



UNIVERSITI PUTRA MALAYSIA

***SCREENING OF ANTAGONISTIC BACTERIA FOR BIOCONTROL
ACTIVITIES AGAINST FUSARIUM OXYSPORUM AND
GANODERMA BONINENSE***

NURUL ASHIQIN BINTI MAT NAWI

FP 2013 83

**SCREENING OF ANTAGONISTIC BACTERIA FOR BIOCONTROL
ACTIVITIES AGAINST *FUSARIUM OXYSPORUM* AND *GANODERMA
BONINENSE***

BY

NURUL ASHIQIN BINTI MAT NAWI

**A project report submitted to Faculty of Agriculture, Universiti Putra Malaysia, in
fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of
the degree of Bachelor of Agricultural Science**

Faculty of Agriculture

University Putra Malaysia

2012/2013

CERTIFICATION FORM

This project report entitled “Screening Antagonistic Bacteria For Biocontrol Activities against *Fusarium oxysporum* and *Ganoderma boninense*” is prepared by Nurul Ashiqin Binti Mat Nawi and submitted to the Faculty of Agriculture in fulfillment of the requirement of PRT 4999 (Final Year Project) for the award of the degree of Bachelor of Agricultural Science.

Student’s name :

NURUL ASHIQIN BINTI MAT NAWI

Student’s signature:

Certified by:

Assoc. Prof. Dr Jugah Bin Kadir

Senior Lecturer

Department of Plant Protection

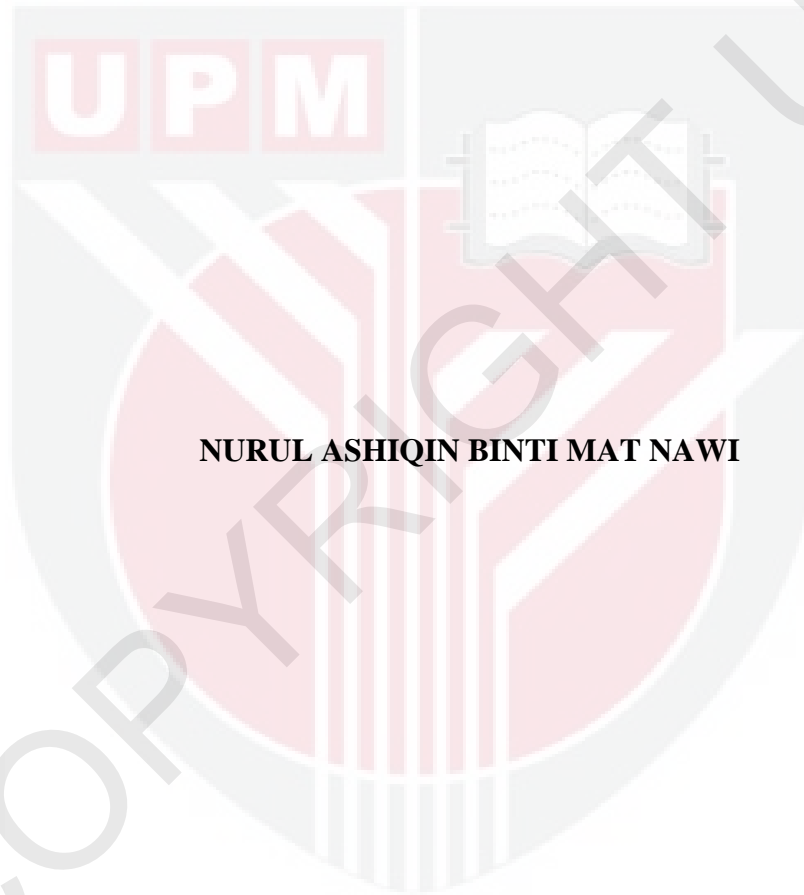
Faculty of Agriculture

University Putra Malaysia

Date: _____

**SCREENING OF ANTAGONISTIC BACTERIA FOR BIOCONTROL
ACTIVITIES AGAINST *FUSARIUM OXYSPORUM* AND *GANODERMA***

BONINENSE



NURUL ASHIQIN BINTI MAT NAWI

FACULTY OF AGRICULTURE

UNIVERSITI PUTRA MALAYSIA

SERDANG, SELANGOR DARUL EHSAN

2012/2013

ACKNOWLEDGMENT

First and foremost, I wish to express my thankfulness to Almighty Allah S.W.T because of His bless fullness and guidance to me from the beginning of this project until now. I take this opportunity to express my appreciation to the people who have been instrumental in the successful completion of this project report.

I would like to express my sincere gratitude to my project supervisor, Assoc. Prof. Dr. Jugah Bin Kadir for his invaluable advice, ideas, encouragement, guidance, discussion and suggestion throughout this project to make sure I can finish this project smoothly. I am also indebted to PhD and Master's student in laboratory of Microbial Biological Control which are Miss Salha Ibnouf, Madam Shamima Akter and Miss Bibi Nazihah Mohd Din for their guidance and help as well as assisting me during handling this project at laboratory. Their guidance and assist are giving me a good knowledge how to work in laboratory.

I also like to show my special thanks to my beloved family especially my parent, Mat Nawi Bin Yahya and Che Rusni Binti Mat Nawi for their pray, support and encouragement for me to finish this final year project. Lastly, I want to say a very thank you to all my friends especially to Muhammad Afandi, Lili Amira Hazinan and Nor Asmah Ismail for their helping and accompany me during handling this project. Without their support and advice I will not able to finish this project in the given period.

ABSTRACT

Ganoderma boninense is a fungus that is pathogenic on oil palms causing basal stem rot and most destructive disease of oil palm in South-East Asia. Meanwhile, *Fusarium oxysporum* is a causal agent of *Fusarium* wilt which affects a broad range of plants including black pepper, cucumber, banana and tomato. This study determined the potential of antagonistic bacteria that inhibited against *G.boninense* and *F.oxysporum* growth. Five types of bacteria isolates, BLH, B38, B40, B41 and B43 were used for screening of their antagonistic activity against *G.boninense* and *F.oxysporum*. The bacteria isolates were screened for *in vitro* antagonistic activity towards both fungal, *G.boninense* and *F.oxysporum* through dual culture and culture filtrate test/ diffusible antibiotic. The result showed that all of the bacteria isolates inhibited the growth of *F.oxysporum* and *G.boninense* with percentage of inhibition radial growth (PIRG) values ranging from 47.57 to 63.78% and 38.8 to 62.0%. Isolates B43 showed highest antagonism against *F.oxysporum* and *G.boninense* mycelia with 83.19 and 97.37% in culture filtrate test. The antifungal activities apparently involved the secretion diffusible bioactive compounds. Information obtained from this study show the potential of B43 as biocontrol agent against *G. boninense* and *F.oxysporum*. It might be an economical as well as environmental save way to suppress the diseases.

ABSTRAK

Ganoderma boninense adalah kulat yang patogenik terhadap kelapa sawit yang menyebabkan reput pangkal batang dan merupakan penyakit yang paling merosakkan kelapa sawit di Asia Tenggara. Sementara itu, *Fusarium oxysporum* adalah agen penyebab bagi penyakit layu *Fusarium* yang memberi kesan penyakit kepada pelbagai tumbuhan termasuk lada hitam, timun, pisang dan tomato. Kajian ini adalah untuk menentukan bakteria antagonis yang berpotensi untuk merencatkan pertumbuhan *G.boninense* dan *F.oxysporum*. Lima jenis pencilan bakteria iaitu BLH, B38, B40, B41 dan B43 telah digunakan untuk menguji aktiviti antagonis terhadap *G.boninense* dan *F.oxysporum*. Setiap bakteria telah disaring bagi aktiviti antagonis secara vitro terhadap kedua-dua kulat melalui kultur dual dan ujian turasan kultur/antibiotik resap. Hasil menunjukkan bahawa semua bakteria telah merencatkan pertumbuhan *F.oxysporum* dan *G.boninense* dengan peratusan pertumbuhan perencatan jejari (PIRG) nilai-nilai diantara 47,57-63,78% dan 38,8-62,0%. Bacteria B43 menunjukkan aktiviti antagonis yang paling tertinggi terhadap miselia *F.oxysporum* dan *G.boninense* dengan 83,19 dan 97.37% bagi ujian turasan kultur. Aktiviti antikulat menunjukkan penglibatan rembesan resap sebatian bioaktif. Maklumat yang diperolehi daripada kajian ini menunjukkan bahawa potensi B43 sebagai agen kawalan biologi terhadap *G. boninense* dan *F.oxysporum*. Ia mungkin menjadi lebih ekonomik terhadap alam sekitar dan merupakan cara yang menjimatkan untuk menyekat penyakit.

TABLE OF CONTENTS

	PAGES
ACKNOWLEDGEMENT	i
ABSTRACT	ii
ABSTRAK	iii
TABLE OF CONTENTS	iv
LIST OF TABLES	vii
LIST OF FIGURES	viii
LIST OF PLATES	ix
LIST OF APPENDICES	x
INTRODUCTION	1
LITERATURE REVIEW	
2.1 Oil Palm Disease Cause by <i>Ganoderma boninense</i>	3
2.2 <i>Ganoderma boninense</i>	4
2.3 Control of <i>Ganoderma boninense</i>	5
2.3.1 Chemical Control of <i>G.boninense</i>	5
2.3.2 Biological Control of <i>G. boninense</i>	6
2.4 <i>Fusarium</i> sp.	7
2.5 Modes of Existence	7
2.6 Diseases Caused by <i>Fusarium oxysporum</i>	8
2.7 Control of <i>F. oxysporum</i>	10
2.7.1 Biological Control	11

MATERIALS AND METHODS

3.1	Experiment Location	13
3.2	<i>Fusarium oxysporum</i> Culture	13
3.3	<i>Ganoderma boninense</i> Culture	14
3.4	Bacteria Isolates	14
3.5	<i>In Vitro</i> Screening Bacteria Against <i>G. boninense</i> and <i>F. oxysporum</i>	16
3.6	Antagonism	
	3.6.1 Dual Culture Assay	16
	3.6.2 Preparation of Bacterial Suspension	17
	3.6.3 Culture Filtrate Test/ Non Volatile Antibiotics	18
	3.6.4 Mycelial Growth Test	18
	3.6.5 Spore Germination Test	19
3.7	Identification of Bacteria	20
3.8	Statistical Analysis	21

RESULTS AND DISCUSSION

4.1	Dual Culture assay	
	4.1.1 <i>F.oxysporum</i>	22
4.2	Culture Filtrate Test / Non Volatile Antibiotic	25
4.3	Antagonistic Activity Isolate Bacteria for <i>Ganoderma boninense</i>	28

CONCLUSIONS AND RECOMMENDATIONS	32
REFERENCES	33
APPENDICES	x



© COPYRIGHT UPM

LIST OF TABLE

Table		Pages
Table 1	Antagonistic Potential of Bacteria Isolates in Dual Culture Against <i>F.oxysporum in vitro</i>	24
Table 2	Antagonistic Potential of Bacteria Isolates in Culture Filtrate Test Against <i>F.oxysporum in vitro</i>	26
Table 3	Antagonistic Potential of Bacteria Isolates in Dual Culture Against <i>G.boninense in vitro</i>	29

LIST OF FIGURES

Figures		Pages
Figure 1	Dual Culture Test. Antagonistic Interaction Between BLH, B38, B40, B41, B43 and <i>F.oxysporum</i> on PDA after Incubation at 28±°C	23
Figure 2	Growth rate of <i>F.oxysporum</i> in Dual Culture Assay	24
Figure 3	Effects of BLH, B38, B40, B41, B43 on the Radial Growth of <i>F.oxysporum</i> In Culture Filtrate Test at Seven Days Incubation.	27
Figure 4	Dual Culture Test. Antagonistic Interaction Between B41 and B43 With <i>G.boninense</i> on PDA after Incubation at 28±°C	29
Figure 5	Effect of B43 on the radial growth of <i>G.boninense</i> in Culture Filtrate test at Seven Days	30
Figure 6	Mycelial Growth Test	30
Figure 7	Observation of Mycelial abnormalities of <i>G.boninense</i> at 400 Magnification as Treated with B43	31

LIST OF APPENDICES

	Pages
Appendix 1 Radial Growth of <i>F.oxysporum</i> by Dual Culture Assay Produced By Antagonistic Bacteria	x
Appendix 2 Radial Growth of <i>G. boninense</i> by Dual Culture Assay Produced by Antagonistic Bacteria	xii
Appendix 3 Diameter Growth Inhibition of <i>F.oxysporum</i> by Culture Filtrate Test/Non Volatile Antibiotics Test Produced by Antagonistic Bacteria	xiii
Appendix 4 Diameter Growth Inhibition of <i>G. boninense</i> by Culture Filtrate Test/ Non Volatile Antibiotics Test Produced by Antagonistic Bacteria	xv
Appendix 5 ANOVA for dual culture of <i>F.oxysporum</i>	xvi
Appendix 6 ANOVA for culture filtrate test of <i>F.oxysporum</i>	xviii



CHAPTER 1

INTRODUCTION

Biological control of plants diseases offers a safer and ecologically acceptable alternative to chemical control. The introduction and establishment of living natural enemies of pathogens is viewed as a powerful alternative to chemical fungicides conventionally used in agriculture, satisfying the preference of consumers for sustainable products with few or no chemical residues (Gnanamanickam *et al.*, 2002). A variety of promising biological control agents belonging to various genera of bacteria such as *Bacillus*, *Pseudomonas*, *Agrobacterium*, *Burkholderia*, other than fungi such as *Trichoderma* have been isolated, characterized and proposed for practical applications and some have been registered as biopesticides in order to reduce biotic damage due to soil borne diseases (Pane *et al.*, 2012). There are many studies that reported the potential of *Bacillus* species as biocontrol agent that inhibit fungal growth. The *Bacillus* spp. produce endospores that are resistant to desiccation, heat, UV irradiation and organic solvents. The qualities make them more resistant to adverse weather conditions.

Fusarium wilt is a fungal disease which affects a broad range of plants such as tomato, banana (*Musa* spp.), cucumber and black pepper (*Piper nigrum*). *Fusarium oxysporum* infects commercial crops such banana (Mohammad *et al.*, 2011), chickpea (Kaur *et al.*, 2007), cumin (Haggag and Abo-Sedera, 2005), kidney bean (El-Mehalawy, 2004) and strawberry (Hyeon *et al.*, 2009) and tomato (Chandel *et al.*, 2009). The soil-borne tomato root-infecting pathogens *Fusarium oxysporum* f.sp.*lycopersici* is particularly difficult to control using standard cultural and chemical methods (Chandel *et*

al., 2009). Wilt resistant varieties of tomato are available, but in the past resistance have been overcome by the appearance of new races of the pathogen, a continuous problem in crop production (Hausbeck and Lamour, 2004). Growing awareness of the potential hazards in using agrochemicals has led to increased investment in research on alternative methods for effective disease control.

Elaeis guineensis Jacq., which is commonly known as the oil palm is the most crucial species in the genus *Elaeis* which belongs to the family *Palmae*. Oil palm is truly “a golden crop of Malaysia” since it generates profitable export earnings for the country and as a nature gift for poverty in Malaysia (Basiron, 2007). According to Principle Statistic of Oil Palm Malaysia, in 2010 the planted area for oil palm in Malaysia is 4.202 million hectares. Production increased from 94000 tones in 1960 to 15 million tones in 2005, or almost 160 times within 45 years. This represents a compound annual growth of 11.93 % per year (Basiron, 2007). Plant health is crucial in obtaining maximum production. In Malaysia, Basal Stem Rot caused by *Ganoderma boninense* is attacking the oil palm. With no known cure at present, it is the major disease of oil palm and therefore of great economic importance to the Malaysian oil palm industry (Sapak *et al*, 2008).

The objective of this study is to screen the bacteria that can inhibit the growth of *Ganoderma boninense* and *Fusarium oxysporum*.

REFERENCES

- Abdul Razak, J., Ahmad, H., Ramdhan, K., Idris, A., Abdul Rahim, S., Aminul, R., & Fauzi, I. (2004). Mechanical trunk injection for control of *Ganoderma*. *MPOB Information Series*, MPOB TT No. 215.
- Ahmed Idris, H., N. Labuschagne and L. Korsten, 2007. Screening rhizobacteria for biological control of *Fusarium* root and crown rot of sorghum in Ethiopia. *Biologi. Cont.*, 40: 97–106
- Ahn, P., Chung, H. S., & Lee, Y. H. (1998). Vegetative compatibility groups and pathogenicity among isolates of *Fusarium oxysporum* f. sp. *cucumerinum*. *Plant Disease*, 82(2), 244-246
- Alabouvette, C., Olivain, C., Migheli, Q., & Steinberg, C. (2009). Microbiological control of soil-borne phytopathogenic fungi with special emphasis on wilt-inducing *fusarium oxysporum*. *New Phytologist*, 184(3), 529-544.
- Anandaraj, M. (2000). Diseases of black pepper. *Black Pepper Piper Nigrum*, , 239-267.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109(4), 289-295.
- Bivi, M. R., Farhana, M., Khairulmazmi, A., & Idris, A. (2010). Control of *Ganoderma boninense*: A causal agent of basal stem rot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture and Biology*, 12(6), 833-839.
- Burgess, L. (1981). General ecology of the fusaria. in ‘*Fusarium*: Diseases, biology, and taxonomy’. eds P.E. Nelson, T.A. Toussoun, R.J. Cook) pp. 225–235.
- Chandel, S., Allan, E. J., & Woodward, S. (2010). Biological control of *Fusarium oxysporum* f.sp. *lycopersici* on tomato by *Brevibacillus brevis*. *Journal of Phytopathology*, 158(7-8), 470-478.

- Chérif, M., & Benhamou, N. (1990). Cytochemical aspects of chitin breakdown during the parasitic action of a *Trichoderma* sp. on *Fusarium oxysporum* f. sp. *radicis-lycopersici*. *Phytopathology*, 80(12), 1406-1414.
- Chung, G., Darus, A., & Sukaimi, J. (1991). Preliminary results on trunk injection of fungicides against *Ganoderma* basal stem rot in oil palm. *Proceedings of Ganoderma Workshop, Bangi, Selangor, Malaysia, 11 September 1990*. 81-97.
- Cook, R. J. (1993). Making greater use of introduced microorganisms for biological control of plant pathogens. *Annual Review of Phytopathology*, 31(1), 53-80.
- Corley, R., & Tinker, P. (2003). The classification and morphology of the oil palm. *The Oil Palm*, pp. 27-50.
- Dharmaputra, O. S., Tjitrosomo, H. S., & Abadi, A. (1989). Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. *Biotropia*, (3), 41-49.
- Edward, E. J., King, W. S., Teck, S. L. C., Jiwan, M., Aziz, Z. F. A., Kundat, F. R., Ahmed, H. A., Majid, N. M. A (2013). Antagonistic activities of endophytic bacteria against *Fusarium* wilt of black pepper (*Piper nigrum*). *International Journal of Agriculture Biology*, (15), 291-296.
- El-Mehalawy, A. A. (2004). The rhizosphere yeast fungi as biocontrol agents for wilt disease of kidney bean caused by *Fusarium oxysporum*., *International Journal of Agriculture Biology*, (6), 310-316.
- Fravel, D., Olivain, C., & Alabouvette, C. (2003). *Fusarium oxysporum* and its biocontrol. *New Phytologist*, 157(3), 493-502.

- Gnanamanickam, S. S., Vasudevan, P., Reddy, M. S., Defago, G., & Kloepper, J. (2002). Principles of biological control. *Biological Control of Crop Diseases*, pp, 1-9. New York. Marcel Dekker.
- Haggag, W. M., & Abo-Sedera, S. (2005). Characteristics of three *Trichoderma* species in peanut haulms compost involved in biocontrol of cumin wilt disease. *International Journal of Agriculture Biology*, (7), 222-229.
- Hausbeck, M. K., & Lamour, K. H. (2004). *Phytophthora capsici* on vegetable crops: Research progress and management challenges. *Plant Disease*, 88(12), 1292-1303.
- Hyeon, N., Park, M. S., Kim, H. G., & Yoo, S. J. (2009). Biological control of strawberry *Fusarium* wilt caused by *Fusarium oxysporum* f. sp. *fragariae* using *Bacillus velezensis* BS87 and RK1 formulation. *Journal of Microbiology and Biotechnology*, 19(5), 520-524.
- Kaur, R., Singh, R., & Alabouvette, C. (2007). Antagonistic activity of selected isolates of fluorescent *Pseudomonas* against *Fusarium oxysporum* f. sp. *ciceri*. *Asian Journal of Plant Sciences*, 6, 446-454.
- Khairudin, H., Darus, A., & Sukaimi, J. (1991). Results of four trials on *Ganoderma* basal stem rot of oil palm in golden hope estates. *Proceedings of Ganoderma Workshop, Bangi, Selangor, Malaysia, 11 September 1990. pp*, 67-80.
- Lemanceau, P., & Alabouvette, C. (1991). Biological control of *Fusarium* diseases by fluorescent *Pseudomonas* and non-pathogenic *Fusarium*. *Crop Protection*, 10(4), 279-286.
- Li, L., Ma, J., Li, Y., Wang, Z., Gao, T., & Wang, Q. (2012). Screening and partial characterization of *Bacillus* with potential applications in biocontrol of cucumber *Fusarium* wilt. *Crop Protection*, 35(0), 29-35.

- Meon, S. (1997). Antagonistic effect of Malaysian isolates of *Trichoderma harzianum* and *gliocladium virens* on *sclerotium rolfsii*. *Pertanika Journal of Tropical Agricultural Science*, 20(1), 35-41.
- Mohammed, A. M., AL-Ani, L. K., Bekbayeva, L., & Salleh, B. (2011). Biological control of *Fusarium oxysporum* f. sp. *cubense* by *Pseudomonas fluorescens* and BABA in vitro. *World Applied Sciences Journal*, 15(2), 189-191.
- Nel, B., Steinberg, C., Labuschagne, N., & Viljoen, A. (2006). The potential of nonpathogenic *Fusarium oxysporum* and other biological control organisms for suppressing *Fusarium* wilt of banana. *Plant Pathology*, 55(2), 217-223.
- Pane, C., Villecco, D., Campanile, F., & Zaccardelli, M. (2012). Novel strains of bacillus, isolated from compost and compost-amended soils, as biological control agents against soil-borne phytopathogenic fungi. *Biocontrol Science and Technology*, 22(12), 1373-1388.
- Paterson, R. R. M. (2000). Ganoderma. A therapeutic fungal biofactory. *Phytochemistry*, 67(18), 1985-2001.
- Paterson, R. R. M. (2006). Fungi and fungal toxins as weapons. *Mycological Research*, 110(9), 1003-1010.
- Paterson, R. R. M. (2007). Ganoderma disease of oil palm—A white rot perspective necessary for integrated control. *Crop Protection*, 26(9), 1369-1376.
- Pavlou, G. C., & Vakilounakis, D. J. (2005). Biological control of root and stem rot of greenhouse cucumber, caused by *Fusarium oxysporum* f. sp. *radicis-cucumerinum*, by lettuce soil amendment. *Crop Protection*, 24(2), 135-140.
- Rahman, M., Kadir, J., Mahmud, T., Rahman, R. A., & Begum, M. (2007). Screening of antagonistic bacteria for biocontrol activities on *Colletotrichum gloeosporioides* in papaya. *Asian Journal of Plant Sciences*, (6), 12-20.

- Rolph, H., Wijesekara, R., Lardner, R., Abdullah, F., Kirk, P., Holderness, M., Flood, J. (2000). Ganoderma isolates from oil palm, coconut and betelnut. *Ganoderma disease of perennial crops*. CABI Publishing, Wallingford, UK, pp. 205-221.
- Sanderson, F., Pilotti, C., & Bridge, P. (2000). Rot of oil palm. *Ganoderma Diseases of Perennial Crops*,. CABI Publishing, Wallingford, UK, pp. 113-119.
- Sapak, Z., Meon, S., Ahmad, M., & Abidin, Z. (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture & Biology*, 10(2), 127-132.
- Sariah, M., 1994. Potential of *Bacillus* spp. as a biocontrol agent for anthracnose fruit rot of chilli. *Malays. Applied Biol.*, 23: 53-60.
- Sariah, M. (2003). The potential of biological management of basal stem rot of oil palm: Issues, challenges and constraints. *Oil Palm Bulletin*, 47, 1-5.
- Sariah, M., & Zakaria, H. (2000). Oil-palm seedlings. *Ganoderma Diseases of Perennial Crops*. CABI Publishing, Wallingford, UK, pp. 89-99.
- Shanmugam, V., & Kanoujia, N. (2011). Biological management of vascular wilt of tomato caused by *Fusarium oxysporum* f.sp. *lycospersici* by plant growth-promoting rhizobacterial mixture. *Biological Control*, 57(2), 85-93.
- Sharples, A. (1928). Palm diseases in malaya. *Malayan Agricultural Journal*, 16(9-10), 313-360.
- Siddiquee, S., Yusuf, U. L, Hossain, K., and Jahan, S (2009). *In vitro studies on the potential Trichoderma harzianum for antagonistic properties against Ganoderma boninense*. *Journal of Food, Agriculture & Environment* , 7 (3-4). pp. 970-976.
- Singh, G. (1991). Ganoderma-the scourge of oil palms in the coastal areas. *The Planter*, 67: 421-444.

- Singh, G., Darus, A., & Sukaimi, J. (1991). Ganoderma-the scourge of oil palm in the coastal area. *Proceedings of Ganoderma Workshop, Bangi, Selangor, Malaysia, 11 September 1990*. pp. 7-35. Bangi: Palm oil Research Institute of Malaysia.
- Soepena, H., Purba, R., & Pawirosukarto, S. (2000). A control strategy for basal stem rot (Ganoderma) on oil palm. *Ganoderma Diseases of Perennial Crops*. CABI Publishing, Wallingford, UK pp. 83-88.
- Suárez-Estrella, F., Vargas-García, C., López, M. J., Capel, C., & Moreno, J. (2007). Antagonistic activity of bacteria and fungi from horticultural compost against *fusarium oxysporum* f. sp. *melonis*. *Crop Protection*, 26(1), 46-53.
- Susanto, A., Sudharto, P., & Purba, R. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159(1), 153-157.
- Suryanto, D., Wibowo, R. H., Siregar, E. B. M., & Munir, E. (2012). A possibility of chitinolytic bacteria utilization to control basal stems disease caused by *Ganoderma boninense* in oil palm seedling. *African Journal of Microbiology Research*, 6(9), 2053-2059.
- Vakalounakis, D. J., Wang, Z., Fragkiadakis, G. A., Skaracis, G. N., & Li, D. B. (2004). Characterization of *Fusarium oxysporum* isolates obtained from cucumber in China by pathogenicity, VCG, and RAPD. *Plant Disease*, 88(6), 645-649.
- Ting, A., Sariah, M., Jugah, K., & Amar, A. (2003). Potential use of suppressive soil in managing *Fusarium* wilt of banana seedlings. *Infomusa*, 12, 33-34.
- Yang, T., & Dai, C. (2013). Interactions of two endophytic fungi colonizing *Atractylodes lancea* and effects on the host's essential oils. *Acta Ecologica Sinica*, 33(2), 87-93.