



UNIVERSITI PUTRA MALAYSIA

***ISOLATION AND CHARACTERIZATION OF ENDOGLUCANASE
PRODUCED BY MICROBES RESIDING IN THE GUT OF COPTOTERMES
CURVIGNATHUS HOLMGREN TERMITE***

MONICA HII HUNG LING

FSPM 2016 2



**ISOLATION AND CHARACTERIZATION OF ENDOGLUCANASE
PRODUCED BY MICROBES RESIDING IN THE GUT OF *COPTOTERMES*
CURVIGNATHUS HOLMGREN TERMITE**

By
MONICA HII HUNG LING

Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Science

January 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
the requirement for the degree of Master of Science

**ISOLATION AND CHARACTERIZATION OF ENDOGLUCANASE
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January 2016

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This research was carried out to isolate and identify endoglucanase producing microbes from the digestive system of wood termite *Coptotermes curvignathus* Holmgren as well as to characterize endoglucanase produced by these microbes based on their optimum pH, temperature, and enzymatic activity. Five endoglucanases producing bacteria were isolated, four were molecularly identified as aerobic *Bacillus spp.* and the other one was an unknown anaerobic bacterium. Based on Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) analysis, endoglucanases produced by these isolates were similar in molecular size, at 11 kDa. These endoglucanases were relatively smaller than the endoglucanases that is produced in *Reticulitermes speratus* salivary glands, which were reported at 41 kDa and 42 kDa. *Reticulitermes speratus* is phylogenetically close to *Coptotermes curvignathus* Holmgren, and both feed on wood. Identities of the endoglucanase producing bacteria were further confirmed using BIOLOG phenotypic analysis. Isolates TG117 was identified as *Bacillus thuringiensis*, NA45/1 as *B. cereus*, TG111 as *B. pseudomycoides*, and TG005 as *B. mycoides*. Among the five endoglucanases tested, endoglucanase produced by *B. cereus* NA45/1 showed the highest enzymatic activity, 0.40 UmL^{-1} at pH9 and 45°C . Endoglucanase *B. cereus* NA45/1 also had significantly higher enzymatic activity when compared to the commercial cellulase from *Aspergillus niger* (C1184 Sigma). Endoglucanase *B. pseudomycoides* TG111 performed optimally at alkaline condition pH9 and 70°C with enzymatic activity at 0.23 UmL^{-1} . Endoglucanase *B. thuringiensis* TG117 had the highest enzymatic activity at 0.21 UmL^{-1} when acted in an acidic condition, pH5 and at temperature, 40°C . Both isolates *B. mycoides* TG005 and unknown anaerobic ST1 has their maximum enzymatic activity at pH6 and temperature at 55°C . This study showed that *C. curvignathus* Holmgren had a wide range of endoglucanases, where optimum temperature and pH for maximum enzymatic activities varies widely. With this array of endoglucanases, *C. curvignathus* Holmgren that feed mainly on living plant-based diet would be able break down cellulose into oligomers and reducing sugars that will subsequently be broken down to fermentable glucoses to sustain life.

Abstrak tesis yang dikemukakan kepada Senat University Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**PEMENCILAN DAN PENCIRIAN ENDOGLUKANASE YANG DIHASILKAN
OLEH MIKROB YANG TINGGAL DI DALAM USUS ANAI *COPTOTERMES
CURVIGNATHUS HOLMGREN***

Oleh

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Kajian ini dijalankan untuk memencarkan dan mengenalpasti endoglukanase yang dihasilkan oleh mikrob dalam sistem pencernaan anai-anai *Coptotermes curvignathus* Holmgren, serta penciran endoglukanase yang dihasilkan oleh mikrob tersebut berdasarkan pH, suhu dan aktiviti enzim optima masing-masing. Antara lima penciran bakteria yang menghasilkan endoglukanase, empat penciran bakteria dikenalpasti secara molekul sebagai aerobik *Bacillus* spp. dan satu lagi penciran adalah bakteria anaerobik yang belum dikenalpasti. Berdasarkan analisis *Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis* (SDS-PAGE), endoglukanase yang dihasilkan oleh penciran ini mempunyai saiz molekul yang sama iaitu 11 kDa. Saiz endoglukanase ini adalah lebih kecil daripada endoglukanase yang dihasilkan dari kelenjar air liur anai-anai spesis *Reticulitermes speratus*, di mana ia dilaporkan bersaiz 41 kDa dan 42 kDa masing-masing. *R. Speratus* adalah spesis terdekat dengan *C. curvignathus* Holmgren secara filogenetik dan kedua-duanya makan kayu. Identiti endoglukanase yang dihasilkan oleh bakteria ditentukan dengan menggunakan BIOLOG fenotip analisis. Penciran TG117 dikenalpasti sebagai *Bacillus thuringiensis*, NA45/1 sebagai *B. cereus*, TG111 sebagai *B. pseudomycooides*, dan TG005 sebagai *B. mycoides*. Di antara lima endoglukanase yang telah dikaji, endoglukanase yang dihasilkan oleh *B. cereus* NA45/1 menunjukkan aktiviti enzim yang paling tinggi, iaitu 0.40 UmL^{-1} pada pH9 dan suhu 45°C . Endoglukanase *B. cereus* NA45/1 menunjukkan aktiviti enzim yang lebih tinggi daripada selulase komersil daripada *Aspergillus niger* (C1184 Sigma). Manakala, endoglukanase *B. pseudomycooides* TG111 mempunyai aktiviti enzim paling tinggi dalam keadaan beralkali, iaitu 0.23 UmL^{-1} pada pH9 dan suhu 70°C . Endoglukanase *B. thuringiensis* TG117 pula mempunyai aktiviti enzim paling tinggi dalam keadaan berasid, iaitu 0.21 UmL^{-1} pada pH5 dan suhu 40°C . Kedua-dua penciran *B. mycoides* TG005 dan anaerobik yang belum dikenalpasti ST1 mempunyai aktiviti enzim yang maksimum pada pH6 dan pada suhu 55°C . Kajian ini menunjukkan bahawa *C. curvignathus* Holmgren mempunyai pelbagai jenis endoglukanase yang berfungsi pada pH dan suhu yang berbeza. Keadaan ini boleh membolehkan *C. curvignathus* Holmgren untuk mencernakan selulosa kepada oligomer dan gula penurun diikuti dengan seterusnya glukosa untuk kemandirian anai-anai yang hanya bergantung kepada tumbuhan sebagai makanan tunggalnya.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the supervisory committee were as follows:

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LIST OF ABBREVIATIONS

| | |
|-------------------|---|
| A | Alpha |
| ANOVA | Analysis of Variance |
| BLAST | Basic Local Alignment Search Tool |
| B | Beta |
| BSA | Bovine Serum Albumin |
| °C | Degree Celsius |
| CAZy | CArbohydrate-Active EnZymes |
| Cm | Centimeter |
| CMC | Carboxymethyl Cellulose |
| dNTP | Deoxy Nucleotide Triphosphate |
| DNA | Deoxy Ribonucleic Acid |
| GOPOD | Glucose oxidase/ peroxidase |
| M | Molar |
| Mg | Milligram |
| MgCl ₂ | Magnesium Chloride |
| Min | Minute |
| mL | Milliliter |
| Mm | Millimeter |
| µL | Microliter |
| µg | Microgram |
| µmol | Micromole |
| Nm | Nanometer |
| NA | Nutrient Agar |
| NaCl | Sodium Chloride |
| NCBI | National Center for Biotechnology Information |
| Native-PAGE | Non-denaturing Polyacrylamide Gel Electrophoresis |
| pH | Potential hydrogen |
| SDS-PAGE | Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis |
| ST | Special Topic |
| TG | Termite Gut |
| % | Percent |

CHAPTER 1

INTRODUCTION

Termites and its gut microbes are believed to share at least 20 million years of relationship as indicated by fossil evidence (Wier *et al.*, 2002). Hongoh *et al.* (2005) reported that diverse gut bacteria have co-evolved with their host termites and became a stable symbiotic complex. Some members of this symbiotic microbiota are found to be host specific at genus level (Hongoh *et al.*, 2005). Although the gut of termite is very small in volume (1 to 2 μL), it contains a rich community of microbes (10^5 to 3×10^7 cells per mL of termite gut) (Breznak & Leadbetter, 2006; Eutick *et al.*, 1978). These gut microbes have a great impact on termites survival and their specialized diet (Radek, 1999). Cooperation of both termite and its gut symbionts enable a high efficiency in biomass conversion into energy and nutrient needed by both host and microbes (Scharf, 2015; Radek, 1999).

Termite feeds solely on lignocellulosic materials (Ohkuma, 2008). They have high lignocellulose degradation efficiency. This ability is very intriguing since lignocellulose is known to be a very robust material. Thus far, several studies have concluded those microbes that residing in the wood feeding termite can degrade lignocellulose and produce large amounts of hydrogen as intermediate (Pester and Brune, 2007). The ability of lower termites in degrading cellulose is well established partly attributed to its symbiotic protists. Other than their symbiont counterparts, termites are also known to produce their own endoglucanases in their salivary gland (Lo *et al.*, 2011; Khademi *et al.*, 2004). Endoglucanase is a cellulase that hydrolyses 1, 4- β bonds of the cellulose chains (Li *et al.*, 2006). The beta acetyl linkage (β -1, 4 glycosidic bonds) gives rigidity property to the cellulose. In order to fully digest cellulose to glucose, it requires three types of enzymes: (1) Endoglucanase to hydrolyse β -1, 4 bonds of cellulose chains, (2) exoglucanase that cleaves cellobiosyl units on their non-reducing ends, and (3) β -glucosidase cleaves on cellobiosyl and other cello-oligosaccharides to glucose (Li *et al.*, 2006).

Termites are known to be economically important insect pests especially in peat soil oil palm plantations in Malaysia covering 5.01 million hectare where 13.3% constitutes peat soil (Kon *et al.*, 2012). One of the common termite species in oil palm plantations is *Coptotermes curvignathus* Holmgren where it can infest as early as 12 months after field planting and may kill more than 5.3% of the palms in a year (Masijan *et al.*, 2006). This termite is among the very few known termite species that has the ability to feed on plant living tissues without the need of their diets to be predigested by other microbes (Chan *et al.*, 2011). This indicates *C. curvignathus* Holmgren have the ability to achieve effective lignocellulose biomass conversion. This fascinating characteristic was investigated in this study with a focus to isolate and characterize different endoglucanases that can be found in the termite digestive system. The knowledge and findings of these endoglucanases can be used to improve present biomass conversion enzyme technology. Thus, the objectives of this study were to: (1) isolate and identify microbes residing in the gut of *C. curvignathus* Holmgren termites that can produce endoglucanases, (2) characterization of these endoglucanases based on their optimum pH and temperature to give the highest enzymatic activity.

REFERENCES

- Aarthi, N., and Ramana, K. V. (2011). Identification and characterization of polyhydroxybutyrate producing *Bacillus cereus* and *Bacillus mycoides* strains. *International Journal of Environmental Sciences*, 1(5), 744-756
- Adams, L., and Boopathy, R. (2005). Isolation and characterization of enteric bacteria from the hindgut of Formosan termite. *Bioresource Technology*, 96 (14), 1592-1598
- Anggoro, H. P. (2002). A survey of the termites (Insecta: Isoptera) of tabalong district South Kalimantan, Indonesia. *The Raffles Bulletin of Zoology*, 50(1), 117-128
- Araujo, R., Casal, M., and Cavaco-Paulo, A. (2008). Application of enzymes for textile fibres processing. *Biocatalysis and Biotransformation*, 25(5), 332-349
- Bajaj, B. K., Pangotra, H., Wani, M. A., Sharma, P., and Sharma, A. (2009). Partial purification and characterization of a highly thermostable and pH stable endoglucanase from a newly isolated *Bacillus* strain M-9. *Indian Journal of Chemical Technology*, 16, 382-387
- Baker, P. B., Marchosky, R. J., and Yelich, A. J. (2005). Arizona termites of economic importance. <http://ag.arizona.edu/pubs/insects/az1369.pdf> accessed on 20/12/2013
- Beeby, M., O'Connor, B. D., Ryttersgaard, C., Boutz, D. R., Perry, L. J., and Yeates, T. O. (2005). The genomic of disulfide bonding and protein stabilization in thermophiles. *PLOS Biology*, 3(9): e309
- Bhat, M. K. (2000). Cellulases and related enzyme in biotechnology. *Biotechnology Advances*, 18(5), 355-383
- BiOLOG, (2008). GEN III MicroPlateTM. <http://www.biolog.com/pdf/milit/00P%20185rA%20GEN%20III%20MicroPlate%20IFU%20Mar2008.pdf> accessed on 09/04/2013
- Bisswanger, H. (2014). Enzyme assay. *Science Direct*, 2014(1), 41-45
- Bollag, D. M., and Edelstein, S. J. (1990). *Protein Methods*. Wiley, Switzerland. Pp 96-115
- Borror, D. J., Delong, D. M., and Triplehorn, C. A. (1981). *An introduction to the study of insects*, (Fifth edition). Saunders College Publishing, Philadelphia, pp. 140-146
- Borror, D. J., and White, R. E. (1970). *Insects*. Houghton Mifflin, New York. pp. 88-92

- Bourguignon, T., Sobotnik, J., Hanus, R., and Roisin, Y. (2009). Developmental pathways of *Glossotermes oculatus* (Isoptera, Serritermitidae): at the cross-roads of worker caste evolution in termites. *Evolution and Development*, 11(6), 659-668
- Bradford, M. M. (1976). A rapid and sensitive method for quantitation of microgram quantities of protein utilizing the principle of protein-dye-binding. *Analytical Biochemistry*, 72, 248-254
- Breznak, J. A., and Leadbetter, J. R. (2006). Termites gut spirochetes. *Prokaryotes*, 7, 318-329
- Breznak, J. A., and Switzer, J. M. (1986). Acetate synthesis from H₂ plus CO₂ by termite gut microbes. *Applied and Environmental Microbiology*, 52(4), 623-630
- Brugerolle, G., and Radek, R. (2006). Symbiotic protozoa of termites. *Soil Biology*, 1 (6), 243-269
- Brune, A., and Friedrich, M. (2000). Microecology of the termite gut: Structure and function on a microscale. *Ecology and Industrial Microbiology*, 3(3), 263-269
- Brune, A., Enerson, D., and Breznak, J. A. (1995). The termite Gut microflora as an oxygen sink: Microelectrode determination of oxygen and pH gradients in guts of lower and higher termites. *American Society for Microbiology*, 61(7), 2681-2687
- Bull, A. T., Ward, A. C., and Goodfellow, M. (2000). Search and discovery strategies for biotechnology: the paradigm shift. *Microbiology and Molecular Biology Reviews*, 64(3), 573-606
- Cabrera, B. J., and Rust, M. K. (1999). Caste differences in feeding and trophallaxis in the western drywood termite, *Incisitermes minor* (Hagen) (Isoptera, Kalotermitidae). *Insectes Sociaux*, 46(3), 244-249.
- Campbell, N. A., and Reece, J. B. (2005). *Biology*, (Seventh edition). Pearson, New York. Pp 70-73
- Chan, S. P., Bong, C. F. J., and Lau, W. H., (2011). Damage pattern and nesting characteristic of *Coptotermes curvignathus* (Isoptera: Rhinotermitidae) in oil palm on peat. *American Journal of Applied Science*, 8(5), 420-427
- Cheng, J., (2010). Biomass to renewable energy processes. <https://books.google.com.my/books?id=wFvNBQAAQBAJ&pg=PA230&lpg=PA230&dq=endoglucanase+can+be+produce+by?&source=bl&ots=VWD248PE9J&sig=vSRMokgvsKQZdjUxzGSeeVYibeU&hl=en&sa=X&ei=sJ-mVJzWKcaPuASDwYGACg&ved=0CEIQ6AEwBTgK#v=onepage&q=endo+glucanase%20can%20be%20produce%20by%3F&f=false> accessed on 2/1/2015

- Chinedu, S. N., Nwinyi, O. C., Okafor, U. A., and Okochi, V. I., (2011). Kinetic study and characterization of 1,4- β -endoglucanase of *Aspergillus niger* ANL301. *Dynamic Biochemistry, Process Biotechnology and Molecular Biology*, 5(2), 41-46
- Das, A., Ghosh, U., Mohapatra, P. K. D., Pati, B. R., and Mondal, K. C. (2012). Study on thermodynamics and absorption kinetics of purified endoglucanase (CMCASE) from *Penicillium Notatum NCIM No-923* produced under mixed solid-state fermentation of waste cabbage and bagasse. *Brazilian Journal of Microbiology*, 2012, 1103-1111
- Deevong, P., Hattori, S., Yamada, A., Trakulnaleamsai, S., Ohkuma, M., Noparatnaraporn, N., et al. (2004). Isolation and detection of Methanogens from the gut of higher termites. *Microbes and Environments*, 19(3), 221-226
- Delmotte, N., Knief, C., Chaffron, S., Innerebner, G., Roschitzki, B., Schlapbach, R. et al. (2009). Community proteogenomics reveals insights into the physiology of phyllosphere bacteria. *PNAS*, 106(38), 16428-16433
- Dobrev, G. T., and Zhekova B. Y. (2012). Biosynthesis, purification and characterization of endoglucanase from a xylanase producing strain *Aspergillus Niger* B03. *Brazilian Journal of Microbiology*, 43(1), 70-77
- Donovan, S. E., Eggleton, P., and Bignell, D. E. (2001). Gut content analysis and a new feeding group classification of termites. *Ecological Entomology*, 26, 356-366
- Dyer, J. and Grosvenor, A. (2011). Protein fiber surface modification. <http://cdn.intechopen.com/pdfs-wm/23055.pdf> accessed on 2/1/2015
- Eggleton, P. (2006). The termites gut habitat: Its evolution and co-evolution. *Soil Biology*, 6(2), 373-404
- Eggleton, P., Homathevi, R., Jones, D. T., MacDonald, J. A., Jeeva, D., Bignell, D. E. et al. (1999). Termite assemblages, forest disturbance and greenhouse gas fluxes in sabah, East Malaysia. *The Royal Society, Philosophical Transactions: Biological Science*, 354(1391), 1791-1802
- Eggleton, P., Homathevi, R., Jeeva, D., Jones, D. T., Davies, R. G., and Maryati, M. (1997). The species richness and composition of termites (Isoptera) in primary and regenerating lowland dipterocarp forest in Sabah, east Malaysia. *Ecotropica*, 3, 119-128
- Ejaz, M., Qidi, z., Gaisheng, Z., Qunzhu, W., Na, N., and Huiyan, Z. (2014). Comparison of small scale methods for the rapid and efficient extraction of mitochondrial DNA from wheat crop suitable for down-stream processes. *Genetics and Molecular Research*, 13(4), 10320-10331
- El-Sersy, N. A., Abd-Elnaby, H., Abou-Elela, G. M., Ibrahim, H. A. H., and El-Toukhy, N. M. K. (2010). Optimization, economization and characterization of cellulase produced by marine *Streptomyces ruber*. *African Journal of Biotechnology*, 9(38), 6355-6364.

- Eutick, M. L., Obrien, R. W., and Slaytor, M. (1978). Bacteria from the gut of Australian Termites. *Applied and Environmental Microbiology*, 35(5), 823-828
- Forschler, B. T., and Jenkins, T. M. (2000). Subterranean termites in the urban landscape: Understanding their social structure is the key to successfully implementing population management using bait technology. *Urban Ecosystems*, 4(3), 231-251
- Freymann, B. P., Buitenwerf, R., Desouza, O., and Olff, H. (2008). The importance of termites (Isoptera) for the recycling of herbivore dung in tropical ecosystems: a review. *European Journal of Entomology*, 105(2), 165-173
- Friedrich, M. W., Wagner, D. S., Lueders, T., and Brune, A. (2001). Axial differences in community structure of *Crenarchaeota* and *Euryarchaeota* in the highly compartmentalized gut of the soil-feeding termite *Cubitermes orthognathus*. *Applied and Environmental Microbiology*, 67(10), 4880-4890
- Galbe, M., and Zacchi, G. (2012). Pretreatment: The key to efficient utilization of lignocellulosic materials. *Biomass and Bioenergy*, 46, 70-78
- Galkiewicz, J. P., and Kellogg, C. A. (2008). Cross-kingdom amplification using bacteri-specific primers: complications for studies of coral microbial ecology. *Applied and Environmental Microbiology*, 74(24), 7828-7831
- Gold, R. E., Howell Jr., H. N., Glenn, G. J. and Engler, K. M. (2005). Subterranean Termites. <https://insects.tamu.edu/extension/publications/epubs/e-368.cfm> accessed on 20/01/2015
- Gomathi, V., Ramasamy, K., Reddy, M. R. V. P., Ramalakshimi, A., and Ramanathan, A. (2009). Methan emission by gut symbionts of termites. *Academic Journal of Plant Sciences*, 2(3), 189-194
- Gupta, S., Adlakha, N., and Yazdani, A. S. (2013). Efficient extracellular secretion of an endoglucanase and a β -glucosidase in *E.coli*. *Protein Expression and Purification*, 88(1), 20-25.
- Hoe, P. K., Bong, C. F. J., Jugah, K., and Rajan, A. (2009). Evaluation of *Metarhizium anisopliae* var. *anisopliae* (Deuteromycotina: Hyphomycete) isolates and their effects on subterranean termite *Coptotermes curvignathus* (Isoptera: Rhinotermitidae). American Journal of Agricultural and Biological Sciences, 4(4), 289-297
- Hogan, M. E., Schulz, M. W., Slaytor, M., Czolij, R. T., and Obrien, R. W. (1988). Components of termite and protozoal cellulases from the lower termite, *Coptotermes lacteus* froggatt. *Insect Biochemistry*, 18(1), 45-51
- Hongoh, Y. (2010). Diversity and genomes of uncultured microbial symbionts in the termite gut. *Bioscience, Biotechnology and Biochemistry*, 72(6), 1145-1151

- Hongoh, Y., Deevong, P., Inoue, T., Moriya, S., Trakulnaleamsai, S., Okhuma, M., et al. (2005). Intra- and Interspecific Comparisons of bacterial diversity and community structure support coevolution of gut microbiota and termite host. *Applied and Environmental Microbiology*, 71(11), 6590-6599
- Hongoh, Y., Yuzawa, H., Ohkuma, M., and Kudo, T. (2006). Evaluation of primers and PCR conditions for the analysis of the 16S rRNA genes from a natural environment. *FEMS Microbiology Letters*, 221(2), 299-304
- Hu, X. P., and Appel, A. G. (2004). Seasonal variation of critical thermal limits and temperature tolerance in formosan and eastern subterranean termites (Isoptera: Rhinotermitidae). *Environment Entomology*, 33(2), 197-205
- Ibrahim, B. U., and Adeboye, D. A. (2012). Appraisal of the economic activities of termites: A review. *Baybero Journal of Pure and Applied Science*, 5(1), 84-89
- Ito, S., (1997). Alkaline cellulases from alkaliphilic *Bacillus*: Enzymatic properties, genetics and application to detergents. *Extremophiles* 1: 61-66
- Jabasingh, S. A. and Nachiyar, C. V. (2012). Process optimization for the biopolishing of jute fibers with cellulases from *Aspergillus Nidulans* AJ SU04. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 2(1), 12-16
- Jarzemowski, E. A. (1981). An early cretaceous termite from southern England (Isoptera: Hodotermitidae). *Systematic Entomology*, 6(1), 91-99
- Karnchanat, A., Petsom, A., Sangvanich, P., Piapukiew J., Whalley, A. J. S., Reynolds, C. D. et al. (2008). A novel thermostable endoglucanase from the wood-decaying fungus *Daldinia eschscholtzii* (Ehrenb. Fr.) Rehm. *Enzyme and Microbial Technology*, 42, 404-413
- Kasana, R. C., Salwan, R., Dhar, H., Dutt, S. and Gulati, A. (2008). A rapid and easy method for the detection of microbial cellulases on agar plates using gram's iodine. *Current Microbiology*, 57(5), 503-507
- Khademi, S., Guarino, L. A., Watanabe, H., Tokuda, G., and Meyer, E. F. (2004). Structure of an endoglucanase from termite, *Nasutitermes takasagoensis*. *Acta Crystallographica Section D*, 58(4), 653-659
- King, J. H. P., Mahadi, N. M., Bong, C. F. J., Ong, K. H., and Hassan, O. (2013). Bacterial microbiome of *Coptotermes curvignathus* (Isoptera: Rhinotermitidae) reflects the coevolution of species and dietary pattern. *Insect science*, 21(5), 1-13
- Korb, J. (2008). The ecology of social evolution in termites. Springer Berlin Heidelberg, German, 151-174
- Kon, T. W., Bong, C. F., King, J. H. P., and Leong, C. T. S. (2012). Biodiversity of termite (Insecta: Isoptera) in tropical peat land cultivated with oil palms. *Pakistan Journal of Biological Science*. 15(3), 108-120

- Konig, H. (2006). *Bacillus* species in the intestine of termites and other soil invertebrates. *Journal of Applied Microbiology*, 101(3), 620-627
- Kuhad, R. M., Gupta, R., and Singh, A. (2011). Microbial cellulases and their industrial applications. *Enzyme Research*, 2011(2011), 1-10
- Lai, P.Y., Tamashiro, M., Yates, J. R., Su, N. Y., Fujii, J. K. and Ebisu, R. H. (1983). Living plants in Hawaii attacked by *Coptotermes formosanus*. *Proceeding, Hawaiian Entomological Society*, 24 (2-3), 283-286
- Lee, H. V., Hamid, S. B. A., and Zain, S. K. (2014). Conversion of lignocellulosic biomass to nanocellulose: structure and chemical process. *The Scientific World Journal*, 2014 (2014), 1-20
- Li, C., Ladisch, C. M., and Ladisch, M. R., (2001). Pore characterization of cellulase enzyme treated cotton fabric. *Textile Research Journal*, 71(5), 407-414
- Li, L., Frohlich, J., and Konig, H. (2006). Cellulose digestion in the termite gut. *Soil Biology*, 6(2), 221-241
- Lo, N., Engel, M. S., Cameron, S., Nalepa, C. A., Tokuda, G., Grimaldi, D., et al. (2007). Save Isoptera: a comment on inward *et al.* *Biology Letters*, 3(5), 562-563
- Lo, N., Tokuda, G., and Watanabe, H. (2011). Evolution and function of endogenous termite cellulases. *Biology of Termites: a Modern Synthesis*, 51-67
- Masijan, Z., Kamarudin, N., Wahid, M. B., Ali, Z., and Junid, K. N. (2006). Rubstake- Rubber wood state for detecting subterranean termites in peat soil. <http://palmoilis.mpob.gov.my/publications/TOT/TT-308.pdf> accessed on 8/06/2015
- Mathew, G. M., Ju, Y. M., Lai, C. Y., Mathew, D. C., and Huang, C. C. (2012). Microbial community analysis in the termite gut and fungus comb of *Odontotermes formosanus*: the implication of *Bacillus* as mutualists. *FEMS Microbiology Ecology*, 79(2), 504-517
- Matsui, T., Tokuda, G., and Shizato, N. (2009). Termites as functional gene resources. *Recent Patents on Biotechnology*, 3(1), 10-18
- McDonald, A. K., Muegge, M. A., and Sansone, C. (2010). Desert termites *Gnathotermes tubiformans*. *Texas A&M Agrilife Extension*, E-258
- Megazyme, (2011). D-glucose assay procedure (GOPOD-FORMAT) Megazyme International Ireland. https://secure.megazyme.com/files/BOOKLET/R-GLC4_1107_DATA.pdf accessed on 18/06/2013
- Millar, S. (2012). Tips and tricks for the lab: column packing. http://www.chemistryviews.org/details/education/2040151/Tips_and_Tricks_for_the_Lab_Column_Packing.html accessed on 9/07/2012

- Nakamura, L. K. (1998). *Bacillus pseudomycooides* sp. nov. *International Journal of Systematic Bacteriology*. 48(3), 1031-1035
- Nation, L. J. (2002). *Insect physiology and biochemistry*. CRC Press LLC: London, New York. Pp 52-53
- Navya, P. N. and Puspha, S. M. (2013). Production, statistical optimization and application of endoglucanase from *Rhizopus stolonifer* utilizing coffee husk. *Bioprocess Biosystems Engineering*, 36(8), 1115-1123
- Ngugi, D. K., Tsanuo, M. K., and Boga H. I., (2007). Benzoic acid-degrading bacteria from the intestinal tract of *Macrotermes michaelensi* Sjostedt. *Journal of Basic Microbiology*, 47(1), 87-92
- Ohkuma, M. (2008). Symbioses of flagellates and prokaryotes in the gut of lower termites. *Trends in Microbiology*, 16(7), 345-352
- Ohkuma, M., Hongoh, Y., and Kudo, T. (2006). Diversity and molecular analyses of yet-uncultivated microorganisms. *Soil Biology*, 6(2), 303-317
- Ong, L. K. (2004). Conversion of lignocellulosic biomass to fuel ethanol- a brief review. *The Planter, Kuala Lumpur* , 80(941), 517-524
- Organic chemistry (2014), column chromatography
<http://orgchem.colorado.edu/Technique/Procedures/Columnchrom/Columnchrom.html> accessed on 30/9/2014
- Orphan, V. J., Taylor, L. T., Hafenbradl, D., and Delong, E. F. (2000). Culture-dependent and culture-independent characterization of microbial assemblages associated with high-temperature petroleum reservoirs. *Applied and Environmental Microbiology*, 66(2), 700-711
- Pearce, D. A., van der Gast, C. J., Lawley, B., and Ellis-Evans, J. C. (2006). Bacaterioplankton community diversity in a maritime Antarctic lake, determined by culture-dependent and culture-independent techniques. *Microbiology Ecology*, 45(1), 59-70
- Peralta-Yahya, P. P., Zhang, F., del Cardayre, S. B., and Keasling, J.D. (2012). Microbial engineering for the production of advanced biofuels. *Nature*, 488 (7411), 320-328
- Pester, M., and Brune, A. (2007). Hydrogen is the central free intermediate during lignocellulose degradation by termite gut symbionts. *International Society for Microbial Ecology*, 1(6), 551-565
- Plantwise (2014), Plantwise knowledge bank
<http://www.plantwise.org/KnowledgeBank/Datasheet.aspx?dsid=15282> accessed on 28/12/2014

- Pokhrel, B., Bashyal, B., and Magar, R. T. (2014). Production, purification and characterization of cellulase from *Bacillus subtilis* isolated from soil. *European Journal of Biotechnology and Bioscience*, 2(5), 31-37
- Pol, D., Laxman, R. S., and Rao, M. (2012). Purification and biochemical characterization of endoglucanase from *Penicillium pinophilum* MS 20. *Indian Journal of Biochemistry & Biophysics*. 49(3), 189-192
- Potrikus, C. J., and Breznak, J. A. (1976). Nitrogen-fixing *Enterobacter agglomerans* isolated from guts of wood-eating termites. *Applied an Environmental Microbiology*, 33(2), 392-399
- Radek, R. (1999). Flagellates, bacteria, and fungi associated with termites: diversity and function in nutrition-a review. *Ecotropica* 5: 183-196
- Rahman, H., and Tawatao, N. (2003). Isoptera (Termite). <http://www.bbdc.sabah.gov.my/overall/bbdc21/isoptera.pdf> accessed on 02/04/2015
- Raina, A., Park, Y. I., and Gelman, D. (2008). Molting in workers of the Formosan subterranean termite *Coptotermes formosanus*. *Journal of Insect Physiology*, 54(1), 155-161
- Rosenthal, A., Zhang, X., Lucey, K. S., Ottesen, E. A., Trivedi, V., Choi, H. M. T., et al.. (2013). Localizing transcripts to single cells suggests an important role of uncultured delta-proteobacteria in the termite gut hydrogen economy. *PNAS*, 110(40), 16163-16168
- Saddler, J., and Kumar, L. (2013). Special issue from the NSERC bioconversion network workshop: pretreatment and fractionation of biomass for biorefinery/biofuels. *Biotechnology for Biofuels*, 6: 17
- Salle, R. D., Gatesy, J., Wheeler, W., and Grimaldi, D. (1992). DNA sequences from a fossil termite in Oligo-Miocene amber and their phylogenetic implications. *Science*, 257(5078), 1933-1936
- Sami, A. J., Akhtar, M. W., Malik, N. N., and Naz, B. A. (1988). Production of free and substrate-bound cellulases of *Cellulomonas flavigena*. *Enzyme and Microbial Technology*, 10, 626-631
- Sami, A. J., and Shakoori, A. R. (2008). Biochemical characterization of endo-1, 4- β -D-glucanase activity of a green insect pest *Aulacophora foveicollis* (Lucas). *Life Science Journal*, 5 (2), 30-36
- Sandgren, M., Stahlberg, J., Mitchinson, C. (2005). Structural and biochemical studies of GH family 12 cellulases: improved thermal stability, and ligand complexes. *Progress in Biophysics and Molecular Biology*, 89(3), 246-291

- Saqib, A. A. N., Farooq, A., Iqbal, M., Hassan, J. U., Hayat, U., and Baig, S. (2012). A thermostable crude endoglucanase produced by *Aspergillus fumigatus* in a novel solid state fermentation process using isolated free water. *Enzyme Research*, 2012(2012), 1-6
- Scharf, M. E. (2015). Omic research in termites: an overview and a roadmap. *Frontiers in Genetics*, 6(76), 1-19
- Scheffrahn, R. H. (2008). Encyclopedia of entomology. *Biomedical and Life Sciences*, 20, 3737-3747
- Scheffrahn, R. H., and Postle A. (2013). New termite species and newly recorded genus for Australia: *Marginitermes absitus* (Isopter: Kalotermitidae). *Australian Journal of Entomology*, 52(3), 199-205
- Shimada, K., and Maekawa, K. (2010). Changes in endogenous cellulase gene expression levels and reproductive characteristics of primary and secondary reproductives with colony development of the termite *Reticulitermes speratus* (Isoptera: Rhinotermitidae). *Journal of Insect Physiology*, 56(9), 1118-1124
- Singh, S., Moholkar, V. S., and Goyal, A. (2013). Isolation, identification, and characterization of a cellulolytic *Bacillus amyloliquefaciens* Strain SS35 from Rhinoceros Dung. *ISRN Microbiology*, 2013(2013), 1-7
- Slama, K., Sobotnik, J., and Hanus, R. (2007). Respiratory concerts revealed by scanning microrespirography in a termite *Prorhinotermes simples* (Isoptera: Rhinotermitidae). *Journal of Insect Physiology*, 53(4), 295-311
- Slaytor, M., Veivers, P. C., and Lo, N. (1997). Aerobic and anaerobic metabolism in the higher termite *Nasutitermes walkeri* (Hill). *Insect Biochemistry and Molecular Biology*, 27(4), 291-303
- Sobotnik, J., Bourguignon, T., Hanus, R., Weyda, F., and Roisin, Y. (2010). Structure and function of defensive glands in soldiers of *Glossotermes oculatus* (Isoptera: Serritermitidae). *Biological Journal of the Linnean Society*, 99, 839-848
- Sun, Y., and Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource technology*, 83(1), 1-11
- Tai, V., James, E. R., Perlman, S. J., and keeling, P. J. (2013). Single-cell DNA barcoding using sequences from the small subunit rRNA and internal transcribed spacer region identifies new species of *Trichonympha* and *Trichomitopsis* from the hindgut of the termite *Zootermopsis angusticollis*. *PLoS ONE*, 8(3), e58728
- Taylor, K. A., Crosby, B., McGavin, M., and Forsberg, C. W. (1987). Characteristics of the endoglucanase encoded by a *cel* gene from *Bacteroides succinogenes* expressed in *Escherichia coli*. *Applied and Environmental Microbiology*, 53(1), 41-46

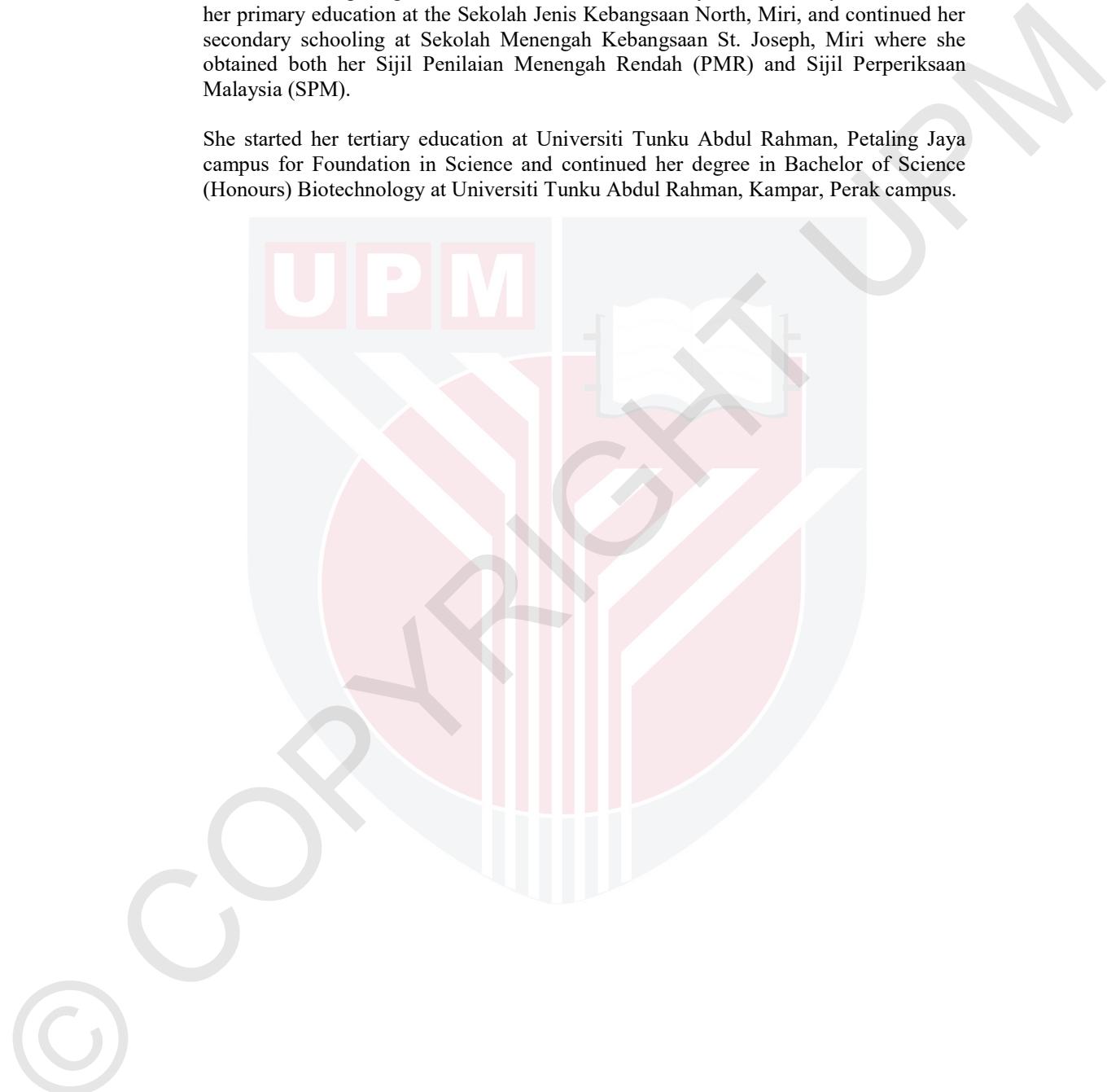
- Teng, Y., Yin, Q., Ding, M., and Zhao, F. (2010). Purification and characterization of a novel endo- β -1,4-glucanase, AFEG22, from the giant snail, *Achatina fulica frussac*. *Acta Biochim Biophys Sin*, 42(10), 729-734
- Thapa, R. S., (1981). Termites of Sabah. Sabah Forest Record Number 12. Sabah Forest Department. Sandakan. pp. 77-80
- Tho, Y. P., (1992). Termites of Peninsular Malaysia. Malaysian Forest Records No. 36, Forest Research Institute Malaysia. Kepong, Kuala Lumpur. pp. 56-61
- Thorne, B. L., (1997). Evolution of eusociality in termites. *Annual Review of Ecology and Systematics*, 28(1), 27-54
- Thorne, B. L., and Carpenter, J. M. (1992). Phylogeny of the Dictyoptera. *Systematic Entomology*, 17, 253-268
- Thorne, B. L., and Haverty, M. I. (1989). Accurate identification of Zootermopsis species (Isoptera: Termopsidae) based on a mandibular character of nonsoldier castes. *Entomological Society of America*, 82(3), 262-266
- Thongaram, T., Hongoh, Y., Kosono, S., Ohkuma, M., Trakulnaleamsai, S., Noparatnaraporn, N., and kudo, T. (2005). Comparison of bacterial communities in the alkaline gut segment among various species of higher termites. *Extremophiles*, 9(3), 229-238
- Tokuda, G., Watanabe, H., Matsumoto T., and Noda, H. (1997). Cellulose digestion in the wood-eating higher termite, *Nasutitermes takasagoensis* (Shiraki): distribution of cellulases and properties of endo- β -1, 4-glucanase. *Zoological Science*, 14(1), 83-93
- Trivedi, D. T. P. (2007). Applied entomology insecticidal methods of pest control. <http://nsdl.niscair.res.in/jspui/bitstream/123456789/457/1/TERMITES%20-%20Formatted.pdf> accessed on 02/04/2015
- Turner, J. S. (2010). Termites as models of swarm cognition. *Swarm Intelligence*. 5(1), 19-43
- Veivers, P. C., Musca, A. M., Obrien, R. W., and Slaytor, M. (1982). Digestive enzymes of the salivary glands and gut of *Mastotermes darwiniensis*. *Insect Biochemistry*, 12(1), 35-40
- Voytas, D. (2001). Agarose gel electrophoresis. *Current Protocols in Immunology*. doi: 10.1002/0471142735
- Walter, P., Metzger, J., Thiel, C., and Helms, V. (2013). Predicting where small molecules bind at protein-protein interfaces. *PLoS ONE*, 8(3), e58583

- Watanabe, H., Nakamura, M., Tokuda, G., Yamaoka, I., Scrivener, A. M., and Noda, H. (1997). Site of secretion and properties of endogenous endo- β -1,4-glucanase components from *Reticulitermes speratus* (Kolbe), a Japanese subterranean termite. *Insect Biochemistry and Molecular Biology*, 27(4), 305-313
- Wenzel, M., Schonig, I., Berchtold, M., Kampfer, P., and König, H. (2002). Aerobic and facultatively anaerobic endoglucanase bacteria from the gut of the termite *Zootermopsis angusticollis*, *Journal of Applied Microbiology*, 92(1) 32-40
- Wier, A., Dolan, M., Grimaldi, D., Guerrero, R., Wagensberg, J., and Margulis, L. (2002). Spirochete and protist symbionts of a termite (*Mastotermes electrodominicus*) in Miocene amber. *PNAS*, 99(3), 1410-1413
- Willis, J. D., Oppert, C., and Jurat-Fuentes, J. L. (2010). Methods for discovery and characterization of cellulolytic enzymes from insects. *Insect Science*, 17(3), 184-189
- Yennamalli, R. M., Rader, A. J., Kenny, A. J., Wolt, J. D. and Sen, T. Z. (2013). Endoglucanases: insights into thermostability for biofuel applications. *Biotechnology for Biofuels*, 6(1): 136
- Yennamalli, R. M., Rader, A. J., Wolt, J. D. and Sen, T. Z. (2011). Thermostability in endoglucanases is fold-specific. *BioMed Central Structural Biology*, 11, 1-15

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