



**UNIVERSITI PUTRA MALAYSIA**

***ISOLATION OF BENEFICIAL BACTERIA AND FEED INCORPORATED  
WITH BENEFICIAL BACTERIA ON GROWTH PERFORMANCE OF  
ASIAN SEA BASS (*Lates calcarifer*, BLOCH) FINGERLING***

**WENDY FERRINA TARRY**

**FSPM 2015 6**



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By

**WENDY FERRINA ANAK TARRY**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the Degree of Master of Science**

**January 2015**

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Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**ISOLATION OF BENEFICIAL BACTERIA AND FEED INCORPORATED WITH BENEFICIAL BACTERIA ON GROWTH PERFORMANCE OF ASIAN SEA BASS (*Lates calcarifer*, BLOCH) FINGERLING**

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**January 2015**

**Chair: Abu Hena Mustafa Kamal, PhD**

**Faculty: Agriculture and Food Sciences (Bintulu)**

The intestinal tract of healthy animals is assumed to be the natural place of 'good' micro-organisms. The present study investigated different parts of intestine of Asian sea bass (*Lates calcarifer*, Bloch) to isolate and identify strains potentially useful for fish probiotics. *In vivo* test was done to validate the performance of isolated *Enterobacter ludwigii* potential probiotic to be used in sea bass fry rearing in aquaculture industry. Five Randomized Complete Block Design (RCBD) treatments for different concentration of *E. ludwigii* were mixed with sea bass feed, i.e., T1 (control; without *E. ludwigii*), T2 ( $1 \times 10^1$  cfu/g of *E. ludwigii*), T3 ( $1 \times 10^3$  cfu/g of *E. ludwigii*), T4 ( $1 \times 10^6$  cfu/g of *E. ludwigii*), T5 ( $1 \times 10^9$  cfu/g of *E. ludwigii*) and fed to the sea bass fry for 28 days. The total culturable aerobic gut bacteria of both live and dead sea bass samples ranged from  $1.17-84.00 \times 10^6$  cfu/g, with counts being higher in posterior intestine ( $1.97-84.00 \times 10^6$  cfu/g) compared to the number of cells occurring in anterior ( $0.21-7.87 \times 10^6$  cfu/g) and middle ( $1.17-3.50 \times 10^6$  cfu/g) parts, although significantly ( $p < 0.05$ ) higher numbers were associated with live fish. The isolate MS32 was selected that inhibit the growth of fish pathogens *Vibrio parahaemolyticus* and *Aeromonas hydrophilla* in disc diffusion, well diffusion assay and cross streak. The isolate MS32 produced gamma-hemolysin and was identified by standard biochemical tests and 16S rDNA sequences as *E. ludwigii*. The species *E. ludwigii* can be grouped as moderately halophile marine bacteria. Storage at 4°C is suitable to store sea bass feed pellet that have been mixed with *E. ludwigii* and can be used for about 20 days.

*In vitro* test showed that *E. ludwigii* formed a clear inhibition zone against 3 fish pathogens at concentration level of  $1 \times 10^9$  cfu/ml via well and disc diffusion method. Presence of *E. ludwigii* in feed was safe to be used to the sea bass fry where no significant difference ( $p > 0.05$ ) among treatments and control. *E. ludwigii* did not improved the growth or increased appetite of sea bass fry with no significant difference ( $p > 0.05$ ) on the growth performance of sea bass fry. The number of *E. ludwigii* in the gastrointestinal tract of sea bass fry and each water sample were also not significantly different ( $p > 0.05$ ). Significant difference ( $p < 0.05$ ) in nitrate, ammonium and phosphate concentrations of rearing water of sea bass fry was observed within the treatments. The concentration of nitrate range from 0.0012–0.0022 mg/L, phosphate concentration from 0.032–0.045 mg/L and ammonium concentrations from 0.981–1.457 mg/L. The presence of *E. ludwigii* can help to improve the water quality compared to control group. Observation after 7 days of challenge test against *V. parahaemolyticus* ( $2 \times 10^8$  cfu/ml) showed that the highest survival percentage (100%) was found for T5 compared to T1 (79%). The findings of the study revealed that *E. ludwigii* could be a potential probiotics for rearing of sea bass fry where improvement of water quality in the rearing tank of sea bass fry and protection against fish pathogen were remarkable.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGASINGAN BAKTERIA BERFAEDAH DAN PENGATURAN MAKANAN  
DENGAN BAKTERIA BAIK TERHADAP PRESTASI PERTUMBUHAN  
BENIH SIAKAP ASIA (*Lates calcarifer*, BLOCH)**

Oleh

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**Januari 2015**

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Saluran perut dan usus haiwan yang sihat adalah tempat semulajadi bagi mikroorganisma baik. Kajian ini adalah untuk mengkaji bahagian yang berbeza pada saluran perut dan usus ikan Siakap (*Lates calcarifer*, Bloch) untuk pengasingan dan mengenalpasti strain yang berpotensi menjadi probiotik yang berguna kepada ikan. Kajian *in vivo* telah dijalankan untuk memastikan strain *Enterobacter ludwigii* sebagai probiotik yang berpotensi untuk digunakan dalam penternakan benih siakap dalam industri akuakultur. Lima rawatan secara 'Randomized Complete Block Design (RCBD)' dengan kepekatan *E. ludwigii* yang berbeza telah dicampur dengan makanan siakap, T1 (kawalan; tanpa *E. ludwigii*), T2 (*E. ludwigii* dengan  $1 \times 10^1$  cfu/g), T3 (*E. ludwigii* dengan  $1 \times 10^3$  cfu/g), T4 (*E. ludwigii* dengan  $1 \times 10^6$  cfu/g), T5 (*E. ludwigii* dengan  $1 \times 10^9$  cfu/g) dan diberikan kepada benih siakap selama 28 hari. Jumlah bakteria aerobik perut dan usus yang terdapat pada ikan segar dan ikan mati adalah dalam lingkungan  $1.17-84.00 \times 10^6$  cfu/g, dimana kiraan yang tertinggi adalah pada bahagian posterior usus ( $1.97-84.00 \times 10^6$  cfu/g) berbanding kiraan pada anterior usus ( $0.21-7.87 \times 10^6$  cfu/g) dan bahagian tengah ( $1.17-3.50 \times 10^6$  cfu/g), dimana dengan perbezaan ketara ( $p < 0.05$ ) kiraan yang tertinggi adalah pada ikan segar. Strain MS32 dipilih dimana dapat menghalang pertumbuhan patogen ikan *Vibrio parahaemolyticus* dan *Aeromonas hydrophilla* melalui cakera serapan, serapan telaga-agar and goresan melintang. MS32 membentuk gamma-hemolisis dan dikenalpasti sebagai *E. ludwigii* menerusi ujian biokimia dan 16S rDNA. Spesies *E. ludwigii* boleh dikelaskan kepada bakteria sederhana 'halophile' marin. Pelet siakap yang telah dicampurkan dengan *E. ludwigii* sesuai disimpan pada suhu 4°C dan tempoh penggunaannya selama 20 hari sahaja.

Kajian *in vitro*, *E. ludwigii* telah membentuk zon perencatan pertumbuhan terhadap 3 patogen ikan pada kepekatan  $1 \times 10^9$  cfu/ml melalui kaedah cakera serapan dan serapan telaga-agar. Kehadiran *E. ludwigii* dalam makanan adalah selamat digunakan untuk benih siakap dimana tiada perbezaan ketara ( $p > 0.05$ ) diantara kumpulan rawatan dan kawalan. *E. ludwigii* tidak meningkatkan prestasi pertumbuhan atau selera benih siakap dimana tiada perbezaan ketara ( $p > 0.05$ ) bagi prestasi pertumbuhan benih siakap. Pengiraan *E. ludwigii* pada saluran perut dan usus benih siakap serta sampel air masing-masing adalah tiada perbezaan ketara ( $p > 0.05$ ). Perbezaan ketara ( $p < 0.05$ ) didapati pada kepekatan nitrat, ammonium dan fosfat dalam sampel air rawatan yang digunakan untuk memternak benih siakap. Kepekatan nitrat adalah di antara 0.0012–0.0022 mg/L, fosfat di antara 0.032–0.045 mg/L dan ammonium antara 0.981–1.457 mg/L. Kehadiran *E. ludwigii* dapat membantu memperbaiki kualiti air jika dibandingkan dengan rawatan kawalan. Pemerhatian selama 7 hari semasa ujian cabaran terhadap *V. parahaemolyticus* ( $2 \times 10^8$  cfu/ml) menunjukkan kadar yang hidup adalah tinggi (100%) pada T5 berbanding dengan T1 (79%). Penemuan kajian ini mengesahkan bahawa *E. ludwigii* boleh menjadi probiotik yang berpotensi untuk membantu dalam penternakan benih siakap dimana peningkatan kualiti air dalam tangki ternakan benih siakap dan perlindungan daripada patogen ikan telah diperhatikan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

RCBD	-	Randomized complete block design
SCT	-	Salinity Conductivity and Temperature meter
TDS	-	Total dissolved solid
SGR	-	Specific growth rate
HIS	-	Hepatosomatic index
VSI	-	Verosomatic index
CI	-	Condition index
FCR	-	Feed conversion rate
PI	-	Protein intake
PER	-	Protein efficiency rate
DO	-	Dissolved oxygen
OD	-	Optical density
MA	-	Marine agar
MB	-	Marine broth
TCBS	-	Thiosulphate citrate bile sucrose
MSA	-	Market siakap anterior
MSM	-	Market siakap middle
MSP	-	Market siakap posterior
LSA	-	Live siakap anterior
LSM	-	Live siakap middle
LSP	-	Live siakap posterior
EDTA	-	Ethylene diamine tetraacetic acid

## CHAPTER 1

### INTRODUCTION

Sea bass is a commercially important coastal, estuarine and freshwater farmed species in the Indo-Pacific region. The culture of this species has extended recently in North America and Europe (Katersky and Carter, 2007). This fish is popular for its delicately-flavored flesh and high market price. It has also fast growth rate and grows to a larger size, thus making it very suitable for aquaculture (Cheong, 1990). In recent years, the culture of sea bass both in pond and cage are in moribund stage due to feeding cost and feed available in the Southeast Asia.

Besides, disease outbreak especially the bacterial diseases has lead to the economic losses in the fish farming (Marzouk *et al.*, 2008). Disease can cause fish mortality and in other hand increase the expenses for treatment, postponement or loss opportunity to sell the fish (Adedeji *et al.*, 2011). The usage of antibiotics to control disease can affect human health and also environment (Birkbeck, 2004). On the other hand, the natural stocks of this fish species have drastically reduced due to natural and manmade catastrophes, degradation of aquatic environment due to climate change and the reduction of many wetlands and water areas in Asia including Malaysia. These factors have created a serious problem to their breeding, migration route, availability and stock, and thus assume this fish become gradually taking place in the list of endangered species.

According to IUCN, there is still data deficiency for this fishery species all over the world. In order to maintain this fish population as well as to conserve their diversity, development of suitable feed component for the rearing and growing of sea bass is very essential both in hatchery, nursery and pond systems. There are also small numbers of sea bass seed from local market with low quality. This may hinder the sustainable production of marine fish culture including sea bass in Malaysia. The use of trash fish as fish feed leads to the breakage of diseases. However, it is difficult to change to pellet where the price is expensive and not readily available compared to trash fish. The mass mortality is also noticed, which is related to the water quality and oxygen depletion elsewhere (Othman, 2008). Studies have been conducted on nutrition, carbohydrate and lipid variation on growth and protein synthesis, feeding frequency on the growth for sea bass (Cuzon *et al.*, 1989; Catacutan and Coloso, 1997; Katersky and Carter, 2007; Biswas *et al.*, 2011) elsewhere, however, no systematic information are available on the diets with bacterial products of this important fishery.

Several studies are conducted on the important roles of probiotics in aquaculture related to the productivity and nutrition of cultured animals, modification of host-associated/ambient microbial community and prevention of diseases and/or improvement of water quality (Verschuere *et al.*, 2000; Wang *et al.*, 2008; Abu Hena *et al.*, 2011). In general, probiotics is harmless to the host and also human being while improved disease resistance for host against diseases. The usage of probiotics is an eco-

friendly alternative measure for sustainable aquaculture (Sihag and Sharma, 2012). Probiotics can help to increase the resistance of host against pathogen, improve their digestion and absorption, and improve feed nutrition as well as maintaining and improving water quality in culture system (Havenaar and Huis, 1992; Gatesoupe, 1999; Verschuere *et al.*, 2000; Cruz *et al.*, 2012).

The isolation and screening of the potential probiotics was done from the gastrointestinal tract of the juvenile sea bass where usually, fish gastrointestinal tract have a high and wide population diversity of microbiota that is mainly dominated by bacteria (Spanggaard *et al.*, 2001; Pond *et al.*, 2006; Denev *et al.*, 2009). The intestinal microbiota is very important since they have their own and specific metabolic, trophic and protective function (Denev *et al.*, 2009; Guarner and Malagelada, 2003). Studies revealed that bacterial products such as probiotics could be useful for both the food and biological control agents of fish disease and activities on the rates of nutrient regeneration for culture organisms (Yasuda and Taga, 1980), hence this study is initiated to investigate on marketed and wild live sea bass from Sarawak region.

The suitable doses of the potential probiotics for the usage towards sea bass culture are an important factor that will influence their effectiveness. A consideration for an appropriate dosage level should be known because it depends on the probiotics species, host fish species, rearing conditions and specific goal of feeding application (Merrifield *et al.*, 2010). The feeding trial by potential probiotics was conducted using sea bass fry since their gastrointestinal tract is still sterile and not fully developed. Early development stages are suitable to apply probiotic treatments since they are highly exposed to gastrointestinal microbiota-associated disorder, because they still have incomplete immune system and their digestive tract is not fully developed (Timmermans, 1987; Vadstein, 1997; Gatesoupe, 1999).

## **Objectives**

The objectives of the study are;

- 1) to isolate the beneficial bacteria (probiotics) from the gastrointestinal tracts of marketed and live sea bass, *Lates calcarifer* (Bloch) and;
- 2) to observe the growth performance and survival of sea bass fry fed with isolated potential beneficial bacteria strain (*E. ludwigii*).

## REFERENCES

- Abu Hena, M.K., Wong, S.K., Idris, M.H. and Aftab Uddin, S. (2011). Pond health management of black tiger shrimp *Penaeus monodon* Fabricius using bacterial products. *Research Journal of Fishery and Hydrobiology*. 6(1):17-21.
- Adedeji, O. B., Adebisi, T. and Emikpe, B. O. (2011). Bacteria load on the skin and stomach of *Clarias gariepinus* and *Oreochromis niloticus* from Ibadan, South West Nigeria: Public health implication. *Journal of Microbiology and Biotechnology Research*. 1(1):52-59.
- Ai, Q., Xu, H., Mai, K., Xu, W. and Wang, J. (2011). Effects of dietary supplementation of *Bacillus subtilis* and fructo-oligosaccharide on growth performance, survival, non-specific immune response and disease resistance of juvenile large yellow croaker, *Larimichthys crocea*. *Aquaculture*. 317:155-161.
- Ajitha, S., Sridhar, M., Sridhar, N., Singh, I.S.B. and Varghese, V. (2004). Probiotics effects of lactic acid bacteria against *Vibrio alginolyticus* in *Penaeus* (Fenneropenaeus) *Indicus* (H. Milne Edwards). *Asian Fisheries Science*. 17: 71-80.
- Al-Dohail, M.A, Hashim, R. and Aliyu-Paiko, M. (2009). Effects of the probiotic, *Lactobacillus acidophilus*, on the growth performance, haematology parameters and immunoglobulin concentration in African Catfish (*Clarias gariepinus*, Burchell 1822) fingerling. *Aquatic Research*. 40:1642-52.
- Al-Harbi, A.H. and Uddin, M.N. (2005). Microbiological quality changes in the intestine of hybrid tilapia (*Oreochromis niloticus* x *Oreochromis aureus*) in fresh and frozen storage condition. *Letters in Applied Microbiology*. 40:486-490.
- Al-Harbi, A.H. and Uddin, M.N. (2012). Bacterial content in the intestine of frozen common carp *Cyprinus carpio*. *African Journal of Biotechnology*. 11(30): 7751-7755.
- Ali, H.M., Yussoff, N.H.N., Om, A.D., Musa, C.U.C. and Othman, M.F. (2008). *Status and prospects of grouper aquaculture in Malaysia*. In *The Aquaculture of Grouper* (pp. 155-175). Liao, I. C. and Leano, E. M (Ed.). Asian Fisheries Society, World Aquaculture Society, Fish Soc. Taiwan and National Taiwan Ocean University.
- Aly, S.M., Abd-El-Rahman, A.M., John, G. and Mohamed, M.F. (2008). Characterization of some bacteria isolated from *Oreochromis niloticus* and their potential use as probiotics. *Aquaculture*. 277:1-6.
- Andani, H.R.R., Tukmechi, A., Meshkini, S. and Sheikhzadeh, N. (2012). Antagonistic activity of two potential probiotic bacteria from fish intestines and investigation of their effects on growth performance and immune response in rainbow trout (*Oncorhynchus mykiss*). *Journal of Applied Ichthyology*. 1-7.

- Austin, B., Stuckey, L.F., Robertson, P.A.W., Effendi, J. and Griffith, D.R.W. (1995). A probiotic strain of *Vibrio alginolyticus* effective in reducing diseases caused by *Aeromonas salmonicida*, *Vibrio anguillarum* and *Vibrio ordalii*. *Journal of Fish Diseases*. 18(1):93-96.
- Austin, B. and Austin, D.A. (1999). Bacterial fish pathogens: *Disease of farmed and wild fish*, 3<sup>rd</sup> (revised) Edn. Godalming, UK: Springer Praxis.
- Austin, B. and Austin, D.A. (2012). Bacterial Fish Pathogens: Disease in Farmed and Wild Fish, 5th Edn. Springer, Dordrecht.
- Axelsson, L. (1998). Lactic acid bacteria: Classification and physiology. In S. Salminen and A. Von Wright. *Lactic Acid Bacteria. Microbiology and Functional Aspects*. (pp. 1-72). Marcel Dekker, New York, NY, USA.
- Ayoola, S.O., Ajani, E.K. and Fashae, O.F. (2013). Effect of probiotics (*Lactobacillus* and *Bifidobacterium*) on growth performance and hematological profile of *Clarias gariepinus* juveniles. *World Journal of Fish and Marine Sciences*. 5(1):01-08.
- Bairagi, A., Ghosh, K.S., Sen, S.K. and Ray, A.K. (2002). Enzyme producing bacterial flora isolated from fish digestive tracts. *Aquaculture International*. 10:109-121.
- Balcázar, J.L., de Blas, I., Ruiz-Zarzuola, I., Cunningham, D.J.L. Vendrell, and Múzquiz, D. (2006). The role of probiotics in aquaculture. *Veterinary Microbiology*. 114:173–186.
- Barlow, C., Williams, K. and Rimmer, M. (1996). Sea bass culture in Australia. *Infofish International*. 2:26-33.
- Baskar, V.P. and Kannan, S. (2009). Marine bacteria as probiotics to control pathogenic *Vibrio* on infected shrimp. *Academic Review*. 16(1):77-85.
- Bergh, O., Naas, K.E. and Harboe, T. (1994). Shift in the intestinal microflora of Atlantic halibut (*Hippoglossus hippoglossus*) larvae during first feeding. *Canadian Journal of Fish Aquatic Sciences*. 51:1899–1903.
- Bernard, V.H., Nurhidayu, A., Ina-Salwany, M.Y. and Abdelhadi, Y. (2012). *Bacillus cereus*; JAQ04 Strain as a potential probiotic for red tilapia; *Oreochromis* species. *Asian Journal of Animal and Veterinary Advances*. 1-6.
- Bimboim, H.C. and Doly, J. (1979). A rapid alkaline extraction procedure for screening recombinant plasmid DNA. *Nucleic Acids Research*. 7(6):1513-1524.
- Birkbeck, T.H. (2004). Current trends in the study of bacterial and viral fish and shrimp diseases - Role of probiotics in fish disease prevention. *Molecular Aspects of Fish and Marine Biology*. 3(14):390-409.
- Biswas, G., Thirunavukkarasu, A.R., Sundaray, J.K. and Kailasam, M. (2011). Culture of Asian seabass *Lates calcarifer* (Bloch) in brackishwater tide-fed ponds:

growth and condition factor based on length and weight under two feeding systems. *Indian Journal Fisheries*. 58(2):53-57.

- Bourouni, O. C., El Bour, M., Calo-Mata, P. and Barros-Velazquez, J. (2012). Antimicrobial Resistance and Potential Probiotic Application of *Enterococcus* spp. in sea bass and sea bream Aquaculture. In P. Marina. *Antibiotic Resistant Bacteria—A Continuous Challenge in the New Millennium*. (pp. 514-525). InTech Europe.
- Bucio, A.G., Hartemink, R., Schrama, J.W., Verreth, J., Bucio, J.G. and Zwietering, M.H. (2009). Kinetics of *Lactobacillus plantarum* 44a in the faeces of tilapia (*Oreochromis niloticus*) after its intake in feed. *Journal of Applied Microbiology*. 107:1967-1975.
- Buntin, N., Chanthachum, S. and Hongpattarakere, T. (2008). Screening of lactic acid bacteria from gastrointestinal tracts of marine fish for their potential use as probiotics. *Songklanakarin Journal of Science and Technology*. 30:141-148.
- Burr, G., Gatlin, III, D.M. and Ricke, S. (2005). Microbial ecology of the gastrointestinal tract of fish and the potential application of prebiotics and probiotics in finfish aquaculture. *Journal of the World Aquaculture Society*. 36:425-436.
- Burbank, D.R., Shah, D.H., LaPatra, S.E., Fornshell, G. and Cain, K.D. (2011). Enhanced resistance to coldwater disease following feeding of probiotic bacterial strains to rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*. 321: 185-190.
- Bromage, E. and Owens, L. (2009). Environmental factors affecting the susceptibility of barramundi to *Streptococcus iniae*. *Aquaculture*. 290:224-228.
- Browdy, C. (1998). Recent developments in penaeid broodstock and seed production technologies: improving the outlook for superior captive stocks. *Aquaculture*. 164:3-21.
- Caipang, C. M., Brinchmann, M. F. and Kiron, V. (2010). Antagonistic activity of bacterial isolates from intestinal microbiota of Atlantic cod, *Gadus morhua*, and an investigation of their immunomodulatory capabilities. *Aquaculture Research*. 41:249-256.
- Capkin, E. and Altinok, I. (2009). Effects of dietary probiotic supplementations on prevention/treatment of yersiniosis disease. *Journal of Applied Microbiology*. 106:1147-1153.
- Cappuccino, J. G. and Sherman, N. (1987). *Microbiology A Laboratory Manual*. Second Edition (pp. 75-80). The Benjamin/Cummings Publishing Company, Inc.
- Carlucci, A.F. and Pramer, D. (1959). Factors affecting the survival of *Escherichia coli* in seawater. *Applied Microbiology*. 7:388-392.



- Catacutan, M. and Coloso, R. (1997). Effect of dietary protein to energy ratios on growth, survival and body composition of juvenile Asian sea bass *Lates calcarifer*. *Aquaculture*. 131:125-133.
- Chabrillón, M., Rico, R.M., Balebona, M.C. and Moríñigo, M.A. (2005). Adhesion to sole, *Solea senegalensis* Kaup, mucus of microorganisms isolated from farmed fish, and their interaction with *Photobacterium damsela* subsp. *piscicida*. *Journal of Fish Diseases*. 28:229-237.
- Chang, C.I., Liu, W.Y., Shyu, C.Z. (2000). Use of prawn blood agar hemolysis to screen for bacteria pathogenic to cultured tiger prawns *Penaeus monodon*. *Diseases of Aquatic Organisms*. 43:153-157.
- Cheng, W. and Chen, J.C. (1999). Effect of cultivation broth pH, temperature and NaCl concentration on virulence of an *Enterococcus*-like bacterium to the giant freshwater prawn *Macrobrachium rosenbergii*. *Disease of Aquatic Organisms*. 36:233-237.
- Cheong, L. (1990). Status of knowledge on farming of sea bass (*Lates calcarifer*) in South East Asia. In: *Advances in Tropical Aquaculture Workshop* held at Tahiti, French Polynesia. 20 February-4 March, 1989, J. Barret, J. Calvas, G. Cuzon, J. Fuchs and M. Weppe (Ed.); *Aquacop*, 9:421-428.
- Chong, Y.C. and Chao, T.M. (1986). Common diseases of marine food fish. *Fish Handbook No. 2* (pp. 34). Primary Production Department, Ministry of National Development, Republic of Singapore.
- Chythanya, R., Karunasagar, I. and Karunasagar, I. (2002). Inhibition of shrimp pathogenic *Vibrios* by a marine *Pseudomonas* 12 strain. *Aquaculture*. 208:1-10.
- Crab, R., Avnimelech, Y., Defoirdt, T., Bossier, P. and Verstraete, W. (2007). Nitrogen removal techniques in aquaculture for a sustainable production. *Aquaculture*. 270 (1-4): 1-14.
- Cross, M.L. (2002). Microbes versus microbes: immune signals generated by probiotic lactobacilli and their role in protection against microbial pathogens. *Immunology and Medical Microbiology*. 34:245-253.
- Cruz, P.M., Ibáñez, A.L., Hermosillo, O.A.M. and Saad, H.C.R. (2012). Use of probiotics in aquaculture. International Scholarly Research Network.
- Cuzon, G., Chou, R. and Fuchs, J. (1989). Nutrition of the seabass *Lates calcarifer*. In: J. Barret, J. Calvas, G. Cuzon, J. Fuchs and M. Weppe (eds.), *Advance in Tropical Aquaculture*, *Aquacop*, 20 February-4 March, 1989. Tahiti, pp. 757-763.
- Dahiya, T.P. and Sihag, R.C. (2009). Incidence of vibriosis in the Indian magur (*Clarias batrachus* L.) in saline water ponds of Haryana-A new report from India. *The Ecotechnology*. 1:116-121.



- De, G.K. (1971). On the biology of post-larval and juvenile stages of *Lates calcarifer* Bloch. *Journal of the Indian Fisheries Association*. 1:51-64.
- De, B. C, Meena, D. K., Behera, B. K., Pronob, D., Mohapatra, P. K. D. and Sharma, A. P. (2014). Probiotics in fish and shellfish culture: immune modulatory and ecophysiological responses. *Fish Physiological Biochemical*. 40: 921-971.
- Denev, S., Staykov, Y., Moutafchieva, R. and Beev, G. (2009). Microbial ecology of the gastrointestinal tract of fish and the potential application of probiotics and prebiotics in finfish aquaculture. *International Aquatic Research*. 1:1-29.
- Decamp, O. and Moriarty. D. (2007). Aquaculture species profit from probiotics. *Feed Mix*. 15(1):20-23.
- Dhevandran, K. and Annie, K. (1999). Antibiotic and L-asparaginase of Streptomyces isolated from fish, shellfish, and sediments of veli estuarine along Kerala coast. *Indian Journal Marine Science*. 28:335-337.
- Divya. K.R, isamma, A., Ramasubramanian, V., Sureshkumat, S. and Arunjith, T.S. (2012). Colonization of probiotic bacteria and its impact on ornamental fish *Puntius conchoniis*. *Journal of Environmental Biology*. 33: 551-555.
- Dopazo, C., Lemos, M., Lodeiros, C., Bolinches, J., Barja, J. and Toranzo, A. (1988). Inhibitory activity of antibiotic-producing marine bacteria against fish pathogens. *Journal of Applied Bacteriology*. 65:97-101.
- El-Dakar, A.Y., Shalaby, S.M. and Saoud, I.P. (2007). Assessing the use of a dietary probiotic/prebiotic as an enhancer of spinefoot rabbitfish *Siganus rivulatus* survival and growth. *Aquaculture Nutrition*. 13:407-412.
- El-Gohary, M. S. and Diab, A. M. (2014). Some Studies on the Effect of Protexin on Immune Status of Cultured Seabass Fingerlings. *Alexandria Journal of Veterinary Sciences*. 41:109-119.
- El-Haroun, E.R., Goda, A.M.A.S. and Kabir Chowdhury, M.A. (2006). Effect of dietary probiotics biogens supplementation as a growth promoter on growth performance and feed utilization of Nile tilapia *Oreochromis niloticus* (L.). *Aquaculture*. 37:1473-1480.
- Esteban, M.A., Cuesta, A., Ortuno, J. and Meseguer, J. (2001). Immunomodulatory effects of dietary intake of chitin on gilthead sea bream (*Sparus aurata* L.) innate immune system. *Fish and Shellfish Immunology*. 11:303-315.
- Evelyn, T.P.T. (1996). Infection and disease. In: Iwama, G., Nakanishi, T. (Ed.), *The Immune System: Organism, Pathogen and Environment* (pp 339-366). Fish Physiological. Series 15, Academic Press, San Diego, CA, USA.
- Far, H.Z., Che Roos, S., Daud, H.M., Harmin, S.A. and Shakibazadeh, S. (2009). Effect of *Bacillus subtilis* on the growth and survival rate of shrimp (*Litopenaeus vannamei*). *African Journal of Biotechnology*. 8(14):3369-3376.

- FAO. (1995). Aquaculture Production Statistics 1984-1993. FAO Fisheries Circular No. 815, Revision 7. Food and Agriculture Organization of the United Nations, Rome.
- FAO. (2001). 1999 Fisheries Statistics: Aquaculture Production. Volume 88/2. FAO, Rome.
- Faramarzi, M., Jafaryan, H., Patimar, R., Kordjazi, Z., Kiaalvandi, S., Iranshahi, F., Bolouki, M.L., Adineh, H., Roozbehfar, R., Malekzadeh, A. and Isari, A. (2011). The potential of *Daphnia magna* bioencapsulated with probiotics bacilli on growth and feeding parameters of Persian Sturgeon (*Acipenser persicus*) larvae. *World Journal of Zoology*. 6(3):268-273.
- Fjellheim, A.J., Playfoot, K.J., Skjermo, J. and Vadstein, O. (2007). *Vibrionaceae* dominates the microflora antagonistic towards *Listonella anguillarum* in the intestine of cultured Atlantic cod (*Gadus morhua* L.) larvae. *Aquaculture*. 269: 98–106.
- Fuller, R. (1989). A review: probiotics in man and animals. *Journal of Applied Bacteriology*. 66:365-378.
- Fuller, R. (1992). History and development of probiotics. In: Fuller, R. (Ed.), *Probiotics: The Scientific Basis*. pp. 1–8. Chapman and Hall, London
- Garrity, G.M., Bell, J.A. and Lilburn, T.G. (2003). *Bergey's Manual of Systematic Bacteriology*. Springer-Verlag, New York.
- Gatesoupe, F. J. (1991). The effect of three strains of lactic bacteria on the production rate of rotifers, *Brachionus plicatilis*, and their dietary value for larval turbot, *Scophthalmus maximus*. *Aquaculture*. 96:335–342.
- Gatesoupe, F.J. (1999). The use of probiotics in aquaculture. *Aquaculture*. 180: 147–165.
- Gatesoupe, F.J. (2010). Probiotics and other microbial manipulations in fish feeds: Prospective health benefits. In *Animal Models to Study Probiotics* (pp. 541-552). Elsevier Inc.
- Gullian, M., Thompson, F. and Rodriguez, J. (2004). Selection of probiotic bacteria and study of their immunostimulatory effect in *Penaeus vannamei*. *Aquaculture*. 233:1–14.
- Ghosh, S., Ringo, E., Deborah, G.S.A., Rahiman, K.M.M. and Hatha, A.A.M. (2011). *Enterobacter hormaechei* Bac 1010 from the gut of flathead grey mullet as probable aquaculture probionat. *Journal of Nature Science and Sustainable Technology*. 5(3):189-199.
- Gildberg, A., Mikkelsen, H., Sandaker, E. and Ringo, E. (1997). Probiotic effect of lactic acid bacteria in the feed on growth and survival of fry of Atlantic cod (*Gadus morhua*). *Hydrobiologia*. 352:279-285.

- Gismondo, M.R., Drago, L. and Lombardi, A. (1999). Review of probiotics available to modify gastrointestinal flora. *International Journal of Antimicrobial Agents*. 12:287-292.
- Govoni, J.J., Boehlert, G.W. and Watanabe, Y. (1986). The physiology of digestion in fish larvae. *Environment Biology of Fish*. 61: 187-201.
- Gram, L., Melchiorson, J., Spanggaard, B., Huber, I. and Nielsen, T. (1999). Inhibition of *Vibrio anguillarum* by *Pseudomonas fluorescens* strain AH2, a possible probiotic treatment of fish. *Applied Environmental Microbiology*. 65:969-973.
- Gram, L., Lovold, T., Nielsen, J., Melchiorson, J. and Spanggaard, B. (2001). *In vitro* antagonism of the probiont *Pseudomonas fluorescens* strain AH2 against *Aeromonas salmonicida* does not confer protection of salmon against furunculosis. *Aquaculture*. 199:1-11.
- Grace, M. (2009). *Taxonomy, identification and biology of Seabass (Lates calcarifer)* (pp. 38-43). Central Marine Fisheries Research Institute. National Fisheries Development Board. National Training on 'Cage Culture of Seabass' held at CMFRI, Kochi. Dec. 14-23.
- Greenwood, P.H. (1976). A review of the Family Centropomidae (*Pisces perciformes*). *Bulletin of the British Museum (Natural History) Zoology*. 29:1-81.
- Gonza'lez, C., Encinas, J., Garcı'a-Lo'pez, M. and Otero, A. (2000). Characterization and identification of lactic acid bacteria from freshwater fish. *Food Microbiology*. 17:383-391.
- Guarner, F. and Malagelada, J.R. (2003). Gut flora in health and disease. *The Lancet*. 360(8):512-519.
- Güroy, D., Deveciler, E., Kut Guroy, B. and Tekinay, A.A. (2005). Influence of feeding frequency on feed intake, growth performance and nutrient utilization in European sea bass (*Dicentrarchus labrax*) fed pelleted or extruded diets. *Turkey Journal Veterinary Animal Science*. 30:171-177.
- Halver, J.E. (1971). The vitamins. In: *Fish nutrition*. Academic Press, London, UK.
- Hansen, G. H., Strøm, E. and Olafsen, J. A. (1992). Effect of different holding regimens on the intestinal microflora of herring (*Clupea harengus*) larvae. *Applied Environment Microbiology*. 58:461-470.
- Harikrishnan, R., Balasundaram, C. and Heo, M.S. (2010). Effect of probiotics enriched diet on *Paralichthys olivaceus* infected with lymphocystis disease virus (LCDV). *Fish and Shellfish Immunology*. 29:868-874.
- Haroun, E., Goda, A. and Kabir, M. (2006). Effect of dietary probiotic Biogen supplementation as agrowth promoter on growth performance and feed utilization of Nile tilapia *Oreochromis niloticus* (L.). *Aquaculture Research*. 37(14):1473-1480.

- Harzevili, A.R.S., Van Duffel, H., Dhert, P., Swings, J. and Sorgeloos, P. (1998). Use of a potential probiotic *Lactobacillus lactis* Ar21 strain for the enhancement of growth in the rotifer *Brachionus plicatilis* (Muller). *Aquaculture Research*. 29:411-417.
- Hassan, R. (1992). Acute ammonia toxicity of red tilapia and seabass. *Fisheries Bulletin of the Department of Fisheries, Kuala Lumpur, Malaysia*, No. 73.
- Havenaar, R. and Huis, in't Veld, M.J.H. (1992). Probiotics: A general view. In: *Lactic acid bacteria in health and disease* (Ed) Wood, J.B.J. Vol. 1. Elsevier Applied Science Publishers, Amsterdam.
- Hoffman, H., Stindl, S., Stumpf, A., Mehlen, A., Monget, D., Heesemann, J., Schleifer, K. H. and Roggenkamp, A. (2005). Description of *Enterobacter ludwigii* sp. nov., a novel *Enterobacter* species of clinical relevance. *Systematic and Applied Microbiology*. 28:206-212.
- Holt, J.G, Krieg, N.R, Sneath, P.H.A., Staley, J.T. and Williams, S.T. (2000). *Bergey's Manual of Determinative Bacteriology*, Ninth Edition (pp. 178). Lippincott Williams and Wilkins.
- Hossain, M.I., Kamal, M.M., Mannan, M.A., Bhuyain, M.A.B. and Hossain, M.I. (2013). Effects of probiotics on growth and survival of shrimp (*Penaeus monodon*) in coastal pond at Khulna, Bangladesh. *Journal of Scientific Research*. 5(2):363-370.
- Huys, L., Dhert, P.H., Robles, R., Ollevier, F., Sorgeloos, P. and Swings, J. (2001) Search for beneficial bacterial strains for turbot (*Scophthalmus maximus* L.) larviculture. *Aquaculture*. 193:25-37.
- Hrenovic, J. and Ivankonic, T. (2009). Survival of *Escherichia coli* and *Acinetobacter junii* at various concentrations of sodium chloride. *EurAsian Journal of BioSciences*. 3:144-151.
- Ige, B.A. (2013). Probiotics use in intensive fish farming. *African Journal of Microbiology Research*. 7(22):2701-2711.
- Iman, M. K. A., Wafaa, T. A., Elham, S. A., Mohammad, M. N. A., El-Shafei, K., Osama, M. S., Gamal, A. I., Zeinab, I. S. and El-Sayed, H. (2013). Evaluation of *Lactobacillus plantarum* as a probiotic in aquaculture: Emphasis on growth performance and innate immunity. *Journal of Applied Sciences Research*. 9 (1):572-582.
- Irianto, A. and Austin, B. (2002). Probiotics in aquaculture. *Journal of Fish Diseases*. 25:1-10.
- Jay, J. M. (1982). Antimicrobial properties of diacetyl. *Applied Environmental Microbiology*. 44:525-532.

- Katersky, R. S. and Carter, C. G. (2007). A preliminary study on growth and protein synthesis of juvenile barramundi, *Lates calcarifer* at different temperature. *Aquaculture*. 267:157-164.
- Kesarcodi-Watson, A., Kaspar, H., Lategan, M.J. and Gibson, L. (2008). Probiotics in aquaculture: The need, principles and mechanisms of action and screening processes. *Aquaculture*. 274:1-14.
- Khattab, Y.A.E., Shalaby, A.M.E., Sharaf Saffa, M., El-Marakby, H. and RizlAlla, E.H. (2004). The physiological changes and growth performance of the Nile Tilapia *Oreochromis niloticus* after feeding with Biogen as growth promoter. *Egyptian Journal of Aquatic Biology Fisheries*. 8(2):145–158.
- Kim, D.H., Brunt, J. and Austin, B. (2007). Microbial diversity of intestinal contents and mucus in rainbow trout (*Oncorhynchus mykiss*). *Journal of Applied Microbiology*. 102:1654–1664.
- Kitamura, H., Ishitani, H., Kuge, Y. and Nakamoto, N. (1982). Determination of nitrate in freshwater and seawater by hydrazine reduction method. *Japan Journal of Water Pollution Research*. 5:35-42.
- Kollath, W. (1953). Ernährung und Zahnsystem. *Deutsch. Zahnaerzt. Z.* 8:7-16.
- Kuangvankij, P., Pudadera, B.J., Tiro, L.B. and Potestas, I.O. (1984). Biology and culture of sea bass (*Lates calcarifer*) (pp 67). NACA Training Manual Series No 3.
- Kumar, D., Suresh, K., Dey, R.K. and Mishra, B.K. (1986). Stress mediated columnaris disease in rohu, *Labeo rohita* (Ham.). *Journal of Fish Diseases*. 9:87-89.
- Laloo, R., Ramchuran, S., Ramduth, D., Gorgens, J. and Gardiner, N. (2007). Isolation and selection of *Bacillus* spp. as potential biological agents for enhancement of water quality in culture of ornamental fish. *Journal of Applied Microbiology*. 1-9.
- Lauzon, H.L., Gudmundsdottir, S., Petursdottir, S.K., Reynisson, E., Steinarsson, A., Oddgeirsson, M., Bjornsdottir, R. and Gudmundsdottir, B.K. (2010). Microbiota of Atlantic cod (*Gadus morhua* L.) rearing systems at pre- and post hatch stages and the effect of different treatments. *Journal of Applied Microbiology*. 109:1775-1789.
- Lertcanawanichakul, M. and Sawangnop, S. (2008). A comparison of two methods used for measuring the antagonistic activity of *Bacillus* species. *Walailak Journal of Science and Technology*. 5(2):1610171.
- Lightner, D.V. (1996). A Handbook of shrimp pathology and diagnostic procedures for diseases of cultures *Penaeid* shrimp. *World Aquaculture Society*, Baton Rouge, LA, USA.

- Lily, D.M. and Stillwell, R.H. (1965). Probiotics. Growth promoting factors produced by microorganisms. *Science*. 147:747-748.
- Lim, H.J., Kapareiko, D., Schott, E.J., Hanif, A., and Wikfors, H. (2011). Isolation and evaluation of new probiotic bacteria for use in shellfish hatcheries: I. Isolation and screening for bioactivity. *Journal of Shellfish Research*. 30(3):609-615.
- Lovatelli, A. (1990). Regional seafarming development and demonstration project. Regional Seafarming Resources Atlas, FAO Corporate Document Repository (pp 83). FAO, Rome.
- Magda, M.E., Sabry, S.E., Mohamed, A.E., Santoch, L., Said, M.D and Neven, A.E. (2011). The viability of probiotics as a factor influencing the immune response in the Nile tilapia, *Oreochromis niloticus*. *Egypt Journal Aquatic Biology and Fishery*. 15(1):105-124.
- Mahdhi, A., Kamoun, F., Messina, C., Santulli, A. and Bakhrouf, A. (2012). Probiotic properties of *Brevibacillus brevis* and its influence on sea bass (*Dicentrarchus labrax*) larval rearing. *African Journal of Microbiology Research*. 6 (32): 6487-6495.
- Maity, J., Kundu, J., Pramanik, A. and Patra, B.C. (2011). Effect of cellulolytic gut bacteria as a feed supplement on the growth performance and nutrient digestibility of Asian Seabass (*Lates calcarifer*). *International Journal of Aquatic Science*. 2(1):3-15.
- Marzouk, M.S., Moustafa, M.M. and Nermeen, M.M. (2008). The influence of some probiotics on the growth performance and intestinal microbial flora of *O. niloticus*. (pp. 1059-1071). 8<sup>th</sup> International Symposium on Tilapia in Aquaculture.
- Mehrabi, Z., Firouzbakhsh, F. and Jafarpour, A. (2011). Effects of dietary supplementation of symbiotic on growth performance, serum biochemical parameters and carcass composition in rainbow trout (*Oncorhynchus mykiss*) fingerlings. *Journal of Animal Physiological Animal Nutrition*. 24:0396-1439.
- Merrifield, D.L., Bradley, G., Baker, R.T.M. and Davies, S.J. (2010). Probiotic applications for rainbow trout (*Oncorhynchus mykiss* Walbaum) II. Effects on growth performance, feed utilization, intestinal microbiota and related health criteria post-antibiotic treatment. *Aquaculture Nutrition*. 16:496-503.
- Mette, H., Oivind, B., Ana, R., Janne, N., Jette, M., Sigmund, J., Hazel, D., Peter, A., Harry, B. and Lone, G. (2004). Selection and identification of autochthonous potential probiotic bacteria from turbot larvae (*Scophthalmus maximus*) rearing units. *Systematic Applied Microbiology*. 27:360-371.
- Michael, T.M. and John, M.M. 2006. Brock Biology of Microorganisms. International Edition. Eleventh Edition (pp.150-152). Pearson Education International.



- Mohapatra, S., Chakraborty, T., Kumar, V., Deboeck, G. and Mohanta, K. N. (2012). Aquaculture and stress management: A review of probiotic intervention. *Journal of Animal Physiology and Animal Nutrition*. 97(3):405-430.
- Moriarty, D.J.W. (1998). Control of luminous *Vibrio* species in penaeid aquaculture ponds. *Aquaculture*. 164:351-358.
- Moriarty, D.J.W. (1990). Interactions of microorganisms and aquatic animals, particularly the nutritional role of the gut flora. In: Lesel, R. (Ed.), *Microbiology of Poecilotherms*. (pp. 217–223). Elsevier Amsterdam, New York.
- Munro, P.D., Birkbeck, T.H. and Barbour, A. (1993). Influence of rate of bacterial colonization of the gut of turbot larvae on larval survival. In: *Fish Farming Technology – Proceedings of the First International Conference of on Fish Farming Technology*. Balkema, Rotterdam. Reinerstein, H., Dahle, L.A., Jørgensen, L. and Tvinnereim, K. (Ed.). pp. 85–92.
- Munro, P.D., Barbour, A. and Birkbeck, T.H. (1994). Comparison of the gut bacterial flora of start-feeding larval turbot reared under different conditions. *Journal of Applied Bacteriology*. 77:560–566.
- Nageswara, P. V. and Babu, D.e. (2006). Probiotics as an alternative therapy to minimize or avoid antibiotics use in aquaculture. *Fishing Chimes*. 26(1):112-114.
- Nair, A. J. (2010). Principles of biochemistry and genetic engineering, 2<sup>nd</sup> Edition. In *Cellular Techniques*. (pp. 292). Laxmi Publications.
- Nansombat, S., Phunpruch, S. and Jaichalad, T. (2012). Screening and identification of lactic acid bacteria from raw seafoods and Thai fermented seafood products for their potential use as starter cultures. *Songklanakarin Journal of Science and Technology*. 34(3):255-262.
- Nayak, S.K. (2010). Probiotics and immunity: A fish perspective. *Fish and Shellfish Immunology*. 29:2-14.
- Nayak S.K., Mukherjee S.C. (2011). Screening of gastrointestinal bacteria of Indian major carps for a candidate probiotic species for aquaculture practices. *Aquaculture Research*. 42:1034-1041.
- Nester, E.W., Anderson, D.G., Roberts Jr.C.E., Pearsall, N.N. and Nester, M.T. (2004). *Microbiology: A human perspective*. McGraw-Hill Inc, New York.
- Nikoskelainen, S., Ouwehand, A.C., Bylund, G., Salminen, S. and Lilius, E.M. (2003). Immune enhancement in rainbow trout (*Oncorhynchus mykiss*) by potential probiotic bacteria (*Lactobacillus rhamnosus*). *Fish and Shellfish Immunology*. 15:443–452.
- Noveirian, H. A. and Nasrollahzadeh, A. (2012). The effects of different levels of biogen probiotic additives on growth indices and body composition of juvenile

- common carp (*Cyprinus carpio* L.). *Journal of Environmental Science*. 10 (1): 115-121.
- Nurhidayu, A., Ina-Salwany, M.Y., Mohd Daud, H. and Harmin, S.A. (2012). Isolation, screening and characterization of potential probiotics from farmed tiger grouper (*Epinephelus fuscoguttatus*). *African Journal of Microbiology Research*. 6(9):1924-1933.
- Oggioni, M.R., Ciabattini, A., Cuppone, A.M. and Pozzi, G. (2003). *Bacillus* spores for vaccine delivery. *Vaccine*. 31:96-101.
- Olafsen, J.A. (2001). Interactions between fish larvae and bacteria in marine aquaculture. *Aquaculture*. 200:223-247.
- Othman, M. F. Challenge ahead in meeting aquaculture production in Malaysia. In *Third National Agriculture Policy, Nap3 (1998-2010)*. 17-03-2008. Brackish Water Aquaculture Research Center (BARC) Ministry of Agricultural and Agro-Based Industry Department of Fisheries Malaysia Johor. (2008).
- Pan, X., Wu, T., Song, Z., Tang, H. and Zhao, Z. (2008). Immune responses and enhanced disease resistance in Chinese drum, *Miichthys miiuy* (Basilewsky), after oral administration of live or dead cells of *Clostridium butyricum* CB2. *Journal of Fish Diseases*. 31:679–686.
- Panigrahi, A., Kiron, V., Kobayashi, T., Puangkaew, J., Satoh, S. and Sugita, H. (2004). Immune responses in rainbow trout, *Onchorhynchus mykiss*, induced by a potential probiotic, *Lactobacillus rhamnosus* JCM1136. *Veterinary Immunology and Immunopathology*. 102:379-388.
- Panigrahi, A. and Azad, I.S. (2007). Microbial invention for better fish health in aquaculture: The Indian scenario. *Fish Physiological Biochemical*. 33:429-758.
- Panneerselvam, A. and Arumugam, G. (2012). Isolation and identification of bacteria from lake water in and around Ranipet Are, Vellore District. *International Journal of Pharmaceutical and Biological Archives*. 3(4):1008-1011.
- Parker, R.B. (1974). Probiotics, the other half of the antibiotic story. *Animal Nutrition Health*. 29:4-8.
- Parsons, T.R., Maita, Y. and Lalli, C.M. (1984). A manual of chemical and biological methods for seawater analysis (pp. 3-38). Pergamon Press, Oxford.
- Parthasarathy, R. and Ravi, D. (2011). Probiotic bacteria as growth promoter and biocontrol agent against *Aeromonas hydrophila* in *Catla catla* (Hamilton, 1822). *Indian Journal of Fisheries*. 58(3):87-93.
- Picchiotti, S., Mazzini, M., Taddei, A.R., Renna, R., Fausto, A.M., Mulero, V., Carnevali, O., Cresci, A. and Abelli, L. (2007). Effects of administration of probiotic strains on GALT of larval gilthead seabream: immunohistochemical and ultrastructural studies. *Fish and Shellfish Immunology*. 22: 57–67.



- Plant, L., Lamy, C., Conway, P.L. and KO'Riordan, K. (2003). Gastrointestinal microbial community shifts observed following oral administration of a *Lactobacillus fermentum* strain to mice. *FEMS Microbiology Ecology*. 43:133-140.
- Pond, M.J., Stone, D.M. and Alderman, D.J. (2006). Comparison of conventional and molecular techniques to investigate the intestinal microflora of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*. 261:194-203.
- Porubcan, R.S. (1991). Reduction of ammonia nitrogen and nitrate in tanks *Penaeus monodon* using floating biofilters containing processed diatomaceous earth media pre-inoculated with nitrifying bacteria. *Journal of World Aquaculture Society*. 22(3):48.
- Purivirojkul, W. (2013). Application of probiotic bacteria for controlling pathogenic bacteria in fairy shrimp *Branchinella thailandensis* culture. *Turkish Journal of Fisheries and Aquatic Sciences*. 13: 187–196.
- Rahman, S., Khan, S. N., Naser, M. N. and Karim, M. M. Probiotic technology for sustainable aquaculture. In: *Advances in Fisheries Research in Bangladesh: Proceeding of 5<sup>th</sup> Fisheries conference and Research Fair 2012*, Bangladesh Agricultural Research Council, Dhaka, Bangladesh Fisheries Research Forum, Dhaka, Bangladesh, 18-19 January 2012. Wahab, M. A., Shah, M. S., Hossain, M. A. R., Barman, B. K. and Hoq, M. E. (Ed.); pp 83-96. 2014.
- Ramirez, R.F. and Dixon, B.A. (2003). Enzyme production by obligate intestinal anaerobic bacteria isolated from oscars (*Astronotus ocellatus*), angelfish (*Pterophyllum scalare*) and southern flounder (*Paralichthys lethostigma*). *Aquaculture*. 227:417–426.
- Rao, A.V. (2001). Bioremediation technology to maintain healthy ecology in aquaculture ponds. *Aquaculture Technology*. 80-84.
- Ravi, A.V., Musthafa, K.S., Jegathammbal, G., Kathiresan, K. and Pandian, S.K. (2007). Screening and evaluation of probiotics as a biocontrol agent against pathogenic *Vibrios* in marine aquaculture. *Letters in Applied Microbiology*. 45:219-223.
- Refstie, S., Landsverk, T., Bakke-McKellep, A.M., Ringø, E., Sundby, A., Shearer, K.D. and Krogdahl, A. (2006). Digestive capacity, intestinal morphology, and microflora of 1 year and 2-year old Atlantic cod (*Gadus morhua*) fed standard or bioprocessed soybean meal. *Aquaculture*. 261:269-284.
- Reitan, K.I., Natvik, C.M. and Vadstein, O. (1998). Drinking rate, uptake of bacteria and microalgae in turbot larvae. *Journal of Fish Biology*. 53:1145-1154.
- Rengpipat, S., Rueangruklikhit, T. and Piyatiratitivorakul, S. (2008). Evaluations of lactic acid bacteria as probiotics for juvenile seabass *Lates calcarifer*. *Aquaculture Research*. 39:134-143.

- Rimmer, M.A. and Russell, D.J. (1998). Aspects of the biology and culture of *Lates calcarifer*. In: S.S. De Silva (ed.). *Tropical Mariculture* (pp 449-476). Academic Press, San Diego, USA.
- Rimmer, M. (2003). Barramundi. In Lucas, J.S. and Southgate, P.C. *Aquaculture farming aquatic animals and plants* (pp 364-381). Blackwell Publishing.
- Ringø, E., Strøm, E. and Tabachek, J.A. (1995). Intestinal micro-fora of salmonids: a review. *Aquaculture Research*. 26:773-789.
- Ringø, E., Birkbeck, T.H, Munro, P.D., Vadstein, O. and Hjelmeland, K. (1996). The effect of early exposure to *Vibrio pelagius* on the aerobic bacterial flora of turbot, *Scophthalmus maximus* larvae. *Journal of Applied Bacteriological*. 81:207-211.
- Ringø, E. and Gatesoupe, F.J. (1998). Lactic acid bacteria in fish: A review. *Aquaculture*. 160:177-203.
- Ringø, E. and O. Vadstein. (1998). Colonization of *Vibrio pelagius* and *Aeromonas caviae* in early developing turbot (*Scophthalmus maximus* L.) larvae. *Journal of Applied Microbiology*. 84:227-233.
- Ringø, E. and Birkbeck, T.H. (1999). Intestinal microflora of fish larvae and fry. *Aquaculture Research*. 30:73-93.
- Ringø, E., Sperstad, S., Myklebust, R., Refstie, S. and Krogdahl, A. (2006). Characterisation of the microbiota associated with intestine of Atlantic cod (*Gadus morhua* L.) The effect of fish meal, standard soybean meal and a bioprocessed soybean meal. *Aquaculture*. 261:829-841.
- Ringø, E., Myklebust, R., Mayhew, T.M. and Olsen, R.E. (2007). Bacterial translocation and pathogenesis in the digestive tract of larvae and fry. *Aquaculture*. 268:251-264.
- Robertson, P.A. W., Odowd, C., Burrells, C., Williams, P. and Austin, B. (2000). Use of *Carnobacterium* sp. as a probiotic for Atlantic salmon (*Salmo salar* L.) and rainbow trout (*Oncorhynchus mykiss*, Walbaum). *Aquaculture*. 185:235-243.
- Roo, F.J., Socorro, J., Izquierdo, M.S., Caballero, M.J., Hernandez-Cruz, C.M., Fernandez, A. and Fernandez-Palacios, H. (1999). Development of red porgy *Pagrus pagrus* visual system in relation with changes in the digestive tract and larval feeding habits. *Aquaculture*. 179:499-512.
- Ruangpan, L. (1987). Diseases of cultured sea bass. In: Training manual on sea bass (*Lates calcarifer*) culture in Thailand. Prepared by Satul Fisheries Station, Thailand, for the Demonstration/Training Course on sea bass culture: 1-22 August 1988 in Satul, Thailand. Training Manual 88/3 (RAS/86/024). Chapter X:69-84.

- Ruangpanit, N. Developing hatchery techniques for sea bass (*Lates calcarifer*): a review. In *Management of wild and cultured sea bass/barramundi (Lates calcarifer)* (pp 132-135). Proceedings of an international workshop, Darwin, N.T., Australia, Sept. 24-30, 1986. Copland J.W. and Grey, D.L. (Ed.); ACIAR Proceedings No. 20. Australian Centre for International Agricultural Research, Canberra, Australia.
- Saenz de Rodriguez, M.A., Diaz-Rosales, P., Chabrillon, M., Smidt, H., Arijo, S., Leon- Rubio, J.M., et al. (2009). Effect of dietary administration of probiotics on growth and intestine functionality of juvenile Senegalese sole (*Solea senegalensis*, Kaup 1858). *Aquatic Nutrition*. 15:177-85.
- Sahu, M.K., Swarnakumar, N.S., Thangaradjou, T. and Kannan, L. (2008). Probiotics in aquaculture: Importance and future perspectives. *Indian Journal Microbiology*. 48:299-308.
- Sakata, T. (1990). Microflora in the digestive tract of fish and shellfish. In R. Lesel. *Microbiology in Poecilotherms* (pp. 171-176). Elsevier, Amsterdam.
- Salinas, I., Cuesta, A., Esteban, M.A. and Meseguer, J. (2005). Dietary administration of *Lactobacillus delbrueckii* and *Bacillus subtilis*, single or combined, on gilthead seabream cellular innate immune responses. *Fish and Shellfish Immunology*. 19:67-77.
- Salminen, S., Ouwehand, A.C., Benno, Y. and Lee, Y.K. (1999). Probiotics: how should they be defined?. *Trends Food Science Technology*. 10:107-110.
- Scholz, U., Garcia Diaz, G., Ricque, D., Cruz Suarez, L.E., Vargas, A.F. and Latchford, J. (1999). Enhancement of vibriosis resistance in juvenile *Penaeus vannamei* by supplementation of diets with different yeast products. *Aquaculture*. 176:271-283.
- Sekar, V.T., Santiago, T.C., Vijayan, K.K., Alavandi, S.V., Raj, V.S., Rajan, J.J.S., Sanjuktha, M. and Kalaimani, N. (2008). Involvement of *Enterobacter cloacae* in the mortality of the fish, *Mugil cephalus*. *Letters in Applied Microbiology*. 46:667-672.
- Shailender, M., Krishna, P.V. and Suresh, B.C. (2012). Effect of probiotics on growth and survival of post larvae of giant freshwater prawn, *Macrobrachium rosenbergii* (De Man). *International Journal of Bioassays*. 01(12):184-190.
- Shakibazadeh, S., Che Roos, S., Hafezieh, M., Christianus, A., Kamaridin, M.S., and Kamaruzaman, S. (2012). A putative probiotic isolated from hatchery reared juvenile *Penaeus monodon*. *Iranian Journal of Fisheries Sciences*. 11(4):849-866.
- Shan, Y., Lai, Y. and Yan, A. (2012). Metabolic Reprogramming Under Microaerobic and Aerobic Conditions in Bacteria. In X. Wang, J. Chen and P. Quinn. *Reprogramming Microbial Metabolic Pathways*. (pp 159-163). Springer Science and Business Media.

- Shapawi, R., Tan, E.T. and Al-Azad, S. (2012). Inclusion of purple non-sulfur bacterial biomass in formulated feed to promote growth, feed conversion ratio and survival of Asian Seabass *Lates calcarifer* juveniles. *Journal of Fisheries and Aquatic Science*. 7(6):475-480.
- Sharifuzzaman, S.M. and Austin, B. (2009). Influence of probiotic feeding duration on disease resistance and immune parameters in rainbow trout. *Fish and Shellfish Immunology*. 27:440-445.
- Sharifuzzaman, S. M., Al-Harbi, A. H. and Austin, B. (2014). Characteristics of growth, digestive system functionality, and stress factors of rainbow trout fed probiotics *Kocuria* SM1 and *Rhodococcus* SM2. *Aquaculture*. 418-419:55-61.
- Sihag, R.C. and Sharma, P. (2012). Probiotics: The new ecofriendly alternative measures of disease control for sustainable aquaculture. *Journal of Fisheries and Aquatic Science*. 7(2):72-103.
- Silvi, S., Nardi, M., Sulpizio, R., Orpianesi, C., Caggiano, M., Carnevali, O. and Cresci, A. (2008). Effect of the addition of *Lactobacillus delbrueckii* subsp. *delbrueckii* on gut microbiota composition and contribution to the well-being of European sea bass (*Dicentrarchus labrax*, L.). *Microbial Ecology in Health and Disease*. 20: 53-59.
- Skrodenyte-Arbaciauskiene, V., Sruoga, A., Butkauskas, D. and Skrupskelis, K. (2008). Phylogenetic analysis of intestinal bacteria of freshwater salmon *Salmo salar* and sea trout *Salmo trutta trutta* and diet. *Fisheries Science*. 74:1307-1314.
- Son, V. M., Chang, C. C., Wu, M. C., Guu, Y. K. and Chiu, C. H. (2009). Dietary administration of the probiotic, *Lactobacillus plantarum*, enhanced the growth, innate immune responses, and disease resistance of the grouper *Epinephelus coioides*. *Fish Shellfish Immunology*. 26:691-698.
- Sorroza, L., Padilla, D., Acosta, F., Grasso, V., Vega, J. and Real, F. (2012). Characterization of the probiotic strain *Vagococcus fluvialis* in the protection of European sea bass (*Dicentrarchus labrax*) against vibriosis by *Vibrio anguillarum*. *Veterinary Microbiology*. 155:369-373.
- Spanggaard, B., Huber I., Nielsen, J., Sick, E.B., Pipper, C.B., Martinussen, T., Slierendrecht, W.J. and Gram, L. (2001). The probiotic potential against vibriosis of the indigenous microflora of rainbow trout. *Environmental Microbiology*. 3:755-765.
- Stofels, G., Nes, I.F. and Guomundsottir, A. (1992). Isolation and properties of a bacteriocin-producing *Carnobacterium piscicola* isolated from fish. *Journal of Applied Bacteriology*. 73:309-316.
- Subasinghe, R. (1997). Fish health and quarantine. In *Review of the State of the World Aquaculture*, FAO Fisheries circular no. 886, pp. 45-49. Food and Agriculture Organization of the United Nations, Rome.

- Sugita, H., Tokuyama, K. and Deguchi, Y. (1985). The intestinal microflora of carp *Cyprinus carpio*, grass carp *Ctenopharyngodonidella* and tilapia *Sarotherodon niloticus*. *Bulletin of the Japanese Society of Scientific Fisheries*. 51:1325-1329.
- Sugita, H., Tsunohara, M., Ohkoshi, T. and Deguchi, Y. (1988). The establishment of an intestinal microflora in developing goldfish (*Carassius auratus*) of culture ponds. *Microbial Ecology*. 15:333-344.
- Sugita, H., Kawasaki, J. and Deguchi, Y. (1997). Production of amylase by the intestinal microflora in cultured freshwater fish. *Letter in Applied Microbiology*. 24:105-108.
- Sugita, H., Hirose, Y., Matsuo, N. and Deguchi, Y. (1998). Production of the antibacterial substances by *Bacillus* sp. starin NM 12, an intestinal bacterium of Japanese coastal fish. *Aquaculture*. 165:269-280.
- Sugita, H., Okano, R., Suzuki, Y., Iwai, D., Mizukami, M., Akiyama, N. and Matsuura, S. (2002). Antibacterial abilities of intestinal bacteria from larval and juvenile Japanese flounder against fish pathogens. *Fishery Science*. 68:1004-1011.
- Suguna, S. and Rajendran, K. (2012). Production of probiotics from *Streptomyces* sp. associated with fresh water fish and its growth evaluation on *Xiphorous helleri*. *International Journal of Pharmaceutical and Biological Archives*. 3(3):601-603.
- Sung, H.H., Hsu, S.F., Chen, C.K., Ting, Y.Y. and Chao, W.L. (2001). Relationship between disease outbreaks in cultured tiger shrimp (*Penaeus monodon*) and the composition of *Vibrio* communities in pond water and shrimp hepatopancreas during cultivation. *Aquaculture*. 192:101-110.
- Sunitha, K. and Padmavathi, P. (2013). Influence of probiotics on water quality and fish yield in fish ponds. *International Journal of Pure and Applied Sciences and Technology*. 19(2):48-60.
- Talpur, A.D., Memon, A.J., Khan, M.I., Ikhwanuddin, M., Danish Danial, M.M. and Abol-Munafi, A.B. (2012). Isolation and screening of lactic acid bacteria from the gut of blue swimming crab, *P. pelagicus*, an *in vitro* inhibition assay and small scale *in vivo* model for validation of isolates as probiotics. *Journal of Fisheries and Aquatic Science*. 7(1):1-28.
- Tanaka, R., Ootsubo, M., Sawabe, T., Ezur, Y. and Tajima, K. (2004). Biodiversity and in situ abundance of gut microflora of abalone (*Haliotis discus hannai*) determined by culture-independent techniques. *Aquaculture*. 241:453-463.
- Temmerman, R., Huy, G. and Swings, J. (2004). Identification of lactic acid bacteria: culture-dependent and culture-independent methods. *Trends Food Science Technology*. 15:348-359.
- Thompson, F.L., Abreu, P.C. and Cavalli, R. (1999). The use of microorganisms as food source for *Penaeus paulensis* larvae. *Aquaculture*. 174:139-153.

- Timmermans, L.P.M. (1987). Early development and differentiation in fish. *Sarsia*. 72:331-339.
- Tovar-Ramirez, D., Infante, J. Z., Cahu, C., Gatesoupe, F. J. and Vazquez-Juarez, R. (2004). Influence of dietary live yeast on European sea bass (*Dicentrarchus labrax*) larval development. *Aquaculture*. 234 (1-4):415-427.
- Vadstein, O. (1997). The use of immunostimulation in marine larviculture: possibilities and challenges. *Aquaculture*. 155:401-417.
- Verner-Jeffreys, D.W., Shields, R.J. and Birkbeck, T.H. Bacterial influences on Atlantic halibut *Hippoglossus hippoglossus* yolk-sac larval survival and start-feed response. *Diseases of Aquatic Organisms*. 56:105-113.
- Verschuere, L., Rombaut, G., Sorgeloos, P., and Verstraete, W. (2000). Probiotic bacteria as biological control agents in aquaculture. *Microbiology and Molecular Biology Reviews*. 64(4):655-671.
- Vine, N.G., Leukes, W.D. and Kaiser, H. (2004). *In vitro* growth characteristics of five candidate aquaculture probiotics and two fish pathogens grown in fish intestinal mucus. *FEMS Microbiology Letters*. 231:145-152.
- Vine, N.G., Leukes, W.D. and Kaiser, H. (2006). Probiotics in marine larviculture. *FEMS Microbiology Reviews*. 30:404-427.
- Wang, Y., Henz, M.E., Fregeau, G.N., Chai, S., Gibbs, A.C., Yan, L.Z., Stiles, M.E., Wishart, D.S. and Vederas, J.C. (1999). Solution structure of carnobacteriocin B2 and implications for structure-activity relationships among type IIa bacteriocins from lactic acid bacteria. *Biochemistry*. 38:15438-15447.
- Wang, Y.B., Xu, Z.R. and Xia, M.S. (2005). The effectiveness of commercial probiotics in northern white shrimp *Penaeus vannamei* ponds. *Fisheries Science*. 71:1036-1041.
- Wang, Y.B., Tian, Z.Q., Yao, J.T. and Li, W.F. (2008). Effect of probiotics, *Enterococcus faecium*, on tilapia (*Oreochromis niloticus*) growth performance and immune response. *Aquaculture*. 277:203-207.
- Wang, Y.B., Li, J.R. and Lin, J. (2008). Probiotics in aquaculture: Challenges and outlook. *Aquaculture*. 281:1-4.
- Weatherburn, M.W. (1967). Phenol-hypochlorite reaction for determination of ammonia. *Analytical Chemistry*. 39:971-974.
- William, A. R., David, J. W., Robert, A. W., Jane, D. S., Michele, L. P., Raymond, Y. W. C., Alfred, DeMaria, A., James, T. L., William, E. S., Beth, H. S. and Marjorie, A. U. (2008). *Guideline for disinfection and sterilization in healthcare facilities, 2008*. pp 39, 58-59. Department of Health and Human Services, USA.



- Woo, N.Y.S. and Chiu, S.F. (1994). Effects of nitrite exposure on growth and survival of sea bass, *Lates calcarifer*, fingerlings in various salinities. *Journal of Application Aquaculture*. 4:45-54.
- Wongsomnuk, S. and Manevonk, S. (1973). Results of experiment on artificial breeding and larval rearing of the sea bass (*Lates calcarifer*, Bloch). Contribution No. 5 Songkla Marine Fish Station, Department of Fisheries, 21 pp. (in Thailand).
- Wu, Z.X., Feng, X., Xie, L.L., Peng, X.Y., Yuan, J. and Chen, X.X. (2012). Effect of probiotic *Bacillus subtilis* Ch9 for grass carp, *Ctenopharyngodon idella* (Valenciennes, 1844), on growth performance, digestive enzyme activities and intestinal microflora. *Journal of Applied Ichthyology*. 28:721-727.
- Xie, F., Zhu, T., Zhang, F., Zhou, K., Zhao, Y. and Li, Z. (2013). Using *Bacillus amuloloquefaciens* for remediation of aquaculture water. *Springer Plus*. 2:119-124.
- Yasuda, K and Taga, N. (1980). A mass culture method for *Artemia salina* using bacteria as food. *Bulletin Society France-Japan Oceanography*. 18:53-63.
- Yang, G., Bao, B., Peatman, E., Li, H., Huang, L. and Ren, D. (2007). Analysis of the composition of the bacterial community in puffer fish *Takifugu obscurus*. *Aquaculture*. 262:183-191.
- Zhou, X.X., Wang, Y.B. and Li, W.F. (2009). Effect of probiotic on larvae shrimp (*Penaeus vannamei*) based on water quality, survival rate and digestive enzyme activities. *Aquaculture*. 287:349-353.
- Zhou, X. X. and Wang, Y. B. (2012). Probiotics in Aquaculture-Benefits to the Health, Technological Application and Safety. InC. Edmir. *Health and Environment in Aquaculture*. (pp 215-221). InTech.
- Zmyslowska, I. and Lewandowska, D. (1999). Survival of bacterial strains in fish feeds stored at different temperatures. *Polish Journal of Environmental Studies*. 8(6):447-451.

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## LIST OF PUBLICATIONS

- Wendy, F.T.**, Abu Hena, M.K., Wong, S.K., Idris, M.H., Sharifuzzaman, S.M., Ina-Salwany, M.Y. (2014). *Enterobacter ludwigii*, a candidate probiont from the intestine of Asian seabass. *Journal of Science and Technology in the Tropics*. 10:5-14.
- Wendy, F.T.**, Abu Hena, M.K., Wong, S.K., Idris, M.H. and Ina-Salwany, M.Y. (2013). In *Isolation and characterization of potential probiotics from gastrointestinal tract of Asian sea bass (Lates calcarifer, Bloch)*. Proceeding of Oceanography and Sustainable Marine Production: A challenge of managing marine resources under climate change, ICOSMaP, Kuantan, Malaysia, October 28-30, 2013.
- Abu Hena, M.K., Johan, I., Idris, M.H., Amy, H.R., Amin, S.M.N., Wong, S.K., Seca, G., Kamil, L., Mimi, A.M.T, Nettley, A.T, **Wendy, T.F.** and Muzammel, H. (2013). In *Fishermen community and knowledge transfer activities at Kuala Nyalau, Bintulu, Sarawak*. Proceeding of 1st National Conference on Knowledge Transfer Program (KTP01), Bangi, Malaysia, August 21-23, 2013. Kementerian Pendidikan Malaysia, 2013.