



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF BACTERIAL QUORUM SENSING DEGRADER ON
Artemia franciscana CULTURE***

LIM YAN LI

FP 2013 92

EFFECTS OF BACTERIAL QUORUM SENSING DEGRADER ON

Artemia franciscana CULTURE



LIM YAN LI

DEPARTMENT OF AQUACULTURE

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

SERDANG, SELANGOR

2013

CERTIFICATION OF APPROVAL
DEPARTMENT OF AQUACULTURE
FACULTY OF AGRICULTURE
UNIVERSITI PUTRA MALAYSIA

Name of student: Lim Yan Li

Matric Number: 159266

Programme: Bachelor of Agriculture (Aquaculture)

Year: 2013

Name of supervisor: Dr. Natrah Fatin Mohd Ikhsan

Title of Project:

Effects of bacterial Quorum Sensing Degradar on *Artemia franciscana* culture.

This is to certify that I have examined the final project report and all corrections have been made as recommended by the panel of examiners. This report complies with the recommended format stipulated in the AKU4999 project guidelines, Department of Aquaculture, Faculty of Agriculture, Universiti Putra Malaysia.

Signature and official stamp of supervisor/co-supervisor:

Dr. Natrah Fatin Mohd Ikhsan

Date:

ACKNOWLEDGEMENT

The first thought that crossed my mind the moment I had completed my Final Year Academic Project (AKU4999) was to contribute my sincere gratitude to my supervisor, Dr. Natrah Fatin Mohd Ikhsan. She had given me a tremendous assistance and support throughout my study and research. She was willing to spend her time and give me guidance. She did every arrangement she could do and prepared all the equipments that I need start from the very beginning just to help me to complete my research. Without her guidance, I would not able to complete my research.

I am grateful to Noorashikin Md Noor who had been helping me a lot during my research. She was the friends that helped me to prepare the equipments I needed. She taught me the knowledge to handle practical work and the safety precaution in laboratory. I would face difficulty without her help because I did not have much practical experience.

I am also grateful to Pn. Nur Shafika Maulad Abd Jalil because she would help me to handle and gather the equipments need in the research. She also taught me how to prepare the chemical solution since it is dangerous or hazardous. She is an experienced officer in dealing with laboratory practical.

Meanwhile, I would like to express my sincere gratitude to lecturer, demonstrator, and staff of Aquaculture Department for helping me, providing me helpful suggestion and guidance. Last but not least, deepest thanks to my beloved parents for giving me support and inner strength.

ABSTRACT

Quorum sensing is a bacterial cell-to-cell communication with small signal molecules such as acyl-homoserine lactones (AHL) that can regulate the virulence of many pathogenic bacteria. Therefore, quorum sensing interference using degrader strains is a biocontrol strategy to fight bacterial infections. In this study, one of the quorum sensing degrader strains, BP-ART/6 was successfully isolated from *Artemia franciscana* by enriching newly hatched *Artemia* nauplii in AHL. Anti-quorum sensing activity of BP-ART/6 was screened using *Chromobacterium violaceum* CV026 biosensor bioassay, and having the best degradation properties which fully degrade 10ppm of AHL in less than 24 hours. The Gram staining test indicated BP-ART/6 as Gram-negative bacteria. Strain BP-ART/6 was resistant to antibiotic kanamycin and rifampicin at 20, 50 and 100ppm, respectively after 24 hours and 48 hours of incubation. Result from antibacterial study showed BP-ART/6 has no antibacterial activity against *Vibrio campbellii* ATCC14126, *Vibrio alginolyticus* ATCC17749 and *Vibrio anguillarum* ATCC43313. Significant differences ($p < 0.05$) of survivability in the presence of the degrader strain can be observed between control and the treatment with the QS degraders at both 10^5 and 10^6 CFU/ml for 6, 12 and 24 hours. At the same time, encapsulation of degrader strains BP-ART/6 by *Artemia* was less than 30 colonies as the bacterial counts lay below the countable range of 30 to 300 colonies.

ABSTRAK

Kuorum penginderaan merupakan komunikasi antara sel bacteria dengan molekul kecil iaitu N-acy-homoserine lakton (AHL) yang boleh mengawal kebiasaan pelbagai bacteria berpatogenik. Oleh itu, penggunaan strain degradasi dalam gangguan kuorum penginderaan merupakan strategi kawalan biologi untuk menentang jangkitan bacteria. Salah satu strain degradasi, BP-ART/6, telah berjaya diasingkan melalui kaedah pengayaan daripada *Artemia franciscana* yang baru menetas. Aktiviti anti-Kuorum penginderaan BP-ART/6 telah diuji menggunakan *Chromobacterium violaceum* CV026 sebagai biosensor bioesei, and mempunyai ciri-ciri degradasi yang baik yang boleh mendegradkan 10ppm AHL dalam masa kurang daripada 24 jam. Ujian Gram stain menunjukkan BP-ART/6 sebagai bacteria Gram-negatif. BP-ART/6 resisten terhadap antibiotic kanamycin and rifampicin pada 20, 50 and 100ppm selepas 24 dan 48 jam inkubasi. BP-ART/6 tidak menunjukkan aktiviti antibakteria terhadap *Vibrio campbellii* ATCC14126, *Vibrio alginolyticus* ATCC17749 and *Vibrio anguillarum* ATCC43313. Perbezaan signifikan ($p < 0.05$) tahap kehidupan dalam kewujudan strain degradasi amat ketara antara kontrol dengan sampel degradasi pada 10^5 and 10^6 CFU/ml untuk 6, 12 and 24 jam. Pada masa yang sama, pengkapsulan strain degradasi oleh *Artemia* adalah rendah disebabkan jumlah kiraan bacteria adalah kurang daripada 30 hingga 300 koloni.

TABLE OF CONTENTS

	Page
Contents	
ACKNOWLEDGEMENT	I
ABSTRACT	II
ABSTRAK	III
TABLE OF CONTENTS	IV
LIST OF TABLES	VII
LIST OF FIGURES	VIII
LIST OF SYMBOLS	IX
1.0 INTRODUCTION	1
2.0 LITERATURE REVIEW	3
2.1 Live feed interaction with aquatic host	3
2.2 Common practices to overcome bacterial disease	13
2.3 Quorum sensing	13
2.3.1 Bacterial quorum sensing	14
2.4 Quorum sensing interference as an alternative	17
2.4.1 AHL degrader interference with quorum sensing	17
3.0 MATERIALS AND METHODS	21
3.1 Enrichment of AHL degraders from <i>Artemia</i>	21
3.1.1 Preparation of microbial communities (MC)	21

3.1.2 Isolation of AHL degrading strain	21
3.2 Quorum sensing degradation assay	22
3.3 Gram's stain of AHL degraders	23
3.4 Antibiotic resistance test for AHL degrading strains	24
3.5 Antibacterial Cross Streak Method (CSM)	24
3.6 <i>In vivo</i> test	25
3.6.1 Bacteria- free <i>Artemia</i> nauplii	25
3.6.2 Test on <i>Artemia</i> encapsulation and survival	26
3.6.3 Bacterial count in <i>Artemia</i>	27
4.0 RESULTS	28
4.1 Enrichment of AHL degraders from <i>Artemia</i>	28
4.2 Quantitative quorum sensing degradation assay	30
4.3 Gram staining	35
4.4 Antibiotic resistance study	37
4.5 Antibacterial activity	39
4.6 <i>In vivo</i> test	41
4.6.1 Preliminary test on gnotobiotic <i>Artemia</i> encapsulation and survival	41
4.6.2 Bacterial count	44

5.0 DISCUSSION	45
5.1 Quorum sensing degradation from bacteria	45
5.2 Gram staining test	48
5.3 Antibiotic resistance study	49
5.4 Antibacterial activity	50
5.5 <i>In vivo</i> test	51
5.5.1 Preliminary test on gnotobiotic <i>Artemia</i> encapsulation and survival	51
5.5.2 Bacterial counts	52
6.0 CONCLUSION	53
REFERENCES	54

LIST OF TABLES

		Page
Table 1	Live feed interaction with bacteria in different culture	6
Table 2	Bacterial enzymes as quorum sensing inhibitors.	20
Table 3	Concentration level of AHL after degradation activity for two different durations.	33
Table 4	Antibiotic resistance of degrader strains BP-ART/6 against 4 different antibiotics.	38
Table 5	Antibacterial activity of degrader samples against three different <i>Vibrio</i> pathogen strains.	40
Table 6	Survival count of <i>Artemia</i> in different concentration of bacteria sample for three different durations.	43

LIST OF FIGURES

		Page
Figure 1	Multi-channel signaling in <i>Vibrio campbellii</i> .	16
Figure 2	Six degrader strains isolated from <i>Artemia</i> with AHL degradation properties.	29
Figure 3	The purple pigmentation for different AHLs concentration for standard curve.	31
Figure 4	Calibration standard curve on the diameter of the ring formed according to the known concentration of AHL for the determination on degradation of AHL degrader strains.	31
Figure 5	Detection of AHLs degradation effect by purple violacein pigmentation.	34
Figure 6	Gram staining for bacteria sample BP-ART/6 under 100x magnification	36
Figure 7	Antibacterial test for degrader strains BP-ART/1 to BP-ART/6 using cross streak method.	40

LIST OF SYMBOLS

%	Percent
°C	Degree Celsius
≈	Approximate
>	Greater than
<	Less than
rpm	Round per minute
ppm	Parts per million
US\$	United States dollar
min	Minutes
x	Times
M	Molar
CFU	Colony-forming unit
g	Gram
kg	Kilogram
mm	Millimetre
cm	Centimetre
ml	milliliter
L	Liter
μm	Micrometre
μl	Microliter

CHAPTER 1

INTRODUCTION

Quorum sensing (QS) is a mechanism in which bacteria are able to coordinate the expression of certain genes in response to the presence of small signal molecules (Hense *et al.*, 2007). It is a type of bacterial cell-cell communication that can regulate the expression of virulence genes in many pathogenic bacteria, and most of them usually use different signal molecules to regulate virulence gene expression (Pande *et al.*, 2013). Bassler (2002) stated that quorum sensing can regulate bioluminescence, virulence factor expression, biofilm formation, sporulation, and mating. According to Natrah (2011b), aquaculture pathogens also use QS to regulate the expression of important virulence phenotypes. High mortality of larvae is observed in *Macrobrachium rosenbergii* (Baruah *et al.*, 2008) and burbot larvae (Natrah *et al.*, 2012) in the presence of pathogen with QS activity.

Fish larvae often undergo heavy mortalities in early stage which is four to nine days after hatching (Kohno *et al.*, 1997), and often exposed to *Vibrio* diseases such as *Vibrio alginolyticus*, *Vibrio harveyi*, *Vibrio vulnificus* and other *Vibrio* spp. (Huang *et al.*, 2012). Among the preventive measures are through the use of antibiotic. However, the frequent use of antibiotic treatment has resulted in the development of antibiotic resistance, causing this compound to be ineffective to control vibriosis (Defoirdt *et al.*, 2007). Since QS is important for the virulence

towards aquatic host, QS degrader could also be an alternative solution for disease control (Defoirdt *et al.*, 2013). Hence, QS degrader strain can be inculcated into live feed as a carrier.

Live prey such as *Artemia* nauplii have successively been used to rear marine fish larvae because it is high in free amino acids with small peptides, or other water-soluble nutrients (Johnson *et al.*, 2009). The research also stated *Artemia* having consistent availability and quality which increased nutrient delivery and addition of probiotic can support the larval growth of a particular species. Hence, the objectives of this research are:

1. To isolate quorum sensing degrader from *Artemia*
2. To test the quorum sensing degrader encapsulated to *Artemia*

REFERENCES

- Asanka Gunasekara, R.A.Y.S., Rekecki, A., Baruah, K., Bossier, P. and Van den Broek, W. (2010). Evaluation of probiotic effect of *Aeromonas hydrophila* on the development of the digestive tract of germ-free *Artemia franciscana* nauplii. *Journal of Experimental Marine Biology and Ecology.*, 393, 78–82.
- Bai, F., Han, Y., Chen, J. and Zhang, X. H. (2008). Disruption of quorum sensing in *Vibrio harveyi* by the AiiA protein of *Bacillus thuringiensis*. *Aquaculture.*, 274(1), 36-40.
- Baruah, K., Cam, D.T.V., Diercken, K., Wille, M., Defoirdt, T., Sorgeloos, P. and Bossier, P. (2008). *In vivo* effects of single or combined N-acyl homoserine lactone quorum sensing signals on the performance of *Macrobrachium rosenbergii* larvae. *Aquaculture.*, 288, 233–238.
- Bassler, B. L. (2002). Small talk: cell-to-cell communication in bacteria. *Cell.*, 109(4), 421-424.
- Binnerup, S. J., Højberg, O. and Sørensen, J. (1998). Gram characteristics determined on single cells and at the microcolony level of bacteria immobilized on polycarbonate membrane filters. *Journal of microbiological methods.*, 31(3), 185-192.
- Cam, D.T.V., Hao, N.V., Dierckens, K., Defoirdt, T., Boon, N., Sorgeloos, P., Bossier, P. (2009a). Novel approach of using homoserine lactone degrading and poly- β -hydroxybutyrate accumulating bacteria to protect *Artemia* from the pathogenic effects of *Vibrio harveyi*. *Aquaculture.*, 291, 23–30.
- Cam, D.T.V., Dinh, T.N., Ceuppens, S., Nguyen, V.H., Dierckens, K., Wille, M., Sorgeloos, P. and Bossier, P. (2009b). Effect of N-acyl homoserine lactone-degrading enrichment cultures on *Macrobrachium rosenbergii* larviculture. *Aquaculture.*, 294 (1-2), 5–13
- Chen, T., Lin, H., Lin, C., Lu, C. and Chen, Y. (2012). *Picochlorum* as an alternative to *Nannochloropsis* for grouper larval rearing. *Aquaculture.*, 338-341, 82–88
- Chiu, C., Cheng, C., Gua, W., Guu, Y. and Cheng, W. (2010). Dietary administration of the probiotic, *Saccharomyces cerevisiae* P13, enhanced the growth, innate immune responses, and disease resistance of the grouper, *Epinephelus coioides*. *Fish & Shellfish Immunology.*, 29, 1053-1059.
- Comps, M., Menu, B., Breuil, G. and Bonami, J. R. (1990). Viral infection associated with rotifer mortalities in mass culture. *Aquaculture.*, 93(1), 1-7.

- David, A. B. (2003). *Status of marine aquaculture in relation to live prey: past, present and future*. In: Josianne, G. S and Lesley, A. M. (Eds.), *Live feeds in marine aquaculture*. Blackwell publishing, UK, pp. 1-16.
- Defoirdt, T., Boon, N., Bossier, P. and Verstraete, W. (2004). Disruption of bacterial quorum sensing: an unexplored strategy to fight infections in aquaculture. *Aquaculture.*, 240, 69–88.
- Defoirdt, T., Bossier, P., Sorgeloos, P. and Verstraete, W. (2005). The impact of mutations in the quorum sensing systems of *Aeromonas hydrophila*, *Vibrio anguillarum* and *Vibrio harveyi* on their virulence towards gnotobiotically cultured *Artemia franciscana*. *Environmental Microbiology.*, 7(8), 1239–1247.
- Defoirdt, T., Halet, D., Vervaeren, H., Boon, N., Van de Wiele, T., Sorgeloos, P., Bossier, P. and Verstraete, W. (2007). The bacterial storage compound poly- β -hydroxybutyrate protects *Artemia franciscana* from pathogenic *Vibrio campbellii*. *Environmental Microbiology.*, 9(2), 445–452.
- Defoirdt, T., Sorgeloos, P. and Bossier, P. (2011a). Alternatives to antibiotics for the control of bacterial disease in aquaculture. *Current Opinion in Microbiology.*, 14, 251–258.
- Defoirdt, T., Thanh, L. D., Delsen, B. V., De Schryver, P., Sorgeloos, P., Boon, N. and Bossier, P. (2011b). N-acylhomoserine lactone-degrading *Bacillus* strains isolated from aquaculture animals. *Aquaculture.*, 311, 258–260.
- Defoirdt, T., Pande, G. S. J., Baruah, K. and Bossier, P. (2013). The Apparent Quorum-Sensing Inhibitory Activity of Pyrogallol Is a Side Effect of Peroxide Production. *Antimicrobial agents and chemotherapy.*, 57(6), 2870–2873.
- Dhara, L. and Tripathi, A. (2013). Antimicrobial activity of eugenol and cinnamaldehyde against extended spectrum beta lactamase producing enterobacteriaceae by in vitro and molecular docking analysis. *European Journal of Integrative Medicine.*, 5(6), 527–536.
- Dhert, P., Rombaut, G., Suantika, G. and Sorgeloos, P. (2001). Advancement of rotifer culture and manipulation techniques in Europe. *Aquaculture.*, 200(1), 129-146.
- Dong, Y. H., Xu, J. L., Li, X. Z. and Zhang, L. H. (2000). AiiA, an enzyme that inactivates the acylhomoserine lactone quorum-sensing signal and attenuates the virulence of *Erwinia carotovora*. *Proceedings of the National Academy of Sciences.*, 97(7), 3526-3531.

- Drillet, G., Frouël, S., Sichlau, M.H., Jepsen, P.M., Højgaard, J.K., Joarder, A.K. and Hansen, B.W. (2011). Status and recommendations on marine copepod cultivation for use as live feed. *Aquaculture.*, 315, 155–166.
- Duray, M.N. (1994). Daily rates of ingestion on rotifers and *Artemia* nauplii by laboratory-reared grouper larvae, *Epinephelus suillus*. *Philipp. Sci.*, 31: 32-41.
- FAO. (2010). *Epinephelus coioides*. Cultured Aquatic Species Information Programme. Rome, FAO. 2010.
- Flagan, S., Ching, W. K. and Leadbetter, J. R. (2003). Arthrobacter strain VAI-A utilizes acyl-homoserine lactone inactivation products and stimulates quorum signal biodegradation by *Variovorax paradoxus*. *Applied and environmental microbiology.*, 69(2), 909-916.
- Gamboa-Delgado, J. and Le Vay, L. (2009). *Artemia* replacement in co-feeding regimes for mysis and postlarval stages of *Litopenaeus vannamei*: Nutritional contribution of inert diets to tissue growth as indicated by natural carbon stable isotopes. *Aquaculture.*, 297(1), 128-135.
- Gomez-Gil, B., Herrera-Vega, M. A., Abreu-Grobois, F. A. and Roque, A. (1998). Bioencapsulation of Two Different *Vibrio* Species in Nauplii of the Brine Shrimp (*Artemia franciscana*). *Applied and environmental microbiology.*, 64(6), 2318-2322.
- Harikrishnan, R., Balasundaram, C. and Heo, M. (2011). Fish health aspects in grouper aquaculture. *Aquaculture.*, 320, 1–21.
- Henke, J.M. and Bassler, B.L. (2004). Three parallel quorum sensing systems regulate gene expression in *V. harveyi*. *J. Bacteriol.*, 186, 6902-6914.
- Hense, B. A., Kuttler, C., Müller, J., Rothballer, M., Hartmann, A. and Kreft, J. U. (2007). Does efficiency sensing unify diffusion and quorum sensing? *Nature Reviews Microbiology.*, 5(3), 230-239.
- Høj, L., Bourne, D. G. and Hall, M. R. (2009). Localization, abundance and community structure of bacteria associated with *Artemia*: Effects of nauplii enrichment and antimicrobial treatment. *Aquaculture.*, 293(3), 278-285.
- Hu, Y. H., Deng, T., Sun, B. G. and Sun, L. (2012). Development and efficacy of an attenuated *Vibrio harveyi* vaccine candidate with cross protectivity against *Vibrio alginolyticus*. *Fish & Shellfish Immunology.*, 32(6), 1155-1161.

- Huang, Z., Tang, J., Fu, Y., Dong, C., Zhong, J.F. and He, J. (2012). Immunological evaluation of *Vibrio alginolyticus*, *Vibrio harveyi*, *Vibrio vulnificus* and infectious spleen and kidney necrosis virus (ISKNV) combined-vaccine efficacy in *Epinephelus coioides*. *Veterinary Immunology and Immunopathology.*, 150, 61– 68.
- Intriago, P. and Jones, D. A. (1993). Bacteria as food for *Artemia*. *Aquaculture.*, 113(1), 115-127.
- Jha, B., Kavita, K., Westphal, J., Hartmann, A. and Schmitt-Kopplin, P. (2013). Quorum Sensing Inhibition by *Asparagopsis taxiformis*, a Marine Macro Alga: Separation of the Compound that Interrupts Bacterial Communication. *Marine drugs.*, 11(1), 253-265.
- Johnson, R. B., Cook, M. A., Nicklason, P. M. and Rust, M. B. (2009). Determination of apparent protein digestibility of live *Artemia* and a microparticulate diet in 8-week-old Atlantic cod *Gadus morhua* larvae. *Aquaculture.*, 288(3), 290-298.
- Kalia, V.C. (2013). Quorum sensing inhibitors: An overview. *Biotechnology Advances.*, 31, 224–245.
- Kohno, H., Ordonio-Aguilar, R.S., Ohno, A. and Taki, Y. (1997). Why is grouper larval rearing difficult?: an approach from the development of the feeding apparatus in early stage larvae of the grouper, *Epinephelus coioides*. *Ichthyol. Res.*, 44 (3), 267-274.
- König, H., Claus, H. and Varma, A. (2010). *Prokaryotic cell wall compounds*. Retrieved from <http://link.springer.com/book/10.1007/978-3-642-05062-6>
- Lee, S. J., Park, S. Y., Lee, J. J., Yum, D. Y., Koo, B. T. and Lee, J. K. (2002). Genes encoding the N-acyl homoserine lactone-degrading enzyme are widespread in many subspecies of *Bacillus thuringiensis*. *Applied and Environmental Microbiology.*, 68(8), 3919-3924.
- Lenz, D.H., Mok, K.C., Lilley, B.N., Kulkarni, R.V., Wingreen, N.S. and Bassler, B.L. (2004). The small RNA chaperone Hfq and multiple small RNAs control quorum sensing in *Vibrio harveyi* and *Vibrio cholerae*. *Cell.*, 118, 69–82.
- Lim, L. C., Dhert, P. and Sorgeloos, P. (2003). Recent developments in the application of live feeds in the freshwater ornamental fish culture. *Aquaculture.*, 227(1), 319-331.
- Lin, Y. H., Xu, J. L., Hu, J., Wang, L. H., Ong, S. L., Leadbetter, J. R. and Zhang, L. H. (2003). Acyl-homoserine lactone acylase from *Ralstonia* strain XJ12B represents a novel and potent class of quorum-quenching enzymes. *Molecular microbiology.*, 47(3), 849-860.

- Liu, C., Chiu, C., Wang, S. and Cheng, W. (2012). Dietary administration of the probiotic, *Bacillus subtilis* E20, enhances the growth, innate immune responses, and disease resistance of the grouper, *Epinephelus coioides*. *Fish & Shellfish Immunology.*, 33, 699-706.
- Makridis, P. and Vadstein, O. (1999). Food size selectivity of *Artemia franciscana* at three developmental stages. *Journal of plankton research.*, 21(11), 2191-2201.
- Mai, M. G., Engrola, S., Morais, S., Portella, M. C., Verani, J. R., Dinis, M. T. and Conceição, L. E. (2009). Co-feeding of live feed and inert diet from first-feeding affects *Artemia* lipid digestibility and retention in Senegalese sole (*Solea senegalensis*) larvae. *Aquaculture.*, 296(3), 284-291.
- Marques, A., Dhont, J., Sorgeloos, P. and Bossier, P. (2004). Evaluation of different yeast cell wall mutants and microalgae strains as feed for gnotobiotically grown brine shrimp *Artemia franciscana*. *Journal of experimental marine biology and ecology.*, 312(1), 115-136.
- Marques, A., Thanh, T. H., Sorgeloos, P. and Bossier, P. (2006). Use of microalgae and bacteria to enhance protection of gnotobiotic *Artemia* against different pathogens. *Aquaculture.*, 258(1), 116-126.
- McClellan, K.H., Winson, M.K., Fish, L., Taylor, A., Chhabra, S.R., Camara, M., ...Williams, P. (1997). Quorum sensing and *Chromobacterium violaceum*: exploitation of violacein production and inhibition for the detection of Nacylhomoserine lactones. *Microbiology.*, 143, 3703–3711.
- Natrah, F.M.I. (2011a). *Role of bacterial quorum sensing and micro-algae in fish and crustacean larviculture*. PhD thesis, Ghent University, Ghent, Belgium.
- Natrah, F.M.I., Kenmegne, M. M., Wiyoto, W., Sorgeloos, P., Bossier, P. and Defoirdt, T. (2011b). Effects of micro-algae commonly used in aquaculture on acyl-homoserine lactone quorum sensing. *Aquaculture.*, 317, 53–57.
- Natrah, F.M.I., Defoirdt, T., Sorgeloos, P. and Bossier, P. (2011c). Disruption of bacterial cell-to-cell communication by marine organisms and its relevance to aquaculture. *Marine Biotechnology.*, 13, 109-126.
- Natrah, F. M. I., Alam, M. I., Pawar, S., Harzevili, A. S., Nevejan, N., Boon and Defoirdt, T. (2012). The impact of quorum sensing on the virulence of *Aeromonas hydrophila* and *Aeromonas salmonicida* towards burbot (*Lota lota* L.) larvae. *Veterinary microbiology.*, 159(1), 77-82.
- Orozco-Medina, C., Maeda-Martínez, A. M. and López-Cortés, A. (2002). Effect of aerobic Gram-positive heterotrophic bacteria associated with *Artemia franciscana* cysts on the survival and development of its larvae. *Aquaculture.*, 213(1), 15-29.

- Pande, G. S. J., Scheie, A. A., Benneche, T., Wille, M., Sorgeloos, P., Bossier, P. and Defoirdt, T. (2013). Quorum sensing-disrupting compounds protect larvae of the giant freshwater prawn *Macrobrachium rosenbergii* from *Vibrio harveyi* infection. *Aquaculture.*, 406–407, 121–124.
- Pintado, J., Pérez-Lorenzo, M., Luna-González, A., Sotelo, C. G., Prol, M. J., and Planas, M. (2010). Monitoring of the bioencapsulation of a probiotic *Phaeobacter* strain in the rotifer *Brachionus plicatilis* using denaturing gradient gel electrophoresis. *Aquaculture.*, 302(3), 182-194.
- Planas, M., Vázquez, J. A., Marqués, J., Pérez-Lomba, R., González, M. P. and Murado, M. (2004). Enhancement of rotifer (*Brachionus plicatilis*) growth by using terrestrial lactic acid bacteria. *Aquaculture.*, 240(1), 313-329.
- Planas, M., Pérez-Lorenzo, M., Antonio Vázquez, J. and Pintado, J. (2005). A model for experimental infections with *Vibrio* (*Listonella*) *anguillarum* in first feeding turbot (*Scophthalmus maximus* L.) larvae under hatchery conditions. *Aquaculture.*, 250(1), 232-243
- Rajkumar, M. (2006). Suitability of the copepod, *Acartia clausi* as a live feed for Seabass larvae (*Lates calcarifer* Bloch): Compared to traditional live-food organisms with special emphasis on the nutritional value. *Aquaculture.*, 261(2), 649-658.
- Romero, M., Martin-Cuadrado, A.B., Roca-Rivada, A., Cabello, A.M. and Otero, A. (2011). Quorum quenching in cultivable bacteria from dense marine coastal microbial communities. *FEMS Microbiol Ecol.*, 75(2), 205-17.
- Saida, H., Maekawa, T., Satake, T., Higashi, Y. and Seki, H. (2000). Gram stain index of a natural bacterial community at a nutrient gradient in the freshwater environment. *Environmental Pollution.*, 109(2), 293-301.
- Son, V.M., Chang, C., Wu, M., Guu, Y., Chiu, C. and Cheng, W. (2009). Dietary administration of the probiotic, *Lactobacillus plantarum*, enhanced the growth, innate immune responses, and disease resistance of the grouper *Epinephelus coioides*. *Fish & Shellfish Immunology.*, 26, 691–698.
- Soto-Rodriguez, S. A., Roque, A., Lizarraga-Partida, M. L., Guerra-Flores, A. L. and Gomez-Gil, B. (2003). Virulence of luminous vibrios to *Artemia franciscana* nauplii. *Diseases of aquatic organisms.*, 53(3), 231-240.
- Sun, Y., Yang, H., Huang, K., Ye, J. and Zhang, C. (2013). Application of autochthonous *Bacillus* bioencapsulated in copepod to grouper *Epinephelus coioides* larvae. *Aquaculture.*, 392-395, 44–50.

- Tinh, N.T., Asanka Gunasekara, R.A., Boon, N., Dierckens, K., Sorgeloos, P. and Bossier, P. (2007). N-acylhomoserine lactone degrading microbial enrichment cultures isolated from *Penaeus vannamei* shrimp gut and their probiotic properties in *Brachionus plicatilis*. *FEMS Microbiol Ecol.*, 62, 45-53.
- Tinh, N. T. N., Yen, V. H. N., Dierckens, K., Sorgeloos, P. and Bossier, P. (2008). An acyl homoserine lactone-degrading microbial community improves the survival of first-feeding turbot larvae *Scophthalmus maximus*. *Aquaculture.*, 285(1), 56-62.
- Velho-Pereira, S. and Kamat, N. M. (2011). Antimicrobial Screening of Actinobacteria using a Modified Cross-Streak Method. *Indian J. Pharm. Sci. Mar-Apr.*, 73(2): 223–228.
- Verdonck, L., Grisez, L., Sweetman, E., Minkoff, G., Sorgeloos, P., Ollevier, F. and Swings, J. (1997). Vibrios associated with routine productions of *Brachionus plicatilis*. *Aquaculture.*, 149(3), 203-214.
- Villamil, L., Figueras, A., Planas, M. and Novoa, B. (2003). Control of *Vibrio alginolyticus* in *Artemia* culture by treatment with bacterial probiotics. *Aquaculture.*, 219, 43–56.
- Vu, M. T., Jepsen, P. M. and Hansen, B. W. (2013). A comprehensive and precise quantification of the calanoid copepod *Acartia tonsa* (Dana) for intensive live feed cultures using an automated ZooImage system. *Aquaculture.*
- Zhao, J., Xu, X., Li, X. and Wang, J. (2014). Promotion of Sn on the Pd/AC catalyst for the selective hydrogenation of cinnamaldehyde. *Catalysis Communications.*, 43, 102-106.