



**UNIVERSITI PUTRA MALAYSIA**

***NUTRIENT UPTAKE AND GROWTH OF IMMOBILIZED  
FILAMENTOUS MICROALGAE *Stigeoclonium nanum* (DILLWYN)  
KÜTZING 1849 FOR IMPROVEMENT OF WATER QUALITY***

**MUHAMMAD KHAIRANI BIN ZULKIFELY**

**FP 2013 104**

**NUTRIENT UPTAKE AND GROWTH OF IMMOBILIZED FILAMENTOUS  
MICROALGAE *Stigeoclonium nanum* (DILLWYN) KÜTZING 1849 FOR  
IMPROVEMENT OF WATER QUALITY**

**MUHAMMAD KHAIRANI BIN ZULKIFELY**

**DEPARTMENT OF AQUACULTURE**

**FACULTY OF AGRICULTURE**

**UNIVERSITI PUTRA MALAYSIA**

**SERDANG, SELANGOR**

**2013**

**Nutrient Uptake and Growth of Immobilized Filamentous Microalgae  
*Stigeoclonium nanum* (Dillwyn) Kützing 1849 for Improvement of Water  
Quality**

**MUHAMMAD KHAIRANI BIN ZULKIFELY**

**164109**

**This project report is submitted in partial fulfillment of the requirements for the  
degree of Bachelor of Agriculture (Aquaculture)**

**DEPARTMENT OF AQUACULTURE**

**FACULTY OF AGRICULTURE**

**UNIVERSITI PUTRA MALAYSIA**

**SERDANG, SELANGOR**

**2013**

**CERTIFICATION OF APPROVAL**  
**DEPARTMENT OF AQUACULTURE**  
**FACULTY OF AGRICULTURE**  
**UNIVERSITI PUTRA MALAYSIA**

Name of student : Muhammad Khairani bin Zulkifely  
Matric number : 164109  
Programme : Bachelor of Agriculture (Aquaculture)  
Name of supervisor : Prof. Dr. Fatimah Md. Yusoff  
Title of project : Nutrient Uptake and Growth of Immobilized filamentous  
microalgae *Stigeoclonium nanum* (Dillwyn) Kützing 1849  
for Improvement of Water Quality

This is to certify that I have examined the final year project report and all corrections have been made as recommended format stipulate in the AKU 4999 project guidelines, Department of Aquaculture, Faculty of Agriculture, and Universiti Putra Malaysia.

Signature and official stamp of supervisor:

---

Prof. Dr. Fatimah Md. Yusoff

Date:

## ACKNOWLEDGEMENT

First of all, Alhamdulillah, praise be to Allah for giving me the strength to complete this project and thesis. Secondly, special thanks to my supervisor Prof. Dr. Fatimah Md. Yusoff for her encouragement and invaluable help, guidance and constructive discussion we had over the experimental period. Great thanks to Pn. Norulhuda Mohamed Ramli for helping and encouraging me to complete this project and thesis. She had patiently guided me and shared with me her knowledge in facilitating me to conduct my project.

I also like to give my special thanks to Assoc. Prof. Dr Yuzine Esa, the coordinator of Final Year Project, who spent time to teach us the proper method to write a thesis. Not to forget, thanks to my fellow friends, especially the seniors from the Laboratory of Marine Biotechnology, Institute of Bioscience, Universiti Putra Malaysia. Last but not least, to my parents and family for their love and support.

## ABSTRACT

In aquaculture practices, a large amount of water has to be exchanged frequently in order to maintain good water quality. This procedure contributes to the eutrophication of aquatic environment due to flushing of nutrient-enriched waters from aquaculture facilities. Furthermore, the process of frequent water exchange will eventually result in lack of good water supply which can also increase the risk of diseases in the hatchery. To overcome eutrophication and the risk of diseases, an alternative eco-friendly method was investigated to decrease harmful compounds especially ammonia and nitrite by using microalgae. In this study, filamentous green microalgae *Stigeoclonium nanum* was used. The first objective of this project was to study growth of the filamentous microalgae (*S. nanum*) immobilized in alginate beads and in free suspension condition. Four experimental treatments, control 1 (no algae and no beads), control 2 (beads with free algae), algae in free suspension and beads with algae were tested. The parameters that were examined included *S. nanum* growth in terms of chlorophyll-a content, and ammonium loading for the microalgae culture. The ammonia loading is ammonia which was added up during experiment when it has zero reading. From the study, significantly higher ( $p < 0.05$ ) chlorophyll-a content was observed in alginate beads ( $5.27 \pm 0.63$  mg/l) on the final day of the experiment than the microalgae cultured in normal suspension culture ( $0.62 \pm 0.20$  mg/l). Significantly higher ( $p < 0.05$ ) total ammonium loading was also observed in the culture of immobilized microalgae. The second objective of the experiment was to measure the uptake of the ammonia-N ( $\text{NH}_4\text{-N}$ ) and nitrate-N ( $\text{NO}_3\text{-N}$ ) by immobilized filamentous microalgae. Five culture media with different concentrations of  $\text{NO}_3\text{-N}$  and  $\text{NH}_4\text{-N}$  were tested in this experiment, treatment A was Bold's Basal Medium (BBM), treatment B was zero concentration of  $\text{NO}_3\text{-N}$  and  $\text{NH}_4\text{-N}$  (control), treatment C was zero concentration of  $\text{NO}_3\text{-N}$  with  $10 \text{ mgL}^{-1}$  concentration of  $\text{NH}_4\text{-N}$ , treatment D was  $10 \text{ mgL}^{-1}$  of  $\text{NO}_3\text{-N}$  with zero concentration  $\text{NH}_4\text{-N}$ , and treatment E was  $10 \text{ mgL}^{-1}$  of  $\text{NO}_3\text{-N}$  with  $10 \text{ mgL}^{-1}$  of  $\text{NH}_4\text{-N}$ . The parameter examined was *S. nanum* uptake of the  $\text{NH}_4\text{-N}$  and  $\text{NO}_3\text{-N}$ . This study illustrated that  $\text{NH}_4\text{-N}$  was preferred than  $\text{NO}_3\text{-N}$  as nitrogen source by the immobilized *S. nanum*.

## ABSTRAK

Dalam amalan akuakultur, sejumlah besar air perlu ditukar kerap untuk mengekalkan kualiti air yang baik. Prosedur ini menyumbang kepada eutrofikasi air di dalam habitat akibat curahan air dengan nutrien diperkaya daripada kemudahan akuakultur. Tambahan pula, proses pertukaran air kerap akhirnya akan mengakibatkan kekurangan bekalan air yang baik yang juga boleh meningkatkan risiko penyakit di tapak penetasan. Untuk mengatasi eutrofikasi dan risiko penyakit, eko- mesra kaedah alternatif telah diselidiki dalam mengurangkan sebatian berbahaya terutamanya ammonia dan nitrat dengan menggunakan mikroalga. Dalam kajian ini, mikroalga hijau berfilamen *Stigeoclonium nanum* telah digunakan. Objektif pertama projek ini adalah untuk mengkaji pertumbuhan mikroalga yang berfilamen (*S. nanum*) bergerak di dalam manik alginat dan dalam keadaan kultur bebas. Empat rawatan eksperimen telah digunakan, kawalan 1 (tiada alga dan tiada manik), kawalan 2 (manik dengan tiada alga), alga dalam penggantungan percuma dan manik dengan alga telah diuji. Parameter yang dikaji adalah pertumbuhan *S. nanum* segi kandungan klorofil-a dan muatan ammonium di dalam didalam kultur mikroalga. Muatan ammonia adalah ammonia yang ditambah semasa eksperimen apabila bacaan ammonia adalah kosong. Hasil kajian tersebut, didapati kandungan klorofil-a lebih tinggi dan ketara ( $p < 0.05$ ) di dalam manik alginat ( $5.27 \pm 0.63$  mg/l) pada hari terakhir daripada mikroalga dikultur dalam kultur penggantungan biasa ( $0.62 \pm 0.20$  mg/l). Muatan ammonium lebih tinggi juga dilihat dalam kultur mikroalga tidak bergerak. Objektif kedua eksperimen ini adalah untuk mengukur pengambilan ammonia-N ( $\text{NH}_4\text{-N}$ ) dan nitrat-N ( $\text{NO}_3\text{-N}$ ) dengan mikroalga berfilamen di dalam manik alginat. Lima media kultur dengan kepekatan yang berbeza  $\text{NO}_3\text{-N}$  dan  $\text{NH}_4\text{-N}$  telah diuji dalam eksperimen ini, rawatan A adalah Bold Basal Media (BBM), rawatan B adalah kepekatan sifar  $\text{NO}_3\text{-N}$  dan  $\text{NH}_4\text{-N}$  (kawalan), rawatan C kepekatan sifar  $\text{NO}_3\text{-N}$  dengan kepekatan  $10 \text{ mgL}^{-1}$   $\text{NH}_4\text{-N}$ , rawatan D adalah  $10 \text{ mgL}^{-1}$   $\text{NO}_3\text{-N}$  dengan kepekatan sifar  $\text{NH}_4\text{-N}$ , dan rawatan E adalah  $10 \text{ mgL}^{-1}$   $\text{NO}_3\text{-N}$  dengan  $10 \text{ mgL}^{-1}$   $\text{NH}_4\text{-N}$ . Parameter yang dikaji adalah pengambilan  $\text{NH}_4\text{-N}$  dan  $\text{NO}_3\text{-N}$  daripada *S. nanum*. Dalam eksperimen ini, kita boleh menyimpulkan bahawa  $\text{NH}_4\text{-N}$  adalah pilihan utama berbanding  $\text{NO}_3\text{-N}$  sebagai sumber nitrogen oleh *S. nanum* didalam manik alginat.

## TABLE OF CONTENTS

<b>Contents</b>	<b>Page</b>
<b>ACKNOWLEDGEMENT</b>	i
<b>ABSTRACT</b>	ii
<b>ABSTRAK</b>	iii
<b>TABLE OF CONTENTS</b>	iv
<b>LIST OF TABLES</b>	vii
<b>LIST OF FIGURES</b>	viii
<b>LIST OF ABBREVIATIONS AND SYMBOLS</b>	ix
<b>1.0 INTRODUCTION</b>	1
1.1 Research objectives	2
<b>2.0 LITERATURE REVIEW</b>	
2.1 Aquaculture waste water	3
2.2 Microalgae for aquaculture	4
2.3 Filamentous microalga <i>Stigeoclonium nanum</i> (Dillwyn) Kützing 1849	6
2.3.1 Taxonomy	6
2.3.2 Morphology	6
2.4 Immobilizing microalgae into alginate beads	9



### 3.0 MATERIALS AND METHODS

3.1	Location of the study	11
3.2	General description of experimental design for measuring the growth of the filamentous microalga ( <i>Stigeoclonium nanum</i> ) immobilized in alginate beads and in free suspension condition	11
3.3	General description of experimental design for measuring the uptake of ammonia-N, and nitrate-N, by immobilized filamentous microalga ( <i>Stigeoclonium nanum</i> )	12
3.4	Source and maintenance of microalgae	13
3.5	Batch culture of microalgae	14
3.6	Preparation of immobilized of microalgae	14
3.7	Culture condition	15
3.8	Bold's Basal Medium (BBM)	16
3.9	Analytical techniques	
3.9.1	Procedure analysis of chlorophyll-a	17
3.9.1.1	Extraction of chloropyll-a	17
3.9.1.2	Spectrophotometric procedure	18
3.9.2	Measuring ammonia-N by using phenate methods	18
3.9.3	Procedure of analysis nitrate-N by using ion chromatography	19
3.10	Statistical procedures	20

<b>4.0</b>	<b>RESULTS AND DISCUSSIONS</b>	
4.1	Result of the first objective: Chlorophyll-a contents and ammonium loading of filamentous microalgae ( <i>Stigeoclonium nanum</i> ) immobilized in alginate beads and in free suspension condition	21
4.1.1	Chlorophyll-a contents analysis	21
4.1.2	Ammonium loading in the culture of filamentous microalgae ( <i>Stigeoclonium nanum</i> ) immobilized in alginate beads and in free suspension condition	23
4.2	Result of the second objective: Measuring the uptake of ammonia-N, NH <sub>4</sub> -N and nitrate-N, NO <sub>3</sub> -N by immobilized filamentous microalgae ( <i>Stigeoclonium nanum</i> )	25
<b>5.0</b>	<b>CONCLUSION</b>	<b>30</b>
	<b>REFERENCES</b>	<b>31</b>
	<b>APPENDIX</b>	

## LIST OF TABLES

		<b>Page</b>
Table 1	The stock solution for Bold's Basal Medium (BBM)	16
Table 2	Means $\pm$ standard errors of microalgae biomass (Chlorophyll-a) in different cultures during the experimental period	22
Table 3	Means $\pm$ standard errors of ammonia-N, (NH <sub>4</sub> -N) level and NH <sub>4</sub> -N percent reduction in the culture of alginate beads during the experimental period	27
Table 4	Means $\pm$ standard errors of nitrate-N, (NO <sub>3</sub> -N) level and NO <sub>3</sub> -N percent reduction in the culture of alginate beads during the experimental period	28

## LIST OF FIGURES

		<b>Page</b>
Figure1	Micrographs show three difference appearance of <i>Stigeoclonium nanum</i> . (A) End tips of the cell (B) Structure of middle cell and (C) Clumping of cells	8
Figure 2	The randomly arrangement of the sample in the incubator	15
Figure 3	Means $\pm$ standard errors of chlorophyll-a contents in free suspension <i>Stigeoclonium nanum</i> and in alginate beads	23
Figure 4	Cummulative ammonia-N loading in the culture of <i>S.nanum</i> (normal culture and immobilized in alginate beads)	24
Figure 5	Ammonia-N and nitrate-N uptake in alginate beads by <i>S.nanum</i>	26
Figure 6	Levels of ammonia-N and nitrate-N in treatment E (%)	29

## LIST OF ABBREVIATIONS AND SYMBOLS

ml	milliliter
g	gram
$\mu$ l	microliter
%	percent
$^{\circ}$ C	degree centigrade
$\text{ml L}^{-1}$	milligram per liter
$\mu\text{mol}$	micro mole
TAN	total ammonia nitrogen
$\text{NO}_3\text{-N}$	nitrate nitrogen
$\text{NH}_4\text{-N}$	ammonia nitrogen

## CHAPTER 1

### 1.0 INTRODUCTION

The aquaculture sector has recorded an annual growth rate of about 10 percent in the last 5 years and has now grown into a profitable and sustainable industry (FAO, 2013). The rapid expansion of intensive aquaculture industries, are often accompanied by waste production mainly from uneaten feed, feces and organic residues. These wastes increase nutrients in water, in particular nitrogen and phosphorous, and water quality can rapidly deteriorates as a result (Beveridge 1996). Microbial decomposition of organic matter leads to increased levels of total ammonia nitrogen (TAN) and nitrite-N, which are very harmful to fish even at low concentrations (Meade, 1985; Jiménez-Montealegre et al., 2002; Torres-Beristain et al., 2006). The TAN present in the water may be transformed into nitrite, nitrate and gaseous nitrogen. Bacteria present in the water and sediment carry out these nitrogen transformations by nitrification and denitrification processes.

In general, both TAN and nitrate can be assimilated by microalgae, present in the water column. The microalgae can be consumed by cultured organisms (Turker et al., 2003). In stagnant ponds TAN tends to accumulate within the system due to insufficient nitrification activity (Grommen et al., 2002). Therefore, presence of microalgae may be helpful to reduce nitrogen in the water. There are many groups of

microalgae which can be found in aquatic ecosystem for example green algae (Chlorophyta), red algae (Rhodophyta), blue-green algae (Cyanobacteria) and diatoms (Bacillariophyta). In this research, *Stigeoclonium nanum*, a green filamentous fresh water microalga, was studied. Immobilization of microalgae in alginate beads was studied because in this form, they are easy to manage, microalgae biomass can be maintained at certain volume, and algae bloom can be prevented in the culture system.

This research investigated the growth of *S.nanum* culture in normal suspension and *S.nanum* immobilized in alginate beads. Futhermore, this research investigated the efficiency of immobilized *S.nanum* to utilize ammonium-N and nitrate-N in water.

### **1.1 Research objectives**

- i. To study the growth of the filamentous microalgae (*Stigeoclonium nanum*) immobilized in alginate beads and in free suspension condition
- ii. To measure the uptake of the ammonia-N and nitrate-N, by immobilized filamentous microalgae (*Stigeoclonium nanum*)

## REFERENCES

- Abdel Hameed M.S. (2002). Effect of Immobilization on growth and photosynthesis of the green alga *Chlorella vulgaris* and its efficiency in heavy metals removal. *Bull. Fac. Sci. Assiut Uni.*, **31**: 233-240.
- Abdel Hameed M.S., Hammouda O (2007). Review: Biotechnological potential uses of immobilized algae. *Int. J. Agric. Biol.* **9**: 183-192.
- APHA (2012). Standard method for the examination of waters and wastewaters, 22nd., Washington, DC, USA. pp654.
- Arabi H, Tabatabaei Yazdi M, Faramarzi M.A (2010) Influences of whole microalgal cell immobilization and organic solvent on the bioconversion of androst-4-en-3,17-dione to testosterone by *Nostoc muscorum*. *J Mol Catal B: Enzym* **62**:213–217.
- Beveridge, M.C.M., (1996). Cage aquaculture, 2nd ed. Fishing News Books, BlackwellScience Ltd, Oxford, UK.pp436.
- Butcher, R..W., (1955). Relation between the biology and the polluted condition of the Trent. *Verb. Int. Vet. Limnol.*, **12**: 823-827.
- Bruns, I., Siebert, A., Baumbach, R., Miersch, J., Gunther, D., Markert, B., Krauss, G.-J., (1995). Analysis of heavy metals and sulphur-rich compounds in the water moss *Fontinalis antipyretica* L. ex Hedw. *Fres. J. Anal. Chem.* **353**, 101–104.
- Cassidy M.B., Lee H, Trevors JT (1996) Environmental applications of immobilized microbial cells: a review. *J Ind Microbiol Biotechnol* **16**:79–101.
- Chappell, K.R., Goulder, R., (1994). Epilithic extracellular enzyme activity in a zinc contaminated stream. *Bull. Environ. Contam. Toxicol.* **52**, 305\_310.
- Chevalier P, de la Noue J (1985). Wastewater nutrient removal with microalgae immobilized in carrageenan. *Enz. Microb. Technol.* **7**: 621-24.
- Crab R, Avnimelech Y, Defoirdt T, Bossier P, Verstraete W (2007) Nitrogen removal techniques in aquaculture for a sustainable production. *Aquaculture* **270**:1–14.
- Craggs R.J., Mc Auley P.J. and Smith V.J., (1997). Waste water nutrient removal by marine algae grown on a corrugated raceway. *Water Res.***31**: 1701-1707.



- de-Bashan L.E., Hernandez J.P., Morey T, Bashan Y (2004) Microalgae growth-promoting bacteria as “helpers” for microalgae: a novel approach for removing ammonium and phosphorus from municipal wastewater. *Water Res* **38**:466–474.
- de-Bashan L.E., Bashan Y (2008) Joint Immobilization of plant growth-promoting bacteria and green microalgae in alginate beads as an experimental model for studying plant–bacterium interactions. *Appl Environ Microbiol* **74**:6797–6802.
- de-Bashan L.E., Bashan Y (2010) Immobilized microalgae for removing pollutants: review of practical aspects. *Bioresour Technol* **101**:1611–1627.
- de la Noue J, Proulx D. (1988). Biological tertiary treatment of urban wastewaters with chitosan-immobilized *Phormidium*. *Appl. Microbiol. Biotechnol.* **29**: 292-297.
- de la Noue J, Laliberte G, Proulx D (1992). Algae and wastewater. *J. Appl. Phycol.* **4**: 247-254.
- Deniseger, J., Austin, A., Lucey, W.P., (1986). Periphyton communities in a pristine mountain stream above and below heavy metal mining operations. *Freshwater Biol.* **16**, 209–218.
- El Samra, M.I., Olàh, J., (1979). Significance of nitrogen fixation in fish ponds. *Aquaculture* **18**, 367–372.
- Ernst, W.H.O., Schat, H., Verkleij, J.A.C., (1990). Evolutionary biology of metal resistance in *Silene bulgaris*. *Evolut. Trends Plants* **4**, 45–51.
- Faafeng B.A., van Donk E., Källqvist ST (1994) In situ measurement of algal growth potential in aquatic ecosystems by immobilized algae. *J Appl Phycol* **6**:301–308.
- Fallowfield H.J., Garrett M.K. (1985). The photosynthetic treatment of pig slurry in temperate climatic conditions: A pilot plant study. *Agric. Waste* **12**: 111–136.
- FAO. 2013 Report of the Fourth Aquaculture Network for Africa (ANAF) Annual Meeting. Entebbe, Uganda, 4–6 December 2012. *FAO Fisheries and Aquaculture Report* No. **1049**. Rome. 82.
- Fierro, S., del Pilar Sánchez-Saavedra, M., Copalcúaa, C., (2008). Nitrate and phosphate removal by chitosan immobilized *Scenedesmus*. *Bioresour. Technol.* **99**, 1274– 1279.

- Fjordingstad, E., (1965). Taxonomy and saprobic valency of benthic phytomicroorganisms. *Int. Rev. ges. Hydrobiol.*, **4**: 548-549.
- Foster, P.L., (1982). Species associations and metal contents of algae from rivers polluted by heavy metals. *Freshwater Biol.* **12**, 17-39.
- Foster, P.L., (1982). Metal resistance of Chlorophyta from rivers polluted by heavy metals. *Freshwater Biol.* **12**, 1-61.
- Francke, J.A., Rhebergen, L.J., (1982). Euryhaline ecotypes in some species of *Stigeoclonium* Kütz. *Br. Phycol. J.* **17**, 135-145.
- Francke, J.A., Ten Cate, H.J., (1980). Ecotypic differentiation in response to nutritional factors in the algal genus *Stigeoclonium* Kütz. (Chlorophyceae). *Br. Phycol. J.* **15**, 343-355.
- Grommen, R., Van Hauteghem, I., Van Wambeke, M., Verstraete, W., (2002). An improved nitrifying enrichment to remove ammonium and nitrite from freshwater aquaria systems. *Aquaculture* **211**, 115-124.
- Hall B.M., McLaughlin A.J., Leung K.T., Trevors J.T., Lee H (1998) Transport and survival of alginate-encapsulated and free Lux-Lac marked *Pseudomonas aeruginosa* UG2Lr cells in soil. *FEMS Microbiol Ecol* **36**:51-61.
- Harding, J.P.C., Whitton, B.A., (1976). Resistance to zinc of *Stigeoclonium tenue* in the field and the laboratory. *Br. Phycol. J.* **11**, 417-426.
- Harding, J.P.C., Whitton, B.A., (1977). Environmental factors reducing the toxicity of zinc to *Stigeoclonium tenue*. *Br. Phycol. J.* **12**, 17-21.
- Hirooka, T., Nagase, H., Uchida, K., Hiroshige, Y., Ehara, Y., Nishikawa, J., Nishihara, T., Miyamoto, K., Hirata, Z., (2005). Biodegradation of bisphenol A and disappearance of its estrogenic activity by the green alga *Chlorella fusca* var. *Vacuolata*. *Environ. Toxicol. Chem.* **24**, 1896-1901.
- Jácome-Pilco, C.R., Cristiani-Urbina, E., Flores-Cotera, L.B., Velasco-García, R., Ponce-Noyola, T., Cañizares-Villanueva, R.O., (2009). Continuous Cr(VI) removal by *Scenedesmus incrassatulus* in an airlift photobioreactor. *Bioresour. Technol.* **100**, 2388-2391.
- Jiménez-Montealegre, R., Verdegem, M.C.J., van Dam, A., Verreth, J.A.J., 2002. Conceptualization and validation of a dynamic model for the simulation of nitrogen transformations and fluxes in fish ponds. *Ecol. Model.* **147**, 123-152.

- Kelly, M.G., Whitton, B.A., (1989a). Interspecific differences in Zn, Cd and Pb accumulation by freshwater algae and bryophytes. *Hydrobiologia* **175**, 1–11.
- Kelly, M.G., Whitton, B.A., (1989). Relationship between accumulation and toxicity of zinc in *Stigeoclonium* (Chaetophorales, Chlorophyta). *Phycologia* **28**, 51–517.
- Kinross, J.H., Read, P.A., Christofi, N., (2000). The influence of pH and aluminium on the growth of filamentous algae in artificial streams. *Arch. Hydrobiol.* **149**, 67–86.
- Kotrba, P., Macek, T., Ruml, T., (1999). Heavy metal-binding peptides and proteins in plants. A review. *Collect. Czech. Chem. Commun.* **64**, 1057–1086.
- Kützing, F.T. (1843). *Phycologia generalis* oder Anatomie, Physiologie und Systemkunde der Tange... Mit 80 farbig gedruckten Tafeln, gezeichnet und gravirt vom Verfasser. pp. [part 1]: [i]-xxxii, [1]-142, [part 2:] 143-458, 1, err.), pls 1-80. Leipzig: F.A. Brockhaus
- Lau P.S., Tam N.F.Y., Wong Y.S. (1995). Effect of algal density on nutrient removal from primary settled wastewater. *Environ. Pollut.*, **89**: 59-66.
- Lau P.S., Tam N.F.Y., Wong Y.S. (1997). Wastewater Nutrients (N and P) Removal by Carrageenan and Alginate Immobilized *Chlorella vulgaris*. *Environ. Technol.*, **18**: 945-951.
- Lau P.S., Tam N.F.Y., Wong Y.S. (1998). Operational optimization of batchwise nutrient removal from wastewater by carrageenan immobilized *Chlorella vulgaris*. *Water Sci. Technol.* **38**: 185–192.
- Lavoie A, de la Noue J. (1985). Hyperconcentrated cultures of *Scenedesmus obliquus*: A new approach for wastewater biological tertiary treatment. *Water Res.*, **19**, 1437-1442.
- Loez, C.R., Topalian, M.L., Salibian, A., (1995). Effects of zinc on the structure and growth dynamics of a natural freshwater phytoplankton assemblage reared in the laboratory. *Environ. Pollut.* **88**, 275–281.
- Lebeau, T., Robert, J.M., (2006). Biotechnology of immobilized micro-algae: a culture technique for the future? In: Rao, S. (Ed.), *Algal Cultures, Analogues of Blooms and Applications*. Science Publishers, Enfield, NH, pp. 801–837.

- Mallick N, Rai LC (1993). Influence of culture density, pH, organic acids and divalent cations on the removal of nutrients and metals by immobilized *Anabaena doliolum* and *Chlorella vulgaris*. *World J. Microb. Biotechnol.* **9**: 196-201.
- Mallick, N., (2002). Biotechnological potential of immobilized algae for wastewater N, P and metal removal: a review. *BioMetals* **15**, 377–390.
- Mallick N., (2003) Biotechnological potential of *Chlorella vulgaris* for accumulation of Cu and Ni from single and binary metal solutions. *World J Microbiol Biotechnol* **19**:695–701.
- McLean, R.O., Benson-Evans, K., (1974). The distribution of *Stigeoclonium tenue* Kütz. in southern Wales in relation to its use as an indicator of organic pollution. *Br. Phycol. J.* **9**, 83–89.
- Meade, J.W., (1985). Allowable ammonia for fish culture. *Prog. Fish-Cult.* **47**, 135–145.
- Mohapatra S, Chakraborty T, Kumar V, Deboeck G, Mohanta KN (2012) Aquaculture and stress management: a review of probiotic intervention. *J Anim Physiol Anim Nutr*:1–26. doi:10.1111/j.1439-0396.2012.01301.x.
- Moreno-Garrido, I., (2008). Microalgae immobilization: current techniques and uses. *Bioresour. Technol.* **99**, 3949–3964.
- Moreno-Garrido I, Codd G.A., Gadd G.M., Lubian L.M. (2002) Acumulación de Cu y Zn por células microalgales marinas *Nannochloropsis gaditana* (Eustigmatophyceae) inmovilizadas en alginato de calcio. *Cienc Mar* **28**:107–119.
- Muthukumaran M, Raghavan BG, Subrahmanian VV and Sivasubrahmanian V (2005). Bioremediation of industrial effluent using micro algae. *Indian Hydrobiology.* **7**: 105 – 122.
- Neumann, D., Zur Nieden, U., Schweiger, W., Leopold, I., Lichtenberger, O., (1997). Heavy metal tolerance of *Minuartia verna*. *J. Plant Physiol.* **151**, 101\_108.
- Nichols H.W., and H.C. Bold (1965) *J. Phycology* **1**, 34-38.
- Ninawe AS, Selvin J (2009) Probiotics in shrimp aquaculture: avenues and challenges. *Crit Rev Microbiol* **35**:43–66.

- Olguin EJ (2003). Phycoremediation: key issues for cost effective nutrient removal Processes. *Biotechnol Adv.* **22**: 81- 90.
- Oswald WJ (1998). Microalgae and wastewater treatment. Microalgal biotechnology, Cambridge University Press, Cambridge. 691 – 707.
- Palmer, C. M., (1959). Algae in water supplies. Publ. U.S. *Public Health Service*, **657**: 1-88.
- Pawlik, B., Skowron´ ski, T., (1994). Transport and toxicity of cadmium: its regulation in the cyanobacterium *Synechocystis aquatilis*. *Environ. Exp. Bot.* **34**, 225–233.
- Pawlik-Skowron´ ska, B., (2001). Phytochelatin production in freshwater algae *Stigeoclonium* in response to heavy metals contained in mining water; effects of some environmental factors. *Aquat. Toxicol.* **52**, 241\_/249.
- Piedrahita, R.H., (2003). Reducing the potential environmental impact of tank aquaculture effluents through intensification and recirculation. *Aquaculture* **226**, 35–44.
- Prasad, K. & Kadokawa, J.-I. (2009). Alginate-based blends and nano / microbeads. *Microbiol. Monographs* **13**:175–210.
- Read, P., Fernandes, T., (2003). Management of environmental impacts of marine aquaculture in Europe. *Aquaculture* **226**, 139–163.
- Rijstenbil, J.W., Poortvliet, T.C.W., (1992). Copper and zinc in estuarine water: chemical speciation in relation to bioavailability to the marine planktonic diatom *Ditylum brightwelli*. *Environ. Toxicol. Chem.* **11**, 1615–1625.
- Robinson P.K., Reeve J.O., Goulding K.H. (1988). Kinetics of phosphorus uptake by immobilized *Chlorella*. *Biotechnol. Lett.* **10**: 17-20.
- Romo S, Perez-Martinez C (1997). The use of immobilization in alginate beads for long-term storage of *Pseudoanabaena galeata* (Cyanobacteria) in the laboratory. *J. Phycol.* **33**: 1073-1076.
- Sanita di Toppi, L., Gabrielli, L., (1999). Response to cadmium in higher plants. *Environ. Exp. Bot.* **41**, 105\_/130.

- Say, P.J., Whitton, B.A., (1981). Chemistry and plant ecology of zinc-rich streams in the Northern Pennines. In: Say, P.J., Whitton, B.A. (Eds.), *Heavy Metals in Northern England: Environmental and Biological Aspects*. Department of Botany, University of Durham, Durham, UK, pp. 55–64.
- Simons, J., Van Beem, A.P., De Vries, P.J.R., (1986). Morphology of the prostrate thallus of *Stigeoclonium* (Chlorophyceae, Chaetophorales) and its taxonomic implications. *Phycologia* **25**, 210–220.
- Silverberg, B.A., (1975). Ultrastructural localisation of lead in *Stigeoclonium tenue* (Chlorophyceae, Ulotrichales) as demonstrated by cytochemical and X-ray microanalysis. *Phycologia* **14**, 265–274.
- Skowronski, T., (1984). Energy-dependent transport of cadmium by *Stichococcus bacillaris*. *Chemosphere* **13**, 1379–1384.
- Skowronski, T., Rzczycka, M., (1980). Effect of high zinc concentrations on the growth of *Stichococcus bacillaris* and *Chlorella vulgaris*. *Acta Microbiol. Polon.* **29**, 389–396.
- Smidsrød, O. & Skjåk-Bræk, G. (1990). Alginate as immobilization matrix for cells. *Trends Biotechnol.* **8**:71–8.
- Stauber, J.L., Florence, T.M., (1990). Mechanism of toxicity of zinc to the marine diatom *Nitzschia closterium*. *Mar. Biol.* **105**, 519–524.
- Stein, J. (ED.) *Handbook of Phycological methods. Culture methods and growth measurements*. Cambridge University Press. 448 pp.
- Stenson L.R., Klaenhammer T.R., Swaisgood H.E. (1987) Calcium alginate-immobilized cultures of lactic Streptococci are protected from bacteriophages. *J Dairy Sci* **70**:1121–1127.
- Sugiura, S.H., Marchant, D.D., Wiggins, T., Ferraris, R.P., (2006). Effluent profile of commercially used low-phosphorus fish feeds. *Environ. Pollut.* **140**, 95–101.
- Talbot P, de la Noue J. (1993). Tertiary treatment of wastewater with *Phormidium bohneri* (Schmidle) under various light and temperature conditions. *Water Res.* **27**: 153-159.
- Takamura, N., Kasai, F., Watanabe, M.M., (1989). Effects of Cu, Cd and Zn on photosynthesis of freshwater benthic algae. *J. Appl. Phycol.* **1**, 39–52.

- Takamura, N., Hatakeyama, S., Sugaya, Y., (1990). Seasonal changes in species composition and production of periphyton in an urban river running through an abandoned copper mining region. *Jpn. J. Limnol.* **4**, 225–235.
- Tam N.F.Y., Wong Y.S. (1989). Wastewater nutrient removal by *Chlorella pyrenoidosa* and *Scenedesmus* sp. *Environ. Pollut.*, **58**: 19-34.
- Tam N.F.Y., Lau P.S., Wong Y.S. (1994). Wastewater inorganic N and P removal by immobilized *Chlorella vulgaris*. *Wat. Sci. Tech.* **30**: 369-374.
- Tanaka H, Ohta T, Harada S, Ogonna JC, Yajima M (1994) Development of a fermentation method using immobilized cells under unsterile conditions. 1. Protection of immobilized cells against anti-microbial substances. *Appl Microbiol Biotechnol* **41**:544–550.
- Thompson, F.L., Abreu, P.C., Wasielesky, W., (2002). Importance of biofilm for water quality and nourishment in intensive shrimp culture. *Aquaculture* **203**, 263–278.
- Torres-Beristain, B., Verdegem, M., Kerepeczki, E., Verreth, J., (2006). Decomposition of high protein aquaculture feed under variable oxic conditions. *Water Res.* **40**, 1341–1350.
- Turker, H., Eversole, A.G., Brune, D., (2003). Comparative Nile tilapia and silver carp filtration rates of Partitioned Aquaculture System phytoplankton. *Aquaculture* **220**, 449–457.
- Travieso, L., Pellón, A., Benítez, F., Sánchez, E., Borja, R., O’Farrille, N., Weiland, P., (2002). BIOALGA reactor: preliminary studies for heavy metals removal. *Biochem. Eng. J.* **12**, 87–91.
- Verschuere L, Rombaut G, Sorgeloos P, Verstraete W (2000) Probiotic bacteria as biological control agents in aquaculture. *Microbiol Mol Biol Rev* **64**:655–671.
- Vilchez C, Garabayo I, Marckvichea E, Galvan F, Leon R (2001). Studies on the suitability of alginate-entrapped *Chlamydomonas reinhardtii* cells for sustaining nitrate consumption processes. *Biores. Technol.* **78**: 55-61.
- Vymazal, J., (1984). Short-term uptake of heavy metals by periphyton algae. *Hydrobiologia* **119**, 171–179.
- Weimann, R., (1952). Abwassertypen in Nordrhein-Westfalen. *Schweiz. Z. Hydrol.*, **14**: 372-433.

Weir S.C., Dupuis S.P., Providenti M.A., Lee H., Trevors J.T. (1995) Nutrient-enhanced survival of and phenanthrene mineralization by alginate-encapsulated and free *Pseudomonas* sp. UG14Lr cells in creosote-contaminated soil slurries. *Appl Microbiol Biotechnol* **43**:946–951.

Whitton, B. A. & SAY, P. J., (1975). Heavy metals. In *River Ecology* (Whitton, B. A., Editor). Pp. 286-311. Blackwell, Oxford.

Zohar-Perez C, Chernin L, Chet I, Nussinovitch A (2003) Structure of dried cellular alginate matrix containing fillers provides extra protection for microorganisms against UVC radiation. *Radiat Res* **160**:198–204

