Journal of the Korean Society for Horticultural Science*, 46(3):217-224.

- Lee, T.C., & Chichester, C.O. (1974). The influence of harvest time on nutritional value. In: *Nutritional qualities of fresh fruits and vegetables*, (Philip, L.W. and Nancy, S.R.D., eds.), p. 111. New York: Futura Publishing Company.
- Legner, N., Fleck, S., & Leuschner, C. (2014). Within-canopy variation in photosynthetic capacity, SLA and foliar N in temperate broad-leaved trees with contrasting shade tolerance. *Trees*. 28(1): 263-280.
- Leong, T. L. (2015). Effects of rice husk ash (RHA) produced from different temperature on the performance of concrete. Bachelor Thesis, Universiti Tunku Abdul Rahman.
- Li, A., Li, S., Wu, X., Zhang, J., He, A., Zhao, G., & Yang, X. (2016). Effect of light intensity on leaf photosynthetic characteristics and accumulation of flavonoids in *Lithocarpus litseifolius* Chun. (Fagaceae). *Journal of Forestry*, 6(5): 445-459.
- Li, Y., Johnson, D. A., Su, Y., Cui, J., & Zhang, T. (2005). Specific leaf area and leaf dry matter content of plants growing in Sand Dunes. *Botanical Bulletin- Academia Sinica Taipei*, 46: 127-134.
- Lichtenthaler, H. K., Ac, A., Marek, M. V., Kalina, J., & Urban, O. (2007). Differences in pigment composition, photosynthetic rates and chlorophyll fluorescence images of sun and shade leaves of four tree species. *Plant Physiology and Biochemistry*, 45(8): 577-588.
- Lim, Y. Y. & Murtijaya, J. (2007). Antioxidant properties of *Phyllanthus amarus* extracts as affected by different drying methods. *LWT-Food Science and Technology*, 40: 1664-1669.
- Liu, W., & Su, J. (2016). Effects of light acclimation on shoot morphology, structure, and biomass allocation of two *Taxus* species in southwestern China. *Scientific reports*, 6, 35384, DOI: 10.1038/srep35384.
- Liu, Y., Dawson, W., Prati, D., Haeuser, E., Feng, Y., & van Kleunen, M. (2016). Does greater specific leaf area plasticity help plants to maintain a high performance when shaded. *Journal of Business Ethics*, 118(7): 1329–1336.
- Liu, C. W., Sung, Y., Cheng, B. C., & Lai, H. Y. (2014). Effects of nitrogen fertilizers on the growth and nitrate content of lettuce (*Lactuca sativa* L.). *International Journal of Environmental Research Public Health*, 11(4): 4427-4440.
- Liu, W. T., Ni, J. C., & Zhou, Q. X. (2013). Uptake of heavy metals by trees. *Prospects*, p 105.

- Liu, T., Song, F., Liu, S., & Zhu, X. (2011). Canopy structure, light interception, and photosynthetic characteristics under different narrow-wide planting patterns in maize at silking stage. *Journal of Agricultural Research*, 9(4): 1249–1261.
- Liu, C. Z., Guo, C., Wang, Y. C., & Ouyang, F. (2002). Effect of light irradiation on hairy root growth and artemisinin biosynthesis of *Artemisia annua*. *Journal of Process Biochemistry*, 38: 581–585.
- Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy Review*, 4(8): 118-126.
- Lochhead, A. G., & Chase, F. E. (1943). Qualitative studies of soil microorganisms: nutritional requirements of the predominant bacterial flora. *Soil Science*, 55(2): 185-196.
- Lodish, H., Berk, A., & Zipursky, S. L. (2000). *Photosynthetic Stages and Light-Absorbing Pigments*. In *Molecular Cell Biology*, New York: Freeman and Co.
- Long, S. P., Humphries, S., & Falkowski, P. G. (1994). Photoinhibition of photosynthesis in nature. *Annual Review of Plant Physiology and Plant Molecular Biology*, 45: 633-662.
- Lutts, S., Lefevre, I., Delperee, C., Kivits, S., Dechamps, C., Robledo, A., & Correale, E. (2004). Heavy metal accumulation by the halophyte species Mediterranean Saltbush. *Journal of Environmental Quality*, 33: 1271-1279.
- Ma, X., Song, L., Yu, W., Hu, Y., Liu, Y., Wu, J., & Ying, Y. (2015). Growth, physiological, and biochemical responses of *Camptotheca acuminata* seedlings to different light environments. *Frontiers in Plant Science*, 6(5): 1–12.
- Magkos, F., Arvaniti, F., & Zampelas, A. (2003). Organic food: nutritious food or food for thought? a review of evidence. *International Journal of Food Science Nutrition*, 54: 357-371.
- Mahlia, T. M. I., Yong, J. H., Safari, A., & Mekhilef, S. (2012). Techno-economic analysis of palm oil mill wastes to generate power for grid-connected utilization. *Energy Science and Research*, 28(2): 1117-1136.
- Man, Y., Liang, G. Li, A., & Pan, L. (2017). Recent advances in mycotoxin determination for food monitoring via microchip. *Journal of Toxicology*, 9(10): 324-327.
- Marenco, R., Antezana Vera, S., & Nascimento, H. C. (2009). Relationship between specific leaf area, leaf thickness, leaf water content and SPAD-502 readings in six Amazonian tree species. *Photosynthetica*, 47(2): 184–190.

- Marjenah, Kiswanto, Purwati, S., & Sofyan, F. P. M. (2016). The effect of biochar, cocopeat and saw dust compost on the growth of two dipterocarps seedlings. *Nusantara Bioscience*, 8(1): 39-44.
- Marschner, H. (1995). Mineral nutrition of higher plants. 2.ed. London: Academic Press, Pp. 889.
- Masand, S., Madan, S., & Balian, S. K. (2014). Modern concept of storage and packaging of raw herbs used in ayurveda. *International Journal of Research in Ayurveda and Pharmacy*, 5(2): 242–245.
- Masarirambi, M. T., Dlamini, P., Wahome, P. K., & Oseni, T. O. (2012). Effects of chicken manure on growth, yield and quality of lettuce (*Lactuca sativa* L.) 'Taina' under a lath house in a semi-arid sub-tropical environment. *American-Eurasian Journal of Agriculture & Environmental Science*, 12(3): 399-406.
- Masarovicova, E., Majekova, M., & Vykoukova, I. (2015). Functional traits and plasticity of plants in ecological research and education. *Chemistry Didactics Ecology Metrology*, 20(1): 59–65.
- Matloobi, M. (2012). Light harvesting and photosynthesis by the canopy. *Advances in Photosynthesis - Fundamental Aspects*, 235–256.
- Mattana, R. S., Vieira, M. A. R., Marchese, J. A., Ming, L. C., & Marques, M. O. M. (2010). Shade level effects on yield and chemical composition of the leaf essential oil of *Pothomorphe umbellata* (L.) Miquel. *Scientia Agricola*, 67(4): 414–418.
- MD Jaafar, N. (2007). Effects of empty fruit bunch compost and arbuscular mycorrhiza on nutrient uptake and growth on grain maize, Master Thesis, Faculty of Agriculture, Universiti Putra Malaysia.
- Mediani, A., Abas, F., Tan, C., & Khatib, A. (2014). Effects of different drying methods and storage time on free radical scavenging activity and total phenolic content of *Cosmos caudatus*. *Antioxidants*, 3(2): 358–370.
- Medina, C. L., Souza, R. P. Machado, E. C., Ribeiro, R. V., & Silva, J. A. B. (2002). Photosynthetic response of citrus grown under reflective aluminized polypropylene shading nets. *Journal of Science Horticulture*, 96: 115–125.
- Megat, S. S., Ebuna, R. M., & Secretaria M. I. (2002). Mid-term yield response (1996-2001) of coconut to the application of coconut coir dust or coco peat in coastal area (Mindanao, Phil.). Paper presented in the World Food Day Celebration, Lecture Series on Soil Conservation & Water Resources Management Technologies. Seminar for Future Soil Scientists. Oct.15, 2002. DA-BSWM, Diliman, Quezon City.
- Meng, W., Xiaoliang, R., Xiumei, G., Vincieri, F. F., & Bilia, A. R. (2009). "Stability of active ingredients of traditional chinese medicine (TCM)". *Natural Product Communications*, 4(2): 1761-1776

- MIDA. (2016). Malaysia Investment Performance Report 2016 (p. 92). Retrieved from <http://www.mida.gov.my/env3/uploads/PerformanceReport/2013/IPR2013>.
- Milenkovic, S. M., Zvezdanovic, J. B., Anelkovic, T. D., & Markovic, D. Z. (2012). The identification of chlorophyll and its derivatives in the pigment mixtures: hplc-chromatography, visible and mass spectroscopy studies. *Advanced Technologies*, 1(1): 16–24.
- Milla, O. V., Rivera, E. B., Huang, W. J., Chien, C. C., & Wang, Y. M. (2013). Agronomic properties and characterization of rice husk and wood biochars and their effect on the growth of water spinach in a field test. *Journal of Soil Science and Plant Nutrition*, 13(2): 251-266.
- Milla, R., Reich, P. B., Niinemets, U., & Castro-Díez, P. (2008). Environmental and developmental controls on specific leaf area are little modified by leaf allometry. *Functional Ecology*, 22(4): 565–576.
- Ministry of Agriculture and Agro-based Industries. (2011). Program booklet NKEA EPP1, *Prosiding Persidangan Kebangsaan Ekonomi Malaysia Ke-10 2015*.
- Mingzhu, H., Zhang, K, Tan, H., Su, J., Wang, J., Huang, L., Zhang, Y., & Li, X. (2015). Nutrient levels within leaves, stems, and roots of xeric species *Reaumuria soongorica* in relation to geographical, climatic, and soil conditions. *Ecology Evolution*, 5(7): 1494-1503.
- Modi, A.T. (2007). Growth temperature and plant age influence on nutritional quality of *Amaranthus* leaves and seed germination capacity. *African Journal of Water SA*, 33(3): 369-375.
- Mohammad Azmin, S. N. H., Abdul Manan, Z., Wan Alwi, S. R., Chua, L. S., Mustafa, A. A., & Yunus, N. A. (2016). Herbal processing and extraction technologies. *Separation and Purification Reviews*, 45(4): 305-320.
- Mohammed, A. A., & Zakariya'u. (2012). Bitter leaf (*Vernonia amygdalina*) as a feed additive in broiler diets. *Research Journal of Animal Sciences*, 6(3): 38-41.
- Mohd Zainal, M.K., Abdul Hamid, A., Abu Bakar, F., & Pak Dek, S. (2009). Effect of different drying methods on the degradation of selected flavonoids in *Centella asiatica*. *Journal of Food Chemistry* 16: 531-537.
- Mohiseni, M. (2017). Medicinal herbs, strong source of antioxidant in aquaculture: a mini review. *Modern Applications in Pharmacy & Pharmacology*, 1(1): 1-5.
- Moniruzzaman, M., Islam, M. S., Hossain, M. M., Hossain, T., & Miah, M. G. (2009). Effects of shade and nitrogen levels on quality Bangladhonia Production. *Bangladesh Journal of Agricultural Research*, 34(2): 205-213.



- Morgan, J. B., & Connolly, E. L. (2013). Plant-soil interactions: Nutrient uptake. *Nature Education Knowledge*, 4(8): 2.
- Mpalantinos, M. A., de Moura, R. S., Parente, J. P., & Kuster, R. M. (1998). Biologically active flavonoids and kava pyrones from the aqueous extract of *Alpinia zerumbet*. *Phytotherapy Research*, 12: 442–444.
- Muchoki, C. N., Lamuka, P. O., & Imungi, J. K. (2010). Reduction of nitrates, oxalates and phenols in fermented solar-dried stored cowpea (*Vigna unguiculata* L.) leaf vegetables. *African journal of food, agriculture, nutrition and development*, 10(11): 4398-4412.
- Musa, A., & Ogbadoyi, E. (2014). Effect of cold storage on the concentrations of some nutrients, anti-nutrients and toxic substances in the leaves of *Vernonia amygdalina* (Bitter leaf). *Advances in Research*, 2(1): 24-39.
- Mutuchelian, K., Paliwal, K., & Gnanam, A. (1989). Influence of shading on net photosynthesis and transpiration rates, stomatal diffusive resistance, nitrate reductase and biomass productivity of a woody legume tree species (*Erythrina variegata* Lam.). *Journal of Plant Science*, 99(6): 539-546.
- Myers, C., Anderson, R., & Byers, D. (2005). Influence of shading on the growth and leaf photosynthesis of the invasive non-indigenous plant garlic mustard [*Alliaria petiolata* (M. Bieb) Cavara and Grande] grown under simulated late-winter to mid-spring conditions. *Journal of the Torrey Botanical Society*, 132(1): 1-10.
- Nadgorska-Socha, A., Kafel, A., Kandziora-Ciupa, M., Gospodarek, J., & Zawisza-Raszka, A. (2013). Accumulation of heavy metals and antioxidant responses in *Vicia faba* plants grown on monometallic contaminated soil. *Environmental Science and Pollution Research*, 20(2): 1124-1134.
- Naser, H. M., Sultana, S., Mahmud, N. U., Gomes, R., & Noor, S. (2012). Heavy metal levels in vegetables with growth stage and plant species variations. *Bangladesh Journal of Agricultural Research*, 36(4): 563-574.
- Nazari, F., Farahmand, H., Khosh-Khui, M., & Salehi, H. (2011). Effects of coir as a component of potting media on growth, flowering and physiological characteristics of hyacinth (*Hyacinthus orientalis* L. cv. *Sonbol-e-Irani*). *International Journal of Agricultural and Food Science*, 1(2): 34-38.
- Nedosa, U., & Anastasia, U. (2011). Mechanisms of anti-cancer effects of *Vernonia amygdalina* leaf extract. *American Journal of Pharmacology and Toxicology*, 6(3): 76-79.
- Noor Ismawaty, N., Ahmad Tarmizi, S., Engku Hasmah, E. A., & Nurulnihar, E. (2015). Effect of storage on phytochemical contents of Misai kucing (*Orthosiphon stamineus* Benth) leaves. *Journal of Tropical Agriculture and Food Science*, 43(2): 165-170.

- Norawanis, A. R., Shaari, A. R., & Yeng, L. Y. (2018). Effect of packaging materials on *Orthosiphon stamineus* dried-leaf quality during storage. *IOP Conference Series: Materials Science Engineering*, 318 012010.
- Nor shariah, S. (2017). *Effect of drying method and storage on phytochemical stability of A. paniculata (hempedu bumi)*. PhD Thesis. Universiti Putra Malaysia, Serdang, Malaysia.
- Nostro, A., Germano, M. P., D'angelo, V., Marino, A., & Cannatelli, M. A. (2000). Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity, *Letters in Applied Microbiology*, 30: 379-384.
- Nur Faezah, O., Siti Aishah, H, Mohd Fauzi, R., Puteri Edaroyati, M. W., & Mohd Puad, A. (2016). Growth and phytochemical responses of *Andrographis paniculata* as influenced by different shade levels and prunings. *Journal of Tropical Plant Physiology*, 8: 61-69.
- Nur Faezah, O., Siti Aishah, H., Edaroyati, Puteri, W. M., Ramlan, M. F., & Puad, M. A. (2015). Growth and dry matter partitioning of *Andrographis paniculata* to different light intensities and pruning. *Global Advance Research Journal of Agricultural Science*, 4(12): 851-857.
- Nwaoguikpe, R. (2010). The effect of extract of bitter leaf (*Vernonia amygdalina*) on blood glucose levels of diabetic rats. *International Journal of Biological and Chemical Sciences*, 4(6): 721–729.
- Nwogwugwu, C.P., Petrus, N. E., Ethelbert, O. C., & Lynda, O. C. (2015). Effect of *Vernonia amygdalina* (Bitter leaf) extract on growth performance, carcass quality and economics of production of broiler chickens. *International Institute of Academic Research and Development*, 1(5): 1-13.
- Ofori, D., Anjarwalla, P., Jamnadass, R., Stevenson, P., & Smith, P. (2013). *Vernonia amygdalina*. *Pesticidal plant leaflet*, Pp. 1-2.
- Ogbulie, J. N., Ogueke, C. C., & Nwannebu, F. C. (2007). Antibacterial properties of *Uvaria chamae*, *Congronema latifolium*, *Garcinia kola*, *Vernonia amygdalina* and *Aframomium melegueta*. *African Journal of Biotechnology*, 6(13): 1549-1553.
- Ogundola, A. F., Bvenura, C., & Afolayan, A. J. (2018). Nutrient and antinutrient compositions and heavy metal uptake and accumulation in *S. nigrum* cultivated on different soil types. *The scientific world Journal*, 1: 1-20.
- Oladele, O. O., & Aborisode, A. T. (2009). Influence of different drying methods and storage on the quality of Indian spinach (*Basella rubra* L.). *American Journal of Food Technology*, 4(2): 66-70.
- Orrono, D. I., & Lavado, R. S. (2009). Heavy metal accumulation in *Pelargonium hortorum*: Effects on growth and development. *Phyton (Buenos Aires)*, 78: 75-82.

- Osinubi, A. A. A. (2007). Effects of *Vernonia amygdalina* and chlorpropamide on blood glucose. *Medicinal Journal of Islamic World Academy of Sciences*, 16(3): 115-119.
- Owen, O. J., Amakiri, A. O., & Karibi-Botoye, T. A. (2011). Sugar-lowering effects of bitter leaf (*Vernonia amygdalina*) in experimental broiler finisher chickens. *Asian Journal of Pharmaceutical and Clinical Research*, 4(1): 19–21.
- Oyeyemi, I. T., Akinlabi, A. A., Adewumi, A., Aleshinloye, A. O., & Oyeyemi, O. T. (2017). *Vernonia amygdalina*: A folkloric herb with anthelmintic properties. *Beni-Suef University Journal of Basic and Applied Sciences*, 1(7): 1–7.
- Patakas, A., Kofidis, G., & Bosabalís, A. M. (2003). The relationship between CO<sub>2</sub> transfer mesophyll resistance and photosynthetic efficiency in grapevine cultivars. *Journal of Science Horticulture*, 97: 255-263.
- Pearcy, R. W., Muraoka, H., & Valladares, F. (2005). Crown architecture in sun and shade environments: Assessing function and trade-offs with a three-dimensional simulation model. *New Phytologist*, 166(3): 791–800.
- Pholphana, N., Rangkadilok, N., Saehun, J., Ritruethai, S., & Satayavivad, J. (2013). Changes in the contents of four active diterpenoids at different growth stages in *A. paniculata* (Burm. f.) Nees. (Chuanxinlian). *Chinese Medicine*, 8(1): 1-13.
- Pons, T. L., Jordi, W., & Kuiper D. (2001). Acclimation of plants to light gradients in leaf canopies: evidence for a possible role for cytokinins transported in the transpiration stream. *Journal of Experimental Botany*, 52: 1563-1574.
- Poorter, H., Niklas, K. J., Reich, P. B., Oleksyn, J., Poot, P., & Mommer, L. (2012). Biomass allocation to leaves, stems and roots: meta-analyses of interspecific variation and environmental control. *Journal of New Phytologist*, 193(1): 30-50.
- Pradubsuk, S., & Davenport, J. R. (2010). Seasonal uptake and partitioning of macronutrients in mature 'Concord grape'. *Journal of the American Society for Horticultural Science*, 135(5): 474-483.
- Priyadharshini, J., & Seran, T. H. (2009). Paddy husk ash as a source of potassium for growth and yield of cowpea (*Vigna unguiculata* L.). *Journal of Agricultural Sciences*, 4(2): 67-76.
- Quintero, Ortega, M. F., D., Valenzuela, J. L., & Guzmán. M. (2013). Variation of hydro-physical properties of burnt rice husk used for carnation crops: Improvement of fertigation criteria. *Science Horticulture*, 154: 82–87.
- Rachmawati, D., Sudjino, & Asiyah, I. J. (2017). Resonse and stevioside levels of stevia (*Stevia rebaudiana bertonii*) grown at different light intensity

and water availability. *Journal of Botany*, 49(5): 1689-1695.

- Ramachendrin, A. D. (2015). *In vitro activity of local plants from Malaysia against chikungunya virus*. Degree Thesis. Universiti Tunku Abdul Rahman, Malaysia.
- Raschke K. (1975). Stomatal action. *Annual Review of Plant Physiology*, 26: 309-340.
- Rao, P. B., & Singh, S. P. (1989). Effect of shade on Central Himalayan species from a successional gradient. *International Journal of Ecology*, 10: 21-33.
- Raya, K. B., Ahmad, S. H., Sanusi, F. F., Mohammad, M., Tajidin, N. E., & Parvez, A. (2015). Changes in phytochemical contents in different parts of *Clinacanthus nutans* (Burm. f.) lindau due to storage duration. *Bragantia, Campinas*, 74(4): 445-452.
- Ren, Z. G., Chen, Y. S., & Tang, F. Q. (1996) Effect of inorganic fertilizer combined with organic manure on the microflora and enzyme activities in paddy soil. *Plant Nutrition and Fertilizer Science*, 2: 279-283.
- Riaz, A., Younis, A., Ghani, I., Tariq, U., & Ahsan, M. (2015). Agricultural waste as growing media component for the growth and flowering of *Gerbera jamesonii* cv. hybrid mix. *International Journal of Recycling of Organic Waste in Agriculture*, 4(3): 197-204.
- Righi, C. A., Bernandes, M. S., Lunz, A. M. P., Pereira, C. R., Neto, D. D., & Favarin, E. L. (2007). Measurement and simulation of solar radiation availability in relation to the growth of coffee plants in an agroforestry system with rubber trees. *Forest Research Society*, 31(2): 195-207.
- Rocha, R. P., Melo, E. C., & Radunz, L. L. (2011). Influence of drying process on the quality of medicinal plants: a review. *Journal of Medicinal Plants Research*, 5: 7076-7084.
- Rodrigo, R., & Bosco, C. (2006). Oxidative stress and protective effect of polyphenols: comparative studies in human and rodent kidney. *Comparative Biochemistry and Physiology*, 142: 317-327.
- Rodriguez, M. J., Villanueva, M. J., & Tenorio, M. D. (1999). Changes in chemical composition during storage of peaches (*Prunus persica*). *European Food Research and Technology*, 209: 135-139.
- Rozendaal, D. M. A., Hurtado, V. H., & Poorter, L. (2006). Plasticity in leaf traits of 38 tropical tree species in response to light; relationships with light demand and adult stature. *Functional Ecology*, 20(2): 207-216.
- Rugna, A. Z., Ricco, R., Gurni, A., & Wagner, M. (2008). Variation in leaves polyphenol content in *Smilax campestris* Griseb. (Smilacaceae) according to their development. *Latin American Journal of Pharmacy*, 27(2): 247-249.

- Saikkonen, K., Koivunen, S., Vuorisalo, T., & Mutikainen, P. (1998). Interactive effects of pollination and heavy metals on resource allocation in *Potentilla anserina* L. *Ecology*, 79: 1620–1629.
- Salata, A., Buczkowska, H., Vicente, S., Galarza, L., & Moreno-ramon, H. (2015). The polyphenolic compounds content of a Cardoon herb depending on length of the vegetation period. *Journal of Biological Sciences*, 14(4): 155-167.
- Salisu, M. A., Daud, W. N., Abdul Halim, R., & Sulaiman, Z. (2016). Effect of soilless media on growth and some physiological traits of rubber (*Hevea brasiliensis*) seedlings. *International Journal of Agriculture, Forestry and Plantation*, 3(6): 95-100.
- Sanullah, M., Rumpel, C., Charrier, X., & Chabbi, A. (2012). How does drought stress influence the decomposition of plant litter with contrasting quality in a grassland ecosystem? *Plant Soil*, 352: 277-288.
- Sardans, J., & Penuelas, J. (2012). The role of plants in the effects of global change on nutrient availability and stoichiometry in the plant-soil system. *Journal of Plant Physiology*, 160(4): 1741-1761.
- Schollenberger, C. J. & Simon, R. H. (1945). Determination of exchange capacity and exchangeable bases in soil-ammonium acetate method. *Soil Science*, 59: 13-23.
- Sarikioti, V., de Visser, P. H. B., Buck-Sorlin, G. H., & Marcelis, L. F. M. (2011). How plant architecture affects light absorption and photosynthesis in tomato: Towards an ideotype for plant architecture using a functionalstructural plant model. *Annals of Botany*, 108(6): 1065–1073.
- Schimel, J., Balsler, T. C., & Wallenstein, M. (2007). Microbial stress-response physiology and its implications for ecosystem function. *Ecology*, 88(6): 1386-1894.
- Scuderi, D., Giuffrida, F., Toscano, S., & Romano, D. (2012). Growth, physiological response, and quality characteristics of weeping fig in response to shading levels and climatic conditions. *Journal of Horticulture Science*, 47(11): 1586–1592.
- Senevirathna, A. M. W. K., Stirling, C. M., & Rodrigo, V. H. L. (2003). Growth, photosynthetic performance and shade adaptation of rubber (*Hevea brasiliensis*) grown in natural shade. *Tree Physiology*, 23(10): 705–712.
- Shaban, N., Abdou, K. A., & Hassan, N. E. Y. (2016). Impact of toxic heavy metals and pesticide residues in herbal products. *Journal of Basic and Applied Sciences*, 5(1): 102–106.
- Shadchina, T. M., & Dmitrieva. V. V. (1995). Leaf chlorophyll content as a possible diagnostic mean for the evaluation of plant nitrogen uptake from the soil. *Journal of Plant Nutrition*, 18: 1427–1437.

- Shara, S. A., Zaharah, S. S., Radziah, O., & Puteri, E. M. W. (2017). Physical, chemical and microbiological properties of different combination of soilless media and their effect on vegetative component and nutrient content of Hempedu bumi (*Andrographis paniculata*). *Pertanika Journal of Tropica Agriculture*, 40(1): 35-52.
- Shiow, Y. W. (2012). Phytonutrients and bioactive compounds. Course on Postharvest Handling of Tropical Produce, Serdang.
- Singh, G., Kawatra, A., Sehgal, S., & Pragati. (2003). Effect of storage and nutritional composition of selected dehydrated leafy vegetable, herb and carrot powders. *Plant Foods for Human Nutrition*, 58: 1-9.
- Sousa Paiva, E., Dos Santos Isaias, R., Aguiar Vale, F., & De Senna Queiroz, C. (2003). The influence of light intensity on anatomical structure and pigment contents of *Tradescantia pallida* (Rose) Hunt. cv. purpurea boom (commelinaceae) leaves. *Brazilian Archives of Biology and Technology*, 46(4): 617–624.
- Sterling, T. M. (2004). Transpiration- Water movement through plants. New Mexico State University.
- Stevic, T., Pavlovic, S., Stankovic, S., & Savikin, K. (2012). Pathogenic microorganisms of medicinal herbal drugs. *Archives of Biological Sciences*, 64(1): 49-58.
- Stojadinov, J. (1998). Investigation of microbial contamination on Pitoma (*Mentha piperita* L.). *Medical matters XVII*, 45-53.
- Sultan, S. E. (2000). Phenotypic plasticity for plant development, function and life history. *Trends in Plant Science*, 5: 537-542.
- Taiz, L., & Zeiger, E. (2006). *Plant Physiology*, 5th ed. Belmont, California: Sinauer Associates, Inc.
- Tajidin, N.E. (2017). *Effects of plant harvest age and plant parts on phytochemical compounds of Andrographis paniculata (Burm. F.) Nees*. PhD Thesis. Universiti Putra Malaysia, Serdang, Malaysia.
- Takenaka, A. (1994). Effects of leaf blade narrowness and petiole length on the light capture efficiency of a shoot. *Ecological Research*, 9: 109-114.
- Tanchuling, M. N. R., Resurreccion, A. C., & Ong, D. (2012). A research project removing heavy metals from wastewater of small-scale gold miners of camarines norte (philippines) using coco-peat as sorbent material. Pp. 23-29.
- Tangahu, B. V., Sheikh Abdullah, S. R., Basri, H., Idris, M., Anuar, N., & Mukhlisin, M. (2011). A Review on heavy metals (As, Pb, and Hg) uptake by plants through phytoremediation. *International Journal of Chemical Engineering*, 1–31.

- Tatiya, A. U., Tapadiya, G. G., Kotecha, S., & Surana, S. J. (2011). Effect of solvents on total phenolics, antioxidant and antimicrobial properties of *Bridelia retusa* Spreng. Stem bark. *Indian Journal of Natural Products and Resources*, 2(4): 442-447.
- Teh, C. B. S., & Jamal, T. (2006). Bulk Density, water content, porosity and water retention. In: *Soil Physic Analyses*, University Putra Malaysia Press, Selangor, Malaysia, Pp. 11-17.
- Theunissen, J., Ndakidemi, P. A., & Laubscher, C. P. (2010). Potential of vermicompost produced from plant waste on the growth and nutrient status in vegetable production. *International Journal of the Physical Sciences*, 5(13): 1964-1973.
- Thi, N. D., & Hwang, E. (2014). Bioactive compound contents and antioxidant activity in Aronia (*Aronia melanocarpa*) leaves collected at different growth stages. *Nutrition and Food Science*, 19(3): 204-212.
- Tiwari, R., & Rana, C.S. (2015). Plant secondary metabolites: A review. *International Journal of Engineering Research and General Science*, 3(5): 661-670.
- Tomsone, L., & Kruma, Z. (2017). Influence of harvest time on the phenolic content of Horseradish leaves. In *Baltic Conference on Food Science and Technology*, Pp. 45-50.
- Tonukari, N., Avwioroko, O., Ezedom, T., & Anigboro, A. (2015). Effect of preservation on two different varieties of *Vernonia amygdalina* Del. (Bitter) leaves. *Journal of Food and Nutrition Sciences*, (6): 633-642.
- Tsado, Lawal, A. N., Santali, B., Shaba, E. S., Chirama, A. M., Balarabe, D. N., Jiya, M. M., & Alkali, H. A. (2015). Effect of different processing methods on nutritional composition of bitter leaf (*Vernonia amygdalina*). *Journal of Pharmacy*, 5(6): 8-14.
- Tsakaldimi, M. (2006). Kenaf (*Hibiscus cannabinus* L.) core and rice hulls as components of container media for growing *Pinus halepensis* M. seedlings. 97(14): 1631-1639.
- Ucar, E., Ozyigit, Y., Eruygur, N., Guven, D., Yur, S., Turgut, K., Ozek, T., & Kutuk, N. (2017). The effect of the plant age and growth period on the nutritional substance, chlorophyll and steviol glycoside rates in stevia (*Stevia rebaudiana* Bertoni) leaves. *Communications in Soil Science and Plant Analysis*, 11: 1-12.
- Uchida, R., & Silva, J. A. (2000). Essential nutrients for plant growth: Nutrient functions and deficiency symptoms. College of Tropical Agriculture and Human Resources, University of Hawaii at Manoa, Pp. 31-55.
- Udochukwu, U., Omeje, F. I., Uloma, I. S., & Oseiwe, F. D. (2015). Phytochemical analysis of *Vernonia amygdalina* and *Ocimum gratissimum* extracts and their antibacterial activity on some drug

- resistant bacteria. *American Journal o Research Communication*, 3(5): 225–235.
- Utumi, M. M., Monnerat P. H., Pereira, P. R. G., Fontes, P. C. R., & Godinho V. (1999). Macronutrient deficiencies in Stevia: Visual and effects on growth, chemical composition, and stevioside production. *Brazilian Agricultural Research*, 34: 1039-1043.
- Valladares, F., & Niinemets, U. (2008). Shade tolerance, a key plant feature of complex nature and consequences. *Annual Review of Ecology, Evolution, and Systematics*, 39: 237-257.
- Vandana & Bhatt, R. K. (1999). Physiological changes in Sesbania species to reducing light intensities. *Journal of Agronomy and Crop Science*, 182(1): 43-47.
- Vicente, R., Morcuende, R., & Babiano, J. (2011). Differences in rubisco and chlorophyll content among tissues and growth stages in two tomato (*Lycopersicon esculentum* Mill.) varieties. *Agronomy Research*, 9(2): 501–507.
- Volf, I., Ignat, I., Neamtu, M., & Popa, V. I. (2014). Thermal stability, antioxidant activity, and photo-oxidation of natural polyphenols, *Chemical Papers*, 68(1): 121–129.
- Wadood, A., Ghufran, M., Jamal, S. B., Naeem, M., Khan, A., Ghaffar, R., & Asnad. (2013). Phytochemical analysis of medicinal plants occurring in local area of Mardan. *Biochemistry & Analytical Biochemistry*, 2(4): 1-4.
- Wang, Y. S., Gao, L. P., Wang, Z. R., Liu, Y. J., Sun, M. L., Yang, D. Q., Wei, C. L., Shan, Y., & Xia, T. (2012). Light-induced expression of genes involved in phenylpropanoid biosynthetic pathways in callus of tea (*Camellia sinensis* (L.) O. Kuntze). *Journal of Science Horticulture*, 133: 72-83.
- Wazis, C. H., Timothy, S. Y., Zakama, S. G., Balla, H. J., & Maspalma, I. D. (2013). Phytochemical screening and purgative activity of ethanolic extracts of *Vernonia amygdalina* Del. leaf. *International Journal of Ayurveda and Pharmacy*, 4(1): 46-49.
- WHO. (2007). Guidelines for assessing quality of herbal medicines with reference to contaminants and residues. WHO Press, Geneva, Switzerland. Pp. 27.
- Williams, W. A., Loomis, R. S., & Lepley, C. R. (1965). Vegetative growth of corn as affected by population density. I. Productivity in relation to interception of solar radiation. *Crop Science*, 5: 211-215.
- Wong, F. C., Woo, C. C., Hsu, A., & Tan, B. K. H. (2013). The anti-cancer activities of *Vernonia amygdalina* extract in human breast cancer cell lines are mediated through caspase-dependent and p53-independent pathways. *PLoS ONE*, 8(10).



- Wu, Y., Gong, W., & Yang, W. (2017). Shade inhibits leaf size by controlling cell proliferation and enlargement in Soybean. *Scientific Reports*, 7(8): 1-10.
- Xiolei, S., & Zhifeng, W. (2002). The optional leaf area index for cucumber photosynthesis and production in plastic greenhouse. Paper presented at the XXVI International Congress: Protected Cultivation 2002: In Search of Structures, Systems and Plant Materials for 633.
- Xu-yang, Y., Xiao-ying, L., Zhi-gang, X., & Xue-lei, J. (2017). Effects of light intensity on leaf microstructure and growth of rape seedlings cultivated under a combination of red and blue LEDs. *Journal of Integrative Agriculture*, 16(1): 97-105.
- Yadav, S. K., Babu, S., Yadav, M. K., Singh, K., Yadav, G. S., & Pal, S. (2013). A review of organic farming for sustainable agriculture in Northern India. *International Journal of Agronomy*, 1(1): 1-8.
- Yang, B., Zheng, J., Laaksonen, O., Tahvonen, R., & Kallio, H. (2013). Effects of latitude and weather conditions on phenolic compounds in currant (*Ribes* spp.) cultivars. *Journal of Agricultural and Food Chemistry*, 61: 3517-3532.
- Yargholi, B., & Azimi, A. A. (2008). Investigation of cadmium absorption and accumulation in different parts of some vegetables. *American-Eurasian Journal of Agricultural & Environmental Sciences*, 3(3): 357-364.
- Yau, P. Y., & Murphy, R. J. (2000). Biodegraded cocopeat as a horticultural substrate. *International Science of horticulture*, 517: 275-278.
- Yeap, S. K., Ho, W. Y., Beh, B. K., Liang, W. S., Ky, H., Hadi, A., & Alitheen, N. B. (2010). Ethnomedical used green vegetable with multiple bio-activities. *Journal of Medicinal Plants Research*, 4(25): 2787-2812.
- Yew, B. S., & Wee, F. H. (2014). Agricultural waste based-coco peat microwave absorber. *International Journal of Engineering Sciences and Emerging Technologies*, 7(2): 547-554.
- Zandonadi, D. B., Santos, M. P., Busato, J. G., Peres, L. E. P., & Façanha, A. R. (2013). Plant physiology as affected by humified organic matter. *Theoretical and Experimental Plant Physiology*, 25(1): 13-25.
- Zervoudakis, G., Salahas, G., Kaspiris, G., & Konstantopoulon, E. (2012). Influence of light intensity on growth and physiology characteristics of Common sage (*Salvia officinalis* L.). *An International Journal of Brazilian Archives of Biology and Technology*, 55(1): 89-95.
- Zhang, W., Wang, C., Shi, B. & Pan, X. (2017). Effect of storage temperature and time on the nutritional quality of walnut male inflorescences. *Journal of Food and Drug Analysis*, 25(2): 374-384.
- Zhang, D., Zhang, L., Liu, J., Han, S., Wand, Q., Evers, J., Liu, J., van de Werf, W., & Li, L. (2014). Plant density affects light interception and yield in

cotton grown as companion crop in young Jujube plantation. *Field Crops Research*, 169(12): 132-139.

Zhang, L., Gao, Y., Zhang, Y., Liu, J., & Yu, J. (2010). Changes in bioactive compounds and antioxidant activities in pomegranate leaves. *Journal of Science Horticulture*, 123: 543-546.

Zhang S, Ma, K., & Chen, L. (2003). Response of photosynthetic plasticity of *Paeonia suffruticosa* to changed light environments. *Environmental and Experimental Botany*, 49: 121-133.

Zulkarimi, B., Ashrafuzzaman, M., & Mohd Razi, I. (2010). Morpho-physiological growth, yield and fruit quality of rock melon as affected by growing media and electrical conductivity. *Journal of Food, Agriculture and Environment*, 8(1): 249-252.