



UNIVERSITI PUTRA MALAYSIA

***NUTRITIONAL ENHANCEMENT OF WHOLE GRAIN BARIO RICE BY
PHYTIC ACID DEGRADATION DURING GERMINATION***

LEE HUEI HONG

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By

LEE HUEI HONG

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of
Doctor of Philosophy**

April 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

NUTRITIONAL ENHANCEMENT OF WHOLE GRAIN BARIO RICE BY PHYTIC ACID DEGRADATION DURING GERMINATION

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April 2015

Chair : Assoc. Prof. Yiu Pang Hung, PhD
Faculty : Agriculture and Food Sciences (Bintulu)

Whole grain rice is suggested to be a better solution against the polished form with more vitamins, minerals and fiber than its processed equivalent. Abundance of phytic acid in whole grain rice demotes the nutritional values of whole grain rice in general, although its beneficial properties were also reported. Various strategies had been conducted to counteract the nutritional limitations in whole grain rice, including germination. These led to the main objective of this study, to determine the nutritional quality in whole grain rice of Bario rice cultivars and later improve the targeted nutritional quality through the germination process.

Thirty rice cultivars named as “Adan” or “Bario” were assessed for nutrient bioaccessibility and antioxidant properties. The studies showed an acceptable level of *in vitro* digestibility in starch (193.77 ± 93.09 mg maltose released/g) and protein ($69.01 \pm 8.06\%$), but were poor in antioxidant properties and bioaccessibility of minerals (Phy/Fe 125.37 ± 38.95 , Phy/Zn 201.97 ± 80.54 and Phy/Ca 11.58 ± 3.35). Most of the lowland Bario rice with medium grains had relatively better nutrient bioaccessibility while pigmented rice had high amounts of phenolic compounds (0.64 ± 0.21 mg GAE/g) and antioxidant properties. The drawback in the whole grains were related to relatively low mineral content (24.30 ± 0.46 g Fe/100g, 1.30 ± 0.35 g Zn/100g, 13.30 ± 2.79 g Ca/100g) and high phytic acid content (24.29 ± 4.07 g/kg) in general, and also low phenolic content in non-pigmented rice (0.10 ± 0.04 mg GAE/g).

The relationship between intrinsic phytic acid content, nutrient bioaccessibility and antioxidant properties was studied. Results suggested that intrinsic phytic acid could be an important chelating antioxidant ($51.92 \pm 8.35\%$, $r = 0.32$) in non-pigmented rice which also reduced mineral bioaccessibility in the whole grains. Phytic acid content significantly influenced mineral bioaccessibility ($r = 0.40$ [Phy/Fe], $r = 0.27$ [Phy/Zn], $r = 0.60$ [Phy/Ca]) especially iron and calcium. *In vitro* digestibility of starch and protein were not affected by intrinsic phytic acid content. The contribution of phytic acid to the antioxidant properties of whole grain rice was low, but phytic acid remains as an important antioxidant in non-pigmented rice. These imply that the antinutrient effects from phytic acid were stronger and significantly contributed to the low mineral bioaccessibility in the collection.

Phytate degradation based on changes in phytic acid content, phytase activity and phytate globoids were investigated under various germination treatments in selected rice cultivars. This study showed that germination treatments facilitated the hydrolysis of phytate complexes in whole grain rice significantly, reducing phytic acid content (0.73 - 99.99% loss), accelerating phytase activity (0.38 – 270 U/kg increment) and releasing minerals from phytate globoids. However, the phytate degradation differed with rice cultivars under the same germination conditions. Germination is a suitable treatment for phytate degradation in Bario rice cultivars with the right germination conditions.

The study was continued with optimization of germination conditions by response surface methodology in the cultivar Tuan. Phytic acid content was significantly reduced (1.24 – 99.99% loss) with facilitation of phytase activity (27.43 – 165.3 U/kg increment) during germination and led to increments in mineral bioaccessibility (8.03 – 239.56% Ca, 25.38% Fe, 1.18 – 45.01% Zn). Soaking for 12 hours under acidic condition (pH 2) was the most suitable for improvement of calcium and zinc bioaccessibility at room temperature (25°C). However, longer germination duration (50 hours) was required for higher iron bioaccessibility. Optimized germination conditions of pH 2.7, at 25°C and 12 hours germination successfully reduced phytic acid content and phytic acid to minerals mole ratio, and led to an increase in mineral bioaccessibility in calcium, iron and zinc.

In conclusion, the whole grain rice of Bario rice cultivars had the advantages in *in vitro* digestibility of starch and protein but with limited mineral bioaccessibility and antioxidant properties. The limitations in mineral bioaccessibility were due to the abundance of phytic acid content in the whole grain rice. Phytate degradation in whole grain rice was cultivar and condition dependent. Optimized germination condition could promote phytate degradation and result in improved mineral bioaccessibility in selected whole grain Bario rice. Germinated whole grain rice with better mineral bioaccessibility can be a good substitution of whole grain rice as part of healthy diets.

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PENINGKATAN NILAI PEMAKANAN BERAS PENUH BARIO DENGAN PENGURAIAN ASID FITIK MELALUI PERCAMBAHAN

Oleh

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Beras penuh mempunyai vitamin, mineral dan serat yang lebih tinggi berbanding dengan beras putih. Kandungan asid fitik yang tinggi dalam beras penuh mengurangkan nilai pemakanan secara umum walaupun faedahnya juga telah dilaporkan. Pelbagai strategi telah dilaksanakan untuk mengatasi kekurangan dalam beras penuh, termasuk proses percambahan. Oleh itu, objektif utama kajian ini adalah menentukan nilai pemakanan dalam beras penuh Bario dan seterusnya meningkatkan nilai pemakanan melalui proses percambahan.

Tiga puluh kultivar beras yang dinamakan “Adan” atau “Bario” telah ditentukan bioaksesibiliti nutrien dan sifat antioksidasi. Koleksi beras ini menunjukkan tahap penghadaman kanji (193.77 ± 93.09 mg maltosa/g) dan protein ($69.01 \pm 8.06\%$) yang baik tetapi sifat antioksidasi dan bioaksesibiliti mineral (Phy/Fe 125.37 ± 38.95 , Phy/Zn 201.97 ± 80.54 and Phy/Ca 11.58 ± 3.35) yang rendah. Kebanyakan beras Bario dari lokasi tanaman tanah rendah dan berbentuk sederhana mempunyai bioaksesibiliti nutrien yang lebih tinggi. Manakala beras penuh berpigmen mengandungi fenolik yang tinggi (0.64 ± 0.21 mg GAE/g) dan sifat antioksidasi yang aktif. Kelemahan yang didapati dalam beras penuh Bario adalah kandungan mineral yang rendah (24.30 ± 0.46 g Fe/100g, 1.30 ± 0.35 g Zn/100g, 13.30 ± 2.79 g Ca/100g), kandungan asid fitik yang tinggi (24.29 ± 4.07 g/kg) dan kandungan fenolik yang rendah (0.10 ± 0.04 mg GAE/g) dalam beras bukan berpigmen.

Hubungan antara kandungan intrinsik asid fitik, bioaksesibiliti nutrien dan sifat antioksidasi telah dikaji. Keputusan mencadangkan asid fitik dalam beras penuh boleh menjadi agen antioksidasi yang penting ($51.92 \pm 8.35\%$, $r = 0.32$) dalam beras bukan berpigmen tetapi ia juga mengurangkan bioaksesibiliti mineral. Kandungan asid fitik mempengaruhi bioaksesibiliti mineral ($r = 0.40$ [Phy/Fe], $r = 0.27$ [Phy/Zn], $r = 0.60$ [Phy/Ca]) terutamanya zat besi dan kalsium. Penghadaman kanji dan protein secara *in vitro* tidak terjejas oleh kandungan asid fitik dalam beras penuh. Sumbangan asid fitik kepada sifat antioksidasi beras penuh adalah rendah, tetapi asid fitik masih merupakan agen antioksidasi yang penting dalam beras tidak berpigmen. Ini menunjukkan bahawa asid fitik memberi sumbangan yang lebih berkesan kepada sifat antinutrien dan memberi bioaksesibiliti mineral yang rendah dalam beras penuh Bario.

Penguraian asid fitik berdasarkan perubahan dalam kandungan asid fitik, aktiviti enzim fitase dan globoid fitik semasa proses percambahan telah dikaji dalam kultivar yang terpilih. Kajian ini menunjukkan bahawa percambahan membantu hidrolisis kompleks asid fitik, dengan mengurangkan kandungan asid fitik (0.73 - 99.99%), mempercepatkan aktiviti fitase (0.38 - 270.00 U/kg) atau penguraian mineral dari globoid. Walau bagaimanapun, perubahan tersebut adalah berbeza dalam setiap kultivar beras yang diuji. Justeru, percambahan merupakan kaedah yang sesuai untuk penguraian asid fitik dalam kultivar beras Bario dengan keadaan percambahan yang betul.

Kajian ini diteruskan dengan pengoptimuman keadaan percambahan dalam kultivar "Tuan". Kandungan asid fitik telah dikurangkan dengan ketara (1.24 - 99.99%) melalui aktiviti fitase (27.43 -165.30 U/kg) semasa percambahan dan seterusnya meningkatkan bioaksesibiliti mineral (8.03 - 239.56% Ca, 25.38% Fe, 1.18 - 45% Zn). Percambahan dalam keadaan berasid (pH 2) untuk 12 jam adalah keadaan yang paling sesuai untuk peningkatan bioaksesibiliti zat kalsium dan zink pada suhu bilik (25°C). Namun demikian, tempoh percambahan yang lebih panjang (50 jam) diperlukan untuk bioaksesibiliti zat besi yang tinggi. Keadaan percambahan yang telah dioptimumkan pada pH 2.7, 25°C dan 12 jam berjaya mengurangkan kandungan asid fitik, nisbah asid fitik kepada mineral dan memberi peningkatan dalam bioaksesibiliti mineral dalam zat kalsium, besi dan zink.

Kesimpulannya, beras penuh Bario mempunyai kelebihan dalam penghadaman kanji dan protein, tetapi bioaksesibiliti mineral dan sifat antioksidan yang terhad. Kelemahan dalam bioaksesibiliti mineral adalah disebabkan kandungan asid fitik yang tinggi dalam beras penuh. Penguraian asid fitik dalam beras penuh Bario adalah bergantung kepada kultivar dan keadaan percambahan. Keadaan percambahan yang optimum dapat membantu penguraian asid fitik dan memberi bioaksesibiliti mineral yang lebih baik dalam beras penuh Bario terpilih. Beras penuh bercambah yang mempunyai bioaksesibiliti mineral yang lebih tinggi boleh dijadikan alternatif kepada beras penuh untuk pemakanan yang sihat.

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TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF APPENDICES	xviii
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Rice	4
2.1.1 Consumption	4
2.1.2 Production	4
2.1.3 Nutritional value	5
2.1.4 Traditional rice in Sarawak	9
2.2 Phytates and phytic acid	10
2.2.1 Occurrence, chemistry and analysis	11
2.2.2 Dietary intake, digestion and absorption	16
2.2.3 Nutritional impacts of phytic acid	17
2.2.4 Phytate degradation	20
2.3 Strategies to overcome nutritional limitations in rice	24
2.3.1 Fortification	24
2.3.2 Bio-fortification	25
2.3.3 Post-harvest processing/ modifications	26
2.3.4 Diet diversification	28
2.3.5 Supplementation	28
2.4 Germination	28
2.4.1 Physiological and metabolic process	28
2.4.2 Application in food products	30
2.4.3 Nutritional benefits	30
2.4.4 Health benefits	32
3 MATERIALS AND METHODS	34
3.1 Overview	34
3.2 Instrumentation	34
3.3 Materials	35
3.3.1 Samples	35
3.3.2 Chemicals	36
3.4 Analytical methods	40

	3.4.1	Physical and genotypic characterization	40
	3.4.2	Nutrient bioaccessibility	41
	3.4.3	Antioxidant properties	43
	3.4.4	Phytic acid content	45
	3.4.5	Phytase activity	46
	3.4.6	Microscopy analysis	46
3.5		Experimental design	47
	3.5.1	Nutritional profile of Bario rice cultivars	47
	3.5.2	Effect of phytic acid content on the nutritional profile	47
	3.5.3	Effect of germination treatments on phytate degradation	48
	3.5.4	Mineral bioaccessibility in germinated whole grain rice	49
4		RESULTS AND DISCUSSION	51
	4.1	Nutrient bioaccessibility and antioxidant properties of Bario rice cultivars	51
	4.1.1	Nutrient bioaccessibility	51
	4.1.2	Antioxidant properties	57
	4.1.3	Nutritional profiles	62
	4.2	Effect of phytic acid content on nutrient bioaccessibility and antioxidant properties	67
	4.2.1	Nutrient bioaccessibility	67
	4.2.2	Antioxidant properties	71
	4.3	Effect of germination treatments on phytate degradation	74
	4.3.1	Phytic acid content	74
	4.3.2	Phytase activity	78
	4.3.3	Elemental and morphological analysis of phytate globoids	82
	4.4	Response surface analysis of mineral bioaccessibility in germinated whole grain rice	87
	4.4.1	Phytic acid content and phytase activity	87
	4.4.2	Phytic acid to mineral mole ratio	93
	4.4.3	<i>In-vitro</i> mineral bioaccessibility	95
	4.4.4	Optimization of germination conditions	98
5		SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	100
	5.1	Summary	100
	5.2	Conclusion	101
	5.3	Novelty and applications	101
	5.4	Recommendations for future research	102

REFERENCES	103
APPENDICES	133
BIODATA OF STUDENT	143
LIST OF PUBLICATIONS	144



LIST OF TABLES

Table		Page
2.1.	Nutrient composition of medium grain rice and Bario rice	7
2.2.	Phytic acid content in rice grains and rice products	12
2.3.	Studies on germination processing in cereals and legumes	31
3.1.	Climatic conditions and soil series of cultivation sites during cropping period of collected samples (2009 – 2010)	38
3.2.	Bario rice samples	39
3.3.	Size classification of whole grain rice according to Juliano (1993)	40
3.4.	Shape classification of whole grain rice according to Juliano (1993)	40
3.5.	Temperature settings for acid digestion process for minerals analysis	41
3.6.	Germination treatments	48
3.7.	Central composite design for response surface modeling of germination treatments	50
3.8.	Levels of independent factors in germination treatments	50
4.1.	Nutrient bioaccessibility of 30 Bario rice cultivars	52
4.2.	Correlations between mineral content and <i>in vitro</i> starch digestibility	54
4.3.	Correlations between protein content and <i>in vitro</i> protein digestibility	55
4.4.	Phytic acid and mineral content in 30 Bario rice cultivars	56
4.5.	Correlations between mineral content and phytate to mineral mole ratio	57
4.6.	Antioxidant properties of 30 Bario rice cultivars	59
4.7.	Correlations between antioxidant properties of whole grain rice	61
4.8.	Correlations between phytic acid content and nutrient bioaccessibility of 30 Bario rice cultivars	68
4.9.	Pearson's correlation coefficients between phytic acid content and	71

in vitro protein digestibility

4.10.	Correlations between phytic acid content and antioxidant properties of 30 Bario rice cultivars	72
4.11.	Correlations between phytic acid content and antioxidant properties of rice cultivars with different bran color	73
4.12.	Phytase content and phytase activity of selected rice cultivars	75
4.13.	Phytic acid content (g/kg) and its percent changes (%) of selected rice cultivars under different germination treatments	76
4.14.	Phytase activity (U/kg) and its percent changes (%) of selected rice cultivars under different germination treatments	79
4.15.	Phytic acid content of fresh and preheated rice samples (mg/g)	81
4.16.	Phytic acid content (g/kg), phytase activity (U/kg), phytic acid to mineral mole ratio and <i>in vitro</i> mineral bioaccessibility (%) of cultivar Tuan under germination conditions in central composite design	88
4.17.	Correlations between mineral content, mole ratio of phytic acid to mineral and <i>in vitro</i> mineral bioaccessibility in germinated whole grain rice (Tuan)	97
4.18.	Predicted responses from optimized germination conditions	99
4.19.	Comparison of phytic acid content, phytase activity, phytic acid to mineral mole ratio and <i>in vitro</i> mineral bioaccessibility between native and germinated whole grain rice treated at optimized conditions	99

LIST OF FIGURES

Figure		Page
2.1.	The anatomy of rice seed	5
2.2.	Chemical structure of phytic acid	10
2.3.	Conformational structures of phytic acid	13
2.4.	Phytate-protein-starch complex molecule: a potential structure	13
2.5.	Enzymatic degradation of phytate involves the hydrolysis of phosphate ester bonds and resulting inositol, phosphate and divalent elements	21
2.6.	Pathways for phytate hydrolysis by cereal and microbial phytases	23
2.7.	Time course events during seed germination	29
3.1.	Study framework	34
3.2.	Laboratory-scale double rubber roller dehusker provided by BERNAS	35
3.3.	Sampling locations of Bario rice cultivars	37
4.1.	The relationship between mineral (iron and zinc) content and <i>in vitro</i> starch digestibility among 30 Bario rice cultivars	54
4.2.	The relationship between total protein content, undigested protein content and <i>in vitro</i> protein digestibility among 30 Bario rice cultivars	55
4.3.	The comparison of antioxidant properties between pigmented and non-pigmented rice among 30 Bario rice cultivars	58
4.4.	Inhibition of lipid peroxidation of selected rice extracts at different incubation period	62
4.5.	Principal component analysis (PCA): factor loading plot (left) and score plot (right) of the 1st principal component (PC1=TPC, total phenolic content) and the 2nd principal component (PC2=Phy/Fe, phytic acid to mineral mole ratio) describing the variation among the Bario rice cultivars.	64
4.6.	The comparison of nutrient bioaccessibility between highland and lowland rice among 30 Bario rice cultivars	65
4.7.	The comparison of nutrient bioaccessibility between short and medium grains rice among 30 Bario rice cultivars	66

4.8.	The relationship between iron and zinc bioaccessibility and phytic acid content among 30 Bario rice cultivars	68
4.9.	The relationship between calcium bioaccessibility and phytic acid content among 30 Bario rice cultivars	69
4.10.	Available mineral content from iron (1ppm Fe), zinc (1 ppm Zn) and calcium (1 ppm Ca) solutions with different amounts of phytic acid	70
4.11.	The relationship between phytic acid, total protein content and undigested protein content.	71
4.12.	The relationship between phytic acid content and ferrous chelating activity among 30 Bario rice cultivars	73
4.13.	Comparison in phytic acid content (mean \pm standard deviation; mg/g) between fresh and preheated rice samples at different treatment conditions (S1-S8, Table 4.15)	81
4.14.	Energy dispersive X-ray spectra of phytate globoids in scutellum cells (left column) and aleurone cells (right column) under germination conditions: (a) ungerminated native samples, (b) pH 2, 30°C, 25.1 hrs, (c) pH 7, 30°C, 25.1 hrs and (d) pH 12, 30°C, 25.1 hrs	83
4.15.	Comparison of phytate globoids (shown by arrows) in scutellum cells (left column) and aleurone cells (right column) under germination conditions at different pH : (a) ungerminated native samples, (b) pH 2, 30°C, 25.1 hrs, (c) pH 7, 30°C, 25.1 hrs and (d) pH 12, 30°C, 25.1 hrs	84
4.16.	Comparison of phytate globoids (shown by arrows) in scutellum cells (left column) and aleurone cells (right column) under acidic germination conditions: (a) ungerminated native samples, (b) pH 4, 20°C, 40 hrs, (c) pH 4, 40°C, 40 hrs and (d) pH 4, 40°C, 10.2 hrs	85
4.17.	Energy dispersive X-ray spectra of phytate globoids in scutellum cells (left column) and aleurone cells (right column) under germination conditions: (a) ungerminated native samples, (b) pH 4, 20°C, 40 hrs, (c) pH 4, 40°C, 40 hrs and (d) pH 4, 40°C, 10.2 hrs	86
4.18.	Response surface curve of phytic acid content (left) and phytase activity (right) in whole grain rice treated under germination conditions at constant time of 25.1 hours	92
4.19.	Response surface curve of mole ratio of phytic acid to calcium (left), iron (middle) and zinc (right) in whole grain rice treated under germination conditions at constant temperature of 20°C	94

- 4.20 Response surface curve of mole ratio of phytic acid to calcium (left), iron (middle) and zinc (right) in whole grain rice treated under germination conditions at constant temperature of 20°C 96



LIST OF APPENDICES

Appendix		Page
A1	Physical grain traits of 30 rice cultivars from northern Sarawak, Malaysia	134
A2	Antioxidant properties of Bario rice cultivars with different genotypes	135
A3	Antioxidant properties of Bario rice cultivars between cultivation location and physical grain traits	136
A4	Nutrient bioaccessibility of Bario rice cultivars with different genotypes	137
A5	Nutrient bioaccessibility of Bario rice cultivars between cultivation location and physical grain traits	138
A6	Analysis of variance on the effect of germination conditions on response variables in cultivar Tuan	139
B1	Genetic clustering of 30 rice cultivars based on multiplex SSR marker panels (UPGMA, NTSYS-Pc)	140
B2	Elemental profiles of rice cultivars analyzed by energy-dispersive X-ray spectroscopy	141
B3	Comparison of phytate globoids in scutellum cells (left) and aleurone cells (right) of rice cultivars (a) Merah, (b) Tuan, (c) Bario A and (d) Bario Pendek	142

LIST OF ABBREVIATIONS

AACC	American Association of Cereal Chemistry
AAE	Ascorbic Acid Equivalent
AAS	Atomic Absorption Spectrometer
ANOVA	Analysis of Variance
AOAC	Association of Analytical Communities
BERNAS	National Paddy and Rice Board
BIRRI	Bangladesh Research Rice Institute
CCD	Central Composite Design
DNMRT	Duncan New Multiple Range Test
DNA	Deoxyribose Nucleic Acid
DPPH	2,2-diphenyl-1-picrylhydrazyl
EDTA	Ethylenediaminetetraacetic acid
EDX	Energy Dispersive X-Ray
FAO	Federal Agriculture Organization
FTIR	Fourier Transform Infrared Spectroscopy
GABA	Gamma Amino Butyric Acid
GAE	Gallic Acid Equivalent
GC-MS	Gas Chromatography Mass Spectrometer
HPIC	High Performance Ion Chromatography
HPLC	High Performance Liquid Chromatography
ICP-AES	Inductively Coupled Plasma Atomic Emission Spectrometer
ICP-MS	Inductively Coupled Plasma Mass Spectrometer
Ins	Inositol
IRRI	International Rice Research Institute

ISGA	International Sprouted Grains Association
IUB	International Union of Biochemistry
IUPAC	International Union of Pure And Applied Chemistry
kDa	kilo Dalton
kV	kilo Volts
M	Molar
MANOVA	Multivariate Analysis Of Variance
mM	mili molar
mRNA	messenger Ribonucleic Acid
MyIPO	Malaysia Intellectual Property Organization
NCCFN	National Coordinating Committee on Food and Nutrition
NMR	Nuclear Magnetic Resonance
PCA	Principal Component Analysis
PDA	Photodiode array
Phy/Fe	Phytic acid to iron mole ratio
Phy/Zn	Phytic acid to zinc mole ratio
Phy/Ca	Phytic acid to calcium mole ratio
PTFE	Polytetrafluoroethylene
QBSD	Quadrant Back Scattering Detector
SAHN	Sequential Agglomerative Hierarchical Nesting
SAS	Statistical Analysis Software
SEM	Scanning Electron Microscope
USDA	U.S. Department of Agriculture
UV-Vis	Ultraviolet Visible light
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

Rice is the primary source of nutrition in many developing countries including Malaysia. It is a readily available medium for transporting the essential nutrients to the local people to tackle nutritional-related diseases. In spite of that, nutritional limitations that underlies within rice grains are contributing to the growing health problems in developing countries (Potrykus, 2003). Rice is consumed mainly as polished rice containing little nutrients, where beneficial nutrients especially those chemical compounds with biomedical values are reduced significantly during the milling process (Champagne *et al.*, 2004).

Whole grain rice can be a substitute to its processed equivalent for daily consumption with more vitamins, minerals and fiber. Positive associations between whole grain rice consumption and lower incidence of many chronic diseases such as heart disease, obesity, cancer and Type 2 diabetes have been reported (Panlasigui and Thompson, 2006; Dinesh Babu *et al.*, 2009; Slavin, 2013). However, whole grain rice is also reported to be low in minerals, starch and protein bioavailability and its antioxidant potential were not as optimistic as reported. The low availability of vitamins, minerals and health promoting phytochemicals in whole grain rice may not be sufficient for effective nutritional consumption (Grusak, 2002).

Rice improvement programs have included nutritional targets for global health development, aside from milling and cooking quality which dictates higher commercial value. Understanding of associations between rice constituents with desired nutritional quality is important to facilitate the production of rice with higher nutritional quality. Phytic acid which is abundantly present in whole grain rice has been considered to be the major contributor to nutritional drawbacks of the whole grain rice. Its strong ability to complex with multi-charged metal ions, proteins and starches reduces the availability of these nutrients (Yoon *et al.*, 1983; Raboy, 2009). Nevertheless, phytic acid also exhibits beneficial properties such as good anti-cancer agent, inhibitor for renal stone development and also behaves as an antioxidant agent (Thompson, 1993). Therefore, phytic acid could play a significant role in the overall nutritional value of whole grain rice depending on its complexing capability with nutrients in different mediums including its formation, release and breakdown.

Strategies have been made to rectify the nutritional limitations present in rice through plant breeding, food processing and nutritional programme, by increasing the targeted nutrients, reducing nutrient inhibitors and increasing enhancers for the release of nutrients. Modern technology enables alteration at certain compound levels through genetic engineering and marker-assisted breeding (Grusak, 2002; Mackey, 2002). These efforts contribute significantly in the production of rice varieties with a relatively high concentration of minerals, vitamins and antioxidants, but low in inhibitors such as phytic acid (Lucca *et al.*, 2001a, 2001b; Glahn *et al.*, 2002). Low phytic acid (*lpa*) mutant rice lines have been generated by chemical mutation through molecular balance mechanisms, in which the seeds contained normal levels of total phosphorus, but reduced levels of phytic acid phosphorus (Raboy *et al.*, 2000). These forms of research require long experimental periods in the selection of suitable rice lines for safe release

of nutritionally improved rice varieties. Conversely, food-processing techniques offer a more practical and consumer friendly method to counteract the nutritional limitations in rice.

Traditional food processing and preparation practices provide a good potential for nutritional improvement in rice. The traditional food processing methods are easily adaptable to poor resource settings and could be optimized to provide better consumer acceptance. These include thermal processing, mechanical processing, soaking, fermentation, and germination. Thermal processing is not an effective approach to tackle the influence of phytic acid in whole grain rice, due to the heat stable nature of phytic acid. Mechanical processing reduces significantly the phytic acid levels but also removes the nutrient rich bran layer. Soaking, germination, and fermentation which involve incubation of rice in water or inoculums in different time frames may bring down the phytic acid content (Nergiz and Gökğöz, 2007; Liang *et al.*, 2009; Luo *et al.*, 2009). Among these practices, germination outstand the latter two with its advantages in low nutrient losses due to leaching compared to soaking and less off-flavor generation compared to fermentation.

Germination is the natural process of plants to provide nutrients to seedlings. The mechanism of germination activates hydrolytic enzymes for the breakdown of starch, non-starch polysaccharides and proteins, and leads to an increase in readily absorbable oligosaccharides and amino acids (Yang *et al.*, 2001; Rimsten *et al.*, 2003; Shu *et al.*, 2008). The germination process also improves protein and carbohydrate digestibility, increases bioactive compounds, denatures amylase inhibitors and other anti-nutritional factors in whole grains (Egli *et al.*, 2002; Tian *et al.*, 2004; Komatsuzaki *et al.*, 2007). Nevertheless, nutrient density within the germinated grains is highly dependent on the germination conditions such as temperature, pH and time during processing. Optimization of the germination process could maximize the nutritional values output in whole grain cereals and legumes. However, this may vary among grains and varieties.

Sarawak, the land of hornbills is blessed with a great diversity of rice. Bario rice is one of the famous traditional premium rice identified by the Department of Agriculture, Sarawak for its soft texture, fine elongated grains and exquisite taste and aroma. Its unique characteristics dictated a geographical indication award from the Intellectual Property Corporation of Malaysia (MyIPO) in 2009. It is originated from the Kelabit's highlands, known as "Bario" and widely adopted by lowland rice growers in northern Sarawak regions. Bario rice cultivars are reported to be good source of protein and thiamine, moderate glycemic index and also low in fat content (Nicholas *et al.* 2014). However, the information on phytic acid level, nutrient bioaccessibility and antioxidant properties in the Bario rice cultivars is lacking.

Hence, the general objectives of the study were to determine nutritional value of Bario rice in the whole grain, and, identify germination condition for nutritional improvement of the grains.

The specific objectives of this study were:

- a) To determine the nutrient bioaccessibility and antioxidant properties in the Bario rice cultivars.
- b) To identify the effect of the intrinsic phytic acid content on nutrient bioaccessibility and antioxidant properties in the rice.

- c) To study the effect of germination on phytic acid content, phytase activity and phytate globoids in the rice.
- d) To optimize the germination conditions for mineral bioaccessibility improvement in the rice.





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REFERENCES

- AACC. The Definition of Dietary Fiber. *Cereal Foods World* **2001**, *46*, 112–126.
- Adam, A.; Crespy, V.; Levrat-Verny, M.-A.; Leenhardt, F.; Leuillet, M.; Demigne, C.; Remesy, C. The Bioavailability of Ferulic Acid Is Governed Primarily by the Food Matrix Rather than Its Metabolism in Intestine and Liver in Rats 1. *J. Nutr.* **2002**, *132*, 1962–1968.
- Afify, A. E.-M. M. R.; El-Beltagi, H. S.; El-Salam, S. M. A.; Omran, A. A. Bioavailability of Iron, Zinc, Phytate and Phytase Activity during Soaking and Germination of White Sorghum Varieties. *PLoS One* **2011**, *6*, e25512.
- Afify, A. E.-M. M. R.; El-Beltagi, H. S.; El-Salam, S. M. A.; Omran, A. A. Protein Solubility, Digestibility and Fractionation after Germination of Sorghum Varieties. *PLoS One* **2012**, *7*, e31154.
- Agte, V.; Tarwadi, K.; Chiplonkar, S. Phytate Degradation During Traditional Cooking: Significance of the Phytic Acid Profile in Cereal-Based Vegetarian Meals. *J. Food Compos. Anal.* **1999**, *12*, 161–167.
- Ahn, D. J.; Won, J. G.; Rico, C. M.; Lee, S. C. Influence of Variety, Location, Growing Year, and Storage on the Total Phosphorus, Phytate-Phosphorus, and Phytate-Phosphorus to Total Phosphorus Ratio in Rice. *J. Agric. Food Chem.* **2010**, *58*, 3008–3011.
- Ahn, H.-J.; Kim, J.-H.; Jo, C.; Kim, M.-J.; Byun, M.-W. Comparison of Irradiated Phytic Acid and Other Antioxidants for Antioxidant Activity. *Food Chem.* **2004**, *88*, 173–178.
- Albarracín, M.; González, R. J.; Drago, S. R. Effect of Soaking Process on Nutrient Bio-Accessibility and Phytic Acid Content of Brown Rice Cultivar. *LWT - Food Sci. Technol.* **2013**, *53*, 76–80.
- Anderson, M. J.; Whitcomb, P. J. *RSM Simplified: Optimizing Processes Using Response Surface Methods for Design of Experiments*; CRC Press Taylor & Francis Group, 2005.
- Anderson, R. A Contribution to the Chemistry of Phytin. *J. Biol. Chem.* **1912**, 171–190.
- Angel, R.; Tamim, N. M.; Applegate, T. J.; Dhandu, A. S.; Ellestad, L. E. Phytic Acid Chemistry : Influence on Phytin-Phosphorus Availability and Phytase Efficacy 1. *J. Appl. Poult. Res.* **2002**, *11*, 471–480.
- Ayet, G.; Burbano, C.; Cuadrado, C.; Pedrosa, M. M.; Robredo, L. M.; Muzquiz, M.; Cuadra, C. De; Castano, A.; Osagie, A. Effect of Germination , under Different Environmental Conditions , on Saponins , Phytic Acid and Tannins in Lentils (*Lens Culinaris*). *J. Sci. Food Agric.* **1997**, *74*, 273–279.

- Banchuen, J.; Thammarutwasik, P.; Oraikul, B.; Wuttijumnong, P.; Sirivongpaisal, P. Effect of Germinating Processes on Bioactive Component of Sangyod Muang Phatthalung Rice. *Thai J. Agric. Sci.* **2009**, *42*, 191–199.
- Barker, C. J.; Berggren, P. O. Inositol Hexakisphosphate and Beta-Cell Stimulus-Secretion Coupling. *Anticancer Res.* **1999**, *19*, 3737–3741.
- Barrientos, L. G.; Murthy, P. P. N. Conformational Studies of Myo-Inositol Phosphates. *Carbohydrates Res.* **1996**, *296*, 39–54.
- Baruah, K.; Sahu, N. P.; Pal, A. K.; Jain, K. K.; Debnath, D.; Yengkokpam, S. Interactions of Dietary Microbial Phytase, Citric Acid and Crude Protein Level on Mineral Utilization by Rohu, Labeo Rohita (Hamilton), Juveniles. *J. World Aquac. Soc.* **2007**, *38*, 238–249.
- Bau, H.; Villaume, C.; Nicolas, J.; Mejean, L. Effect of Germination on Chemical Composition, Biochemical Constituents and Antinutritional Factors of Soya Bean (Glycine Max) Seeds. *J. Sci. Food Agric.* **1997**, *73*, 1–9.
- Bergman, C.; Gualberto, D. G.; Weber, C. W. Mineral Binding Capacity of Dephytinized Insoluble Fiber from Extruded Wheat, Oat and Rice Brans. *Plant Foods Hum. Nutr.* **1997**, *51*, 295–310.
- Bergman, C.; Xu, Z. Genotype and Environment Effects on Tocopherol, Tocotrienol, and Gamma-Oryzanol Contents of Southern US Rice. *Cereal Chem.* **2003**, *80*, 446–449.
- Bergman, E.-L.; Autio, K.; Sandberg, A. Optimal Conditions for Phytate Degradation, Estimation of Phytase Activity, and Localization of Phytate in Barley (cv. Blenheim). *J. Agric. Food Chem.* **2000**, *48*, 4647–4655.
- Bewley, J. D. Seed Germination and Dormancy. *Plant Cell* **1997**, *9*, 1055–1066.
- Bewley, J. D.; Bradford, K. J.; Hilhorst, H. W. M.; Nonogaki, H. Germination. In *Seeds: Physiology of development, germination and dormancy*; Springer New York: New York, NY, 2013; pp. 133–181.
- Blaabjerg, K.; Hansen-Møller, J.; Poulsen, H. D. High-Performance Ion Chromatography Method for Separation and Quantification of Inositol Phosphates in Diets and Digesta. *J. Chromatogr. B. Analyt. Technol. Biomed. Life Sci.* **2010**, *878*, 347–354.
- Blatny, P.; Kvasnicka, F.; Kenndler, E. Phytic Acid in Cereal Grains, Legumes, and Feeds by Capillary Isotachopheresis. *J. Agric. Food Chem.* **1995**, *43*, 129–133.
- Bohn, L.; Meyer, A. S.; Rasmussen, S. K. Phytate: Impact on Environment and Human Nutrition. A Challenge for Molecular Breeding. *J. Zhejiang Univ. Sci. B* **2008**, *9*, 165–191.

- Bohn, T.; Davidsson, L.; Walczyk, T.; Hurrell, R. F. Phytic Acid Added to White-Wheat Bread Inhibits Fractional Apparent Magnesium Absorption in Humans. *Am. J. Clin. Nutr.* **2004**, *79*, 418–423.
- Bong, W. S.; Tu, P. J.; Yiu., P. H. Physicochemical Properties of 53 Sarawak Rice Cultivars. In *Proceedings of Sarawak Rice Conference 2011*; 2011; p. 26.
- Brinch-Pedersen, H.; Borg, S.; Tauris, B.; Holm, P. B. Molecular Genetic Approaches to Increasing Mineral Availability and Vitamin Content of Cereals. *J. Cereal Sci.* **2007**, *46*, 308–326.
- Bronstein, Y. L.; Cummings, J. L. Neurochemistry of Frontal–subcortical Circuits. In *Frontal– Subcortical Circuits in Psychiatry and Neurological Disorders*; Lichter, D. G.; Cummings, J. L., Eds.; Guilford Press: New York, 2001; pp. 59–91.
- Brune, M.; Rossander-Hulthen, L.; Hallberg, L.; Gleeup, A.; Sandberg, A. Iron Absorption from Bread in Humans: Inhibiting Effects of Cereal Fiber, Phytate and Inositol Phosphates with Different Numbers of Phosphate Groups. *J. Nutr.* **1992**, *122*, 442–449.
- Burbano, C.; Muzquiz, M.; Osagie, a.; Ayet, G.; Cuadrado, C. Determination of Phytate and Lower Inositol Phosphates in Spanish Legumes by HPLC Methodology. *Food Chem.* **1995**, *52*, 321–325.
- Butsat, S.; Siriamornpun, S. Antioxidant Capacities and Phenolic Compounds of the Husk, Bran and Endosperm of Thai Rice. *Food Chem.* **2010**, *119*, 606–613.
- Bye, J. W.; Cowieson, N. P.; Cowieson, A. J.; Selle, P. H.; Falconer, R. J. Dual Effects of Sodium Phytate on the Structural Stability and Solubility of Proteins. *J. Agric. Food Chem.* **2013**, *61*, 290–295.
- Camire, A. L.; Clydesdale, F. M. Effect of pH and Heat Treatment on the Binding of Calcium, Magnesium, Zinc, and Iron to Wheat Bran and Fractions of Dietary Fiber. *J. Food Sci.* **1981**, *46*, 548–551.
- Canan, C.; Delarozza, F.; Casagrande, R.; Baracat, M. M.; Shimokomaki, M.; Ida, E. I. Antioxidant Capacity of Phytic Acid Purified from Rice Bran. *Acta Sci. Technol.* **2012**, *34*, 457–463.
- Capanzana, M. V.; Buckle, K. a. Optimisation of Germination Conditions by Response Surface Methodology of a High Amylose Rice (*Oryza Sativa*) Cultivar. *LWT - Food Sci. Technol.* **1997**, *30*, 155–163.
- Carnovale, E.; Lugaro, E.; Lombardi-boccia, G. Phytic Acid in Faba Bean and Pea: Effect on Protein Availability. *Cereal Chem.* **1988**, *65*, 114–117.
- Centeno, C.; Viveros, A.; Brenes, A.; Canales, R.; Lozano, A.; de la Cuadra, C. Effect of Several Germination Conditions on Total P, Phytate P, Phytase, and Acid

Phosphatase Activities and Inositol Phosphate Esters in Rye and Barley. *J. Agric. Food Chem.* **2001**, *49*, 3208–3215.

Champagne, E. T.; Phillippy, B. Q. Effects of pH on Calcium, Iron and Zinc Solubilities Following *in vitro* Digestion of Soy Protein Isolate. *J. Food Sci.* **1989**, *54*, 587–592.

Champagne, E. T.; Wood, D.; Juliano, B. O.; Bechtel, D. The Rice Grain and Its Gross Composition. In *Rice*; Champagne, E. T., Ed.; American Association of Cereal Chemistry: St. Paul, USA, 2004; pp. 77–107.

Chang, T. Origin, Domestication and Diversification. In *Rice: Origin, History, Technology and Production*; Smith, C. W., Ed.; John Wiley & Sons, Inc., 2003.

Charoenthaikij, P.; Jangchud, K.; Jangchud, A.; Piyachomkwan, K.; Tungtrakul, P.; Prinyawiwatkul, W. Germination Conditions Affect Physicochemical Properties of Germinated Brown Rice Flour. *J. Food Sci.* **2010**, *74*, C658–C665.

Chee, Y. M.; Wong, S. Y.; Yiu, P. H.; Wong, S. C.; Rajan, A. Quality Characteristics and Nutritional Potential of Selected Rice Cultivars from Southern Sarawak. In *9th TWAS Young Scientist Conference 2009 Programme and Abstracts*; 2009; p. 92.

Chen, C. Evaluation of Air Oven Moisture Content Determination Methods for Rough Rice. *Biosyst. Eng.* **2003**, *86*, 447–457.

Chen, Q. C.; Li, B. W. Separation of Phytic Acid and Other Related Inositol Phosphates by High-Performance Ion Chromatography and Its Applications. *J. Chromatogr. A* **2003**, *1018*, 41–52.

Chen, Q. Determination of Phytic Acid and Inositol Pentakisphosphates in Foods by High-Performance Ion Chromatography. *J. Agric. Food Chem.* **2004**, *52*, 4604–4613.

Cheryan, M. Phytic Acid Interactions in Food Systems. *Crit. Rev. Food Sci. Nutr.* **1980**, *13*, 297–335.

Chiou, R. Y.-Y.; Ku, K.-L.; Chen, W.-L. Compositional Characterization of Peanut Kernels after Subjection to Various Germination Times. *J. Agric. Food Chem.* **1997**, *45*, 3060–3064.

Chitra, U.; Singh, U.; Rao, P. V. Phytic Acid, *in vitro* Protein Digestibility, Dietary Fiber, and Minerals of Pulses as Influenced by Processing Methods. *Plant Foods Hum. Nutr.* **1996**, *49*, 307–316.

Chotimarkorn, C.; Benjakul, S.; Silalai, N. Antioxidant Components and Properties of Five Long-Grained Rice Bran Extracts from Commercial Available Cultivars in Thailand. *Food Chem.* **2008**, *111*, 636–641.

- Claver, I. P.; Zhang, H.; Li, Q.; Zhou, H.; Zhu, K. Optimized Conditions of Steeping and Germination and Their Effect on Sorghum [Sorghum Bicolor (L.) Moench] Composition. *Pakistan J. Nutr.* **2010**, *9*, 686–695.
- Dai, F.; Wang, J.; Zhang, S.; Xu, Z.; Zhang, G. Genotypic and Environmental Variation in Phytic Acid Content and Its Relation to Protein Content and Malt Quality in Barley. *Food Chem.* **2007**, *105*, 606–611.
- Debnath, D.; Sahu, N. P.; Pal, A. K.; Baruah, K.; Yengkokpam, S.; Mukherjee, S. C. Present Scenario and Future Prospects of Phytase in Aqua Feed—review. *Asian-Australian J. Anim Sci.* **2005**, *18*, 1800–1812.
- Del Mundo, A. M.; Felix, A.; Narsio, M. H. *Brown Rice Assessment and the Filipino Milled Rice Consumers*; Los Banos, Philippines, 2001.
- Deliliers, L. G.; Servida, G.; Fracchiolla, N. S.; Ricci, C.; Borsotti, C.; Colombo, G. Effects of Inositol Hexaphosphate (IP6) on Human Normal and Leukaemic Hematopoietic Cells. *Br. J. Haematol.* **2002**, *117*, 577–587.
- Deshpande, S. S.; Damodaran, S. Effect of Phytate on Solubility, Activity and Conformation of Trypsin and Chymotrypsin. *J. Food Sci.* **1989**, *54*, 695–699.
- Dexter, P. B. *Rice Fortification For Developing Countries*; Fayetteville, 1998.
- Dinesh Babu, P.; Subhasree, R. S.; Bhagyaraj, R.; Vidhyalakshmi, R. Brown Rice—Beyond the Color Reviving a Lost Health Food - A Review. *Am. J. Agron.* **2009**, *2*, 67–72.
- Dipti, S. S.; Bergman, C.; Indrasari, S. D.; Herath, T.; Hall, R.; Lee, H. H.; Habibi, F.; Bassinello, P. Z.; Graterol, E.; Ferraz, J. P.; *et al.* The Potential of Rice to Offer Solutions for Malnutrition and Chronic Diseases. *Rice (N. Y.)* **2012**, *5*, 16.
- Dost, K.; Tokul, O. Determination of Phytic Acid in Wheat and Wheat Products by Reverse Phase High Performance Liquid Chromatography. *Anal. Chim. Acta* **2006**, *558*, 22–27.
- Dvořáková, M.; Guido, L. F.; Dostálek, P.; Skulilová, Z.; Moreira, M. M.; Barros, A. A. Antioxidant Properties of Free, Soluble Ester and Insoluble-Bound Phenolic Compounds in Different Barley Varieties and Corresponding Malts. *Inst. Brew. Distill.* **2008**, *114*, 27–33.
- Ebizuka, H.; Ihara, M.; Arita, M. Antihypertensive Effect of Alcohol in Spontaneously Hypertensive Rats. *Food Sci. Technol. Res.* **2009**, *15*, 625–630.
- Ebrahimzadeh, M. A.; Nabavi, S. M.; Nabavi, S. F.; Bahramian, F.; Bekhradnia, A. R. Antioxidant and Free Radical Scavenging Activity of *H. Officinalis* L. Var. *Angustifolius*, v. *Odorata*, B. *Hyrcana* and C. *Speciosum*. *Pak. J. Pharm. Sci.* **2008**, *23*, 29–34.

- Egli, I.; Davidsson, L.; Juilerat, M.-A.; Barclay, D.; Hurrell, R. Phytic Acid Degradation in Complementary Foods Using Phytase Naturally Occurring in Whole Grain Cereals. *J. Food Sci.* **2003**, *68*, 1855–1859.
- Egli, I.; Davidsson, L.; Juillerat, M. A.; Barclay, D.; Hurrell, R. F. The Influence of Soaking and Germination on the Phytase Activity and Phytic Acid Content of Grains and Seeds Potentially Useful for Complementary Feeding. *J. Food Sci.* **2002**, *67*, 3484–3488.
- Ekholm, P.; Virkki, L.; Ylinen, M.; Johansson, L. The Effect of Phytic Acid and Some Natural Chelating Agents on the Solubility of Mineral Elements in Oat Bran. *Food Chem.* **2003**, *80*, 165–170.
- Ellis, R.; Kelsay, J. L.; Reynolds, R. D.; Morris, E. R.; Moser, P. B.; Frazer, C. W. Phytate:zinc and Phytate X Calcium:zinc Millimolar Ratios in Self Selected Diets of Americans.,Asian Indian and Nepales. *J. Am. Diet. Assoc* **1987**, *87*, 1043–1047.
- Ellis, R.; Morris, E. R. Relation between Phytate and Trace Metals in Wheat Bran and Soybean. *Cereal Chem.* **1981**, *58*, 367–370.
- Espín, J. C.; García-Conesa, M. T.; Tomás-Barberán, F. a. Nutraceuticals: Facts and Fiction. *Phytochemistry* **2007**, *68*, 2986–3008.
- Fardet, A.; Rock, E.; Rémésy, C. Is the *in vitro* Antioxidant Potential of Whole-Grain Cereals and Cereal Products Well Reflected in Vivo? *J. Cereal Sci.* **2008**, *48*, 258–276.
- Ferry, S.; Matsuda, M.; Yoshida, H.; Hirata, M. Inositol Hexakisphosphate Blocks Tumor Cell Growth by Activating Apoptotic Machinery as Well as by Inhibiting the Akt/NFkappaB-Mediated Cell Survival Pathway. *Carcinogenesis* **2002**, *23*, 2031–2041.
- Ficco, D. B. M.; Riefolo, C.; Nicastro, G.; De Simone, V.; Di Gesù, A. M.; Beleggia, R.; Platani, C.; Cattivelli, L.; De Vita, P. Phytate and Mineral Elements Concentration in a Collection of Italian Durum Wheat Cultivars. *F. Crop. Res.* **2009**, *111*, 235–242.
- Finch-Savage, W. E.; Leubner-Metzger, G. Seed Dormancy and the Control of Germination. *New Phytol.* **2006**, *171*, 501–523.
- Finney, P. L. Effect of Germination on Cereal and Legume Nutrient Changes and Food or Feed Value : A Comprehensive Review. *Recent Adv. Phytochem.* **1982**, *17*, 229–305.
- Fox, C. H.; Eberl, M. Phytic Acid (IP6), Novel Broad Spectrum Anti-Neoplastic Agent: A Systematic Review. *Complement. Ther. Med.* **2002**, *10*, 229–234.

- Fredlund, K.; Isaksson, M.; Rossander-Hulthén, L.; Almgren, A.; Sandberg, A.-S. Absorption of Zinc and Retention of Calcium: Dose-Dependent Inhibition by Phytate. *J. Trace Elem. Med. Biol.* **2006**, *20*, 49–57.
- Friedman, M. Nutritional Value of Proteins from Different Food Sources . A Review. *J. Agric. Food Chem.* **1996**, *44*, 6–29.
- Frontela, C.; García-Alonso, F. J.; Ros, G.; Martínez, C. Phytic Acid and Inositol Phosphates in Raw Flours and Infant Cereals: The Effect of Processing. *J. Food Compos. Anal.* **2008b**, *21*, 343–350.
- Frontela, C.; Haro, J. F.; Ros, G.; Martínez, C. Effect of Dephytinization and Follow-on Formula Addition on *in vitro* Iron, Calcium, and Zinc Availability from Infant Cereals. *J. Agric. Food Chem.* **2008a**, *56*, 3805–3811.
- Galicia, L.; Nurit, E.; Rosales, A.; Palacios-Rojas, N. *Maize Nutrition Quality and Plant Tissue Analysis Laboratory Laboratory Protocols 2008*; International Maize and Wheat Improvement Center (CIMMYT): Mexico, 2009.
- García-Villanova, R.; García-Villanova, R.; Lope, C. R. de. Determination of Phytic Acid by Complexometric Titration of Excess of Iron (III). *RSC* **1982**, 1503–1506.
- Gbadamosi, S. O.; Abiose, S. H.; Aluko, R. E. Amino Acid Profile, Protein Digestibility, Thermal and Functional Properties of Conophor Nut (*Tetracarpidium Conophorum*) Defatted Flour, Protein Concentrate and Isolates. *Int. J. Food Sci. Technol.* **2012**, *47*, 731–739.
- Gibson, R. S. Content and Bioavailability of Trace Elements in Vegetarian Diets. *Am. J. Clin. Nutr.* **1994**, *59*, 1223–1232.
- Gibson, R. S.; Ferguson, E. L. Assessment of Dietary Zinc in a Population. *Am. J. Clin. Nutr.* **1998**, *68*, 430–434.
- Gibson, R. S.; Hotz, C. Dietary Diversification/modification Strategies to Enhance Micronutrient Content and Bioavailability of Diets in Developing Countries. *Br. J. Nutr.* **2007**, *85*, S159.
- Gilani, G. S.; Chao, W. X.; Cockell, K. A. Impact of Antinutritional Factors in Food Proteins on the Digestibility of Protein and the Bioavailability of Amino Acids and on Protein Quality. *Br. J. Nutr.* **2012**, *108 Suppl*, S315–S332.
- Gilani, G. S.; Cockell, K. A.; Sepehr, E. Effects of Antinutritional Factors on Protein Digestibility and Amino Acid Availability in Foods. *J. AOAC Int.* **2005**, *88*, 967–987.
- Glahn, R. P.; Cheng, Z.; Welch, R. M.; Gregorio, G. B. Comparison of Iron Bioavailability from 15 Rice Genotypes: Studies Using an *in vitro* Digestion/caco-2 Cell Culture Model. *J. Agric. Food Chem.* **2002**, *50*, 3586–3591.

- Gordon, D. T.; Stoops, D.; Ratliff, V. Dietary Fiber and Mineral Nutrition. In *Dietary fiber in health and disease.*; Kritchevsky, D.; Bonfield., C., Eds.; 1995.
- Goto, H.; Tomono, Y.; Ajiro, K.; Kosako, H.; Fujita, M.; Sakurai, M.; Okawa, K.; Iwamatsu, A.; Okigaki, T.; Takahashi, T.; *et al.* Identification of a Novel Phosphorylation Site on Histone H3 Coupled with Mitotic Chromosome Condensation. *J. Biol. Chem.* **1999**, *274*, 25543–25549.
- Graf, E.; Eaton, J. W. J. W. Antioxidant Functions of Phytic Acid. *Free Radic. Biol. Med.* **1990**, *8*, 61–69.
- Graf, E.; Eaton, J. W. J. W. Dietary Suppression of Colonic Cancer: Fiber or Phytate. *Cancer* **1985**, *56*, 717–718.
- Grases, F.; Costa-Bauza, A.; Prieto, R. M. Renal Lithiasis and Nutrition. *Nutr. J.* **2006**, *5*, 23.
- Grases, F.; Perelló, J.; Isern, B.; Prieto, R. M. Determination of *Myo*-Inositol Hexakisphosphate (phytate) in Urine by Inductively Coupled Plasma Atomic Emission Spectrometry. *Anal. Chim. Acta* **2004**, *510*, 41–43.
- Grases, F.; Simonet, B. M.; Prieto, R. M.; March, J. G. Dietary Phytate and Mineral Bioavailability. *J. Trace Elem. Med. Biol.* **2001**, *15*, 221–228.
- Gregorio, G. B.; Senadhira, D.; Htut, H.; Graham, R. D. Breeding for Trace Mineral Density in Rice. *Food Nutr. Bull.* **2000**, *21*, 382–386.
- Greiner, R.; Konietzny, U.; Jany, K. D. Purification and Characterization of Two Phytases from *Escherichia Coli*. *Arch. Biochem. Biophys.* **1993**, *303*, 107–113.
- Greiner, R.; Konietzny, U.; Jany, K. Phytate – an Undesirable Constituent of Plant-Based Foods ? *J. für Ernährungsmedizin* **2006**, *8*, 18–28.
- Grewal, P. K.; Sangha, J. K. Effect of Processing on Thiamin and Riboflavin Contents of Some High Yielding Rice Varieties of Punjab. *J. Sci. Food Agric.* **1990**, *52*, 387–391.
- Grusak, M. A. Enhancing Mineral Content in Plant Food Products. *J. Am. Coll. Nutr.* **2002**, *21*, 178S – 183S.
- Haard, N. F.; Chism, G. W. Characteristics of Edible Plant Tissues. In *Food Chemistry*; Fennema, O. W., Ed.; Marcel Dekker: New York, 1996; pp. 944–1011.
- Hallberg, L.; Brune, M.; Rossander, L. Iron Absorption in Man: Ascorbic Acid and Dose-Dependent Inhibition by Phytate. *Am. J. Clin. Nutr.* **1989**, *49*, 140–144.
- Hallberg, L.; Rossander, L.; Skanberg, A. B. Phytates and the Inhibitory Effect of Bran on Iron Absorption in Man. *Am. J. Clin. Nutr.* **1987**, *45*, 988–996.

- Hamada, J. S. Scale-up Potential of Ion-Pair High-Performance Liquid Chromatography Method to Produce Biologically Active Inositol Phosphates. *J. Chromatogr. A* **2002**, *944*, 241–248.
- Hapuarachchi, S.; Gooneratne, J.; Kumarapperuma, S. C. *Effect of Parboiling on the Reduction of Phytates in Different Varieties of Rice.*; Sri Lanka, 2003; p. 208.
- Harland, B. F.; Morris, E. R. Phytate: A Good or A Bad Food Component? *Nutr. Res.* **1995**, *15*, 733–754.
- Harland, B. F.; Oberleas, D. Anion-Exchange Method for Determination of Phytate in Foods: Collaborative Study. *J. Assoc. Off. Anal. Chem.* **1986**, *69*, 667–670.
- Hayakawa, T.; Toma, Y.; Igaue, I. Purification and Characterization of Acid Phosphatases with or without Phytase Activity from Rice Bran. *Agric. Biol. Chem.* **1989**, *53*, 1475–1483.
- He, D.; Yang, P. Proteomic of Rice Seed Germination. *Front. Plant Sci.* **2013**, *4*, 1–9.
- He, Z.; Honeycutt, C. W.; Zhang, T.; Bertsch, P. M. Preparation and FT-IR Characterization of Metal Phytate Compounds. *J. Environ. Qual.* **2006**, *35*, 1319–1328.
- Hemalatha, S.; Gautam, S.; Platel, K.; Srinivasan, K. Influence of Exogenous Iron, Calcium, Protein and Common Salt on the Bioaccessibility of Zinc from Cereals and Legumes. *J. Trace Elem. Med. Biol.* **2009**, *23*, 75–83.
- Hemalatha, S.; Platel, K.; Srinivasan, K. Influence of Heat Processing on the Bioaccessibility of Zinc and Iron from Cereals and Pulses Consumed in India. *J. Trace Elem. Med. Biol.* **2007a**, *21*, 1–7.
- Hemalatha, S.; Platel, K.; Srinivasan, K. Zinc and Iron Contents and Their Bioaccessibility in Cereals and Pulses Consumed in India. *Food Chem.* **2007b**, *102*, 1328–1336.
- Hotz, C.; McClafferty, B. From Harvest to Health: Challenges for Developing Biofortified Staple Foods and Determining Their Impact on Micronutrient Status. *Food Nutr. Bull.* **2007**, *28*, 271–279.
- Houde, R. L.; Alli, I.; Kermasha, S. Purification and Characterisation of Canola Seed (Brassica Sp.) Phytase. *J. Food Biochem.* **1990**, *114*, 331–351.
- Howell, K. a; Narsai, R.; Carroll, A.; Ivanova, A.; Lohse, M.; Usadel, B.; Millar, a H.; Whelan, J. Mapping Metabolic and Transcript Temporal Switches during Germination in Rice Highlights Specific Transcription Factors and the Role of RNA Instability in the Germination Process. *Plant Physiol.* **2009**, *149*, 961–980.
- Hsu, T.; Kise, M.; Wang, M.; Ito, Y.; Yang, M.; Aoto, H.; Yoshihara, R.; Yokoyama, J.; Kunii, D. Effects of Pre-Germinated Brown Rice on Blood Glucose and Lipid

Levels in Free-Living Patients with Impaired Fasting Glucose or Type 2 Diabetes. *J. Nutr. Sci. Vitaminol. (Tokyo)*. **2008**, *54*, 163–168.

Hu, H.; Wise, A.; Henderson, C. Hydrolysis of Phytate and Inositol Tri-, Tetra- and Penta- Phosphates by the Intestinal Mucosa of the Pig. *Nutr. Res.* **1996**, *16*, 781–787.

Huang, D.; Ou, B.; Prior, R. L. The Chemistry behind Antioxidant Capacity Assays. *J. Agric. Food Chem.* **2005**, *53*, 1841–1856.

Hudson, E.; Dinh, P.; Kokubun, T.; Simmonds, M.; Gescher, A. Characterization of Potentially Chemopreventive Phenols in Extracts of Brown Rice That Inhibit the Growth of Human Breast and Colon Cancer Cells. *Cancer Epidemiol Biomarkers Prev* **2000**, *9*, 1163–1170.

Imam, M. U.; Azmi, N. H.; Bhangar, M. I.; Ismail, N.; Ismail, M. Antidiabetic Properties of Germinated Brown Rice: A Systematic Review. *Evidence-based Complement. Altern. Med.* **2012**, 2012.

Imam, M. U.; Ismail, M. Nutrigenomic Effects of Germinated Brown Rice and Its Bioactives on Hepatic Gluconeogenic Genes in Type 2 Diabetic Rats and HEPG2 Cells. *Mol. Nutr. Food Res.* **2013**, *57*, 401–411.

Iqbal, S.; Bhangar, M. I.; Anwar, F. Antioxidant Properties and Components of Some Commercially Available Varieties of Rice Bran in Pakistan. *Food Chem.* **2005**, *93*, 265–272.

Iqbal, T. H.; Lewis, K. O.; Cooper, B. T. Phytase Activity in the Human and Rat Small Intestine. *Gut* **1994**, *35*, 1233–1236.

Isabelle, M.; Chan, P.; Wijaya, S. Y. *Report on Regulatory Status of Micronutrient Fortification in Southeast Asia*; Singapore, 2011.

ISGA. The History of Sprouts and Their Nutritional Value <http://www.isga-sprouts.org/about-sprouts/sprout-history/> (accessed Jan 10, 2015).

Islam, M. A.; Becerra, J. X. Analysis of Chemical Components Involved in Germination Process of Rice Variety Jhapra. *J. Sci. Res.* **2011**, *4*, 251–262.

Iwai, T.; Takahashi, M.; Oda, K.; Terada, Y.; Yoshida, K. T. Dynamic Changes in the Distribution of Minerals in Relation to Phytic Acid Accumulation during Rice Seed Development. *Plant Physiol.* **2012**, *160*, 2007–2014.

Izquierdo-Pulido, M. I.; Haard, T. A.; Hung, J.; Haard, N. F. Oryzacystatin and Other Protease Inhibitors in Rice Grain: Potential Use in Preventing Proteolysis in Surimi and Other Fish Products. *J. Agric. Food Chem.* **1994**, *42*, 616.

Jacobs, D.; Pereira, M.; Meyer, K.; Kushi, L. Fiber from Whole Grains, but Not Refined Grains, Is Inversely Associated with All Cause Mortality in Older

- Women: The Iowa Women's Health Study. *J. Am. Coll. Nutr.* **2000**, *19*, 326–330.
- Jariwalla, R. J.; Sabin, R.; Lawson, S.; Herman, Z. S. Lowering of Serum Cholesterol and Triglycerides and Modulation of Divalent Cations by Dietary Phytate. *J. Appl. Nutr.* **1990**, *42*, 18–28.
- Jayadeep, A.; Malleshi, N. G. Nutrients, Composition of Tocotrienols, Tocopherols, and Γ -Oryzanol, and Antioxidant Activity in Brown Rice before and after Biotransformation Nutrientes, Composición de Tocotrienoles, Tocoferoles Y Γ -Oryzanol, Y Actividad Antioxidante Del Arroz Integra. *CyTA - J. Food* **2011**, *9*, 82–87.
- Jenab, M.; Thompson, L. U. Phytic Acid in Wheat Bran Affects Colon Morphology , Cell Differentiation and Apoptosis Wheat Bran (WB) and Its Component Phytic Acid (PA) Have Carcinogenesis , I . E . the PCNA Labeling Index of Cell Proliferation and Certain Aberrant Crypt Foci Para. *Carcinogenesis* **2000**, *21*, 1547–1552.
- Jiang, S. L.; Wu, J. G.; Feng, Y.; Yang, X. E.; Shi, C. H. Correlation Analysis of Mineral Element Contents and Quality Traits in Milled Rice (*Oryza sativa* L.). *J. Agric. Food Chem.* **2007**, *55*, 9608–9613.
- Jiang, S. L.; Wu, J. G.; Thang, N. B.; Feng, Y.; Yang, X. E.; Shi, C. H. Genotypic Variation of Mineral Elements Contents in Rice (*Oryza sativa* L.). *Eur. Food Res. Technol.* **2008**, *228*, 115–122.
- John, A. Why is upland rice farming is a big deal. *Oryza*. 2014. <http://www.oryza.com>
- Jonnalagadda, S. S.; Harnack, L.; Liu, R. H.; Mckeown, N.; Seal, C.; Liu, S.; Fahey, G. C. Putting the Whole Grain Puzzle Together : Health Benefits Associated with Whole Grains — Summary of American Society for Nutrition 2010 Satellite Symposium. *J. Nutr.* **2011**, 1011–1022.
- Juliano, B. O. *Rice Chemistry and Quality*; Philippine Rice Research Institute, 2007.
- Juliano, B. O. *Rice in Human Nutrition*; Food and Agriculture Organization of the United States: Rome, Italy, 1993.
- Kahlon, T. S.; Chow, F. I.; Chiu, M. M.; Hudson, C. C.; Sayre, R. N. Cholesterol-Lowering by Rice Bran and Rice Bran Oil Unsaponifiable Matter in Hamsters. *Cereal Chem.* **1996**, *73*, 69–74.
- Karladee, D.; Suriyong, S. Γ -Aminobutyric Acid (GABA) Content in Different Varieties of Brown Rice during Germination. *ScienceAsia* **2012**, *38*, 13–17.
- Kasim, A. B.; Edwards Jr, H. M. The Analysis for Inositol Phosphate Forms in Feed Ingredients. *J. Sci. Food Agric.* **1998**, *76*, 1–9.

- Katayama, T. Effects of Dietary *Myo*-Inositol or Phytic Acid on Hepatic Concentrations of Lipids and Hepatic Activities of Lipogenic Enzymes in Rats Fed on Corn Starch or Sucrose. *Nutr. Res.* **1997**, *17*, 721–728.
- Kaufman, H. W.; Kleinberg, I. Effect of pH on Calcium Binding by Phytic Acid and Its Inositol Phosphoric Acid Derivatives and on the Solubility of Their Calcium Salts. *Arch. Oral Biol.* **1971**, *16*, 445–460.
- Khattak, A. B.; Zeb, A.; Bibi, N.; Khalil, S. A.; Khattak, M. S. Influence of Germination Techniques on Phytic Acid and Polyphenols Content of Chickpea (*Cicer Arietinum* L.) Sprouts. *Food Chem.* **2007**, *104*, 1074–1079.
- Kies, A. K.; De Jonge, L. H.; Kemme, P. a; Jongbloed, A. W. Interaction between Protein, Phytate, and Microbial Phytase. *In vitro* Studies. *J. Agric. Food Chem.* **2006**, *54*, 1753–1758.
- Kies, A. K.; Selle, P. A Review of the Antinutritional Effects of Phytic Acid on Protein Utilisation by Broilers. In *Proc. Aust. Poult. Sci. Sym.* 1998; 1998.
- Kiing, S.-C.; Yiu, P.-H.; Rajan, A.; Wong, S.-C. Effect of Germination on γ -Oryzanol Content of Selected Sarawak Rice Cultivars. *Am. J. Appl. Sci.* **2009**, *6*, 1658–1661.
- Kim, T. W.; Lei, X. G. An Improved Method for a Rapid Determination of Phytase Activity in Animal Feed. *J. Anim. Sci.* **2005**, *83*, 1062–1067.
- Komatsuzaki, N.; Tsukahara, K.; Toyoshima, H.; Suzuki, T.; Shimizu, N.; Kimura, T. Effect of Soaking and Gaseous Treatment on GABA Content in Germinated Brown Rice. *J. Food Eng.* **2007**, *78*, 556–560.
- Konietzny, U.; Greiner, R. Molecular and Catalytic Properties of Phytate-Degrading Enzymes (phytases). *Int. J. Food Sci. Technol.* **2002**, *37*, 791–812.
- Koornneef, M.; Bentsink, L.; Hilhorst, H. Seed Dormancy and Germination. *Curr. Opin. Plant Biol.* **2002**, *5*, 33–36.
- Kouakou, B.; Sallert Alexis, K. K.; Adjehi, D.; Marcelin, D. K.; Dago, G. Biochemical Changes Occurring During Germination and Fermentation of Millet and Effect of Technological Processes on Starch Hydrolysis by the Crude Enzymatic Extract of Millet. *J. Appl. Sci. Res.* **2008**, *4*, 1502–1510.
- Kumar, V.; Sinha, A. K.; Makkar, H. P. S.; Becker, K. Dietary Roles of Phytate and Phytase in Human Nutrition: A Review. *Food Chem.* **2010**, *120*, 945–959.
- Lai, K.; Kueh. Food Security in Sarawak-Malaysia. In *ASEAN Regional Conference on Food Security*; Penang, Malaysia, 2013.
- Lantsch, H. J.; Scheuermann, S. E.; Menke, K. H. Gastrointestinal Hydrolysis of Phytate from Wheat, Barley and Corn in Young-Pigs. *J. Anim. Physiol. Anim. Nutr. (Berl)*. **1988**, *59*, 273–284.

- Laokuldilok, T.; Shoemaker, C. F.; Jongkaewwattana, S.; Tulyathan, V. Antioxidants and Antioxidant Activity of Several Pigmented Rice Brans. *J. Agric. Food Chem.* **2011**, *59*, 193–199.
- Larsson, M.; Rossander-Hulthén, L.; Sandström, B.; Sandberg, A. Improved Zinc and Iron Absorption from Breakfast Meals Containing Malted Oats with Reduced Phytate Content. *Br. J. Nutr.* **1996**, *76*, 677–688.
- Latifah, S. Y.; Armania, N.; Tze, T. H.; Azhar, Y.; Nordiana, A. H.; Norazalina, S.; Hairuszah, I.; Saidi, M.; Maznah, I. Germinated Brown Rice (GBR) Reduces the Incidence of Aberrant Crypt Foci with the Involvement of Beta-Catenin and COX-2 in Azoxymethane-Induced Colon Cancer in Rats. *Nutr. J.* **2010**, *9*, 16.
- Latta, M.; Eskin, M. A Simple and Rapid Colorimetric Method for Phytate Determination. *J. Agric. Food Chem.* **1980**, *28*, 1313–1315.
- Lee, K. Y.; Lee, S.; Lee, H. G. Effect of the Degree of Enzymatic Hydrolysis on the Physicochemical Properties and *in vitro* Digestibility of Rice Starch. *Food Sci. Biotechnol.* **2010**, *19*, 1333–1340.
- Lee, Y. R.; Kim, C. E.; Kang, M. Y.; Nam, S. H. Cholesterol-Lowering and Antioxidant Status-Improving Efficacy of Germinated Giant Embryonic Rice (*Oryza Sativa* L.) in High Cholesterol-Fed Rats. *Ann. Nutr. Metab.* **2007**, *51*, 519–526.
- Lee, Y. R.; Nam, S. H.; Kang, M. Y. Hypoglycemic Effect of the Giant Embryonic Rice Supplementation on Streptozotocin-Induced Diabetic Rats. *Korean J. Food Sci. Technol.* **2006**, *38*, 427–431.
- Lee, Y. R.; Nam, S. H.; Kang, M. Y. Hypoglycemic Effect of the Giant Embryonic Rice Supplementation on Streptozotocin-Induced Diabetic Rats. *Korean J. Food Sci. Technol.* **2006**, *38*, 427–431.
- Leenhardt, F.; Levrat-Verny, M.-A.; Chanliaud, E.; Rémésy, C. Moderate Decrease of pH by Sourdough Fermentation Is Sufficient to Reduce Phytate Content of Whole Wheat Flour through Endogenous Phytase Activity. *J. Agric. Food Chem.* **2005**, *53*, 98–102.
- Leeson, S. Recent Advances in Fat Utilisation by Poultry. In *Recent Advances in Animal Nutrition in Australia*; The University of New England: Armidale, NSW, 1993; pp. 1700–1981.
- Lehrfeld, J. High-Performance Liquid Chromatography Analysis of Phytic Acid on a pH Stable, Macroporous Polymer Column. *Cereal Chem.* **1989**, *66*, 510–515.
- Lehrfeld, J. HPLC Separation and Quantitation of Phytic Acid and Some Inositol Phosphates in Foods: Problems and Solutions. *J. Agric. Food Chem.* **1994**, *42*, 2726–2731.

- Lei, X. G.; Porres, J. M. Phytase Enzymology, Applications, and Biotechnology. *Biotechnol. Lett.* **2003**, *25*, 1787–1794.
- Lestienne, I.; Besançon, P.; Caporiccio, B.; Lullien-Péllierin, V.; Tréche, S. Iron and Zinc *in vitro* Availability in Pearl Millet Flours (*Pennisetum Glaucum*) with Varying Phytate, Tannin, and Fiber Contents. *J. Agric. Food Chem.* **2005**, *53*, 3240–3247.
- Li, J.; Chen, Z.; Guan, X.; Liu, J.; Zhang, M.; Xu, B. Optimization of Germination Conditions to Enhance Hydroxyl Radical Inhibition by Water-Soluble Protein from Stress Millet. *J. Cereal Sci.* **2008**, *48*, 619–624.
- Liang, J.; Han, B.; Han, L.; Nout, M. J. R.; Hamer, R. J. Iron , Zinc and Phytic Acid Content of Selected Rice Varieties from China. *J. Sci. Food Agric.* **2007**, *87*, 504–510.
- Liang, J.; Han, B.-Z.; Nout, M. J. R.; Hamer, R. J. Effect of Soaking and Phytase Treatment on Phytic Acid, Calcium, Iron and Zinc in Rice Fractions. *Food Chem.* **2009**, *115*, 789–794.
- Liang, J.; Han, B.-Z.; Nout, M. J. R.; Hamer, R. J. Effects of Soaking, Germination and Fermentation on Phytic Acid, Total and *in vitro* Soluble Zinc in Brown Rice. *Food Chem.* **2008**, *110*, 821–828.
- Liang, J.; Han, B.-Z.; Nout, M. J. R.; Hamer, R. J. Effects of Soaking, Germination and Fermentation on Phytic Acid, Total and *in vitro* Soluble Zinc in Brown Rice. *Food Chem.* **2008a**, *110*, 821–828.
- Liang, J.; Li, Z.; Tsuji, K.; Nakano, K.; Nout, M. J. R.; Hamer, R. J. Milling Characteristics and Distribution of Phytic Acid and Zinc in Long-, Medium- and Short-Grain Rice. *J. Cereal Sci.* **2008b**, *48*, 83–91.
- Liu, K.; Gu, Z. Selenium Accumulation in Different Brown Rice Cultivars and Its Distribution in Fractions. *J. Agric. Food Chem.* **2009**, *57*, 695–700.
- Liu, Q.; Yao, H. Antioxidant Activities of Barley Seeds Extracts. *Food Chem.* **2007**, *102*, 732–737.
- Liu, Z.; Cheng, F.; Zhang, G. Grain Phytic Acid Content in Japonica Rice as Affected by Cultivar and Environment and Its Relation to Protein Content. *Food Chem.* **2005**, *89*, 49–52.
- Loewus, F. a.; Murthy, P. P. N. *Myo*-Inositol Metabolism in Plants. *Plant Sci.* **2000**, *150*, 1–19.
- Lonnerdal, B. Phytic Acid – Trace Element (Zn , Cu , Mn) Interactions. *Int. J. Food Sci. Technol.* **2002**, *37*, 749–758.

- Lopez, H. W.; Leenhardt, F.; Coudray, C.; Remesy, C. Minerals and Phytic Acid Interactions: Is It a Real Problem for Human Nutrition? *Int. J. Food Sci. Technol.* **2002**, *37*, 727–739.
- Lott, J. N. A.; Ockenden, I.; Raboy, V.; Batten, G. D. A Global Estimate of Phytic Acid and Phosphorus in Crop Grains, Seeds, and Fruits. In *Food Phytates*; 2002.
- Lott, J. N. A.; Spitzer, E. X-Ray Analysis Studies of Elements Stored in Protein Body Globoid Crystals of Triticum Grains. *Plant Physiol.* **1980**, *66*, 494–499.
- Lu, B.; Snow, A. Gene Flow from Genetically Modified Rice and Its Environmental Consequences. *Bioscience* **2005**, *55*, 669–678.
- Lu, L.; Tian, S.; Liao, H.; Zhang, J.; Yang, X.; Labavitch, J. M.; Chen, W. Analysis of Metal Element Distributions in Rice (*Oryza Sativa* L.) Seeds and Relocation during Germination Based on X-Ray Fluorescence Imaging of Zn, Fe, K, Ca, and Mn. *PLoS One* **2013**, *8*, e57360.
- Lu, Z.-H.; Zhang, Y.; Li, L.-T.; Curtis, R. B.; Kong, X.-L.; Fulcher, R. G.; Zhang, G.; Cao, W. Inhibition of Microbial Growth and Enrichment of Gamma-Aminobutyric Acid during Germination of Brown Rice by Electrolyzed Oxidizing Water. *J. Food Prot.* **2010**, *73*, 483–487.
- Lucca, P.; Hurrell, R.; Potrykus, I. Approaches to Improving the Bioavailability and Level of Iron in Rice Seeds. *J. Sci. Food Agric.* **2001a**, *81*, 828–834.
- Lucca, P.; Hurrell, R.; Potrykus, I. Genetic Engineering Approaches to Improve the Bioavailability and the Level of Iron in Rice Grains. *TAG Theor. Appl. Genet.* **2001b**, *102*, 392–397.
- Luo, Y.; Gu, Z.; Han, Y.; Chen, Z. The Impact of Processing on Phytic Acid, *in vitro* Soluble Iron and Phy/Fe Molar Ratio of Faba Bean (*Vicia Faba* L.). *J. Sci. Food Agric.* **2009**, *89*, 861–866.
- Luo, Y.; Xie, W.; Jin, X.; Zhang, B.; Wang, Q.; He, Y. The Impact of Processing on Phytic Acid, *in vitro* Soluble Zinc and Phy / Zn Molar Ratio of Faba Bean (*Vicia Faba* L.). *Int. Food Res. J.* **2013**, *20*, 1285–1291.
- Ma, G.; Jin, Y.; Piao, J.; Kok, F.; Guusje, B.; Jacobsen, E. Phytate, Calcium, Iron, and Zinc Contents and Their Molar Ratios in Foods Commonly Consumed in China. *J. Agric. Food Chem.* **2005**, *53*, 10285–10290.
- Mackey, M. The Application of Biotechnology to Nutrition: An Overview. *J. Am. Coll. Nutr.* **2002**, *21*, 157S – 160S.
- Maenz, D. D.; Engele-Schaan, C. M.; Newkirk, R. W.; Classen, H. L. The Effect of Minerals and Mineral Chelators on the Formation of Phytase-Resistant and Phytase-Susceptible Forms of Phytic Acid in Solution and in a Slurry of Canola Meal. *Anim. Feed Sci. Technol.* **1999**, *81*, 177–192.

- Maga, J. A. Phytate: Its Chemistry, Occurrence, Food Interactions, Nutritional Significance and Methods of Analysis. *J. Agric. Food Chem.* **1982**, *30*, 1–9.
- Mamiya, T.; Kise, M.; Morikawa, K.; Aoto, H.; Ukai, M.; Noda, Y. Effects of Pre-Germinated Brown Rice on Depression-like Behavior in Mice. *Pharmacol. Biochem. Behav.* **2007**, *86*, 62–67.
- March, J. G.; Simonet, B. M.; Grases, F. Determination of Phytic Acid by Gas Chromatography-Mass Spectroscopy: Application to Biological Samples. *J. Chromatogr. B. Biomed. Sci. Appl.* **2001**, *757*, 247–255.
- Martin, R. C.; Pluskota, W. E.; Nonogaki, H. Plant Developmental Biology - Biotechnological Perspectives. In *Plant Developmental Biology - Biotechnological Perspectives: Volume 1*; Pua, E. C.; Davey, M. R., Eds.; Springer Berlin Heidelberg: Berlin, Heidelberg, 2010; Vol. 1, pp. 383–404.
- Martinez-Camacho, J.; Vara, L. G. Ia; Hamabata, A.; Mora-Escobedo, R.; Calderon-Salinas, V. A pH-Stating Mechanism in Isolated Wheat (*Triticum Aestivum*) Aleurone Layers Involves Malic Acid Metabolism. *J. Plant Physiol.* **2004**, *161*, 1289–1298.
- Matsuyama, A.; Yoshimura, K.; Shimizu, C.; Murano, Y.; Takeuchi, H.; Ishimoto, M. Characterization of Glutamate Decarboxylase Mediating Gamma-Amino Butyric Acid Increase in the Early Germination Stage of Soybean (*Glycine Max* [L.] Merr). *J. Biosci. Bioeng.* **2009**, *107*, 538–543.
- Maugenest, S.; Martinez, I.; Godin, B.; Perez, P.; Lescure, A. M. Structure of Two Maize Phytase Genes and Their Spatio-Temporal Expression during Seedling Development. *Plant Mol. Biol.* **1999**, *39*, 503–514.
- Mazzola, E. P.; Phillippy, B. Q.; Harland, B. F.; Miller, T. H.; Potemra, J. M.; Katsimpiris, E. W. Phosphorus-31 Nuclear Magnetic Resonance Spectroscopic Determination of Phytate in Foods. *J. Agric. Food Chem.* **1986**, *34*, 60–62.
- McDonough, F. E.; Sarwar, G.; Steinke, F. H.; Slump, P.; Garcia, S.; Boisen, S. *In vitro* Assay for Protein Digestibility: Interlaboratory Study. *J. Assoc. Off. Anal. Chem.* **1990**, *73*, 622–625.
- Megat Rusydi, M.; Azrina, A. Effect of Germination on Total Phenolic , Tannin and Phytic Acid Contents in Soy Bean and Peanut. *Int. Food Res. J.* **2012**, *19*, 673–677.
- Megat Rusydi, M.; Noraliza, C.; Azrina, A.; Zulkhairi, A. Nutritional Changes in Germinated Legumes and Rice Varieties. *Int. Food Res. J.* **2011**, *696*, 688–696.
- Megat Rusydi, M.; Noraliza, C.; Azrina, A.; Zulkhairi, A. Nutritional Changes in Germinated Legumes and Rice Varieties. *Int. Food Res. J.* **2011**, *696*, 688–696.
- Mehansho, H. Iron Fortification Technology Development: New Approaches. *J. Nutr.* **2006**, 1059–1063.

- Metheson, N. K.; Strother, S. The Utilization of Phytate by Germinating Wheat. *Phytochemistry* **1969**, *8*, 1349–1356.
- Midorikawa, K.; Murata, M.; Oikawa, S.; Hiraku, Y.; Kawanishi, S. Protective Effect of Phytic Acid on Oxidative DNA Damage with Reference to Cancer Chemoprevention. *Biochem. Biophys. Res. Commun.* **2001**, *288*, 551–557.
- Miller, A.; Engel, K. H. Content of Γ -Oryzanol and Composition of Steryl Ferulates in Brown Rice (*Oryza Sativa* L.) of European Origin. *J. Agric. Food Chem.* **2006**, *54*, 8127–8133.
- Miller, A.; Engel, K. H. Content of Γ -Oryzanol and Composition of Steryl Ferulates in Brown Rice (*Oryza Sativa* L.) of European Origin. *J. Agric. Food Chem.* **2006**, *54*, 8127–8133.
- Min, B.; McClung, A.; Chen, M.-H. Effects of Hydrothermal Processes on Antioxidants in Brown, Purple and Red Bran Whole Grain Rice (*Oryza Sativa* L.). *Food Chem.* **2014**, *159*, 106–115.
- Mohamed Arshad, F.; Alias, E. F.; Noh, K. M.; Tasrif, M. Food Security: Self-sufficiency of Rice in Malaysia 1. *IJMS* **2011**, *18*, 83–100.
- Mohan, B. H.; Malleshi, N. G.; Koseki, T. *Physico-Chemical Characteristics and Non-Starch Polysaccharide Contents of Indica and Japonica Brown Rice and Their Malts*; 2010; Vol. 43, pp. 784–791.
- Mohanty, S.; Wassmann, R.; Nelson, A.; Moya, P.; Jagadish, S. V. K. *Rice and Climate Change: Significance for Food Security and Vulnerability*; 49; Los Banos (Phillippines), 2013; p. 14.
- Montgomery, D. C. *Design and Analysis of Experiments*; Wiley Press: New York, 1991.
- Moongnarm, A.; Khomphiphatkul, E. Germination Time Dependence of Bioactive Compounds and Antioxidant Activity in Germinated Rough Rice (*Oryza Sativa* L.). *Am. J. Appl. Sci.* **2011**, *8*, 15–25.
- Moongnarm, A.; Saetung, N. Comparison of Chemical Compositions and Bioactive Compounds of Germinated Rough Rice and Brown Rice. *Food Chem.* **2010**, *122*, 782–788.
- Morris, E. R.; Ellis, R. Bioavailability of Dietary Calcium. In *Nutritional bioavailability of calcium*; Kies, C., Ed.; American Chemical Society: Washington DC, 1985; pp. 63–72.
- Muñoz, J. A.; Valiente, M. Determination of Phytic Acid in Urine by Inductively Coupled Plasma Mass Spectrometry. *Anal. Chem.* **2003**, *75*, 6374–6378.

- Nakano, T.; Joh, T.; Tokumoto, E.; Hayakawa, T. Purification and Characterization of Phytase from Bran of *Triticum Aestivum* L. Cv. Nourin #61. *Food Sci. Technol. Res.* **1999**, *5*, 18–23.
- Nanua, J. N.; Mcgregor, J. U.; Godber, J. S. Influence of High-Oryzanol Rice Bran Oil on the Oxidative Stability of Whole Milk Powder. *J. Dairy Sci.* **2000**, *83*, 2426–2431.
- NCCFN. *Recommended Nutrient Intakes for Malaysia*; Putrajaya, 2005.
- Nergiz, C.; Gökgöz, E. Effects of Traditional Cooking Methods on Some Antinutrients and *in vitro* Protein Digestibility of Dry Bean Varieties (*Phaseolus Vulgaris* L.) Grown in Turkey. *Int. J. Food Sci. Technol.* **2007**, *42*, 868–873.
- Nicholas, D.; Hazila, K. K.; Chua, H. P.; Rosniyana, A. Nutritional Value and Glycemic Index of Bario Rice Varieties. *J. Trop. Agric. Food Sci.* **2014**, *42*, 1–8.
- Nicolosi, R. J.; Wilson, T. A.; Lawton, C.; Handelman, G. J.; Garry, J. Dietary Effects on Cardiovascular Disease Risk Factors: Beyond Saturated Fatty Acids and Cholesterol. *J. Am. Coll. Nutr.* **2001**, *20*, 421S – 427S.
- Nielsen, M. M.; Hansen, A. Stability of Vitamin E in Wheat Flour and Whole Wheat Flour during Storage. *Cereal Chem.* **2008**, *87*, 716–720.
- Ning, H.; Liu, Z.; Wang, Q.; Lin, Z.; Chen, S.; Li, G.; Wang, S.; Ding, Y. Effect of Nitrogen Fertilizer Application on Grain Phytic Acid and Protein Concentrations in Japonica Rice and Its Variations with Genotypes. *J. Cereal Sci.* **2009**, *50*, 49–55.
- Nonogaki, H.; Bassel, G. W.; Bewley, J. D. Germination—Still a Mystery. *Plant Sci.* **2010**, *179*, 574–581.
- Norazalina, S.; Norhaizan, M. E.; Hairuszah, I.; Norashareena, M. S. Anticarcinogenic Efficacy of Phytic Acid Extracted from Rice Bran on Azoxymethane-Induced Colon Carcinogenesis in Rats. *Exp. Toxicol. Pathol.* **2010**, *62*, 259–268.
- Norazalina, S.; Norhaizan, M. E.; Hairuszah, I.; Sabariah, A. R.; S. Nurul Husna; Norsharina, S. Antiproliferation and Apoptosis Induction of Phytic Acid in Hepatocellular Carcinoma (HEPG2) Cell Lines. *Afr. J. Biotechnol.* **2011**, *10*, 16646–16653.
- Norhaizan, M. E.; Ng, S.; Norashareena, M.; Abdah, M. A. Antioxidant and Cytotoxicity Effect of Rice Bran Phytic Acid as an Anticancer Agent on Ovarian, Breast and Liver Cancer. *Malays. J. Nutr.* **2011**, *17*, 367–375.
- Norhaizan, M. E.; Nor Faizadatul Ain, a W. Determination of Phytate, Iron, Zinc, Calcium Contents and Their Molar Ratios in Commonly Consumed Raw and Prepared Food in Malaysia. *Malays. J. Nutr.* **2009**, *15*, 213–222.

- Nurul-Husna, S.; Norhaizan, M. E.; Hairuszah, I.; Abdah, M. A.; Norazalina, S.; Norsharina, I. Rice Bran Phytic Acid (IP6) Induces Growth Inhibition, Cell Cycle Arrest and Apoptosis on Human Colorectal Adenocarcinoma Cells. *J. Med. Plants Res.* **2010**, *4*, 2283–2289.
- O'Dell, B. L.; De-Boland, A. R.; Koirtyohann, S. R. Distribution of Phytate and Nutritionally Important Elements among the Morphological Components of Cereal Grains. *J. Agric. Food Chem.* **1972**, *20*, 3–6.
- Oatway, L.; Vasanthan, T.; Helm, J. H. Phytic Acid. *Food Rev. Int.* **2001**, *17*, 419–431.
- Oberleas, D. The Role of Phytate in Zinc Bioavailability and Homeostasis. In *Nutritional bioavailability of zinc*; Inglett, G. E., Ed.; American Chemical Society: Washington DC, 1983; pp. 145–158.
- Oberleas, D.; Harland, B. F. Newer Methods for Phytate Analysis. In *Handbook of Dietary Fiber in Human Nutrition*; American Chemical Society: Washington DC, 2001; pp. 113–126.
- Ockenden, I.; Dorsch, J. A.; Reid, M. M.; Lin, L.; Grant, L. K.; Raboy, V.; Lott, J. N. A. Characterization of the Storage of Phosphorus, Inositol Phosphate and Cations in Grain Tissues of Four Barley (*Hordeum Vulgare* L.) Low Phytic Acid Genotypes. *Plant Sci.* **2004**, *167*, 1131–1142.
- Ogawa, M.; Tanaka, K.; Kasai, Z. Note on the Phytin-Containing Particles Isolated from Rice Scutellum. *Cereal Chem.* **1977**, *54*, 1029–1034.
- Oh, C. S.; Choi, Y. H.; Lee, S. J.; Yoon, D. B.; Moon, H. P.; Ahn, S. N. Mapping of Quantitative Trait Loci for Cold Tolerance in Weedy Rice. *Breed. Sci.* **2004**, *54*, 373–380.
- Oh, C.-H.; Oh, S.-H. Effects of Germinated Brown Rice Extracts with Enhanced Levels of GABA on Cancer Cell Proliferation and Apoptosis. *J. Med. Food* **2004**, *7*, 19–23.
- Ohtsubo, K.; Suzuki, K.; Yasui, Y.; Kasumi, T. Bio-Functional Components in the Processed Pre-Germinated Brown Rice by a Twin-Screw Extruder. *J. Food Compos. Anal.* **2005**, *18*, 303–316.
- Ohtsubo, K.; Suzuki, K.; Yasui, Y.; Kasumi, T. Bio-Functional Components in the Processed Pre-Germinated Brown Rice by a Twin-Screw Extruder. *J. Food Compos. Anal.* **2005**, *18*, 303–316.
- Okada, T.; Sugishita, T.; Murakami, T.; Murai, H.; Saikusa, T.; Horio, T. Effect of the Defatted Rice Germ Enriched with GABA for Sleeplessness, Depression, Autonomic Disorder by Oral Administration. *Nippon Shokuhin Kagaku Kougaku Kaishi* **2000**, *47*, 596–603.

Onomi, S.; Okazaki, Y.; Katayama, T. Effect of Dietary Level of Phytic Acid on Hepatic and Serum Lipid Status in Rats Fed a High-Sucrose Diet. *Biosci. Biotechnol. Biochem.* **2004**, *68*, 1379–1381.

Oryza.com. Groups: It's time Malaysia became self-sufficient.

Ou, K.; Cheng, Y.; Xing, Y.; Lin, L.; Nout, R.; Liang, J. Phytase Activity in Brown Rice during Steeping and Sprouting. *J. Food Sci. Technol.* **2010**, *48*, 598–603.

Ou, S.; Kwok, K.-C. Ferulic Acid: Pharmaceutical Functions, Preparation and Applications in Foods. *J. Sci. Food Agric.* **2004**, *84*, 1261–1269.

Paddy Statistics of Malaysia; Department of Agriculture Peninsular Malaysia, 2013; pp. 1–105.

Panchan, K.; Naivikul, O. Effect of Pre-Germination and Parboiling on Brown Rice Properties. *Asian J. Food Agro-Industry* **2009**, *2*, 515–524.

Panlasigui, L. N.; Thompson, L. U. Blood Glucose Lowering Effects of Brown Rice in Normal and Diabetic Subjects. *Int. J. Food Sci. Nutr.* **2006**, *57*, 151–158.

Panlasigui, L. N.; Thompson, L. U.; Juliano, B. O.; Perez, C. M.; Yiu, S. H.; Greenberg, G. R. Rice Varieties with Similar Amylose Content Differ in Starch Digestibility and Glycemic Response in Humans. *Am. J. Clin. Nutr.* **1991**, *54*, 871–877.

Pascoe, D. A.; Fulcher, R. G. Biochemistry and Compartmentalization of Cereal Grain Components and Their Functional Relationship to Mammalian Health. In *Whole Grains and Health*; Blackwell Publishing Professional, 2007; pp. 89–114.

Patil, S. B.; Khan, M. K. Germinated Brown Rice as a Value Added Rice Product: A Review. *J. Food Sci. Technol.* **2011**, *48*, 661–667.

Persson, H.; Turk, M.; Nyman, M.; Sandberg, A. Binding of Cu²⁺, Zn²⁺, and Cd²⁺ to Inositol Tri-, Tetra-, Penta-, and Hexaphosphates. *Agric. Food Chem.* **1998**, *46*, 3194–3200.

Phillippy, B. Q.; Bland, J. M. Gradient Ion Chromatography of Inositol Phosphates. *Anal. Biochem.* **1988**, *175*, 162–166.

Phillippy, B. Q.; Graf, E. Antioxidant Functions of Inositol 1,2,3- Trisphosphate and Inositol 1,2,3,6-Tetrakisphosphate. *Free Radic. Biol. Med.* **1997**, *22*, 939–946.

Phillippy, B. Q.; Johnston, M. . Determination of Phytic Acid in Foods by Ion Chromatography with Postcolumn Derivatization. *J. Food Sci.* **1985**, *50*, 541–542.

Pinkaew, S.; Winichagoon, P.; Hurrell, R. F.; Wegmuller, R. Extruded Rice Grains Fortified with Zinc, Iron, and Vitamin A Increase Zinc Status of Thai School

- Children When Incorporated into a School Lunch Program. *J. Nutr.* **2013**, *143*, 362–368.
- Pisani, F.; Livermore, T.; Rose, G.; Chubb, J.; Gaspari, M. Analysis of Dictyostelium Discoideum Inositol Pyrophosphate Metabolism by Gel Electrophoresis. *PLoS One* **2014**, *9*, e85533.
- Potrykus, I. Nutritionally Enhanced Rice to Combat Malnutrition Disorders of the Poor. *Nutr. Rev.* **2003**, *61*, S101–S104.
- Qi, X.; Wilson, K.; Tan-Wilson, A. Characterization of the Major Protease Involved in the Soybean Beta-Conglycinin Storage Protein Mobilization. *Plant Physiol.* **1992**, *99*, 725–733.
- Raboy, V. Approaches and Challenges to Engineering Seed Phytate and Total Phosphorus. *Plant Sci.* **2009**, *177*, 281–296.
- Raboy, V. Seeds for a Better Future: “ Low Phytate ” Grains Help to Overcome Malnutrition and Reduce Pollution. *Trends Plant Sci.* **2001**, *6*, 458–462.
- Raboy, V.; Gerbasi, P. F.; Young, K. a; Stoneberg, S. D.; Pickett, S. G.; Bauman, a T.; Murthy, P. P.; Sheridan, W. F.; Ertl, D. S. Origin and Seed Phenotype of Maize Low Phytic Acid 1-1 and Low Phytic Acid 2-1. *Plant Physiol.* **2000**, *124*, 355–368.
- Raes, K.; Knockaert, D.; Strujis, K.; Camp, J. Van. Roles of Processing on Bioaccessibility of Minerals: Influence of Localization of Minerals and Antinutritional Factors in Plant. *Trends Food Sci. Technol.* **2014**, *37*, 32–41.
- Ramesh, S. A.; Choimes, S.; Schachtman, D. P. Over-Expression of an Arabidopsis Zinc Transporter in Hordeum Vulgare Increases Short-Term Zinc Uptake after Zinc Deprivation and Seed Zinc Content. *Plant Mol. Biol.* **2004**, *54*, 373–385.
- Ramli, S.; Ismail, N.; Mubarek Alkarkhi, A. F.; Easa, A. M. The Use of Principal Component and Cluster Analysis to Differentiate Banana Peel Flours Based on Their Starch and Dietary Fibre Components. *Trop. Life Sci. Res.* **2010**, *21*, 91–100.
- Rattanachitthawat, S. Phenolic Content and Antioxidant Activities in Red Unpolished Thai Rice Prevents Oxidative Stress in Rats. *J. Med. Plants Res.* **2010**, *4*, 796–801.
- Reddy, N. R. Dietary Intake of Phytate. In *Food Phytates*; 2002.
- Reddy, N. R.; Pierson, M. Reduction in Antinutritional and Toxic Components in Plant Foods by Fermentation. *Food Res. Int.* **1994**, *27*, 281.
- Reddy, N. R.; Pierson, M.; Sathe, S.; Salunkhe, D. *Phytate Digestion and Bioavailability. In Phytates in Cereals and Legumes.*; CRC Press: Florida, 1989a; pp. 71–80.

- Reddy, N. R.; Pierson, M.; Sathe, S.; Salunkhe, D. *Phytate in Cereals and Legumes*; CRC Press: Boca Raton, FL, 1989b.
- Reddy, N. R.; Sathe, S. *Food Phytates*; Rukma Reddy, N.; Sathe, S., Eds.; CRC Press, 2002.
- Rickard, S. E.; Thompson, L. U. Interactions and Biological Effects of Phytic Acid. In *Antinutrients and phytochemicals in food*; American Chemical Society: Washington DC, 1997; pp. 292–312.
- Rimsten, L.; Stenberg, T.; Andersson, R.; Andersson, A.; Aman, P. Determination of Beta-Glucan Molecular Weight Using SEC with Calcofluor Detection in Cereal Extracts. *Cereal Chem.* **2003**, *80*, 485–490.
- Rohlf, F. J. Numerical Taxonomy and Multivariate's Analysis System NTSys-PC, 2005.
- Roohinejab, S. Effects of Germination Time and Varieties of Unmilled Rice on Blood Cholesterol in Sprague-Dawley Male Rats, Univerisiti Putra Malaysia, 2009.
- Roohinejab, S. Effects of Germination Time and Varieties of Unmilled Rice on Blood Cholesterol in Sprague-Dawley Male Rats, Univerisiti Putra Malaysia, 2009.
- Roohinejad, S.; Omidzadeh, A.; Mirhosseini, H.; Saari, N.; Mustafa, S.; Yusof, R. M.; Hussin, A. S. M.; Hamid, A.; Abd Manap, M. Y. Effect of Pre-Germination Time of Brown Rice on Serum Cholesterol Levels of Hypercholesterolaemic Rats. *J. Sci. Food Agric.* **2010**, *90*, 245–251.
- Roohinejad, S.; Omidzadeh, A.; Mirhosseini, H.; Saari, N.; Mustafa, S.; Yusof, R. M.; Hussin, A. S. M.; Hamid, A.; Abd Manap, M. Y. Effect of Pre-Germination Time of Brown Rice on Serum Cholesterol Levels of Hypercholesterolaemic Rats. *J. Sci. Food Agric.* **2010**, *90*, 245–251.
- Roy, P.; Orikasa, T.; Okadome, H.; Nakamura, N.; Shiina, T. Processing Conditions, Rice Properties, Health and Environment. *Int. J. Environ. Res. Public Health* **2011**, *8*, 1957–1976.
- Sabirov, K. A.; Demkina, N. V; Igamnazarov, R. P.; Yuldashev, B. T.; Sultanov, K.; Rakhimov, M. M. Action of Rice Bran Phytase on Hydrolysis of Phytin. In: 1990; pp. 599–602.
- Sagum, R.; Arcot, J. Effect of Domestic Processing Methods on the Starch, Non-Starch Polysaccharides and *in vitro* Starch and Protein Digestibility of Three Varieties of Rice with Varying Levels of Amylose. *Food Chem.* **2000**, *70*, 107–111.
- Sakac, M.; Canadanovic-Brunet, J.; Misan, A.; Tumbas, V.; Medic, D. Antioxidant Activity of Phytic Acid in Lipid Model System. *Food Technol. Biotechnol.* **2010**, *48*, 524–529.

- Sakamoto, K.; Venkatraman, G.; Shamsuddin, A. M. Growth Inhibition and Differentiation of HT-29 Cells *in vitro* by Inositol Hexaphosphate (phytic Acid). *Carcinogenesis* **1993**, *14*, 1815–1819.
- Salunkhe, D. K.; Chavan, J. K.; Kadam, S. S. *Dietary Tannins: Consequences and Remedies.*; CRC Press: Boca Raton, FL, 1990.
- Saman, P.; Vázquez, J. A.; Pandiella, S. S. Controlled Germination to Enhance the Functional Properties of Rice. *Process Biochem.* **2008**, *43*, 1377–1382.
- Sandberg, A. *In vitro* and *in Vivo* Degradation of Phytate. In *Food Phytates*; 2002.
- Sandberg, A.; Andersson, H.; Carlsson, N. G. G.; Sandstrom M, B.; Sandström, B. Degradation Products of Bran Phytate Formed during Digestion in the Human Small Intestine. Effect of Extrusion Cooking on Digestibility. *J. Nutr.* **1987**, *117*, 2061–2065.
- Sandberg, A.; Larsen, T.; Sandstrom, B. High Dietary Calcium Level Decreases Colonic Phytate Degradation in Pigs Fed a Rapeseed Diet. *J. Nutr.* **1993**, *123*, 559–566.
- Sandberg, A.-S.; Andlid, T. Phytogenic and Microbial Phytases in Human Nutrition. *Int. J. Food Sci. Technol.* **2002**, *37*, 823–833.
- Sandstrom, B.; Bugel, S.; McGaw, B. A.; Price, J.; Reid, M. D. Nutrient Metabolism A High Oat-Bran Intake Does Not Impair Zinc Absorption in Humans When Added to a Low-Fiber Animal Protein-Based Diet. *J. Nutr.* **2000**, *130*, 594–599.
- Sandstrom, B.; Bugel, S.; McGaw, B. A.; Price, J.; Reid, M. D. Nutrient Metabolism A High Oat-Bran Intake Does Not Impair Zinc Absorption in Humans When Added to a Low-Fiber Animal Protein-Based Diet. *J. Nutr.* **2000**, *130*, 594–599.
- Sandström, B.; Cederblad, A.; Stenquist, B.; Andersson, H. Effect of Inositol Hexaphosphate on Retention of Zinc and Calcium from the Human Colon. *Eur. J. Clin. Nutr.* **1990**, *44*, 705–708.
- Sarawak Agriculture Statistics*; Department of Agriculture Sarawak, Malaysia., 2013; pp. 1–9.
- Sareepuang, K.; Siriamornpun, S.; Wiset, L.; Meeso, N. Effect of Soaking Temperature on Physical , Chemical and Cooking Properties of Parboiled Fragrant Rice. *World J. Agric. Sci.* **2008**, *4*, 409–415.
- Schlemmer, U.; Frølich, W.; Prieto, R. M.; Grases, F. Phytate in Foods and Significance for Humans : Food Sources , Intake , Processing , Bioavailability , Protective Role and Analysis. *Mol. Nutr. Food Res.* **2009**, *53*, 330–375.
- Schlemmer, U.; Miller, H.; Jany, K. D. The Degradation of Phytic Acid in Legumes Prepared by Different Methods. *Eur. J. Clin. Nutr.* **1995**, *49*, 207–210.

- Selle, P. H.; Gill, R. J.; Scott, T. A. Effects of Pre-Pelleted Wheat and Phytase Supplementation on Broiler Growth Performance and Nutrient Utilisation. In *Proceedings of Australian Poultry Science Symposium*; 2007; pp. 182–185.
- Selle, P. H.; Ravindran, V. Microbial Phytase in Poultry Nutrition. *Anim. Feed Sci. Technol.* **2007**, *135*, 1–41.
- Selle, P. H.; Ravindran, V.; Caldwell, R. A.; Bryden, W. L. Phytate and Phytase: Consequences for Protein Utilisation. *Nutr. Res. Rev.* **2000**, *13*, 255–278.
- Shafie, B.; Cheng, S. C.; Yiu, P. H. Pasting Properties of Sarawak Rice Cultivars. In *Proceedings of Sarawak Rice Conference 2011*; 2011; p. 25.
- Shafie, N. H.; Esa, N. M.; Ithnin, H.; Akim, A.; Saad, N.; Pandurangan, A. K. Preventive Inositol Hexaphosphate Extracted from Rice Bran Inhibits Colorectal Cancer through Involvement of Wnt / -Catenin and COX-2 Pathways. **2013a**, 2013.
- Shafie, N. H.; Esa, N. M.; Ithnin, H.; Saad, N.; Pandurangan, A. K. Pro-Apoptotic Effect of Rice Bran Inositol Hexaphosphate (IP6) on HT-29 Colorectal Cancer Cells. *Int. J. Mol. Sci.* **2013b**, *14*, 23545–23558.
- Shamsuddin, A. M.; Yang, G. Y.; Vucenic, I. Novel Anti-Cancer Functions of IP6: Growth Inhibition and Differentiation of Human Mammary Cancer Cell Lines *in vitro*. *Anticancer Res.* **1996**, *16*, 3287–3292.
- Shao, Y.; Ga, Z.; Bao, J. Total Phenolic Content and Antioxidant Capacity of Rice Grains with Extremely Small Size. *African J. Agric. Res.* **2011**, *6*, 2289–2293.
- Sharma, P.; Gujral, H. S.; Singh, B. Antioxidant Activity of Barley as Affected by Extrusion Cooking. *Food Chem.* **2012**, *131*, 1406–1413.
- Shears, S. B.; Turner, B. L. Nomenclature and Terminology of Inositol Phosphates: Clarification and a Glossary of Terms. In *Inositol Phosphates: Linking Agriculture and the Environment*; Turner, B. L.; Richardson, A. E.; Mullaney, E. J., Eds.; CABI: Wallingford, UK, 2007; pp. 1–7.
- Shen, Y.; Jin, L.; Xiao, P.; Lu, Y.; Bao, J. Total Phenolics, Flavonoids, Antioxidant Capacity in Rice Grain and Their Relations to Grain Color, Size and Weight. *J. Cereal Sci.* **2009**, *49*, 106–111.
- Shibuya, N. Phenolic Acids and Their Carbohydrate Esters in Rice Endosperm Cell Walls. *Phytochemistry* **1984**, *23*, 2233–2237.
- Shu, X.-L.; Frank, T.; Shu, Q.-Y.; Engel, K.-H. Metabolite Profiling of Germinating Rice Seeds. *J. Agric. Food Chem.* **2008**, *56*, 11612–11620.
- Siddhuraju, P.; Becker, K. Nutritional and Antinutritional Composition, *in vitro* Amino Acid Availability, Starch Digestibility and Predicted Glycemic Index of

- Differentially Processed Mucuna Beans (*Mucuna Pruriens* Var. *Utilis*): An under-Utilised Legume. *Food Chem.* **2005**, *91*, 275–286.
- Sing, S. X.; Wong, T. S.; Wong, L. L.; Lee, H. H.; Yiu, P. H. Health Promoting Phytochemicals In Sarawak Pigmented Rice. In *Proceedings of Sarawak Rice Conference 2011*; 2011; p. 27.
- Singh, J.; Dartois, A.; Kaur, L. Starch Digestibility in Food Matrix: A Review. *Trends Food Sci. Technol.* **2010**, *21*, 168–180.
- Singh, R. P.; Agarwal, C.; Agarwal, R. Inositol Hexaphosphate Inhibits Growth, and Induces G1 Arrest and Apoptotic Death of Prostate Carcinoma DU145: Modulation of CDKI-CDK-Cyclin and pRb-Related Protein-E2F Complexes. *Carcinogenesis* **2003**, *24*, 555–563.
- Skoglund, E.; Carlsson, N.; Sandberg, A. High-Performance Chromatographic Separation of Inositol Phosphate Isomers on Strong Anion Exchange Columns. *J. Agric. Food Chem.* **1998**, *46*, 1877–1882.
- Skoglund, E.; Larsen, T.; Sandberg, A.-S. Comparison between Steeping and Pelleting a Mixed Diet at Different Calcium Levels on Phytate Degradation in Pigs. *Can. J. Anim. Sci.* **1997**, *77*, 471–477.
- Skoglund, E.; Lönnerdal, B.; Sandberg, A. Inositol Phosphates Influence Iron Uptake in Caco-2 Cells. *J. Agric. Food Chem.* **1999**, *47*, 1109–1113.
- Slavin, J. Fiber and Prebiotics: Mechanisms and Health Benefits. *Nutrients* **2013**, *5*, 1417–1435.
- Slavin, J. Whole Grains and Human Health. *Nutr. Res. Rev.* **2004**, *17*, 99–110.
- Slavin, J. Why Whole Grains Are Protective: Biological Mechanisms. *Proc. Nutr. Soc.* **2003**, *62*, 129–134.
- Slavin, J.; Savarino, V.; Paredes-Diaz, a.; Fotopoulos, G. A Review of the Role of Soluble Fiber in Health with Specific Reference to Wheat Dextrin. *J. Int. Med. Res.* **2009**, *37*, 1–17.
- Sögnen, E. Effects of Calcium-Binding Substances on Gastric Emptying as Well as Intestinal Transit and Absorption in Intact Rats. *Acta Pharmacol. Toxicol. (Copenh)*. **1965**, *22*, 31–48.
- Soi-ampornkul, R. Protective Effects of Pre-Germinated Brown Rice Extract against Amyloid B-Peptide Induced Neurotoxicity in Neuronal SK-N-SH Cells. *Int. J. Nutr. Food Sci.* **2013**, *2*, 167.
- Spallholz, J. E.; Boylan, L. M.; Driskel, J. A. *Nutrition: Chemistry and Biology*; CRC Press: Canada, 1999.

- Srikaeo, K.; Sopade, P. a. Functional Properties and Starch Digestibility of Instant Jasmine Rice Porridges. *Carbohydr. Polym.* **2010**, *82*, 952–957.
- Sripriya, G.; Antony, U.; Chandra, T. S. Changes in Carbohydrate, Free Amino Acids, Organic Acids, Phytate and HCl Extractability of Minerals during Germination and Fermentation of Finger Millet (Eleusine Coracana). *Food Chem.* **1997**, *58*, 345–350.
- Srivastava, L. M. Seed Germination, Mobilization of Food Reserves, and Seed Dormancy. In; Srivastava, L. M., Ed.; Academic Press: San Diego, 2002; pp. 447–471.
- Subudhi, P. K.; Sasaki, T.; Khush, G. S. Rice. In *Genome Mapping and Molecular Breeding in Plants, Cereals and Millets.*; Kole, C., Ed.; Springer: Verlag Berlin Heidelberg, 2006; pp. 264–270.
- Suliburska, J.; Krejpcio, Z. Evaluation of the Content and Bioaccessibility of Iron, Zinc, Calcium and Magnesium from Groats, Rice, Leguminous Grains and Nuts. *J. Food Sci. Technol.* **2011**.
- Syahariza, Z. a; Sar, S.; Hasjim, J.; Tizzotti, M. J.; Gilbert, R. G. The Importance of Amylose and Amylopectin Fine Structures for Starch Digestibility in Cooked Rice Grains. *Food Chem.* **2013**, *136*, 742–749.
- Tajima, M.; Horino, T.; Maeda, M.; Rok Son, J. Maltooligosaccharides Extracted from Outer Layer of Rice Grain. *Nippon Shokuhin Kogyo Gakkaishi* **1992**, *39*, 857–861.
- Tanaka, K.; Kasai, Z. Phytic Acid in Rice Grains. In *Antinutrients and natural toxicants in foods*; Ory, R. L., Ed.; Food Nutrition Press, Inc.: Westport, CT, 1981; pp. 239–260.
- Tang, G.; Qin, J.; Dolnikowski, G. G.; Russell, R. M.; Grusak, M. A. Golden Rice Is an Effective Source of Vitamin A. *Ame* **2009**, *89*, 1776–1783.
- Tan-Wilson, A. L.; Wilson, K. A. Mobilization of Seed Protein Reserves. *Physiol. Plant.* **2011**, *145*, 140–153.
- Teo, G. K. *Rice Biodiversity*; Semongok, Sarawak, Malaysia, 2010.
- Thompson, L. U. Antinutrients and Blood Glucose. *Food Technol.* **1988**, *42*, 123–132.
- Thompson, L. U. Potential Health Benefits and Problems Associated with Antinutrients in Foods. *Food Res. Int.* **1993**, *26*, 131–149.
- Thompson, L. U.; Button, C. L.; Jenkins, D. J. A. Phytic Acid and Calcium Affect the *in vitro* Rate of Navy Bean Starch Digestion and Blood Glucose Response in Humans. *Am. J. Clin. Nutr.* **1987**, *46*, 467–473.

- Tian, S.; Nakamura, K.; Kayahara, H. Analysis of Phenolic Compounds in White Rice, Brown Rice, and Germinated Brown Rice. *J. Agric. Food Chem.* **2004**, *52*, 4808–4813.
- Tian, S.; Nakamura, K.; Kayahara, H. Analysis of Phenolic Compounds in White Rice, Brown Rice, and Germinated Brown Rice. *J. Agric. Food Chem.* **2004**, *52*, 4808–4813.
- Tizazu, S.; Urga, K.; Belay, A.; Abuye, C.; Retta, N. Effect of Germination on Mineral Bioavailability of Sorghum-Based Complementary Foods. *African J. Food, Agric. , Nutr. Dev.* **2011**, *11*, 5083–5095.
- Towo, E.; Mgoba, C.; Ndossi, G. D.; Kimboka, S. Effect of Phytate and Iron-Binding Phenolics on the Content and Availability of Iron and Zinc in Micronutrients Fortified Cereal Flours. *African J. Food, Agric. , Nutr. Dev.* **2006**, *6*, 1–14.
- Trugo, L. C.; Donangelo, C. M.; Trugo, N. M.; Bach Knudsen, K. E. Effect of Heat Treatment on Nutritional Quality of Germinated Legume Seeds. *J. Agric. Food Chem.* **2000**, *48*, 2082–2086.
- Tulyathan, V.; Leeharatanaluk, B. Changes in Quality of Rice (*Oryza Sativa* L.) CV. Khao Dawk Mali 105 during Storage. *J. Food Biochem.* **2006**, *31*, 415–425.
- Tungtrakul, P.; Varayanond, W.; Surojanametakul, V.; Wattanasiritham, L. Accumulation of Gamma-Aminobutyric Acid (GABA) in Non-Waxy and Waxy Rice Germ during Water Soaking. *Thai J. Agric. Sci.* **2005**, 1–4.
- Tungtrakul, P.; Varayanond, W.; Surojanametakul, V.; Wattanasiritham, L. Accumulation of Gamma-Aminobutyric Acid (GABA) in Non-Waxy and Waxy Rice Germ during Water Soaking. *Thai J. Agric. Sci.* **2005**, 1–4.
- Turnlund, J. R.; King, J. C.; Keyes, W. R.; Gong, B.; Michel, M. C. A Stable Isotope Study of Zinc Absorption in Young Men: Effects of Phytate and A-Cellulose. *Am. J. Clin. Nutr.* **1984**, *40*, 1071–1077.
- USDA. *USDA National Nutrient Database for Standard Reference*; Beltsville, MD, USA, 2007.
- Usuki, S.; Ito, Y.; Morikawa, K.; Kise, M.; Ariga, T.; Rivner, M.; Yu, R. K. Effect of Pre-Germinated Brown Rice Intake on Diabetic Neuropathy in Streptozotocin-Induced Diabetic Rats. *Nutr. Metab. (Lond).* **2007**, *4*, 25.
- Varayanond, W.; Tungtrakul, P.; Surojanametakul, V.; Wattanasiritham, L.; Luxiang, W. Effects of Water Soaking on Gamma-Aminobutyric Acid (GABA) in Germ of Different Thai Rice Varieties. *Kasetsart J. (Natural Sci.* **2005**, *39*, 411–415.
- Vatanasuchart, N.; Niyomwit, B.; Wongkrajang, K. Resistant Starch Contents and the *in vitro* Starch Digestibility of Thai Starchy Foods. *Kasetsart J. (Natural Sci.* **2009**, *43*, 178–186.

- Vatanasuchart, N.; Niyomwit, B.; Wongkrajang, K. Resistant Starch Content , *in vitro* Starch Digestibility and Physico-Chemical Properties of Flour and Starch from Thai Bananas. *Maejo Int. J. Sci. Technol.* **2012**, *6*, 259–271.
- Vohra, A.; Satyanarayana, T. Statistical Optimization of the Medium Components by Response Surface Methodology to Enhance Phytase Production by *Pichia Anomala*. *Process Biochem.* **2002**, *37*, 999–1004.
- Vucenik, I.; Podczasy, J. J.; Shamsuddin, A. M. Antiplatelet Activity of Inositol Hexaphosphate (IP6). *Anticancer Res.* **1999**, *19*, 3689–3693.
- Vucenik, I.; Shamsuddin, A. M. Protection Against Cancer by Dietary IP 6 and Inositol. *Nutr. Cancer* **2009**, *55*, 109–125.
- Wada, T.; Lott, J. N. A. Light and Electron Microscopic and Energy Dispersive X-Ray Microanalysis Studies of Globoids in Protein Bodies of Embryo Tissues and the Aleurone Layer of Rice (*Oryza Sativa* L.) Grains. *Can. J. Bot.* **1997**, *75*, 1137–1147.
- Walker, A. R. P.; Fox, F. W.; Irving, K. T. Studies in Human Mineral Metabolism. The Effect of Bread Rich in Phytate-Phosphorus on the Metabolism of Certain Mineral Salts with Special Reference to Calcium. *Biochem. J.* **1948**, *42*, 452–462.
- Wang, K. M.; Wu, J. G.; Li, G.; Zhang, D. P.; Yang, Z. W.; Shi, C. H. Distribution of Phytic Acid and Mineral Elements in Three Indica Rice (*Oryza Sativa* L.) Cultivars. *J. Cereal Sci.* **2011**, *54*, 116–121.
- Wei, Y.; Shohag, M. J. I.; Wang, Y.; Lu, L.; Wu, C.; Yang, X. Effect of Zinc Sulfate Fortification in Germinated Brown Rice on Seed Zinc Concentration, Bioavailability, and Seed Germination. *J. Agric. Food Chem.* **2012**, *60*, 1871–1879.
- Wen, H.; Cao, X.; Gu, Z.; Tang, J.; Han, Y. Effects of Components in the Culture Solution on Peptides Accumulation during Germination of Brown Rice. *Eur. Food Res. Technol.* **2009**, *228*, 959–967.
- Weremko, D.; Fandrejowski, H.; Zebrowska, T.; Han, K.; Kim, J. H.; Cho, W. T. Bioavailability of Phosphorus in Feeds of Plant Origin for Pigs – Review. *Asian-Australians J. Anim. Sci.* **1997**, *10*, 551–566.
- Wichamanee, Y.; Teerarat, I. Production of Germinated Red Jasmine Brown Rice and Its Physicochemical Properties. *Int. Food Res. J.* **2012**, *19*, 1649–1654.
- Wilhelmson, A.; Oksman-Caldentey, K.; Suortti, T.; Kaukovirta-Norja, A.; Poutanen, K. Development of a Germination Process for Producing High B-Glucan Whole Grain Food Ingredients from Oat. *Cereal Chem.* **2001**, *78*, 715–720.
- Wong, J. M. W.; Jenkins, D. J. A. Carbohydrate Digestibility and Metabolic Effects. *J. Nutr.* **2007**, *137*, 2539S – 2546S.

- Wu, F.; Yang, N.; Touré, A.; Jin, Z.; Xu, X. Germinated Brown Rice and Its Role in Human Health. *Crit. Rev. Food Sci. Nutr.* **2013**, *53*, 451–463.
- Wu, P.; Tian, J.-C.; Walker, C. E. (Chuck); Wang, F.-C. Determination of Phytic Acid in Cereals - a Brief Review. *Int. J. Food Sci. Technol.* **2009**, *44*, 1671–1676.
- Wu, W.; Cheng, F.; Liu, Z.; Wei, K. Difference of Phytic Acid Content and Its Relation to Four Protein Composition Contents in Grains of Twenty-Nine Japonica Rice Varieties from Jiangsu and Zhejiang Provinces, China. *Rice Sci.* **2007**, *14*, 311–314.
- Xu, J.; Zhang, H.; Guo, X.; Qian, H. The Impact of Germination on the Characteristics of Brown Rice Flour and Starch. *J. Sci. Food Agric.* **2012**, *92*, 380–387.
- Xu, J.-G.; Hu, Q.-P.; Duan, J.-L.; Tian, C.-R. Dynamic Changes in Γ -Aminobutyric Acid and GAD Activity during Steeping and Germination. *J. Agric. Food Chem.* **2010**, *58*, 9759–9763.
- Xu, P.; Price, J.; Wise, A.; Aggett, P. J. Interaction of Inositol Phosphates with Calcium, Zinc and Histidine. *J. Inorg. Biochem.* **1992**, *47*, 119–130.
- Yamaguchi, R.; Tatsumi, M.; Kato, K.; Yoshimitsu, U. Effect of Metal Salts and Fructose on the Autoxidation of Methyl Linoleate in Emulsions. *Agric. Biol. Chem.* **1988**, *52*, 849–850.
- Yang, F.; Basu, T. K.; Ooraikul, B. Studies on Germination Conditions and Antioxidant Contents of Wheat Grain. *Int. J. Food Sci. Nutr.* **2001**, *52*, 319–330.
- Yanke, L. J.; Bae, H. D.; Selinger, L. B.; Cheng, K. J. Phytase Activity of Anaerobic Ruminant Bacteria. *Microbiol.* **1998**, *144*, 1565–1573.
- Ye, X.; Al-Babili, S.; Klotti, A.; Zhang, J.; Lucca, P.; Beyer, P.; Potrykus, I. Engineering the Provitamin A (beta-Carotene) Biosynthetic Pathway into (Carotenoid-Free) Rice Endosperm. *Science (80-.)*. **2000**, *287*, 303–305.
- Yoon, J. H.; Thompson, L. U.; Jenkins, D. J. The Effect of Phytic Acid on *in vitro* Rate of Starch Digestibility and Blood Glucose Response. *Am. J. Clin. Nutr.* **1983**, *38*, 835–842.
- Yoshida, T.; Tanaka, K.; Kasai, Z. Phytase Activity Associated with Isolated Aleurone Particles of Rice Grains. *Agric. Biol. Chem.* **1975**, *39*, 289–290.
- Yousif, N. E.; Tinay, A. H. E. L. Effect of Natural Fermentation on Protein Fractions and *in vitro* Protein Digestibility of Rice. *Plant Foods Hum. Nutr.* **2004**, *58*, 1–8.
- Yuan, J.-P.; Liu, Y.-B.; Peng, J.; Wang, J.-H.; Liu, X. Changes of Isoflavone Profile in the Hypocotyls and Cotyledons of Soybeans during Dry Heating and Germination. *J. Agric. Food Chem.* **2009**, *57*, 9002–9010.

- Zakaria, S.; Matsuda, T.; Nitta, Y. Morphological Studies on the Mobilization of Reserves in Germinating Rice Seed. *Plant Prod. Sci.* **2000**, *3*, 152–160.
- Zeng, Y.; Zhang, H.; Wang, L.; Pu, X.; Du, J.; Yang, S.; Liu, J. Genotypic Variation in Element Concentrations in Brown Rice from Yunnan Landraces in China. *Environ. Geochem. Health* **2010**, *32*, 165–177.
- Zhang, C.; Shen, Y.; Chen, J.; Xiao, P.; Bao, J. Nondestructive Prediction of Total Phenolics, Flavonoid Contents, and Antioxidant Capacity of Rice Grain Using near-Infrared Spectroscopy. *J. Agric. Food Chem.* **2008**, *56*, 8268–8272.
- Zhang, M. W.; Zhang, R. F.; Zhang, F. X.; Liu, R. H. Phenolic Profiles and Antioxidant Activity of Black Rice Bran of Different Commercially Available Varieties. *J. Agric. Food Chem.* **2010a**, *58*, 7580–7587.
- Zhang, W.; Liu, S.; Wang, Y.; Zheng, L.; Liu, F.; Han, X.; Ren, Y.; Long, Q.; Zhao, Z.; Jiang, L.; *et al.* A Study of the *in vitro* Protein Digestibility of Indica and Japonica Cultivars. *Food Chem.* **2010b**, *122*, 1199–1204.
- Zhao, H.; Fan, W.; Dong, J.; Lu, J.; Chen, J.; Shan, L.; Lin, Y.; Kong, W. Evaluation of Antioxidant Activities and Total Phenolic Contents of Typical Malting Barley Varieties. *Food Chem.* **2008**, *107*, 296–304.
- Zhu, K.-X.; Lian, C.-X.; Guo, X.-N.; Peng, W.; Zhou, H.-M. Antioxidant Activities and Total Phenolic Contents of Various Extracts from Defatted Wheat Germ. *Food Chem.* **2011**, *126*, 1122–1126.

BIODATA OF STUDENT

Lee Hwei Hong was born in 24 August 1985 and raised up in Kuala Lumpur, Malaysia. She attended her primary education in S.R.J.K. (C) Lai Chee, Kuala Lumpur (1991), secondary education in S.M.K.P. Jalan Ipoh, Kuala Lumpur (1997), followed by pre-university education in S.M.K. Maxwell, Kuala Lumpur (2003). She pursued her Bachelor study in Bioindustry Sciences in Universiti Putra Malaysia Bintulu Sarawak Campus (UPMKB), and graduated with first class honors in 2009.

Her interest in scientific research leads her to continue her postgraduate study in the Faculty of Agriculture and Food Sciences, UPMKB. She was offered the PhD studentship by the School of Graduate Studies, UPM and received a scholarship award (National Science Fellowship Batch1/2009) from the Ministry of Science, Technology and Innovation (MOSTI), Malaysia. She was also supported by the special graduate research assistant scheme offered by UPM and through research grant from the Ministry of Education.

Throughout her PhD candidature, she actively participated in various research activities at national and international levels. These include outbound research attachment at International Rice Research Institute (IRRI), Philippines and rice conferences at state level (SARICON 2011), national level (NRC2011) and international level (IRC2014). Exhibitions participated include Malaysia Innovations Expo 2013 (MIExpo 2013) and BioMalaysia and Bioeconomy Asia Pacific Conference and Exhibition 2013. The patent filed, awards received and published journal articles and conference proceedings from her PhD program are presented in the next page.

LIST OF PUBLICATIONS

Patent Filed

Yiu, P. H.; Lee, H. H.; Wong, S. C.; Rajan, A. Multiplex Assay for Detection of Bario Rice Samples. PI2011005993, 2011.

Journal Papers

Lee, H.H., Loh, S.P., Bong, C.F.J., Sarbini, S.R. and Yiu, P.H. Impact of phytic acid on nutrients bioaccessibility and antioxidant properties of dehusked rice. *J. Food Sci. Technol.* **2015**. DOI 10.1007/s13197-015-1918-9.

Lee, H. H.; Bong, C. F. J.; Loh, S. P.; Sarbini, S. R.; Yiu, P. H. Genotypic , Grain Morphological and Locality Variation in Rice Phytate Content and Phytase Activity. *Emirates J. Food Agric.* **2014**, *26*, 844–852.

Dipti, S. S.; Bergman, C.; Indrasari, S. D.; Herath, T.; Hall, R.; Lee, H. H.; Habibi, F.; Bassinello, P. Z.; Graterol, E.; Ferraz, J. P.; *et al.* The Potential of Rice to Offer Solutions for Malnutrition and Chronic Diseases. *Rice (N. Y).* **2012**, *5*, 16.

Calingacion, M.; Laborte, A.; Nelson, A.; Resurreccion, A.; Concepcion, J. C. T.; Daygon, V. D.; Mumm, R.; Reinke, R.; Dipti, S.; Bassinello, P. Z.; *et al.* Diversity of Global Rice Markets and the Science Required for Consumer-Targeted Rice Breeding. *PLoS One* **2014**, *9*, e85106.

Conference Proceedings/Abstracts

Lee, H. H.; Loh, S. P.; Bong, C. F. J.; Yiu, P. H. Dietary Roles Of Intrinsic Phytate In Whole Grains Of Bario Rice. In *4th International Rice Congress 2014*; International Rice Research Institute (IRRI): Bangkok, Thailand, 2014.

Lee, H. H.; Bong, C. F. J.; Loh, S. P.; Yiu, P. H. Nutritional Quality Of Whole Grain Bario Rice. In *Sarawak Rice Conference 2011*; Sarawak Institute of Agriculture Scientist (SIAS): Kuching, Malaysia, 2011; p. 26.

Lee, H. H.; Wong, S. C.; Rajan, A.; Bong, C. F. J.; Yiu, P. H. Multiplex SSR Panels as Rapid Rice Genotyping Tool in Sarawak. In *Strengthening Food Security Through Sustainable Rice Production*; Othman, S.; Man, A.; Mokhtar, A.; Abd. Rahman, Z.; Abdul Razak, A.; Hashim, S.; Said, W., Eds.; Malaysian Agricultural Research and Development Institute (MARDI): Perak, Malaysia, 2010; pp. 390–393.

Awards

Young Rice Scientist Award on “Dietary roles of intrinsic phytate in whole grains of Bario rice”, given by International Rice Research Institute (IRRI) at 4th International Rice Congress 2014, Bangkok, Thailand, 30 Oct 2014).

Bioinnovation Award in Agriculture/Agro-based Industry (Bronze Medal) on “Multiplex Assay for Detection of Bario Rice Samples”, given by the organizers of BioMalaysia and Bioeconomy Asia Pacific Conferences and Exhibition 2013 (21 Oct 2013).

