

# **UNIVERSITI PUTRA MALAYSIA**

EVALUATION OF POTENTIAL PROBIOTIC BACTERIA FOR MICROALGAE PROPAGATION AND Artemia fransiscana (Kellog, 1906) BIOENCAPSULATION

NUR NATASYA AIN BT ROSLAND

IB 2018 37



### EVALUATION OF POTENTIAL PROBIOTIC BACTERIA FOR MICROALGAE PROPAGATION AND Artemia fransiscana (Kellog, 1906) BIOENCAPSULATION



NUR NATASYA AIN BT ROSLAND

Thesis Submitted to School of Graduate Studies, University Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

November 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

 $\mathbf{G}$ 



Abstract of thesis prepared to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

### EVALUATION OF POTENTIAL PROBIOTIC BACTERIA FOR MICROALGAE PROPAGATION AND Artemia fransiscana (Kellog, 1906) BIOENCAPSULATION

By

### NUR NATASYA AIN BT ROSLAND

November 2018

### Chairman : Murni Marlina Bt Abd Karim, PhD Faculty : Institute of Bioscience

The emergence of aquaculture industry is in need for non-antibiotic based disease control approach to minimize risk of antibiotic-resistance bacteria. Bacterial infections mainly caused by *Vibrio* spp. have caused mass mortalities to fish especially during larval stages. The potential of microalgae as future feedstocks in aquaculture sector has prompted to a new area of research in finding potential probiotics that can enhance algal biomass and at the same time, suppress fish larval pathogens. The current research was undertaken to discover new potential probiont strains isolated from microalgae and elucidate their interaction. The capability of the potential probionts in protecting *Artemia* from *Vibrio* spp. was also determined in this study.

A total of 27 strains were successfully isolated from different species of fresh microalgae cultures (*Amphora* sp., *Chaetoceros* sp., *Chlorella* sp., *Nannochlorum* sp. and *Spirulina* sp.). All the isolated strains were screened for antibacterial activity against fish pathogens, *Vibrio harveyi* and *Vibrio parahaemolyticus* in *in vitro* study. Results of the agar well diffusion assay showed that 4 out of 27 strains were able to inhibit the growth of *V. harveyi* and *V. parahaemolyticus*. Strains labeled as A-1, A-2 and Cl-3 showed inhibition towards both pathogens tested while strain S-2 showed inhibition towards *V. parahaemolyticus* only. These four potential probionts were identified as *Lysinibacillus fusiformis* strain A-1 (isolated from *Amphora* sp.), *Bacillus* sp. strain A-2 (isolated from *Amphora* sp.), *Lysinibacillus fusiformis* strain Cl-3 (isolated from *Chlorella* sp.) and *Bacillus pocheonensis* strain S-2 (isolated from *Spirulina* sp.) using 16s rRNA molecular method.

Biochemical assay demonstrated that all these probionts had the ability in excreting catalase, gelatinase, lipase, oxidase, protease, and anti-quorum sensing. In biofilm

inhibition assay, the highest percentage of inhibition of *V. harveyi* and *V. parahaemolyticus* were demonstrated by *B. pocheonensis* strain S-2 (70.36 ± 4.84 %) and *L. fusiformis* strain A-1 (67.79 ± 4.70 %), respectively. Through microalgae propagation experiment, the inoculation ratio of bacteria and microalgae was fixed to 1:4, and co-culture incubation was monitored for 7 to 8 days. *Bacillus* sp. strain A-2 and *L. fusiformis* strain Cl-3 were able to promote the growth of *Amphora* sp., and *Chlorella* sp., respectively. *Bacillus* sp. strain A-2 showed a good correlation after co-cultured with *Amphora* sp. evidenced by a steep growth of *Amphora* sp. (8.5 x 10<sup>5</sup> cells ml<sup>-1</sup>) in comparison with the control (1.4 x 10<sup>5</sup> cells ml<sup>-1</sup>) ) at day 8 of co-culture. The cell density of *Chlorella* sp. was promoted more than two times due to *L. fusiformis* strain Cl-3 supplementation (9.9 x 10<sup>6</sup> cells ml<sup>-1</sup>) compared with the control (4.2 x 10<sup>6</sup> cells ml<sup>-1</sup>), observed at day 7 of co-culture incubation.

A preliminary *in vivo* assay was carried out on *Artemia fransiscana* nauplii against *V*. *harveyi* and *V*. *parahaemolyticus*. Probionts were supplied to *Artemia* as single strain and in multiple strains known as MIX (combinations of all potential probionts) along with respective microalgae. Results revealed that *Artemia* bioencapsulated with MIX + *Amphora* sp. showed the highest survival rate  $(60 \pm 4\%)$  when compared with the control  $(13 \pm 2\%)$ , after challenged with *V*. *harveyi*. A significant reduction of vibrios load was observed in *Artemia* bioencapsulated with MIX + *Amphora* sp.  $(7.29 \pm 0.02 \text{ CFU ml}^{-1})$  compared with the control  $(7.31 \pm 0.05 \text{ CFU ml}^{-1})$ .

Meanwhile, for the Artemia challenged with V. parahaemolyticus, a significant survival was observed in Artemia bioencapsulated with L. fusiformis strain A-1 + Amphora sp.  $(78 \pm 2 \%)$ ; Bacillus sp. strain A-2 + Amphora sp.  $(78 \pm 1 \%)$ ; L. fusiformis strain Cl-3 + Chlorella sp.  $(78 \pm 2 \%)$  and MIX + Amphora sp.  $(78 \pm 2 \%)$  in comparison with the control  $(32 \pm 1 \%)$ . The number of vibrios in Artemia bioencapsulated with B. pocheonensis strain S-2 + Spirulina sp. showed significant reduction  $(7.16 \pm 0.09 \text{ CFU ml}^{-1})$  after challenged with V. parahaemolyticus. In conclusion, the present study proposed that all the potential probionts isolated from microalgae exhibit essential probiotics properties which worth to be studied for application in aquaculture. Two potential probionts (Bacillus sp. strain A-2 and L. fusiformis strain Cl-3) were found to be good promoting bacteria for microalgal growth. Moreover, mixture of all potential probionts is beneficial for Artemia bioencapsulation in conferring protection to Artemia nauplii against pathogenic vibrios in the culture environment.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

### PENILAIAN BAKTERIA BERPOTENSI YANG MEMPUNYAI CIRI-CIRI PROBIOTIK UNTUK PERTUMBUHAN MIKROALGA DAN PENGKAYAAN DIDALAM Artemia fransiscana (Kellog, 1906)

Oleh

#### NUR NATASYA AIN BT ROSLAND

November 2018

### Pengerusi : Murni Marlina Bt Abd Karim, PhD Fakulti : Institut Biosains

Kepesatan industri akuakultur memerlukan satu pendekatan dalam usaha mengawal penyakit yang tidak berasaskan antibiotik bagi mengurangkan risiko bakteria yang mempunyai daya tahan yang tinggi terhadap antibiotik. Jangkitan bakteria yang disebabkan oleh spesies *Vibrio* telah menyebabkan kematian yang tinggi terhadap ikan terutama sewaktu peringkat larva. Potensi microalga sebagai stok makanan pada masa hadapan dalam sektor akuakultur telah mendorong kepada bidang penyelidikan baru dalam mencari probiotik yang berpotensi bagi meningkatkan biojisim algal dan pada masa yang sama dapat mengurangkan patogen yang biasa menyerang larva ikan. Penyelidikan semasa dijalankan untuk mengkaji potensi baru probiotik dari microalga dan interaksinya dengan microalga. Kemampuan probiotik tersebut dalam melindungi *Artemia* dari *Vibrio* spp. turut dijalankan.

Sejumlah 27 strain bakteria yang berbeza telah berjaya diasingkan dari pelbagai spesies mikroalga (*Amphora* sp., *Chaetoceros* sp., *Chlorella* sp., *Nannochlorum* sp. dan *Spirulina* sp.). Bakteria tersebut telah menjalani ujian saringan *in vitro* dalam usaha mengenalpasti bakteria yang mempunyai kebolehan menghalang pertumbuhan patogen, *Vibrio harveyi* dan *Vibrio parahaemolyticus*. Hasil ujian *in vitro* menunjukkan bahawa 4 dari 27 strain mampu merencatkan pertumbuhan V. *harveyi* dan V. *parahaemolyticus*. Strain A-1, A-2 dan Cl-3 mampu menghalang pertumbuhan kedua - dua patogen yang diuji manakala strain S-2 tidak menunjukkan sebarang perencatan terhadap V. *harveyi*. Keempat-empat bakteria berpotensi ini dikenalpasti sebagai strain Lysinibacillus fusiformis A-1 (diasingkan dari *Amphora* sp.), *Bacillus* sp. strain A-2 (diasingkan dari *Amphora* sp.), *Lysinibacillus fusiformis* strain Cl-3 (diasingkan dari *Chlorella* sp.) dan *Bacillus pocheonensis* (diasingkan daripada *Spirulina* sp.) menggunakan 16s rRNA.

Ujian biokimia menunjukkan bahawa semua bakteria yang berpotensi sebagai probiotik mempunyai kemampuan dalam merembeskan enzim catalase, gelatinase, lipase, oxidase,

protease, dan anti-kuorum. Ujian perencatan biofilem menunjukkan bahawa B. pocheonensis strain S-2 mampu merencatkan V. harveyi paling tinggi  $(70.36 \pm 4.84 \%)$ manakala V. parahaemolyticus direncatkan oleh L. fusiformis strain A-1, dengan kadar perencatan tertinggi iaitu  $67.69 \pm 4.70$  %. Melalui eksperimen pertumbuhan mikroalga, nisbah inokulasi bakteria dan mikroalga telah ditetapkan pada 1:4, dan inkubasi bersama dijalankan selama 7 hingga 8 hari. Bacillus sp. strain A-2 dan L. fusiformis strain Cl-3 dapat meningkatkan pertumbuhan Amphora sp. dan Chlorella sp. masing-masing. Ini dibuktikan dengan peningkatan sel densiti yang tinggi oleh Amphora sp. (8.5 x 10<sup>5</sup> sel ml<sup>-1</sup>) berbanding dengan kawalan (1.4 x 10<sup>5</sup> sel ml<sup>-1</sup>), dilihat pada hari ke-8 pengkulturan bersama. Sel Chlorella sp. telah meningkat lebih dua kali ganda disebabkan oleh L. fusiformis strain Cl-3 (9.9 x 10<sup>6</sup> sel ml<sup>-1</sup>) berbanding dengan kawalan (4.2 x  $10^6$  sel ml<sup>-1</sup>), diperhatikan pada hari ketujuh pengkulturan bersama. Ujian *in vivo* telah dilakukan terhadap nauplii Artemia fransiscana menentang V. harveyi dan V. parahaemolyticus. Bakteria berpotensi ini dibekalkan kepada Artemia dalam satu strain tunggal dan kombinasi semua strain yang terpilih, dikenali sebagai MIX bersama-sama dengan mikroalga. Keputusan in vivo menunjukkan bahawa Artemia yang diperkayakan dengan MIX + Amphora sp.  $(60 \pm 4\%)$  menunjukkan kadar kelangsungan hidup tertinggi berbanding dengan kumpulan kawalan (13  $\pm$  2 %). Penurunan yang ketara dalam bilangan vibrio diperhatikan dalam Artemia yang telah diperkayakan dengan MIX + Amphora sp.  $(7.29 \pm 0.02 \text{ CFU ml}^{-1})$  berbanding dengan kumpulan kawalan  $(7.31 \pm 0.05)$  $CFU ml^{-1}$ ).

Dalam kes Artemia yang dicabar dengan V. parahaemolyticus, kelangsungan hidup yang ketara diperhatikan dalam Artemia yang diperkayakan dengan L. fusiformis strain A-1 + Amphora sp.  $(78 \pm 2 \%)$ ; Bacillus sp. strain A-2 + Amphora sp.  $(78 \pm 1 \%)$ ; L. fusiformis strain Cl-3 + Chlorella sp.  $(78 \pm 2 \%)$  dan MIX + Amphora sp.  $(78 \pm 2 \%)$  berbanding dengan kumpulan kawalan  $(32 \pm 1 \%)$ . Bilangan vibrio di dalam Artemia yang diperkayakan dengan B. pocheonensis strain S-2 + Spirulina sp. juga menunjukkan penurunan yang ketara  $(7.16 \pm 0.09 \text{ CFU ml}^{-1})$ , selepas dicabar oleh V. parahaemolyticus. Kesimpulannya, kajian ini menyarankan agar keempat-empat bakteria berpotensi yang diasingkan dari mikrolaga ini dikaji dengan lebih mendalam kerana ia berpotensi dijadikan probiotik untuk kegunaan dalam akuakultur. Dua bakteria berpotensi (Bacillus sp. strain A-2 dan L. fusiformis strain Cl-3) didapati mempunyai kebolehan dalam meningkatkan pertumbuhan mikroalga. Kajian ini mencadangkan bahawa kombinasi semua probiotik terpilih boleh menjadi bahan yang baik dalam memperkayakan Artemia dan melindunginya dari patogenik Vibrio didalam kultur persekitaran tersebut.

#### ACKNOWLEDGEMENTS

#### In the Name of Allah the Most Beneficent, the Most Merciful

The ultimate gratitude, devotion and dedication surely belongs to Allah SWT. Alhamdulillah for granting us safe passage through the vicissitude of life to see this day in a great state of iman, a'mal and health. Alhamdulillah for providing me the strength, ability and opportunity to undertake this research study and to complete it successfully. Without His blessings and help, this achievement would not have been possible.

I would like to acknowledge the vast contributions and amazing spirit of teamwork, diligence, sharing and kindness of many wonderful people. In my journey, I have found a supervisor, a teacher, an inspiration, and a pillar of support, Dr. Murni Marlina Abd Karim. She has been there providing her support and guidance at all times and has given me invaluable inspiration, suggestions and guidance in my research journey. She has given me all the freedom to pursue my research but at the same time always keep me on track and reminds me to graduate on time. Without her excellent guidance and patience, this thesis would not have been possible and I shall be grateful having her as my supervisor. May Allah SWT ease your journey as an academician and keep being a great source of inspirations for all students.

I would also like to express my gratitude to my committee members, Dr. Natrah Fatin Mohd Ikhsan and Dr. Chong Chou Min for sparing their valuable time whenever I approached them and showing me the way ahead. A big thank also goes to my internal examiner, Dr. Suriana Sabri for her big contribution in correcting this thesis.

I take pride in acknowledging the insightful guidance of my labmates, Jasmin, Fathiah, Puvi and Sofea for being there at all times assisting me in collation of data for my research. Their support, credible ideas and motivation have been great contributors in the completion of this thesis. I would like to thank entire staff of Aquaculture Department, UPM especially Mrs. Shafika who has been so helpful and cooperative in giving her support at all times to help me accomplish my research.

My acknowledgement would be incomplete without thanking my colleague; Aini, Arina, Izzah, Atikah, Ifa, Laisha, Atiyah, Ain, Syuhada, Jannah, Nisa, Dilla, Nur Ain, Sarmila, and Sheri in ensuring that the fighting spirit keeps burning, keep me going on my path to success and being there in every single time I required motivation and moral support. A very special word of thanks goes for my bestfriend, A who has always been a major source of support, help me countless time just to ensure I can finish everything on time, and always being there when I needed someone to talk to. I will forever be grateful for having all of you as my friends.

I would like to dedicate this piece of work to my family members for being the biggest source of my strength. The blessings of my mother, Mrs. Fadilah and the love of my

siblings, Azwa Afifie, Aliff Asyraf, Khoirun Nisa' and my silent supporter would be my aunt, Miss Zurina and my grandparents, Mr. Ishak, Mrs. Robiah and Mrs. Aminah. All of them have made a tremendous contribution in helping me reach this stage in my life. I am indebted to them for always stay with me in thick and thin and lift me up just to make sure that I can achieve my dreams. This would not have been possible without their love and support given to me through this entire journey.

Thank you everyone!

G



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

### Murni Marlina Abd Karim, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

### Natrah Fatin Mohd Ikhsan, PhD Associate Professor

Faculty of Agriculture Universiti Putra Malaysia (Member)

### Chong Chou Min, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

# ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date :

### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisors and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed, or in electric form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature :	Date :	
<i>c</i> =		

Name and Matric No. : Nur Natasya Ain Binti Rosland, GS47713

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

C)

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	PM 1831
Signature: Name of Member of Supervisory Committee:	

### TABLE OF CONTENTS

				Page
	STRAC			1
	STRAK KNOW	VLEDGEMENTS		iii v
-	ROV			vii
		ATION		viii
		ΓABLES		xv
		FIGURES		xvi
LIS	T OF A	ABBREVIATIONS		xviii
СН	АРТЕІ	R		
1	INT	RODUCTION		1
	1.1	Background of the study		1
	1.2	Problem statement		3
	1.3	Significant of study		3
	1.4	Objectives of study		4
	1.5	Hypothesis of study		4
2		ERATURE REVIEW		5
	2.1	Aquaculture status		5
	2.2	Bacterial diseases in aquaculture		5
	2.3	Fish health management		6
	2.4	Probiotics		7
		2.4.1 Probiotics as an alternative method		7
		2.4.2 Properties of probiotics		7
	2.5	2.4.3 Mechanisms of probiotics		8
	2.5	Community of bacteria in microalgae		9
	2.6 2.7	Microalgae and bacteria interaction Beneficial effect of live feed enriched with probiotics		9 12
	2.8	Importance of Artemia in aquaculture		12
	ISOI	LATION, SCREENING AND IDENTIFICATION	OF	14
3		LATION, SCREENING AND IDENTIFICATION TENTIAL BACTERIA FROM MICROALGAE	AS	14
		DBIOTICS		
	3.1	Introduction		14
	3.2	Materials and methods		15
		3.2.1 Culture and maintenance of microalgae		15
		3.2.2 Culture of pathogenic strains		15
		3.2.3 Isolation of potential bacteria isolated from microa	lgae	15
		3.2.3.1 Isolation process		15
			igae	-

- 3.2.3.2 Elimination of pathogenic strains 16
- 3.2.4 Preliminary *in vitro* screening of potential strains as 16 probiotics

 $\bigcirc$ 

		3.2.4.1	Agar well diffus	sion method		16
	3.2.5	Identific	ation of potential	probionts		16
		3.2.5.1	Gram staining			16
		3.2.5.2		ification using 16	S rRNA	gene 17
3.3	Result	s	unurjene			18
	3.3.1		n and purification	of potential prob	ionts fro	
	3.3.2		tion of pathogeni	c strains		18
	3.3.3		ng of antibacterial		tial prob	
		against	<i>Vibrio harveyi</i> an	d Vibrio parahae		ıs
	3.3.4		logy of potential			21
	3.3.5	Gram-st	taining of potentia	al probionts		22
	3.3.6	PCR Ar	nplification of 16	S rRNA gene seq	uence	24
	3.3 <mark>.</mark> 7	Molecu	lar identification	using 16S rRNA	gene ana	lysis 25
3.4	Discus	sion				26
3.5	Conclu	ision				28
4 DD	ODEDTI	EC OF		DROBIONES		<b>1756 20</b>
	OPERTI		POTENTIAL ROWTH OF M		AND	<b>ITS</b> 29
4.1	Introd			ICROALGAE		29
4.2		als and m	ethods			30
1.2	4.2.1		of probionts			30
	4.2.2		lular enzymes pro	duction		30
	7.2.2	4.2.2.1	Preparation of n			30
		4.2.2.2		liculu		30
		4.2.2.3				31
		4.2.2.4				31
			Lipase test			31
		4.2.2.6	Protease test			31
	4.2.3	Anti qu	orum-sensing effe	ects of potential p	robionts	31
	4.2.4	Biofilm	inhibition assay			32
	4.2.5		lgal growth condi	tions		32
	4.2.6		and bacterial conc			33
	4.2.7	Evaluat	tion of potential p llgae		rowth of	
		4.2.7.1	Evaluation of I	<i>Lysinibacillus fus</i> he growth of <i>Am</i>		33
		4.2.7.2		Bacillus sp. strain		
		4.2.7.3	Evaluation of L	<i>Lysinibacillus fusi</i> he growth of <i>Chl</i>	·	33
		4.2.7.4		Bacillus pocheone wth of Spirulina s		in 34
	4.2.3	Statistic	al analysis	•	-	34
4.3	Result		÷			35

		4.3.1 Extracellul	ar enzymes production	35
		4.3.1.1 An	nylase test	35
		4.3.1.2 Ca	talase test	35
		4.3.1.3 Ge	elatinase test	35
		4.3.1.4 Lip	pase test	37
		4.3.1.5 Ox	idase test	38
		4.3.1.6 Pro	otease test	39
		4.3.2 Anti quoru	m-sensing effects of potential probiotics	40
		4.3.3 Biofilm inh	nibition assay	41
		4.3.4 Evaluation microalgae	of potential probionts on the growth of e	42
			valuation of <i>Lysinibacillus fusiformis</i> strain -1 on the growth of <i>Amphora</i> sp.	42
			valuation of <i>Bacillus</i> sp. strain A-2 on the rowth of <i>Amphora</i> sp.	43
			valuation of <i>Lysinibacillus fusiformis</i> strain I-3 on the growth of <i>Chlorella</i> sp.	44
			valuation of <i>Bacillus pocheonensis</i> strain S-2 on e growth of <i>Spirulina</i> sp.	45
	4.4	Discussion		46
	4.5	Conclusion		49
5	PRC	<b>BIONTS ISOL</b>	ON IN Artemia fransiscana NAUPLII WITH ATED FROM MIROALGAE AND ITS INST PATHOGENIC VIBRIOS	50
	5.1	Introduction		50
	5.2	Materials and me	ethods	51
		5.2.1 Probiont	and pathogen culture	51
		5.2.2 Culture	of axenic Artemia	51
			tion on axenity	52
			ation of potential probionts	52
		Nauplii I	ary <i>in vivo</i> assay using <i>Artemia fransiscana</i> bioencapsulated with probionts and fed with gae against <i>Vibrio</i> spp.	52
			culture of <i>Artemia</i>	52
		5.2.5.2 E	xperimental design	52
			ibrios count in Artemia	54
		5.2.6 Statistical	analysis	54
	5.3	Results		55
		5.3.1 Colonizat 12 h inte	tion of probionts in gnotobiotic <i>Artemia</i> at every erval	55
		5.3.2 Artemia	in vivo challenge test	58
		v	Survival of <i>Artemia franciscana</i> bioencapsulated with potential probionts and fed with nicroalgae without challenged with pathogen	58

		5.3.2.2 Survival of <i>Artemia fransiscana</i> bioencapsulate with potential probionts and fed with microalga after challenged with <i>Vibrio harveyi</i>	
		5.3.2.3 Vibrios count in <i>Artemia</i> after challenged with <i>Vibrio harveyi</i>	60
		5.3.2.4 Survival of <i>Artemia fransiscana</i> bioencapsulate with potential probionts and fed with microalga after challenged with <i>Vibrio parahaemolyticus</i>	
		5.3.2.5 Vibrios count in <i>Artemia</i> after challenged with <i>Vibrio parahaemolyticus</i>	62
	5.4	Discussion	63
	5.5	Conclusion	66
6	SUN	AMARY, CONCLUSION AND RECOMMENDATION	67
v	6.1	Summary	67
	6.2	Conclusion	69
	6.3	Recommendation for future research	70
REI	FERE	NCES	71
APF	PENDI	ICES	92
		A OF STUDENT	101
LIS	TOF	PUBLICATIONS	102

 $(\mathbf{C})$ 

# LIST OF TABLES

Table		Page
2.1	Literature on bacteria-algae interaction	11
3.1	The number of different colonies isolated from each group	18
	of microalgae	
3.2	Diameter of potential probionts isolated from microalgae	19
	against Vibrio harveyi and Vibrio parahaemolyticus	
3.3	The morphology of potential probionts observed with	22
	naked eyes	
3.4	Results of BLAST analysis	25
4.1	The reading of excitation and emission using Varioskan	32
	Lux microplate reader	
4.2	The measurement of degrading zone by probionts in anti-	40
	quorum sensing assay	
4.3	Percentage inhibition of pathogens after co-cultivated with	41
	the potential probiont cultures	
5.1	Treatments in in vivo Artemia challenge test	53
	(a) Unchallenged group	53
	(b) Group challenged with Vibrio harveyi	53
	(c) Group challenged with Vibrio parahaemolyticus	54
5.2	Mean bacterial count in gnotobiotic Artemia fransiscana	57
	nauplii enriched with $10^6$ CFU ml <sup>-1</sup> of probionts,	
	recorded at every 12 h interval	
5.3	Vibrios count in Artemia fransiscana nauplii after	60
	challenged with Vibrio harveyi	
5.4	Vibrios count in Artemia fransiscana nauplii after	62
	challenged with Vibrio pararahaemolyticus	

# LIST OF FIGURES

Figure		Page
3.1	The inhibition zone produced by potential probionts against Vibrio harveyi	20
3.2	The inhibition zone produced by potential probionts against Vibrio parahaemolyticus	21
3.3	Gram-staining of potential probiont A-1 under light microscope with 100x magnification	22
3.4	Gram-staining of potential probiont A-2 under light microscope with 100x magnification	23
3.5	Gram-staining of potential probiont Cl-3 under light microscope with 100x magnification	23
3.6	Gram-staining of potential probiont S-2 under light microscope with 100x magnification	24
3.7	Agarose gel electrophoresis analysis (0.8% agarose gel) of PCR amplification of the 16S rRNA of potential probionts strain A-1, A-2, Cl-3 and S-2 with expected band of 1000 bp	24
4.1	The quantitative assay of amylase enzyme production by the potential probionts	35
4.2	The quantitative assay of gelatinase enzyme production by the potential probionts carried out in Bijou bottles	36
4.3	The quantitative assay of gelatinase enzyme production by the potential probionts carried out in test tubes	37
4.4	The quantitative assay of lipase enzyme production by the potential probionts	37
4.5	The quantitative assay of oxidase enzyme production by the potential probionts	38
4.6	The quantitative assay of protease enzyme production by the potential probionts	39

4.7	The quantitative assay of anti-quorum sensing enzyme	40
	production by the potential probionts against	
	Chromobacterium violacein	
4.8	The growth of Lysinibacillus fusiformis strain A-1 co-	42
	cultured with Amphora sp. (red line) compared with	
	single Amphora sp. (blue line) as the control	
4.9	The growth of Bacillus sp. strain A-2 co-cultured with	43
	Amphora sp. (red line) compared with single Amphora	
	sp. (blue line) as the control	
4.10	The growth of Lysinibacillus fusiformis strain Cl-3 co-	44
	cultured with Chlorella sp. (red line) compared with	
	single <i>Chlorella</i> sp. (blue line) as the control	
4.11	The growth of Bacillus pocheonensis strain S-2 co-	45
	cultured with Spirulina sp. (red line) compared with	
	single Spirulina sp. (blue line) as the control	
5.1	Survival of Artemia fransiscana after 24h of	58
	bioencapsulation with potential probionts and fed with	
	microalgae without challenged with pathogen	
5.2	Survival of Artemia fransiscana after 24 h of	59
	bioencapsulation with potential probionts and fed with	
	microalgae after challenged with Vibrio harveyi	
5.3	Survival of Artemia fransiscana after 24 h of	61
	bioencapsulation with potential probionts and fed with	
	microalgae after challenged with Vibrio	
	parahaemolyticus	

# LIST OF ABBREVIATIONS

% CFU ml <sup>-1</sup>	Percentage Colony forming units per millilitre
Cells ml <sup>-1</sup>	Cells per millilitre
mmt	Million metric ton
mm	Millimetre
ml	Millilitre
min	Minute
NaOH	Sodium hydroxide
NaOCl	Sodium hypochlorite
Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub>	Sodium thiosulphate
$g l^{-1}$	Gram per litre
L	Litre
h	Hour
ppt	Part per thousand
TSA	Thiosulphate Soy Agar
TCBS	Thiosulphate Citrate Bile Salt
rfu	Relative forming unit
μΙ	Microlitre
um	Micrometre
°C	Degree Celcius

 $\mathbf{G}$ 

### **CHAPTER 1**

#### INTRODUCTION

### 1.1 Background of study

The extensive of industrial development has caused severe disruption to the worldwide aquatic environment. Various problems have occurred, for instance, the widespread of aquatic diseases and the deterioration of water quality have become a major threat in aquaculture.

The accumulation of drug residues in aquatic products has raising a great concern in term of food safety due to the use of antibiotics in aquaculture practices. Furthermore, several pathogens have also resist to drug. Antibiotics must be replaced or reduced with new and environmental friendly products since the food quality demands keep increasing and the negative impacts of antibiotics have caused a risk to humans (Quesada et al., 2013; Arias-Andres et al., 2014; Siriyappagouder et al., 2014).

Live microalgae have been considered as an essential element in aquaculture diet. For instance, microalgae serve as diet component for marine bivalve, mollusc, larvae of some gastropods, fish and shrimp species which, later on will be used as live feeds to cultivate carnivorous larvae of many shrimp and fish species currently farmed (Muller-Feuga, 2000; Naumann et al., 2013). Several tests on non-living diets of algal or non-algal origin have been studied over the years in an attempt providing cost-effective approaches to live microalgae feeds (Hemaiswarya et al., 2011). Unfortunately, Aji (2011) claimed that non-living diets was not able to completely replace live microalgae as feed in aquaculture. Thus, live microalgae remain as primary choice in marine aquaculture feeding (Hemaiswarya et al., 2011) and the management of microalgae production is fundamental in most aquaculture operations, especially in hatcheries (Naumann et al., 2013).

As aquaculture is facing problem due to massive loss because of spreading of diseases there are numerous approaches available to protect aquatic animals against pathogens. Of these approaches, probiotics have become widely used for disease control. Probiotics are defined as live microorganisms where it can confer a health benefit to the host when consume in adequate amount (Reid et al., 2003). Although probiotics offer a promising alternative in aquaculture, the ways that probiotics are applied need to take into consideration to prevent any negative impacts. Probiotics applications can be either in mono or multiple strains. Probiotics also can be a combination of probiotic strains with immunostimulants or prebiotic (Hai, 2015).

Most of the current outbreaks are due to intracellular pathogens, and the production of vaccines against these pathogens has not been an easy task. Vaccines have been reported only can control a specific disease. Due to this problem, vaccinations alone is not possible

in combating all the diseases that occurred in aquatic environments. Thus, the production of non-toxic and non-polluting biological agents for antibiotics substitutions and vaccines has become the main purpose in aquaculture research and development (Wang et al., 2017).

Therefore, the use of probiotics in replacement to antibiotics could be a great alternative in order to improve the safety and quality of aquaculture products. Probiotics are harmless bacteria that improve the well being of an organism or host and contribute either directly or indirectly in protecting the host against harmful bacterial population (Pandiyan et al. 2013). Numerous studies demonstrated that some bacteria have been used as probiotics in aquatic culture to combat bacterial diseases or to improve growth and survival. Moreover, the alteration of microbial community in culture environment also contribute to the reduction of potentially pathogenic microbial population, and enhancement of microalgae as well as live feed cultures (Planas et al., 2004; Villamil et al., 2003; Zink et al., 2013). Planas et al. (2015) pointed out that the selection and application of bacteria as probiotic in microalgae cultures are required to eliminate potentially inhibiting strains or strains that might cause detrimental effects to the cultures. It is well known that bacteria was able to increase or supress the growth of certain microalgae species.

The symbiotic consortium plays an important role in aquatic ecosytems. A greater insight of these associations between microalgae and bacteria in natural ecosystems is very important aspect that has not been thoroughly investigated (Tandon and Jin, 2017). The effects of single bacterial strain on microalgal growth were well documented in several studies. For instance, *Rhizobium* sp. enhanced the growth of *Botrycoccus braunii* and *Candidatus phycosocius bacilliformis* (Rivas, 2010; Tanabe et al., 2015), *Pelagibaca bermudensis* enhanced the growth of *Tetraselmis striata* (Park et al., 2017), and the growth of *Chlamydomonas reinhardtii*, *Chlorella vulgaris*, *Scenedesmus* sp. and *B. braunii* was enhanced by *Rhizobium* sp. (Kim et al., 2014). In previous study, de-Bashan (2016) also reported that associations of *Azospirillium brasilense* and *Bacillus pumilus* to the *Chlorella* sp. culture significantly increased its growth rate.

The occurrence of pathogenic and opportunistic bacteria in aquatic culture can be controlled by incorporation of probiotics via *Artemia*. Preventing colonization of pathogenic bacteria with selected probiotic strains has been proposed as a valuable approach for microbial control during *Artemia* culture. Another studies demonstrated that probiotics bacteria (*Aeromonas* sp. strain LVS3 and *V. alginolyticus* strain LVS8) can hinder the growth of *Vibrio proteolyticus* (Verschuere et al., 2000) while lactic acid bacteria can reduce the number of *Vibrio alginolyticus* in *Artemia* culture (Villamil et al., 2002). Application of probiotics in rotifers culture exhibit inhibitory effects towards *Vibrio anguillarum* (Harzevili et al., 1988).

### 1.2 Problem statement

An utmost bottleneck in aquaculture is the sudden reduction of fish larvae caused by pathogenic bacteria (Reid et al., 2009). Among the groups of diseases in aquaculture, vibriosis is the well-known cause of massive economic losses and responsible for high mortality in cultured shrimp, fish and shellfish. Global losses resulting from shrimp diseases are estimated around US\$3 billion where *Vibrio harveyi* has been identified as the main causative agent, in which it caused 100 % losses in larval production in many countries (Lavilla-Pitogo et al., 1990; Vinod et al. 2006). *Vibrio campbelli, V. parahaemolyticus, V. harveyi* and *V. tubiashii* were found to predominate in the west coast of Peninsular Malaysia (You et al., 2016). According to Ransangan et al. (2013), the three most dominant species found in shrimp hatchery (*P. monodon*) in Sabah, Malaysia were *V. harveyi* (22.2%), *V. parahaemolyticus* (22.2%) and *V. alginolyticus* (19.4%).

In addition, the diseases and sudden change of environmental conditions often lead to serious economic losses. The use of antibiotics to control these crashes however has led to spreading and developing of antibiotic-resistance bacteria. Several alternatives have been implemented to improve the sustainability and quality of aquaculture products. Of those methods, the most promising alternatives is the use of probiotics in aquatic animals (Rekiel et al., 2007).

Low concentration of microalgae biomass is one of the serious problems face by most hatcheries production. Thus, the enhancement of microalgae cultures is one of the crucial things that need to be considered for high production of microalgae.

### 1.3 Significant of the study

Aquaculture as the science of cultivating aquatic animals, plants and organisms like fish, shellfish, microalgae and seaweed, for human consumption and use. In future, microalgae are considered to be one of the most promising feed stocks for aquaculture. One of the effective way to enhance microalgae biomass is through mutualistic interaction between microalgae-bacteria symbiosis. In this study, probionts were isolated from microalgae and co-cultured again with the microalgae. Beneficial bacteria or probionts with antibacterial properties that able to enhance the growth of microalgae would be valuable in aquaculture industry. Besides increasing the algal biomass, the probionts are able to supress fish larval pathogens in aquaculture system thus decreasing the prevalence of diseases caused by *Vibrio* spp..

### 1.4 Objectives of the study

Hence, the objectives of this study were :

- 1. To isolate, screen and identify bacteria with antibacterial activity as probiotics from different species of microalgae against *Vibrio harveyi* and *Vibrio parahaemolyticus*
- 2. To determine the properties of potential probionts in producing extracellular enzymes, anti-quorum sensing, biofilm inhibition and its effect on the growth of respective microalgae.
- 3. To determine the capability of potential probionts to colonize in *Artemia* and its protection effect against *Vibrio* spp. in preliminary *in vivo* assay.

### 1.5 Hypothesis of the study

The hypothesis of the study :

**Null hypothesis :** The isolated strains are unable to inhibit the growth of *V. harveyi* and *V. parahaemolyticus* in *in vitro* assay, do not show any probiotic properties, do not able to increase the growth of microalgae and do not provide protection to *Artemia fransiscana* from vibrios infection.

Alternative hypothesis : The isolated strains are able to inhibit the growth of *V. harveyi* and *V. parahaemolyticus* in *in vitro* assay, portray probiotic properties, can increase the growth of microalgae and able to protect *Artemia fransiscana* from vibrios infection.

#### REFERENCES

- Abudoleh, S. M. and Mahasneh, A. M. (2016). Quorum Sensing Inhibitors: The novel bacterial infection disrupting agents. Journal of *British Microbiology Research*. 15(6): 1-18.
- Aguirre-Guzman, G., Ruiz, H.M. and Asxencio, F. (2004). A review of extracellular virulence product of *Vibrio* species important in disease of cultivated shrimp. *Aquaculture Research*. 35: 1395-1404.
- Al-Ajlani, M. M. and Hasnain, S. (2010). Bacteria exhibiting antimicrobial activities; screening for antibiotics and the associated genetic studies. In *Open Conference Proceedings Journal*. 1: 230-238.
- Ali, A. (2000). Application of immunostimulants in aquaculture: Current knowledge and future perspectives. *Aquaculture Research*. 48: 1-23.
- Ali, W. and Sultana, R. (2011). Response of *Artemia franciscana* fed with *Aeromonas hydrophilla* and challenged with *Vibrio campbellii*. *Pakistan Journal of Biochemistry and Molecular Biology*. 44(2): 77-80.
- Aji, L. P. (2011). The use of algae concentrates, dried algae and algal substitutes to feed bivalves. *Makara Journal of Science*. 15(1): 1-8.
- Amavizca, E., Bashan, Y., Ryu, C. M., Farag, M. A., Bebout, B. M. and de-Bashan, L.
  E. (2016). Enhanced performance of the microalga *Chlorella sorokiniana* remotely induced by the plant growth-promoting bacteria *Azospirillum* brasilense and Bacillus pumilus. Nature. 7: 1-11.
- Amin, M., Rakhisi, Z. and Ahmady, A. Z. (2015). Isolation and identification of *Bacillus* species from soil and evaluation of their antibacterial properties. *Avicenna Journal of Clinical Microbiology and Infection*. 2(1): 1-4.
- Anh, N. T. N., Van Hoa, N., Van Stappen, G. and Sorgeloos, P. (2009). Effect of different supplemental feeds on proximate composition and *Artemia* biomass production in salt ponds. *Aquaculture*. 286(3-4): 217-225.
- Arias, A., and Carrera-Parra, L. F. (2014). First record of the genus *Kuwaita* (Annelida: *Lumbrineridae*) in Europe with the description of a new species and new ultramorphological data for the genus. *Zootaxa*. 3887(1): 68-78.
- Avella, M. A., Gioacchini, G., Decamp, O., Makridis, P., Bracciatelli, C. and Carnevali, O. (2010). Application of multi-species of *Bacillus* in sea bream larviculture. *Aquaculture*. 305(1-4): 12-19.
- Austin, B. and Austin, D.A. (2007). Bacterial fish pathogens: disease of farmed and wild fish. Springer Praxis Publishing, Chichester, United Kingdom. 4:522.Azam, F. and Malfatti, F. (2007). Microbial structuring of marine ecosystems. *Nature Reviews Microbiology*. 5: 789-791.

- Azirah,, M. Z., Karim, M., and Harmin, S. A. (2016). Enrichment of brine shrimp (Artemia franciscana) nauplii with potential probiont strains, Bacillus JAQ04 and Micrococcus JAQ07. International proceeding: The 4<sup>th</sup> International Seminar on Fisheries and Marine Science. 21-27.
- Balcázar, J. L., De Blas, I., Ruiz-Zarzuela, I., Cunningham, D., Vendrell, D., and Múzquiz, J. L. (2006). The role of probiotics in aquaculture. *Veterinary Microbiology*. 114 (3-4): 173-186.
- Barman, P., Banerjee, A., Bandyopadhyay, P., Chandra, K., Kumar, P. and Mohapatra, D.(2011). Isolation, identification and molecular characterization of potential probiotic bacterium, *Bacillus subtilis* PPP 13 from *Penaeus monodon. Journal* of Biotechnology, Bioinformatics and Bioengineering. 1(4): 473-482.
- Basurto-Cadena, M. G. L., Vázquez-Arista, M., García-Jiménez, J., Salcedo-Hernández, R., Bideshi, D. K. and Barboza-Corona, J. E. (2012). Isolation of a new Mexican strain of *Bacillus subtilis* with antifungal and antibacterial activities. *The Scientific World Journal*. 2012: 1-7.
- Ben-Haim, Y., Thompson, F. L., Thompson, C. C., Cnockaert, M. C., Hoste, B., Swings, J.,and Rosenberg, E. (2003). Vibrio corallilyticus sp. nov., a temperaturedependent pathogen of the coral Pocillopora damicornis. International Journal of Systematic and Evolutionary Microbiology. 53(1): 309-315.
- Brennan, L. and Owende, P. (2010). Biofuels from microalgae a review of technologies for production, processing and extractions of biofuels and co-products. *Renew Sustain Energy Review*. 14: 557-577.
- Bhatia, D., Mittal, A. and Malik, D. K. (2016). Antimicrobial activity of PVP coated silver nanoparticles synthesized by *Lysinibacillus varians*. *3 Biotech*. 6(2): 196.
- Blythe, J., Sulu, R., Harohau, D., Weeks, R., Schwarz, A. M., Mills, D. and Phillips, M. (2017). Social dynamics shaping the diffusion of sustainable aquaculture innovations in the Solomon Islands. *Sustainability*. 9(1): 126.
- Browdy, C. (1998). Recent developments in penaeid broodstock and seed production technologies: improving the outlook for superior captive stocks. *Aquaculture*.164: 3-21.
- Brown, M. R. (2002). Nutritional value and use of microalgae in aquaculture. *CSIRO Marine Research*. 282-289.
- Brown, M. R., Jeffrey, S. W., Volkman, J. K., and Dunstan, G. A. (1997). Nutritional properties of microalgae for mariculture. *Aquaculture*. 151: 315-331.
- Brown, M. (2011). Modes of action of probiotics: recent developments. *Journal of Animal and Veterinary Advances*. 10(14): 1895-1900.
- Bruckner, C.G., Rehm, C., Grossart, H-P., Kroth, P.G. (2011). Growth and release of extracellular organic compounds by benthic diatoms depend on interactions with bacteria. *Environmental Microbiology*. 13: 1052-1063.

- Bruckner, C. G., Bahulikar, R., Rahalkar, M., Schink, B., and Kroth, P. G. (2008). Bacteria associated with benthic diatoms from Lake Constance: phylogeny and influences on diatom growth and secretion of extracellular polymeric substances. *Applied and Environmental Microbiology*. 74(24): 7740-7749.
- Bull, A. T., Ward, A. C., and Goodfellow, M. (2000). Search and discovery strategies for biotechnology: the paradigm shift. *Microbiology and Molecular Biology Reviews*. 64(3): 573-606.
- Burkholder, J. M., Gilbert, P. M., Skelton, H. M. (2008). Mixotrophy, a major mode of nutrition for harmful algal species in eutrophic waters. *Harmful Alga*. 8: 77-93.
- Chang, C. and Liu W. (2002). An evaluation of two probiotic bacterial strains, *Enterococcus faccium* SF68 and *Bacillus toyoi*, for reducing *Edwardsiellosis* in cultured European eel, *Anguilla anguilla L. Journal of Fish Diseases*. 25: 311-315.
- Chen, J. Y., Chen, J. C., Chang, C. Y., Shen, S. C., Chen, M. S. and Wu, J. L. (2000). Expression of recombinant tilapia insulin-like growth factor-I and stimulation of juvenile tilapia growth by injection of recombinant IGFs polypeptides. *Aquaculture*. 181(3-4): 347-360.
- Chi, C., Bing, J., Xiao, B. Y., Tian, Q. L., Lei, X. and Gao, X. W. (2014). Effects of three strains of intestinal autochtholonous bacteria and their extracellular products on the immune response and disease resistance of common carp, *Cyprinus carpio. Fish and Shellfish Immunology.* 36: 9-18.
- Cooper, M. B., and Smith, A. G. (2015). Exploring mutualistic interactions between microalgae and bacteria in the omics age. *Current Opinion in Plant Biology*. 26: 147-153.
- Compaoré, C. S., Jensen, L. B., Diawara, B., Ouédraogo, G.A., Jakobsen, M., and Ouoba, LII. (2013). Resistance of antimicrobials and acid and bile tolerance of *Bacillus* spp. isolated from *Bikalga*, fermented seeds of *Hibiscus sabdariffa*. African Journal of Food Science. 7(11): 408–14.
- Collado, M. C., Isolauri, E., Salminen, S. and Sanz, Y. (2009). The impact of probiotic on gut health. *Current Drug Metabolism*. 10(1): 68-78.
- Cotner, J. B., and Biddanda, B.A. (2002). Small player, large role: microbial influence on biogeochemical processes in pelagic aquatic ecosystems. *Ecosystems*. 5: 105-121.
- Claus, D. (1992). A standardized Gram staining procedure. World journal of Microbiology and Biotechnology. 8(4): 451-452.
- Croft, M. T., Lawrence, A. D., Raux-Deery, E., Warren, M. J., Smith, A. G. (2005). Algae acquire vitamin  $B^{12}$  through a symbiosis relationship with bacteria. *Nature*. 438: 90-93.

- Chevanton, M., Garnier, M., Bougaran, G., Schreiber, N., Lukomska, E., Bérard, J. B. and Cadoret, J. P. (2013). Screening and selection of growth-promoting bacteria for *Dunaliella* cultures. *Algal Research*. 2(3): 212-222.
- Christova, N., Tuleva, B., and Nikolova-Damyanova, B. (2004). Enhanced hydrocarbon biodegradation by a newly isolated *Bacillus subtilis* strain. *Journal of Bioscience*. 59(3-4): 205-208.
- Daniels, C. L., Merrifield, D. L., Ringø, E., and Davies, S. J. (2013). Probiotic, prebiotic and synbiotic applications for the improvement of larval European lobster (*Homarus gammarus*) culture. Aquaculture. 416: 396-406.
- Das, S., Mondal, K., and Haque, S. (2017). A review on application of probiotic, prebiotic and synbiotic for sustainable development of aquaculture. *Growth.* 14: 15.
- Dalmin, G., Kathiresan, K., and Purushothaman, A. (2001). Effect of probiotics on bacterial population and health status of shrimp in culture pond ecosystem. *Indian Journal of Experimental Biology*. 39(9): 939-942.
- De-Bashan, L. E. and Bashan, Y. (2008). Joint immobilization of plant-growth promoting bacteria and green microalgae in alginate beads as an experimental model for studying plant-bacterium interactions. *Applied and Environmental Microbiology*, 74: 6797-6802.
- De-Bashan, L.E., Bashan, Y., Moreno, M., Lebsky, V. K. and Bustillos, J. J. (2002). Increased pigmentation and lipid content, lipid variety, and cell and population size of the microalgae *Chlorella spp*. When co-immobilized in alginate beads with the microalgae-growth-promoting bacterium *Azospirillium brasilense*. *Canadian Journal of Microbiology*. 102: 88-93.
- De-Bashan, L. E., Mayali, X., Bebout, B. M., Weber, P. K., Detweiler, A. M., Hernandez, J. P. and Bashan, Y. (2016). Establishment of stable synthetic mutualism without co-evolution between microalgae and bacteria demonstrated by mutual transfer of metabolites (NanoSIMS isotopic imaging) and persistent physical association (Fluorescent in situ hybridization). *Algal Research*. 15: 179-186.
- Decamp, O., Moriarty, D. J., and Lavens, P. (2008). Probiotics for shrimp larviculture: review of field data from Asia and Latin America. *Aquaculture Research*. 39(4): 334-338.
- Demirbas, M. F. (2011). Biofuels from algae for sustainable development. *Applied Energy*. 88(10): 3473-3480.
- Dhont, J., Dierckens, K., Støttrup, J., Van Stappen, G., Wille, M., and Sorgeloos, P. (2013). Rotifers, *Artemia* and copepods as live feeds for fish larvae in aquaculture. *Advances in Aquaculture Hatchery Technology*. 157-202.

- Dimitroglou, A., Merrifield, D. L., Carnevali, O., Picchietti, S., Avella, M., Daniels, C., and Davies, S. J. (2011). Microbial manipulations to improve fish health and production – a Mediterranean perspective. *Fish and Shellfish Immunology*. 30(1): 1-16.
- Dixon, B., Van Poucke, S., Chair, M., Demasque, M., Nelis, H., Sorgeloos, P. and De Leenheer, A. (1995). Bioencapsulation of the antibacterial drug sarafloxacin in nauplii of the brine shrimp Artemia fransiscana. Journal of Aquatic Animal Health. 7: 42-45.
- Drancourt, M., Bollet, C., Carlioz, A., Martelin, R., Gayral, J. P. and Raoult D. (2000). 16S Ribosomal DNA sequence analysis of a large collection of environmental and clinical unidentifiable bacterial isolates. *Journal of Clinical Microbiology*. 38: 3623-3630.
- Duc, L. H., Hong, H. A., Barbosa, T. M., Henriques, A. O. and Cutting, S. M. (2004). Characterization of *Bacillus* probiotics available for human use. *Applied and Environmental Microbiology*. 70(4): 2161-2171.
- El-Rahman, A. M. A., Khattab, Y. A. E. and Shalaby, A. M. E. (2009). *Micrococcus luteus* and *Pseudomonas* species as probiotics for promoting the growth of performance and health of Nile tilapia, *Oreochromis niloticus*. *Fish and Shellfish Immunology*. 27: 175-180.
- Espeland, E. M. and Wetzel, R. G. (2001). Effects of photosynthesis on bacterial phosphatase production in biofilms. *Microbial Ecology*. 42: 328-337.
- Ethier, S., Woisard, K., Vaughan, D. and Wen, Z. (2011). Continuous culture of the microalgae *Schizochytrium limacinum* or biodiesel-derived crude glycerol for producing docosahexaenoic acid. *Journal of Bioresource Technology*. 102: 88-93.
- Farahi, A., Kasiri, M., Sudagar, M. and Alamshahi, F. (2011). The effects on growth, survival, and tolerance against environmental stressor (high temperature) of different concentrations Probiotic *Bacillus* sp., fed to angelfish (*Pterophyllum* scalare Schultze, 1823) larvae. Journal of Animal and Veterinary Advances. 10(17): 2305-2311.
- Farzanfar, A. (2006). The use of probiotics in shrimp aquaculture. Journal of Immunology and Medical Microbiology. 48: 149-158.
- F.A.O. (2014). The state of world fisheries and aquaculture 2012, Rome:F.A.O..
- Fontana, L., Bermudez-Brito, M., Plaza-Diaz, J., Munoz-Quezada, S. and Gil, A. (2013). Sources, isolation, characterisation and evaluation of probiotics. *British Journal of Nutrition*. 109(2): 35-50.
- Fouilland, E. (2012). Biodiversity as a tool for waste phycoremediation and biomass production. *Reviews in Environmental Science and Biotechnology*. 11(1): 1-4.

- Fulbright, S. P., Chisholm, S., Reardon, K. F. (2016). Growth inhibition of Nannochloropsis species by Bacillus pumilus. Algal Research. 20: 70-76.
- Führ, F., Tesser, M. B., Rodrigues, R. V., Pedron, J., and Romano, L. A. (2016). Artemia enriched with hydrolyzed yeast improves growth and stress resistance of marine pejerrey Odontesthes argentinensis larvae. Aquaculture. 450: 173-181.
- Gärdes, A., Iversen, M. H., Grossart, H. P., Passow, U. and Ullrich, M. S. (2011). Diatom-associated bacteria are required for aggregation of *Thalassiosira weissflogii. Journal of Microbial Ecology.* 5: 436-445.
- Garge, S. S., and Nerurkar, A. S. (2016). Attenuation of quorum sensing regulated virulence of *Pectobacterium carotovorum* subsp. *carotovorum* through an AHLlactonase produced by *Lysinibacillus* sp. Gs50. *PloS One*. 11(12): e0167344.
- Gavin, P. (2016). Testing the efficacy of probiotics for disease control in aquaculture. *Microbiology Australia*. 122-123.
- Gatesoupe, F. (1994). Lactic acid bacteria increase the resistance of turbot larvae (*Scophthalmus maximus*) against pathogenic *Vibrio. Aquatic Living Resources*. 7: 277-282.
- Gatesoupe, F. J. (1999). The use of probiotics in aquaculture. *Aquaculture*. 180: 147-165.
- Gonzalez, L. E. and Bashan, Y. (2000). Increased growth of the microalga *Chlorella vulgaris* when coimmobilized and cocultured in alginate beads with the plantgrowth-promoting bacterium *Azospirillum brasilense*. *Applied and Environmental Microbiology*. 66(4): 1527-1531.
- González-Escalona, N., Cachicas, V., Acevedo, C., Rioseco, M. L., Vergara, J. A., Cabello, F. and Espejo, R. T. (2005). *Vibrio parahaemolyticus* diarrhea, Chile, 1998 and 2004. *Emerging Infectious Diseases*. 11(1): 129.
- Ghosh, S., Sinha, A. and Sahu, C. (2008). Dietary probiotic supplementation in growth and health of live- bearing ornamental fishes. *Aquaculture Nutrition*. 14(4): 289-299.
- Gilbert, J. J. (1988). Suppression of rotifer populations by *Daphnia*: A review of the evidence, the mechanisms, and the effects on zooplankton community structure. *Limnology and Oceanography.* 33(6): 1286-1303.
- Giri, S. S., Sen, S. S., Jun, J. W., Sukumaran, V. and Park, S. C. (2016). Role of *Bacillus subtilis* VSG-4 derived biosurfactant in mediating immune response in *Labeo rohita*. Fish and Shellfish Immunology. 54: 220-229.
- Grossart, H. P., Levold, F., Allgaier, M., Simon, M., Brinkhoff, T. (2005). Marine diatom species harbour distinct bacterial communities. *Environmental Microbiology*. 7: 860-873.

- Guo, Z. and Tong, Y. W. (2014). The interactions between *Chlorella vulgaris* and algal symbiotic bacteria under photoautotrophic and photoheterotrophic conditions. *Journal of Applied Phycology*. 26(3): 1483-1492.
- Gurung, T. B., Urabe, J. and Nakanishi, M. (1999). Regulation of the relationship between phytoplankton Scenedesmus acutus and heterotrophic bacteria by the balance of light and nutrients. *Aquatic Microbial Ecology*. 17(1): 27-35.
- Hai, N., Fotedar, R. and Buller, N. (2007). Selection of probiotics by various inhibition test methods for use in the culture of western king prawns, *Penaeus latisulcatus* (*Kishinouye*). Aquaculture. 272(1-4): 231-239.
- Hai, N., Buller, N. and Fotedar, R. (2010). Encapsulation capacity of Artemia nauplii with customized probiotics for use in the cultivation of western king prawns (*Penaeus latisulcatus Kishinouye*, 1896). Aquaculture Research. 41(6): 893-903.
- Hai, N. V. (2015). The use of probiotics in aquaculture. *Journal of Applied Microbiology*. 119(4): 917-935.
- Han, Q., Keesing, J. K. and Liu, D. (2016). A review of sea cucumber aquaculture, ranching, and stock enhancement in China. *Reviews in Fisheries Science and Aquaculture*. 24(4): 326-341.
- Hannah, C., Mani, M. and Ramasamy, R. (2013). Evaluation of the Biochemical Composition of Four Marine Algae and Its Nutritional Value for Brine Shrimp. *IOSR Journal of Pharmacy and Biological Sciences*. 6(3): 47-51.
- Hatha, A. A. M., Mujeeb Rahiman, K. M., Jasmine, B. and Suresh Kumar, S. (2014). Growth enhancement of micro algae, *Chaetoceros calcitrans* and *Nannochloropsis oculata*, using selected bacterial strains. *International Journal* of Current Microbiolgy and Applied Sciences. 3(4): 352-359.
- Smith Jr, H. L. and Goodner, K. (1958). Detection of bacterial gelatinases by gelatinagar plate methods. *Journal of Bacteriology*. 76(6): 662.
- Harzevili, A. R. S., Van Duffel, H., Dhert, P., Swings, J. and Sogerloos, P., (1998). Use of a potential probiotic *Lactococcus lactis* AR21 strain for the enhancement of growth in the rotifer *Brachionus plicatilis*. *Aquaculture Resource*. 29: 411-417.
- Heatley, N. G. (1944). A method for the assay of penicillin. *Biochemical Journal*. 38(1): 61.
- Hemaiswarya, S., Raja, R., Kumar, R. R., Ganesan, V. and Anbazhagan, C. (2011). Microalgae: a sustainable feed source for aquaculture. World Journal of Microbiology and Biotechnology. 27(8): 1737-1746.
- Hemraj, V., Diksha, S. and Avneet, G. (2013). A review on commonly used biochemical test for bacteria. *Innovare Journal of Life Sciences*. 1(1): 1-7.

- Hidalgo, M. C., Skalli, A., Abellán, E., Arizcun, M. and Cardenete, G. (2006). Dietary intake of probiotics and maslinic acid in juvenile dentex (*Dentex dentex* L.): effects on growth performance, survival and liver proteolytic activities. *Aquaculture nutrition*. 12(4): 256-266.
- Hong, H. A., Duc, L. H. and Cutting, S. M. (2005). The use of bacterial spore formers as probiotics. *Microbiology Reviews*. 29(4): 813-835.
- Ibrahem, M. D. (2015). Evolution of probiotics in aquatic world: potential effects, the current status in Egypt and recent prospectives. *Journal of Advanced Research*. 6(6): 765-791.
- Iguchi, K. I., Ogawa, K., Nagae, M. and Ito, F. (2003). The influence of rearing density on stress response and disease susceptibility of ayu (*Plecoglossus altivelis*). *Aquaculture*. 220(1-4): 515-523.
- Irianto, A. and Austin, B. (2002). Probiotics in aquaculture. *Journal of Fish Diseases*. 25: 633-642.
- Janarthanam, K., George, M. R., John, K. R. and Jeyaseelan, M. J. (2012). In vitro and in vivo biocontrol of Vibrio harveyi using indigenous bacterium, Bacillus spp. Indian journal of Geo-Marine Sciences. 41(1): 83-89.
- Jasmin, M. Y., Wagaman, H., Yin, T. A., Ina-Salwany, M. Y., Daud, H. M., and Karim, M. (2016). Screening and evaluation of local bacteria isolated from shellfish as potential probiotics against pathogenic Vibrios. *Journal of Environmental Biology*. 37(4): 801-809.
- Joint, I., Henriksen, P., Fonnes, G. A., Bourne, D., Thingstad, T. F. and Riemann, B. (2002). Competition for inorganic nutrients between phytoplankton and bacterioplankton in nutrient manipulated mesocosms. *Aquatic Microbial Ecology*. 29(2): 145-159.
- Jung, S. W., Kim, B. H., Katano, T., Kong, D. S. and Han, M. S. (2008). Pseudomonas fluorescens HYK0210- SK09 offers species- specific biological control of winter algal blooms caused by freshwater diatom Stephanodiscus hantzschii. Journal of Applied Microbiology. 105(1): 186-195.
- Jayathilakan, K., Sultana, K., Radhakrishna, K. and Bawa, A. S. (2012). Utilization of byproducts and waste materials from meat, poultry and fish processing industries: A review. *Journal of Food Science and Technology*. 49(3): 278-293.
- Kazamia, E., Czesnick, H., Nguyen, T. T. V., Croft, M. T., Sherwood, E., Sasso, S. and Smith, A. G. (2012). Mutualistic interactions between vitamin B12- dependent algae and heterotrophic bacteria exhibit regulation. *Environmental Microbiology*. 14(6): 1466-1476.
- Kelleher, K., Willmann, R. and Arnason, R. (2009). The sunken billions: the economic justification for fisheries reform. The World Bank.

- Kesarcodi-Watson, A., Kaspar, H., Lategan, M.J. and Gibson, L. (2008). Probiotics in aquaculture: the need, principles and mechanisms of action screening processes. *Aquaculture*. 274: 1-14.
- Kesarcodi-Watson, A., Kaspar, H., Lategan, M. J. and Gibson, L. (2012). Performance of single and multi-strain probiotics during hatchery production of Greenshell<sup>TM</sup> mussel larvae, *Perna canaliculus. Aquaculture*. 354: 56-63.
- Kim, B. H., Ramanan, R., Cho, D. H., Oh, H. M. and Kim, H. S. (2014). Role of *Rhizobium*, a plant growth promoting bacterium, in enhancing algal biomass through mutualistic interaction. *Biomass and Bioenergy*. 69: 95-105.
- Khochamit, N., Siripornadulsil, S., Sukon, P., and Siripornadulsil, W. (2014). Antibacterial activity and genotypic–phenotypic characteristics of bacteriocinproducing *Bacillus subtilis* KKU213: potential as a probiotic strain. *Microbiological Research*. 170: 36-50.
- Kuberan, T., Sangaralingam, S. and Thirumalaiarasu, V. (2010). Isolation and optimization of protease producing bacteria from halophilic soil. *Journal of Bioscience*. 1(3): 163-174.
- Kumar, A., Ergas, S., Yuan, X., Sahu, A., Zhang, Q. and Dewulf, J. (2010). Enhanced carbon dioxide fixation and biofuel production via microalgae: recent development and future directions. *Trends in Biotechnology*. 28: 371-80.
- Kumar, G., Zhen, G., Sivagurunathan, P., Bakonyi, P., Nemestóthy, N., Bélafi-Bakó, K. and Xu, K. Q. (2016). Biogenic H2 production from mixed microalgae biomass: impact of pH control and methanogenic inhibitor (BESA) addition. *Journal* of Biofuel Research. 3(3): 470-474.
- Kushmaro, A., Banin, E., Loya, Y., Stackebrandt, E. and Rosenberg, E. (2001). Vibrio shiloi sp. nov., the causative agent of bleaching of the coral Oculina patagonica. International Journal of Systematic and Evolutionary Microbiology. 51(4): 1383-1388.
- Lafferty, K. D., Harvell, C. D., Conrad, J. M., Friedman, C. S., Kent, M. L., Kuris, A. M. and Saksida, S. M. (2015). Infectious diseases affect marine fisheries and aquaculture economics. *The Annual Review of Marine Science*. 7: 96-471
- Lam, K. S. (2006). Discovery of novel metabolites from marine actinomycetes. Current Opinion in Microbiology. 9(3): 245-251.
- Lalloo, R., Ramchuran, S., Ramduth, D., Görgens, J. and Gardiner, N. (2007). Isolation and selection of *Bacillus* spp. as potential biological agents for enhancement of water quality in culture of ornamental fish. *Journal of Applied Microbiology*. 103(5): 1471-1479.
- Lategan, M. J., Booth, W., Shimmon, R. and Gibson, L. F. (2006). An inhibitory substance produced by *Aeromonas* media A199, an aquatic probiotic. *Aquaculture*. 254(1-4): 115-124.

- Lazado, C. C. and Caipang, C. M. A. (2014). Atlantic cod in the dynamic probiotics research in aquaculture. *Aquaculture*. 424: 53-62.
- Lazado, C. C., Caipang, C. M. A. and Estante, E. G. (2015). Prospects of host-associated microorganisms in fish and penaeids as probiotics with immunomodulatory functions. *Fish and Shellfish Immunology*. 45(1): 2-12.
- Le, Chevanton, M., Garnier, M., Bougaran, G., Schreiber, N., Lukomska, E., Bérard, J. B. and Cadoret, J. P. (2013). Screening and selection of growth-promoting bacteria for *Dunaliella* cultures. *Algal Research*. 2(3): 212-222.
- Lee, Y. K. (2001). Microalgal mass culture systems and methods: their limitations and potential. *Journal of Applied Phycology*. 13: 307-315.
- Lee, C. S. (2015). Dietary nutrients, additives and fish health. John Wiley and Sons. Leifert, C., Li, H., Chidburee, S., Hampson, S., Workman, S., Sigee, D. and Harbour, A. (1995). Antibiotic production and biocontrol activity by *Bacillus subtilis* CL27 and *Bacillus pumilus* CL45. *Journal of Applied Bacteriology*. 78(2): 97-108.
- Li, P., Wen, Q. and Gatlin, D.M. (2009). Dose-dependent influences of dietary β-1,3glucan on innate immunity and disease resistance of hybrid striped bass *Morone chrysops* x *Morone saxatilis*. *Aquaculture Research*. 40: 1578-1584.
- Li, Y., Horsman, M., Wu, N., Lan, C. Q. and Dubois- Calero, N. (2008). Biofuels from microalgae. *Biotechnology Progress*. 24(4): 815-820.
- Lorincz, Z., Preininger, E., Kosa, A., Ponyi, T., Nyitrai, T. and Sarkadi, L. (2010). Artificial tripartite symbiosis involving a green alga (*Chlamadomonas*), a bacterium (*Azotobacter*) and a fungus (*Alternaria*): morphological and physiological characterization. *Folia Microbial (Praha.)*. 55: 393-400.
- Lisboa, M. P., Bonatto, D., Bizani, D., Henriques, J. A. and Brandelli, A. (2006). Characterization of a bacteriocin-like substance produced by *Bacillus amyloliquefaciens* isolated from the Brazilian Atlantic forest. *International Microbiology*. 9(2): 111-118.
- Loh, J. Y. and Ting, A. S. Y. (2016). Effects of potential probiotic *Lactococcus lactis* subsp. lactis on digestive enzymatic activities of live feed *Artemia franciscana*. *Aquaculture International*. 24(5): 1341-1351.
- Loh, J. Y., Kay, G. L. and Ting, A. S. Y. (2018). Bioencapsulation and colonization characteristics of *Lactococcus lactis* subsp. *lactis* CF4MRS in *Artemia franciscana*: A biological approach for the control of *Edwardsiellosis* in larviculture. *Marine Biotechnology*. 20(3): 353-362.
- Liang, Y., Sarkany, N. and Cui, Y. (2009). Biomass and lipid productivities of *Chlorella vulgaris* under autotrophic, heterotrophic and mixotrophic growth conditions. *Biotechnology Letters*. 31: 1043-1049.

- Lavilla-Pitogo, C. R., Baticados, M. C. L., Cruz-Lacierda, E. R. and Leobert, D. (1990). Occurrence of luminous bacterial disease of *Penaeus monodon* larvae in the Philippines. *Aquaculture*. 91(1-2): 1-13.
- Lin, C.K. (1995). Progression of intensive marine shrimp culture in Thailand. In C.L. Browdy and J.S. Hopkins (ed.), Swimming through troubled water. Proceedings of the Special Session on Shrimp Farming, Aquaculture '95. World Aquculture Society. 13-23.
- Liu, J., Lewitus, A. J., Brown, P. and Wilde, S. B. (2008). Growth-promoting effects of a bacterium on raphidophytes and other phytoplankton. *Harmful Algae*. 7(1): 1-10.
- Mata, T. M., Martins, A. A., and Caetano, N. S. (2010). Microalgae for biodiesel production and other applications: a review. *Renewable and Sustainable Energy Reviews*. 14(1): 217-232.
- Mahdhi, A., Hmila, Z., Chaieb, K., Kamoun, F. and Bakhrouf, A. (2011). Probiotic properties of halophilic *Bacillus* strains enhance protection of *Artemia* culture against pathogenic *Vibrio*. *Aquatic Biology*. 13(3): 225-231.
- Marques, A., Thanh, T. H., Sorgeloos, P. and Bossier, P. (2006). Use of microalgae and bacteria to enhance protection of gnotobiotic *Artemia* against different pathogens. *Aquaculture*. 258(1-4): 116-126.
- Masurkar, A. A. K., Datar, A. G. and Vakil, B. V. (2015). New microbial sources of PUFA from the Arabian Sea and fresh water habitats of Indian Territory. *International Journal of Pharmaceutical Sciences and Research*. 6(1): 349-360.
- Matsui, K., Nobuyoshi, I. and Kawabata, Z. (2003). Release of extracellular transformable plasmid DNA from *Escherichia coli* co-cultivated with algae. *Applied and Environmental Microbiology*. 69: 2399-2404.
- Mayali, X. and Doucette, G. J. (2002). Microbial community interactions and population dynamics of an algicidal bacterium active against *Karenia brevis* (Dinophyceae). *Harmful Algae*. 1(3): 277-293.
- McClean, K. H., Winson, M. K., Fish, L., Taylor, A., Chhabra, S. R., Camara, M. and Stewart, G. S. (1997). Quorum sensing and *Chromobacterium violaceum*: exploitation of violacein production and inhibition for the detection of Nacylhomoserine lactones. *Microbiology*. 143(12): 3703-3711.
- McKeen, C. D., Reilly, C. C., & Pusey, P. L. (1986). Production and partial characterization of antifungal substances antagonistic to *Monilinia fructicola* from *Bacillus subtilis*. *Phytopathology*. 76(2): 136-139.
- Merchie, G., Lavens, P., Dhert, P. H., Dehasque, M., Nelis, H., De Leenheer, A. and Sorgeloos, P. (1995). Variation of ascorbic acid content in different live food organisms. *Aquaculture*. 134: 325-337.

- Mohebbi, F. (2010). The Brine Shrimp *Artemia* and hypersaline environments microalgal composition: a mutual interaction. *International Journal of Aquatic Science*. 1(1): 19-27.
- Mohebbi, F., Mohsenpour Azari, A., Ahmadi, R., Seidgar, M., Mostafazadeh, B. and Ganji, S. (2015). The Effects of *Dunaliella tertiolecta*, *Tetraselmis suecica* and *Nannochloropsis oculata* as food on the growth, survival and reproductive characteristics of *Artemia urmiana*. *Environmental Resources Research*. 3(2): 111-120.
- Moriarty, D. J. W. (1998). Control of luminous Vibrio species in penaeid aquaculture ponds. Aquaculture. 164(1-4): 351-358.
- Mouget, J. L., Dakhama, A., Lavoic, M.C. and de la Noüe, J. (1995). Algal growth enhancement by bacteria: is consumption of photosynthetic oxygen involved? *Microbiology Ecology*. 18: 35-44.
- Muñoz-Atienza, E., Gómez-Sala, B., Araújo, C., Campanero, C., Del Campo, R., Hernández, P. E. and Cintas, L. M. (2013). Antimicrobial activity, antibiotic susceptibility and virulence factors of lactic acid bacteria of aquatic origin intended for use as probiotics in aquaculture. *Journal of Microbiology*. 13(1): 15.
- Merrifield, D. L., Dimitroglou, A., Foey, A., Davies, S. J., Baker, R. T., Bøgwald, J. and Ringø, E. (2010). The current status and future focus of probiotic and prebiotic applications for salmonids. *Aquaculture*. 302(1-2): 1-18.
- Murray, R. E., Cooksey, K. E., Priscu, J. C. (1996). Simulation of bacterial DNA synthesis by algal exudates in attached algal-bacterial consortia. *Applied and Environmental Microbiology*. 52: 1777-1182.
- Murray, H. M., Gallant, J. W., Perez- Casanova, J. C., Johnson, S. C. and Douglas, S. E. (2003). Ontogeny of lipase expression in winter flounder. *Journal of Fish Biology*. 62(4): 816-833.
- Munirasu, S., Ramasubramaniam, V. and Venkatachalam, U. (2013). Bioenrichment of live feed *Daphnia magna* for the survival and growth of freshwater *Catla catla*. *International Journal of Current* Research and Academic Review. 5(8): 20-31.
- Munoz, R. and Guieyse, B. (2006) Algal-bacterial processes for the treatment of hazardous contaminants: a review. *Water Research*. 40(15): 2799-2815.
- Miao, X. and Wu, Q. (2006). Biodiesel production from heterotrophic microalgal oil. *Bioresource Technology*. 97(6): 841-846.
- Muller-Feuga, A. (2000). The role of microalgae in aquaculture: situation and trends. *Journal of Applied Phycology*. 12(3-5): 527-534.
- Mustafa, M. G. (1995). A review: Dietary benefits of algae as an additive in fish feed. *The Israeli Journal of Aquaculture*. 47: 155-162.

- Nakano, S. (1996). Bacterial response to extracellular dissolved organic carbon released from healthy and senescent *Flagilaria crotonensis* (*Bacillariophyceae*) in experimental systems. *Hydrobiologia*. 339: 47-55.
- Nakayama, T., Lu, H. and Nomura, N. (2009). Inhibitory effects of *Bacillus* probionts on growth and toxin production of *Vibrio harveyi* pathogens of shrimp. *Letters in Applied Microbiology*. 49(6): 679-684.
- Natrah, F. M. I., Bossier, P., Sorgeloos, P., Yusoff, F. M. and Defoirdt, T. (2013). Significance of microalgal-bacterial interactions for aquaculture. *Reviews in Aquaculture*. 5:1-14.
- Naumann, T., Çebi, Z., Podola, B. and Melkonian, M. (2013). Growing microalgae as aquaculture feeds on twin-layers: a novel solid-state photobioreactor. *Journal of Applied Phycology*. 25(5): 1413-1420.
- Nayak, S. K. (2010). Probiotics and immunity: a fish perspective. *Fish and Shellfish Immunology*. 29(1): 2-14.
- Newaj-Fyzul, A., Al-Harbi, A. H. and Austin, B. (2014). Developments in the use of probiotics for disease control in aquaculture. *Aquaculture*. 431: 1-11.
- Nithya, C. and Pandian, S. K. (2010). The *in vitro* antibiofilm activity of selected marine bacterial culture supernatants against *Vibrio* spp. *Archives of Microbiology*. 192(10): 843-854.
- Niu, Y. (2014). Host-microbe interactions in brine shrimp Artemia cultured under gnotobiotic and conventional rearing conditions (Doctoral dissertation, Ghent University). 1-238.
- Nguyen, T. H. and Nguyen, V. D. (2017). Characterization and applications of marine microbial enzymes in biotechnology and probiotics for animal health. *Advances in Food and Nutrition Research*. 80: 37-74.
- Nwogu, N. A., Olaji, E. D. and Egomwanre, A. F. (2011). Application of probiotics in Nigerian Aquaculture: current status, challenges and prospects. *International Research Journal of Microbiology*. 2(7): 215-219.
- Ortiz-Marquez, J.C., Do, Nascimento, M., Dublan, Mde, L. and Curatti, L. (2012). Association with an ammonium-excreting bacterium allows diazotrophic culture of oil-rich eukaryotic microalgae. *Applied and Environmental Microbiology*. 78: 2345-2352.
- Oviya, M., Giri, S. S., Sukumaran, V. and Natarajan, P. (2012). Immobilization of carbonic anhydrase enzyme purified from *Bacillus subtilis* VSG-4 and its application as *CO*<sup>2</sup> sequesterer. Preparative *Biochemistry and Biotechnology* 42: 462-475.

- Pan, X., Wu, T. and Song, Z. (2008). Immune responses and enhanced disease resistance in Chinese drum, *Miichthys miiuy* (Basilewsky), after oral administration of live or dead cells of *Clostridium butyrium* CB2. *Journal of Fish Disease*. 31: 679-686.
- Pandiyan, P., Balaraman, D., Thirunavukkarasu, R., George, E. G. J., Subaramaniyan, K., Manikkam, S. and Sadayappan, B. (2013). Probiotics in aquaculture. *Drug Invention Today*. 5(1): 55-59.
- Panigrahi, A. and Azad, I. S. (2007). Microbial intervention for better fish health in aquaculture: the Indian scenario. *Fish Physiology and Biochemistry*. 33(4): 429-440.
- Pande, G. S. J., Scheie, A. A., Benneche, T., Wille, M., Sorgeloos, P., Bossier, P. and Defoirdt, T. (2013). Quorum sensing-disrupting compounds protect larvae of the giant freshwater prawn *Macrobrachium rosenbergii* from *Vibrio harveyi* infection. *Aquaculture*. 406: 121-124.
- Panel, E. F. (2012). Guidance on the assessment of bacterial susceptibility to antimicrobials of human and veterinary importance. *Journal of Food Safety and Authority*. 10(6): 2740.
- Park, Y., Je, K-W., Lee, K., Jung, S-E. and Choi, T-J. (2008). Growth promotion of *Chlorella ellipsoiden* by co-inoculation with *Brevundimonas sp.* isolated from the microalga. *Hydrobiologia*. 598: 219-228.
- Park, J., Park, B. S., Wang, P., Patidar, S. K., Kim, J. H., Kim, S. H. and Han, M. S. (2017). Phycospheric native bacteria *Pelagibaca bermudensis* and *Stappia* sp. ameliorate biomass productivity of *Tetraselmis striata* (KCTC1432BP) in cocultivation system through mutualistic interaction. *Frontiers in Plant Science*. 8: 289.
- Patil, J. S. and Anil, A. C. (2005). Influence of diatom exopolymers and biofilms on metamorphosis in the barnacle *Balanus amphitrite*. *Marine Ecology Progress Series*. 301: 231-245.
- Pradhan, A. K., Pradhan, N., Sukla, L. B., Panda, P. K. and Mishra, B. K. (2014). Inhibition of pathogenic bacterial biofilm by biosurfactant produced by *Lysinibacillusfusiformis* S9. *Bioprocess and Biosystems Engineering*. 37(2): 139-149.
- Paul, C. and Pohnert, G. (2011). Interactions of the algicidal bacterium Kordia algicida with diatoms: regulated protease excretion for specific algal lysis. *PloS One*. 6(6): e21032.
- Planas, M., Vázquez, J. A., Marqués, J., Pérez-Lomba, R., González, M. and Murado, M. (2004). Enhancement of rotifer (*Brachionus plicatilis*) growth by using terrestrial lactic acid bacteria. *Aquaculture*. 240(1-4): 313-329.

- Planas, M., Vázquez, J. A. and Novoa, B. (2015). Stimulative effect of lactic acid bacteria in the growth of the microalgae *Isochrysis galbana*. *Journal of Coastal Life Medicine*. 3(12): 925-930.
- Prabha, M. S., Divakar, K., Priya, J. D. A., Selvam, G. P., Balasubramanian, N. and Gautam, P. (2015). Statistical analysis of production of protease and esterase by a newly isolated *Lysinibacillus fusiformis* AU01: purification and application of protease in sub-culturing cell lines. *Annals of Microbiology*. 65(1): 33-46.
- Priya, T. and Usharani, G. (2009). Comparative study for biosurfactant production by using *Bacillus subtilis* and *Pseudomonas aeruginosa*. *Botany Research International*. 2(4): 284-287.
- Pulz, O. and Gross, W. (2004). Valuable products from biotechnology of microalgae. *Applied Microbiology and Biotechnology*. 65: 635-648.
- Quesada, S. P., Paschoal, J. A. R. and Reyes, F. G. (2013). A simple method for the determination of fluoroquinolone residues in tilapia (*Oreochromis niloticus*) and pacu (*Piaractus mesopotamicus*) employing LC-MS/MS QToF. *Food Additives and Contaminants*: 30(5): 813-825.
- Rabin, N., Zheng, Y., Opoku-Temeng, C., Du, Y., Bonsu, E. and Sintim, H. O. (2015). Agents that inhibit bacterial biofilm formation. *Future Medicinal Chemistry*. 7(5): 647-671.
- Raja, R.A. and Jithendran, K.P. (2015). Aquaculture disease diagnosis and health management. *Advances in Marine and Brackishwater Aquaculture*. 247-255.
- Ransangan, J., Imm, L. K. L., Lal, T. M. and Sade, A. (2013). Phenotypic characterization and antibiotic susceptibility of *Vibrio* spp. isolated from aquaculture waters on the west coast of Sabah, Malaysia. *International Journal* of Research in Pure and Applied Microbiology. 3(3): 58-66.
- Reda, R. M., Selim, K. M., El-Sayed, H. M. and El-Hady, M. A. (2017). In vitro selection and identification of potential probiotics isolated from the gastrointestinal tract of nile Tilapia, Oreochromis niloticus. Probiotics and Antimicrobial Proteins. 1-12.
- Reid, G., Sanders, M. E., Gaskins, H. R., Gibson, G. R., Mercenier, A., Rastall, R. and Klaenhammer, T. R. (2003). New scientific paradigms for probiotics and prebiotics. *Journal of Clinical Gastroenterology*. 37(2): 105-118.
- Reid, H. I., Adam, B. and Birkbeck, T. H. (2009). Analysis of bacterial populations in the gut of developing cod larvae and identification of *Vibrio logei*, *Vibrio anguillarum* and *Vibrio splendidus* as pathogens of cod larvae. *Aquaculture*. 288(1-2): 36-43.
- Ren, D., Bedzyk, L. A., Rick, W. Y., Thomas, S. M. and Wood, T. K. (2004). Stationaryphase quorum-sensing signals affect autoinducer-2 and gene expression in *Escherichia coli*. Applied and Environmental Microbiology. 70(4): 2038-2043.

- Rengipipat, S., Rukpratanporn, S., Piyatiratitivorakul, S. and Menasaveta, P. (2000). Immunity enhancement in black tiger shrimp (*Penaeus monodon*) by a probiont bacterium (*Bacillus* S11). *Aquaculture*. 191(4): 271-288.
- Rehberg-Haas, S., Meyer, S., Tielmann, M., Lippemeier, S., Vadstein, O., Bakke, I. and Schulz, C. (2015). Use of the microalga *Pavlova viridis* as enrichment product for the feeding of Atlantic cod larvae (*Gadus morhua*). *Aquaculture*. 438: 141-150.
- Rekiel, A., Wiecek, J., Bielecki, W., Gajewska, J., Cichowicz, M., Kulisiewicz, J. and Beyga, K. (2007). Effect of addition of feed antibiotic flavomycin or prebiotic BIO-MOS on production results of fatteners, blood biochemical parameters, morphometric indices of intestine and composition of microflora. *Archives of Animal Breeding*. 50: 172-180.
- Rosenberg, J.N., Oyler, G.A., Wilkinson, L. and Betenbaugh, M.J. (2008). A green light for engineered algae: directing metabolism to fuel and biotechnology revolution. *Current Opinion in Biotechnology*. 19: 430-436.
- Roy, S. S. and Pal, R. (2013). Microalgae in aquaculture: a review with special references to nutritional value and fish dietetics. In *Proceedings of The Zoological Society*. 68(1): 1-8.
- Ribalet, F., Intertaglia, L., Lebaron, P. and Casotti, R. (2008). Differential effect of three polyunsaturated aldehydes on marine bacterial isolates. *Aquatic Toxicology*. 86: 249-255.
- Reid, G., Dols, J. and Miller, W. (2009). Targeting the vaginal microbiota with probiotics as a means to counteract infections. *Current Opinion in Clinical Nutrition and Metabolic Care.* 12(6): 583-587.
- Rico-Mora, R., Voltolina, D. and Villaescusa-Celaya, J. A. (1998). Biological control of *Vibrio alginolyticus* in *Skeletonema costatum* (*Bacillariophyceae*) cultures. *Aquacultural Engineering*. 19(1): 1-6.
- Rier, S. T. and Stevenson, R. J. (2002). Effects of light, dissolved organic carbon, and inorganic nutrients [2pt] on the relationship between algae and heterotrophic bacteria in stream periphyton. *Hydrobiologia*. 489(1-3): 179-184.
- Rivas, M. O., Vargas, P. and Riquelme, C. E. (2010). Interactions of *Botryococcus braunii* cultures with bacterial biofilms. *Microbial Ecology*. 60: 628-635.
- Ruwandeepika Hettipala Arachchige, D. (2010). Expression of virulence genes of vibrios belonging to the *Harveyi* clade in the brine shrimp *Artemia* (Doctoral dissertation, Ghent University). Retrieved from <u>https://biblio.ugent.be/publication/1081411</u>.
- Santhakumari, S., Kannappan, A., Pandian, S. K., Thajuddin, N., Rajendran, R. B. and Ravi, A. V. (2016). Inhibitory effect of marine cyanobacterial extract on biofilm formation and virulence factor production of bacterial pathogens causing vibriosis in aquaculture. *Journal of Applied Phycology*. 28(1): 313-324.

- Sayes, C., Leyton, Y. and Riquelme, C. (2018). Probiotic bacteria as an healthy alternative for fish aquaculture. *InTech.* 7: 115-132.
- Seenivasan, C., Bhavan, P. S., Radhakrishnan, S. and Shanthi, R. (2012). Enrichment of Artemia nauplii with Lactobacillus sporogenes for enhancing the survival, growth and levels of biochemical constituents in the post-larvae of the freshwater prawn Macrobrachium rosenbergii. Turkish Journal of Fisheries and Aquatic Sciences. 12(1).
- Servin, A. L. (2004). Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. *Microbiology Reviews*. 28(4): 405-440.
- Silva, E. F., Soares, M. A., Calazans, N. F., Vogeley, J. L., do Valle, B. C., Soares, R. and Peixoto, S. (2013). Effect of probiotic (*Bacillus* spp.) addition during larvae and postlarvae culture of the white shrimp *Litopenaeus vannamei*. *Aquaculture Research*. 44(1): 13-21.
- Silo-Suh, L. A., Lethbridge, B. J., Raffel, S. J., He, H., Clardy, J. and Handelsman, J. (1994). Biological activities of two fungistatic antibiotics produced by *Bacillus cereus* UW85. *Applied and Environmental Microbiology*. 60(6): 2023-2030.
- Singh, R. P., Mantri, V. A., Reddy, C. R. K. and Jha, B. (2011). Isolation of seaweedassociated bacteria and their morphogenesis-inducing capability in axenic cultures of the green alga *Ulva fasciata*. *Aquatic Biology*. 12(1): 13-21.
- Singh, S. K., Major, S. R., Cai, H., Chen, F., Hill, R. T. and Li, Y. (2018). Draft Genome Sequences of *Cloacibacterium normanense* IMET F, a Microalgal Growth-Promoting Bacterium, and *Aeromonas jandaei* IMET J, a Microalgal Growth-Inhibiting Bacterium. *Genome announcements*. 6(24): 1-18.
- Siriyappagouder, P., Shankar, K. M., Kumar, B. N., Patil, R. and Byadgi, O. V. (2014). Evaluation of biofilm of *Aeromonas hydrophila* for oral vaccination of *Channa striatus*. *Fish and Shellfish Immunology*. 41(2): 581-585.
- Snieszko, S. F. (1974). The effects of environmental stress on outbreaks of infectious diseases of fishes. *Journal of Fish Biology*. 6(2): 197-208.
- Soltani, N., Latifi, A.M., Alnajar, N., Dezfulian, M., Shokarvi, S., Heydari I. and Choopani, A. (2016). Biochemical and physiological characterization of tree microalgae spp. as candidates for food supplement. *Journal of Applied Biotechnology*. 3: 377-381.
- Sorgeloos, P., Lavens, P., Leger, P. and Tackaert, W. (1991). State of the art in larviculture of fish and shellfish. *Larviculture*. 91: 3-5.
- Sorgeloos, P., Dhert, P. and Candreva, P. (2001). Use of brine shrimp, *Artemia* sp., in marine fish larviculture. *Aquaculture*. 200: 147-159.
- Subasinghe, R. (1997). Fish health and quarantine; In reviews of the state of the world aquaculture. FAO Fisheries circular no. 886. *Food and Agriculture Organization of the United Nations, Rome, Italy.* 45-49.

- Song, S. K., Beck, B. R., Kim, D., Park, J., Kim, J., Kim, H. D. and Ringo, E. (2014). A review: Prebiotics as immunostimulants in aquaculture. *Fish and Shellfish Immunology*. 40: 40-48.
- Skjelbred, B., Edvardsen, B. and Andersen, T. (2012). A high-throughput method for measuring growth and loss rates in microalgal cultures. *Journal of Applied Phycology*. 24(6): 1589-1599.
- Stanković, S., Mihajlović, S., Draganić, V., Dimkić, I., Vukotić, G., Berić, T. and Fira, Đ. (2012). Screening for the presence of biosynthetic genes for antimicrobial lipopeptides in natural isolates of *Bacillus* sp. *Archives of Biological Sciences*. 64(4): 1425-1432.
- Subashchandrose, S. R., Ramakrishnan, B., Megkaraj, M., Venkateswarlu, K. and Naidu, R. (2011). Consortia of cyanobacteria, microalgae and bacteria: biotechnological potential. *Biotechnology Advances*. 29: 896-907.
- Sugita, H., Hirose, Y., Matsuo, N. and Deguchi, Y. (1998). Production of the antibacterial substance by *Bacillus* sp. strain NM 12, an intestinal bacterium of Japanese coastal fish. *Aquaculture*. 165(3-4): 269-280.
- Sun, Y. Z., Yang, H. L., Ma, R. L. and Lin, W. Y. (2010). Probiotic applications of two dominant gut *Bacillus* strains with antagonistic activity improved the growth performance and immune responses of grouper *Epinephelus coioides*. *Fish and Shellfish Immunology*. 29(5): 803-809.
- Talpur, A. D., Memon, A. J., Khan, M. I., Ikhwanuddin, M., Daniel, M. D. and Abol-Munafi, A. B. (2011). Supplementation of indigenous *Lactobacillus* bacteria in live prey and as water additive to larviculture of *Portunus pelagicus* (Linnaeus, 1758). Advance Journal of Food Science and Technology. 3(5): 390-398.
- Tandon, P. and Jin, Q. (2017). Microalgae culture enhancement through key microbial approaches. *Renewable and Sustainable Energy Reviews*. 80: 1089-1099.
- Tanabe, Y., Okazaki, Y., Yoshida, M., Matsuura, H., Kai, A., Shiratori, T. and Watanabe, M. M. (2015). A novel alphaproteobacterial ectosymbiont promotes the growth of the hydrocarbon-rich green alga *Botryococcus braunii*. *Nature*. 5: 10467.
- Toi, H. T., Boeckx, P., Sorgeloos, P., Bossier, P. and Van Stappen, G. (2014). Co-feeding of microalgae and bacteria may result in increased N assimilation in *Artemia* as compared to mono-diets, as demonstrated by a <sup>15</sup> N isotope uptake laboratory study. *Aquaculture*. 422: 109-114.
- Toyama, T., Hanaoka, T., Tanaka, Y., Morikawa, M. and Mori, K. (2018). Comprehensive evaluation of nitrogen removal rate and biomass, ethanol, and methane production yields by combination of four major duckweeds and three types of wastewater effluent. *Bioresource Technology*. 250: 464-473.

- Tinh, N. T. N., Dierckens, K., Sorgeloos, P. and Bossier, P. (2007). A review of the functionality of probiotics in the larviculture food chain. *Marine Biotechnology*. 10(1): 1-12.
- Thenmozhi, R., Nithyanand, P., Rathna, J. and Karutha Pandian, S. (2009). Antibiofilm activity of coral-associated bacteria against different clinical M serotypes of *Streptococcus pyogenes*. *Immunology and Medical Microbiology*. 57(3): 284-294.
- Thomas, T. E. and Robinson, M. G. (1987). The role of bacteria in the metal tolerance of the fouling diatom Amphora coffeaeformis Ag. Journal of Experimental Marine Biology and Ecology. 107(3): 291-297.
- Thompson, F. L., Lida, T. and Swings, J. (2004). Biodiversity of Vibrios. Microbiology and Molecular Biology Reviews. 68: 403-431.
- Tran, L., Nunan, L., Redman, R. M., Mohney, L. L., Pantoja, C. R., Fitzsimmons, K. and Lightner, D. V. (2013). Determination of the infectious nature of the agent of acute hepatopancreatic necrosis syndrome affecting penaeid shrimp. *Diseases* of Aquatic Organisms. 105(1): 45-55.
- Troell, M., Naylor, R., Metian, M., Beveridge, M., Tyedmers, P. H., Folke, C., Arrow, K. J., Barrett, S., Crepin, A., Ehrlich, P. R., Gren, A., Kautsky, N., Levin, S.A., Nyborg, K., Osterblom, H., Polasky, S., Scheffer, M., Walker, B. H., Xepapadeas, T.and Zeeuw, A. (2014). Does aquaculture add resilience to global food system? *Proceedings of the National Academy of Sciences of the United States of Amerika*. 111: 13257-13263.
- Tuan, T. N., Duc, P. M. and Hatai, K. (2013). Overview of the use of probiotics in aquaculture. *International Journal of Research in Fisheries and Aquaculture*. 3(3): 89-97.
- Vaseeharan, B. A. R. P. and Ramasamy, P. (2003). Control of pathogenic Vibrio spp. by Bacillus subtilis BT23, a possible probiotic treatment for black tiger shrimp Penaeus monodon. Letters in Applied Microbiology. 36(2): 83-87.
- Vázquez-Silva, G., Ramírez-Saad, H. C., Aguirre-Garrido, J. F., Mayorga-Reyes, L., Azaola-Espinosa, A. and Morales-Jiménez, J. (2017). Effect of bacterial probiotics bio-encapsulated into Artemia franciscana on weight and length of the shortfin silverside (*Chirostoma humboldtianum*), and PCR-DGGE characterization of its intestinal bacterial community. Latin American Journal of Aquatic Research. 45(5): 1031-1043.
- Vezzulli, L., Colwell, R. R. and Pruzzo, C. (2012). Ocean warming and spread of pathogenic vibrios in the aquatic environment. *Microbial Ecology*. 65(4): 817-825.
- Vezzulli, L. I., Brettar, E., Pezzati, P. C., Reid, R. R., Colwell, M. and Ghofle (2013). Long term effects of ocean warming on the prokaryotic community: evidence from the vibriosis. *Journal of Microbial Ecology*. 6: 21-30.

- Villamil, L., Figueras, A., Planas, M. and Novoa, B. (2010). *Pediococcus acidilactici* in the culture of turbot (*Psetta maxima*) larvae: administration pathways. *Aquaculture*. 307(1-2): 83-88.
- Villamil, L., Figueras, A., Planas, M. and Novoa, B. (2003). Control of Vibrio alginolyticus in Artemia culture by treatment with bacterial probiotics. Aquaculture. 219(1-4): 43-56.
- Villamil, L., Tafalla, C., Figueras, A. and Novoa, B. (2002). Evaluation of immunomodulatory effects of lactic acid bacteria in turbot (*Scophthalmus* maximus). Clinical and Diagnostic Laboratory Immunology. 9(6): 1318-1323.
- Vinod, M. G., Shivu, M. M., Umesha, K. R., Rajeeva, B. C., Krohne, G., Karunasagar, I.and Karunasagar, I. (2006). Isolation of *Vibrio harveyi* bacteriophage with a potential for biocontrol of luminous vibriosis in hatchery environments. *Aquaculture*. 255(1-4): 117-124.
- Verschuere, L., Rombaut, G., Sorgeloos, P. and Verstraete, W. (2000). Probiotic bacteria as biological control agents in aquaculture. *Microbiology and Molecular Biology Reviews*. 64: 655-671.
- Verschuere, L., Heang, H., Criel, G., Sorgeloos, P. and Verstraete, W. (2000). Selected bacterial strains protect Artemia spp. from the pathogenic effects of Vibrio proteolyticus CW8T2. Applied and Environmental Microbiology. 66(3): 1139-1146.
- Volkman, J. K. and Brown, M. R. (2006). Nutritional value of microalgae and applications. Algal cultures, Analogues of blooms and Applications. 1: 407-457.
- Vu, H.T., Otsuka, S., Ueda, H. and Senoo, K. (2010). Cocultivated bacteria can increase or decrease the culture lifetime of *Chlorella vulgaris*. Journal of General and Applied Microbiology. 65:2527-2533.
- Wang, H., Hill, R. T., Zheng, T., Hu, X. and Wang, B. (2016). Effects of bacterial communities on biofuel-producing microalgae: stimulation, inhibition and harvesting. *Critical Reviews in Biotechnology*. 36(2): 341-352.
- Wang, W., Sun, J., Liu, C. and Xue, Z. (2017). Application of immunostimulants in aquaculture: current knowledge and future perspectives. *Aquaculture Research*. 48: 1-23.
- Watanabe, K., Takihana, N., Aoyagi, H., Hanada, S., Watanabe, Y. and Ohmura, N. (2005). Symbiotic association in *Chlorella* culture. *Microbiology Ecology*. 51: 187-196.
- Witte, W., Klare, I. and Werner, G. (1990). Selective pressure by antibiotics as feed additives. *Infection*. 272: 35-38.

- Yang, C., Hua, Q. and Shimuzu, K. (2000). Energetics and carbon metabolism during growth of microalgal cells under photoautotrophic, mixotrophic and cyclic light-autotrophic I and dark heterotrophic conditiond. *Journal of Biochemical Engineering*. 6: 87-102.
- Yu, M. C., Li, Z. J., Lin, H. Z., Wen, G. L. and Ma, S. (2009). Effects of dietary medicinal herbs and *Bacillus* on survival, growth, body composition, and digestive enzyme activity of the white shrimp *Litopenaeus* vannamei. Aquaculture International. 17(4): 377-384.
- You, J., Xue, X., Cao, L., Lu, X., Wang, J., Zhang, L. and Zhou, S. (2007). Inhibition of Vibrio biofilm formation by a marine actinomycete strain A66. Applied Microbiology and Biotechnology. 76(5): 1137-1144.
- You, K. G., Bong, C. W. and Lee, C. W. (2016). Antibiotic resistance and plasmid profiling of *Vibrio* spp. in tropical waters of Peninsular Malaysia. *Environmental Monitoring and Assessment*. 188(3): 171.
- Yousef N. M. H. (2014) Characterization and antimicrobial activity of silver nanoparticles synthesized by rice straw utilizing bacterium (*Lysinibacillus fusiformis*). International Journal of Dental Research. 4(9): 1875–1879
- Zacarias- Soto, M., Lazo, J. P. and Viana, M. T. (2011). Effect of three probiotics administered through live feed on digestive enzyme activity in California halibut, *Paralichthyscalifornicus*, larvae. *Journal of the World Aquaculture Society*. 42(3): 321-331.
- Zink, I. C., Douillet, P. A. and Benetti, D. D. (2013). Improvement of rotifer *Brachionus plicatilis* population growth dynamics with inclusion of *Bacillus* spp. probiotics. *Aquaculture Research*. 44(2): 200-211.
- Zokaeifar, H., Babaei, N., Saad, C. R., Kamarudin, M. S., Sijam, K. and Balcazar, J. L. (2014). Administration of *Bacillus subtilis* strains in the rearing water enhances the water quality, growth performance, immune response, and resistance against *Vibrio harveyi* infection in juvenile white shrimp, *Litopenaeus vannamei*. Fish and Shellfish Immunology. 36(1): 68-74.