



***EFFECTS OF JASMONIC ACID IN REDUCING CUCUMBER MOSAIC
VIRUS INFECTION AND IMPROVING GROWTH PERFORMANCE OF
CHILI (*Capsicum annuum* L.)***

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By

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**Thesis Submitted to the School of Graduate Studies, Universiti
Putra Malaysia, in Fulfilment of the Requirements for the Degree of
Master of Science**

April 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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Chair: Prof. Mohd Razi Ismail, PhD

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The high demand for chili has made agriculture entirely dependent on application of chemical pesticide to control pests and diseases. Results demonstrated that plants treated with pesticide significantly affected the number of pests such as aphids, thrips and mites which reduced the percentage of disease incidence up to 80%. However, regarding the long-term effect of pesticide toxicity, many researchers have developed alternative approaches which could improve plant productivity and give benefits to human and environmental health. In this study, jasmonic acid (JA), a potential plant elicitor had been tested using different concentrations (0.6 mM and 1.0 mM) and spray frequencies (single, double and triple application) to evaluate the efficacy in promoting growth performances, physiological responses, improving yield and reducing disease incidence. Results demonstrated treatment of 0.6 mM JA with double spray showed the outstanding positive effect compared to others in lowering the disease incidence by 50%. Despite of insignificant in concentration and spray frequencies factors, concentration of JA at 0.6 mM with double spray succeeded in improving high fruit yield which produced similar results to plants treated with concentration 1.0 mM of JA in double and triple sprays. Therefore, the next study to evaluate the effect of JA on chili plant infected with Cucumber Mosaic Virus (CMV) was carried out using concentration of 0.6 mM with double spray. CMV was reported to be one of the most prevalent cucumovirus in Malaysia due to its large host range and insect vectors. The findings revealed that CMV-infected plant treated with JA had the highest inhibitory effect on CMV infection at 30 days post inoculation (dpi) compared to untreated and pesticide-treated plants that were severely damaged. Due to low percentage of disease severity in JA application, the chili yield increased up to 95% and significantly raised the dry matter accumulation in leaves, stems and roots. The elicitation of JA had a significant impact on activities of catalase (CAT), ascorbate peroxidase (APX), guaiacol peroxidase (GPX), ascorbic acid (AsA), α -tocopherol and carotenoids (CAR) in leaves and fruits. In the JA-treated plants a significant increase in AsA, CAT, APX and GPX activities showed that these

antioxidants were essential to detoxify reactive oxygen species (ROS) at 14 dpi (flowering stage) while AsA and α -tocopherol was suggested to become the selective protection against CMV infection at 30 dpi (fruiting stage). Among the enzymatic antioxidants, GPX had the highest activity in chili fruit of JA-treated plants followed by CAT and APX. These results suggested that exogenous application of JA could effectively reduce CMV infection by enhancing activities of enzymatic antioxidants and concentration of non-antioxidant to quench excessive ROS. So, it can be concluded that the optimum concentration and spray frequencies of JA can become a promising tool for reducing CMV disease thus improving the growth performances in chili plants as an alternative method to replace or reduce the pesticide use.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN ASID JASMONIK DALAM MENGURANGKAN JANGKITAN VIRUS
TIMUN MOZEK DAN MENINGKATKAN PRESTASI PERTUMBUHAN CILI
(*Capsicum annuum L.*)**

Oleh

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Permintaan yang tinggi untuk cili menyebabkan sektor pertanian bergantung sepenuhnya pada penggunaan racun perosak untuk mengawal serangga dan penyakit. Hasil kajian ini mendapati penggunaan racun perosak pada pokok telah memberi kesan yang signifikan terhadap bilangan serangga perosak seperti afid, kutu trip dan hamama yang telah mengurangkan sehingga 80% peratusan kadar penyakit. Walau bagaimanapun, disebabkan kesan jangka masa yang panjang penggunaan racun perosak, ramai pengkaji telah menghasilkan pendekatan alternatif untuk memperbanyakkan produktiviti dan memberi kesan yang baik kepada kesihatan manusia dan alam sekitar. Dalam kajian ini, asid jasmonik (JA) yang merupakan elisitor telah diujikaji dengan penggunaan kepekatan (0.6 mM dan 1.0 mM) dan kekerapan semburan (sekali, dua kali dan tiga kali) yang berbeza untuk mengkaji kesan dalam menggalakkan tumbesaran pokok, tindakbalas fisiologi, meningkatkan hasil dan mengurangkan peratusan penyakit. Hasil kajian menunjukkan bahawa rawatan menggunakan kepekatan 0.6 mM sebanyak dua kali semburan memberi kesan yang sangat positif berbanding rawatan yang lain dengan mengurangkan kejadian penyakit pada 50%. Walaupun faktor kepekatan dan kekerapan semburan tidak signifikan, namun kepekatan JA pada 0.6 mM sebanyak dua kali semburan berjaya meningkatkan hasil yang hampir sama seperti kepekatan JA pada 1.0 mM sebanyak dua dan tiga kali semburan. Oleh itu, kajian seterusnya untuk mengkaji kesan JA pada pokok yang dijangkiti dengan Virus Timun Mozek (CMV) telah dijalankan dengan menggunakan kepekatan 0.6 mM sebanyak dua kali semburan. CMV telah dilaporkan sebagai salah satu daripada cucumovirus yang sangat lazim di Malaysia berikutan kepelbagaian sel perumah dan vektor serangga. Hasil kajian mendapati bahawa pokok yang telah dijangkiti dengan CMV yang dirawat menggunakan JA menunjukkan kesan perencatan terhadap jangkitan CMV pada hari ke-30 inokulasi berbanding dengan kerosakan yang teruk pada pokok yang tidak dirawat dan dirawat dengan racun perosak. Disebabkan peratusan keterukan penyakit yang rendah dalam penggunaan JA, hasil cili telah meningkat sebanyak 95% dan menambahkan pengeluaran bahan kering dalam daun, dahan dan akar. Aplikasi JA juga memberi kesan yang ketara terhadap aktiviti katalase (CAT), askorbat peroksida (APX), guaiacol peroksida (GPX), asid askorbik

(AsA), α -tokoferol dan karotenoid (CAR) di dalam daun dan buah. Peningkatan yang signifikan dalam AsA dan aktiviti-aktiviti CAT, APX dan GPX pada pokok yang dirawat dengan JA menunjukkan antioksidan ini sangat penting untuk menyahtoksikan spesies oksigen reaktif (ROS) pada 14 hari selepas inokulasi (dpi) (peringkat berbunga) manakala AsA dan α -tokoferol terpilih untuk melindungi daripada jangkitan CMV pada 30 dpi (peringkat berbuah). Antara antioksidan enzimatik, GPX mempunyai aktiviti yang tertinggi di dalam buah cili yang dirawat dengan JA, diikuti CAT and APX. Penggunaan JA juga telah meningkatkan jumlah AsA di dalam buah cili berbanding rawatan yang lain. Hasil kajian ini mencadangkan bahawa penggunaan JA secara luaran telah mengurangkan jangkitan CMV secara efektif dengan meningkatkan aktiviti antioksidan enzimatik dan kandungan antioksidan bukan enzimatik untuk menghapuskan ROS. Oleh itu, secara kesimpulannya, kepekatan dan kekerapan semburan yang optima boleh menjadikannya sebagai satu cara untuk mengurangkan CMV sekaligus meningkatkan kadar tumbesaran pokok cili serta sebagai salah satu langkah alternatif untuk menggantikan atau mengurangkan penggunaan racun perosak.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

3T3	Mouse Embryonic Fibroblast Cell line
ABA	Abscisic Acid
ANOVA	Analysis of Variance
AP	Antibody-alkaline Phosphate
AOS	Allene Oxide Synthase
APX	Ascorbate Peroxidase
ArOU	Aryloxy radical
AsA	Ascorbic Acid
BSA	Bovine Serum Albumin
CAR	Carotenoid
CAT	Catalase
CCD	Coconut Coir Dust
CD	Cluster Designation
CEC	Cation Exchange Capacity
CLCV	Chili Leaf Curl Virus
CMV	Cucumber Mosaic Virus
CVMV	Chili Veinal Mottle Virus
DAS-ELISA	Double Antibody Sandwich-Enzyme Linked Immunosorbent Assay
DAT	Day After Transplanting
DHA	Dehydroascorbate
DI	Disease Incidence
dpi	Days Post Inoculation
DMSO	Dimethyl Sulfoxide
DNA	Deoxyribonucleic Acid
EC	Electrical Conductivity
ECE	Ethanol Crude-dried Extract
EDTA	Ethylenediaminetetraacetic Acid
EFB	Empty Fruit Bunch
F1	First Generation
GA	Gibberellic Acid
GGPP	Geranylgeranyl Diphosphate
GPX	Guaiacol Peroxidase
GR	Glutathione Reductase
GSH	Glutathione
H ₂ O ₂	Hydrogen Peroxide
Ha	Hectare
HepG2	Hepatocellular Carcinoma Cell Line
hpi	Hours Post Inoculation
HO·	Hydroxyl Radical
HOCl	Hypochlorous Acid
IAA	Indole Acetic Acid
IUE	Irrigation Use Deficiency
JA	Jasmonic Acid
LA	Linolenic Acid
LOX	Lipoxygenase
LSD	Least Significance Difference
MARDI	Malaysian Agricultural Research and Development Institute

MD	Monodehydroascorbate Radical
MDAR	Monodehydroascorbate Reductase
MEP	Methyerythritol 4-phosphate
MeJA	Methyl Jasmonate
MRL	Maximum Residue Level
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium Bromide
n	Number of Plants
NADPH	Nicotinamide Adenine Dinucleotide Phosphate
NO·	Nitric Oxide
NO ₂	Nitrogen Dioxide
¹ O ₂	Singlet Oxygen
O ₃	Ozone
O ²⁻	Superoxide Anion
OD	Optical Density
ONOO·	Peroxynitrite
OPDA	Cyclopentanone 12-oxophytodienoic Acid
PHI	Pre-harvested Interval
ppm	Parts Per Million
PPO	Polyphenol Oxidase
PPR	Portable Photosynthesis Rate
PS	Photosensitizer
PS II	Photosystem II
RCBD	Randomized Complete Block Design
RNA	Ribonucleic Acid
RNS	Reactive Nitrogen Species
RO ₂ ·	Peroxyl
RO·	Alkoxy
ROS	Reactive Oxygen Species
RONs	Reactive Oxygen Nitrogen Species
Rpm	Rotation Per Minute
S.E	Standard Error
SA	Salicylic Acid
SAS	Statistical Analysis System
SChE	Serum-cholinesterase
SOD	Superoxide Dismutase
TCA	Trichloroacetic Acid
TCV	Turnip Crinkle Virus
TMV	Tobacco Mosaic Virus
ULVC	Urdbean Leaf Crinkle Virus



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CHAPTER 1

INTRODUCTION

Chili (*Capsicum annum L.*) is one of the most valuable spice crops in Malaysia that occupied about 3523 ha area in major producing states such as Kelantan, Pahang, Johor and Perak in 2017 (DOA, 2017). Although local chili production reaches approximately 50,299 metric tons annually, the yield produced is deficient to meet domestic demands (2.0 kg/year per capita consumption) and therefore additional import dependency ratio (IDR) had raised from 38.7% to 52.9% in 2013 to 2014 (DOSM, 2017). Thailand becomes the leading exporter which represent 89% in fresh chili supply followed by China, India, Vietnam and Indonesia (APEDA, 2007). Besides limited cultivation areas, other predominant factor contributing to low chili production are the diseases infected by fungus, bacteria and virus. Among these, viral diseases are considered to be the major limiting factor causing yield losses based on surveys conducted by (Schreinemachers *et al.*, 2015) tropical and subtropical Asia.

Cucumber Mosaic Virus (CMV) is one of the important viral pathogens commonly found in chili plants in Malaysia which cause the severest damage and contribute to yield loss (Roff, 1992). According to field crop surveys, CMV was discovered to be the most prevalent on chili with higher disease incidence that can reach up to 80% (Iqbal *et al.*, 2012; Simón *et al.*, 2016). In addition, CMV damage on chili are influenced differently by time of infection as early and late infection cause yield losses at 10-15% and exceed 60% respectively (Rafidah *et al.*, 2016). The severely infected plants with mosaic disease produce less flowers and deformed fruits (Khan *et al.*, 2006) which result in drastic reduction in marketable yield and fruit quality. Consequently, chili being sold at higher prices due to low production and higher demands.

Regarding these problems, farmers are entirely relying on chemical pesticides application including insecticides, fungicides, molluscicide and weedicide to control the pest and diseases. About 78.4% of chili growers mentioned the input cost including pesticides and other chemicals greatly increase in order to reduce numbers of pest (Arumugam *et al.*, 2012). Besides the rising disease management cost, the inaccurate in handling the register pesticide and illegally use of unregistered pesticide can cause severe environmental consequences and produce carcinogenic chili (Mesnage *et al.*, 2014). In Malaysia, based on Food Safety Act 1983, pesticide can be applied by farmers without reaching the Maximum Residue Level (MRL), as overuse will cause harm to human health. Despite that, most of the farmers disobey the rule without considering the consequences of active ingredients that will remain in the fruits.

In order to overcome the disease, many researchers have developed eco-friendly approaches like cultural methods, use of chemical and biological fungicide and resistant cultivars (Than *et al.*, 2008). However, most of these options are still unable to control the diseases. Jasmonic acid (JA) was first reported by Farmer and Ryan (1992) to be a potential elicitor to induce disease resistance against insect and pathogen attack. The effectiveness of JA in defence is also proved in many crops such as alfalfa and tobacco

(Farmer and Ryan, 1992), arabidopsis (Turner *et al.*, 2002), grapes (Omer *et al.*, 2000), tomato (Cooper and Rieske, 2008) and chili (Awang *et al.*, 2013). The positive effects of JA on antioxidant capacity in many studies demonstrated that JA application are able to increase antioxidants thereby causing alleviation of oxidative stress in plants (Qiu *et al.*, 2014; Asghari and Hasanlooe, 2015). The reports regarding JA treatment succeeded in inhibiting CMV (Luo *et al.*, 2011) suggested JA can improve plant resistance and become capable applicator to give plant protection in fields (Shang *et al.*, 2011).

Although the use of pesticides facilitate the disease management strategy, it leads to the accumulation of toxic residues in crop which affect human health and the ecosystems (Williamson *et al.*, 2008; Zhou and Jin, 2009). So, in this study, we would like to develop an eco-friendly control strategy to manage virus diseases and improve yields in chili plants. Besides, this study can help the farmers to reduce the maintenance cost and make pesticide-free chili production. The main objective of this study is to evaluate the effect of jasmonic acid as elicitor to reduce the disease severity in CMV infection, increase antioxidants activities for plant defence which in turn improve the yield production of chili.

The three main hypotheses to be tested in the present study were:

- 1) Cultivation of chili without pesticides application increase number of pest and diseases severely affect plant growth and cause greater yield loss.
- 2) Application of jasmonic acid with optimum concentration and spray frequencies enhance growth performance, chili yields, physiological responses and reduce disease incidence.
- 3) Selected concentration and spray frequencies of jasmonic acid reduce Cucumber Mosaic Virus (CMV) infection and improving growth performance by increasing biochemical responses.

The objectives of the present study were:

- 1) To evaluate the growth performances of chili plants with and without pesticide application.
- 2) To determine the concentrations and spray frequencies of jasmonic acid application in chili plants.
- 3) To study the effect of jasmonic acid on chili plant infected with Cucumber Mosaic Virus (CMV).

REFERENCES

- Abdel-Hameed, E.-S. S., Bazaid, S. A., Shohayeb, M. M. & El-Sayed, M. M. (2012). Phytochemical Studies and Evaluation of Antioxidant, Anticancer and Antimicrobial Properties of *Conocarpus erectus* L. Growing in Taif, Saudi Arabia. *European Journal of Medicinal Plants*, 2(2): 93-112.
- Agrawal, R. & Patwardhan, M. V. (1993). Production of peroxidase enzyme by callus cultures of *Citrus aurantifolia* S. *Journal of the Science of Food and Agriculture*, 61(3): 377-378.
- Agriculture and Processed Food Products Export Development Authority. (2007). Market Fresh Fruit & Vegetables, Malaysia. Retrieved from <http://apeda.gov.in/apedawebsite/Archive/ViewAll.htm>
- Agrios, G. N. (2005). Plant diseases caused by viruses. In G. N. Agrios (Ed.), *Plant Pathology. Fifth Edition*. (pp. 724–820). San Diego CA: Elsevier Academia Press.
- Ahmad, A., Ismail, M. R., Yusop, M. K., Mahmood, M. & Mohd, S. (2004). Physical and chemical properties of coconut coir dust and oil palm empty fruit bunch and the growth of hybrid heat tolerant cauliflower plant. *Pertanika Journal of Tropical Agricultural Science*, 27(2): 121-133.
- Ahmad, P., Jaleel, C. A., Salem, M. A., Nabi, G. & Sharma, S. (2010). Roles of enzymatic and nonenzymatic antioxidants in plants during abiotic stress. *Critical Reviews in Biotechnology*, 30(3): 161-175.
- Ahmad, P., Rasool, S., Gul, A., Sheikh, S. A., Akram, N. A., Ashraf, M., Kazi A. M. & Guzel, S. (2016). Jasmonates: multifunctional roles in stress tolerance. *Frontiers in Plant Science*, 7(813): 1-15.
- Ahmed, P., Bhagawati, P., Das, S. K., Kalita, M. C. & Das, S. (2013). Hypersensitive reaction and anatomical changes of young tea leaf (*Camellia sinensis*, clone TV1) during feeding by tea mosquito bug (*Helopeltis theivora* Waterhouse: Hemiptera: Miridae). *International Journal of Current Microbiology and Applied Science*, 2(8): 187-195.
- Ahuja, K. D. K. & Ball, M. J. (2006). Effects of daily ingestion of chilli on serum lipoprotein oxidation in adult men and women. *British Journal of Nutrition*, 96(2): 239-242.
- Akula, R. & Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signaling & Behavior*, 6(11): 1720-1731.
- Alagarmalai, J., Grinberg, M., Perl-Treves, R. & Soroker, V. (2009). Host selection by the herbivorous mite *Polyphagotarsonemus latus* (Acari: Tarsonemidae). *Journal of Insect Behavior*, 22(5): 375-387.
- Alavanja, M. C. R. (2009). Introduction: Pesticides use and exposure, extensive worldwide. *Reviews on Environmental Health*, 24(4): 303-310.

- Alegbejo, M. D. & Abo, M. E. (2002). Ecology, epidemiology and control of pepper vein mottle virus (PVMV), genus Potyvirus, in West Africa. *Journal of Sustainable Agriculture*, 20(2): 5-16.
- Ali, A. & Kobayashi, M. (2010). Seed transmission of Cucumber mosaic virus in pepper. *Journal of Virological Methods*, 163(2): 234-237.
- Amako, K., Chen, G.-X. & Asada, K. (1994). Separate assays specific for ascorbate peroxidase and guaiacol peroxidase and for the chloroplastic and cytosolic isozymes of ascorbate peroxidase in plants. *Plant and Cell Physiology*, 35(3): 497-504.
- Amoako, S., Yahaya, A., & Sarfo, J. K. (2015). Catalase activity of cassava (*Manihot esculenta*) plant under African cassava mosaic virus infection in Cape coast, Ghana. *African Journal of Biotechnology*, 14(14): 1201-1206.
- Anand, P., Kwak, Y., Simha, R. & Donaldson, R. P. (2009). Hydrogen peroxide induced oxidation of peroxisomal malate synthase and catalase. *Archives of Biochemistry and Biophysics*, 491(1): 25-31.
- Andrews, P. K., Fahy, D. A. & Foyer, C. H. (2004). Relationships between fruit exocarp antioxidants in the tomato (*Lycopersicon esculentum*) high pigment-1 mutant during development. *Physiologia Plantarum*, 120(4): 519-528.
- Apel, K. & Hirt, H. (2004). Reactive Oxygen Species: Metabolism, Oxidative Stress, and Signal Transduction. *Annual Review of Plant Biology*, 55(1): 373-399.
- Arumugam, N., Wan Muda, W. M. & Aisyah, N. (2012). Chili Growers' Experience with Contract Farming in Malaysia. *Journal of International Food & Agribusiness Marketing*, 24(2): 137-147.
- Asghari, M. & Hasanlooe, A. R. (2015). Interaction effects of salicylic acid and methyl jasmonate on total antioxidant content, catalase and peroxidase enzymes activity in "Sabrosa" strawberry fruit during storage. *Scientia Horticulturae*, 197: 490-495.
- Asghari, M. & Hasanlooe, A. R. (2016). Methyl jasmonate effectively enhanced some defense enzymes activity and Total Antioxidant content in harvested "Sabrosa" strawberry fruit. *Food Science & Nutrition*, 4(3): 377-383.
- Ashfaq, M., Khan, M. A., Javed, N., Mughal, S. M., Shahid, M. & Sahi, S. T. (2010). Effect of urdbean leaf crinkle virus infection on total soluble protein and antioxidant enzymes in blackgram plants. *Pakistan Journal of Botany*, 42(1): 447-454.
- Ashfaq, M., Iqbal, S., Mukhtar, T., Shah, H., Agricultural, N. & Road, P. (2014). Screening For Resistance To Cucumber Mosaic Cucumovirus In Chilli. *Journal of Animal and Plant Sciences*, 24(3): 791-795.
- Aslam, M. N., Mukhtar, T., Ashfaq, M. & Hussain, M. A. (2017). Evaluation of chili germplasm for resistance to bacterial wilt caused by *Ralstonia solanacearum*. *Australasian Plant Pathology*, 46(3): 289-292.

- Awang, N. A., Islam, M. R., Ismail, M. R., Zulkarami, B. & Omar, D. (2013). Effectiveness of different elicitors in inducing resistance in chilli (*Capsicum annuum* L.) against pathogen infection. *Scientia Horticulturae*, 164: 461-465.
- Azidah, A. (2011). Thripidae (Thysanoptera) species collected from common plants and crops in Peninsular Malaysia. *Scientific Reseach and Essays*, 6(24): 5107-5113.
- Barros, A. I., Nunes, F. M., Gonçalves, B., Bennett, R. N. & Silva, A. P. (2011). Effect of cooking on total vitamin C contents and antioxidant activity of sweet chestnuts (*Castanea sativa* Mill.). *Food Chemistry*, 128(1):165-172.
- Basu, S. K. & De, A. K. (2003). Capsicum. In A. K. De (Ed.), *Capsicum* (pp. 3-5). London: CRC Press.
- Beale, S. I. (1978). δ -Aminolevulinic acid in plants: its biosynthesis, regulation, and role in plastid development. *Annual Review of Plant Physiology*, 29(1): 95-120.
- Berger, S., Sinha, A. K. & Roitsch, T. (2007). Plant physiology meets phytopathology: plant primary metabolism and plant-pathogen interactions. *Journal of Experimental Botany*, 58(15-16): 4019-4026.
- Bhatnagar, V. K. (2001). Pesticides pollution: trends and perspectives. *ICMR Bulletin*, 31(9): 87-88.
- Bhattacharya, A., Chattopadhyay, A., Mazumdar, D., Chakravarty, A. & Pal, S. (2010). Antioxidant Constituents and Enzyme Activities in Chilli Peppers. *International Journal of Vegetable Science*, 16(3): 201-211.
- Black, C. A., Karban, R., Godfrey, L. D., Granett, J. & Chaney, W. E. (2003). Jasmonic acid: a vaccine against leafminers (Diptera: Agromyzidae) in celery. *Environmental Entomology*, 32(5): 1196-1202.
- Black, L. L., Green, S. K., Hartman, G. L. & Poulos, M. (1993). *Pepper diseases: a field guide*. CTA.
- Blackman, R. L. & Eastop, V. F. (2007). Taxonomic issues. In H. F. Van Emden & R. Harrington (Eds.), *Aphids as Crop Pests* (pp. 717). Trowbridge: CABI.
- Blokhina, O., Virolainen, E. & Fagerstedt, K. V. (2003). Antioxidants, oxidative damage and oxygen deprivation stress: a review. *Annals of Botany*, 91(2): 179-194.
- Bonfig, K. B., Schreiber, U., Gabler, A., Roitsch, T. & Berger, S. (2006). Infection with virulent and avirulent *P. syringae* strains differentially affects photosynthesis and sink metabolism in Arabidopsis leaves. *Planta*, 225(1): 1-12.
- Borland, A., Elliott, S., Patterson, S., Taybi, T., Cushman, J., Pater, B. & Barnes, J. (2006). Are the metabolic components of crassulacean acid metabolism up-regulated in response to an increase in oxidative burden? *Journal of Experimental Botany*, 57(2): 319-328.

- Bosland, P. W. (1994). Chiles: history, cultivation, and uses. In: G. Charalambous (Ed.), *Spices, Herbs, And Edible Fungi* (pp. 347-366). Amsterdam: Elsevier Science BV.
- Bostock, R. M. (2005). Signal crosstalk and induced resistance: straddling the line between cost and benefit. *Annual Reviews of Phytopathology*, 43: 545-580.
- Boukaew, S., Chuenchit, S. & Petcharat, V. (2011). Evaluation of *Streptomyces* spp. for biological control of *Sclerotium* root and stem rot and *Ralstonia* wilt of chili pepper. *BioControl*, 56(3): 365-374.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2): 248-254.
- Braut, V., Uzest, M., Monsion, B., Jacquot, E. & Blanc, S. (2010). Aphids as transport devices for plant viruses. *Comptes Rendus Biologies*, 333(6): 524-538.
- Britton, G. (1995). Structure and properties of carotenoids in relation to function. *The FASEB Journal*, 9(15): 1551-1558.
- Burton, G. W. & Ingold, K. U. (1984). β -Carotene: an unusual type of lipid antioxidant. *Science*, 224: 569-574.
- Cai, J. H., Xie, K., Lin, L., Qin, B. X., Chen, B. S., Meng, J. R. & Liu, Y. L. (2010). Cotton leaf curl Multan virus newly reported to be associated with cotton leaf curl disease in China. *Plant Pathology*, 59(4): 794-795.
- Cai, K. Z., Dong, T. X. & Xu, T. (2006). The physiological roles and resistance control in stress environment of jasmonates. *Ecology and the Environment*, 15: 397-404.
- Cao, S., Zheng, Y., Wang, K., Jin, P. & Rui, H. (2009). Methyl jasmonate reduces chilling injury and enhances antioxidant enzyme activity in postharvest loquat fruit. *Food Chemistry*, 115(4): 1458-1463.
- Castillo, M. C., Martínez, C., Buchala, A., Métraux, J.-P. & León, J. (2004). Gene-specific involvement of β -oxidation in wound-activated responses in *Arabidopsis*. *Plant Physiology*, 135(1): 85-94.
- Castro-Concha, L. A., Canche-Chuc, I. & Miranda-Ham, M. de L. (2012). Determination of antioxidants in fruit tissues from three accessions of habanero pepper (*Capsicum chinense* Jacq.). *Journal of the Mexican Chemical Society*, 56(1): 15-18.
- Cazzonelli, C. I. & Pogson, B. J. (2010). Source to sink: regulation of carotenoid biosynthesis in plants. *Trends in Plant Science*, 15(5): 266-274.
- Cenzano, A., Vigliocco, A., Kraus, T. & Abdala, G. (2003). Exogenously applied jasmonic acid induces changes in apical meristem morphology of potato stolons. *Annals of Botany*, 91: 915-919.
- Cerda, R., Avelino, J., Gary, C., Tixier, P., Lechevallier, E. & Allinne, C. (2017). Primary and secondary yield losses caused by pests and diseases: Assessment and

modeling in coffee. *Public Library of Science One*, 12(1): e0169133.

- Cesari, I. M., Carvalho, E., Figueiredo Rodrigues, M., Mendonça, B. Dos S., Amôedo, N. D. & Rumjanek, F. D. (2014). Methyl jasmonate: putative mechanisms of action on cancer cells cycle, metabolism, and apoptosis. *International Journal of Cell Biology*, 2014: 572097.
- Chaim, A. Ben, Grube, R. C., Lapidot, M., Jahn, M. & Paran, I. (2001). Identification of quantitative trait loci associated with resistance to cucumber mosaic virus in *Capsicum annuum*. *TAG Theoretical and Applied Genetics*, 102(8): 1213-1220.
- Camps, J., Burger, A.-S., Mitsiadis, T. A., Butler, W. T. & Franquin, J.-C. (2005). Polymerized bonding agents and the differentiation in vitro of human pulp cells into odontoblast-like cells. *Dental Materials*, 21(2): 156-163.
- Charles, D. J. (2012). Capsicum. In D.J. Charles (Ed.), *Antioxidant Properties of Spices, Herbs and Other Sources* (pp. 189-197). Norway: Springer.
- Cheah, U., Kirkwood, R. C. & Lum, K. (1997). Adsorption, desorption and mobility of four commonly used pesticides in Malaysian agricultural soils. *Pest Management Science*, 50(1): 53-63.
- Chen, J., Jarvi, M., Lo, P.-C., Stefflova, K., Wilson, B. C. & Zheng, G. (2007). Using the singlet oxygen scavenging property of carotenoid in photodynamic molecular beacons to minimize photodamage to non-targeted cells. *Photochemical & Photobiological Sciences*, 6(12): 1311-1317.
- Chen, J., Yan, Z. & Li, X. (2014). Effect of methyl jasmonate on cadmium uptake and antioxidative capacity in *Kandelia obovata* seedlings under cadmium stress. *Ecotoxicology and Environmental Safety*, 104: 349-356.
- Chen, Z., Young, T. E., Ling, J., Chang, S.-C. & Gallie, D. R. (2003). Increasing vitamin C content of plants through enhanced ascorbate recycling. *Proceedings of the National Academy of Sciences*, 100(6): 3525-3530.
- Chitarrini, G., Zulini, L., Masuero, D. & Vrhovsek, U. (2017). Lipid, phenol and carotenoid changes in 'Bianca' grapevine leaves after mechanical wounding: a case study. *Protoplasma*, 254(6): 2095-2106.
- Choe, E. & Min, D. B. (2009). Mechanisms of antioxidants in the oxidation of foods. *Comprehensive Reviews in Food Science and Food Safety*, 8(4): 345-358.
- Choudhury, S., Panda, P., Sahoo, L. & Panda, S. K. (2013). Reactive oxygen species signaling in plants under abiotic stress. *Plant Signaling & Behavior*, 8(4): e23681.
- Claiborne, A. (1985). Catalase activity. In: R. A. Greenwald (Ed.), *Handbook of methods for oxygen free radical research* (pp.283-284). Boca Raton, FL: CRC Press.
- Clark, M. F. & Adams, A. N. (1977). Characteristics of the microplate method of enzyme-linked immunosorbent assay for the detection of plant viruses. *Journal of General Virology*, 34(3): 475-483.

- Colombo, M. L. (2010). An update on vitamin E, tocopherol and tocotrienol—perspectives. *Molecules*, 15(4): 2103-2113.
- Colville, L., & Smirnov, N. (2008). Antioxidant status, peroxidase activity, and PR protein transcript levels in ascorbate-deficient *Arabidopsis thaliana* vtc mutants. *Journal of Experimental Botany*, 59(14): 3857–3868.
- Conconi, A., Miquel, M. & Ryan, C. A. (1996). Intracellular levels of free linolenic and linoleic acids increase in tomato leaves in response to wounding. *Plant Physiology*, 111(3): 797-803.
- Conklin, P. L. (2001). Recent advances in the role and biosynthesis of ascorbic acid in plants. *Plant, Cell & Environment*, 24(4): 383-394.
- Conklin, P. L. & Barth, C. (2004). Ascorbic acid, a familiar small molecule intertwined in the response of plants to ozone, pathogens, and the onset of senescence. *Plant, Cell & Environment*, 27(8): 959-970.
- Cooper, J. & Dobson, H. (2007). The benefits of pesticides to mankind and the environment. *Crop Protection*, 26(9): 1337-1348.
- Cooper, W. R. & Rieske, L. K. (2008). Differential responses in American (*Castanea dentata* Marshall) and Chinese (*C. mollissima* Blume) chestnut (Fales: Fagaceae) to foliar application of jasmonic acid. *Chemoecology*, 18(2): 121-127.
- Costache, M. A., Campeanu, G. & Neata, G. (2012). Studies concerning the extraction of chlorophyll and total carotenoids from vegetables. *Romanian Biotechnological Letters*, 17(5): 7703-7708.
- Creelman, R. A. & Mullet, J. E. (1995). Jasmonic acid distribution and action in plants: regulation during development and response to biotic and abiotic stress. *Proceedings of the National Academy of Sciences*, 92(10): 4114-4119.
- Creelman, R. A. & Mullet, J. E. (1997). Biosynthesis and Action of Jasmonates in Plants. *Annual Review of Plant Physiology and Plant Molecular Biology*, 48: 355-381.
- Creelman, R. A., Tierney, M. L. & Mullet, J. E. (1992). Jasmonic acid/methyl jasmonate accumulate in wounded soybean hypocotyls and modulate wound gene expression. *Proceedings of the National Academy of Sciences*, 89(11): 4938-4941.
- Cruz de Carvalho, M. H. (2008). Drought stress and reactive oxygen species: production, scavenging and signaling. *Plant Signaling & Behavior*, 3(3): 156-165.
- Csiszár, J., Gallé, Á., Horváth, E., Dancsó, P., Gombos, M., Váry, Z., Erdei, L., Györgyey, J. & Tari, I. (2012). Different peroxidase activities and expression of abiotic stress-related peroxidases in apical root segments of wheat genotypes with different drought stress tolerance under osmotic stress. *Plant Physiology and Biochemistry*, 52: 119-129.
- da Silva, E., Lourenco, E. J. & Neves, V. A. (1990). Soluble and bound peroxidases from papaya fruit. *Phytochemistry*, 29(4): 1051-1056.

- Dai, F., Chen, W. F. & Zhou, B. (2008). Antioxidant synergism of green tea polyphenols with α -tocopherol and l-ascorbic acid in SDS micelles. *Biochimie*, 90(10): 1499-1505.
- Damalas, C. A. (2009). Understanding benefits and risks of pesticide use. *Scientific Research and Essays*, 4(10): 945-949.
- Das, K. & Roychoudhury, A. (2014). Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants, *Frontiers of Environmental Science*, 2(53): 1-13.
- Dave, R. (2009). In vitro models for antioxidant activity evaluation and some medicinal plants possessing antioxidant properties: An overview. *African Journal of Microbiology Research*, 3(13): 981-996.
- Dayan, N. (2008). Prevention and Treatment of Aging Skin with Topical Antioxidants. In N. Dayan (Ed.), *Skin aging handbook: an integrated approach to biochemistry and product development* (pp.149-155). Norwich:William Andrew.
- De Gara, L., de Pinto, M. C. & Tommasi, F. (2003). The antioxidant systems vis-à-vis reactive oxygen species during plant-pathogen interaction. *Plant Physiology and Biochemistry*, 41(10): 863-870.
- De Leonardis, S., Dipierro, N. & Dipierro, S. (2000). Purification and characterization of an ascorbate peroxidase from potato tuber mitochondria. *Plant Physiology and Biochemistry*, 38(10): 773-779.
- De Schepper, V., De Swaef, T., Bauweraerts, I. & Steppe, K. (2013). Phloem transport: a review of mechanisms and controls. *Journal of Experimental Botany*, 64(16): 4839-4850.
- Dedryver, C.-A., Le Ralec, A. & Fabre, F. (2010). The conflicting relationships between aphids and men: a review of aphid damage and control strategies. *Comptes Rendus Biologies*, 333(6): 539-553.
- Déllano-Frier, J. P., Martínez-Gallardo, N. A., Martínez-de La Vega, O., Salas-Araiza, M. D., Barbosa-Jaramillo, E. R., Torres, A., Vargas, P. & Borodanenko, A. (2004). The effect of exogenous jasmonic acid on induced resistance and productivity in amaranth (*Amaranthus hypochondriacus*) is influenced by environmental conditions. *Journal of Chemical Ecology*, 30(5): 1001-1034.
- DellaPenna, D. (2005). A decade of progress in understanding vitamin E synthesis in plants. *Journal of Plant Physiology*, 162(7): 729-737.
- Demole, E., Lederer, E. & Mercier, D. (1962). Isolement et détermination de la structure du jasmonate de méthyle, constituant odorant caractéristique de l'essence de jasmin. *Helvetica Chimica Acta*, 45(2): 675-685.
- Denness, L., McKenna, J. F., Segonzac, C., Wormit, A., Madhou, P., Bennett, M., Mansfield, J. & Hamann, T. (2011). Cell wall damage-induced lignin biosynthesis

is regulated by a reactive oxygen species-and jasmonic acid-dependent process in *Arabidopsis*. *Plant Physiology*, 156(3): 1364-1374.

Department of Agriculture. (2016). Buku Perangkaan Tanaman 2015. Retrieved from <http://www.doa.gov.my/>

Department of Statistic Malaysia. (2016). Supply and Utilization Accounts Selected Agricultural Commodities, Malaysia 2010-2014. Retrieved from <http://www.statistics.gov.my/>

Dermastia, M., Ravnikar, M., Vilhar, B. & Kovač, M. (1994). Increased level of cytokinin ribosides in jasmonic acid-treated potato (*Solanum tuberosum*) stem node cultures. *Physiologia Plantarum*, 92(2): 241-246.

Desikan, R., Last, K., Harrett-Williams, R., Tagliavia, C., Harter, K., Hooley, R., Hancock, J.T. & Neill, S. J. (2006). Ethylene-induced stomatal closure in *Arabidopsis* occurs via AtrbohF-mediated hydrogen peroxide synthesis. *The Plant Journal*, 47(6): 907-916.

Devoto, A. & Turner, J. G. (2005). Jasmonate-regulated *Arabidopsis* stress signalling network. *Physiologia Plantarum*, 123(2): 161-172.

Dhandhukia, P. C. & Thakkar, V. R. (2008). Response surface methodology to optimize the nutritional parameters for enhanced production of jasmonic acid by *Lasiodiplodia theobromae*. *Journal of Applied Microbiology*, 105(3): 636-643.

Dhawan, V. (2014). Reactive Oxygen and Nitrogen Species: General Considerations. In N. K. Ganguly, S. K. Jindal, S. Biswal, P. J. Barnes & R. Pawankar (Eds.) *Studies on Respiratory Disorders* (pp. 27-47). New York: Springer.

Dheepa, R. & Paranjothi, S. (2010). Transmission of Cucumber Mosaic Virus (CMV) infecting banana by aphid and mechanical methods. *Emirates Journal of Food and Agriculture*, 22(2), 117-129.

Diaz, M. & Pavan, C. (1965). Chromosomal anomalies due to virus infection in different crop plants. *Proceedings of the National Academy of Sciences*, 54: 1321-1327.

Díaz, J. & Merino, F. (1998). Wound-induced shikimate dehydrogenase and peroxidase related to lignification in pepper (*Capsicum annuum* L.) leaves. *Journal of Plant Physiology*, 152(1): 51-57.

Díaz, J., Pomar, F., Bernal, Á. & Merino, F. (2004). Peroxidases and the metabolism of capsaicin in *Capsicum annuum* L. *Phytochemistry Reviews*, 3(1-2): 141-157.

Dikilitas, M., Guldur, M. E., Deryaoglu, A. & Erel, O. (2011a). A novel method of measuring oxidative stress of pepper (*Capsicum annuum* var. Charlee) infected with tobacco mosaic virus. *Journal of Applied Biosciences*, 37: 2425-2433.

Dikilitas, M., Guldur, M. E., Deryaoglu, A. & Erel, O. (2011b). Antioxidant and Oxidant Levels of Pepper (*Capsicum annuum* cv. 'Charlee') Infected with Pepper Mild

- Mottle Virus. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 39(2): 58-63.
- Dolatabadian, A. & Jouneghani, R. S. (2009). Impact of exogenous ascorbic acid on antioxidant activity and some physiological traits of common bean subjected to salinity stress. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 37(2): 165-172.
- Dordas, C. (2009). Role of nutrients in controlling plant diseases in sustainable agriculture: A review. In E. Lichtfouse, M. Hamelin, M. Navarrete & P. Debaek (Eds.), *Sustainable agriculture* (pp. 443-460). New York: Springer.
- Drażkiewicz, M. & Baszyński, T. (2005). Growth parameters and photosynthetic pigments in leaf segments of *Zea mays* exposed to cadmium, as related to protection mechanisms. *Journal of Plant Physiology*, 162(9): 1013-1021.
- Duarte, T. L. & Lunec, J. (2005). When is an antioxidant not an antioxidant? A review of novel actions and reactions of vitamin C. *Free Radical Research*, 39(7): 671-686.
- El-Sayed, O. M., El-Gammal, O. H. M. & Salama, A. S. M. (2014). Effect of ascorbic acid, proline and jasmonic acid foliar spraying on fruit set and yield of Manzanillo olive trees under salt stress. *Scientia Horticulturae*, 176: 32-37.
- Elstner, E. F. (1987). Metabolism of activated oxygen species. In: D.D. Davies (Ed.), *The Biochemistry of Plants: A Comprehensive Treatise* (pp. 253-315). San Diego: Academic Press.
- Estrada, B., Bernal, M. A., Díaz, J., Pomar, F. & Merino, F. (2000). Fruit Development in *Capsicum annuum*: Changes in Capsaicin, Lignin, Free Phenolics, and Peroxidase Patterns. *Journal of Agricultural and Food Chemistry*, 48(12): 6234-6239.
- Etim, U. J., Umoren, S. A. & Eduok, U. M. (2016). Coconut coir dust as a low cost adsorbent for the removal of cationic dye from aqueous solution. *Journal of Saudi Chemical Society*, 20: 67-76.
- Evans, M. D., Dizdaroglu, M. & Cooke, M. S. (2004). Oxidative DNA damage and disease: induction, repair and significance. *Mutation Research/Reviews in Mutation Research*, 567(1): 1-61.
- Fain, O. (2005). Musculoskeletal manifestations of scurvy. *Joint Bone Spine*, 72(2): 124-128.
- Farmer, E. & Ryan, C. (1992). Octadecanoid Precursors of Jasmonic Acid Activate the Synthesis of Wound-Inducible Proteinase Inhibitors. *The Plant Cell*, 4(2): 129-134.
- Farris, P. K. (2005). Topical vitamin C: a useful agent for treating photoaging and other dermatologic conditions. *Dermatologic Surgery*, 31: 814-818.

- Fayazi, S., Farokhpyam, M. & Talali, S. (2015). Effect of Capsaicin Cream on Chronic Low Back Pain in Patients With Inter-Vertebral Disc Herniation. *Jundishapur Journal of Chronic Disease Care*, 4(3): 3-7.
- Ferlay, J., Shin, H.-R., Bray, F., Forman, D., Mathers, C. & Parkin, D. M. (2010). Estimates of worldwide burden of cancer in 2008:GLOBOCAN 2008. *International Journal of Cancer*, 127(2): 2893-2917.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin D. M. Forman D. & Bray, F. (2015). Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*, 136(5): 359-386.
- Fingrut, O. & Flescher, E. (2002). Plant stress hormones suppress the proliferation and induce apoptosis in human cancer cells. *Leukemia*, 16(4): 608.
- Fogelman, E., Kaplan, A., Tanami, Z. & Ginzberg, I. (2011). Antioxidative activity associated with chilling injury tolerance of muskmelon (*Cucumis melo* L.) rind. *Scientia Horticulturae*, 128(3): 267-273.
- Foyer, C. H. & Halliwell, B. (1976). The presence of glutathione and glutathione reductase in chloroplasts: a proposed role in ascorbic acid metabolism. *Planta*, 133(1): 21-25.
- Foyer, C. H. & Noctor, G. (2003). Redox sensing and signalling associated with reactive oxygen in chloroplasts, peroxisomes and mitochondria. *Physiologia Plantarum*, 119(3): 355-364.
- Foyer, C. H. & Noctor, G. (2005). Oxidant and antioxidant signalling in plants: a re-evaluation of the concept of oxidative stress in a physiological context. *Plant, Cell & Environment*, 28(8): 1056-1071.
- Foyer, C. H. & Noctor, G. (2011). Ascorbate and glutathione: the heart of the redox hub. *Plant Physiology*, 155(1): 2-18.
- Foyer, C. H. & Shigeoka, S. (2011). Understanding oxidative stress and antioxidant functions to enhance photosynthesis. *Plant Physiology*, 155(1): 93-100.
- Freeman, B. C. & Beattie, G. A. (2008). An overview of plant defenses against pathogens and herbivores. *The Plant Health Instructor*. doi:10.1094/PHI-I-2008-0226-01.
- Freeman, L. E. B., Bonner, M. R., Blair, A., Hoppin, J. A., Sandler, D. P., Lubin, J. H., Dosemeci, M., Lynch, C. F., Knott, C. & Alavanja, M. C. R. (2005). Cancer incidence among male pesticide applicators in the Agricultural Health Study cohort exposed to diazinon. *American Journal of Epidemiology*, 162(11): 1070-1079.
- Fujimoto, T., Tomitaka, Y., Abe, H., Tsuda, S., Futai, K. & Mizukubo, T. (2011). Expression profile of jasmonic acid-induced genes and the induced resistance against the root-knot nematode (*Meloidogyne incognita*) in tomato plants (*Solanum lycopersicum*) after foliar treatment with methyl jasmonate. *Journal of*

Plant Physiology, 168(10): 1084-1097.

- Galanihe, L. D., Priyantha, M., Yapa, D. R., Bandara, H. M. S. & Ranasinghe, J. (2004). Insect pest and disease incidences of exotic hybrids chilli pepper varieties grown in the low country dry zone of Sri Lanka. *Annals of Sri Lanka*, 6: 99-106.
- Gallie, D. R. (2012). The role of L-ascorbic acid recycling in responding to environmental stress and in promoting plant growth. *Journal of Experimental Botany*, 64(2): 433-443.
- Gallitelli, D. (2000). The ecology of Cucumber mosaic virus and sustainable agriculture. *Virus Research*, 71(1): 9-21.
- Gao, Y., Lei, Z. & Reitz, S. R. (2012). Western flower thrips resistance to insecticides: detection, mechanisms and management strategies. *Pest Management Science*, 68(8): 1111-1121.
- Gill, S. S. & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48(12): 909-930.
- Goel, A. K., Lundberg, D., Torres, M. A., Matthews, R., Akimoto-Tomiya, C., Farmer, L., Dang, J. L. & Grant, S. R. (2008). The *Pseudomonas syringae* type III effector HopAM1 enhances virulence on water-stressed plants. *Molecular Plant-Microbe Interactions*, 21(3): 361-370.
- Gols, R., Roosjen, M., Dijkman, H. & Dicke, M. (2003). Induction of direct and indirect plant responses by jasmonic acid, low spider mite densities, or a combination of jasmonic acid treatment and spider mite infestation. *Journal of Chemical Ecology*, 29(12): 2651-2666.
- Gomez, J. M., Jimenez, A., Olmos, E. & Sevilla, F. (2004). Location and effects of long-term NaCl stress on superoxide dismutase and ascorbate peroxidase isoenzymes of pea (*Pisum sativum* cv. Puget) chloroplasts. *Journal of Experimental Botany*, 55(394): 119-130.
- González-Pérez, J. L., Espino-Gudiño, M. C., Torres-pacheco, I., Guevara-González, R. G., Herrera-Ruiz, G. & Rodríguez-Hernández, V. (2011). Quantification of virus syndrome in chili peppers. *African Journal of Biotechnology*, 10(27): 5236-5250.
- Goverse, A. & Bird, D. (2011). The role of plant hormones in nematode feeding cell formation. In J. Jones, G. Gheysen, and C. Fenoll (Eds.), *Genomics and Molecular Genetics of Plant-Nematode Interactions* (pp. 325-347). Netherland: Springer.
- Grabber, J. H. (2005). How do lignin composition, structure, and cross-linking affect degradability? A review of cell wall model studies. *Crop Science*, 45(3): 820-831.
- Grant, M. & Lamb, C. (2006). Systemic immunity. *Current Opinion in Plant Biology*, 9(4): 414-420.

- Greer, L. & Dole, J. M. (2003). Aluminum foil, aluminium-painted, plastic, and degradable mulches increase yields and decrease insectvectored viral diseases of vegetables. *HortTechnology*, 13(2): 276-284.
- Grinberg, M., Perl-Treves, R., Palevsky, E., Shomer, I. & Soroker, V. (2005). Interaction between cucumber plants and the broad mite, *Polyphagotarsonemus latus*: from damage to defense gene expression. *Entomologia Experimentalis et Applicata*, 115(1): 135-144.
- Grube, R. C., Zhang, Y. & Graduate, F. (2000). New Source of Resistance to Cucumber mosaic virus in *Capsicum frutescens*. *Plant Disease*, 84(8): 885-891.
- Güldür, M. E. & Çağlar, B. K. (2006). Outbreaks of Pepper mild mottle virus in greenhouses in Sanliurfa, Turkey. *Journal of Plant Pathology*, 88(3): 339-342.
- Gumiero, A., Murphy, E. J., Metcalfe, C. L., Moody, P. C. E. & Raven, E. L. (2010). An analysis of substrate binding interactions in the heme peroxidase enzymes: a structural perspective. *Archives of Biochemistry and Biophysics*, 500(1): 13-20.
- Guo, L., Wang, P., Gu, Z., Jin, X. & Yang, R. (2017). Proteomic analysis of broccoli sprouts by iTRAQ in response to jasmonic acid. *Journal of Plant Physiology*, 218: 16-25.
- Guo, Y. & Gan, S. (2012). Convergence and divergence in gene expression profiles induced by leaf senescence and 27 senescence-promoting hormonal, pathological and environmental stress treatments. *Plant, Cell & Environment*, 35(3): 644-655.
- Ha, S.-H., Kim, J.-B., Park, J.-S., Lee, S.-W. & Cho, K.-J. (2007). A comparison of the carotenoid accumulation in *Capsicum* varieties that show different ripening colours: deletion of the capsanthin-capsorubin synthase gene is not a prerequisite for the formation of a yellow pepper. *Journal of Experimental Botany*, 58(12): 3135-3144.
- Halim, V. A., Hunger, A., Macioszek, V., Landgraf, P., Nürnberger, T., Scheel, D. & Rosahl, S. (2004). The oligopeptide elicitor Pep-13 induces salicylic acid-dependent and-independent defense reactions in potato. *Physiological and Molecular Plant Pathology*, 64(6): 311-318.
- Halimatunsadiah, A. B., Norida, M., Omar, D. & Kamarulzaman, N. H. (2016). Application of pesticide in pest management: The case of lowland vegetable growers. *International Food Research Journal*, 23(1):85-94.
- Han, C., Liu, Q. & Yang, Y. (2009). Short-term effects of experimental warming and enhanced ultraviolet-B radiation on photosynthesis and antioxidant defense of *Picea asperata* seedlings. *Plant Growth Regulation*, 58(2): 153-162.
- Han, R., Tian, Y., Wu, Y., Wang, P., Ai, X., Zhang, J. & Skibsted, L. H. (2006). Mechanism of radical cation formation from the excited states of zeaxanthin and astaxanthin in chloroform. *Photochemistry and Photobiology*, 82(2): 538-546.

- Harms, K., Atzorn, R., Brash, A., Kuhn, H., Wasternack, C., Willmitzer, L. & Pena-Cortes, H. (1995). Expression of a flax allene oxide synthase cDNA leads to increased endogenous jasmonic acid (JA) levels in transgenic potato plants but not to a corresponding activation of JA-responding genes. *The Plant Cell*, 7(10): 1645-1654.
- Hasanuzzaman, M., Nahar, K. & Fujita, M. (2014). Role of tocopherol (vitamin E) in plants: Abiotic stress tolerance and beyond. In P. Ahmad & S. Rasool (Eds.), *Emerging Technologies and Management of Crop Stress Tolerance* (pp. 267-289). San Diego: Academic Press.
- Hase, S., Takahashi, S., Takenaka, S., Nakaho, K., Arie, T., Seo, S., Ohashi, Y. & Takahashi, H. (2008). Involvement of jasmonic acid signalling in bacterial wilt disease resistance induced by biocontrol agent *Pythium oligandrum* in tomato. *Plant Pathology*, 57(5): 870-876.
- Haskell, M. J. (2013). Provitamin A carotenoids as a dietary source of vitamin A. In S. A. Tanumihardjo (Ed.), *Carotenoids and human health* (pp. 249-260). Madison: Humana Press.
- He, Y., Fukushige, H., Hildebrand, D. F. & Gan, S. (2002). Evidence supporting a role of jasmonic acid in Arabidopsis leaf senescence. *Plant Physiology*, 128(3): 876-884.
- Heinrich, M., Hettenhausen, C., Lange, T., Wünsche, H., Fang, J., Baldwin, I. T. & Wu, J. (2013). High levels of jasmonic acid antagonize the biosynthesis of gibberellins and inhibit the growth of *Nicotiana attenuata* stems. *Plant Journal*, 73(4): 591-606.
- Hernández, J. A., Rubio, M., Olmos, E., Ros-Barceló, A. & Martínez-Gómez, P. (2004). Oxidative stress induced by long-term plum pox virus infection in peach (*Prunus persica*). *Physiologia Plantarum*, 122(4): 486-495.
- Hidayat, S. H., Opriana, E., Manzila, I. & Sujiprihati, S. (2012). Occurrence of chilli veinal mottle virus (ChiVMV) in Indonesia and response of chilli germplasm to ChiVMV infection. *The Journal of International Society for Southeast Asian Agricultural Sciences*, 18(2): 55-61.
- Hodges, D. M., Andrews, C. J., Johnson, D. A. & Hamilton, R. I. (1996). Antioxidant compound responses to chilling stress in differentially sensitive inbred maize lines. *Physiologia Plantarum*, 98(4): 685-692.
- Hodgson, E. W., Rice, M. E. & O'Neal, M. (2011). Management Recommendations for Soybean Aphid (Hemiptera: Aphididae) in the United States. *Journal of Integrated Pest Management* 3(2012): 1-10.
- Hofius, D. & Sonnewald, U. (2003). Vitamin E biosynthesis: biochemistry meets cell biology. *Trends in Plant Science*, 8(1): 6-8.

- Hogenhout, S. A., Ammar, E.-D., Whitfield, A. E. & Redinbaugh, M. G. (2008). Insect vector interactions with persistently transmitted viruses. *Annual Review of Phytopathology*, 46: 327-359.
- Hojo, E. T. D., Durigan, J. F., Hojo, R. H., Donadon, J. R. & Martins, R. N. (2011). Uso de tratamento hidrotérmico e ácido clorídrico na qualidade de lichia 'Bengal'. *Revista Brasileira de Fruticultura*, 33(2): 386-393.
- Holmes, F. O. (1929). Local lesions in tobacco mosaic. *Botanical Gazette*, 87(1): 39-55.
- Hou, L., Lee, W. J., Rusiecki, J. A., Hoppin, J. A., Blair, M. R., Lubin, I., Samanic, C., Sandler, D. P., Blair, A. & Alavanja, M. (2006). Cancer incidence, pendimethatin exposure among pesticide applicators: A report from the Agricultural Health Study. *Epidemiology*, 17(3): 302-307.
- Howe, G. A. & Schillmiller, A. L. (2002). Oxylin metabolism in response to stress. *Current Opinion in Plant Biology*, 5(3): 230-236.
- Howitt, C. A. & Pogson, B. J. (2006). Carotenoid accumulation and function in seeds and non-green tissues. *Plant, Cell & Environment*, 29(3): 435-445.
- Hu, X., Wansha, L., Chen, Q. & Yang, Y. (2009). Early signals transduction linking the synthesis of jasmonic acid in plant. *Plant Signaling & Behavior*, 4(8): 696-697.
- Huang, R., Xia, R., Hu, L., Lu, Y. & Wang, M. (2007). Antioxidant activity and oxygen-scavenging system in orange pulp during fruit ripening and maturation. *Scientia Horticulturae*, 113(2): 166-172.
- Huang, S.-P., Chen, J.-C., Wu, C.-C., Chen, C.-T., Tang, N.-Y., Ho, Y.-T., Lo, C., Lin, J.-P., Chung, J.-G. & Lin, J.-G. (2009). Capsaicin-induced apoptosis in human hepatoma HepG2 cells. *Anticancer Research*, 29(1): 165-174.
- Hultén, K., Van Kappel, A. L., Winkvist, A., Kaaks, R., Hallmans, G., Lenner, P. & Riboli, E. (2001). Carotenoids, alpha-tocopherols, and retinol in plasma and breast cancer risk in northern Sweden. *Cancer Causes & Control*, 12(6): 529-537.
- Hussain, F. & Abid, M. (2011). Pest and diseases of chilli crop in Pakistan: A review. *International Journal of Biology and Biotechnology*, 8(2): 325-332.
- Iloki-Assanga, S. B., Lewis-Luján, L. M., Lara-Espinoza, C. L., Gil-Salido, A. a., Fernandez-Angulo, D., Rubio-Pino, J. L. & Haines, D. D. (2015). Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras* L. and *Phoradendron californicum*. *BMC Research Notes*, 8(1): 396-409.
- Imlay, J. A. (2003). Pathways of oxidative damage. *Annual Reviews in Microbiology*, 57(1): 395-418.
- Iqbal, J., Abbasi, B. A., Mahmood, T., Kanwal, S., Ali, B. & Khalil, A. T. (2017). Plant-derived anticancer agents: A green anticancer approach. *Asian Pacific Journal of Tropical Biomedicine*, 7(12): 1129-1150.

- Iqbal, S., Ashfaq, M. & Shah, H. (2012). Prevalence and Distribution of Cucumber mosaic virus (CMV) in major Chilli Growing Areas of Pakistan. *Pakistan Journal of Botany*, 44(5): 1749–1754.
- Ishikawa, T., & Shigeoka, S. (2008). Recent advances in ascorbate biosynthesis and the physiological significance of ascorbate peroxidase in photosynthesizing organisms. *Bioscience, Biotechnology, and Biochemistry*, 72(5): 1143-1154.
- Jacob, R. A. & Sotoudeh, G. (2002). Vitamin C function and status in chronic disease. *Nutrition in Clinical Care*, 5(2): 66-74.
- Jagota, S. K. & Dani, H. M. (1982). A new colorimetric technique for the estimation of vitamin C using Folin phenol reagent. *Analytical Biochemistry*, 127(1): 178-182.
- Jayasinghe, C., Gotoh, N. & Wada, S. (2013). Pro-oxidant/antioxidant behaviours of ascorbic acid, tocopherol, and plant extracts in n-3 highly unsaturated fatty acid rich oil-in-water emulsions. *Food Chemistry*, 141(3): 3077-3084.
- Jeyaseeli, M. D. & Raj, S. P. (2010). Chemical characteristics of coir pith as a function of its particle size to be used as soilless medium. *The Ecoscan*, 4(2-3): 163-169.
- Johnson, K. D., O’Neal, M. E., Bradshaw, J. D. & Rice, M. E. (2008). Is preventative, concurrent management of the soybean aphid (Hemiptera: Aphididae) and bean leaf beetle (Coleoptera: Chrysomelidae) possible?. *Journal of Economic Entomology*, 101(3): 801-809.
- Jones, R. A. C. (2000). Determining ‘threshold’ levels for seed-borne virus infection in seed stocks. *Virus Research*, 71(1-2): 171-183.
- Jones, R. A. C., Coutts, B. A., Latham, L. J. & McKirdy, S. J. (2008). Cucumber mosaic virus infection of chickpea stands: temporal and spatial patterns of spread and yield-limiting potential. *Plant Pathology*, 57(5): 842-853.
- Kachaiyaphum, P., Howteerakul, N., Sujirarat, D., Siri, S. & Suwannapong, N. (2010). Serum cholinesterase levels of Thai chilli-farm workers exposed to chemical pesticides: prevalence estimates and associated factors. *Journal of Occupational Health*, 52(1): 89-98.
- Kamffer, Z., Bindon, K. A. & Oberholster, A. (2010). Optimization of a method for the extraction and quantification of carotenoids and chlorophylls during ripening in grape berries (*Vitis vinifera* cv. Merlot). *Journal of Agricultural and Food Chemistry*, 58(11): 6578-6586.
- Kanno, C. & Yamauchi, K. (1977). Application of a new iron reagent, 3-(2-pyridyl)-5, 6-diphenyl-1, 2, 4-triazine, to spectrophotometric determination of tocopherols. *Agricultural and Biological Chemistry*, 41(3): 593-596.
- Kanwischer, M., Porfirova, S., Bergmüller, E. & Dörmann, P. (2005). Alterations in tocopherol cyclase activity in transgenic and mutant plants of Arabidopsis affect tocopherol content, tocopherol composition, and oxidative stress. *Plant Physiology*, 137(2): 713-723.

- Karagounis, C., Kourdoumbalos, A. K., Margaritopoulos, J. T., Nanos, G. D. & Tsitsipis, J. A. (2006). Organic farming-compatible insecticides against the aphid *Myzus persicae* (Sulzer) in peach orchards. *Journal of Applied Entomology*, 130(3): 150-154.
- Karuppanapandian, T., Moon, J.-C., Kim, C., Manoharan, K. & Kim, W. (2011). Reactive oxygen species in plants: their generation, signal transduction, and scavenging mechanisms. *Australian Journal of Crop Science*, 5(6): 709-725.
- Kaur, S. & Singh, S. (2013). Efficacy of some insecticides and botanicals against sucking pests on capsicum under net house. *Agriculture for Sustainable Development*, 1(1): 25-29.
- Kaya, A. & Doganlar, Z. B. (2016). Exogenous jasmonic acid induces stress tolerance in tobacco (*Nicotiana tabacum*) exposed to imazapic. *Ecotoxicology and Environmental Safety*, 124: 470-479.
- Khan, S. M., Raj, S. K., Bano, T. & Garg, V. K. (2006). Incidence and management of mosaic and leaf curl diseases in cultivars of chilli (*Capsicum annuum*). *Journal of Food Agriculture and Environment*, 4(1): 171-174.
- Khoo, H.-E., Prasad, K. N., Kong, K.-W., Jiang, Y. & Ismail, A. (2011). Carotenoids and their isomers: color pigments in fruits and vegetables. *Molecules*, 16(2): 1710-1738.
- Kiffin, R., Bandyopadhyay, U. & Cuervo, A. M. (2006). Oxidative stress and autophagy. *Antioxidants & Redox Signaling*, 8(1-2): 152-162.
- Kim, J., Ahn, J., Lee, S., Moon, B., Ha, T. & Kim, S. (2011). Phytochemicals and antioxidant activity of fruits and leaves of paprika (*Capsicum annuum* L. var. Special) cultivated in Korea. *Journal of Food Science*, 76(2): 193-198.
- Kiokias, S. & Gordon, M. H. (2004). Antioxidant properties of carotenoids in vitro and in vivo. *Food Reviews International*, 20(2): 99-121.
- Király, L., Barna, B. & Király, Z. (2007). Plant resistance to pathogen infection: forms and mechanisms of innate and acquired resistance. *Journal of Phytopathology*, 155(7-8): 385-396.
- Király, L., Hafez, Y. M., Fodor, J. & Király, Z. (2008). Suppression of tobacco mosaic virus-induced hypersensitive-type necrotization in tobacco at high temperature is associated with downregulation of NADPH oxidase and superoxide and stimulation of dehydroascorbate reductase. *Journal of General Virology*, 89(3): 799-808.
- Koleva-Gudeva, L., Mitrev, S., Maksimova, V., & Spasov, D. (2013). Content of capsaicin extracted from hot pepper (*Capsicum annuum* ssp. *microcarpum* L.) and its use as an ecopesticide. *Hemijaska industrija*, 67(4): 671-675.

- Kopsell, D. A. & Kopsell, D. E. (2010). Carotenoids in vegetables: biosynthesis, occurrence, impacts on human health, and potential for manipulation. In: Ross, R. & Preedy, V. R. (Eds.), *Bioactive Foods in Promoting Health: Fruits and Vegetables* (pp. 645-662). St. Louis: Elsevier.
- Kramell, R., Miersch, O., Atzorn, R., Parthier, B. & Wasternack, C. (2000). Octadecanoid-derived alteration of gene expression and the "oxylipin signature" in stressed barley leaves. Implications for different signaling pathways. *Plant Physiology*, 123(1): 177-188.
- Krishnamurthi, K. (2007). 17-screening of natural products for anticancer and antidiabetic properties. *Cancer*, 3(1-2): 69-75.
- Kumar, S., Kumar, S., Singh, M., Singh, A. K. & Rai, M. (2006). Identification of host plant resistance to pepper leaf curl virus in chilli (*Capsicum* species). *Scientia Horticulturae*, 110(4): 359-361.
- Kumar, S., Chauhan, P. S., Agrawal, L., Raj, R., Srivastava, A., Gupta, S., Mishra, S. K., Yadav, S., Singh, P. C., Raj, S. K. & Nautiyal, C. S. (2016). *Paenibacillus lentimorbus* inoculation enhances tobacco growth and extenuates the virulence of Cucumber mosaic virus. *PLoS One*, 11(3): e0149980.
- Kunkaliker, S. R., Poojari, S., Arun, B. M., Rajagopalan, P. A., Chen, T.-C., Yeh, S.-D., Naidu R.A., Zehr U.B. & Ravi, K. S. (2011). Importance and genetic diversity of vegetable-infecting tospoviruses in India. *Phytopathology*, 101(3): 367-376.
- Kurutas, E. B. (2015). The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: current state. *Nutrition Journal*, 15(1): 71.
- Laloi, C., Apel, K. & Danon, A. (2004). Reactive oxygen signalling: the latest news. *Current Opinion in Plant Biology*, 7(3): 323-328.
- Lang, Y.-Q., Yanagawa, S., Sasanuma, T. & Sasakuma, T. (2004). Orange fruit color in *Capsicum* due to deletion of capsanthin-capsorubin synthesis gene. *Breeding Science*, 54(1): 33-39.
- Lantos, C., Juhász, A. G., Vági, P., Mihály, R., Kristóf, Z. & Pauk, J. (2012). Androgenesis induction in microspore culture of sweet pepper (*Capsicum annuum* L.). *Plant Biotechnology Reports*, 6(2): 123-132.
- Lapidot, M., Ben-Joseph, R., Cohen, L., Machbash, Z. & Levy, D. (2006). Development of a scale for evaluation of Tomato yellow leaf curl virus resistance level in tomato plants. *Phytopathology*, 96(12): 1404-1408.
- Ledford, H. K. & Niyogi, K. K. (2005). Singlet oxygen and photo-oxidative stress management in plants and algae. *Plant, Cell & Environment*, 28(8): 1037-1045.
- Lee, C. C. & Houghton, P. (2005). Cytotoxicity of plants from Malaysia and Thailand used traditionally to treat cancer. *Journal of Ethnopharmacology*, 100(3): 237-243.

- Lee, M.-W., Qi, M. & Yang, Y. (2001). A novel jasmonic acid-inducible rice myb gene associates with fungal infection and host cell death. *Molecular Plant-Microbe Interactions*, 14(4): 527-535.
- Lewsey, M., Surette, M., Robertson, F. C., Ziebell, H., Choi, S. H., Ryu, K. H., Canto, T., Palukaitis, P., Payne, T., Walsh, J.A. & Carr, J. P. (2009). The role of the Cucumber mosaic virus 2b protein in viral movement and symptom induction. *Molecular Plant-Microbe Interactions*, 22(6): 642-654.
- Li, G., Wan, S., Zhou, J., Yang, Z. & Qin, P. (2010). Leaf chlorophyll fluorescence, hyperspectral reflectance, pigments content, malondialdehyde and proline accumulation responses of castor bean (*Ricinus communis* L.) seedlings to salt stress levels. *Industrial Crops and Products*, 31(1): 13-19.
- Li, C., Liu, G., Xu, C., Lee, G. I., Bauer, P., Ling, H. Q., Ganai, M. W. & Howe, G. A. (2003). The tomato suppressor of prosystemin-mediated responses2 gene encodes a fatty acid desaturase required for the biosynthesis of jasmonic acid and the production of a systemic wound signal for defense gene expression. *The Plant Cell*, 15(7): 1646-1661.
- Lichtenthaler, H. K. (1987). Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. *Methods in Enzymology*, 148: 350-382.
- Lichtenthaler, H. K., & Buschmann, C. (2001). Chlorophylls and carotenoids: Measurement and characterization by UV-VIS spectroscopy. *Current Protocols in Food Analytical Chemistry*.
- Lichtenthaler, H. K., Buschmann, C., Döll, M., Fietz, H.-J., Bach, T., Kozel, U., Meier D. & Rahmsdorf, U. (1981). Photosynthetic activity, chloroplast ultrastructure, and leaf characteristics of high-light and low-light plants and of sun and shade leaves. *Photosynthesis Research*, 2(2): 115-141.
- Liebler, D. C. (1993). Antioxidant reactions of carotenoids. *Annals of the New York Academy of Sciences*, 691(1): 20-31.
- Linser, P. J. & Dinglasan, R. R. (2014). Insect gut structure, function, development and target of biological toxins. *Advances in Insect Physiology*, 47: 1-37.
- Lisenbee, C. S., Heinze, M. & Trelease, R. N. (2003). Peroxisomal ascorbate peroxidase resides within a subdomain of rough endoplasmic reticulum in wild-type Arabidopsis cells. *Plant Physiology*, 132(2): 870-882.
- Liu, L., Wei, J., Zhang, M., Zhang, L., Li, C., & Wang, Q. (2012). Ethylene independent induction of lycopene biosynthesis in tomato fruits by jasmonates. *Journal of Experimental Botany*, 63(16): 5751-5761.
- Liu, X., Chi, H., Yue, M., Zhang, X., Li, W. & Jia, E. (2012). The regulation of exogenous jasmonic acid on UV-B stress tolerance in wheat. *Journal of Plant Growth Regulation*, 31(3): 436-447.

- López-Martínez, N., Colinas-León, M. T., Peña-Valdivia, C. B., Salinas-Moreno, Y., Fuentes-Montiel, P., Biesaga, M. & Zavaleta-Mejía, E. (2011). Alterations in peroxidase activity and phenylpropanoid metabolism induced by *Nacobbus aberrans* Thorne and Allen, 1944 in chilli (*Capsicum annuum* L.) CM334 resistant to *Phytophthora capsici* Leo. *Plant and Soil*, 338(1-2): 399–409.
- Lou, Y.-G., Du, M.-H., Turlings, T. C. J., Cheng, J.-A. & Shan, W.-F. (2005). Exogenous application of jasmonic acid induces volatile emissions in rice and enhances parasitism of *Nilaparvata lugens* eggs by the Parasitoid *Anagrus nilaparvatae*. *Journal of Chemical Ecology*, 31(9): 1985-2002.
- Lowe, G., Woodward, M., Rumley, A., Morrison, C., Tunstall-Pedoe, H. & Stephen, K. (2003). Total tooth loss and prevalent cardiovascular disease in men and women: possible roles of citrus fruit consumption, vitamin C, and inflammatory and thrombotic variables. *Journal of Clinical Epidemiology*, 56(7): 694-700.
- Luo, Y., Shang, J., Zhao, P., Xi, D., Yuan, S. & Lin, H. (2011). Application of Jasmonic Acid Followed by Salicylic Acid Inhibits Cucumber mosaic virus Replication. *The Plant Pathology Journal*, 27(1): 53-58.
- Maeda, H., Song, W., Sage, T. L. & DellaPenna, D. (2006). Tocopherols play a crucial role in low-temperature adaptation and phloem loading in Arabidopsis. *The Plant Cell Online*, 18(10): 2710-2732.
- Mahan, J. R. & Wanjura, D. F. (2005). Seasonal patterns of glutathione and ascorbate metabolism in field-grown cotton under water stress. *Crop Science*, 45(1): 19-201.
- Majumder, T., Sakina, P., Suresh, P., Ashok, K., Nagulmeera, S. & Rajesh, K. (2013). Estimation Of Total Phenolic Content And Evaluation Of In-Vitroantioxidant Activity *Capsicum annuum* Linn. Leaves. *International Research Journal of Pharmaceutical and Applied Sciences*, 3(2):180-185.
- Maksymiv, I. V. (2015). Pesticides: Benefits and Hazards. *Journal of Vasyl Stefanyk Precarpathian National University*, 2(1): 71-77.
- Mallick, N. & Mohn, F. H. (2000). Reactive oxygen species: response of algal cells. *Journal of Plant Physiology*, 157(2): 183-193.
- Manas, D., Bandopadhyay, P. K., Chakravarty, A., Pal, S. & Bhattacharya, A. (2014). Influence of foliar applications of chelator and micronutrients on antioxidants in green chilli. *International Journal of Nutrition and Metabolism*, 6(2): 18-27.
- Mani, M., Shivaraju, C. & Kulkarni, N. S. (2014). Pests. In M. Mani, C. Shivaraju & N. S. Kulkarni (Eds.), *The grape entomology* (pp. 9-166). New Delhi: Springer.
- Mariyono, J. & Bhattarai, M. (2009). Chili production practices in Central Java, Indonesia: A Baseline Report. *AVRDC-The World Vegetable Center, Taiwan*.
- Martí, M. C., Camejo, D., Vallejo, F., Romojaro, F., Bacarizo, S., Palma, J. M., Sevilla, F. & Jiménez, A. (2011). Influence of fruit ripening stage and harvest period on the antioxidant content of sweet pepper cultivars. *Plant Foods for Human*

Nutrition, 66(4): 416-423.

- Masia, A. (1998). Superoxide dismutase and catalase activities in apple fruit during ripening and post-harvest and with special reference to ethylene. *Physiologia Plantarum*, 104(4): 668-672.
- Mateos, R. M., Jiménez, A., Román, P., Romojaro, F., Bacarizo, S., Corpas, F. J. & Palma, J. M. (2013). Antioxidant Systems from Pepper (*Capsicum annuum* L.): Involvement in the Response to Temperature Changes in Ripe Fruits. *International Journal of Molecular Sciences*, 14(5): 9556-9580.
- Mateos, R. M., León, A. M., Sandalio, L. M., Gómez, M., Luis, A. & Palma, J. M. (2003). Peroxisomes from pepper fruits (*Capsicum annuum* L.): purification, characterisation and antioxidant activity. *Journal of Plant Physiology*, 160(12): 1507-1516.
- Mathé, C., Barre, A., Jourda, C. & Dunand, C. (2010). Evolution and expression of class III peroxidases. *Archives of Biochemistry and Biophysics*, 500(1): 58-65.
- Mauch-Mani, B. & Mauch, F. (2005). The role of abscisic acid in plant-pathogen interactions. *Current Opinion in Plant Biology*, 8(4): 409-414.
- Maucher, H., Hause, B., Feussner, I., Ziegler, J. & Wasternack, C. (2000). Allene oxide synthases of barley (*Hordeum vulgare* cv. *Salome*): tissue specific regulation in seedling development. *The Plant Journal*, 21(2): 199-213.
- Mazidah, M., Yusoff, K., Habibuddin, H., Tan, Y. H. & Lau, W. H. (2012). Characterization of cucumber mosaic virus (CMV) causing mosaic symptom on (*Catharanthus roseus* L.) G. Don in Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 35(1): 41-53.
- McDougall, S., Watson, A., Stodart, B., Kelly, G., Troidahl, D. & Tesoriero, L. (2013). *Tomato, capsicum, chilli and eggplant: a field guide for the identification of insect pests, beneficials, diseases and disorders in Australia and Cambodia*. Canberra: Australian Centre for International Agricultural Research (ACIAR).
- Meena, R. S., Ameta, O. P. & Meena, B. L. (2013). Population dynamics of sucking pests and their correlation with weather parameters in chilli, (*Capsicum annuum* L.) crop. *The Bioscan*, 8(1): 177-180.
- Mehlhorn, H., Lelandais, M., Korth, H. G. & Foyer, C. H. (1996). Ascorbate is the natural substrate for plant peroxidases. *FEBS Letters*, 378(3): 203-206.
- Mehta, S. K. & Gowder, S. J. T. (2015). Members of Antioxidant Machinery and Their Functions. In S. Joghi & T. Gowder (Eds.), *Basic Principles and Clinical Significance of Oxidative Stress* (pp. 59-77). Croatia: InTech.
- Melanie, L. L. I. & Sally, A. M. (2004). Anthracnose Fruit Rot of Pepper. Extension Fact sheet, Ohio State University. Retrieved from <http://ohioline.osu.edu>

- Memelink, J. (2009). Regulation of gene expression by jasmonate hormones. *Phytochemistry*, 70(13): 1560-1570.
- Menzel, T. R., Weldegergis, B. T., David, A., Boland, W., Gols, R., van Loon, J. J. A. & Dicke, M. (2014). Synergism in the effect of prior jasmonic acid application on herbivore-induced volatile emission by Lima bean plants: transcription of a monoterpene synthase gene and volatile emission. *Journal of Experimental Botany*, 65(17): 4821-4831.
- Mesnager, R., Defarge, N., Spiroux De Vendômois, J. & Séralini, G. E. (2014). Major pesticides are more toxic to human cells than their declared active principles. *BioMed Research International*, 43:122-125.
- Mhamdi, A., Noctor, G. & Baker, A. (2012). Plant catalases: peroxisomal redox guardians. *Archives of Biochemistry and Biophysics*, 525(2): 181-194.
- Miersch, O., Weichert, H., Stenzel, I., Hause, B., Maucher, H., Feussner, I. & Wasternack, C. (2004). Constitutive overexpression of allene oxide cyclase in tomato (*Lycopersicon esculentum* cv. *Lukullus*) elevates levels of some jasmonates and octadecanoids in flower organs but not in leaves. *Phytochemistry*, 65(7): 847-856.
- Milrot, E., Jackman, A., Flescher, E., Gonen, P., Kelson, I., Keisari, Y. & Sherman, L. (2013). Enhanced killing of cervical cancer cells by combinations of methyl jasmonate with cisplatin, X or alpha radiation. *Investigational New Drugs*, 31(2): 333-344.
- Mishra, S., Jha, A. B. & Dubey, R. S. (2011). Arsenite treatment induces oxidative stress, upregulates antioxidant system, and causes phytochelatin synthesis in rice seedlings. *Protoplasma*, 248(3): 565-577.
- Moloi, M. J., Mwenye, O. J. & van der Merwe, R. (2016). Differential involvement of ascorbate and guaiacol peroxidases in soybean drought resistance. *South African Journal of Science*, 112(9-10): 1-4.
- Mondal, K., Malhotra, S. P., Jain, V. & Singh, R. (2009). Oxidative stress and antioxidant systems in Guava (*Psidium guajava* L.) fruits during ripening. *Physiology and Molecular Biology of Plants*, 15(4): 327-334.
- Mondal, K., Sharma, N. S., Malhotra, S. P., Dhawan, K. & Singh, R. (2004). Antioxidant systems in ripening tomato fruits. *Biologia Plantarum*, 48(1): 49-53.
- Moore, J. P., Paul, N. D., Whittaker, J. B. & Taylor, J. E. (2003). Exogenous jasmonic acid mimics herbivore-induced systemic increase in cell wall bound peroxidase activity and reduction in leaf expansion. *Functional Ecology*, 17(4): 549-554.
- Mordecai, E. A. (2011). Pathogen impacts on plant communities: unifying theory, concepts, and empirical work. *Ecological Monographs*, 81(3): 429-441.
- Moreira, X., Sampedro, L. & Zas, R. (2009). Defensive responses of *Pinus pinaster* seedlings to exogenous application of methyl jasmonate: concentration effect and

- systemic response. *Environmental and Experimental Botany*, 67(1): 94-100.
- Mortensen, A. & Skibsted, L. H. (1997). Real time detection of reactions between radicals of lycopene and tocopherol homologues. *Free Radical Research*, 27(2): 229-234.
- Mortensen, A., Skibsted, L. H., Willnow, A. & Everett, S. A. (1998). Re-appraisal of the tocopheroxyl radical reaction with β -carotene: evidence for oxidation of vitamin E by the β -carotene radical cation. *Free Radical Research*, 28(1): 69-80.
- Moulding, P. H., Grant, H. F., McLellan, K. M. & Robinson, D. S. (1987). Heat stability of soluble and ionically bound peroxidases extracted from apples. *International Journal of Food Science & Technology*, 22(4): 391-397.
- Mukai, K., Tokunaga, A., Itoh, S., Kanesaki, Y., Ohara, K., Nagaoka, S. & Abe, K. (2007). Structure Activity Relationship of the Free-Radical-Scavenging Reaction by Vitamin E (α -, β -, γ -, δ -Tocopherols) and Ubiquinol-10: pH Dependence of the Reaction Rates. *The Journal of Physical Chemistry B*, 111(3): 652-662.
- Munne-Bosch, S. (2005). The role of α -tocopherol in plant stress tolerance. *Journal of Plant Physiology*, 162(7): 743-748.
- Munné-Bosch, S. & Alegre, L. (2002). The function of tocopherols and tocotrienols in plants. *Critical Reviews in Plant Sciences*, 21(1): 31-57.
- Munné-Bosch, S. & Falk, J. (2004). New insights into the function of tocopherols in plants. *Planta*, 218(3): 323-326.
- Munné-Bosch, S., Weiler, E. W., Alegre, L., Müller, M., Düchting, P. & Falk, J. (2007). α -Tocopherol may influence cellular signaling by modulating jasmonic acid levels in plants. *Planta*, 225(3): 681-691.
- Munné-Bosch, S. (2005). Linking tocopherols with cellular signaling in plants. *New Phytologist*, 166(2): 363-366.
- Munné-Bosch, S. & Penuelas, J. (2003). Photo-and antioxidative protection during summer leaf senescence in *Pistacia lentiscus* L. grown under mediterranean field conditions. *Annals of Botany*, 92(3): 385-391.
- Muñoz-Espinoza, V. A., López-Climent, M. F., Casaretto, J. A. & Gómez-Cadenas, A. (2015). Water stress responses of tomato mutants impaired in hormone biosynthesis reveal abscisic acid, jasmonic acid and salicylic acid interactions. *Frontiers in Plant Science*, 6(997):1-14.
- Murphy, J. F., Reddy, M. S., Ryu, C.-M., Kloepper, J. W. & Li, R. (2003). Rhizobacteria-mediated growth promotion of tomato leads to protection against Cucumber mosaic virus. *Phytopathology*, 93(10): 1301-1307.
- Murray, R. R., Emblow, M. S. M., Hetherington, A. M. & Foster, G. D. (2016). Plant virus infections control stomatal development. *Scientific Reports*, 6 (34507): 1-7.

- Nakano, Y. & Asada, K. (1981). Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts. *Plant and Cell Physiology*, 22(5): 867-880.
- Namitha, K. & Negi, P. S. (2010). Chemistry and biotechnology of carotenoids. *Critical Reviews in Food Science and Nutrition*, 50(8): 728-760.
- Nauen, R., Reckmann, U., Thomzik, J. & Thielert, W. (2008). Biological profile of spirotetramat (Movento®)—a new two-way systemic (ambimobile) insecticide against sucking pest species. *Bayer Crop Science Journal*, 61(2): 245-278.
- Navarro, L., Bari, R., Achard, P., Lisón, P., Nemri, A., Harberd, N. P. & Jones, J. D. G. (2008). DELLAs control plant immune responses by modulating the balance of jasmonic acid and salicylic acid signaling. *Current Biology*, 18(9): 650-655.
- Ng, J. C. K. & Perry, K. L. (2004). Transmission of plant viruses by aphid vectors. *Molecular Plant Pathology*, 5(5): 505-511.
- Nimse, S. B. & Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *Rsc Advances*, 5(35): 27986-28006.
- Nishikawa, F., Kato, M., Hyodo, H., Ikoma, Y., Sugiura, M. & Yano, M. (2003). Ascorbate metabolism in harvested broccoli. *Journal of Experimental Botany*, 54(392): 2439-2448.
- Noguera, P., Abad, M., Puchades, R., Maquieira, A. & Noguera, V. (2003). Influence of particle size on physical and chemical properties of coconut coir dust as container medium. *Communications in Soil Science and Plant Analysis*, 34(3-4): 593-605.
- Nsabiya, V., Ochwo-Ssemakula, M. & Sseruwagi, P. (2012). Hot pepper reaction to field diseases. *African Crop Science Journal*, 20(1): 72-97.
- Nwose, E. U., Jelinek, H. F., Richards, R. S., Tinley, P. & Kerr, P. G. (2009). Atherothrombosis and oxidative stress: the connection and correlation in diabetes. *Redox Report*, 14(2): 55-60.
- O'Neal, M. E. & Johnson, K. D. (2010). 14 Insect Pests of Soybean and Their Management. In G. Singh (Ed.) *The Soybean: Botany, Production and Uses* (pp. 300-324). Cambridge: CABI.
- Oboh, G., Ademiluyi, A. O. & Faloye, Y. M. (2011). Effect of combination on the antioxidant and inhibitory properties of tropical pepper varieties against α -amylase and α -glucosidase activities in vitro. *Journal of Medicinal Food*, 14(10): 1152-1158.
- Oerke, E. C. (2006). Crop losses to pests. *The Journal of Agricultural Science*, 144(1): 31-43.
- Okwu, D. E. & Emenike, I. N. (2006). Evaluation of the phytonutrients and vitamins content of citrus fruits. *International Journal of Molecular Medicine and Advance Sciences*, 2(1): 1-6.

- Omer, A. D., Thaler, J. S., Granett, J. & Karban, R. (2000). Jasmonic acid induced resistance in grapevines to a root and leaf feeder. *Journal of Economic Entomology*, 93(3): 840-845.
- Orabi, S. A. & Abdelhamid, M. T. (2016). Protective role of α -tocopherol on two *Vicia faba* cultivars against seawater-induced lipid peroxidation by enhancing capacity of anti-oxidative system. *Journal of the Saudi Society of Agricultural Sciences*, 15(2): 145-154.
- Orozco-Cárdenas, M. L., Narváez-Vásquez, J. & Ryan, C. A. (2001). Hydrogen peroxide acts as a second messenger for the induction of defense genes in tomato plants in response to wounding, systemin, and methyl jasmonate. *The Plant Cell*, 13(1): 179-191.
- Orozco-Cardenas, M. & Ryan, C. A. (1999). Hydrogen peroxide is generated systemically in plant leaves by wounding and systemin via the octadecanoid pathway. *Proceedings of the National Academy of Sciences*, 96(11): 6553-6557.
- Ortuño, J., Cuesta, A., Esteban, M. A. & Meseguer, J. (2001). Effect of oral administration of high vitamin C and E dosages on the gilthead seabream (*Sparus aurata* L.) innate immune system. *Veterinary Immunology and Immunopathology*, 79(3): 167-180.
- Oukabli, A., Bartolini, S. & Viti, R. (2003). Anatomical and morphological study of apple (*Malus × domestica* Borkh.) flower buds growing under inadequate winter chilling. *The Journal of Horticultural Science and Biotechnology*, 78(4): 580-585.
- Overmyer, K., Brosché, M. & Kangasjärvi, J. (2003). Reactive oxygen species and hormonal control of cell death. *Trends in Plant Science*, 8(7): 335-342.
- Ozaslan, M., Bas, B., Aytakin, T. & Sigirci, Z. (2006). Identification of Pepper Viruses by Das-elisa Assays in Gaziantep-Turkey. *Plant Pathology Journal*. 5(1): 11-14.
- Ozawa, R., Shiojiri, K., Sabelis, M. W., Arimura, G.-I., Nishioka, T. & Takabayashi, J. (2004). Corn plants treated with jasmonic acid attract more specialist parasitoids, thereby increasing parasitization of the common armyworm. *Journal of Chemical Ecology*, 30(9): 1797-1808.
- Page, T., Griffiths, G. & Buchanan-Wollaston, V. (2001). Molecular and biochemical characterization of postharvest senescence in broccoli. *Plant Physiology*, 125(2): 718-727.
- Pakdevaraporn, P., Wasee, S., Taylor, P. W. J. & Mongkolporn, O. (2005). Inheritance of resistance to anthracnose caused by *Colletotrichum capsici* in Capsicum. *Plant Breeding*, 124(2): 206-208.
- Pallas, V. & García, J. A. (2011). How do plant viruses induce disease? Interactions and interference with host components. *Journal of General Virology*, 92(12): 2691-2705.

- Palma, J. M., Jiménez, A., Corpas, F. J., Mateos, R. M., Martí, M. C., Sevilla, F. & del Rfo, L. A. (2011). Role of ascorbate on the fruit physiology of pepper (*Capsicum annuum* L.). *Functional Plant Science and Biotechnology*, 5: 56-61.
- Parker, J. E. (2009). The quest for long-distance signals in plant systemic immunity. *Science Signalling*, 2(70): 31. Doi: 10.1126/scisignal.270pe31
- Parsons, K. K., Maeda, N., Yamauchi, M., Banes, A. J. & Koller, B. H. (2006). Ascorbic acid-independent synthesis of collagen in mice. *American Journal of Physiology-Endocrinology and Metabolism*, 290(6): 1131-1139.
- Pastori, G. M., Kiddle, G., Antoniow, J., Bernard, S., Veljovic-Jovanovic, S., Verrier, P. J., Noctor G. & Foyer, C. H. (2003). Leaf vitamin C contents modulate plant defense transcripts and regulate genes that control development through hormone signaling. *The Plant Cell*, 15(4): 939-951.
- Pavet, V., Olmos, E., Kiddle, G., Mowla, S., Kumar, S., Antoniow, J., Alvarez, M. E. & Foyer, C. H. (2005). Ascorbic acid deficiency activates cell death and disease resistance responses in Arabidopsis. *Plant Physiology*, 139(3): 1291-1303.
- Pazdro, R. & Burgess, J. R. (2010). The role of vitamin E and oxidative stress in diabetes complications. *Mechanisms of Ageing and Development*, 131(4): 276-286.
- Pedranzani, H., Sierra-de-Grado, R., Vigliocco, A., Miersch, O. & Abdala, G. (2007). Cold and water stresses produce changes in endogenous jasmonates in two populations of *Pinus pinaster* Ait. *Plant Growth Regulation*, 52(2): 111-116.
- Perz, J. F., Armstrong, G. L., Farrington, L. A., Hutin, Y. J. F. & Bell, B. P. (2006). The contributions of hepatitis B virus and hepatitis C virus infections to cirrhosis and primary liver cancer worldwide. *Journal of Hepatology*, 45(4): 529-538.
- Petrova, V. Y., Drescher, D., Kujumdzieva, A. V. & Schmitt, M. J. (2004). Dual targeting of yeast catalase A to peroxisomes and mitochondria. *Biochemical Journal*, 380(2): 393-400.
- Piotrowska, A., Bajguz, A., Godlewska-Żyłkiewicz, B., Czerpak, R. & Kamińska, M. (2009). Jasmonic acid as modulator of lead toxicity in aquatic plant *Wolffia arrhiza* (Lemnaceae). *Environmental and Experimental Botany*, 66(3): 507-513.
- Poonam, S., Kaur, H. & Geetika, S. (2013). Effect of jasmonic acid on photosynthetic pigments and stress markers in *Cajanus cajan* (L.) Millsp. seedlings under copper stress. *American Journal of Plant Sciences*, 4(4): 817-823.
- Popovich, D. G., Sia, S. Y., Zhang, W. & Lim, M. L. (2014). The color and size of chili peppers (*Capsicum annuum*) influence Hep-G2 cell growth. *International Journal of Food Sciences and Nutrition*, 65(7): 881-885.
- Prakash, S. & Singh, S. J. (2006). Insect transmitted viruses of peppers: A review. *Vegetable Science*, 33: 109-116.

- Pré, M., Atallah, M., Champion, A., De Vos, M., Pieterse, C. M. J. & Memelink, J. (2008). The AP2/ERF domain transcription factor ORA59 integrates jasmonic acid and ethylene signals in plant defense. *Plant Physiology*, 147(3): 1347-1357.
- Prochazkova, D., Sairam, R. K., Srivastava, G. C. & Singh, D. V. (2001). Oxidative stress and antioxidant activity as the basis of senescence in maize leaves. *Plant Science*, 161(4): 765-771.
- Qiu, Z., Guo, J., Zhu, A., Zhang, L. & Zhang, M. (2014). Exogenous jasmonic acid can enhance tolerance of wheat seedlings to salt stress. *Ecotoxicology and Environmental Safety*, 104: 202-208.
- Quisenberry, S. S. & Ni, X. (2007). Feeding Injury. In H. F. van Emden & R. Harrington (Eds.), *Aphids as Crop Pests* (pp. 331-352). Oxfordshire: CABI Bioscience.
- Radwan, D. E. M., Fayez, K. A., Mahmoud, S. Y. & Lu, G. (2010). Modifications of antioxidant activity and protein composition of bean leaf due to Bean yellow mosaic virus infection and salicylic acid treatments. *Acta Physiologiae Plantarum*, 32(5): 891-904.
- Rafidah, A., Faridah, S., Shahrul, A. A., Mazidah, M. & Zamri, I. (2016). Chronoamperometry Measurement for Rapid Cucumber Mosaic Virus Detection in Plants. *Procedia Chemistry*, 20: 25-28.
- Rahman, T., Roff, M. N. M. & Ghani, I. B. A. (2010). Within-field distribution of *Aphis gossypii* and aphidophagous lady beetles in chili, *Capsicum annuum*. *Entomologia Experimentalis et Applicata*, 137(3): 211-219.
- Rao, V. G. & Ramana, P. V. (2017). Natural Occurrence Of Cucumber Mosaic Virus In Chilli (*Capsicum annuum* L.) In Southern Telangana Zone Of Andhra Pradesh. *Global Journal of Bio-Science and Biotechnology*, 6(2): 221-223.
- Reische, D. W., Lillard, D. A. & Eitenmiller, R. R. (2008). Antioxidants. In C. C. Ahoh & D. B. Min (Eds.), *Food Lipids: Chemistry, Nutrition, and Biotechnology* (pp.409-428). Boca Raton: CRC Press.
- Riccioni, G., D’Orazio, N., Salvatore, C., Franceschelli, S., Pesce, M. & Speranza, L. (2012). Carotenoids and vitamins C and E in the prevention of cardiovascular disease. *International Journal for Vitamin and Nutrition Research*, 82(1): 15-26.
- Rice, M. E., O’Neal, M. E. & Pedersen, P. (2007). Soybean aphids in Iowa—2007. Reasearch Report SP 247. Agriculture and Environment Extension Publications. 89
- Rietjens, I. M. C. M., Boersma, M. G., de Haan, L., Spenkelink, B., Awad, H. M., Cnubben, N. H. P., van Zanden J. J., van der Woude H., Alink G. M. & Koeman, J. H. (2002). The pro-oxidant chemistry of the natural antioxidants vitamin C, vitamin E, carotenoids and flavonoids. *Environmental Toxicology and Pharmacology*, 11(3): 321-333.

- Roff, M. (1992). Epidemiology of aphidborn virus disease of chilli in Malaysia and their management. In B. H. Chew, W.H. Loke, R. Melor & A. R. Syed (Eds.), *Proceedings of the Conference on Chilli Pepper Production in the Tropics* (pp. 13-14). Kuala Lumpur: Malaysian Agricultural Research and Development Institute.
- Roggero, P., Masenga, V. & Tavella, L. (2002). Field isolates of Tomato spotted wilt virus overcoming resistance in pepper and their spread to other hosts in Italy. *Plant Disease*, 86(9): 950-954.
- Rogiers, S. Y., Kumar, G. N. M. & Knowles, N. R. (1998). Maturation and ripening of fruit of *Amelanchier alnifolia* Nutt. are accompanied by increasing oxidative stress. *Annals of Botany*, 81(2): 203-211.
- Rubbo, H., Radi, R., Anselmi, D., Kirk, M., Barnes, S., Butler, J., Eiserich, J. P. & Freeman, B. A. (2000). Nitric Oxide Reaction with Lipid Peroxyl Radicals Spares α -Tocopherol during Lipid Peroxidation Greater Oxidant Protection From The Pair Nitric Oxide/ α -Tocopherol Than α -tocopherol/ascorbate. *Journal of Biological Chemistry*, 275(15): 10812-10818.
- Ruiz-May, E., De-la-Pena, C., Galaz-Avalos, R. M., Lei, Z., Watson, B. S., Sumner, L. W. & Loyola-Vargas, V. M. (2011). Methyl jasmonate induces ATP biosynthesis deficiency and accumulation of proteins related to secondary metabolism in *Catharanthus roseus* (L.) G. hairy roots. *Plant and Cell Physiology*, 52(8): 1401-1421.
- Ryu, C., Murphy, J. F., Mysore, K. S. & Kloepper, J. W. (2004). Plant growth-promoting rhizobacteria systemically protect *Arabidopsis thaliana* against Cucumber mosaic virus by a salicylic acid and NPR1-independent and jasmonic acid-dependent signaling pathway. *The Plant Journal*, 39(3): 381-392.
- Salguero, A., de la Morena, B., Vigarra, J., Vega, J. M., Vilchez, C. & León, R. (2003). Carotenoids as protective response against oxidative damage in *Dunaliella bardawil*. *Biomolecular Engineering*, 20(4): 249-253.
- Samuel, G. & Bald, J. G. (1933). On the use of the primary lesions in quantitative work with two plant viruses. *Annals of Applied Biology*, 20(1): 70-99.
- Sandorf, I. & Holländer-Czytko, H. (2002). Jasmonate is involved in the induction of tyrosine aminotransferase and tocopherol biosynthesis in *Arabidopsis thaliana*. *Planta*, 216(1): 173-179.
- Sastry, K. S. (2013). *Seed-borne plant virus diseases*. India: Springer.
- Sathishkumar, R., Lakshmi, P. T. V. & Annamalai, A. (2010). Comparative analyses of non enzymatic and enzymatic antioxidants of *Encicostemma littorale* Blume. *International Journal of Pharma and Bio Sciences*, 1(2):1-16.
- Sathua, S. K., Reddy, M. S. S., Sulagitti, A. & Singh, R. N. (2017). Bio-efficacy of various insecticides and botanicals against chilli thrips (*S. dorsalis* Hood) and their comparative cost: Benefit analysis in chilli crop. *Journal of Entomology and*

Zoology Studies, 5(2): 130-134.

- Saxena, A., Raghuwanshi, R., Gupta, V. K. & Singh, H. B. (2016). Chilli anthracnose: The epidemiology and management. *Frontiers in Microbiology*, 7(1527):1-18.
- Schafer, F. Q., Qian, S. Y. & Buettner, G. R. (2000). Iron and free radical oxidations in cell membranes. *Cellular and Molecular Biology (Noisy-Le-Grand, France)*, 46(3): 657-662.
- Schaller, A. & Stintzi, A. (2009). Enzymes in jasmonate biosynthesis—structure, function, regulation. *Phytochemistry*, 70(13): 1532-1538.
- Scharte, J., Schön, H. & Weis, E. (2005). Photosynthesis and carbohydrate metabolism in tobacco leaves during an incompatible interaction with *Phytophthora nicotianae*. *Plant, Cell & Environment*, 28(11): 1421-1435.
- Schreinemachers, P., Balasubramaniam, S., Boopathi, N. M., Ha, C. V., Kenyon, L., Praneetvatakul, S., Sirijinda, A., Le, N. T. Srinivasan, R. & Wu, M. H. (2015). Farmers' perceptions and management of plant viruses in vegetables and legumes in tropical and subtropical Asia. *Crop Protection*, 75: 115-123.
- Schreiber, U. (2004). Pulse-amplitude-modulation (PAM) fluorometry and saturation pulse method: an overview. *Chlorophyll a Fluorescence*, 122: 279-319.
- Shyu, C. & Brutnell, T. P. (2015). Growth–defence balance in grass biomass production: the role of jasmonates. *Journal of Experimental Botany*, 66(14): 4165-4176.
- Seal, T. (2012). Antioxidant activity of some wild edible plants of Meghalaya state of India: A comparison using two solvent extraction systems. *International Journal of Nutrition and Metabolism*, 4(3): 51-56.
- Semida, W. M., Taha, R. S., Abdelhamid, M. T. & Rady, M. M. (2014). Foliar-applied α -tocopherol enhances salt-tolerance in *Vicia faba* L. plants grown under saline conditions. *South African Journal of Botany*, 95: 24-31.
- Sen, G., Eryilmaz, I. E. & Ozakca, D. (2014). The effect of aluminium-stress and exogenous spermidine on chlorophyll degradation, glutathione reductase activity and the photosystem II D1 protein gene (psbA) transcript level in lichen *Xanthoria parietina*. *Phytochemistry*, 98: 54-59.
- Seo, J.-K., Kwon, S.-J., Choi, H.-S. & Kim, K.-H. (2009). Evidence for alternate states of Cucumber mosaic virus replicase assembly in positive-and negative-strand RNA synthesis. *Virology*, 383(2): 248-260.
- Seo, M. & Koshiba, T. (2002). Complex regulation of ABA biosynthesis in plants. *Trends in Plant Science*, 7(1), 41-48.
- Serrano-Martínez, A., Fortea, M. I., del Amor, F. M. & Núñez-Delicado, E. (2008). Kinetic characterisation and thermal inactivation study of partially purified red pepper (*Capsicum annuum* L.) peroxidase. *Food Chemistry*, 107(1): 193-199.

- Sevik, M. A. (2012). Natural occurrence of *Cucumber mosaic virus* infecting water mint (*Mentha aquatica*) in Antalya and Konya, Turkey. *Acta Botanica Croatica*, 71(1): 187-193.
- Shan, C. & Liang, Z. (2010). Jasmonic acid regulates ascorbate and glutathione metabolism in *Agropyron cristatum* leaves under water stress. *Plant Science*, 178(2): 130-139.
- Shang, J., Xi, D.-H., Xu, F., Wang, S.-D., Cao, S., Xu, M.-Y., Zhaou, P.-P., Wang, J. -H., Jia, S. -D., Zhang, Z. -W., Yuan, S. & Zhang, Z.-W. (2011). A broad-spectrum, efficient and nontransgenic approach to control plant viruses by application of salicylic acid and jasmonic acid. *Planta*, 233(2): 299–308.
- Shanmugapriya, K., Saravana, P. S., Payal, H., Mohammed, S. P. & Williams, B. (2012). Antioxidant potential of pepper (*Piper nigrum* Linn.) leaves and its antimicrobial potential against some pathogenic microbes. *Indian Journal of Natural Products and Resources* 3(12): 570-577.
- Shao, H. -B., Chu, L. -Y., Lu, Z. -H. & Kang, C.-M. (2008). Primary antioxidant free radical scavenging and redox signaling pathways in higher plant cells. *International Journal of Biological Sciences*, 4(1): 8-14.
- Sharma, P., Jha, A. B., Dubey, R. S. & Pessarakli, M. (2012). Reactive Oxygen Species, Oxidative Damage, and Antioxidative Defense Mechanism in Plants under Stressful Conditions. *Journal of Botany*, 1-26.
- Sheteawi, S. A. (2007). Improving growth and yield of salt-stressed soybean by exogenous application of jasmonic acid and ascorbin. *International Journal of Agriculture and Biology (Pakistan)*, 9:473-478.
- Shigeoka, S., Ishikawa, T., Tamoi, M., Miyagawa, Y., Takeda, T., Yabuta, Y. & Yoshimura, K. (2002). Regulation and function of ascorbate peroxidase isoenzymes. *Journal of Experimental Botany*, 53(372): 1305-1319.
- Shumbe, L., Bott, R. & Havaux, M. (2014). Dihydroactinidiolide, a high light-induced β -carotene derivative that can regulate gene expression and photoacclimation in *Arabidopsis*. *Molecular Plant*, 7(7): 1248-1251.
- Siegel, R. L., Miller, K. D. & Jemal, A. (2016). Cancer statistics, 2016. *CA: A Cancer Journal for Clinicians*, 66(1): 7-30.
- Sies, H. & Stahl, W. (1995). Vitamins E and C, beta-carotene, and other carotenoids as antioxidants. *The American Journal of Clinical Nutrition*, 62(6): 1315-1321.
- Siger, A., Michalak, M., Cegielska-Taras, T., Szala, L., Lembicz, J. & Nogala-Kalucka, M. (2015). Genotype and environment effects on tocopherol and plastochromanol-8 contents of winter oilseed rape doubled haploid lines derived from F1 plants of the cross between yellow and black seeds. *Industrial Crops and Products*, 65: 134-141.

- Simón, A., García, C., Pascual, F., Ruiz, L. & Janssen, D. (2016). The Influence of Crop Habitat and Control Strategies on Pepper Viruses in Andalusia (Spain). *Horticulturae*, 2(4): 1-9.
- Singh, R. & Dwivedi, U. N. (2008). Effect of ethrel and 1-methylcyclopropene (1-MCP) on antioxidants in mango (*Mangifera indica* var. Dashehari) during fruit ripening. *Food Chemistry*, 111(4): 951-956.
- Singh, S., Pandey, V. P., Naaz, H. & Dwivedi, U. N. (2012). Phylogenetic analysis, molecular modeling, substrate-inhibitor specificity, and active site comparison of bacterial, fungal, and plant heme peroxidases. *Biotechnology and Applied Biochemistry*, 59(4): 283-294.
- Sivaci, A. & Duman, S. (2014). Evaluation of seasonal antioxidant activity and total phenolic compounds in stems and leaves of some almond (*Prunus amygdalus* L.) varieties. *Biological Research*, 47(1): 2-5.
- Skibsted, L. H. (2012). Carotenoids in antioxidant networks. Colorants or radical scavengers. *Journal of Agricultural and Food Chemistry*, 60(10): 2409-2417.
- Smirnoff, N. (2000). Ascorbic acid: metabolism and functions of a multi-faceted molecule. *Current Opinion in Plant Biology*, 3(3): 229-235.
- Smirnoff, N. & Wheeler, G. L. (2000). Ascorbic acid in plants: biosynthesis and function. *Critical Reviews in Biochemistry and Molecular Biology*, 35(4): 291-314.
- Sofo, A., Scopa, A., Nuzzaci, M. & Vitti, A. (2015). Ascorbate peroxidase and catalase activities and their genetic regulation in plants subjected to drought and salinity stresses. *International Journal of Molecular Sciences*, 16(6): 13561-13578.
- Song, F., & Swinton, S. M. (2009). Returns to integrated pest management research and outreach for soybean aphid. *Journal of Economic Entomology*, 102(6): 2116-2125.
- Song, X., Wang, Y., Mao, W., Shi, K., Zhou, Y., Nogués, S. & Yu, J. (2009). Effects of cucumber mosaic virus infection on electron transport and antioxidant system in chloroplasts and mitochondria of cucumber and tomato leaves. *Physiologia Plantarum*, 135(3): 246-257.
- Sorial, M. E., El-Gamal, S. M. & Gendy, A. A. (2010). Response of sweet basil to jasmonic acid application in relation to different water supplies. *Bioscience Research*, 7: 39-47.
- Sreelatha, S. & Padma, P. R. (2009). Antioxidant activity and total phenolic content of *Moringa oleifera* leaves in two stages of maturity. *Plant Foods for Human Nutrition*, 64(4): 303-311.
- Srivalli, B. & Khanna-Chopra, R. (2001). Induction of new isoforms of superoxide dismutase and catalase enzymes in the flag leaf of wheat during monocarpic senescence. *Biochemical and Biophysical Research Communications*, 288(4): 1037-1042.

- Staswick, P. E. (2009). The tryptophan conjugates of jasmonic and indole-3-acetic acids are endogenous auxin inhibitors. *Plant Physiology*, 150(3): 1310-1321.
- Staswick, P. E. & Tiryaki, I. (2004). The oxylipin signal jasmonic acid is activated by an enzyme that conjugates it to isoleucine in Arabidopsis. *The Plant Cell*, 16(8): 2117-2127.
- Stelmach, B. A., Müller, A., Hennig, P., Gebhardt, S., Schubert-Zsilavecz, M. & Weiler, E. W. (2001). A novel class of oxylipins, sn1-O-(12-oxophytodienoyl)-sn2-O-(hexadecatrienoyl)-monogalactosyl diglyceride, from *Arabidopsis thaliana*. *Journal of Biological Chemistry*, 276(16): 12832-12838.
- Stevens, R., Page, D., Gouble, B., Garchery, C., Zamir, D. & Causse, M. (2008). Tomato fruit ascorbic acid content is linked with monodehydroascorbate reductase activity and tolerance to chilling stress. *Plant, Cell & Environment*, 31(8): 1086-1096.
- Sunitha, T. R. (2007). Insect pests of *Capsicum annuum* var. *frutescens* (L.) and their management. (Published master dissertation). University Of Agricultural Sciences, Dharwad.
- Swarbrick, P. J., Schulze-Lefert, P. & Scholes, J. D. (2006). Metabolic consequences of susceptibility and resistance (race-specific and broad-spectrum) in barley leaves challenged with powdery mildew. *Plant, Cell & Environment*, 29(6): 1061-1076.
- Taiwo, M. A. & Akinjogunla, O. J. (2006). Cowpea viruses: quantitative and qualitative effects of single and mixed viral infections. *African Journal of Biotechnology*, 5(19):1749-1756.
- Takahama, U. (2004). Oxidation of vacuolar and apoplastic phenolic substrates by peroxidase: physiological significance of the oxidation reactions. *Phytochemistry Reviews*, 3(1): 207-219.
- Takahashi, S. & Badger, M. R. (2011). Photoprotection in plants: A new light on photosystem II damage. *Trends in Plant Science*, 16(1): 53-60.
- Tan, X., Zhang, D., Wintgens, C., Willingmann, P., Adam, G. & Heinze, C. (2012). A comparative testing of Cucumber mosaic virus (CMV)-based constructs to generate virus resistant plants. *American Journal of Plant Sciences*, 3(4): 461-472.
- Thackray, D. J., Diggle, A. J., Berlandier, F. A. & Jones, R. A. C. (2004). Forecasting aphid outbreaks and epidemics of Cucumber mosaic virus in lupin crops in a Mediterranean-type environment. *Virus Research*, 100(1): 67-82.
- Thaler, J. S., Owen, B. & Higgins, V. J. (2004). The role of the jasmonate response in plant susceptibility to diverse pathogens with a range of lifestyles. *Plant Physiology*, 135(1): 530-538.
- Thaler, J. S., Stout, M. J., Karban, R. & Duffey, S. S. (2001). Jasmonate-mediated induced plant resistance affects a community of herbivores. *Ecological Entomology*, 26(3): 312-324.

- Than, P. P., Prihastuti, H., Phoulivong, S., Taylor, P. W. J., & Hyde, K. D. (2008). Chilli anthracnose disease caused by *Colletotrichum* species. *Journal of Zhejiang University. Science. B*, 9(10): 764-778.
- Thiyam, U., Stöckmann, H. & Schwarz, K. (2006). Antioxidant activity of rapeseed phenolics and their interactions with tocopherols during lipid oxidation. *Journal of the American Oil Chemists' Society*, 83(6): 523-528.
- Thomas, C. E., McLean, L. R., Parker, R. A. & Ohlweiler, D. F. (1992). Ascorbate and phenolic antioxidant interactions in prevention of liposomal oxidation. *Lipids*, 27(7): 543-550.
- Thonemann, B., & Schmalz, G. (2000). Immortalization of bovine dental papilla cells with simian virus 40 large t antigen. *Archives of Oral Biology*, 45(10): 857-869.
- Tierranegra-García, N., Salinas-Soto, P., Torres-Pacheco, I., Ocampo-Velázquez, R. V., Rico-García, E., Mendoza-Díaz, S. O., Feregrino-Pérez, A.A., Mercado-Luna, A., Vargas-Hernandez, M., Guevara-González, R.G. & Soto-Zarazúa, G. M. (2011). Effect of foliar salicylic acid and methyl jasmonate applications on protection against pill-bugs in lettuce plants (*Lactuca sativa*). *Phytoparasitica*, 39(2): 137-144.
- Torrigiani, P., Fregola, F., Ziosi, V., Ruiz, K. B., Kondo, S. & Costa, G. (2012). Differential expression of allene oxide synthase (AOS), and jasmonate relationship with ethylene biosynthesis in seed and mesocarp of developing peach fruit. *Postharvest Biology and Technology*, 63(1): 67-73.
- Touhidur, M. R., Idris, A. B. & Roff, M. N. M. (2006). Population abundance and spatial distribution of *Aphis gossypii* Glover (Homoptera : Aphididae) and coccinellids on chilli (*Capsicum annum* L.). *Journal of Tropical Agriculture and Food Sciences*, 34(2): 393-403.
- Traber, M. G. & Atkinson, J. (2007). Vitamin E, antioxidant and nothing more. *Free Radical Biology and Medicine*, 43(1): 4-15.
- Trebst, A., Depka, B. & Holländer-Czytko, H. (2002). A specific role for tocopherol and of chemical singlet oxygen quenchers in the maintenance of photosystem II structure and function in *Chlamydomonas reinhardtii*. *FEBS Letters*, 516(1-3): 156-160.
- Truman, W., Bennett, M. H., Kubigsteltig, I., Turnbull, C. & Grant, M. (2007). Arabidopsis systemic immunity uses conserved defense signaling pathways and is mediated by jasmonates. *Proceedings of the National Academy of Sciences*, 104(3): 1075-1080.
- Turner, J. G., Ellis, C. & Devoto, A. (2002). The jasmonate signal pathway. *The Plant Cell*, 14(1): 153-164.
- Ulloa, R. M., Raíces, M., MacIntosh, G. C., Maldonado, S. & Téllez-Iñón, M. T. (2002). Jasmonic acid affects plant morphology and calcium-dependent protein kinase

- expression and activity in *Solanum tuberosum*. *Physiologia Plantarum*, 115(3), 417-427.
- Umoren, S. A., Eduok, U. M., Israel, A. U., Obot, I. B. & Solomon, M. M. (2012). Coconut coir dust extract: a novel eco-friendly corrosion inhibitor for Al in HCl solutions. *Green Chemistry Letters and Reviews*, 5(3): 303-313.
- Uttara, B., Singh, A. V, Zamboni, P. & Mahajan, R. T. (2009). Oxidative Stress and Neurodegenerative Diseases: A Review of Upstream and Downstream Antioxidant Therapeutic Options. *Current Neuropharmacol*, 7:65-74.
- van der Ent, S., Van Wees, S. C. M. & Pieterse, C. M. J. (2009). Jasmonate signaling in plant interactions with resistance-inducing beneficial microbes. *Phytochemistry*, 70(13): 1581-1588.
- van der Geest, L. P. S., Elliot, S. L., Breeuwer, J. A. J. & Beerling, E. A. M. (2000). Diseases of mites. *Experimental & Applied Acarology*, 24(7): 497-560.
- van Doorn, W. G. & Ketsa, S. (2014). Cross reactivity between ascorbate peroxidase and phenol (guaiacol) peroxidase. *Postharvest Biology and Technology*, 95: 64-69.
- van Eenennaam, A. L., Lincoln, K., Durrett, T. P., Valentin, H. E., Shewmaker, C. K., Thorne, G. M., Jiang, J., Baszis, S. R. Levering, C. K., Aasen, E. D., Hao, H., Stein, J. C., Norris, S. R. & Last, R.L. (2003). Engineering vitamin E content: from Arabidopsis mutant to soy oil. *The Plant Cell*, 15(12): 3007-3019.
- Vichitbandha, P. & Chandrapatya, A. (2011). Broad mite effects on chili shoot damage and yields. *Pakistan Journal of Zoology*, 43: 637-649.
- Vick, B. A. & Zimmerman, D. C. (1983). The biosynthesis of jasmonic acid: a physiological role for plant lipoxygenase. *Biochemical and Biophysical Research Communications*, 111(2): 470-477.
- Wang, L., Allmann, S., Wu, J. & Baldwin, I. T. (2008). Comparisons of Lipoxygenase3- and Jasmonate-Resistant4/6-silenced plants reveal that jasmonic acid and jasmonic acid-amino acid conjugates play different roles in herbivore resistance of *Nicotiana attenuata*. *Plant Physiology*, 146(3): 904-915.
- Wang, W.-B., Kim, Y.-H., Lee, H.-S., Kim, K.-Y., Deng, X.-P. & Kwak, S.-S. (2009). Analysis of antioxidant enzyme activity during germination of alfalfa under salt and drought stresses. *Plant Physiology and Biochemistry*, 47(7): 570-577.
- Wasternack, C. & Hause, B. (2002). Jasmonates and octadecanoids: signals in plant stress responses and development. *Progress in Nucleic Acid Research and Molecular Biology*, 72: 165-221.
- Wasternack, C. & Hause, B. (2013). Jasmonates: biosynthesis, perception, signal transduction and action in plant stress response, growth and development. An update to the 2007 review in *Annals of Botany*, 111(6): 1021-1058.

- Wei, J., Wen, X. & Tang, L. (2017). Effect of methyl jasmonic acid on peach fruit ripening progress. *Scientia Horticulturae*, 220: 206-213.
- Weintraub, P. G., Kleitman, S., Alchanatis, V. & Palevsky, E. (2007). Factors affecting the distribution of a predatory mite on greenhouse sweet pepper. *Experimental and Applied Acarology*, 42(1): 23-35.
- Weiss, E. A. (2002). *Spice Crops*. Wallington, England New York: CABI.
- Westwood, J. H., Lewsey, M. G., Murphy, A. M., Tungadi, T., Bates, A., Gilligan, C. A. & Carr, J. P. (2014). Interference with jasmonic acid-regulated gene expression is a general property of viral suppressors of RNA silencing but only partly explains virus-induced changes in plant–aphid interactions. *Journal of General Virology*, 95(3): 733-739.
- Westwood, J. H. & Stevens, M. (2010). Resistance to aphid vectors of virus disease. *Advances in Virus Research*, 76: 179-210.
- Whitham, S. A. & Wang, Y. (2004). Roles for host factors in plant viral pathogenicity. *Current Opinion in Plant Biology*, 7(4): 365-371.
- Wild, C. P. & Gong, Y. Y. (2009). Mycotoxins and human disease: a largely ignored global health issue. *Carcinogenesis*, 31(1): 71-82.
- Williamson, S., Ball, A. & Pretty, J. (2008). Trends in pesticide use and drivers for safer pest management in four African countries. *Crop Protection*, 27(10): 1327-1334.
- Wintermantel, W. M., Wisler, G. C., Anchieta, A. G., Liu, H.-Y., Karasev, A. V. & Tzanetakis, I. E. (2005). The complete nucleotide sequence and genome organization of Tomato chlorosis virus. *Archives of Virology*, 150(11): 2287-2298.
- Wira, A. B., Razi, I. M. & Jamil, Z. A. (2011). Composts as additives in coconut coir dust culture for growing rockmelon (*Cucumis melo* L.). *Journal of Tropical Agriculture and Food Science*, 39(2): 229-237.
- Wolucka, B. A., Goossens, A. & Inzé, D. (2005). Methyl jasmonate stimulates the de novo biosynthesis of vitamin C in plant cell suspensions. *Journal of Experimental Botany*, 56(419): 2527-2538.
- Wolucka, B. A. & Van Montagu, M. (2003). GDP-mannose 3', 5'-epimerase forms GDP-L-gulose, a putative intermediate for the de novo biosynthesis of vitamin C in plants. *Journal of Biological Chemistry*, 278(48): 47483-47490.
- World Health Organization. (2015). Cancer. Retrieved from <http://www.who.int/mediacentre/factsheets/fs297/en>.
- Wu, J., Wang, L. & Baldwin, I. T. (2008). Methyl jasmonate-elicited herbivore resistance: does MeJA function as a signal without being hydrolyzed to JA? *Planta*, 227(5): 1161-1168.
- Xi, D., Liu, W., Yang, G., Wu, C. & Zheng, C. (2010). Seed-specific overexpression of

antioxidant genes in Arabidopsis enhances oxidative stress tolerance during germination and early seedling growth. *Plant Biotechnology Journal*, 8(7): 796-806.

- Xu, P., Chen, F., Mannas, J. P., Feldman, T., Sumner, L. W. & Roossinck, M. J. (2008). Virus infection improves drought tolerance. *New Phytologist*, 180(4): 911-921.
- Yadav, H. & Yadav, P. K. (2010). A Study of Mitotic Cell Division in *Capsicum annum* L induced by Chilli Mottle Virus Disease. *Asian Journal of Experimental Biological Sciences*, 1(2): 445-447.
- Yadegari, M. & Shakerian, A. (2014). Journal of Applied Science and Agriculture The Effect Salicylic Acid and Jasmonic Acid Foliar Applications on Essence and Essential Oil of Salvia (*Salvia Officinalis* L.). *Journal of Applied Science and Agriculture*, 9(4): 1578-1584.
- Yamauchi, Y., Furutera, A., Seki, K., Toyoda, Y., Tanaka, K. & Sugimoto, Y. (2008). Malondialdehyde generated from peroxidized linolenic acid causes protein modification in heat-stressed plants. *Plant Physiology and Biochemistry*, 46(8): 786-793.
- Yan, Z., Li, X., Chen, J. & Tam, N. F.-Y. (2015). Combined toxicity of cadmium and copper in *Avicennia marina* seedlings and the regulation of exogenous jasmonic acid. *Ecotoxicology and Environmental Safety*, 113: 124-132.
- Yang, T. & Poovaiah, B. W. (2002). Hydrogen peroxide homeostasis: activation of plant catalase by calcium/calmodulin. *Proceedings of the National Academy of Sciences*, 99(6): 4097-4102.
- Yao, H. & Tian, S. (2005). Effects of pre-and post-harvest application of salicylic acid or methyl jasmonate on inducing disease resistance of sweet cherry fruit in storage. *Postharvest Biology and Technology*, 35(3): 253-262.
- Yeruva, L., Elegbede, J. A. & Carper, S. W. (2008). Methyl jasmonate decreases membrane fluidity and induces apoptosis via tumor necrosis factor receptor 1 in breast cancer cells. *Anti-Cancer Drugs*, 19(8): 766-776.
- Yoshida, K., Kaothien, P., Matsui, T., Kawaoka, A. & Shinmyo, A. (2003). Molecular biology and application of plant peroxidase genes. *Applied Microbiology and Biotechnology*, 60(6): 665-670.
- Yoshida, Y., Niki, E. & Noguchi, N. (2003). Comparative study on the action of tocopherols and tocotrienols as antioxidant: chemical and physical effects. *Chemistry and Physics of Lipids*, 123(1): 63-75.
- Yoshioka, H., Mase, K., Yoshioka, M., Kobayashi, M. & Asai, S. (2011). Regulatory mechanisms of nitric oxide and reactive oxygen species generation and their role in plant immunity. *Nitric Oxide*, 25(2): 216-221.
- Young, A. J. & Lowe, G. M. (2001). Minireview Antioxidant and Prooxidant Properties of Carotenoids. *Archives of Biochemistry and Biophysics*, 385(1): 20-27.

- Zakaria, Z., Heng, L. Y., Abdullah, P., Osman, R. & Din, L. (2003). The environmental contamination by organochlorine insecticides of some agricultural areas in Malaysia. *Malaysian Journal of Chemistry*, 5(1): 78-85.
- Yuan, X. K., Yang, Z. Q., Li, Y. X., Liu, Q., & Han, W. (2016). Effects of different levels of water stress on leaf photosynthetic characteristics and antioxidant enzyme activities of greenhouse tomato. *Photosynthetica*, 54(1): 28-39.
- Zhou, J. & Jin, S. (2009). Safety of vegetables and the use of pesticides by farmers in China: Evidence from Zhejiang province. *Food Control*, 20(11): 1043-1048.
- Zhu, F., Xi, D.-H., Yuan, S., Xu, F., Zhang, D.-W. & Lin, H.-H. (2014). Salicylic acid and jasmonic acid are essential for systemic resistance against tobacco mosaic virus in *Nicotiana benthamiana*. *Molecular Plant-Microbe Interactions*, 27(6): 567-577.
- Zimmermann, P. & Zentgraf, U. (2005). The correlation between oxidative stress and leaf senescence during plant development. *Cellular and Molecular Biology Letters*, 10(3): 515-534.
- Zitter, T. A. & Murphy, J. F. (2009). Cucumber mosaic virus. *Annals of Applied Biology*, 66: 381-386.
- Zollinger, P. E., Tuinebreijer, W. E., Breederveld, R. S. & Kreis, R. W. (2007). Can Vitamin C Prevent Complex Regional Pain Syndrome in Patients with Wrist Fractures?: A Randomized, Controlled, Multicenter Dose-Response Study. *Journal of Bone & Joint Surgery*, 89(7): 1424-1431.