

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF Momordica cochinchinensis SPRENG, FRUIT ON THE PROLIFERATION AND ANGIOGENESIS BIOMARKERS OF HUMAN RETINAL PIGMENT EPITHELIAL CELLS UNDER HIGH GLUCOSE CONDITIONS

ALI ABDULQADER MAHDI MAHDI

FPSK(m) 2019 6



EFFECTS OF Momordica cochinchinensis SPRENG, FRUIT ON THE PROLIFERATION AND ANGIOGENESIS BIOMARKERS OF HUMAN RETINAL PIGMENT EPITHELIAL CELLS UNDER HIGH GLUCOSE CONDITIONS

By

ALI ABDULQADER MAHDI MAHDI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF Momordica cochinchinensis SPRENG, FRUIT ON THE PROLIFERATION AND ANGIOGENESIS BIOMARKERS OF HUMAN RETINAL PIGMENT EPITHELIAL CELLS UNDER HIGH GLUCOSE CONDITIONS

Ву

ALI ABDULQADER MAHDI MAHDI

January 2019

Chairman : Professor Amin bin Ismail, PhD Faculty : Medicine and Health Sciences

Tropical fruits have been reported to contain health-promoting compounds for human benefits. Fruit of *Momordica cochinchinensis* (M. cochinchinensis) is one of these fruits that believed to be rich source of bioactive constitutes. However, the studies on its phytochemical composition and biological activities against hyperglycaemia-related eye disease were found to be limited. Thus, this study attempted to investigate the phytochemical composition, antioxidant activities and the effects of M. cochinchinensis parts extracts grown in Malaysia on cell viability and angiogenic activity of human retinal pigment epithelial (ARPE-19) cells under high glucose (HG) conditions. UV-Vis was used for the estimation of phenolics, flavonoids, and carotenoids contents. Lycopene and β-carotene were quantified using a HPLC technique. LC-MS/MS was used for the identification of antioxidant compounds. DPPH and FRAP assays were employed to evaluate the antioxidant capacities. MTT assay, morphological observations and Trypan blue dye were employed for the cell viability evaluation. Enzyme linked immunosorbent-based assay was performed the evaluate the effect of the fruit extracts on the reactive oxygen species (ROS), vascular endothelial growth factor (VEGF) and pigment epithelium-derived factor (PEDF) secretions. The results showed carotenoids, phenolics and flavonoids were present in all fruit parts. Comparatively, the fruit aril showed the highest carotenoids and phenolics content ($1106 \pm 2.1 \,\mu\text{g/g}$, $308 \pm 2.7 \,\mu\text{g/g}$) respectively, whereas the peel showed the highest flavonoid content (381 \pm 2.2 μ g/g). Lycopene and β-carotene were found to be significantly high (p < 0.05) in aril (579.3 ± 22.7 and 621 \pm 35 μ g/g DW) and relatively high in peel and pulp. Furthermore, rutin, quercetin, linolenic acid and cinnamic acid were identified in peel, seed, pulp and aril. The antioxidant capacity evaluations revealed that, aril possessed the highest scavenging activity $IC_{50} = 865 \mu g/mL$, while the peel appeared to possess the highest ferric reducing power of 140 µmol FeSO₄/µg. Cell culture experiments demonstrated HG condition at (30 mmol/L) condition increased the proliferation of ARPE-19 cell proliferation, ROS

and VEGF secretions compared to low glucose (LG) at (5.5 mmol/L). While the exposure of ARPE-19 cells in HG conditions to M. cochinchinensis led to inhibition of cell viability, induced morphological changes, decreased ROS and VEGF, and increased PEDF levels. M. cochinchinensis pulp, seed, and aril at 1 000 μg/mL showed significant inhibition activities $[(7.5 \pm 5.1)\%, (2.7 \pm 0.5)\%, (3.2 \pm 1.1)\%$, respectively] against HGinduced ARPE-19 cell viability. The findings also demonstrated that aril at 250 μg/mL significantly decreased ROS and VEGF levels [(40.6 ± 3.3) pg/mL, (107.4 ± 48.3) pg/mL, respectively] compared to ROS [(71.7 ± 2.9) pg/mL] and VEGF [(606.9 ± 81.1) pg/mL] in HG untreated cells. Moreover, 250 µg/mL of Gac peel dramatically increased PEDF level [(18.2 ± 0.3) ng/mL] compared to that in HG untreated cells [(0.48 ± 0.39) ng/mL]. The current results demonstrated that M. cochinchinensis fruit was found to be rich source of phytochemical compounds especially carotenoids and to possess antioxidant capacity. In addition, M. cochinchinensis fruit extracts reduced ARPE-19 cell viability, minimized ROS generations and showed angiogenic activities. Therefore, our findings open new insights towards the potentiality of utilizing M. cochinchinensis fruit as a source of antioxidant plant and against HG-related diabetic retinopathy disease.

KESAN BUAH Momordica cochinchinensis SPRENG. KE ATAS PROLIFERASI DAN BIOPENANDA ANGIOGENESIS BAGI SEL EPITELIAL PIGMEN RETINA MANUSIA DI BAWAH KEADAAN GLUKOSA TINGGI

Oleh

ALI ABDULQADER MAHDI MAHDI

Januari 2019

Pengerusi : Profesor Amin bin Ismail, PhD Fakulti : Perubatan dan Sains Kesihatan

Buah-buahan tropika telah dilaporkan mengandungi sebatian mempromosikan kesihatan bagi manfaat manusia. Buah Momordica cochinchinensis (M. cochinchinensis) merupakan antara spesies buah yang dikatakan kaya dengan pelbagai sumber bahan aktif. Walau bagaimanapun, kajian ke atas komposisi fitokimianya dan aktiviti biologikalnya terhadap penyakit mata berkaitan hiperglisemia adalah terhad. Oleh sebab itu, kajian ini dijalankan untuk menilai komposisi fitokimia, aktiviti antioksida dan kesan ekstrak bahagian M. cochinchinensis yang ditanam di Malaysia ke atas viabiliti sel dan aktiviti angiogeni sel epitelial manusia (ARPE-19) di bawah keadaan glukosa tinggi (HG). UV-Vis telah digunakan bagi menganggarkan kandungan fenolik, flavonoid, dan Penentuan kandungan likopen dan β-karotena telah dijalankan menggunakan teknik HPLC. LC-MS/MS telah digunakan untuk mengenal pasti sebatian antioksida. Asai DPPH dan FRAP telah digunakan untuk menilai kapasiti antioksida. Asai MTT, pemerhatian morfologikal dan pencelup biru Trypan telah digunakan bagi menilai viabiliti sel. Asai imunoserap terangkai enzim telah dijalankan bagi menilai kesan ekstrak buah ke atas spesies oksigen reaktif (ROS), faktor pertumbuhan endolial vaskular (VEGF) dan rembesan faktor perolehan epitelium pigmen (PEDF). Keputusan menunjukkan karotenoid, fenolik dan flavonoid wujud dalam semua bahagian buah. Secara bandingan, bahagian aril buah memperlihatkan kandungan karotenoid dan fenolik paling tinggi, iaitu (1106 \pm 2.1 µg/g dan 308 \pm 2.7 µg/g), manakala bahagian kulit memperlihatkan kandungan flavonoid paling tinggi, iaitu (381 \pm 2.2 μ g/g). Likopen dan β-karotena didapati tinggi (p< 0.05) dalam bahagian aril (579.3 ± 22.7 dan 621 ± 35 μg/g DW) dan secara relatif adalah tinggi dalam bahagian kulit dan pulpa. Di samping itu, rutin, asid kuersetin, asid linolenik dan asid sinamik telah dikenal pasti dalam kulit, biji, pulpa dan aril. Penilaian kapasiti antioksida memperlihatkan bahawa, aril mengandungi aktiviti hapus-sisa paling tinggi, iaitu IC₅₀ = 865 μg/mL, manakala kulit didapati mengandungi kuasa pengurangan ferik paling tinggi, iaitu 140 μmol FeSO₄/μg. Eksperimen kultur sel mempamerkan keadaan HG (30 mmol/L) dan meningkatkan

proliferasi sel ARPE-19, hapus-sisa ROS dan VEGF berbanding dengan glukosa rendah (LG) pada (5.5 mmol/L). Manakala pendedahan sel ARPE-19 dalam keadaan HG kepada M. cochinchinensis mengakibatkan perencatan viabiliti sel, mencetuskan perubahan morfologikal, mengurangkan ROS dan VEGF, dan meningkatkan tahap PEDF. Pulpa, biji dan aril M. cochinchinensis pada 1000 µg/mL menunjukkan aktiviti perencatan yang signifikan $[(7.5 \pm 5.1)\%, (2.7 \pm 0.5)\%, (3.2 \pm 1.1)\%]$ terhadap viabiliti sel ARPE-19 tercetus HG. Keputusan juga mempamerkan bahawa aril pada 250 µg/mL, secara signifikan mengurangkan tahap ROS dan VEGF [(40.6 ± 3.3) pg/mL, $(107.4 \pm$ 48.3) pg/mL] berbanding dengan ROS [(71.7 ± 2.9) pg/mL] dan VEGF [(606.9 ± 81.1) pg/mL] dalam sel tidak terawat HG. Tambahan pula, 250 µg/mL kulit M. cochinchinensis secara dramatik meningkatkan tahap PEDF [(18.2 ± 0.3) ng/mL] berbanding dengan dangan sel tidak terawat HG [(0.48 ± 0.39) ng/mL]. Keputusan kajian ini mempamerkan bahawa buah M. cochinchinensis merupakan sumber sebatian fitokimia yang kaya terutama karotenoid dan mengandungi kapasiti antioksida. Di samping itu, ekstrak buah M. cochinchinensis mengurangkan viabiliti sel ARPE-19, meminimumkan penjanaan ROS dan menunjukkan aktiviti angiogenik. Oleh itu, keputusan kajian ini, membuka penemuan baharu terhadap potensi penggunaan buah M. cochinchinensis sebagai sumber antioksida dan terhadap penyakit retinopati diabetik berkaitan HG.

ACKNOWLEDGEMENTS

I would love to express my sincere gratitude and thankful to the chairman of my supervisory committee Prof. Amin Ismail for his invaluable guidance and ideal leadership through the whole way of this research. I would also appreciate the guidance and supervision of Dr. Norhaizan Mohd Esa. In addition, many thanks to all the staffs and students in the laboratory of nutrition and dietetics department and other faculty departments for their friendly attitude and assistance. Bunch of thanks goes to Dr. Faisal Ali for the support and assistance from the beginning of this research until the end.

I would like to take this opportunity to gratefully thank my beloved mother and other family members for their kind support, encouragements, understandings and patience during all this journey. I would also like to offer my thanks to all my friends for their sustained support.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Amin Bin Ismail, PhD

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Norhaizan Mohd Esa, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:	Date:

Name and Matric No.: Ali Abdulgader Mahdi Mahdi/GS46163

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory	
Committee:	Professor Dr. Amin Bin Ismail
	P
Signature:	
Name of Member of Supervisory	
Committee:	Associate Professor Dr. Norhaizan Mohd Esa

TABLE OF CONTENTS

				Page
ABSTR	ACT			i
ABSTR	4 <i>K</i>			iii
ACKNO	OWLED	GEMEN	VTS	v
APPRO	VAL			vi
DECLA	RATIO	N		viii
LIST O	F TABI	LES		xiii
LIST O	F FIGU	RES		xiv
LIST O	F ABBI	REVIATI	IONS	xvi
CHAPT	ER			
1	INTR	ODUCTI	ION	1
	1.1	Backgr	ound of the study	1
	1.2		m statements	2
	1.3	Signifi	cance of the study	2 2 3
	1.4	Object		3
2			EREVIEW	4
	2.1		ications of DM	4
	2.2		d its classification	4
	2.3		evalence of DR	5
	2.4		thogenesis of DR	6
	2.5	Current treatments of DR and their drawbacks 7		
	2.6	Human retinal pigmented epithelial cells and its prominence in		
			ogression	8
	2.7		f bioactive compounds in the prevention DR	10
	2.8		noids and their potentiality in the prevention of DR	11
	2.9		hinchinensis Spreng	12
	2.10		noids in <i>M. cochinchinensis</i>	14
	2.11		ic and flavonoid compounds in M. cochinchinensis	15
	2.12		niology and traditional use of M. cochinchinensis	15
	2.13		idant activities of M. cochinchinensis	16
	2.14	Antica	ncer and other health benefits of <i>M. cochinchinensis</i>	16
3	MATI	FRIAIS	AND METHODS	19
3	3.1		als and chemicals	19
	3.1		Chemicals and reagents	19
			Fruit collection and preparation	19
		3.1.3	Cell culture	20
	3.2	Method		20
	5.4	3.2.1	Morphology and weight distributing of Malaysian	20
		5.4.1	M. cochinchinensis fruit	20
		3.2.2	Proximate analysis	20
		3.2.2	Extraction for the determination of total phenolics	20
		5.4.5	and flavonoids	20
			and navonoids	∠∪

		3.2.4	Determination of total phenolics	21
		3.2.5	Determination of total flavonoids	21
		3.2.6	Extraction and determination of total carotenoids	21
		3.2.7	Extraction and quantification of individual	
			carotenoids (β-carotene and lycopene) using high	
			performance liquid chromatography analysis	22
		3.2.8	Identification of chemical compounds using LC-MS	
			/MS	22
		3.2.9	Extraction for antioxidant and cell culture experiments	23
		3.2.10	DPPH free radical scavenging assay	23
		3.2.11	Ferric reducing antioxidant power assay	24
		3.2.12	Examination of cell proliferation under various	
			glucose conditions	24
		3.2.13	Effects of <i>M. cochinchinensis</i> on ARPE-19 cell proliferation	24
		3.2.14	Cell morphology examination	25
		3.2.15	Trypan blue dye assay for cell viability	25
		3.2.16	Measurement of reactive oxygen species level	26
		3.2.17		
			level	26
		3.2.18	Measurement of pigment epithelium-derived factor	
			level	27
	3.3	Statistic	cal and analysis	27
4	RESU	LTS		28
	4.1	Fruit m	orphology and its description	28
	4.2	Proxim	ate composition	29
	4.3	Yield e	xtract of <i>M. cochinchinensis</i> fruit parts	29
	4.4	Total ca	arotenoids, phenolics and flavonoids contents	29
	4.5	β-carote	ene and lycopene contents	30
	4.6	Identifi	cation of chemical compounds	31
	4.7	DPPH 1	free radical scavenging activities	33
	4.8	Ferric r	reducing antioxidant activities	33
	4.9	Cell via	ability under different glucose concentrations	34
	4.10	Effect of viability	of extracts from M. cochinchinensis on ARPE-19 cell	35
	4.11		of extracts from <i>M. cochinchinensis</i> on ARPE-19	33
	4.11		morphology	36
	4.12		tion of ARPE-19 viable cells by Trypan blue dye	38
	4.12		of extracts from <i>M. cochinchinensis</i> on ROS level	39
	4.13			39 40
	4.14	Effect	of extracts <i>M. cochinchinensis</i> on VEGF level	40
5	DISCU	USSION		42
	5.1	M. coch	hinchinensis fruit morphology	42
	5.2	Proxim	ate value of M. cochinchinensis fruit	42
	5.3	Phenoli	ics, flavonoids and carotenoids in M. cochinchinensis	
		fruit		42
	5.4	B-carote	ene and lycopene concentrations	44

	5.5	Identification of the chemical compounds among A	И.
		cochinchinensis parts	44
	5.6	Antioxidant activities of M. cochinchinensis fruit parts	45
	5.7	High glucose induced ARPE-19 cell proliferation	45
	5.8	Anti-proliferative of <i>M. cochinchinensis</i> fruit parts extract ar	nd
		morphological changes of ARPE-19 cells	46
	5.9	Anti-ROS activity of <i>M. cochinchinensis</i> fruit parts extracts	47
	5.10	Angiogenic activities of M. cochinchinensis fruit parts extrac	ets
			47
6	CONC	CLUSIONS, LIMITATIONS AND RECOMMENDATIONS	S 49
	6.1	Conclusions	49
	6.2	Limitations	49
	6.3	Recommendations	50
REF	FERENCE	S	51
APP	PENDICES	I married arrange	69
BIO	DATA OF	STUDENT	75
T TC	T OF DIID	LICATIONS	76

LIST OF TABLES

Table		Page
2.1	Prevalence of DR among Asian populations	5
2.2	Carotenoids distribution among M. cochinchinensis fruit parts	14
2.3	Biological activities of <i>M. cochinchinensis</i> fruit	17
4.1	Proximate composition of <i>M. cochinchinensis</i> fruit (g per 100 g)	29
4.2	Carotenoids, phenolics, and flavonoids contents in <i>M. cochinchinensis</i> fruit parts (peel, pulp, seed, and aril)	30
43	Chemical compounds in M. cochinchinensis fruit parts	32

LIST OF FIGURES

Figure		Page
2.1	Common complications of DM	4
2.2	Stages development of DR disease from no DR to PDR	5
2.3	The possible mechanisms underlying DR pathogenesis	7
2.4	Mammalian retina morphology	10
2.5	M. cochinchinensis fruit parts (1) Seed, (2) Pulp, (3) Aril, (4) Peel) and morphology adapted from	13
2.6	Biological activities and health benefits of M. cochinchinensis fruit	18
4.1	The morphology of <i>M. cochinchinensis</i> fruit Malaysian cultivar	28
4.2	Weight distribution of M. cochinchinensis fruit parts	29
4.3	Quantification of β -carotene and lycopene in M . $cochinchinensis$ fruit parts	31
4.4	Scavenging activity of M. cochinchinensis fruit parts	33
4.5	Reducing power capacities of M . cochinchinensis fruit parts and BHT as a positive control at different concentrations (150, 300, 600, 1200 μ g/ml)	34
4.6	ARPE-19 cell proliferation activity exposed to different concentrations of glucose (5.5, 30, 50, 70, 100 mmol/L) and incubated for 48 hr using MTT assay	34
4.7	Anti-proliferative activities of <i>M. cochinchinensis</i> parts extracts against ARPE-19 cell proliferation in high glucose (HG) 30 mmol/L conditions for 48 hr. ARPE-19 plated in 96-well plates and treated with different concentrations (31.2-1000 µg/mL) of fruit parts	35
4.8	Effects of <i>M. cochinchinensis</i> parts (peel, pulp, seed and aril) at different concentrations (62.5-1000 μ g/mL) on cellular morphology of ARPE-19 in high glucose conditions for 48 h	37
4.9	Trypan blue dye assay for cell viability of ARPE-19 cells treated with different concentrations of (31.2-1000 μg/mL) of <i>M. cochinchinensis</i> parts extracts for 48 hr in high glucose conditions	38

4.10	ROS level of untreated ARPE-19 cells in low glucose (5.5 mmol/L) and high glucose (30 mmol/L), and in ARPE-19 cells treated with different concentrations (50, 100 and 250 μ g/mL) of <i>M. cochinchinensis</i> parts extracts for 48 hr in high glucose conditions	39
4.11	VEGF level of untreated ARPE-19 in low glucose (5.5 mmol/L) and high glucose (30 mmol/L) and in ARPE-19 in high glucose (HG) conditions treated with different concentrations (50, 100 and 250 µg/mL) of <i>M. cochinchinensis</i> parts extracts for 48 hr	40
4.12	PEDF level of untreated ARPE-19 in low glucose (5.5 mmol/L) and high glucose (30 mmol/L) and in ARPE-19 in high glucose conditions treated with different concentrations (50, 100 and 250 μg/mL) of <i>M. cochinchinensis</i> parts extracts for 48 hr	41

LIST OF ABBREVIATIONS

DR Diabetic retinopathy

HG High glucose

LG Low glucose

AGEs Advance glycation end products

PKC Protein kinase C

PDR Proliferative diabetic retinopathy

VEGF Vascular Endothelial Growth Factor

IGF Insulin Growth Factor

PEDF Pigmented Epithelium Derived Factor

PlGF Placenta like Growth Factor

FGF Fibroblast Growth Factor

T1DM Type 1 Diabetes Mellitus

T2DM Type 2 Diabetes Mellitus

ARPE-19 Human Retinal Pigment Epithelial Cells

Momordica M. cochinchinensis

cochinchinensis

NPDR Non-Proliferative Diabetic Retinopathy

IDF International Diabetes Federation

NHMS National Health and Morbidity Survey

ELISA Enzyme Linked Immunosorbent Assay

AMD Age-related Macular Degradation

PDGF Platelet Derived Growth Factor

NGF Nerve Growth Factor

TNF-α Tumor Necrosis Factor α

HIF-1α Hypoxia-Inducible Factor 1

MMP Matrix Metalloproteinase

HPLC High Performance Liquid Chromatography

UV/Vis Ultra Violet Visible

LC-MS Liquid Chromatography-Mass Spectrum

FW Fresh Weight

DW Dry Weight

FRAP Ferric Reducing Antioxidant Power

MTT 3-(4,5-Dimethylthiazol-2-yl)-2,5-

diphenyltetrazolium bromide

DPPH 2,2-Diphenyl-1-picrylhydrazyl

TPTZ 2,4,6-tripyridyl-s-triazine

BHT Butylated Hydroxytoluene

W/V Weigh/Volume

V/V Volume/Volume

DMSO Dimethyl Sulphate

PBS Phosphate Buffer Saline

LG Low Glucose

HG High Glucose

ROS Reactive Oxygen Species

ATCC American Type Culture Collection

DMEM Dulbecco's Modified Eagle's Medium

FBS Fetal Bovine Serum

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Diabetes Mellitus (DM) is an epidemic concern and one of the most growing disease. Diabetes characterized by chronic hyperglycaemia (World Health Organization, 2016). Overtime diabetes can lead to several complications in both macro and micro vascular which finally lead to organs failure. Vascular diseases are the leading cause of morbidity and mortality among both type 1 diabetes mellitus (T1DM) and type 2 diabetes mellitus (T2DM) (Abdul-Ghani *et al.*, 2017; Kibel *et al.*, 2017; Kayama *et al.*, 2015). In particular, diabetic retinopathy (DR) is one of the most serious complication of diabetes. DR is an ocular disease and one of the leading cause of blindness worldwide (Boscia, 2010). It has established that chronic exposure to hyperglycaemia by the retina gives rise to cause many pathophysiological changes. These changes are including accumulation of advance glycation end products (AGEs), activation of protein kinase C (PKC), increase levels of sorbitol, reactive oxygen species (ROS) and inflammation factors, homeostasis of angiogenic markers (Zong *et al.*, 2011; Brownlee, 2005; Ahsan, 2015).

DR classified according to its severity into non-proliferative DR (NPDR) and proliferative DR (PDR) (Wilkinson et al., 2003; Al-Jarrah & Shatnawi, 2017). NPDR characterized by the presence of vascular abnormalities and PDR known by the excessive formation of abnormal blood vessels (Babapoor-Farrokhran et al., 2015). The formation of new blood vessels known as neovascularization and its tightly regulated by angiogenic markers (Bussolino et al., 1997). Angiogenesis is the process of formation new blood vessels from exciting vessels. Many growth factors are mainly involved in the regulation of the angiogenesis process including but not limited vascular endothelial growth factor (VEGF) and pigmented epithelium-derived factor (PEDF) (Paine et al., 2017; Al Kahtani et al., 2017; Behl & Kotwani, 2015).

Natural sources (plants, fruits and vegetables) are rich sources of phytochemicals and bioactive compounds such as; carotenoids and polyphenols (Singh *et al.*, 2013). These constitutes have gained attention due their pharmaceutical properties such as; antidiabetic, antioxidant, cancer chemoprevention, anti-inflammatory and angiogenic activities (Kuppusamy *et al.*, 2014; Kaur *et al.*, 2017; Cicero *et al.*, 2015). Studies have shown that, high levels of phytochemicals and carotenoids have positively correlated with reverse DR progression (Singh *et al.*, 2013). However, the information regarding tropical-underutilized fruit and its roles in the prevention of DR were found to be very limited. Among all the tropical fruits, *Momordica cochinchinensis* Spreng fruit (*M. cochinchinensis*) was claimed to be an exceptional source of phytochemicals especialy carotenoids (Aoki *et al.*, 2002; Ishida *et al.*, 2004; Kubola & Siriamornpun, 2011). This fruit commonly known as Gac in Vietnam and found to be grown in tropical area such as; Thai, Malaysia and Laos. Traditionally, *M. cochinchinensis* fruit used in folk and

ancestral medicine. However, phytochemical composition and biological activities of *M. cochinchinensis* fruit have not well illustrated, also no studies have found on *M. cochinchinensis* fruit grown in Malaysia. Extraction process was found to play a key role in the recovering of bioactive compounds of plant extracts. Ethanol is commonly used in the extraction process due to its nontoxic, availability and ability to extract polar and non-polar bioactive compounds (Chuyen *et al.*, 2017).

1.2 Problem statements

DR is one of the leading cause of loss vision. Reports showed that, one third of diabetes patients may develop signs of DR (Lee *et al.*, 2015). Thus, continuously increasing level of diabetes will be accompanied with increasing rates of DR to become a series burden for the community that affected their life style and currency. Globally, 126.6 million were living under the risk of DR in 2010 (Zheng *et al.*, 2012). Particularly, based on the report of Malaysian National Health and Morbidity Survey (NHMS) diabetes trend in Malaysia has been elevated through the last two decades 6.3 % in 1986 to 17.5 % in 2015 (Hussein *et al.*, 2015; Yen *et al.*, 2017; Tee & Yap, 2017).

Currently, DR treatment strategies are included VEGF inhibitors drugs (anti-VEGF), laser photocoagulation and vitreous surgery (Duh *et al.*, 2017; El-Asrar *et al.*, 2009; Joussen & Joeres, 2007; Parikh *et al.*, 2017). In spite of the effective role of the mentioned patterns to cure DR, these patterns are extremely expensive and must be performed by specialist and experts (Dakin *et al.*, 2014). In addition, these strategies might have several side effects. For example, laser photocoagulation considers as a destructive tool that do not work to recover the lose vision and cannot be fully safe (Selvaraj *et al.*, 2017; Stitt *et al.*, 2016). Moreover, these treatments may lead to some undesired results such as; retinal damage, retinal detachment, vitreous haemorrhage, endophthalmitis, retinal tears, and intraocular haemorrhage (Simó & Hernandez, 2008; Austeng *et al.*, 2016; Ventrice *et al.*, 2013). Furthermore, Dr is a complex and multi factorial disease which need for new and novel treatments that able to have multi-target activity. Thus, finding new strategies that might be available, cost-effective and safer to treat/prevent or slow the progression of DR are highly required.

1.3 Significance of the study

The role of plants and their bioactive compounds in the prevention of several diseases have been established as alternative potential strategy in folk medicine. Out of the plants, *M. cochinchinensis* fruits were found to be extremely high source of phytochemicals especially carotenoids. Carotenoids and other phytochemicals have known about their abilities to improve overall health vision, safety and to possess multi-target activity which might potentially be a good alternative treatment compare to DR treatments that mostly work on one target such as anti-VEGF injections that only work to reduce VEGF secretions in addition to its risk to cause retinal detachment. Retinal pigmented epithelial (RPE) cells were found to play a crucial role in the progression of DR due to its

anatomical position and its multi functions. According to the literature, there have been no studies investigated the role of *M. cochinchinensis* fruit extracts on hyperglycaemia-related DR disease using RPE cells or other retina cells as a model for DR. The current study is the first to investigate the effect of *M. cochinchinensis* extracts on human retinal pigment epithelial (ARPE-19) under high glucose conditions. Thus, this study might open new insights into the potentiality of using *M. cochinchinensis* fruit as alternative medicine in ethno-pharmacology and biomedicine applications. More specifically, the data of this research can be exploiting to formulate a new dietary supplement or nutraceuticals from natural sources as a remedy to prevent/treat or delay DR.

Furthermore, the majority of the previous studies have only given attention to the aril part of fruit with insignificant concern about other fruit parts which may contribute to waste issues. Therefore, the present study was aimed to study all *M. cochinchinensis* fruit parts which may encourage researches to consider all *M. cochinchinensis* parts as a significant sources of phytochemicals and thus lead to minimize disposal and waste problems. To the best of our knowledge, this study is the first to study *M. cochinchinensis* fruit grown in Malaysia. Thus, the findings of this study might appeal attention towards to study and utilize *M. cochinchinensis* fruit.

1.4 Objectives

The main objective of this research was to study the effect of *M. cochinchinensis* parts extracts on the antioxidant, proliferation and angiogenesis process using human ARPE-19 cell line as a model for DR in high glucose condition. This objective included few other specific objectives that were the following:

- 1. To estimate total carotenoid, phenolic and flavonoid contents, and to quantify β-carotene and lycopene concentrations in the fruit part.
- 2. To identify unknown chemical compounds and to evaluate the antioxidant activities of the fruit parts.
- 3. To assess the effect of the fruit parts extracts on ARPE-19 cell proliferation under high glucose conditions.
- 4. To evaluate the effect of the fruit parts on the secretion of reactive oxygen species of ARPE-19 cells under high glucose conditions.
- 5. To evaluate the effect of the fruit parts extracts on angiogenesis (VEGF and PEDF) markers of ARPE-19 cells under high glucose conditions.

REFERENCES

- Addoor, K. R., Bhandary, S. V., Khanna, R., Rao, L. G., Lingam, K. D., Binu, V. S., Shivaji, S., & Nandannaver, M. (2011). Assessment of awareness of diabetic retinopathy among the diabetics attending the peripheral diabetic clinics in melaka, malaysia. Med J Malaysia 66: 48-52.
- Agarwal, S., & Rao, A. V. (2000). Tomato lycopene and its role in human health and chronic diseases. Canadian Medical Association Journal, 163, 739-744.
- Ahsan, H. (2015). Diabetic retinopathy–biomolecules and multiple pathophysiology. Diabetes & Metabolic Syndrome: Clinical Research & Reviews 9: 51-54.
- Akeem, A., Mohamed, K. B., Asmawi, M. Z., & Sofiman, O. A. (2011). Mutagenic and antimutagenic potentials of fruit juices of five medicinal plants in Allium cepa L.: Possible influence of DPPH free radical scavengers. African Journal of Biotechnology, 10, 10248-10257.
- Al Kahtani, E., Xu, Z., Al Rashaed, S., Wu, L., Mahale, A., Tian, J., Abboud, B. E., Ghazi, G. N., Kozaki, I., Gupta, V., Arevalo, F. J., & Arevalo, J. F. (2017). Vitreous levels of placental growth factor correlate with activity of proliferative diabetic retinopathy and are not influenced by bevacizumab treatment. Eye 31: 529-536.Behl
- Ali, F., Ismail, A., Esa, N. M., & Pei, C. (2016). Cocoa polyphenols treatment ameliorates visceral obesity by reduction lipogenesis and promoting fatty acid oxidation genes in obese rats through interfering with AMPK pathway. European Journal of Lipid Science and Technology 118: 564-575.
- Ali, F., Ranneh, Y., Ismail, A., & Esa, N. M. (2015). Identification of phenolic compounds in polyphenols-rich extract of Malaysian cocoa powder using the HPLC-UV-ESI—MS/MS and probing their antioxidant properties. Journal of Food Science and Technology, 52, 2103-2111.
- Al-Jarrah, M. A., & Shatnawi, H. (2017). Non-proliferative diabetic retinopathy symptoms detection and classification using neural network. Journal of medical engineering & technology 41: 498-505.
- Alothman, M., Bhat, R., & Karim, A. A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. Food Chemistry, 115, 785-788.
- Al-Shabrawey, M., & Smith, S. (2010). Prediction of diabetic retinopathy: role of oxidative stress and relevance of apoptotic biomarkers. EPMA Journal, 1, 56-72.

- Amchova, P., Kotolova, H., & Ruda-Kucerova, J. (2015). Health safety issues of synthetic food colorants. Regulatory Toxicology and Pharmacology, 73, 914-922.
- Antonetti, D. A., Barber, A. J., Bronson, S. K., Freeman, W. M., Gardner, T. W., Jefferson, L. S., ... & Norbury, C. C. (2006). Diabetic retinopathy: seeing beyond glucose-induced microvascular disease. Diabetes, 55, 2401-2411.
- AOAC International (2000). Official methods of analysis of AOAC International. Maryland, USA: AOAC International.
- Aoki, H., Kieu, N. T. M., Kuze, N., Tomisaka, K., & Chuyen, N. V. (2002). Carotenoid pigments in GAC fruit (Momordica cochinchinensis SPRENG). Bioscience, biotechnology, and biochemistry 66: 2479-2482.
- Arnal, E., Johnsen-Soriano, S., Lopez-Malo, D., Perez-Pastor, G. M., Vidal-Gil, L., Morillas, N., Sancho-Pelluz, J., Romero, F.J., & Barcia, J. M. (2016). Docosahexaenoic Acid Protects against High Glucose-Induced Oxidative Stress in Human Retinal Pigment Epithelial Cells. Reactive Oxygen Species 2: 298-307.
- Ashwaq, A. A. S., Al-Qubaisi, M. S., Rasedee, A., Abdul, A. B., Taufiq-Yap, Y. H., & Yeap, S. K. (2016). Inducing G2/M Cell Cycle Arrest and Apoptosis through Generation Reactive Oxygen Species (ROS)-Mediated Mitochondria Pathway in HT-29 Cells by Dentatin (DEN) and Dentatin Incorporated in Hydroxypropyl-β-Cyclodextrin (DEN-HPβCD). International journal of molecular sciences 17: 1653.
- Austeng, D., Morken, T. S., Bolme, S., Follestad, T., & Halsteinli, V. (2016). Nurse-administered intravitreal injections of anti-VEGF: study protocol for noninferiority randomized controlled trial of safety, cost and patient satisfaction. BMC ophthalmology 16: 169.
- Babapoor-Farrokhran, S., Jee, K., Puchner, B., Hassan, S. J., Xin, X., Rodrigues, M., Kashiwabuchi, F., Ma, T., Hu, K., Deshpande, M., Daoud, Y., Solomon, S., Wenick, A., Lutty, G. A., Semenza, G. L., Montaner, S., & Sdhi, A. (2015). Angiopoietin-like 4 is a potent angiogenic factor and a novel therapeutic target for patients with proliferative diabetic retinopathy. Proceedings of the National Academy of Sciences 112: E3030-E3039.
- Bakar, M. F. A., Ahmad, N. E., Karim, F. A., & Saib, S. (2014). Phytochemicals and antioxidative properties of Borneo indigenous liposu (Baccaurea lanceolata) and tampoi (Baccaurea macrocarpa) fruits. Antioxidants, 3, 516-525.
- Bakar, M. F. A., Mohamed, M., Rahmat, A., & Fry, J. (2009). Phytochemicals and antioxidant activity of different parts of bambangan (Mangifera pajang) and tarap (Artocarpus odoratissimus). Food chemistry, 113, 479-483.

- Barot, M., Gokulgandhi, M. R., Patel, S., & Mitra, A. K. (2013). Microvascular complications and diabetic retinopathy: recent advances and future implications. Future medicinal chemistry, 5, 301-314.
- Behl, T., & Kotwani, A. (2015). Exploring the various aspects of the pathological role of vascular endothelial growth factor (VEGF) in diabetic retinopathy. Pharmacological research 99: 137-148.
- Behl, T., Kaur, I., & Kotwani, A. (2016). Implication of oxidative stress in progression of diabetic retinopathy. Survey of ophthalmology, 61, 187-196.
- Benzie, I. F., & Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. Analytical biochemistry, 239, 70-76.
- Bharathi, L. K., Singh, H. S., Shivashankar, S., Ganeshamurthy, A. N., & Sureshkumar, P. (2014). Assay of nutritional composition and antioxidant activity of three dioecious Momordica species of South East Asia. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences 84: 31-36.
- Boscia, F. (2010). Current approaches to the management of diabetic retinopathy and diabetic macular oedema. Drugs 70: 2171-2200.
- Brazionis, L., Rowley, K., Itsiopoulos, C., & O'Dea, K. (2008). Plasma carotenoids and diabetic retinopathy. British journal of nutrition 101: 270-277.
- Bringmann, A., & Wiedemann, P. (2012). Müller glial cells in retinal disease. Ophthalmologica, 227(1), 1-19.
- Britton G, Liaaen-Jensen S, Pfander H SpringerLink. Nutrition and health. Basel, London: Birkhauser; 2009.
- Brownlee, M. (2001). Biochemistry and molecular cell biology of diabetic complications. Nature 414: 813-820.
- Brownlee, M. (2005). The pathobiology of diabetic complications. Diabetes 54: 1615-1625.
- Bussolino, F., Mantovani, A., & Persico, G. (1997). Molecular mechanisms of blood vessel formation. Trends in biochemical sciences 22: 251-254.
- Calderon, G. D., Juarez, O. H., Hernandez, G. E., Punzo, S. M., & De la Cruz, Z. D. (2017). Oxidative stress and diabetic retinopathy: development and treatment. Eye, 31, 1122.
- Capitão, M., & Soares, R. (2016). Angiogenesis and inflammation crosstalk in diabetic retinopathy. Journal of cellular biochemistry 117: 2443-2453.

- Caprnda, M., Kubatka, P., Saxena, S., Valaskova, J., Stefanickova, J., Kobyliak, N., Zulli, A., & Kruzliak, P. (2017). The Impact of Hyperglycemia on VEGF Secretion in Retinal Endothelial Cells. Folia medica 59: 183-189.
- Chen, H. Y., Huang, S. M., Yang, C. M., & Hu, M. L. (2012a). Diverse effects of β-carotene on secretion and expression of VEGF in human hepatocarcinoma and prostate tumor cells. Molecules, 17(4), 3981-3988.
- Chen, M. L., Lin, Y. H., Yang, C. M., & Hu, M. L. (2012b). Lycopene inhibits angiogenesis both in vitro and in vivo by inhibiting MMP□2/uPA system through VEGFR2□mediated PI3K□Akt and ERK/p38 signaling pathways. Molecular nutrition & food research, 56, 889-899.
- Chen, P., Zhang, W., Wang, X., Zhao, K., Negi, D. S., Zhuo, L., ... & Zhang, X. (2015). Lycopene and risk of prostate cancer: a systematic review and meta-analysis. Medicine, 94.
- Chen, Y. H., Chou, H. C., Lin, S. T., Chen, Y. W., Lo, Y. W., & Chan, H. L. (2012). Effect of high glucose on secreted proteome in cultured retinal pigmented epithelium cells: its possible relevance to clinical diabetic retinopathy. Journal of proteomics, 77, 111-128.
- Cheung, N. Mitchell, P. Wong, T, Y. (2010). Diabetic retinopathy. The Lancet 9735: 124–136.
- Chung, Y. M., Kim, J. S., & Do Yoo, Y. (2006). A novel protein, Romo1, induces ROS production in the mitochondria. Biochemical and biophysical research communications, 347, 649-655.
- Chuyen, H. V., Nguyen, M. H., Roach, P. D., Golding, J. B., & Parks, S. E. (2015). Gac fruit (Momordica cochinchinensis Spreng.): a rich source of bioactive compounds and its potential health benefits. International Journal of Food Science & Technology 50: 567-577.
- Chuyen, H. V., Roach, P. D., Golding, J. B., Parks, S. E., & Nguyen, M. H. (2017a). Optimisation of extraction conditions for recovering carotenoids and antioxidant capacity from Gac peel using response surface methodology. International Journal of Food Science & Technology 52: 972-980.
- Chuyen, H. V., Tran, X. T., Nguyen, M. H., Roach, P. D., Parks, S. E., & Golding, J. B. (2017b). Yield of carotenoids, phenolic compounds and antioxidant capacity of extracts from Gac peel as affected by different solvents and extraction conditions. Journal of Advanced Agricultural Technologies, 4, 87-91.
- Cicero, A. F., Parini, A., & Rosticci, M. (2015). Nutraceuticals and cholesterol-lowering action. IJC Metabolic & Endocrine 6: 1-4.

- D Pawar, N., S Panchal, S., Shahmalak, T., & Gadgoli, C. (2016). Crocin Rich Extract of Nyctanthes arbor-tristis Flower Calyx Induces Anti-angiogenic Activity. The Natural Products Journal, 6, 40-48.
- da Silva, L. M. R., de Figueiredo, E. A. T., Ricardo, N. M. P. S., Vieira, I. G. P., de Figueiredo, R. W., Brasil, I. M., & Gomes, C. L. (2014). Quantification of bioactive compounds in pulps and by-products of tropical fruits from Brazil. Food chemistry, 143, 398-404.
- Dakin, H. A., Wordsworth, S., Rogers, C. A., Abangma, G., Raftery, J., Harding, S. P., Lotery, A. J., Downes, S. M., Chakravarthy, U., Reeves, B. C., & IVAN Study Investigators. (2014). Cost-effectiveness of ranibizumab and bevacizumab for age-related macular degeneration: 2-year findings from the IVAN randomised trial. BMJ open 4: e005094.
- deS Senanayake, P., Calabro, A., Hu, J. G., Bonilha, V. L., Darr, A., Bok, D., & Hollyfield, J. G. (2006). Glucose utilization by the retinal pigment epithelium: evidence for rapid uptake and storage in glycogen, followed by glycogen utilization. Experimental eye research, 83, 235-246.
- Duh, E. J., Sun, J. K., & Stitt, A. W. (2017). Diabetic retinopathy: current understanding, mechanisms, and treatment strategies. JCI insight, 2(14).
- Dunn, K. C., Aotaki-Keen, A. E., Putkey, F. R., & Hjelmeland, L. M. (1996). ARPE-19, a human retinal pigment epithelial cell line with differentiated properties. Experimental eye research, 62, 155-170.
- Dunn, K. C., Marmorstein, A. D., Bonilha, V. L., Rodriguez-Boulan, E., Giordano, F., & Hjelmeland, L. M. (1998). Use of the ARPE-19 cell line as a model of RPE polarity: basolateral secretion of FGF5. Investigative ophthalmology & visual science 39: 2744-2749.
- El-Asrar, A. M. A., Al-Mezaine, H. S., & Ola, M. S. (2009). Changing paradigms in the treatment of diabetic retinopathy. Current opinion in ophthalmology 20: 532-538.
- Ellong, E. N., Billard, C., Adenet, S., & Rochefort, K. (2015). Polyphenols, carotenoids, vitamin C content in tropical fruits and vegetables and impact of processing methods. Food and Nutrition Sciences, 6, 299.
- FAO Food and Nutrition paper 14/7, Manuals of Food Quality Control, 7. Food Analysis: General techniques, additives, contaminants, and composition (1986), p 210 213
- Flores, P., Sánchez, E., Fenoll, J., & Hellín, P. (2017). Genotypic variability of carotenoids in traditional tomato cultivars. Food Research International, 100, 510-516.

- Fu, D., & Tian, X. (2017). Effect of astaxanthin on retinal pigment epithelial cells in high glucose: an in-vitro study. Biomedical Research, 28.
- Gao, X., Zhang, H., Zhuang, W., Yuan, G., Sun, T., Jiang, X., ... & Dong, H. (2014). PEDF and PEDF-derived peptide 44mer protect cardiomyocytes against hypoxia-induced apoptosis and necroptosis via anti-oxidative effect. Scientific reports, 4, 5637.
- Gariano, R. F., & Gardner, T. W. (2005). Retinal angiogenesis in development and disease. Nature, 438, 960.
- Giovannucci, E., Rimm, E. B., Liu, Y., Stampfer, M. J., & Willett, W. C. (2002). A prospective study of tomato products, lycopene, and prostate cancer risk. Journal of the National Cancer Institute, 94, 391-398.
- Goh, P. P. (2008). Status of diabetic retinopathy among diabetics registered to the Diabetic Eye Registry, National Eye Database, 2007. The Medical Journal of Malaysia, 63, 24-28.
- Gong, X., & Rubin, L. P. (2015). Role of macular xanthophylls in prevention of common neovascular retinopathies: retinopathy of prematurity and diabetic retinopathy. Archives of biochemistry and biophysics 572: 40-48.
- Gong, X., Draper, C. S., Allison, G. S., Marisiddaiah, R., & Rubin, L. P. (2017). Effects of the macular carotenoid lutein in human retinal pigment epithelial cells. Antioxidants, 6, 100.
- Gorusupudi, A., Nelson, K., & Bernstein, P. S. (2017). The Age-Related Eye Disease 2 Study: Micronutrients in the Treatment of Macular Degeneration. Advances in Nutrition: An International Review Journal 8: 40-53.
- Grigsby, J., Betts, B., Vidro-Kotchan, E., Culbert, R., & Tsin, A. (2012). A possible role of acrolein in diabetic retinopathy: involvement of a VEGF/TGFβ signaling pathway of the retinal pigment epithelium in hyperglycemia. Current eye research, 37, 1045-1053.
- Grodowska, K., & Parczewski, A. (2010). Organic solvents in the pharmaceutical industry. Acta Poloniae Pharmaceutica, 67, 3-12.
- Hamidpour, R., Hamidpour, M., Hamidpour, S., & Shahlari, M. (2015). Cinnamon from the selection of traditional applications to its novel effects on the inhibition of angiogenesis in cancer cells and prevention of Alzheimer's disease, and a series of functions such as antioxidant, anticholesterol, antidiabetes, antibacterial, antifungal, nematicidal, acaracidal, and repellent activities. Journal of traditional and complementary medicine, 5, 66-70.
- Hammes, H. P. (2018). Diabetic retinopathy: hyperglycaemia, oxidative stress and beyond. Diabetologia, 61, 29-38.

- Handelman, G. J., Snodderly, D. M., Adler, A. J., Russett, M. D., & Dratz, E. A. (1992).
 [19] Measurement of carotenoids in human and monkey retinas. Methods in enzymology 213: 220-230.
- Haque, R., Iuvone, P. M., He, L., Hur, E. H., Chung Choi, K. S., Park, D., ... & Kumar, B. (2017). Prorenin receptor (PRR)-mediated NADPH oxidase (Nox) signaling regulates VEGF synthesis under hyperglycemic condition in ARPE-19 cells. Journal of Receptors and Signal Transduction, 37, 560-568.
- Heimsath Jr, E. G., Unda, R., Vidro, E., Muniz, A., Villazana-Espinoza, E. T., & Tsin, A. (2006). ARPE-19 cell growth and cell functions in euglycemic culture media. Current eye research, 31, 1073-1080.
- Hollborn, M., Tenckhoff, S., Seifert, M., Köhler, S., Wiedemann, P., Bringmann, A., & Kohen, L. (2006). Human retinal epithelium produces and responds to placenta growth factor. Graefe's Archive for Clinical and Experimental Ophthalmology 244: 732-741.
- Hu, B. J., Hu, Y. N., Lin, S., Ma, W. J., & Li, X. R. (2011). Application of lutein and zeaxanthin in nonproliferative diabetic retinopathy. International journal of ophthalmology 4: 303-306.
- Huang, O. S., Tay, W. T., Ong, P. G., Sabanayagam, C., Cheng, C. Y., Tan, G. S., Gheung, G. C., Lamoureux, E. L., & Wong, T. Y. (2015). Prevalence and determinants of undiagnosed diabetic retinopathy and vision-threatening retinopathy in a multiethnic Asian cohort: the Singapore Epidemiology of Eye Diseases (SEED) study. British Journal of Ophthalmology 99: 1614-1621.
- Huber, M., & Wachtlin, J. (2012). Vitreous levels of proteins implicated in angiogenesis are modulated in patients with retinal or choroidal neovascularization. Ophthalmologica, 228, 188-193.
- Hussein, Z., Taher, S. W., Singh, H. K. G., & Swee, W. C. S. (2015). Diabetes care in Malaysia: problems, new models, and solutions. Annals of global health 81: 851-862.
- Ikram, E. H. K., Eng, K. H., Jalil, A. M. M., Ismail, A., Idris, S., Azlan, A., ... & Mokhtar, R. A. M. (2009). Antioxidant capacity and total phenolic content of Malaysian underutilized fruits. Journal of food Composition and Analysis, 22, 388-393.
- Ishida, B. K., Turner, C., Chapman, M. H., & McKeon, T. A. (2004). Fatty acid and carotenoid composition of gac (Momordica cochinchinensis Spreng) fruit. Journal of agricultural and food chemistry 52: 274-279.
- Jo, D. H., Cho, C. S., Kim, J. H., Jun, H. O., & Kim, J. H. (2013). Animal models of diabetic retinopathy: doors to investigate pathogenesis and potential therapeutics. Journal of biomedical science 20: 20-38.

- Jomova, K., & Valko, M. (2013). Health protective effects of carotenoids and their interactions with other biological antioxidants. European journal of medicinal chemistry 70: 102-110.
- Joussen, A. M., & Joeres, S. (2007). Benefits and limitations in vitreoretinal surgery for proliferative diabetic retinopathy and macular edema. In Diabetic Retinopathy 39, pp. 69-87. Karger Publishers.
- Jung, K., Chin, Y. W., Chung, Y. H., Park, Y. H., Yoo, H., Min, D. S., Lee, B., & Kim, J. (2013a). Anti-gastritis and wound healing effects of Momordicae Semen extract and its active component. Immunopharmacology and immunotoxicology 35: 126-132.
- Jung, K., Chin, Y. W., Yoon, K. D., Chae, H. S., Kim, C. Y., Yoo, H., & Kim, J. (2013b). Anti-inflammatory properties of a triterpenoidal glycoside from Momordica cochinchinensis in LPS-stimulated macrophages. Immunopharmacology and immunotoxicology 35: 8-14.
- Jung, K., Lee, D., Yu, J. S., Namgung, H., Kang, K. S., & Kim, K. H. (2016). Protective effect and mechanism of action of saponins isolated from the seeds of gac (Momordica cochinchinensis Spreng.) against cisplatin-induced damage in LLC-PK1 kidney cells. Bioorganic & medicinal chemistry letters 26: 1466-1470
- Kaur, A., Gupta, V., Christopher, A. F., Malik, M. A., & Bansal, P. (2017). Nutraceuticals in prevention of cataract—An evidence based approach. Saudi Journal of Ophthalmology 31: 30-37.
- Kayama, Y., Raaz, U., Jagger, A., Adam, M., Schellinger, I. N., Sakamoto, M., Hirofumi, S., Toyama, K., Spin, J. M., & Tsao, P. S. (2015). Diabetic cardiovascular disease induced by oxidative stress. International journal of molecular sciences 16: 25234-25263.
- Kha, T. C., Nguyen, M. H., Roach, P. D., Parks, S. E., & Stathopoulos, C. (2013). Gac fruit: nutrient and phytochemical composition, and options for processing. Food reviews international 29: 92-106.
- Khoo, H. E., Ismail, A., Mohd-Esa, N., & Idris, S. (2008). Carotenoid content of underutilized tropical fruits. Plant Foods for Human Nutrition, 63, 170-175.
- Kibel, A., Selthofer-Relatic, K., Drenjancevic, I., Bacun, T., Bosnjak, I., Kibel, D., & Gros, M. (2017). Coronary microvascular dysfunction in diabetes mellitus. Journal of International Medical Research 45: 1901-1929.
- Kim, B. K., Kim, B. R., Lee, H. J., Lee, S. A., Kim, B. J., Kim, H., ... & Kim, B. J. (2014). Tumor-suppressive effect of a telomerase-derived peptide by inhibiting hypoxia-induced HIF-1α-VEGF signaling axis. Biomaterials, 35, 2924-2933.

- Kim, D. I., Park, M. J., Choi, J. H., Lim, S. K., Choi, H. J., & Park, S. H. (2015). Hyperglycemia-induced GLP-1R downregulation causes RPE cell apoptosis. The international journal of biochemistry & cell biology, 59, 41-51.
- Kim, Y. S., Lee, H. A., Lim, J. Y., Kim, Y., Jung, C. H., Yoo, S. H., & Kim, Y. (2014). β-Carotene inhibits neuroblastoma cell invasion and metastasis in vitro and in vivo by decreasing level of hypoxia-inducible factor-1α. The Journal of nutritional biochemistry, 25, 655-664.
- King, G. L., Kunisaki, M., Nishio, Y., Inoguchi, T., Shiba, T., & Xia, P. (1996). Biochemical and molecular mechanisms in the development of diabetic vascular complications. Diabetes, 45, S105-S108.
- Klungsupya, P., Saenkhum, J., Muangman, T., Rerk-Am, U., Laovitthayanggoon, S., & Leelamanit, W. (2012). Non-Cytotoxic Property and DNA Protective Activity against H2O2 and UVC of Thai GAC Fruit Extracts in Human TK6 Cells. Journal of Applied Pharmaceutical Science, 2, 4.
- Krinsky, N. I., & Johnson, E. J. (2005). Carotenoid actions and their relation to health and disease. Molecular aspects of medicine 26: 459-516.
- Kubola, J., & Siriamornpun, S. (2011). Phytochemicals and antioxidant activity of different fruit fractions (peel, pulp, aril and seed) of Thai gac (Momordica cochinchinensis Spreng). Food chemistry 127: 1138-1145.
- Kuppusamy, P., Yusoff, M. M., Maniam, G. P., Ichwan, S. J. A., Soundharrajan, I., & Govindan, N. (2014). Nutraceuticals as potential therapeutic agents for colon cancer: a review. Acta Pharmaceutica Sinica B 4: 173-181.
- Leasher, J. L., Bourne, R. R., Flaxman, S. R., Jonas, J. B., Keeffe, J., Naidoo, K., Pesudovs, K., Price, H., White, R. A., Wong, T. Y., Resnikoff, S., & Taylor, H. R. (2016). Global estimates on the number of people blind or visually impaired by diabetic retinopathy: a meta-analysis from 1990 to 2010. Diabetes care 39: 1643-1649.
- Lee, R., Wong, T. Y., & Sabanayagam, C. (2015). Epidemiology of diabetic retinopathy, diabetic macular edema and related vision loss. Eye and vision 2: 17.
- Lee, S. H., Lee, J. S., Lee, E. J., Min, K. H., Hur, G. Y., Lee, S. H., ... & Shim, J. J. (2014). Serum reactive oxygen species modulator 1 (Romo1) as a potential diagnostic biomarker for non-small cell lung cancer. Lung Cancer, 85, 175-181.
- Li, Z. Z., Lu, X. Z., Ma, C. C., & Chen, L. (2010). Serum lycopene levels in patients with diabetic retinopathy. European journal of ophthalmology 20: 719-723.

- Lian, J. X., Gangwani, R. A., McGhee, S. M., Chan, C. K., Lam, C. L. K., & Wong, D. S. H. (2015). Systematic screening for diabetic retinopathy (DR) in Hong Kong: prevalence of DR and visual impairment among diabetic population. British Journal of Ophthalmology 100: 151-155.
- Lim, T. K. (2012). Edible medicinal and non-medicinal plants, pp. 656-687. New York, NY, USA: Springer.
- Liu, H. R., Meng, L. Y., Lin, Z. Y., Shen, Y., Yu, Y. Q., & Zhu, Y. Z. (2012). Cochinchina momordica seed extract induces apoptosis and cell cycle arrest in human gastric cancer cells via PARP and p53 signal pathways. Nutrition and cancer 64: 1070-1077.
- Liu, L., Zuo, Z., Lu, S., Liu, A., & Liu, X. (2017). Naringin attenuates diabetic retinopathy by inhibiting inflammation, oxidative stress and NF-κB activation in vivo and in vitro. Iranian journal of basic medical sciences 20: 813-821.
- Malaysian Standard (MS 1191:1991), UDC 642.2:641.13, SIRIM, 1991, pg 8.
- Mallika, P. S., Lee, P. Y., Cheah, W. L., Wong, J. S., Alwi, S. S., Hayati, H. N., & Tan, A. K. (2011). Risk factors for diabetic retinopathy in diabetics screened using fundus photography at a primary health care setting in East Malaysia. Malaysian family physician: the official journal of the Academy of Family Physicians of Malaysia 6: 60-65.
- Maminishkis, A., Chen, S., Jalickee, S., Banzon, T., Shi, G., Wang, F. E., Ehalt, T., Hammer, J. A., & Miller, S. S. (2006). Confluent monolayers of cultured human fetal retinal pigment epithelium exhibit morphology and physiology of native tissue. Investigative ophthalmology & visual science 47: 3612-3624.
- Mannermaa, E., Reinisalo, M., Ranta, V. P., Vellonen, K. S., Kokki, H., Saarikko, A., Kaarniranta., & Urtti, A. (2010). Filter-cultured ARPE-19 cells as outer blood-retinal barrier model. European Journal of Pharmaceutical Sciences 40: 289-296.
- Mateos, M. V., Tenconi, P. E., Giusto, N. M., & Salvador, G. A. (2015). Inflammation and oxidative stress in retinal diseases: the role of intracellular signaling in the retinal pigment epithelium. International Journal of Ophthalmology and Clinical Research 8:1-7.
- Milani, A., Basirnejad, M., Shahbazi, S., & Bolhassani, A. (2017). Carotenoids: biochemistry, pharmacology and treatment. British journal of pharmacology, 174, 1290-1324.
- Mohamed, Q., & Wong, T. Y. (2008). Emerging drugs for diabetic retinopathy. Expert opinion on emerging drugs 13: 675-694.

- Mohan, N., Monickaraj, F., Balasubramanyam, M., Rema, M., & Mohan, V. (2012). Imbalanced levels of angiogenic and angiostatic factors in vitreous, plasma and postmortem retinal tissue of patients with proliferative diabetic retinopathy. Journal of Diabetes and its Complications, 26, 435-441.
- Moo-Huchin, V. M., Estrada-Mota, I., Estrada-León, R., Cuevas-Glory, L., Ortiz-Vázquez, E., y Vargas, M. D. L. V., ... & Sauri-Duch, E. (2014). Determination of some physicochemical characteristics, bioactive compounds and antioxidant activity of tropical fruits from Yucatan, Mexico. Food chemistry, 152, 508-515.
- Müller-Maatsch, J., Sprenger, J., Hempel, J., Kreiser, F., Carle, R., & Schweiggert, R. M. (2016). Carotenoids from gac fruit aril (Momordica cochinchinensis [Lour.] Spreng.) are more bioaccessible than those from carrot root and tomato fruit. Food Research International 99: 928-935.
- Nagineni, C. N., Kutty, V., Detrick, B., & Hooks, J. J. (2005). Expression of PDGF and their receptors in human retinal pigment epithelial cells and fibroblasts: Regulation by TGF β. Journal of cellular physiology 203: 35-43.
- Nhi, T. T. Y., & Tuan, D. Q. (2016). Enzyme Assisted Extraction of GAC Oil (Momordica cochinchinensis Spreng) from Dried Aril. Journal of Food and Nutrition Sciences 4: 1-6.
- Nhung, D. T. T., Bung, P. N., Ha, N. T., & Phong, T. K. (2010). Changes in lycopene and beta carotene contents in aril and oil of gac fruit during storage. Food Chemistry 121: 326-331.
- Niranjana, R., Gayathri, R., Mol, S. N., Sugawara, T., Hirata, T., Miyashita, K., & Ganesan, P. (2015). Carotenoids modulate the hallmarks of cancer cells. Journal of Functional Foods 18: 968-985.
- Nishino, A., Yasui, H., & Maoka, T. (2017). Reaction and scavenging mechanism of β-carotene and zeaxanthin with reactive oxygen species. Journal of oleo science, 66, 77-84.
- Paine, S. K., Mondal, L. K., Borah, P. K., Bhattacharya, C. K., & Mahanta, J. (2017). Pro-and antiangiogenic VEGF and its receptor status for the severity of diabetic retinopathy. Molecular vision 23: 356-356.
- Parikh, R. N., Traband, A., Kolomeyer, A. M., VanderBeek, B. L., Kim, B. J., Maguire, A. M., & Brucker, A. J. (2017). Intravitreal bevacizumab for the treatment of vitreous hemorrhage due to proliferative diabetic retinopathy. American Journal of Ophthalmology 176: 194-202.
- Parks, S. E., Murray, C. T., Gale, D. L., Al-Khawaldeh, B., & Spohr, L. J. (2013). Propagation and production of Gac (Momordica cochinchinensis Spreng.), a greenhouse case study. Experimental agriculture, 49, 234-243.

- Pavan, B., Capuzzo, A., & Forlani, G. (2014). High glucose-induced barrier impairment of human retinal pigment epithelium is ameliorated by treatment with Goji berry extracts through modulation of cAMP levels. Experimental eye research 120: 50-54.
- Peris-Tortajada, M. (2004). Carbohydrates and starch. In M. L. N. Leo (Ed.), Handbook of food analysis: Physical characterization and nutrient analysis (pp. 392). USA: Marcel Dekker, Inc.
- Petchsak, P., & Sripanidkulchai, B. (2015). Momordica cochinchinensis aril extract induced apoptosis in human MCF-7 breast cancer cells. Asian Pac J Cancer Prev 16: 5507-5513.
- Ponnalagu, M., Subramani, M., Jayadev, C., Shetty, R., & Das, D. (2017). Retinal pigment epithelium-secretome: A diabetic retinopathy perspective. Cytokine 95: 126-135.
- Poulaki, V., Joussen, A. M., Mitsiades, N., Mitsiades, C. S., Iliaki, E. F., & Adamis, A. P. (2004). Insulin-like growth factor-I plays a pathogenetic role in diabetic retinopathy. The American journal of pathology 165: 457-469.
- Prayong, P., Barusrux, S., & Weerapreeyakul, N. (2008). Cytotoxic activity screening of some indigenous Thai plants. Fitoterapia, 79, 598-601.
- Premanand, C., Rema, M., Sameer, M. Z., Sujatha, M., & Balasubramanyam, M. (2006). Effect of curcumin on proliferation of human retinal endothelial cells under in vitro conditions. Investigative ophthalmology & visual science, 47, 2179-2184.
- Puwastien, P., Siong, T. E., Kantasubrata, J., Caven, G., Felicionoand, R. R., & Judprasong, K. (2011). Asean Manual of Food Analysis. Regional centre of Asean Network of Food Data System. Institute of Nutrition, Mahidol University Thailand, 1-190.
- Rahimi-Madiseh, M., Malekpour-Tehrani, A., Bahmani, M., & Rafieian-Kopaei, M.
 (2016). The research and development on the antioxidants in prevention of diabetic complications. Asian Pacific journal of tropical medicine 9: 825-831.
- Rahmatullah, M., Biswas, A., Haq, W. M., Seraj, S., & Jahan, R. (2012). An ethnomedicinal survey of cucurbitaceae family plants used in the folk medicinal practices of Bangladesh. Chronicles of Young Scientists 3: 212-222.
- Ranneh, Y., Ali, F., Akim, A. M., Hamid, H. A., Khazaai, H., & Fadel, A. (2017). Crosstalk between reactive oxygen species and pro-inflammatory markers in developing various chronic diseases: a review. Applied Biological Chemistry 60: 327-338.

- Ranneh, Y., Ali, F., Zarei, M., Akim, A. M., Hamid, H. A., & Khazaai, H. (2018). Malaysian stingless bee and Tualang honeys: A comparative characterization of total antioxidant capacity and phenolic profile using liquid chromatographymass spectrometry. LWT-Food Science and Technology, 89, 1-9.
- Rao, A. V., & Rao, L. G. (2007). Carotenoids and human health. Pharmacological research 55: 207-216.
- Ren, J. G., Jie, C., & Talbot, C. (2005). How PEDF prevents angiogenesis: a hypothesized pathway. Medical hypotheses, 64, 74-78.
- Rodriguez-Amaya, D. B. (2018). Natural food pigments and colorants. In Bioactive Molecules in Food (pp. 1-35). Springer, Cham.
- Roglic, G., Varghese, C., & Thamarangsi, T. (2016). Diabetes in South-East Asia: burden, gaps, challenges and ways forward. WHO South East Asia J Public Health 5:1-4.
- Ruiz, R. B., & Hernández, P. S. (2016). Cancer chemoprevention by dietary phytochemicals: Epidemiological evidence. Maturitas 94: 13-19.
- Saini, R. K., Nile, S. H., & Park, S. W. (2015). Carotenoids from fruits and vegetables: chemistry, analysis, occurrence, bioavailability and biological activities. Food Research International 76: 735-750.
- Samuel, W., Jaworski, C., Postnikova, O. A., Kutty, R. K., Duncan, T., Tan, L. X., ... & Redmond, T. M. (2017). Appropriately differentiated ARPE-19 cells regain phenotype and gene expression profiles similar to those of native RPE cells. Molecular vision, 23, 60.
- Sasongko, M. B., Widyaputri, F., Agni, A. N., Wardhana, F. S., Kotha, S., Gupta, P., Widayanti, T. W., Haryanto, S., Widyaningrum, R., Wong, T. Y., & Kawasaki, R. (2017). Prevalence of diabetic retinopathy and blindness in Indonesian adults with type 2 diabetes. American journal of ophthalmology 181: 79-87.
- Schweiggert, R. M., Kopec, R. E., Villalobos-Gutierrez, M. G., Högel, J., Quesada, S., Esquivel, P., ... & Carle, R. (2014). Carotenoids are more bioavailable from papaya than from tomato and carrot in humans: a randomised cross-over study. British Journal of Nutrition 111: 490-498.
- Scripsema, N. K., Hu, D. N., & Rosen, R. B. (2015). Lutein, zeaxanthin, and meso-zeaxanthin in the clinical management of eye disease. Journal of ophthalmology 2015: 865179.
- Selvaraj, K., Gowthamarajan, K., Karri, V. V. S. R., Barauah, U. K., Ravisankar, V., & Jojo, G. M. (2017). Current treatment strategies and nanocarrier based approaches for the treatment and management of diabetic retinopathy. Journal of Drug Targeting 25: 386-405.

- Sharma, Y., Saxena, S., Mishra, A., Saxena, A., & Natu, S. M. (2017). Nutrition for diabetic retinopathy: plummeting the inevitable threat of diabetic vision loss. European Journal of Nutrition 56: 2013-2027.
- Shaw, J. E., Sicree, R. A., & Zimmet, P. Z. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. Diabetes research and clinical practice 87: 4-14.
- Shen, M., Zhou, X. Z., Ye, L., Yuan, Q., Shi, C., Zhu, P. W., ... & Shao, Y. (2018). Xanthatin inhibits corneal neovascularization by inhibiting the VEGFR2 mediated STAT3/PI3K/Akt signaling pathway. International journal of molecular medicine, 42, 769-778.
- Shen, Y., Meng, L., Sun, H., Zhu, Y., & Liu, H. (2015). Cochinchina momordica seed suppresses proliferation and metastasis in human lung cancer cells by regulating multiple molecular targets. The American journal of Chinese medicine 43: 149-166.
- Shen, Y., Zhang, H., Cheng, L., Wang, L., Qian, H., & Qi, X. (2016). In vitro and in vivo antioxidant activity of polyphenols extracted from black highland barley. Food chemistry 194: 1003-1012.
- Simó, R., & Hernandez, C. (2008). Intravitreous anti-VEGF for diabetic retinopathy: hopes and fears for a new therapeutic strategy. Diabetologia 51: 1574.
- Simó, R., & Hernández, C. (2009). Advances in the medical treatment of diabetic retinopathy. Diabetes Care 32: 1556-1562.
- Simó, R., Villarroel, M., Corraliza, L., Hernández, C., & Garcia-Ramírez, M. (2010). The retinal pigment epithelium: something more than a constituent of the blood-retinal barrier—implications for the pathogenesis of diabetic retinopathy. BioMed Research International 2010: 190724.
- Singh, R., Kaur, N., Kishore, L., & Gupta, G. K. (2013). Management of diabetic complications: a chemical constituents based approach. Journal of ethnopharmacology 150: 51-70.
- Slomiany, M. G., & Rosenzweig, S. A. (2004). Autocrine effects of IGF-I-induced VEGF and IGFBP-3 secretion in retinal pigment epithelial cell line ARPE-19. American Journal of Physiology-Cell Physiology 287: C746-C753.
- Soares, N. D. C. P., Machado, C. L., Trindade, B. B., do Canto Lima, I. C., Gimba, E. R. P., Teodoro, A. J., ... & Borojevic, R. (2017). Lycopene Extracts from Different Tomato-Based Food Products Induce Apoptosis in Cultured Human Primary Prostate Cancer Cells and Regulate TP53, Bax and Bcl-2 Transcript Expression. Asian Pacific journal of cancer prevention: APJCP, 18, 339.

- Stitt, A. W., Curtis, T. M., Chen, M., Medina, R. J., McKay, G. J., Jenkins, A., ... & Lois, N. (2016). The progress in understanding and treatment of diabetic retinopathy. Progress in retinal and eye research 51: 156-186.
- Strauss, O. (2005). The retinal pigment epithelium in visual function. Physiological reviews 85: 845-881.
- Sukhorum, W., Sampannang, A., Sripanidkulchai, B., Iamsaard, S., SUKHORUM, W., SAMPANNANG, A., ... & IAMSAARD, S. (2016). Momordica cochinchinensis (L.) Spreng. Aril Extract Prevents Adverse Reproductive Parameters of Male Rats Induced with Valproic Acid. Int. J. Morphol 34: 870-876.
- Sulaiman, S. F., & Ooi, K. L. (2014). Antioxidant and α-glucosidase inhibitory activities of 40 tropical juices from Malaysia and identification of phenolics from the bioactive fruit juices of Barringtonia racemosa and Phyllanthus acidus. Journal of agricultural and food chemistry, 62, 9576-9585.
- Sun, J., Xu, Y., Sun, S., Sun, Y., & Wang, X. (2010). Intermittent high glucose enhances cell proliferation and VEGF expression in retinal endothelial cells: the role of mitochondrial reactive oxygen species. Molecular and cellular biochemistry, 343, 27-35.
- Tajunisah, I., Wong, P. S., Tan, L. T., Rokiah, P., & Reddy, S. C. (2011). Awareness of eye complications and prevalence of retinopathy in the first visit to eye clinic among type 2 diabetic patients. International journal of ophthalmology 4: 519-524.
- Talvas, J., Caris-Veyrat, C., Guy, L., Rambeau, M., Lyan, B., Minet-Quinard, R., ... & Rock, E. (2010). Differential effects of lycopene consumed in tomato paste and lycopene in the form of a purified extract on target genes of cancer prostatic cells—. The American journal of clinical nutrition, 91, 1716-1724.
- Tee, E. S., & Yap, R. W. K. (2017). Type 2 diabetes mellitus in Malaysia: current trends and risk factors. European Journal of Clinical Nutrition 71: 844-849.
- Tien, P. G., Kayama, F., Konishi, F., Tamemoto, H., Kasono, K., Hung, N. T. K., Kuroki, M., Ishikawa, S. E., Van, C. N., & Kawakami, M. (2005). Inhibition of tumor growth and angiogenesis by water extract of Gac fruit (Momordica cochinchinensis Spreng). International journal of oncology 26: 881-889.
- Ting, D. S. W., Cheung, G. C. M., & Wong, T. Y. (2016). Diabetic retinopathy: global prevalence, major risk factors, screening practices and public health challenges: a review. Clinical & experimental ophthalmology, 44, 260-277.
- Tinrat, S., Akkarachaneeyakorn, S., & Singhapol, C. (2014). Evaluation of antioxidant and antimicrobial activities of Momordica Cochinchinensis Spreng (Gac fruit) ethanolic extract. International Journal of Pharmaceutical Sciences and Research, 5, 3163-3169.

- Tolentino, M. S., Tolentino, A. J., & Tolentino, M. J. (2016). Current and investigational drugs for the treatment of diabetic retinopathy. Expert opinion on investigational drugs 25: 1011-1022.
- Tran, X. T., Parks, S. E., Roach, P. D., Golding, J. B., & Nguyen, M. H. (2016). Effects of maturity on physicochemical properties of Gac fruit (Momordica cochinchinensis Spreng.). Food science & nutrition 4: 305-314.
- Trial, C. (2008). Prolonged Effect of Intensive Therapy on the Risk of Retinopathy Complications in Patients With Type 1 Diabetes Mellitus. Arch Ophthalmol 126: 1707-1715.
- Tsai, F. J., Li, T. M., Ko, C. H., Cheng, C. F., Ho, T. J., Liu, X., ... & Huang, S. M. (2017). Effects of Chinese herbal medicines on the occurrence of diabetic retinopathy in type 2 diabetes patients and protection of ARPE-19 retina cells by inhibiting oxidative stress. Oncotarget 8: 63528-63550.
- Tsoi, A. Y. K., Ng, T. B., & Fong, W. P. (2005). Antioxidative effect of a chymotrypsin inhibitor from Momordica cochinchinensis (Cucurbitaceae) seeds in a primary rat hepatocyte culture. Journal of Peptide Science 11: 665-668.
- Tsoi, A. Y., Wong, R. C., Ng, T. B., & Fong, W. P. (2004). First report on a potato I family chymotrypsin inhibitor from the seeds of a Cucurbitaceous plant, Momordica cochinchinensis. Biological chemistry 385: 185-189.
- Tundis, R., Menichini, F., Bonesi, M., Conforti, F., Statti, G., Menichini, F., & Loizzo, M. R. (2013). Antioxidant and hypoglycaemic activities and their relationship to phytochemicals in Capsicum annuum cultivars during fruit development. LWT-Food Science and Technology 53: 370-377.
- Ventrice, P., Leporini, C., Jose'Francisco Aloe, E. G., Leuzzi, G., Marrazzo, G., Scorcia, G. B., Bruzzichesi, D., Nicola, V., & Scorcia, V. (2013). Anti-vascular endothelial growth factor drugs safety and efficacy in ophthalmic diseases. Journal of pharmacology & pharmacotherapeutics 4: S38-S42.
- Vuong, L. T. (2000). Underutilized β-carotene–rich crops of Vietnam. Food and Nutrition Bulletin 21: 173-181.
- Vuong, L. T. (2003). Gac: a fruit from heaven. Vietnam Journal, October 1-22.
- Vuong, L. T., Dueker, S. R., & Murphy, S. P. (2002). Plasma β-carotene and retinol concentrations of children increase after a 30-d supplementation with the fruit Momordica cochinchinensis (gac). The American journal of clinical nutrition 75: 872-879.
- Vuong, L. T., Franke, A. A., Custer, L. J., & Murphy, S. P. (2006). Momordica cochinchinensis Spreng.(gac) fruit carotenoids reevaluated. Journal of Food Composition and Analysis 19: 664-668.

- Wang, D., Wang, L., Gu, J., Yang, H., Liu, N., Lin, Y., Li, X., & Shao, C. (2014). Scutellarin Inhibits High Glucose–induced and Hypoxia-mimetic Agent–induced Angiogenic Effects in Human Retinal Endothelial Cells through Reactive Oxygen Species/Hypoxia-inducible Factor-1α/Vascular Endothelial Growth Factor Pathway. Journal of cardiovascular pharmacology 64: 218-227.
- Wang, L. Z., Cheung, C. Y., Tapp, R. J., Hamzah, H., Tan, G., Ting, D., Lamoureux, E., & Wong, T. Y. (2017). Availability and variability in guidelines on diabetic retinopathy screening in Asian countries. British Journal of Ophthalmology 101: 1352-1360.
- Wankun, X., Wenzhen, Y., Min, Z., Weiyan, Z., Huan, C., Wei, D., Lvzhen, H., Xu, Y., & Xiaoxin, L. (2011). Protective effect of paeoniflorin against oxidative stress in human retinal pigment epithelium in vitro. Molecular vision 17: 3512-3522.
- Wilkinson, C. P., Ferris, F. L., Klein, R. E., Lee, P. P., Agardh, C. D., Davis, M., Kampik,
 A., Pararajasegaram, R., Verdaguer, J. T., & Group, G. D. R. P. (2003).
 Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. Ophthalmology 110: 1677-1682.
- Wimalasiri, D., Piva, T., & Huynh, T. (2016). Diversity in Nutrition and Bioactivity of Momordica cochinchinensis. International Journal on Advanced Science, Engineering and Information Technology 6: 378-380.
- Wirostko, B., Wong, T. Y., & Simó, R. (2008). Vascular endothelial growth factor and diabetic complications. Progress in retinal and eye research 27: 608-621.
- World Health Organization. (2016). Global report on diabetes. World Health Organization.
- Wu, J., Cho, E., Willett, W. C., Sastry, S. M., & Schaumberg, D. A. (2015). Intakes of lutein, zeaxanthin, and other carotenoids and age-related macular degeneration during 2 decades of prospective follow-up. JAMA ophthalmology 133: 1415-1424.
- Xia, T., & Rizzolo, L. J. (2017). Effects of diabetic retinopathy on the barrier functions of the retinal pigment epithelium. Vision research, 139, 72-81.
- Xiao, C., Hu, S., & Rajput, Z. I. (2007). Adjuvant effect of an extract from Cochinchina momordica seeds on the immune responses to ovalbumin in mice. Frontiers of Agriculture in China 1: 90-95.
- Xing, Q., Zhang, G., Kang, L., Wu, J., Chen, H., Liu, G., ... & Lu, P. (2017). The Suppression of Kallistatin on High-Glucose-Induced Proliferation of Retinal Endothelial Cells in Diabetic Retinopathy. Ophthalmic research, 57, 141-149.

- Xu, H. Z., Song, Z., Fu, S., Zhu, M., & Le, Y. Z. (2011). RPE barrier breakdown in diabetic retinopathy: seeing is believing. Journal of ocular biology, diseases, and informatics 4: 83-92.
- Xu, X. H., Zhao, C., Peng, Q., Xie, P., & Liu, Q. H. (2017). Kaempferol inhibited VEGF and PGF expression and in vitro angiogenesis of HRECs under diabetic-like environment. Brazilian Journal of Medical and Biological Research 50: e5396.
- Yamagata, K., Tagami, M., & Yamori, Y. (2015). Dietary polyphenols regulate endothelial function and prevent cardiovascular disease. Nutrition 31: 28-37.
- Yau, J. W., Rogers, S. L., Kawasaki, R., Lamoureux, E. L., Kowalski, J. W., Bek, T., ... & Haffner, S. (2012). Global prevalence and major risk factors of diabetic retinopathy. Diabetes care 35: 556-564.
- Yen, S. T., Tan, A. K., & Mustapha, F. I. (2017). Awareness of diabetes, hypertension, and hypercholesterolemia in Malaysia. Journal of diabetes 9: 874-883.
- Zang, J., & Guan, G. (2015). Study of pigment epithelium-derived factor in pathogenesis of diabetic retinopathy. Journal of Ophthalmology, 30, 81-88.
- Zhang, S. X., & Ma, J. X. (2007). Ocular neovascularization: Implication of endogenous angiogenic inhibitors and potential therapy. Progress in retinal and eye research 26: 1-37.
- Zheng, L., Zhang, Y. M., Zhan, Y. Z., & Liu, C. X. (2014). Momordica cochinchinensis seed extracts suppress migration and invasion of human breast cancer ZR-75-30 cells via down-regulating MMP-2 and MMP-9. Asian Pac J Cancer Prev 15: 1105-10.
- Zheng, Y., He, M., & Congdon, N. (2012). The worldwide epidemic of diabetic retinopathy. Indian journal of ophthalmology 60: 428-431.
- Zong, H., Ward, M., & Stitt, A. W. (2011). AGEs, RAGE, and diabetic retinopathy. Current diabetes reports 11: 244-252. Abdul-Ghani, M., DeFronzo, R. A., Del Prato, S., Chilton, R., Singh, R., & Ryder, R. E. (2017). Cardiovascular disease and type 2 diabetes: has the dawn of a new era arrived?. Diabetes Care 40: 813-820.

BIODATA OF STUDENT

Ali Abdulqader Mahdi was born in Baghdad, Iraq on 18 October of 1989. He graduated with a bachelor degree from the University of Technology, Baghdad, Iraq in biochemical technology, department of biotechnology, faculty of applied science since 2012. He worked in the medical and laboratory tools and instruments and also in medical laboratory for three years. Then he applied to study Masters of Science in the Faculty of Medicine and Health Sciences, UPM, Malaysia since 2016. Currently, he is submitting his final project as a requirement to get master's degree from University Putra Malaysia.



LIST OF PUBLICATIONS

- Abdulqader, A., Ali, F., Ismail, A., & Esa, N. M. (2018). Gac fruit extracts ameliorate proliferation and modulate angiogenic markers of human retinal pigment epithelial cells under high glucose conditions. Asian Pacific Journal of Tropical Biomedicine, 8(12), 571.
- Abdulqader, A., Ali, F., Ismail, A., & Esa, N. M. (2018). Gac (Momordica cochinchinensis Spreng.) Fruit and its Potentiality and Superiority in-Health Benefits. Journal of Contemporary Medical Sciences, 4(4).
- Abdulqader, A., Ali, F., Ismail, A., & Esa, N. M. (2019). Antioxidants compounds and capacities of Gac (*Momordica cochinchinensis* Spreng) fruits". Research paper-accepted for publication in the Asian Pacific Journal of Tropical Biomedicine.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

	ACADE	MIC SESSION :				
TITLE O	F THESIS / PROJECT RE	EPORT :				
EFFECT	EFFECTS OF Momordica cochinchinensis SPRENG, FRUIT ON THE PROLIFERATION					
	AND ANGIOGENESIS BIOMARKERS OF HUMAN RETINAL PIGMENT EPITHELIAL					
	INDER HIGH GLUCOSE					
NAME O	F STUDENT: ALI ABDUL	QADER MAHDI MAHDI				
belonged	ledge that the copyright to Universiti Putra Mala the library under the follo	and other intellectual property in the thesis/project report ysia and I agree to allow this thesis/project report to be wing terms:				
1. This th	esis/project report is the p	property of Universiti Putra Malaysia.				
2. The like purpos	orary of Universiti Putra ses only.	Malaysia has the right to make copies for educational				
3. The lil acade	orary of Universiti Putra mic exchange.	Malaysia is allowed to make copies of this thesis for				
l declare	that this thesis is classifie	d as:				
*Please tid	k (√)					
	CONFIDENTIAL	(Contain confidential information under Official Secret Act 1972).				
	RESTRICTED	(Contains restricted information as specified by the organization/institution where research was done).				
	OPEN ACCESS	I agree that my thesis/project report to be published as hard copy or online open access.				
This thesi	s is submitted for :					
	PATENT	Embargo from until (date)				
		Approved by:				
	of Student) / Passport No.:	(Signature of Chairman of Supervisory Committee) Name:				
Date :		Date:				

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]