



UNIVERSITI PUTRA MALAYSIA

***METABOLOMIC PROFILING OF ANTIOXIDANT AND ANTI
INFLAMMATORY PROPERTIES IN DIATOM *Chaetoceros calcitrans*
EXTRACTS USING NMR AND UHPLC-MS COUPLED WITH
CHEMOMETRIC ANALYSIS***

AWANIS BINTI AZIZAN

IB 2019 10



**METABOLOMIC PROFILING OF ANTIOXIDANT AND ANTI-
INFLAMMATORY PROPERTIES IN DIATOM *Chaetoceros calcitrans*
EXTRACTS USING NMR AND UHPLC-MS COUPLED WITH
CHEMOMETRIC ANALYSIS**

By

AWANIS BINTI AZIZAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science**

April 2019

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

METABOLOMIC PROFILING OF ANTIOXIDANT AND ANTI-INFLAMMATORY PROPERTIES IN DIATOM *Chaetoceros calcitrans* EXTRACTS USING NMR AND UHPLC-MS COUPLED WITH CHEMOMETRIC ANALYSIS

By

AWANIS BINTI AZIZAN

April 2019

Chairman : Associate Professor Faridah Abas, PhD
Institute : Bioscience

Chaetoceros calcitrans is a diatom microalga that is known to be rich of amino acids, lipids, fatty acids and natural pigments identified as potentially important natural antioxidant and anti-inflammation. Nevertheless, little is known about the metabolome and the antioxidative and anti-inflammatory ability of the indigenous microalga, *C. calcitrans*. The main objectives of this study were to evaluate the metabolites that contributed to the antioxidant activity (DPPH*), nitric oxide (NO) inhibitory activity and total phenolic content (TPC) of *C. calcitrans*, extracted with different solvent polarities, including 70% ethanol, methanol, hexane, acetone, and chloroform using multi-platform metabolomics approaches. Nuclear magnetic resonance (NMR) coupled to multivariate data analysis (MVDA) was applied for the metabolomics profiling and relative quantification of the extracts. Further confirmation of the metabolites identification and quantitation were performed using ultra-high performance liquid chromatography mass spectrometry (UHPLC-MS). The results showed that acetone and chloroform (CHCl₃) extracts of *C. calcitrans* revealed higher levels of TPC with 30.79 and 25.41 mg GAE/g dw, respectively. Both extracts also displayed moderate activity of DPPH radical scavenging inhibition with 43.01 and 35.03% at concentration 333 µg/ml. Furthermore, the CHCl₃ extract inhibited the release of NO production from the LPS-activated RAW 264 cells with an IC₅₀ value of 3.46 µg/ml. Twenty-nine metabolites were identified via NMR analyses from *C. calcitrans* extracts including 6 fatty acids, cholesterol, 11 amino acids, 2 sugars and 1 sugar-alcohol, 6 carotenoids and 2 chlorophylls. The structures of the compounds were also confirmed using tandem mass spectrometry. The main identified secondary metabolites were carotenoids including fucoxanthin, lutein, astaxanthin, canthaxanthin, zeaxanthin and violaxanthin. Comparison of different extracts revealed clear differences in the metabolite profiles and the partial least square (PLS) model indicated that the carotenoids were significantly associated with the tested

bioactivities. The results suggested CHCl_3 and acetone extracts of *C. calcitrans* showed the abundance of high-value metabolites as markers for antioxidant and anti-inflammatory activities. The findings from this research may serve as a benchmark for future extraction processes particularly in recovering antioxidant and anti-inflammatory metabolites derived from diatom. These metabolites can be important active ingredients for medicinal preparation, functional foods, and cosmeceutical and nutraceutical applications.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PEMPROFILAN METABOLOM CIRI ANTIOKSIDAN DAN ANTI-RADANG
DALAM EKSTRAK MIKROALGA DIATOM *Chaetoceros calcitrans*
MENGUNAKAN GABUNGAN NMR DAN UHPLC-MS DENGAN ANALISIS
KEMOMETRIK**

Oleh

AWANIS BINTI AZIZAN

April 2019

Pengerusi : Professor Madya Faridah Abas, PhD
Institut : Biosains

Chaetoceros calcitrans merupakan sejenis diatom mikroalga yang kaya dengan asid amino, lipid, asid lemak, pigmen semulajadi yang dikenalpasti mempunyai potensi sebagai sumber penting antioksidan dan anti-radang semula jadi. Walau bagaimanapun, hanya sedikit maklumat yang diketahui mengenai metabolom dan kemampuan antioksidasi dan anti-radang oleh mikroalga tempatan, *C. calcitrans*. Objektif utama kajian ini adalah untuk menilai metabolit yang menyumbang terhadap aktiviti antioksidasi (DPPH*), aktiviti rencatan nitrik oksida (NO) dan juga jumlah kandungan fenolik (TPC) bagi *C. calcitrans* yang diekstrak dengan beberapa pelarut berketub yang berbeza seperti 70% etanol, metanol, heksana, aseton, dan klorofom dengan menggunakan pendekatan multi-platfom metabolomik. Resonans magnet nukleus (NMR) digabungkan dengan analisis data multivariat (MVDA) telah digunakan untuk memprofil metabolomik dan kuantifikasi relatif terhadap ekstrak. Pengesanan lanjut mengenai metabolit yang dikenal pasti dan dikuantifikasi telah dilakukan dengan menggunakan kromatografi cecair prestasi tinggi-spektrometri jisim (UHPLC-MS). Keputusan menunjukkan ekstrak aseton dan klorofom (CHCl₃) untuk sampel *C. calcitrans* memiliki tahap TPC yang tertinggi sebanyak 30.79 and 25.41 mg GAE/g dw setiap satu. Kedua ekstrak ini juga telah mempamerkan aktiviti antioksidasi yang sederhana melalui aktiviti pemerangkap radikal bebas DPPH sebanyak 43.01 and 35.03% pada kepekatan 333 µg/ml. Tambahan pula, ekstrak CHCl₃ telah merencatkan pembebasan NO dari dalam sel RAW 264 yang diaktifkan oleh LPS dengan nilai IC₅₀ sebanyak 3.46 µg/ml. Dua puluh sembilan metabolit telah dikenal pasti melalui analisis NMR dari ekstrak *C. calcitrans* yang terdiri daripada 6 asid lemak, kolestrol, 11 asid amino, 2 gula dan 1 gula-beralkohol, 6 karotenoid dan 2 klorofil. Struktur sebatian juga disahkan dengan menggunakan spektrometri jisim bergandingan. Metabolit sekunder utama yang telah dikenal pasti adalah karotenoid antaranya fucoxanthin, lutein, astaxanthin, canthaxanthin, zeaxanthin and violaxanthin. Perbandingan dengan ekstrak yang berbeza mendedahkan perbezaan

ketara profil metabolit dan model analisa separa persegi (PLS) menunjukkan karotenoid mempunyai kaitan yang signifikan dengan bioaktiviti yang dikaji. Keputusan kajian ini mencadangkan ekstrak CHCl_3 dan aseton untuk sampel *C. calcitrans* mempunyai kelimpahan metabolit yang bernilai tinggi yang bertindak sebagai penanda yang bertanggungjawab ke atas aktiviti antioksidan dan anti-radang. Hasil kajian dari penyelidikan ini boleh dijadikan penanda aras untuk proses pengekstrakan pada masa akan datang terutamanya dalam perolehan semula metabolit antioksidan dan anti-radang yang berasal dari diatom. Metabolit ini juga boleh dijadikan bahan aktif penting untuk diaplikasikan di dalam proses penyediaan ubat, makanan fungsian, kosmeseutikal dan nutraseutikal.



ACKNOWLEDGEMENTS

I am grateful for my Creator for always being there with me and giving me hope, love, healthy, functioning body and mind to work for this MSc project, hence completing my research project. The completion of this thesis project is made possible through the meaningful contribution of some people.

I wish to express my sincere thanks to Dr Faridah Abas, Prof. Khozirah Shaari, and Prof. Philip James Harris, my supervisor, co-supervisor and Honours supervisor for providing me endless guidance and encouragement during my research life. With their expertise in scientific research, funding, scientific writing, sincere, enthusiasms, research life have become more interesting and smooth for me. Working on journal writing is quite challenging, but, with their continuous efforts, brilliant ideas and encouragement, my skills in scientific writing are improving day by day.

Special thanks and love to Dr Maulidiani, who is formerly working as a post-doctoral candidate in LHS, IBS who has generously given her time and provided me much assistance with the metabolomics tools, statistical analysis and editing.

I place on record, my sincere thank you to Siti Zulaikha, Khaleeda Zulaikha, Nur Ashikin and Nawal Bubaker in the laboratory of LHS for their continuous support throughout my research. They helped me to overcome challenges and difficulties in carrying out the research work.

Big thanks to Muhammad Safwan Bustamam and the LHS staff for their knowledge and assistance in helping me with the identification and collection of microalga samples as well on laboratory techniques.

Last but not least, I would like to extend my special thank you to my dearest parents and my siblings for always being there for me during the ups and downs in my entire journey to finish my thesis, their continuous prayers and constant encouragements. It's really hard to adapt to the new working environment when I first arrived here in UPM. May Allah shower them with good health and happy life. And may Allah allowed me to repay their kindness and support in the near future. InsyaAllah.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Faridah Abas, PhD

Associate Professor
Institute Bioscience
Universiti Putra Malaysia
(Chairman)

Khozirah Shaari, PhD

Professor
Institute Bioscience
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No.: Awanis Binti Azizan GS48378

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory

Committee: Associate Professor Dr. Faridah Abas

Signature: _____

Name of Member
of Supervisory

Committee: Professor Dr. Khozirah Shaari

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Problem statement	2
1.3 Scope and Objectives	3
2 LITERATURE REVIEW	4
2.1 Microalgae	4
2.2 Utilization of microalgae	5
2.2.1 Commercial uses	6
2.2.2 Food for human	8
2.2.3 Food supplements for animals	8
2.2.4 Cosmeceuticals	9
2.2.5 Biofertilizers	10
2.2.6 Traditional medicinal uses	10
2.3 Microalgae components	11
2.3.1 Polyunsaturated fatty acids	11
2.3.2 Vitamins and minerals	14
2.3.3 Carotenoids	14
2.3.4 Chlorophylls	15
2.3.5 Phycobiliproteins	16
2.3.6 Polysaccharides	16
2.3.7 Sterols	17
2.3.8 Proteins	17
2.4 The health-promoting properties of microalga derived antioxidants	18
2.5 Diatom as sources of marine natural products	19
2.5.1 <i>Chaetoceros calcitrans</i>	21
2.5.2 Previous phytochemical and biological activity on <i>C. calcitrans</i>	22
2.6 Extraction of antioxidant metabolites from microalgae	23
2.6.1 Solvent extraction (SE)	23
2.7 Metabolomics	24
2.7.1 Overview of metabolomics and its application	24
2.7.2 Metabolomics workflow	25

2.7.3	Metabolomics in studying oxidative stress disorders	26
2.7.4	Use of NMR and UHPLC-MS-based metabolomics in studying antioxidant compounds	26
2.7.5	Comparison of two different stationary phases: method development to improve metabolome analysis for UHPLC-MS	28
2.7.6	Data preprocessing	29
2.7.7	Statistical analysis in metabolomics	30
2.8	Measurement of <i>in vitro</i> antioxidant activity	31
2.8.1	2,2-diphenyl-1-picrylhydrazyl (DPPH) assay	33
2.8.2	Nitric oxide (NO) inhibitory assay	34
2.8.3	Total phenolic content (TPC)	35
3	MATERIALS AND METHODS	36
3.1	Chemicals, reagents and standards	36
3.2	Microalga material	37
3.2.1	Microalga and growth conditions	37
3.2.2	Harvesting of microalga biomass	38
3.3	Extraction of the microalga	38
3.4	Biological and biocompositional assays on <i>Chaetoceros calcitrans</i>	39
3.4.1	Determination of the total phenolic content	39
3.4.2	DPPH free radical scavenging assay	39
3.4.3	Nitric oxide (NO) inhibitory assay	40
3.5	Nuclear magnetic resonance (NMR) analysis	41
3.5.1	Sample preparation for NMR analysis	41
3.5.2	NMR spectroscopy equipment settings	41
3.5.3	NMR preprocessing	42
3.5.4	Metabolite identification and relative quantification for NMR analyses	42
3.5.5	Chemometric analysis strategy	43
3.6	Ultra-high performance liquid chromatography-mass spectrometry (UHPLC-MS)	43
3.6.1	Equipment, column, mass spectrometric condition and software used for UHPLC-ESI-Orbitrap MS analysis	43
3.6.2	Sample preparation of UHPLC-ESI-Orbitrap MS analysis	45
3.6.3	Preparation of standard solution of the targeted metabolites	45
3.6.4	UHPLC-ESI-Orbitrap MS data processing and analysis	46
3.6.5	Metabolites identification from samples for UHPLC-ESI-Orbitrap MS analysis	46
3.6.6	Chemometric analysis strategy for UHPLC-ESI-Orbitrap MS	47

4	RESULTS AND DISCUSSION	48
4.1	Effect of different solvent extractions on extraction yield, total phenolic content (TPC), 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging and nitric oxide (NO) inhibitory activities of <i>Chaetoceros calcitrans</i>	48
4.2	Metabolite profiling of diatom microalga <i>Chaetoceros calcitrans</i> extracted with five different solvents and correlation with antioxidant and NO inhibitory activities using ¹ H NMR-based metabolomics	52
4.2.1	Assignments of metabolites by 1D nuclear magnetic resonance (NMR) and 2D NMR spectra in microalgal crude extracts	52
4.2.2	Classification of different solvent extracts by principal component analysis (PCA)	61
4.2.3	Relative quantification of metabolites identified from different solvent extractions	64
4.2.4	The correlation study between the metabolites and biological activities in <i>C. calcitrans</i> extracts	68
4.2.5	Metabolite network analysis in diatom <i>C. calcitrans</i>	69
4.3	Method development of an UHPLC-ESI-Orbitrap Mass Spectrometry for identification of metabolites in <i>Chaetoceros calcitrans</i>	73
4.3.1	Evaluation on ionisation of metabolites using BEH C ₁₈ and HSS T3 (C ₁₈)	74
4.3.2	Peak tailing and quality of separation	74
4.3.3	Effect of different high-collisional dissociation (HCD) energies on fragmentation patterns	77
4.4	Metabolite characterization and quantitative analysis of the microalgal extracts of <i>C. calcitrans</i> extract by UHPLC-ESI-Orbitrap MS	81
4.4.1	Identification of metabolites in <i>C. calcitrans</i> extract by UHPLC-ESI-Orbitrap MS	81
4.4.2	Chemometric analysis of MS data	99
4.4.3	Quantitative analysis of selected metabolites in microalgal diatom <i>C. calcitrans</i>	109
5	CONCLUSION AND RECOMMENDATION	114
5.1	Conclusion	114
5.2	Recommendation for future research	116
	REFERENCES	117
	BIODATA OF STUDENT	132
	LIST OF PUBLICATIONS	133

LIST OF TABLES

Table		Page
2.1	Bioactive metabolites derived from microalgae and its health benefits	13
2.2	Scientific classification of <i>Chaetoceros calcitrans</i>	21
2.3	Application of metabolomics in various science fields	25
3.1	Mobile phase flow gradient	44
3.2	Mass spectrometric settings for UHPLC-ESI-Orbitrap MS	44
4.1	Percentage of extraction yield (%) of the <i>C. calcitrans</i> extracted with acetone, CHCl ₃ , 70% ethanol, methanol and hexane	48
4.2	Identified metabolites and its ¹ H NMR (500 MHz, acetone-d ₆) assignment of <i>C. calcitrans</i> in five different solvent extractions	56
4.3	Relative quantification of compounds in the extracts of <i>C. calcitrans</i>	67
4.4	Metabolites identified from <i>C. calcitrans</i> by UHPLC-ESI-Orbitrap MS	85
4.5	Relative quantification of compounds in the extracts of <i>Chaetoceros calcitrans</i>	106
4.6	Analytical parameters and validation results for linearity, LOQ, LOD, accuracy and precision of UHPLC-ESI-Orbitrap MS method	111
4.7	The concentration of targeted metabolites in chloroform and acetone extracts of microalgal diatom <i>C. calcitrans</i>	112

LIST OF FIGURES

Figure		Page
2.1	Phylogenic relationship of marine algae based on the analysis of ribosomal RNS sequence	4
2.2	Potential products from microalgae biomass	6
2.3	<i>Chlorella</i> culture in pond at Yaeyama, Japan	7
2.4	Guacamole and pasta made from Spirulina biomass	8
2.5	Cosmeceutical products made by Euglena company in Japan which derived from microalgae	10
2.6	Chemical structures of fatty acids from microalgae	12
2.7	Chemical structure of tocopherol	14
2.8	Structure of some carotenoids commonly found in microalgae	15
2.9	Chemical structures of chlorophyll- <i>a</i> and chlorophyll- <i>c</i> 2	16
2.10	Chemical structure of cholesterol	17
2.11	<i>Chaetoceros calcitrans</i>	22
2.12	Solvent extraction method	23
2.13	Metabolomics pipeline	25
2.14	Framework for metabolite identification using <i>m/z</i> values and MS/MS	28
2.15	Chemical properties of (A) BEH C ₁₈ and (B) HSS T3 (C ₁₈) columns	29
2.16	The mechanism of DPPH radical scavenged by hydrogen donating atom	34
2.17	Antioxidant effects of nitric oxide	35
3.1	The overall outline of this research	36
4.1	Total phenolic content (A), the percentage of DPPH free radical scavenging activity (B), and NO inhibitory IC ₅₀ (C) of the diatom <i>C. calcitrans</i>	50

4.2	Representative 500 MHz ¹ H nuclear magnetic resonance (NMR) spectra of <i>C. calcitrans</i> in different solvents extraction	55
4.3	2D NMR ¹ H (<i>J</i> -resolved) spectrum of chloroform extract of <i>C. calcitrans</i>	59
4.4	2D NMR ¹ H- ¹³ C (HMBC) spectrum of the chloroform extract of <i>C. calcitrans</i> . (A) at 3.5 – 10 ppm region. Metabolite assignments	60
4.5	2D NMR ¹ H- ¹³ C (HMBC) spectrum of the chloroform extract of <i>C. calcitrans</i> (B) at 0.30- 3.00 ppm region. Metabolite assignments: 15, oleic acid; 16, linolenic acid; 17, α -linolenic acid; 22, fucoxanthin; 23, astaxanthin; 28, chlorophyll <i>c</i> ₁	62
4.6	Score plot of principal component analysis (PC1 versus PC2) in (A) and loading plot (B) obtained from five different solvent extractions of <i>C. calcitrans</i>	63
4.7	Relative quantification of the identified compounds (A) chlorophyll -c ₁ , chlorophyll-a, arachidic acid, α -linolenic acid, astaxanthin, canthaxanthin, lutein, sucrose, palmitic acid and fucoxanthin, (B) stearic acid, isoleucine, violaxanthin, zeaxanthin, cholesterol, leucine, glucose, proline, myo-inositol and glycine of different extraction solvents from <i>C. calcitrans</i> based on the mean peak area of ¹ H NMR signals	66
4.8	Partial least square (PLS) loading biplot of bioactivities represented by DPPH and NO inhibitory activities	68
4.9	Validation of the PLS model using permutation test (200 permutations) of NO (A) and DPPH (B) inhibitory activity	70
4.10	PLS derived relationship between observed vs predicted of NO (A) and DPPH (B) activity	71
4.11	Metabolic map of different biosynthetic pathways (amino acids, carbohydrates, fatty acids, cholesterol, photosynthetic pigments) for various functions in the diatom <i>C. calcitrans</i>	72
4.12	Representatives UHPLC-ESI-Orbitrap MS base peak and UV-vis chromatograms of CHCl ₃ extract of <i>C. calcitrans</i> analysed on two different columns	76
4.13	Extracted chromatograms of fucoxanthin, astaxanthin, lutein and zeaxanthin in the CHCl ₃ extract <i>C. calcitrans</i> on the A) HSS T3 (C ₁₈) and BEH C ₁₈ columns	78
4.14	Fucoxanthin identification from a standard sample	79

4.15	Astaxanthin identification from a standard sample	80
4.16	UHPLC-ESI-Orbitrap MS/ base peak chromatogram of the chloroform extract (A) Retention time 0-15 minutes (B) Retention time (15-30 minutes) obtain from microalga diatom <i>C. calcitrans</i>	84
4.17	(A) MS, MSMS data of fucoxanthin (C ₄₂ H ₅₈ O ₆ , MW=658.4228) (B) Fragmentation pathway of fucoxanthin	90
4.18	(A) MS, MSMS data of fucoxanthinol (C ₄₀ H ₅₆ O ₅ , MW=616.4122) (B) Fragmentation pathway of fucoxanthinol	91
4.19	(A) MS, MSMS data of Pheophytin <i>a</i> (C ₅₅ H ₇₄ N ₄ O ₅ , MW = 870.5654) (B) Fragmentation pathway of Pheophytin <i>a</i>	93
4.20	(A) MS, MSMS data of eicosapentaenoic acid (C ₂₀ H ₃₀ O ₂ , MW = 302.2240) (B) Fragmentation pathway of eicosapentaenoic acid	95
4.21	(A) MS, MSMS data of PG (18:3(9Z,12Z,15Z)/13:0) (C ₃₇ H ₆₇ O ₁₀ P, MW = 702.4466) (B) Fragmentation pathway of PG(18:3(9Z,12Z,15Z)/13:0)	96
4.22	(A) MS, MSMS data of crocetin dialdehyde (C ₂₀ H ₂₄ O ₂ , MW = 296.1771) (B) Fragmentation pathway of crocetin dialdehyde	99
4.23	(A) Score plot (B) loading plots of two-dimensional principal component analysis (2D-PCA) in the microalga diatom <i>C. calcitrans</i> in positive ion mode	100
4.24	Partial least square (PLS) of the UV scaling of the tentatively identified metabolites and bioactivities (NO and DPPH)	102
4.25	Validation of the PLS model using permutation test (100 permutations) of DPPH (A) and NO (B) inhibitory activities	103
4.26	PLS derived relationship between observed vs predicted of DPPH (A) and NO (B) activities. (Ac) Acetone, (CHCl ₃) Chloroform, (Hex) Hexane, (MeOH) Methanol and (70EtOH) 70% Ethanol	104
4.27	Variable importance projection (VIP) plot of the tentatively identified metabolites that influenced the separation in PLS	105
4.28	Relative quantification of the identified compounds (A) Docosahexaenoic acid, 15-HEPE, arachidonic acid, fucoxanthin, 5-HEPE, stearidonic acid, eicosapentaenoic acid, DG(20:3(8Z,11Z,14Z)/20:5(5Z,8Z,11Z,14Z,17Z)/0:0), α-linolenic acid (B) (E)- 3- Hexadecenoic acid, crocetin dialdehyde, PA(18:1(9Z)	

/12:0), PC(20:5(5Z,8Z,11Z,14Z,17Z)/15:0), linoleic acid, fucoxanthinol, N-palmitoyl proline, palmitic acid and lutein 108

4.29 Correlogram summarizing the correlation of significantly influenced metabolites based on PLS VIP (> 0.7) with the tested bioactivities of NO inhibitory and DPPH radical scavenging 109



LIST OF ABBREVIATIONS

a.u	Arbitrary unit
amu	Atomic mass unit
AA	Arachidonic acid
ABTS	2,2-azinobis(3-ethyl-benzothiazoline-6-sulfonic acid)
ACN	Acetonitrile
Ac	Acetone extract
ALA	α -linolenic acid
ANOVA	Analysis of variance
APCI	Atmospheric pressure chemical ionization
API	Atmospheric ionization
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
BPC	Base Peak Chromatogram
C40	40 carbon atoms
CCl ₄	Carbon tetrachloride
(CD ₃) ₂ CO	Acetone-d ₆ , Deuterated acetone
CHCl ₃	Chloroform extract
Chl <i>a</i>	Chlorophyll <i>a</i>
Chl <i>c</i>	Chlorophyll <i>c</i>
CO ₂	Carbon dioxide
CODA	Component Detection Algorithm
COX	Cyclooxygenase

CS	Calibration standard
CUPRAC	Cupric reducing antioxidant capacity
d	Doublet
DAD	Diode array detector
DA	Domoic acid
dd	Doublet of doublet
DPPH	2,2-diphenyl-1-picrylhydrazyl free radicals
DHA	Docosahexaenoic acid
DMEM	Dulbecco's Modified Eagle's Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DSS	Sodium 2,2-dimethylsilapentane sulphonate
EDTA	Ethylenediaminetetraacetic acid
EPA	Eicosapentaenoic acid
EPS	Exopolysaccharides
EtOH	Ethanol extract
ET	Electron transfer
ESI	Electrospray ionization
FA	Fatty acids
FBS	Fetal bovine serum
FC	Follin-Ciocalteu
FRAP	Ferric reducing antioxidant power
FTC	Ferric thiocyanate
GAE	Gallic acid equivalent

HCD	High-collisional dissociation
GABA	γ -Amino butyric acid
GLA	γ -Linolenic acid
H ₃ PO ₄	Phosphoric acid
HAT	Hydrogen atom transfer
HCA	Hierarchical cluster analysis
HCN	Hydrogen cyanide
Hex	Hexane extract
HESI	Heat electrospray ionization
HMBC	Heteronuclear Multiple Bond Correlation
HMDB	Human metabolome databases
HSQC	Heteronuclear single quantum correlation
HPLC	High performance liquid chromatography
HRAM	High resolution accurate mass
IFN- γ	Interferon gamma
iNOS	Inducible nitric oxide synthase
IL	Interleukins
<i>J</i>	Coupling Constant
<i>J</i> -res	<i>J</i> -resolved
kV	kiloVolt
LC	Liquid chromatography
LC/MS	Liquid chromatography/mass spectrometry
LOD	Limit of detection
LOQ	Limit of quantitation

LOX	Lipoxygenase
LOOH	Lipid hydroperoxides
LPS	lipopolysaccharides
m	Multiplet
<i>m/z</i>	Mass-to-charge ratio
MeOH	Methanol extract
MS	Mass spectrometry
MTT	3-(4,5-dimethylthiazol2-yl)-2,5-diphenyl tetrazolium bromide
MVDA	Multivariate data analysis
N.D	Not determined
Na ₂ CO ₃	Sodium carbonate
NCE	Normalized collision energy
NMR	Nuclear magnetic resonance
NO	Nitric oxide
ORAC	Oxygen radical absorption capacity
OPLS-DA	Orthogonal partial least square discrimination analysis
PBS	Phosphate buffered saline
PCA	Principle component analysis
PLS	Partial least squares
PLS-DA	Partial least squares–discriminant analysis
PNP	p-nitrophenol
PNPG	p-nitrophenyl- α -D-glucofuranose
ppm	Part per million
PGs	Prostaglandins

PUFAs	Polyunsaturated fatty acids
QTOF	Quadruple-time of flight
RNS	Reactive nitrogen species
Rs	Resolution
RSD	Relative standard deviation
RT	Retention times
ROS	Reactive oxygen species
S/N	Signal to noise ratio
s	Singlet
SD	Standard deviation
SIM	Single ion monitoring
SIMCA	Soft independent modelling in class analogues
t	Triplet
t_R	Retention time
TAG	Triacylglycerols
TBA	Thiobarbituric acid
TEAC	Trolox equivalent antioxidant capacity
TIC	Total ion chromatogram
TLC	Thin layer chromatography
TNF- α	Tumor necrosis factor alpha
TMS	Tetramethylsilane
TOF	Time of flight
TPC	Total phenolic content
TSP	Sodium 3-(trimethylsilyl) propionate-2,2,3,3-d4

Tukey's-HSD	Tukey's honest significant difference
TW	Time warping
UHPLC-MS	Ultra-high performance liquid chromatography mass spectrometry
UHPLC-MS/MS	Ultra-high performance liquid chromatography tandem mass spectrometry analysis
UPM	Universiti Putra Malaysia
UV	Ultraviolet
VIP	Variable importance in the projection
WHO	World Health Organization
δ	Chemical shift in ppm
1D	One-dimensional
^1H	Proton
2D	Two-dimensional
^{13}C	Carbon-13
O_2^-	Superoxide anion
$\bullet\text{OH}$	Hydroxyl
$\text{ROO}\bullet$	Peroxyl
$\text{RO}\bullet$	Alkaloyl radicals
H_2O_2	Hydrogen peroxide
$^1\text{O}_2$	singlet oxygen

CHAPTER 1

INTRODUCTION

1.1 Background

Oxygen free radicals known as reactive oxygen species (ROS) formed through oxygen poisoning and radiation injury lead to many deleterious effects (Devasagayam et al., 2004). It is increasingly reported that this kind of oxygen free radical plays a key role in approximately hundreds of oxidative stress disorders including cancer, inflammatory bowel disease, cardiovascular diseases, Alzheimer's disease, rheumatoid arthritis, stroke and septic shock (Halliwell, 1996; Wiseman and Halliwell, 1996). Therefore, removal of the harmful radicals exists are indispensable, via antioxidants defence system that capable of blocking the generation of ROS. Since then, most biomedical research has been circulating on free radical chemistry, oxidative pathology, and leading to translate the knowledge from the laboratory into new-drugs discovery.

In recent years, the limited land for growing terrestrial crops, worrying levels of global warming and enlarging numbers of the human population, resulting in the urgent need for sustainable biomass resources from marine ecology (Gaurav et al., 2017). Hence, over the past years have witnessed that marine organisms have received increasing attention from researchers in the various fields of industries to meet the demands. Marine organisms including marine algal species are considered as “barely tapped source” (Stengel and Connan, 2015). They have dominated many kinds of marine environment from oceans, seas and also in the coastal areas (Kim, 2015). Many therapeutic metabolites derived from marine algae have responsible to aid human being in curing diseases and reducing aches, simultaneously, improved human life quality (Kim, 2015). Furthermore, these valuable metabolites including natural pigments, phycobiliproteins, lipids, polyunsaturated fatty acids (PUFAs), polyphenols and polysaccharides also have other enormous potentials such as nutritive feed for mariculture animal species, as ingredients in cosmeceutical products, as lipid-based bioproducts and also as an alternative power sources (biofuels) (Gouveia et al., 2010).

Owing to the variety of their pharmacological properties including antiradical and anti-inflammatory, microalgae carotenoids have been considered as a source of therapeutic metabolites to oxidative stress besides their role in photosynthesis and protection against UV solar radiation (Gong and Bassi, 2016; El Gamal, 2010; Zuluaga et al., 2017). Antioxidant activity of microalgae is frequently measured by several assays including 2,2-azino-bis (3-ethyl-benzothiazoline-6-sulfonic acid) (ABTS) (Goiris et al., 2012; Foo et al., 2015; Foo et al., 2017), 2,2-diphenyl-1-picrylhydrazyl (DPPH) (Goiris et al., 2012; Foo et al., 2017) ferric reducing antioxidant power (FRAP) and thiobarbituric acid (TBA) (Pangestuti and Kim, 2011). Carotenoids not only function as scavengers, but they also have ability to modulate the macrophages function as secretory of a vast array of mediators and cytokines including nitric oxide (NO), prostaglandins (PGs), tumour

necrosis factor alpha (TNF- α), interleukins (IL), lipoxygenase (LOX) and cyclooxygenases (COX) (Pangestuti and Kim, 2011). Although microalgae-derived carotenoids have promising anti-inflammatory activities, little research has been performed on this activity and only a few studies were reported on microalgae (Pangestuti and Kim, 2011).

Chaetoceros calcitrans is a microscopic and fast growth microalga diatom which offers higher production of potent biological sources. It is known as de novo producers of antioxidants and immune stimulants metabolites such as carotenoids, lipids, polyunsaturated fatty acids (PUFAs) and vitamins (Salas-Leiva and Dupré, 2011; Foo et al., 2015). This species is frequently used as food source for feeding the maricultured animal species including bivalve molluscs (etc. mussels), echinoderms (e.g. copepods), crustaceans (e.g. penaeid shrimps) and also zooplankton (e.g. brine shrimp *Artemia*) (Becker, 2004). Although it was claimed as essential food sources for marine animal studies, its application in human consumption still remains elusive.

To develop effective medications for oxidative stress disorders, a better understanding of the metabolic changes caused by stress conditions through metabolomics tools predominantly based on nuclear magnetic resonance (NMR) and mass spectrometry (MS) may facilitate in finding of potential biomarkers for early detection of abnormalities associated with chronic oxidative diseases (Andrisic et al., 2018). Although MS sensitivity in detecting metabolites is much higher (femtomolar to attomolar) compared to NMR as an analytical tool, we believe that the weaknesses of NMR are the strengths of MS spectrometry (Veenstra, 2012). Since NMR and MS have easy sample preparation steps, and capable in detecting of primary and secondary metabolites, these approaches were often selected in previous studies of plant extracts for understanding the dynamical processes regarding interacting biomolecules involved in antioxidant activity. At present, the complete profile and metabolic network of this diatom *C. calcitrans* have not been fully characterized. A sound understanding of the *C. calcitrans* metabolome will further contribute to give a holistic overview of its system biology, lead to new applications and promote this diatom as attractive prolific producers of bioactive metabolites.

1.2 Problem statement

Chaetoceros calcitrans is being widely used to provide a direct source of nutrition for marine fish and shellfish aquaculture study especially as an alternative to conventional feed. Only a few bioactivities related to this species have been reported including antioxidant (Foo et al., 2015) and anti-cancer (Nigjeh et al., 2013). Even though the presence of carotenoids, chlorophylls, phenolic, amino acids, sterol, fatty acids, oxylipins and lipids had been reported in this diatom, comprehensive metabolic identification and correlation for the metabolites that could contribute to the claimed bioactivities have not much explored.

The inherent metabolic variation in the microalgae is associated by the post-harvest handling, including extraction process. Owing to great economic stakes for bioactive production from microalgae, researchers and scientific community have targeted to find efficient and economical ways to recover the metabolites. Besides, extraction is a pivotal step for assessing the deeply inaccessible metabolites from the complex matrix of a given system like microalgae. It was crucial to recovering all classes of metabolites at one time of extraction due to the substantial level of complexity of microalga metabolites. Therefore, extracting the metabolite in an efficient way should be investigated and optimized.

1.3 Scope and Objectives

The main goal of this study is to profile the chemical scaffolds of *C. calcitrans* extracts using NMR and UHPLC-MS metabolomics approaches that allow efficient and accurate identification of the wide spectrum of interesting compounds resulting from the effect of different solvent extractions. Consequently, correlation of the metabolite profile of *C. calcitrans* extracts obtained from both high power metabolomic tools with the antioxidant and anti-inflammatory activities including DPPH free radical scavenging and nitric oxide (NO) inhibitory activities was performed. Also, the correlation will provide information on the effects of type of solvents as a parameter to improve the extraction efficiency of bioactive metabolites from *C. calcitrans*. In this thesis, the reports are presented and discussed in three parts. First part of the work aimed to screen the biological activities of *C. calcitrans* extracted using five different solvent polarities for antioxidant and NO inhibitory activities (Chapter 4, part 4.1). Subsequently, different crude extracts of *C. calcitrans* will be further characterized, quantified and then correlated their biological activities using NMR (Chapter 4, part 4.2) and method development for UHPLC-MS followed by the UHPLC-MS-based metabolomics approaches (Chapter 4, part 4.3 and part 4.4). Lastly, an attempt was made to quantify the compounds in the active extract by using UHPLC-MS.

The specific objectives for this study are:

1. To screen the five different solvent extracts including 70% ethanol, methanol, chloroform, acetone and hexane of *C. calcitrans* for antioxidant, nitric oxide (NO) inhibitory activity and total phenolic content (TPC).
2. To determine the effect of different solvent extractions on the metabolome of *C. calcitrans* and the correlation with antioxidant, nitric oxide (NO) inhibitory activity and total phenolic content (TPC) using NMR based metabolomics.
3. To characterize the different solvent extractions of *C. calcitrans* and correlation with antioxidant and nitric oxide (NO) inhibitory activity using UHPLC-MS-based metabolomics.
4. To quantify the metabolites present in the *C. calcitrans* extract using NMR and UHPLC-MS.

REFERENCES

- Abd El Baky, H. H., & El-Baroty, G. S. (2013). Healthy benefit of microalgal bioactive substances. *Journal of Aquatic Science*, 1(1), 11-23.
- Adebayo, S. A., Dzoyem, J. P., Shai, L. J., & Eloff, J. N. (2015). The anti-inflammatory and antioxidant activity of 25 plant species used traditionally to treat pain in southern African. *BMC Complementary and Alternative medicine*, 15(1), 159.
- Agnolet, S., Jaroszewski, J. W., Verpoorte, R., & Staerk, D. (2010). ¹H NMR-based metabolomics combined with HPLC-PDA-MS-SPE-NMR for investigation of standardized *Ginkgo biloba* preparations. *Metabolomics*, 6(2), 292-302.
- Aktan, F. (2004). iNOS-mediated nitric oxide production and its regulation. *Life sciences*, 75(6), 639-653.
- Algae Industry Magazine. (2011). Development of a Spirulina Industry-Production Retrieved 2018, May 17, from <http://www.algencal.i/microalgae/?lang>
- Algencal Bioenergy. (2010). Microalage. Retrieved from <http://www.algencal.i/microalgae/?lang>
- Alonso, D. L., Belarbi, E. H., Rodríguez-Ruiz, J., Segura, C. I., Giménez, A. (1998) Acyl lipids of three microalgae. *Photochemistry*. 47, 1473-1481.
- Andrisic, L., Dudzik, D., Barbas, C., Milkovic, L., Grune, T., & Zarkovic, N. (2018). Short overview on metabolomics approach to study pathophysiology of oxidative stress in cancer. *Redox biology*, 14, 47-58.
- Ansari, F. A., Shriwastav, A., Gupta, S. K., Rawat, I., Guldhe, A., & Bux, F. (2015). Lipid extracted algae as a source for protein and reduced sugar: A step closer to the biorefinery. *Bioresource technology*, 179, 559-564.
- Antolovich, M., Prenzler, P. D., Patsalides, E., McDonald, S., & Robards, K. (2002). Methods for testing antioxidant activity. *Analyst*, 127(1), 183-198.
- Aruoma, O. I. (1998). Free radicals, oxidative stress, and antioxidants in human health and disease. *Journal of the American oil chemist's society*, 75(2), 199-212.
- Barofsky, A., & Pohnert, G. (2007) Biosynthesis of polyunsaturated short chain aldehydes in the diatom *Thalassiosira rotula*. *Organic Letter*, 1017-1020.
- Bataglion, G. A., da Silva, F. M., Eberlin, M. N., & Koolen, H. H. (2015). Determination of the phenolic composition from Brazilian tropical fruits by UHPLC-MS/MS. *Food Chemistry*, 180, 280-287.

- Becker, W. (2004). Microalgae in human and animal nutrition. *Handbook of microalgal culture: biotechnology and applied phycology* (2nd ed.). Blackwell: Oxford.
- Ben-Amotz, A., & Avron, M. (1992). *Dunaliella*: physiology, biochemistry, and biotechnology. Retrieved from: <https://books.google.com.my/books> = Dunaliella:+physiology,+biochemistry,+and+biotechnology
- Benzie, I. F., & Strain, J. J. (1999). Ferric reducing/antioxidant power assay: Direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methods in enzymology*, 299, 15-27.
- Begum, H., Yusoff, F. M., Banerjee, S., Khatoon, H., & Shariff, M. (2016) Availability and utilization of pigments from microalgae. *Critical Reviews in Food Science and Nutrition*, 56, 2209-2222.
- Bhakta, S., Sahu, E., & Bastia, A. (2014) Microalgae: A potential source of bioactive metabolites. *Natural Products-Drugs Development*, 2, 1-39
- Bijttebier, S. K.; D'Hondt, E.; Hermans, N.; Apers, S.; & Voorspoels, S. (2013) Unravelling ionization and fragmentation pathways of carotenoids using orbitrap technology: a first step towards identification of unknowns. *Journal of Mass Spectrometry*, 48, 740-754
- Bijttebier, S., D'Hondt, E., Noten, B., Hermans, N., Apers, S., & Voorspoels, S. (2014). Ultra high performance liquid chromatography versus high performance liquid chromatography: stationary phase selectivity for generic carotenoid screening. *Journal of Chromatography A*, 1332, 46-56.
- Bird, S. S., Marur, V. R., Sniatynski, M. J., Greenberg, H. K., & Kristal, B. S. (2011). Lipidomics profiling by high resolution LC-MS and HCD fragmentation: focus on characterization of mitochondrial cardiolipins and monolysocardiolipins. *Analytical chemistry*, 83(3), 940
- Boroujerdi, A.F.B., Lee, P.A., DiTullio, G.R., Janech, M.G., Vied, S.B., & Bearden, D.W (2012). Identification of isethionic acid and other small molecule metabolites of *Fragilariopsis cylindrus* with nuclear magnetic resonance. *Analytical and Bioanalytical Chemistry*. 404, 777-784
- Borowitzka, M. A. (1995). Microalgae as sources of pharmaceuticals and other biologically active compounds. *Journal of Applied Phycology*, 7(1), 3-15
- Boussiba, S., & Vonshak, A. (1991). Astaxanthin accumulation in the green alga *Haematococcus pluvialis*. *Plant and cell Physiology*, 32(7), 1077-1082.
- Brennan, L., & Owende, P. (2010). Biofuels from microalgae—a review of technologies for production, processing, and extractions of biofuels and co-products. *Renewable and sustainable energy reviews*, 14(2), 557-577.

- Bromke, M.A (2013). Amino acid biosynthesis pathways in diatoms. *Metabolites*, 3, 294–311.
- Bryan, N. S., & Grisham, M. B. (2007). Methods to detect nitric oxide and its metabolites in biological samples. *Free Radical Biology and Medicine*, 43(5), 645-657.
- Bryk, J., Ochoa, J.B., Correia, M.I.T.D., Munera-Seeley, V., & Popovic, P.J. (2008). Effect of citrulline and glutamine on nitric oxide production in RAW 264.7 cells in an arginine-depleted environment. *Journal of Parenteral and Enteral Nutrition*, 32, 377–383
- Cadenas, E. (1997). Basic mechanisms of antioxidant activity. *Biofactors*, 6(4), 391-397.
- Chacón-Lee, T. L., & González-Mariño, G. E. (2010). Microalgae for “healthy” foods-possibilities and challenges. *Comprehensive reviews in food science and food safety*, 9(6), 655-675.
- Campenni, L., Nobre, B. P., Santos, C. A., Oliveira, A. C., Aires-Barros, M. R., Palavra, A. M. F., & Gouveia, L. (2013). Carotenoid and lipid production by the autotrophic microalga *Chlorella protothecoides* under nutritional, salinity, and luminosity stress conditions. *Applied microbiology and biotechnology*, 97(3), 1383-1393.
- Chauton, M.S., Størseth, T.R., & Johnsen, G. (2003) High-resolution magic angle spinning ^1H NMR analysis of whole cells of *Thalassiosira pseudonana* (Bacillariophyceae): broad range analysis of metabolic composition and nutritional value. *Journal of Applied Phycology*, 15, 533–542.
- Chauton, M.S., Størseth, T.R., & Krane, J. (2004) High-resolution magic angle spinning NMR analysis of whole cells of *Chaetoceros muelleri* (Bacillariophyceae) and comparison with ^{13}C -NMR and distortion less enhancement by polarization transfer ^{13}C -NMR analysis of lipophilic extracts. *Journal Phycology*, 40, 611-618.
- Chauveau-Duriot, B., Doreau, M., Noziere, P., & Graulet, B. (2010). Simultaneous quantification of carotenoids, retinol, and tocopherols in forages, bovine plasma, and milk: validation of a novel UPLC method. *Analytical and bioanalytical chemistry*, 397(2), 777-790.
- Chu, W. L. (2012). Biotechnological applications of microalgae. *International eJournal of Science Medicine and Education*, 6(1), S24-S37.
- Christin, C., Hoefsloot, H. C., Smilde, A. K., Suits, F., Bischoff, R., & Horvatovich, P. L. (2010). Time alignment algorithms based on selected mass traces for complex LC-MS data. *Journal of proteome research*, 9(3), 1483-1495.

- Cohen, Z., Vonshak, A., & Richmond, A. (1988). Effect of environmental conditions on fatty acid composition of the red alga *Porphyridium cruentum*: correlation to growth rate. *Journal of Phycology*, 24(3), 328-332.
- da Costa, E., Silva, J., Mendonça, S. H., Abreu, M. H., & Domingues, M. R. (2016). Lipidomic approaches towards deciphering glycolipids from microalgae as a reservoir of bioactive lipids. *Marine Drugs*, 14(5), 101.
- Danielewicz, M. A., Anderson, L. A., & Franz, A. K. (2011) Triacylglycerol profiling of marine microalgae by mass spectrometry. *Journal of Lipid Research*, D018408, 1-26.
- Dawes, C. J. (1998). *Marine botany*. New York:John Wiley and Sons, Inc.
- de Jesus Raposo, M. F., de Morais, R. M. S. C., & de Morais, A. M. M. B. (2013). Health applications of bioactive compounds from marine microalgae. *Life Sciences*, 93(15), 479-486
- de Morais, M. G., Vaz, B. D. S., de Morais, E. G., & Costa, J. A. V. (2015). Biologically active metabolites synthesized by microalgae. *Biomed Research International*, 4, 835761.
- Desbois, A. P., Mearns-Spragg, A., & Smith, V. J. (2009). A fatty acid from the diatom *Phaeodactylum tricorutum* is antibacterial against diverse bacteria including multi-resistant *Staphylococcus aureus* (MRSA). *Marine Biotechnology*, 11(1), 45-52.
- Devaraj, S., Jialal, I., & Vega-López, S. (2004). Plant sterol-fortified orange juice effectively lowers cholesterol levels in mildly hypercholesterolemic healthy individuals. *Arteriosclerosis, thrombosis, and vascular biology*, 24(3), e25-e28.
- Devasagayam, T. P. A., Tilak, J. C., Bloor, K. K., Sane, K. S., Ghaskadbi, S. S., & Lele, R. D. (2004). Free radicals and antioxidants in human health: current status and future prospects. *Journal of Association of Physicians India*, 52, 794–803.
- Dolan, J. W. (2002). Peak tailing and resolution. *LC GC North America*, 20(5), 430-437.
- Dominguez, H. (Ed.). (2013). *Functional ingredients from algae for foods and nutraceuticals*. Elsevier.
- Dona, A. C., Kyriakides, M., Scott, F., Shephard, E. A., Varshavi, D., Veselkov, K., & Everett, J. R. (2016). A guide to the identification of metabolites in NMR-based metabolomics/metabolomics experiments. *Computational and structural biotechnology journal*, 14, 135-153.
- Englert, G., Bjørnland, T., & Liaaen-Jensen, S. (1990) 1D and 2D NMR study of some allenic carotenoids of the fucoxanthin series. *Magnetic Resonance in Chemistry*, 28, 519-28.

- El Gamal, A. A. (2010). Biological importance of marine algae. *Saudi Pharmaceutical Journal*, 18(1), 1-25.
- Foo, S. C., Yusoff, F. M., Ismail, M., Basri, M., Khong, N. M. H., Chan, K. W., & Yau, S. K. (2015). Efficient solvent extraction of antioxidant-rich extract from a tropical diatom, *Chaetoceros calcitrans* (Paulsen) Takano 1968. *Asian Pacific Journal of Tropical Biomedicine*, 5(10), 834-840.
- Foo, S. C., Yusoff, F. M., Ismail, M., Basri, M., Yau, S. K., Khong, N. M., Chan, K. W., & Ebrahimi, M. (2017). Antioxidant capacities of fucoxanthin-producing algae as influenced by their carotenoid and phenolic contents. *Journal of Biotechnology*, 241, 175-183.
- Fu, W., Magnúsdóttir, M., Brynjólfson, S., Palsson, B. Ø., & Paglia, G. (2012) UPLC-UV-MS^E analysis for quantification and identification of major carotenoid and chlorophyll species in algae. *Analytical and Bioanalytical in Chemistry*, 404, 3145-3154.
- Galindo-Prieto B, Eriksson L, Trygg J (2014) Variable influence on projection (VIP) for orthogonal projections to latent structures (OPLS). *Journal of Chemometrics*, 28:623-632
- García, J. L., Vicente, M., & Galan, B. (2017). Microalgae, old sustainable food and fashion nutraceuticals. *Microbial biotechnology*, 10(5), 1017-1024.
- Gaurav, N., Sivasankari, S., Kiran, G. S., Ninawe, A., & Selvin, J. (2017). Utilization of bioresources for sustainable biofuels: A review. *Renewable and Sustainable Energy Reviews*, 73, 205-214.
- Gerschman, R., Gilbert, D., Nye, S. W., Dwyer, P., & Fenn, W. O. (2001). Oxygen poisoning and X-irradiation: a mechanism in common. 1954. *Nutrition (Burbank, Los Angeles County, Calif.)*, 17(2), 162.
- Ginsburg, S., Tiwari, P., Kurhanewicz, J., Madabhushi, A. (September, 2011) *Variable ranking with pca: Finding multiparametric MR imaging markers for prostate cancer diagnosis and grading*. Proceedings of the International Workshop on Prostate Cancer Imaging Held in Conjunction with MICCAI 2011, Toronto, Canada. Retrieved from: <https://link.springer.com/chapter/10>.
- Glauser, G., Veyrat, N., Rochat, B., Wolfender, J. L., & Turlings, T. C. (2013). Ultra-high pressure liquid chromatography–mass spectrometry for plant metabolomics: A systematic comparison of high-resolution quadrupole-time-of-flight and single stage Orbitrap mass spectrometers. *Journal of Chromatography A*, 1292, 151-159.
- Goiris, K., Muylaert, K., Fraeye, I., Foubert, I., De Brabanter, J., & De Cooman, L. (2012). Antioxidant potential of microalgae in relation to their phenolic and carotenoid content. *Journal of Applied Phycology*, 24(6), 1477-1486.

- Gong, M., & Bassi, A. (2016). Carotenoids from microalgae: A review of recent developments. *Biotechnology Advances*, 34(8), 1396-1412.
- Goodacre, R., Vaidyanathan, S., Dunn, W. B., Harrigan, G. G., & Kell, D. B. (2004). Metabolomics by numbers: acquiring and understanding global metabolite data. *Trends in Biotechnology*, 22(5), 245-252.
- Gouveia, L., Marques, A. E., Sousa, J. M., Moura, P., & Bandarra, N. M. (2010). Microalgae-source of natural bioactive molecules as functional ingredients. *Food Science Technology Bulletin Functional Foods*, 7(2), 21.
- Gowda, G. N., Zhang, S., Gu, H., Asiago, V., Shanaiah, N., & Raftery, D. (2008). Metabolomics-based methods for early disease diagnostics. *Expert review of molecular diagnostics*, 8(5), 617-633.
- Gromski, P. S., Muhamadali, H., Ellis, D. I., Xu, Y., Correa, E., Turner, M. L., & Goodacre, R. (2015). A tutorial review: Metabolomics and partial least squares-discriminant analysis—a marriage of convenience or a shotgun wedding. *Analytica chimica acta*, 879, 10-23.
- Guillard, R.R.L., & Ryther, J.H. (1962). Studies of marine planktonic diatoms. *I. Cyclotella nana Hustedt and Detonula confervacea* (Cleve). *Canadian Journal of Microbiology*, 8: 229-239.
- Guiry, M. D., & Guiry, G. M. (2012). *Chaetoceros Ehrenberg*, 1844, 198.
- Halliwell, B. (1996). Antioxidants in human health and disease. *Annual review of nutrition*, 16(1), 33-50.
- Hanhineva, K., Lankinen, M. A., Pedret, A., Schwab, U., Kolehmainen, M., Paananen, J., de Mello, V., Sola, R., Lehtonen, M., Poutanen, K., Mykkänen, H., & Uusitupa, M. (2014). Nontargeted metabolite profiling discriminates diet-specific biomarkers for consumption of whole grains, fatty fish, and bilberries in a randomized controlled trial—3. *The Journal of nutrition*, 145(1), 7-17.
- Hasle, G. R., Syvertsen, E. E., Steidinger, K. A., Tangen, K., & Tomas, C. R. (1996). *Identifying marine diatoms and dinoflagellates*. Retrieved from: <https://www.sciencedirect.com/book/9780126930153/identifying-marine-diatoms-and-dinoflagellates#book-description>
- Huleihel, M., Ishanu, V., Tal, J., & Arad, S. M. (2001). Antiviral effect of red microalgal polysaccharides on *Herpes simplex* and *Varicella zoster* viruses. *Journal of applied phycology*, 13(2), 127-134.
- Huseby, S., Degerlund, M., Zingone, A., & Hansen, E. (2012). Metabolic fingerprinting reveals differences between northern and southern strains of the cryptic diatom *Chaetoceros socialis*. *European journal of phycology*, 47(4), 480-489.

- Jensen, K. G., & Moestrup, Ø. (1998). The genus *Chaetoceros* (Bacillariophyceae) in inner Danish coastal waters. *Nordic Journal of Botany*, 18(1), 88-88.
- Kaiser, B. K.; Carleton, M.; Hickman, J. W.; Miller, C.; Lawson, D.; Budde, M.; Cross, F. (2013). Fatty aldehydes in cyanobacteria are a metabolically flexible precursor for a diversity of biofuel products. *PLoS One*, 8, e58307.
- Katz, J.J.; Brown, C.E. (1983) Nuclear magnetic resonance spectroscopy of chlorophylls and corrins. *Structure*, 5, 3-49.
- Karleskint, G., Turner, R., & Small, J. (2012). *Introduction to marine biology*. Retrieved from: https://books.google.com.my/books/about/Introduction_to_Marine_Biology.html?id=0JkKOFIj5pgC&redir_esc=y
- Khymenets, O., Vázquez-Fresno, R., Palau-Rodriguez, M., Llorach, R., Urpí-Sardà, M., Garcia-Aloy, M., Tulipani, S., Lupianez-Barbero, A., & Andres-Lacueva, C. (2016). Metabolomic approaches in the study of wine benefits in human health. In *Wine Safety, Consumer Preference, and Human Health* (pp. 293-317). Cham: Springer.
- Kim, H. K., Khan, S., Wilson, E. G., Kricun, S. D. P., Meissner, A., Goral, S., Deelder, A.M., Choi, Y.H., & Verpoorte, R. (2010). Metabolic classification of South American *Ilex* species by NMR-based metabolomics. *Phytochemistry*, 71(7), 773-784.
- Kim, S.K. (2015) *Handbook of marine microalgae: Biotechnology advances*: Waltham, MA, USA: Academic Press.
- Kuczynska, P., Jemiola-Rzeminska, M., & Strzalka, K. (2015). Photosynthetic pigments in diatoms. *Marine Drugs*, 13(9), 5847-5881.
- Kobayashi, M., Kakizono, T., & Nagai, S. (1991). Astaxanthin production by a green alga, *Haematococcus pluvialis* accompanied with morphological changes in acetate media. *Journal of Fermentation and Bioengineering*, 71(5), 335-339.
- Kopec, R. E., Cooperstone, J. L., Cichon, M. J., & Schwartz, S. J. (2012). Analysis methods of carotenoids. *Analysis of antioxidant-rich phytochemicals*. Hoboken: Chichester: Wiley-Blackwell, 105-149.
- Kordalewska, M., & Markuszewski, M. J. (2015). Metabolomics in cardiovascular diseases. *Journal of pharmaceutical and biomedical analysis*, 113, 121-136.
- Laing, I. (1991). *Cultivation of marine unicellular algae* (p. 31). Conwy: Ministry of Agriculture, Fisheries and Food.
- Lang, I., Hodac, L., Friedl, T., & Feussner, I. (2011) Fatty acid profiles and their distribution patterns in microalgae: a comprehensive analysis of more than 2000 strains from the SAG culture collection. *BMC Plant Biology*, 11, 124.

- Lê Cao, K. A., Rossouw, D., Robert-Granié, C., & Besse, P. (2008). A sparse PLS for variable selection when integrating omics data. *Statistical applications in genetics and molecular biology*, 7(1).
- Lee, Y. K. (1997). Commercial production of microalgae in the Asia-Pacific rim. *Journal of Applied Phycology*, 9(5), 403-411.
- Lee, M. Y., Kim, H. Y., Lee, S., Kim, J. G., Suh, J. W., & Lee, C. H. (2015). Metabolomics-based chemotaxonomic classification of *Streptomyces spp.* and its correlation with antibacterial activity. *J Microbiol Biotechnol*, 25(8), 1265-74.
- Lee, S., Oh, D. G., Lee, S., Kim, G. R., Lee, J. S., Son, Y. K., Bae, C.H., Yeo, J., & Lee, C. H. (2015). Chemotaxonomic metabolite profiling of 62 indigenous plant species and its correlation with bioactivities. *Molecules*, 20(11), 19719-19734.
- Lee, S. Y., Abas, F., Khatib, A., Ismail, I. S., Shaari, K., & Zawawi, N. (2016). Metabolite profiling of *Neptunia oleracea* and correlation with antioxidant and α -glucosidase inhibitory activities using ^1H NMR-based metabolomics. *Phytochemistry Letters*, 16, 23-33.
- Leopoldini, M., Marino, T., Russo, N., & Toscano, M. (2004). Antioxidant properties of phenolic compounds: H-atom versus electron transfer mechanism. *The Journal of Physical Chemistry A*, 108(22), 4916-4922.
- Li, H. B., Cheng, K. W., Wong, C. C., Fan, K. W., Chen, F., & Jiang, Y. (2007). Evaluation of antioxidant capacity and total phenolic content of different fractions of selected microalgae. *Food chemistry*, 102(3), 771-776.
- Li, J., Liu, Y., Cheng, J. J., Mos, M., & Daroch, M. (2015). Biological potential of microalgae in China for biorefinery-based production of biofuels and high value compounds. *New biotechnology*, 32(6), 588-596.
- Liu, B., Vieler, A., Li, C., Jones, A. D., & Benning, C.(2013). Triacylglycerol profiling of microalgae *Chlamydomonas reinhardtii* and *Nannochloropsis oceanica*. *Bioresour Technol*, 146, 310-316.
- Liu, X., Ser Z., & Locasale J.W. (2014). Development and quantitative evaluation of a high-resolution metabolomics technology. *Analytical Chemistry*, 86(4), 2175-2184
- Liu, Z., Zeng, Y., & Hou, P. (2018). Metabolomic evaluation of *Euphorbia pekinensis* induced nephrotoxicity in rats. *Pharmaceutical biology*, 56(1), 145-153.
- Liu, H., Tayyari, F., Khoo, C., & Gu, L. (2015). A ^1H NMR-based approach to investigate metabolomic differences in the plasma and urine of young women after cranberry juice or apple juice consumption. *Journal of Functional Foods*, 14, 76-86.

- Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy reviews*, 4(8), 118.
- Loewus, F.A., & Loewus, M.W. (1983). Myo-Inositol: its biosynthesis and metabolism. *Annual Review Plant Physiology*, 34, 137–161
- Lu, S., Wang, J., Ma, Q., Yang, J., Li, X., & Yuan, Y. J. (2013) Phospholipid metabolism in an industry microalga *Chlorella sorokiniana*: the impact of inoculum sizes. *PloS one*, 8(8), e70827.
- Maadane, A., Merghoub, N., Ainane, T., El Arroussi, H., Benhima, R., Amzazi, S., Bakri, Y., & Wahby, I. (2015). Antioxidant activity of some Moroccan marine microalgae: Pufa profiles, carotenoids and phenolic content. *Journal of Biotechnology*, 215, 13–19.
- Mansour, M. P., Frampton, D. M., Nichols, P. D., Volkman, J. K., & Blackburn, S. I. (2005). Lipid and fatty acid yield of nine stationary-phase microalgae: applications and unusual C24–C28 polyunsaturated fatty acids. *Journal of Applied Phycology*, 17, 287-300.
- Martins, D., Custódio, L., Barreira, L., Pereira, H., Ben-Hamadou, R., Varela, J., & Abu-Salah, K. (2013). Alternative sources of n-3 long-chain polyunsaturated fatty acids in marine microalgae. *Marine drugs*, 11(7), 2259-2281.
- Mathew, B., Sankaranarayanan, R., Nair, P. P., Varghese, C., Somanathan, T., Amma, B. P., Amma, N.S., & Nair, M. K. (1995). Evaluation of chemoprevention of oral cancer with *Spirulina fusiformis*. *Taylor & Francis*, 197-202.
- Mehta, Mendes, A., da Silva, T. L., & Reis, A. (2007). DHA concentration and purification from the marine heterotrophic microalga *Cryptocodinium cohnii* CCMP 316 by winterization and urea complexation. *Food technology and biotechnology*, 45(1), 38.
- Michalak, I., & Chojnacka, K. (2015). Algae as production systems of bioactive compounds. *Engineering in Life Sciences*, 15(2), 160-176.
- Mimouni, V., Ulmann, L., Pasquet, V., Mathieu, M., Picot, L., Bougaran, G., Cadoret, J.P., Morant-Manceau, A., & Schoefs, B. (2012). The potential of microalgae for the production of bioactive molecules of pharmaceutical interest. *Current pharmaceutical biotechnology*, 13(15), 2733-2750.
- Moco, S., Vervoort, J., Bino, R. J., De Vos, R. C., & Bino, R. (2007). Metabolomics technologies and metabolite identification. *Trends in Analytical Chemistry*, 26(9), 855-866.
- Morimoto, T., Nagatsu, A., Murakami, N., Sakakibara, J., Tokuda, H., Nishino, H., & Iwashima, A. (1995). Anti-tumour-promoting glyceroglycolipids from the green alga, *Chlorella vulgaris*. *Phytochemistry*, 40(5), 1433-1437.

- Mourelle, M., Gómez, C., & Legido, J. (2017). The potential use of marine microalgae and cyanobacteria in cosmetics and thalassotherapy. *Cosmetics*, 4(4), 46.
- Natrah, F. M. I., Yusoff, F. M., Shariff, M., Abas, F., & Mariana, N. S. (2007). Screening of Malaysian indigenous microalgae for antioxidant properties and nutritional value. *Journal of Applied Phycology*, 19(6), 711-718.
- New, L. S., & Chan, E. C. (2008). Evaluation of BEH C₁₈, BEH HILIC, and HSS T3 (C₁₈) column chemistries for the UPLC-MS-MS analysis of glutathione, glutathione disulfide, and ophthalmic acid in mouse liver and human plasma. *Journal of chromatographic science*, 46(3), 209-214.
- Nicoletti, M. (2016). Microalgae nutraceuticals. *Foods*, 5(3), 54.
- Nigjeh, E.S., Yusoff, F. M., Alitheen, M., Banu, N., Rasoli, M., Keong, Y. S., & Omar, A. R. B. (2013). Cytotoxic effect of ethanol extract of microalga, *Chaetoceros calcitrans*, and its mechanisms in inducing apoptosis in human breast cancer cell line. *BioMed Research International*, 783690, 1-8.
- Nimse, S. B., & Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *Rsc Advances*, 5(35), 27986-28006.
- Ohta, S., Chang, T., Ikegami, N., Kondo, M., & Miyata, H. (1993). Antibiotic substance produced by a newly isolated marine microalga, *Chlorococcum* HS-101. *Bulletin of environmental contamination and toxicology*, 50(2), 171-178.
- Orset, S., & Young, A. J. (1999). Low-temperature-induced synthesis of α -carotene in the microalga *Dunaliella salina* (Chlorophyta). *Journal of Phycology*, 35(3), 520-527.
- Pacini, T., Fu, W., Gudmundsson, S., Chiaravalle, A. E., Brynjolfson, S., Palsson, B. O., Astarita, G., & Paglia, G. (2015). Multidimensional analytical approach based on UHPLC-UV-ion mobility-MS for the screening of natural pigments. *Analytical chemistry*, 87(5), 2593-2599.
- Pangestuti, R., & Kim, S. K. (2011). Biological activities and health benefit effects of natural pigments derived from marine algae. *Journal of functional foods*, 3(4), 255-266.
- Patil, V., Källqvist, T., Olsen, E., Vogt, G., & Gislerød, H. R. (2007). Fatty acid composition of 12 microalgae for possible use in aquaculture feed. *Aquaculture International*, 15(1), 1-9.
- Parul, R., Kundu, S. K., & Saha, P. (2013). In vitro nitric oxide scavenging activity of methanol extracts of three Bangladeshi medicinal plants. *The pharma innovation*, 1(12, Part A), 83.

- Peinado, N. K., Abdala Díaz, R. T., Figueroa, F. L., & Helbling, E. W. (2004). Ammonium and UV radiation stimulate the accumulation of mycosporin-like amino acids in *Porphyra columbina* (Rhodophyta) from Patagonia, Argentina. *Journal of Phycology*, 40(2), 248-259.
- Peatix. (2017). A Healthier Earth & Mankind with Euglena, Japan's Microalgae Biotechnology. Retrieved 2018, May 17, from <https://wasabi-discovery-euglena2.peatix.com/>
- Peng, J., Yuan, J. P., & Wang, J. H. (2012). Effect of diets supplemented with different sources of astaxanthin on the gonad of the sea urchin *Anthocidaris crassispinata*. *Nutrients*, 4(8), 922-934.
- Planet Organic. (2017). Profusion Organic Sprouted Spelt *Spirulina Tagliatelle*. Retrieved from <https://www.planetorganic.com/profusion-organic-sprouted-spelt-spirulina-tagliatelle-250g/23269/>
- Priyadarshani, I., & Rath, B. (2012). Commercial and industrial applications of microalgae—A review. *Journal Algal Biomass Utilization*, 3(4), 89-100.
- Pujos-Guillot, E., Hubert, J., Martin, J. F., Lyan, B., Quintana, M., Claude, S., Chabanas, B., Rothwell, J.A., Bennetau-Pelissero, C., Scalbert, A., Comte, B., Hercberg, S., Morand, C., Galan, P., & Manach, C. (2013). Mass spectrometry-based metabolomics for the discovery of biomarkers of fruit and vegetable intake: citrus fruit as a case study. *Journal of proteome research*, 12(4), 1645-1659.
- Pulz, O., & Gross, W. (2004). Valuable products from biotechnology of microalgae. *Applied microbiology and biotechnology*, 65(6), 635-648.
- Rajkumar, R., & Takriff, M. S. (2016). Prospects of algae and their environmental applications in Malaysia: a case study. *J Bioremed Biodeg*, 7, 321.
- Rivera, S. M., & Canela-Garayoa, R. (2012). Analytical tools for the analysis of carotenoids in diverse materials. *Journal of Chromatography A*, 1224, 1-10.
- Roberts, G.C. & Jardetzky, O. (1970). Nuclear magnetic resonance spectroscopy of amino acids, peptides, and proteins. *Advances in Protein Chemistry*, 24, 447-545.
- Román, R. B., Alvarez-Pez, J. M., Fernández, F. A., & Grima, E. M. (2002). Recovery of pure B-phycoerythrin from the microalga *Porphyridium cruentum*. *Journal of Biotechnology*, 93(1), 73-85.
- Romano, G., Costantini, M., Sansone, C., Lauritano, C., Ruocco, N., & Ianora, A. (2017). Marine microorganisms as a promising and sustainable source of bioactive molecules. *Marine environmental research*, 128, 58-69.

- Romay, C. H., Armesto, J., Ramirez, D., Gonzalez, R., Ledon, N., & Garcia, I. (1998). Antioxidant and anti-inflammatory properties of C-phycoerythrin from blue-green algae. *Inflammation Research*, 47(1), 36-41.
- Round, F. E., Crawford, R. M., & Mann, D. G. (1990). *Diatoms: biology and morphology of the genera*. Cambridge: Cambridge University Press.
- Rothwell, J. A., Madrid-Gambin, F., Garcia-Aloy, M., Andres-Lacueva, C., Logue, C., Gallagher, A. M., Mack, C., Kulling, S.E., Gao, Q., Praticò, G., Dragsted, L.O., & Scalbert, A. (2018). Biomarkers of intake for coffee, tea, and sweetened beverages. *Genes & nutrition*, 13(1), 15.
- Rubingh, C. M., Bijlsma, S., Derks, E. P., Bobeldijk, I., Verheij, E. R., Kochhar, S., & Smilde, A. K. (2006). Assessing the performance of statistical validation tools for megavariable metabolomics data. *Metabolomics*, 2(2), 53-61.
- Ryckebosch, E., Muylaert, K., & Foubert, I. (2012). Optimization of an analytical procedure for extraction of lipids from microalgae. *Journal of the American Oil Chemists' Society*, 89(2), 189-198.
- Roberts, G.C., & Jardetzky, O. (1970) Nuclear magnetic resonance spectroscopy of amino acids, peptides, and proteins. *Advances in Protein Chemistry*, 24, 447-545.
- Safar, F. (1975) Algoterapie. *Prakt. Lek (Praha)* 55, 641-648
- Safar, H., Van Wagenen, J., Møller, P., & Jacobsen, C. (2015). Carotenoids, phenolic compounds and tocopherols contribute to the antioxidative properties of some microalgae species grown on industrial wastewater. *Marine drugs*, 13(12), 7339-7356.
- Sambamurty, A. V. S. S. (2005). *A textbook of algae*. New Delhi: K. International Pvt. Ltd.
- Samarakoon, K., & Jeon, Y. J. (2012). Bio-functionalities of proteins derived from marine algae—A review. *Food Research International*, 48(2), 948-960.
- Salas-Leiva, J. S., & Dupré, E. (2011). Cryopreservation of the microalgae *Chaetoceros calcitrans* (Paulsen): Analysis of the effect of DMSO temperature and light regime during different equilibrium periods. *Latin american journal of aquatic research*, 39(2), 271-279.
- Sas, K. M., Karnovsky, A., Michailidis, G., & Pennathur, S. (2015). Metabolomics and diabetes: analytical and computational approaches. *Diabetes*, 64(3), 718-732.
- Scott, G. (1988). Antioxidants. *Bulletin of the Chemical Society of Japan*, 61(1), 165-170.

- Servel, M. O., Claire, C., Derrien, A., Coiffard, L., & De Roeck-Holtzhauer, Y. (1994). Fatty acid composition of some marine microalgae. *Phytochemistry*, 36(3), 691-693.
- Shahidi, F. (1997). *Natural antioxidants: chemistry, health effects, and applications*. The American Oil Chemists Society. Champaign, Illinois: AOC Press.
- Shamsudin, L. (1992). Lipid and fatty acid composition of microalgae used in Malaysian aquaculture as live food for the early stage of penaeid larvae. *Journal of Applied Phycology*, 4(4), 371-378.
- Shin, S. Y., Bajpai, V. K., Kim, H. R., & Kang, S. C. (2007). Antibacterial activity of bioconverted eicosapentaenoic (EPA) and docosahexaenoic acid (DHA) against foodborne pathogenic bacteria. *International journal of food microbiology*, 113(2), 233-236.
- Shimizu, Y., Gupta, S., Masuda, K., Maranda, L., Walker, C. K., & Wang, R. (1989). Dinoflagellate and other microalgal toxins: chemistry and biochemistry. *Pure and Applied Chemistry*, 61(3), 513-516.
- Singleton, V. L., Orthofer, R., & Lamuela-Raventós, R. M. (1999). Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. In *Methods in enzymology* (Vol. 299, pp. 152-178). Cambridge, Massachusetts: Academic press.
- Simonsen, R. (1974) The diatom plankton of the Indian Ocean Expedition of R/V "Meteor" 1964- 1965. In *Meteor Forschungsergebnisse* (Vol. 19, pp 1-107). Gebruder Borntraeger, Berlin.
- Sims, P. A., & Witkowski, J. (2012). Generic limits within the Eupodiscaceae: I. Observations on three unusual species of *Cerataulus*, with reference to the type species, *C. turgidus*. *Diatom research*, 27(4), 223-236.
- Şirin, S., Clavero, E., & Salvadó, J. (2015). Efficient harvesting of *Chaetoceros calcitrans* for biodiesel production. *Environmental technology*, 36(15), 1902-1912.
- Sousa, I., Gouveia, L., Batista, A. P., Raymundo, A., & Bandarra, N. M. (2008). Microalgae in novel food products. *Food chemistry research developments*, 75-112.
- Spolaore, P., Joannis-Cassan, C., Duran, E., & Isambert, A. (2006). Commercial applications of microalgae. *Journal of bioscience and bioengineering*, 101(2), 87-96.
- Stengel, D. B., & Connan, S. (2015). *Natural Products from Marine Algae: Methods and Protocols* (pp. 1-37). New York, USA: Springer.

- Stonik, V., & Stonik, I. (2015). Low-molecular-weight metabolites from diatoms: structures, biological roles and biosynthesis. *Marine drugs*, 13(6), 3672-3709.
- Sui, X., Niu, X., Shi, M., Pei, G., Li, J., Chen, L., Wang, J., & Zhang, W. (2014). Metabolomic analysis reveals mechanism of antioxidant butylated hydroxyanisole on lipid accumulation in *Cryptocodinium cohnii*. *Journal of agricultural and food chemistry*, 62(51), 12477-12484
- Tokushima, H., Inoue-Kashino, N., Nakazato, Y., Masuda, A., Ifuku, K., & Kashino, Y. (2016). Advantageous characteristics of the diatom *Chaetoceros gracilis* as a sustainable biofuel producer. *Biotechnology for biofuels*, 9(1), 235.
- Tzoulaki, I., Ebbels, T. M., Valdes, A., Elliott, P., & Ioannidis, J. P. (2014). Design and analysis of metabolomics studies in epidemiologic research: a primer on-omic technologies. *American journal of epidemiology*, 180(2), 129-139.
- van Meerloo, J., Kaspers, G. J., & Cloos, J. (2011). Cell sensitivity assays: the MTT assay. In *Cancer cell culture*, (Vol 731: pp. 237-245). Amsterdam, The Netherlands: Humana Press.
- Veenstra, T. D. (2012). Metabolomics: the final frontier? *Genome Medicine*, 4(4),40.
- Volkman, J. K., Jeffrey, S. W., Nichols, P. D., Rogers, G. I., & Garland, C. D. (1989). Fatty acid and lipid composition of 10 species of microalgae used in mariculture. *Journal of Experimental Marine Biology and Ecology*, 128(3), 219-240.
- Waters. (2018a). HSS (High Strength Silica) Technology. Retrieved from http://www.waters.com/waters/en_MY/HSS-%28High-Strength-Silica%29-Technology/nav.htm?cid=134618105&locale=en_MY
- Waters. (2018b). BEH (Ethylene Bridged Hybrid) Technology. Retrieved from http://www.waters.com/waters/en_MY/BEH-%28Ethylene-Bridged-Hybrid%29-Technology/nav.htm?cid=134618172&locale=en_MY
- Werner, D. (Ed.). (1977). *The biology of diatoms* (Vol. 13). University of California Press.
- Westerhuis, J. A., Hoefsloot, H. C., Smit, S., Vis, D. J., Smilde, A. K., van Velzen, E. J., van Duinhoven, J. P. M., & van Dorsten, F. A. (2008). Assessment of PLS-DA cross validation. *Metabolomics*, 4(1), 81-89.
- Wei, X., Shi, X., Kim, S., Zhang, L., Patrick, J. S., Binkley, J., McClain, C., & Zhang, X. (2012). Data preprocessing method for liquid chromatography–mass spectrometry based metabolomics. *Analytical chemistry*, 84(18), 7963-7971.

- Wink, D. A., Miranda, K. M., Espey, M. G., Pluta, R. M., Hewett, S. J., Colton, C., Vitek, M., Feelisch, M., & Grisham, M. B. (2001). Mechanisms of the antioxidant effects of nitric oxide. *Antioxidants and redox signaling*, 3(2), 203-213.
- Wiseman, H., & Halliwell, B. (1996). Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer. *Biochemical Journal*, 313, 17.
- Worley, B., & Powers, R. (2013). Multivariate analysis in metabolomics. *Current Metabolomics*, 1(1), 92-107.
- Xiao, J. F., Zhou, B., & Ransom, H. W. (2012). Metabolite identification and quantitation in LC-MS/MS-based metabolomics. *Trends in Analytical Chemistry*, 32, 1-14.
- Yao, L., Gerde, J. A., Lee, S. L., Wang, T. & Harrata, K. A. (2015). Microalgae lipid characterization. *Journal of Agriculture and Food Chemistry*, 63, 1773-1787.
- Young, I. S. (2001). Measurement of total antioxidant capacity. *Journal of Clinical Pathology*, 54(5), 339.
- Zuluaga M, Gueguen V, Pavon-Djavid G, & Letourneur D. (2017). Carotenoids from microalgae to block oxidative stress. *Bioimpacts*, 7(1): 1–3.
- Zuo, L., Zhou, L., Xu, T., Li, Z., Liu, L., Shi, Y., Kang, J., Gao, G., Du, S., Sun, Z., Zhang X & Zhang, X. (2018). Antiseptic activity of ethnomedicinal *Xuebijing* revealed by the metabolomics analysis using UHPLC-Q-Orbitrap HRMS. *Frontiers in pharmacology*, 9, 300.