



**UNIVERSITI PUTRA MALAYSIA**

***MOLECULAR DETECTION, SEROTYPING AND PHYLOGENETIC  
ANALYSIS OF *Haemophilus parasuis* IN PENINSULAR MALAYSIA***

**LEE CHEE YIEN**

**FPV 2019 4**



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By

**LEE CHEE YIEN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
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Master of Veterinary Science**

**April 2019**

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## DEDICATION

My family,  
Father and Mother  
who have constantly love me and support me

Brother, Sister  
who have kept me entertained all this time

Frankie Lau  
who never complain about me

My PG comrades  
Li Ping, Vynter, Michelle, Vivian, Daniel, Zi Herk, Rathiy, Peggy, Xiao Fen

Friends

And to all those who have made me who I am today.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Veterinary Science

**MOLECULAR DETECTION, SEROTYPING AND PHYLOGENETIC ANALYSIS OF *Haemophilus parasuis* IN PENINSULAR MALAYSIA**

By

**LEE CHEE YIEN**

**April 2019**

**Chairman : Associate Professor Ooi Peck Toung, DVM, PhD**  
**Faculty : Veterinary Medicine**

*Haemophilus parasuis* infections, what is known to cause Glässer's disease has been a nuisance for all farms. The lack on the information of *H. parasuis* serotype complicates vaccination as current available commercial vaccines are serotype-based and efficacy is serotype-dependent. Hence, this study carried the objectives to detect *H. parasuis* in clinical samples from weaner pigs in Peninsular Malaysia with conventional polymerase chain reaction (PCR) assay; to detect *H. parasuis* serotype 4, 5 or 12 and 13 using multiplex PCR as these serotypes were prevalent in other countries; to detect the outer membrane protein P2 (*OMPP2*) gene and further investigate the phylogenetic relatedness of Peninsular Malaysia's *H. parasuis* with reference strains. Throughout October 2016 to June 2018, samples were collected from nine farms from northern, seven farms from central and four farms from southern regions of Peninsular Malaysia using convenience sampling method. A total of 87 pigs was sampled and from there, 323 samples were collected. The number of pigs and samples collected in each farm varied depending case availability and gross post mortem manifestation. Samples collected included lung, brain, bodily fluids and fibrins of various sites. Samples were screened for *H. parasuis* using conventional PCR assay and positive samples were serotyped by multiplex PCR. Finally, representative samples of every states and serotypes were subjected to conventional PCR assay for *OMPP2* gene detection; followed by sequencing and phylogenetic analysis on sequences obtained. Using conventional PCR assay, 58/87 (66.7%) of the pigs sampled were positive of *H. parasuis*. The region with highest detection of *H. parasuis* in pigs was the southern region (4/5), followed by northern region (12/18) and central region (42/64). Pigs apparent of *H. parasuis* infection (Category A, B and C) had a higher detection percentage compared to those that were unapparent (Category D, E and F). Among all of the sample types collected, the highest detection percentage was obtained in the pleural fibrin samples (85.7%).

Generally, multiplex PCR assay on *H. parasuis*-positive samples revealed that serotype 4, 5 or 12 and 13 were detected with predominance order of NT, 5 or 12 and followed equally by serotype 4 and 13. Furthermore, comparison of the partial *H. parasuis* *OMPP2* nucleotide sequence of 15 Malaysian isolates and 16 reference strains showed Malaysia isolates were closely related to China strains. There was no direct correlation between serotype and *OMPP2* gene. Moreover, phylogenetic analysis of *OMPP2* gene revealed that majority of Peninsular Malaysia isolates was found to belong to *H. parasuis* genetic type III group. In conclusion, *H. parasuis* and the three important serotypes 4, 5 or 12 and 13 were detected using PCR technique. The phylogenetic relatedness based on the *OMPP2* gene and its correlation with serotype was identified. The *OMPP2* gene served to be an important virulence gene and future studies should focus on this gene. Equip with the knowledge of this study, more effective control measures could be designed and implemented to control *H. parasuis* infections in Peninsular Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Master Sains Veterinar

**PENGESANAN MOLEKULAR, PENCIRIAN SEROTIP DAN ANALISA  
FILOGENI *Haemophilus parasuis* DI SEMENANJUNG MALAYSIA**

Oleh

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Infeksi *Haemophilus parasuis*, atau dikenali sebagai penyebab penyakit Glässer's telah menjadi gangguan kepada semua ladang. Kekurangan maklumat mengenai serotip *H. parasuis* menyukarkan vaksinasi kerana vaksin komersial yang ada sekarang adalah berdasarkan serotip dan efikasinya adalah bergantung kepada serotip. Oleh itu, objektif kajian ini adalah untuk mengesan *H. parasuis* dalam sampel klinikal babi di Semenanjung Malaysia dengan kaedah PCR konvensional; mengesan serotip utama *H. parasuis* ia adalah serotip 4, 5 atau 12, dan 13 menggunakan kaedah PCR multipleks; mengesan gen protein selaput luar P2 (*OMPP2*) dan selanjutnya menyiasat Keterkaitan filogeni *H. parasuis* penciran Semenanjung Malaysia dengan penciran rujukan. Sepanjang Oktober 2016 sehingga Jun 2018, sampel adalah dikumpul daripada sembilan ladang di kawasan utara, tujuh ladang di kawasan tengah dan empat ladang di kawasan selatan Semenanjung Malaysia menggunakan kaedah persampelan mudah. Sejumlah 87 babi telah dikumpul dan daripada itu, 323 sampel telah dikumpul. Jumlah babi dan sampel yang dikumpul adalah berbeza untuk setiap ladang bergantung kepada kes dan manifestasi lesi. Sampel yang dikumpul meliputi paru-paru, otak, cecair badan dan fibrin dari pelbagai organ. Sampel adalah dikesan untuk *H. parasuis* menggunakan esei PCR konvensional dan sampel positif adalah ditip menggunakan PCR multipleks. Akhirnya, sampel yang mewakili setiap negeri dan serotip adalah dikesan untuk gen *OMPP2* menggunakan PCR konvensional; diikuti dengan penjujukan dan analisa filogeni ke atas jujukan yang diperolehi. Menggunakan PCR konvensional, 58/87 (66.7%) babi yang disampel adalah positif kepada *H. parasuis*. Kawasan yang mempunyai pengesanan *H. parasuis* yang tertinggi adalah kawasan selatan (4/5), diikuti dengan kawasan utara (12/18) dan kawasan tengah (42/64). Babi yang ketara dengan infeksi *H. parasuis* (Kategori A, B, C) mempunyai pengesanan yang

lebih tinggi berbanding babi yang tidak ketara (Kategori D, E, F). Antara semua jenis sampel yang dikumpul, peratus pengesanan yang tertinggi adalah diperoleh daripada sampel fibrin pleura (85.7%). Keseluruhannya, esei PCR multipleks ke atas sampel positif *H. parasuis* menunjukkan bahawa serotip 4, 5 atau 12 dan 13 adalah dikesan dengan turutan dominasi NT, 5 atau 12 diikuti dengan jumlah yang sama adalah serotip 4 dan 13. Tambahan pula, perbandingan 15 jujukan nukleotid separa gen *OMPP2* *H. parasuis* pencilan Malaysia dengan 16 jujukan rujukan menunjuk bahawa pencilan Malaysia adalah berkait rapat dengan strain China. Tiada korelasi terus antara serotip dengan gen *OMPP2*. Selain itu, analisis filogeni mendedahkan bahawa majoriti strain Malaysia tergolong dalam jenis genetik III. Kesimpulannya, *H. parasuis* dan tiga serotip penting iaitu serotip 4, 5 atau 12 dan 13 telah dikesan menggunakan teknik PCR. Kerterkaitan filogeni berdasarkan gen *OMPP2* dan kolerasi dengan serotip telah dikenal pasti. Gen *OMPP2* berfungsi sebagai gen virulen yang penting dan kajian seharusnya menumpukan kepada gen ini. Berdasarkan pengetahuan dari kajian ini, langkah-langkah kawalan efektif boleh direkabentuk dan dilaksanakan untuk mengawal jangkitan *H. parasuis* di Semenanjung Malaysia.



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I certify that a Thesis Examination Committee has met on 18 April 2019 to conduct the final examination of Lee Chee Yien on her thesis entitled "Molecular Detection, Serotyping And Phylogenetic Analysis of *Haemophilus parasuis* in Peninsular Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Veterinary Science.

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## LIST OF ABBREVIATIONS

%	Percent
>	More than
<	Less than
μl	Microliter
μM	Micromolar
°C	Degree Celsius
AFLP	Amplified fragments length polymorphism
AGD	Agar gel diffusion
AMR	Antimicrobial resistant
BLAST	Basic Local Alignment Search Tool
bp	Base pairs
C	Cytosine
CDS	Coding sequence
CFU	Colony forming unit
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
dNTP	Deoxyribonucleotide triphosphate
EMBL- EBI	European Molecular Biology Laboratory-European Bioinformatics Institute
ERIC-PCR	enterobacterial repetitive intergenic consensus-based PCR
etc	et cetera
g	Gram
G	guanine

HPS	<i>Haemophilus parasuis</i>
IACUC	Institutional Animal Care and Use Committee
IHA	indirect hemagglutination
mg	Milligram
MgCl <sub>2</sub>	Magnesium chloride
min	Minutes
ml	Milliliter
MLEE	Multi-locus enzyme electrophoresis
mm	Milimeter
mM	Millimolar
mPCR	Multiplex Polymerase Chain Reaction
mRNA	Messenger ribonucleic acid
NCBI	National Center for Biotechnology Information
ng/μL	Nanogram per microliter
ng/reaction	Nanogram per reaction
NJ	Neighbour-joining
NT	Non-typeable
NTC	No template control
OMP	Outer membrane protein
OMPP2	Outer membrane protein P2
OMPP5	Outer membrane protein P5
PBS	Phosphate buffer saline
PCR	Polymerase Chain Reaction
PCV2	Porcine circovirus type 2
PRDC	Porcine Respiratory Disease Complex

PRRSV	Porcine reproductive and respiratory syndrome virus
PFGE	Pulsed field gel electrophoresis
rep-PCR	Repetitive element based-polymerase chain reaction
RNA	Ribonucleic acid
RNase	Ribonuclease
sec	Second
TAE	Tris-acetate-ethylenediaminetetraacetic acid
UK	United Kingdom
UPM	Universiti Putra Malaysia
USA	United States of America
UV	Ultraviolet
v	Version
vtaA	Virulence associated trimeric autotransporter
× <i>g</i>	Relative centrifugal force

## CHAPTER 1

### INTRODUCTION

In Malaysia, the pig industry is nearly self-sustainable and has remained stagnant for the past few years amounting at around 1.4 million head of pig production each year (Malaysia Department of Veterinary Services, 2018). A slight drop of pig production was observed in the recent three years due to the closing of numerous small-scale farms. Farms of Peninsular Malaysia account most of the Malaysia's pig production in which farms in Sarawak also contributed a significant number.

Swine respiratory diseases are common among pigs especially in the weaners group (Oliveira, 2002). Piglets are exposed to the respiratory pathogens as early since the first week of age and cause high mortality during outbreaks (Choi et al., 2003; Turni and Blackall, 2007). Porcine circovirus type 2 (PCV2) and porcine reproductive and respiratory syndrome virus (PRRSV) are few viruses affecting the respiratory system of pigs and is often complicated with secondary bacterial infections such as *Haemophilus parasuis*, *Mycoplasma hyopneumoniae* and *Streptococcus suis* (Thacker, 2001; Kim, Chung and Chae, 2003; Hansen et al., 2010; Opriessnig et al., 2011). As a matter of fact, appearing as a complex of multiple agents causing respiratory problem in pigs, it is termed as porcine respiratory disease complex (PRDC).

Commercial vaccines have been widely developed to target porcine viral diseases and have proved to be effective in controlling diseases. On the other hand, with the exception of *M. hyopneumoniae*, researchers still face hardship in developing the most ideal vaccines for swine bacterial diseases due to their vast genotypic and phenotypic variability (Dekker et al., 2012; Liu et al., 2016). In Malaysia, neither active study is being carried out nor a proper diagnostic tool that fully differentiating each bacterial pathogen in pigs is available. Most of the time, diagnosis was based on presumptive observation of post mortem lesions which most of the time might not be true.

Important bacterial pathogens of swine were often difficult to isolate (Olvera et al., 2007; Cook et al., 2016). Without proper sampling conditions and appropriate culture methods, the suspected disease would have been misdiagnosed. *Haemophilus parasuis* is one important disease of the swine and is ubiquitous (Miani et al., 2017). Not only causing Glässer's disease on its own, it also contributes to the PRDC, which is one of the leading cause for the mortality in grower and finisher pigs (Aragon et al., 2012; Howell et al., 2015). Glässer's disease is present in all major pig farming countries and the prevalence is increasing (Aragon et al., 2012). Nonetheless, Malaysia having its own pig population is not exempted from this issue. Over the fifty years of pig farming in Malaysia, Glässer's disease has persisted and yet efforts in

defining this bacteria is absent and strategies to control this disease often come to no avail.

*Haemophilus parasuis* is a pathogen of great economic impact. Based on United States statistic, it is the second major causes of mortality in prewean pigs (Holtkamp, Rotto, and Garcia, 2007). Infection rate can go up to 50-75% and mortality rate ranges above 10% (Bak and Riising, 2002; Aragon et al., 2012). Nowadays, it was not only common in the weaner pigs but also the grower pigs. This is a significant loss to the production due to the great number of mortality and unthriftiness (Castilla et al., 2012). This disease is currently considered a menace to the industry since high cost of antimicrobial usage are inflicted and affected animals are disposed, and this occurs even in farms with good husbandry management (Nedbalcova et al., 2006). According to the National Animal Disease Information Service (NADIS) of UK, losses in an outbreak farm were estimated at £14000 in a 300-sow farm per year alone (White, 2012). Therefore, effective and time saving methods of pathogen detection were extremely crucial in disease management. Prompt diagnosis will prevent further disease spread and minimize losses from the negative impacts of disease.

Currently, the available commercial *H. parasuis* vaccines are serotype-based. Undesirable results of vaccination were seen in some farms, providing the idea that vaccine strains are heterologous of field strain and the immunity triggered was not sufficient for protection (Liu et al., 2016). Although cross-protection among serotypes has been portrayed, yet complete protection is still impossible as the neutralizing antibodies production against heterologous strains were low (Miniats, Smart and Rosendal, 1991; Nielsen, 1993; Rapp-Gabrielson et al., 1997; Bak and Riising, 2002). Therefore, the knowledge of the serotype distribution is indeed of paramount importance for the best vaccine selection for disease control. In long term, it is hope that the *H. parasuis* can be effectively control via strategic vaccination in turn minimizing disease occurrence; ultimately reducing the dependent of antimicrobial in disease management.

Despite frequent occurrence of *H. parasuis* in Malaysia pig herds, the nature of these bacteria remains unknown. Therefore, the objectives of this study are:

1. to detect presence of *H. parasuis* in Peninsular Malaysia clinical samples/pigs using PCR technique.
2. to detect major serotypes of *H. parasuis* namely serotype 4, 5 or 12 and 13 in using multiplex PCR assay.
3. to detect the *OMPP2* gene of *H. parasuis* using PCR technique.
4. to determine the phylogenetic relationship of Peninsular Malaysia *H. parasuis* and other reference strains by *OMPP2* virulence gene.

It is hypothesize that:

1. *Haemophilus parasuis* is present in Peninsular Malaysia and is detectable by using PCR technique.
2. serotypes 4, 5 or 12 and 13 are present in Peninsular Malaysia.
3. *OMPP2* gene is detected in *H. parasuis*.
4. there is a close relationship between Peninsular Malaysia *H. parasuis* and other reference strains.





## REFERENCES

- Aarestrup, F. M., Seyfarth, A. M., & Angen, Ø. (2004). Antimicrobial susceptibility of *Haemophilus parasuis* and *Histophilus somni* from pigs and cattle in Denmark. *Veterinary Microbiology*, 101(2), 143-146.
- Abd-Elsalam, K. A. (2003). Bioinformatic tools and guideline for PCR primer design. *African Journal of Biotechnology*, 2(5), 91-95.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215(3), 403-410.
- Amano H., Shibata M., Kajio N., & Morozumi T. (1994): Pathologic observations of pigs intranasally inoculated with serovar 1, 4 and 5 of *Haemophilus parasuis* using immunoperoxidase method. *Journal of Veterinary Medical Science*, 56, 639-644.
- Amano, H., Shibata, M., Takahashi, K., & Sasaki, Y. (1997). Effects on endotoxin pathogenicity in pigs with acute septicemia of *Haemophilus parasuis* infection. *Journal of Veterinary Medical Science*, 59(6), 451-455.
- Andersen, C., Maier, E., Kemmer, G., Benz, J., Hilpert, A. K., Benz, R., & Reidl, J. (2003). Porin *OmpP2* of *Haemophilus influenzae* shows substrate specificity towards nicotinamide-derived nucleotide substrates. *Journal of Biological Chemistry*. 278(27), 24269-24276.
- Angen, O., Oliveira, S., Ahrens, P., Svensmark, B., & Leser, T. D. (2007). Development of an improved species specific PCR test for detection of *Haemophilus parasuis*. *Veterinary Microbiology*, 119(2-4), 266-276.
- Angen, O., Svensmark, B., & Mittal, K. R. (2004). Serological characterization of Danish *Haemophilus parasuis* isolates. *Veterinary Microbiology*, 103(3-4), 255-258.
- Aragon, V., Bouchet, B., & Gottschalk, M. (2010). Invasion of endothelial cells by systemic and nasal strains of *Haemophilus parasuis*. *The Veterinary Journal*, 186(2), 264-267.
- Aragon, V., Cerdà-Cuellar, M., Fraile, L., Mombarg, M., Nofrarias, M., Olvera, A., Sibila, M., Solanes, D., & Segalés, J. (2010). Correlation between clinico-pathological outcome and typing of *Haemophilus parasuis* field strains. *Veterinary Microbiology*, 142(3-4), 387-393.
- Aragon, V., Segalés, J., & Oliveira, S. (2012). Glässer's Disease. In J. J. Zimmerman, L. A. Karriker, A. Ramirez, K. J. Schwartz, & G. W. Stevenson (Eds.), *Diseases of Swine, 10th Edition* (pp. 760-769). Ames, Iowa: Wiley-Blackwell.
- Avadhanula, V., Rodriguez, C. A., Ulett, G. C., Bakaletz, L. O., & Adderson, E. E. (2006). Nontypeable *Haemophilus influenzae* adheres to intercellular

adhesion molecule 1 (ICAM-1) on respiratory epithelial cells and upregulates ICAM-1 expression. *Infection and immunity*, 74(2), 830-838.

- Bak, H., & Riising, H. J. (2002). Protection of vaccinated pigs against experimental infections with homologous and heterologous *Haemophilus parasuis*. *The Veterinary Record*, 151(17), 502-505.
- Bello-Ortí, B., Deslandes, V., Tremblay, Y.D., Labrie, J., Howell, K.J., Tucker, A.W., Maskell, D.J., Aragon, V., & Jacques, M. (2014). Biofilm formation by virulent and non-virulent strains of *Haemophilus parasuis*. *Veterinary Research*, 45(1), 104.
- Benson, D. A., Cavanaugh, M., Clark, K., Karsch-Mizrachi, I., Lipman, D. J., Ostell, J., & Sayers, E. W. (2012). GenBank. *Nucleic acids research*, 41, D36-42.
- Biberstein, E. L., & White, D. C., (1969). A proposal for the establishment of two new *Haemophilus* species. *Journal Medical Microbiology*, 2, 75-78.
- Blackall, P. J., Rapp-gabrielson, V. J., & Hampson, D. J. (1996). Serological characterisation of *Haemophilus parasuis* isolates from Australian pigs, *Australian Veterinary Journal*, 73(3), 93-95.
- Blackall, P. J., Trott, D. J., Rapp-Gabrielson, V., & Hampson, D. J. (1997). Analysis of *Haemophilus parasuis* by multilocus enzyme electrophoresis. *Veterinary Microbiology*, 56(1-2), 125-134.
- Bouchet, B., Vanier, G., Jacques, M., & Gottschalk, M. (2008). Interactions of *Haemophilus parasuis* and its LOS with porcine brain microvascular endothelial cells. *Veterinary Research*, 39(5), 1.
- Bouchet, B., Vanier, G., Jacques, M., Auger, E., & Gottschalk, M. (2009). Studies on the interactions of *Haemophilus parasuis* with porcine epithelial tracheal cells: limited role of LOS in apoptosis and pro-inflammatory cytokine release. *Microbial Pathogenesis*, 46(2), 108-113.
- Buckles, E.L., Wang, X., Lane, M.C., Lockett, C.V., Johnson, D.E., Rasko, D.A., Mobley, H.L. and Donnenberg, M.S. (2009). Role of the K2 capsule in *Escherichia coli* urinary tract infection and serum resistance. *The Journal of Infectious Diseases*, 199(11),1689-1697.
- Cai, X., Chen, H., Blackall, P. J., Yin, Z., Wang, L., Liu, Z., & Jin, M. (2005). Serological characterization of *Haemophilus parasuis* isolates from China. *Veterinary microbiology*, 111(3-4), 231-236.
- Calsamiglia, M., Pijoan, C., Solano, G., & Rapp-Gabrielson, V. (1999). Development of an oligonucleotide-specific capture plate hybridization assay for detection of *Haemophilus parasuis*. *Journal of Veterinary Diagnostic Investigation*, 11(2), 140-145.

- Castilla, K. S., de Gobbi, D. D. S., Moreno, L. Z., Paixão, R., Coutinho, T. A., dos Santos, J. L., & Moreno, A. M. (2012). Characterization of *Haemophilus parasuis* isolated from Brazilian swine through serotyping, AFLP and PFGE. *Research in Veterinary Science*, 92(3), 366-371.
- Choi, Y. K., Goyal, S. M., & Joo, H. S. (2003). Retrospective analysis of etiologic agents associated with respiratory diseases in pigs. *The Canadian Veterinary Journal*, 44(9), 735.
- Cook, B. S., Beddow, J. G., Manso-Silvan, L., Maglennon, G. A., & Rycroft, A. N. (2016). Selective medium for culture of *Mycoplasma hyopneumoniae*. *Veterinary Microbiology*, 195, 158-164.
- Costa-Hurtado, M., & Aragon, V. (2013). Advances in the quest for virulence factors of *Haemophilus parasuis*. *The Veterinary Journal*, 198(3), 571-576.
- Dabo, S. M., Confer, A. W., & Quijano-Blas, R. A. (2003). Molecular and immunological characterization of *Pasteurella multocida* serotype A: 3 OmpA: evidence of its role in *P. multocida* interaction with extracellular matrix molecules. *Microbial pathogenesis*, 35(4), 147-157.
- Davies, R. L., & Lee, I. (2004). Sequence diversity and molecular evolution of the heat-modifiable outer membrane protein gene (ompA) of *Mannheimia (Pasteurella) haemolytica*, *Mannheimia glucosida*, and *Pasteurella trehalosi*. *Journal of Bacteriology*, 186(17), 5741-5752.
- Dayao, D. A., Kienzle, M., Gibson, J. S., Blackall, P. J., & Turni, C. (2014). Use of a proposed antimicrobial susceptibility testing method for *Haemophilus parasuis*. *Veterinary Microbiology*, 172, 586-589.
- de la Fuente, A. M., Tucker, A. W., Navas, J., Blanco, M., Morris, S. J., & Gutierrez-Martın, C. B. (2007). Antimicrobial susceptibility patterns of *Haemophilus parasuis* from pigs in the United Kingdom and Spain. *Veterinary Microbiology*, 120(1-2), 184-191.
- Dekker, C. N. T., Bouma, A., Daemen, A. J. J. M., van Leengoed, L. A. M. G., Jonker, F. H., Wagenaar, J. A., & Stegeman, J. A. (2012). Homologous whole bacterin vaccination is not able to reduce *Streptococcus suis* serotype 9 strain 7997 transmission among pigs or colonization. *Vaccine*, 30(7), 1379-1387.
- Department of Veterinary Services Malaysia, (2018). Livestock Statistic. Retrieved 20 January 2019 from <http://www.dvs.gov.my/index.php/pages/view/1847>
- Drolet, R., Germain, M. C., Tremblay, C., & Higgins, R. (2000). Ear panniculitis associated with *Haemophilus parasuis* infection in growing-finishing pigs. Proceeding from 16th International Pig Veterinary Society Congress (pp.528). Melbourne, Australia.

- Eaves, L. E., Blackall, P. J., & Fegan, M. (1989). Characterisation and antimicrobial sensitivity of haemophili isolated from pigs. *Australian Veterinary Journal*, 66(1), 1-4.
- Fablet, C., Marois, C., Dorenlor, V., Eono, F., Eveno, E., Jolly, J. P., Le Devendec, L., Kobisch, M., Madec, F., & Rose, N. (2012). Bacterial pathogens associated with lung lesions in slaughter pigs from 125 herds. *Research in Veterinary Science*, 93(2), 627-630.
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution*, 39(4), 783-791.
- Frndoloso, R., Martinez-Martinez, S., Gutierrez-Martin, C. B., Rodriguez-Ferri, E. F. (2012). *Haemophilus parasuis* serovar 5 Nagasaki strain adheres and invades PK-15 cells. *Veterinary Microbiology*, 154, 347–352.
- Galofré-Milà, N., Correa-Fiz, F., Lacouture, S., Gottschalk, M., Strutzberg-Minder, K., Bensaid, A., Pina-Pedrero, S., & Aragon, V. (2017). A robust PCR for the differentiation of potential virulent strains of *Haemophilus parasuis*. *BMC Veterinary Research*, 13(1), 124.
- Glässer, K. (1910). Untersuchungen über die Schweineseuche mit besonderer Berücksichtigung ihrer Ätiologie und Pathologie [Studies on swine fever especially regarding etiology and pathology of the disease]. *Deutsche Tierärztliche Wochenschrift*, 18, 729-733.
- Hansen, M. S., Pors, S. E., Jensen, H. E., Bille-Hansen, V., Bisgaard, M., Flachs, E. M., & Nielsen, O. L. (2010). An investigation of the pathology and pathogens associated with porcine respiratory disease complex in Denmark. *Journal of Comparative Pathology*, 143(2-3), 120-131.
- Harper, M., Cox, A. D., St Michael, F., Wilkie, I. W., Boyce, J. D., Adler, B. (2004). A heptosyltransferase mutant of *Pasteurella multocida* produces a truncated lipopolysaccharide structure and is attenuated in virulence. *Infectious Immunology*. 72, 3436–3443.
- Hibbing, M. E., Fuqua, C., Parsek, M. R., & Peterson, S. B. (2010). Bacterial competition: surviving and thriving in the microbial jungle. *Nature Reviews Microbiology*, 8(1), 15.
- Hiltke, T. J., Schiffmacher, A. T., Dagonese, A. J., Sethi, S., & Murphy, T. F. (2003). Horizontal transfer of the gene encoding outer membrane protein P2 of nontypeable *Haemophilus influenzae* in a patient with chronic obstructive pulmonary disease. *The Journal of Infectious Diseases*, 188(1), 114-117.
- Hiltke, T. J., Sethi, S., & Murphy, T. F. (2002). Sequence stability of the gene encoding outer membrane protein P2 of nontypeable *Haemophilus influenzae* in the human respiratory tract. *The Journal of Infectious Diseases*, 185(5), 627-631.

- Hjärre, A., & Wramby, G. (1943). über die fibrinöse Serosa-Gelenkentzündung (Glässer) beim Schwein. *Zeitschrift für Infektionskrankheiten, Parasitäre Krankheiten und Hygiene der Haustiere*, 60.
- Hoefling, D. C. (1991). Acute myositis associated with *Hemophilus parasuis* in primary SPF sows. *Journal of Veterinary Diagnostic Investigation*, 3, 354-355.
- Holtkamp, D., Rotto, H., & Garcia, R. (2007). Economic cost of major health challenges in large US swine production systems. *Swine news*, 30, 85-89.
- Hopkins Lab. n.d. PCR Primer Design Basic Guidelines [PDF file], Retrieved 20 January 2019 from [https://www.researchgate.net/profile/Laurence\\_Dawkins-Hall/post/What\\_is\\_the\\_acceptable\\_penalty\\_value\\_for\\_sequencing\\_primer\\_designed\\_by\\_Primer3/attachment/59d631d179197b807798f956/AS%3A367738649366534%401464687134546/download/PCR+Primer+Design+Guidelines+1\\_4.pdf](https://www.researchgate.net/profile/Laurence_Dawkins-Hall/post/What_is_the_acceptable_penalty_value_for_sequencing_primer_designed_by_Primer3/attachment/59d631d179197b807798f956/AS%3A367738649366534%401464687134546/download/PCR+Primer+Design+Guidelines+1_4.pdf)
- Howell, K. J., Peters, S. E., Wang, J., Hernandez-Garcia, J., Weinert, L. A., Luan, S. L., Chaudhuri, R. R., Angen, Ø., Aragon, V., Williamson, S. M., & Parkhill, J. (2015). Development of a multiplex PCR for rapid molecular serotyping of *Haemophilus parasuis*. *Journal of Clinical Microbiology*, 53(12), 3812-3821.
- Inzana, T. J., Dickerman, A. W., & Bandara, A. B. (2016). Taxonomic reclassification of "*Haemophilus parasuis*" to *Glaesserella parasuis*. Proceeding from *The Prato Conference on the Pathogenesis of Bacterial Infections of Animals 2016*. Prato, Italy. Retrieved 20 January 2019 from <http://vetpath-2016.p.asnevents.com.au/days/2016-10-12/abstract/38912>
- Jia, A. Q., Li, C. L., Wang, G. P., & Yang, D. X. (2009). Institute Veterinary Medicine, Guangdong Academy of Agricultural Sciences. Retrieved 5<sup>th</sup> August 2019 from <https://www.ncbi.nlm.nih.gov/genbank/>
- Jukes, T. H., & Cantor, C. R. (1969). Evolution of Protein Molecules. In H. N. Munro (Ed.), *Mammalian Protein Metabolism* (pp. 21-132). New York: Academic Press.
- Kielstein, P., & Rapp-Gabrielson, V. J. (1992). Designation of 15 serovars of *Haemophilus parasuis* on the basis of immunodiffusion using heat-stable antigen extracts. *Journal of Clinical Microbiology*, 30(4), 862–865.
- Kielstein, P., Wuthe, H. H., Angen, Ø., Mutters, R., & Ahrens, P. (2001). Phenotypic and genetic characterization of NAD-dependent Pasteurellaceae from the respiratory tract of pigs and their possible pathogenetic importance. *Veterinary Microbiology*, 81(3), 243-255.
- Kim, J., Chung, H. K., & Chae, C. (2003). Association of porcine circovirus 2 with porcine respiratory disease complex. *The Veterinary Journal*, 166(3), 251-256.

- Köfer, J., Hinterdorfer, F., & Awad-Masalmeh, M. (1992). Occurrence and drug resistance of bacteria pathogenic to the lungs from autopsy material of swine. *Tierärztliche Praxis*, 20(6), 600-604.
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33(7), 1870-1874.
- Lacouture, S., Rodriguez, E., Strutzberg-Minder, K., & Gottschalk, M. (2017). Canada: serotyping of *Haemophilus parasuis* field isolates from diseased pigs in Quebec by indirect hemagglutination assay and multiplex polymerase chain reaction (PCR). *The Canadian Veterinary Journal*, 58(8), 802.
- Lawrence, P. K., Wiener, B. L., Kolander-Bremer, T., Bey, R. F., Stine, D. L., Kittichotirat, W., & Bumgarner, R. E. (2014). Genome-wide association studies of virulent and avirulent *Haemophilus parasuis* serotype 4 strains. *Genome Announcements*, 2(5), e00884-14.
- Lewis, P. A., & Shope, R. E. (1931). Swine influenza: II. A hemophilic bacillus from the respiratory tract of infected swine. *Journal of Experimental Medicine*, 54(3), 361-371.
- Li, J. X., Jiang, P., Wang, Y., Li, Y. F., Chen, W., Wang, X. W., & Li, P. (2009). Genotyping of *Haemophilus parasuis* from diseased pigs in China and prevalence of two coexisting virus pathogens. *Preventive Veterinary Medicine*, 91(2-4), 274-279.
- Li, P., Bai, J., Li, J. X., Zhang, G. L., Song, Y. H., Li, Y. F., Wang, X. W., & Jiang, P. (2012). Molecular cloning, sequencing, and expression of the outer membrane protein P2 gene of *Haemophilus parasuis*. *Research in Veterinary Science*, 93(2), 736-742.
- Lin, W. H., Shih, H. C., Lin, C. F., Yang, C. Y., Chang, Y. F., Lin, C. N., & Chiou, M. T. (2018). Molecular serotyping of *Haemophilus parasuis* isolated from diseased pigs and the relationship between serovars and pathological patterns in Taiwan. *PeerJ*, 6, e6017.
- Lorenson, M. S., Miani, M., Guizzo, J. A., Barasoul, B., Martínez-Martínez, S., Rodríguez-Ferri, E. F., Gutiérrez-Martín, C. B., Kreutz, L. C., & Frandoloso, R. (2017). Altered indirect hemagglutination method for easy serotyping of *Haemophilus parasuis*. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 69(1), 15-21.
- Lacouture, S., Rodriguez, E., Strutzberg-Minder, K., & Gottschalk, M. (2017). Canada: serotyping of *Haemophilus parasuis* field isolates from diseased pigs in Quebec by indirect hemagglutination assay and multiplex polymerase chain reaction (PCR). *The Canadian Veterinary Journal*, 58(8), 802.

- Lin, W. H., Shih, H. C., Lin, C. F., Yang, C. Y., Chang, Y. F., Lin, C. N., & Chiou, M. T. (2018). Molecular serotyping of *Haemophilus parasuis* isolated from diseased pigs and the relationship between serovars and pathological patterns in Taiwan. *PeerJ*, 6, e6017.
- Liu, H., Xue, Q., Zeng, Q., & Zhao, Z. (2016). *Haemophilus parasuis* vaccines. *Veterinary Immunology and I*, 180, 53-58.
- Luppi, A., Bonilauri, P., Dottori, M., Iodice, G., Gherpelli, Y., Merialdi, G., Maioli, G., & Martelli, P. (2013). *Haemophilus parasuis* serovars isolated from pathological samples in Northern Italy. *Transboundary and Emerging Diseases*, 60(2), 140-142.
- Ma, L., Wang, L., Chu, Y., Li, X., Cui, Y., Chen, S., Zhou, J., Li, C., Lu, Z., Liu, J., & Liu, Y. (2016). Characterization of Chinese *Haemophilus parasuis* isolates by traditional serotyping and molecular serotyping methods. *PLoS one*, 11(12), e0168903.
- Macedo, N., Rovira, A., & Torremorell, M. (2015). *Haemophilus parasuis*: infection, immunity and enrofloxacin. *Veterinary Research*, 46(1), 128.
- Macedo, N. R., Oliveira, S. R., Lage, A. P., Santos, J. L., Araújo, M. R., & Guedes, R. M. C. (2011). ERIC-PCR genotyping of *Haemophilus parasuis* isolates from Brazilian pigs. *The Veterinary Journal*, 188(3), 362-364.
- MacInnes, J. I., & Desrosiers, R. (1999). Agents of the " suis-ide diseases" of swine: *Actinobacillus suis*, *Haemophilus parasuis*, and *Streptococcus suis*. *Canadian Journal of Veterinary Research*, 63(2), 83.
- Macpherson, M. R., & Hodges, R. T. (1985). The occurrence of mycoplasmas in the lungs of pigs in New Zealand. *New Zealand Veterinary Journal*, 33(11), 194-197.
- Menard, J., & Moore, C. (1990). Epidemiology and management of Glasser's disease in SPF herds. Proceeding from 21st Annual Meeting of American Association of Swine Practitioner (pp.187-200). Denver, Colorado.
- Menin, A., Gava, D., & Vaz, E. K. (2005). Aspectos gerais sobre a infecção por *Haemophilus parasuis* em suínos-Revisão. *Rev Ciên Agrovet*, 4, 148-156.
- Miani, M., Lorenson, M. S., Guizzo, J. A., Espíndola, J. P., Rodríguez-Ferri, E. F., Gutiérrez-Martín, C. B., Kreutz, L. C., & Frondoloso, R. (2017). Antimicrobial susceptibility patterns of Brazilian *Haemophilus parasuis* field isolates. *Pesquisa Veterinária Brasileira*, 37(11), 1187-1192.
- Miniats, O. P., Smart, N. L., & Rosendal, S. (1991). Cross protection among *Haemophilus parasuis* strains in immunized gnotobiotic pigs. *Canadian Journal of Veterinary Research*, 55(1), 37.

- Molitor, T. (1993). Secondary infections associated with porcine reproductive and respiratory syndrome. Proceeding from Allen D. Leman Conference (pp.236-237). St. Paul, Minnesota.
- Møller, K., & Kilian, M. (1990). V factor-dependent members of the family Pasteurellaceae in the porcine upper respiratory tract. *Journal of Clinical Microbiology*, 28(12), 2711-2716.
- Møller, K., Andersen, L. V., Christensen, G., & Kilian, M. (1993). Optimization of the detection of NAD dependent Pasteurellaceae from respiratory tract of slaughterhouse pigs. *Veterinary Microbiology*, 36, 261-271.
- Møller, K., Fussing, V., Grimont, D., Paster, J., Dewhirst, E., & Kilian, M. (1996). *Actinobacillus minor* sp. nov., *Actinobacillus porcinus* sp. nov. and *Actinobacillus indolicus* sp. nov., three new V-factor dependent species from respiratory tract of pigs. *International Journal of Systematic Bacteriology*, 46, 951-956.
- Morikoshi, T., Kobayashi, K., Kamino, T., Owaki, S., Hayashi, S., & Hirano, S. (1990). Characterization of *Haemophilus parasuis* isolated in Japan. *The Japanese Journal of Veterinary Science*, 52(3), 667-669.
- Morozumi, T., & Nicolet, J. (1986). Morphological variations of *Haemophilus parasuis* strains. *Journal of Clinical Microbiology*, 23(1), 138-142.
- Mullins, M. A., Register, K. B., Bayles, D. O., Loving, C. L., Nicholson, T. L., Brockmeier, S. L., Dyer, D. W., & Phillips, G. J. (2009). Characterization and comparative analysis of the genes encoding *Haemophilus parasuis* outer membrane proteins P2 and P5. *Journal of Bacteriology*, 191, 5988–6002.
- Mullis, K. B., & Faloona, F. A. (1987). Specific synthesis of DNA in vitro via a polymerase-catalyzed chain reaction. In R. Wu (Ed.), *Methods in Enzymology Vol. 155* (pp. 335-350). Academic Press.
- Nedbalcova, K., Kucerova, Z., Krejci, J., Tesarik, R., Gopfert, E., Kummer, V., Leva, L., Kudlackova, H., Ondriasova, R., & Faldyna, M. (2011). Passive immunisation of post-weaned piglets using hyperimmune serum against experimental *Haemophilus parasuis* infection. *Research in Veterinary Science*, 91(2), 225-229.
- Nedbalcova, K., Satran, P., Jaglic, Z., Ondriasova, R., & Kucerova, Z. (2006). *Haemophilus parasuis* and Glässer's disease in pigs: a review. *Veterinarni Medicina*, 51(5), 168-179.
- Nicolet, J. (1992). *Haemophilus parasuis*. In A. D. Leman, B. E. Straw, W. L. Mengeling, S. D'Allaire, & D. J. Taylor (Eds.), *Diseases of Swine*, 7<sup>th</sup> Edition (pp. 526-528). Iowa, USA: Iowa State University Press.
- Nielsen, R. (1993). Pathogenicity and immunity studies of *Haemophilus parasuis* serotypes. *Acta Veterinaria Scandinavica*, 34(2), 193-198.



- Nielsen, R., & Danielson, V. (1975). An outbreak of Glässer's disease. Studies on etiology, serology and the effect of vaccination. *Nordisk Veterinaermedicin*, 27(1), 20-25.
- Oliveira, S. (2007). *Haemophilus parasuis* diagnostics. *Journal of Swine Health and Production*, 15(2), 99-103.
- Oliveira, S., Blackall, P. J., & Pijoan, C. (2003). Characterization of the diversity of *Haemophilus parasuis* field isolates by use of serotyping and genotyping. *American Journal of Veterinary Research*, 64(4), 435-442.
- Oliveira, S., Galina, L., & Pijoan, C. (2001). Development of a PCR test to diagnose *Haemophilus parasuis* infections. *Journal of Veterinary Diagnostic Investigation*, 13(6), 495-501.
- Oliveira, S., Pijon, C., & Morrison, R. (2002). The role of *Haemophilus parasuis* in nursery mortality. Proceeding from Allen D. Leman Swine Conference (pp. 111-113). Minnesota: University of Minnesota Digital Conservancy.
- Oliveira, S., & Pijoan, C. (2004). *Haemophilus parasuis*: new trends on diagnosis, epidemiology and control. *Veterinary Microbiology*, 99, 1-12.
- Olvera, A., Calsamiglia, M., & Aragon, V. (2006). Genotypic diversity of *Haemophilus parasuis* field strains. *Applied and Environmental Microbiology*, 72(6), 3984-3992.
- Olvera, A., Cerdà-Cuéllar, M., & Aragon, V. (2006). Study of the population structure of *Haemophilus parasuis* by multilocus sequence typing. *Microbiology*, 152(12), 3683-3690.
- Olvera, A., Segalés, J., & Aragón, V. (2007). Update on the diagnosis of *Haemophilus parasuis* infection in pigs and novel genotyping methods. *Veterinary Journal*, 174(3), 522-529.
- Opriessnig, T., Giménez-Lirola, L. G., & Halbur, P. G. (2011). Polymicrobial respiratory disease in pigs. *Animal Health Research Reviews*, 12(2), 133-148.
- Pearson, W. R. (2013). An introduction to sequence similarity ("homology") searching. *Current Protocols in Bioinformatics*, 42(1), 3-1.
- Pereira, D. A., Dalla-Costa, F. A., Ferroni, L. B., Moraes, C. N., Schocken-Iturrino, R. P., & Oliveira, L. G. (2017). The challenges with Glässer's disease in technified pig production. *Austral Journal of Veterinary Sciences*, 49(2), 63-69.
- Rafiee, M., & Blackall, P. J. (2000). Establishment, validation and use of the Kielstein-Rapp-Gabrielson serotyping scheme for *Haemophilus parasuis*. *Australian Veterinary Journal*, 78, 172-174.

- Rapp-Gabrielson, V. J., & Gabrielson, D. A. (1992). Prevalence of *Haemophilus parasuis* serovars among isolates from swine. *American Journal of Veterinary Research*, 53(5), 659-664.
- Rapp-Gabrielson, V. J., Kocur, G. J., Clark, J. T., & Muir, S. K. (1997). *Haemophilus parasuis*: immunity in swine after vaccination. *Veterinary Medicine*, 92(1), 83-90.
- Rapp-Gabrielson, V. J., Oliveira, S. R., & Pijoan, C. (2006). *Haemophilus parasuis*. In B. E. Straw, J. J. Zimmerman, S. D'Allaire, D. J. Taylor (Eds.), *Diseases of Swine 9<sup>th</sup> Edition* (pp.681-690). Ames, Iowa: Blackwell Publishing
- Río, M. L. D., Gutiérrez, B., Gutiérrez, C. B., Monter, J. L., & Ferri, E. F. R. (2003). Evaluation of survival of *Actinobacillus pleuropneumoniae* and *Haemophilus parasuis* in four liquid media and two swab specimen transport systems. *American Journal of Veterinary Research*, 64(9), 1176-1180.
- Rúbies, X., Kielstein, P., Costa, L., Riera, P., Artigas, C., & Espuña, E. (1999). Prevalence of *Haemophilus parasuis* serovars isolated in Spain from 1993 to 1997. *Veterinary Microbiology*, 66(3), 245-248.
- Sack, M., Buettner, F. F., & Baltes, N. (2009). Variability of the *Haemophilus parasuis* OMP P2 protein. *National Center for Biotechnology Information*. Retrieved 5th August 2018 from <https://www.ncbi.nlm.nih.gov/genbank/>
- Saitou, N., & Nei, M. (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4, 406-425.
- Santos, J. L., Sobestiansky, J., & Santos, L. F. (2012). Doença de Glässer. In J. Sobestiansky (Ed). *Doenças dos Suínos 1st Edition* (pp.135-140). Goiânia, Brazil: Cãnone Editorial.
- Segales, J., Domingo, M., Solano, G. I., & Pijoan, C. (1997). Immunohistochemical detection of *Haemophilus parasuis* serovar 5 in formalin-fixed, paraffin-embedded tissues of experimentally infected swine. *Journal of Veterinary Diagnostic Investigation*, 9, 237-243.
- Smart, N. L., Miniats, O. P., & McInnes, J. I. (1988). Analysis of *Haemophilus parasuis* isolates from southern Ontario swine by restriction endonuclease fingerprinting. *Canadian Journal of Veterinary Research*, 52, 319-324.
- Sikkema, D. J., & Murphy, T. F. (1992). Molecular analysis of the P2 porin protein of nontypeable *Haemophilus influenzae*. *Infectious Immunology*, 60, 5204– 5211.
- Solano, G. I., Segales, J., Collins, J. E., Molitor, T. W., & Pijoan, C. (1997). Porcine reproductive and respiratory syndrome virus (PRRSv) interaction with *Haemophilus parasuis*. *Veterinary Microbiology*, 55, 247-257.

- Tadjine, M., Mittal, K. R., Bourdon, S., & Gottschalk, M. (2004). Development of a new serological test for serotyping *Haemophilus parasuis* isolates and determination of their prevalence in North America. *Journal of Clinical Microbiology*, 42(2), 839-840.
- Takahashi, K., Naga, S., Yagihashi, T., Ikehata, T., Nakano, Y., Senna, K., Maruyama, T., & Murofushi, J. (2001) A cross-protection experiment in pigs vaccinated with *Haemophilus parasuis* serovars 2 and 5 bacterins, and evaluation of a bivalent vaccine under laboratory and field conditions. *Journal of Veterinary Medical Science*, 63, 487-491.
- Tang, C., Zhang, B., Yue, H., Yang, F., Shao, G., Hai, Q., Chen, X., Guo, D. (2010). Characteristics of the molecular diversity of the outer membrane protein A gene of *Haemophilus parasuis*. *Canadian Journal of Veterinary Research*, 74, 233-236.
- Teh, S. W., & Ooi, P. T. (2011). Detection of Glasser's Disease in Clinical Samples using Polymerase Chain Reaction. Proceeding from 6th *Proceedings of the Seminar on Veterinary Sciences* (pp.99-103). Serdang, Malaysia.
- Thacker, E. L., & Minion, F. C. (2012). Mycoplasmosis. In J. J. Zimmerman, L. A. Karriker, A. Ramirez, K. J. Schwartz, & G. W. Stevenson (Eds.), *Diseases of Swine, 10th Edition* (pp. 779-797). Ames, Iowa: Wiley-Blackwell.
- Thacker, E. L. (2001). Immunology of the porcine respiratory disease complex. *Veterinary Clinics: Food Animal Practice*, 17(3), 551-565.
- The Straits Time. (2017). Singapore receives first import of live pigs from Malaysia in 18 years. Retrieved 20 January 2019 from <https://www.straitstimes.com/singapore/singapore-receives-first-import-of-live-pigs-from-malaysia-in-18-years>
- Torres, A. G., & Kaper, J. B. (2003). Multiple elements controlling adherence of enterohemorrhagic *Escherichia coli* O157: H7 to HeLa cells. *Infection and Immunity*, 71(9), 4985-4995.
- Turni, C., & Blackall, P. J. (2007). Comparison of sampling sites and detection methods for *Haemophilus parasuis*. *Australian Veterinary Journal*, 85(5), 177-184.
- Turni, C., Pyke, M., & Blackall, P. J. (2010). Validation of a real time PCR for *Haemophilus parasuis*. *Journal of Applied Microbiology*, 108(4), 1323-1331.
- Vahle, J. L., Haynes, J. S., & Andrews, J. J. (1995). Experimental reproduction of *Haemophilus parasuis* infection in swine: clinical, bacteriological, and morphologic findings. *Journal of Veterinary Diagnostic Investigation*, 7, 476-480.

- Vanier, G., Szczotka, A., Friedl, P., Lacouture, S., Jacques, M., & Gottschalk, M. (2006). *Haemophilus parasuis* invades porcine brain microvascular endothelial cells. *Microbiology*, *152*(1), 135-142.
- Wang, Y., Liu, C., Fang, Y., Liu, X., Li, W., Liu, S., Liu, Y., Charreyre, C., Audonnet, J.C., Chen, P., He, Q. (2012). Transcription analysis on response of porcine alveolar macrophages to *Haemophilus parasuis*. *BMC Genomics*, *13*, 86.
- Wang, Z., Zhao, Q., Wei, H., Wen, X., Cao, S., Huang, X., Wu, R., Yan, Q., Huang, Y., & Wen, Y. (2017). Prevalence and seroepidemiology of *Haemophilus parasuis* in Sichuan province, China. *PeerJ*, *5*, e3379.
- Weng, J. K. & Cruz, N. (2015). 7.15 Experimental Molecular Genetics. *Massachusetts Institute of Technology: MIT OpenCourseWare*, Retrieved 5 August 2018 from <https://ocw.mit.edu>
- White, M. (2012). Glässers Disease. National Animal Disease Information Service. Retrieved 30 January 2019 from <http://www.nadis.org.uk/disease-a-z/pigs/glaessers-disease/>
- Wu, X., Xiao, L., Wang, Y., Yao, X., & Yang, Z. (2017). Genetic variants and phylogenetic analysis of *Haemophilus parasuis* (HPS) OMPP2 detected in Sichuan, China from 2013 to 2015. *Journal of Veterinary Medical Science*, *79*(10), 1648-1651.
- Xu, C., Zhang, J., Zhao, Z., Guo, L., Zhang, B., Feng, S., Zhang, L., & Liao, M. (2011). Antimicrobial susceptibility and PFGE genotyping of *Haemophilus parasuis* isolates from pigs in South China (2008-2010). *Journal of Veterinary Medical Science*, *73*(8), 1061-1065.
- Xu, Z., Yue, M., Zhou, R., Jin, Q., Fan, Y., Bei, W. & Chen, H. (2011). Genomic characterization of *Haemophilus parasuis* SH0165, a highly virulent strain of serovar 5 prevalent in China. *PLoS ONE*, *6*, e19631.
- Yue, M., Jin, Q., & Chen, H. (2010). Genetic relatedness of *Haemophilus parasuis* among reference strains and Chinese epidemic isolates. Retrieved <https://www.ncbi.nlm.nih.gov/genbank/>
- Yue, M., Yang, F., Yang, J., Bei, W., Cai, X., Chen, L., Dong, J., Zhou, R., Jin, M., Jin, Q., & Chen, H. (2009). Complete genome sequence of *Haemophilus parasuis* SH0165. *Journal of Bacteriology*, *191*(4), 1359-1360.
- Zhang, B., Feng, S., Xu, C., Zhou, S., He, Y., Zhang, L., Zhang, J., Guo, L., & Liao, M. (2011). Serum resistance in *Haemophilus parasuis* SC096 strain requires outer membrane protein P2 expression. *FEMS Microbiology letters*, *326*(2), 109-115.
- Zhang, B., He, Y., Xu, C., Xu, L., Feng, S., Liao, M., & Ren, T. (2012a). Cytolethal distending toxin (CDT) of the *Haemophilus parasuis* SC096

strain contributes to serum resistance and adherence to and invasion of PK-15 and PUVeC cells. *Veterinary Microbiology*, 157(1-2), 237-242.

Zhang, J., Xu, C., Guo, L., Shen, H., Deng, X., Ke, C., Ke, B., Zhang, B., Li, A., Ren, T., & Liao, M. (2012b). Prevalence and characterization of genotypic diversity of *Haemophilus parasuis* isolates from southern China. *Canadian Journal of Veterinary Research*, 76(3), 224-229.

Zhou, M. G., Guo, Y., Zhao, J. P., Hu, Q. Y., Hu, Y., Zhang, A., Chen, H. C., & Jin, M. L. (2009). Identification and characterization of novel immunogenic outer membrane proteins of *Haemophilus parasuis* serovar 5. *Vaccine*, 27, 5271-5277.

Zhou, S., He, X., Xu, C., Zhang, B., Feng, S., Zou, Y., Li, J., & Liao, M. (2014). The outer membrane protein P2 (*OmpP2*) of *Haemophilus parasuis* induces proinflammatory cytokine mRNA expression in porcine alveolar macrophages. *The Veterinary Journal*, 199(3), 461-464.

Zhou, X., Xu, X., Zhao, Y., Chen, P., Zhang, X., Chen, H. & Cai, X. (2010). Distribution of antimicrobial resistance among different serovars of *Haemophilus parasuis* isolates. *Veterinary Microbiology*, 141(1-2), 168-173.

Zhou, M., Zhang, Q., Zhao, J., & Jin, M. (2012). *Haemophilus parasuis* encodes two functional cytolethal distending toxins: CdtC contains an atypical cholesterol recognition/interaction region. *PLoS one*, 7(3), e32580.