



**UNIVERSITI PUTRA MALAYSIA**

***EFFECTS OF ETHANOLIC AZADIRACHTA EXCELSA (JACK) JACOBS.  
LEAF EXTRACT ON BLOOD PRESSURE AND OXIDATIVE STRESS IN  
THE SPONTANEOUSLY HYPERTENSIVE RAT MODEL***

**NUR FARHANA AHMAD SOPIAN**

**FPV 2017 24**



**UPM**  
UNIVERSITI PUTRA MALAYSIA  
BERILMU BERBAKTI

**EFFECTS OF ETHANOLIC *AZADIRACHTA EXCELSA* (JACK) JACOBS. LEAF  
EXTRACT ON BLOOD PRESSURE AND OXIDATIVE STRESS IN THE  
SPONTANEOUSLY HYPERTENSIVE RAT MODEL**

By

**NUR FARHANA AHMAD SOPIAN**

**Thesis Submitted to the School of Graduate Studies,  
Universiti Putra Malaysia, in Fulfilment of the  
Requirement for the Degree of Master of Science**

**August 2017**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## **DEDICATIONS**

This thesis is especially dedicated to my beloved mother, Salmah Ismail and to my late father Allahyarham Ahmad Sopian Omar. A lot of thanks to my siblings, family and friends for all the tremendous support, prayers and encouragement during my entire postgraduate life.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**EFFECTS OF ETHANOLIC *Azadirachta excelsa* (Jack) Jacobs. LEAF EXTRACT ON BLOOD PRESSURE AND OXIDATIVE STRESS IN THE SPONTANEOUSLY HYPERTENSIVE RAT MODEL**

By

**NUR FARHANA AHMAD SOPIAN**

**August 2017**

**Chair: Hafandi Ahmad, PhD**

**Faculty: Veterinary Medicine**

Oxidative stress is implicated in the development and maintenance of hypertension. To date the available therapies and oral antihypertensive drugs could only help in treating hypertension regardless of oxidative stress status and organ protective effect in the hypertensive patients. Hence, researchers are searching for antihypertensive plants that could complement oral drugs in hypertension. *Azadirachta excelsa* or locally known as “sentang” tree is one of the plants that have been used in Malay traditional medicine to treat diabetes. However, there were only a few studies regarding its antihypertensive and antioxidant activity on mammals. Therefore, this research aimed to evaluate the antihypertensive effects of *A. excelsa* by measuring the blood pressure of rats, serum analysis on aspartate aminotransferase (AST), alanine aminotransferase (ALT), creatinine phosphokinase (CPK) and lactate dehydrogenase (LDH). Meanwhile, its antioxidant effect on oxidative stress status was determined through organ analysis on malondialdehyde (MDA) level, antioxidant enzyme activity [Superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx)] and histopathological changes of tissue structures in the liver, kidney, and heart of SHR. In this study, twenty-four male SHR of 12-14 weeks old were assigned randomly into four different groups (n=6) as followed; Group I, SHR control received distilled water; Group II, SHR received 250 mg/kg of *A. excelsa*; Group III, SHR received 10 mg/kg of quercetin; and Group IV, SHR received 40 mg/kg of captopril. Meanwhile, male Wistar-Kyoto (WKY; n=6) rats were used as normotensive control also received distilled water. The treatments were given in 28 days where the blood pressure was measured weekly. Initially, systolic blood pressure (SBP) and diastolic blood pressure (DBP) of SHR were high significantly ( $P < 0.01$ ) compared to WKY. At the end of experiment, the SBP and DBP in SHR were alleviated by all treatment groups ( $P < 0.01$ ). Meanwhile, only quercetin-treated SHR showed significantly reduced serum enzymes of AST, ALT, LDH, and CPK. MDA level in the liver, kidney, and heart

of SHR were significantly higher ( $P < 0.05$ ) compared to WKY indicating impaired organ in these rats. Interestingly, treatment with *A. excelsa* and quercetin lower the MDA level in all organs ( $P < 0.05$ ). Furthermore, SOD activity in the heart from all treated SHR was decreased while only GPx activity in the liver and heart of *A. excelsa* treatment was increased. *A. excelsa* and quercetin treatments managed to improve hepatocyte structure. Also, all treatments managed to ameliorate glomerular and renal tubular lesions of the kidney, while only captopril treatment retained normal architecture of myocardial tissue of the heart. In conclusion, current findings suggested that *A. excelsa* ethanolic leaf extract could attenuate hypertension via amelioration of oxidative stress and improved tissue structure in SHR.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**KESAN EKSTRAK ETANOL DAUN *Azadirachta excelsa* (Jack) Jacobs.  
TERHADAP TEKANAN DARAH DAN TEGASAN OKSIDATIF KE ATAS  
MODEL TIKUS HIPERTENSI**

Oleh

**NUR FARHANA AHMAD SOPIAN**

**Ogos 2017**

**Pengerusi : Hafandi Ahmad, PhD  
Fakulti : Perubatan Veterinar**

Tegasan oksidatif dikaitkan dengan perkembangan dan penyelenggaraan tekanan darah tinggi. Setakat ini, terapi dan ubatan antihipertensi oral yang sedia ada hanya mampu membantu merawat hipertensi tanpa mengira status tegasan oksidatif dan melindungi organ pesakit hipertensi. Oleh itu, para penyelidik mencari pokok antihipertensi yang mampu bertindak sebagai pelengkap kepada ubatan oral di dalam hipertensi. *Azadirachta excelsa* atau pokok yang dikenali sebagai "sentang" adalah salah satu pokok yang digunakan dalam perubatan tradisional Melayu untuk merawat kencing manis. Walau bagaimanapun, terdapat hanya beberapa kajian sahaja mengenai aktiviti antihipertensi dan antioksidan pada mamalia. Oleh itu, kajian ini bertujuan untuk menilai kesan antihipertensi *A. excelsa* dengan mengukur tekanan darah tikus, analisis serum terhadap paras aspartate aminotransferase (AST), alanina aminotransferase (ALT), kreatina fosfokinase (CPK) dan laktat dehidrogenase. Sementara itu kesan antioksidan ke atas tegasan oksidatif diukur melalui analisis organ terhadap paras malondialdehyde (MDA) dan aktiviti enzim antioksidan [Superoksida dismutase (SOD), katalase (CAT) dan glutathione peroksidase (GPx)] dan perubahan histopatologi struktur tisu di dalam hati, ginjal, dan jantung daripada SHR. Di dalam kajian ini, dua puluh empat tikus SHR jantan berumur 12-14 minggu dibahagikan kepada empat kumpulan yang berbeza secara rawak (n=6) seperti berikut; Kumpulan I, SHR kawalan menerima air suling; Kumpulan II, SHR menerima 250 mg/kg *A.excelsa*; Kumpulan III, SHR menerima 10 mg/kg quercetin; dan Kumpulan IV, SHR menerima 40 mg/kg captopril. Sementara itu, tikus Wistar-Kyoto (WKY; n=6) dijadikan sebagai kawalan normotensif juga menerima air suling. Rawatan diberikan dalam masa 28 hari di mana tekanan darah diambil setiap minggu. Pada mulanya, tekanan sistolik (SBP) dan diastolik (DBP) dari tikus SHR adalah sangat tinggi ( $P<0.01$ ) berbanding tikus WKY. Pada akhir rawatan, semua rawatan telah berjaya mengurangkan tekanan SBP dan DBP ( $P<0.01$ ). Sementara itu, hanya tikus SHR yang dirawat dengan quercetin menunjukkan pengurangan signifikan terhadap paras serum AST, ALT, LDH dan CPK. Paras MDA diukur dalam hati, ginjal dan jantung kawalan SHR menunjukkan peningkatan signifikan ( $P<0.05$ ) berbanding dengan kawalan WKY,

menandakan organ yang terjejas dalam tikus ini. Menariknya, rawatan dengan *A.excelsa* dan quercetin mengurangkan paras MDA dalam semua organ ( $P < 0.05$ ). Tambahan lagi, aktiviti SOD dalam jantung daripada semua SHR yang dirawat telah menurun, sementara itu hanya aktiviti GPx di dalam hati dan jantung daripada rawatan *A.excelsa* telah meningkat. Rawatan *A.excelsa* dan quercetin berjaya memperbaiki struktur hepatosit. Dan juga, semua rawatan berjaya mengurangkan luka-luka tiub glomerular pada ginjal, manakala hanya rawatan captopril membaiki struktur normal tisu miokardium jantung. Kesimpulannya, penemuan terkini menunjukkan bahawa ekstrak *A.excelsa* boleh merawat tekanan darah tinggi dengan menangani tegasan oksidatif dan menambah baik struktur tisu dalam tikus SHR.





## ACKNOWLEDGEMENTS

In the name of Allah S. W. T, the most gracious and most merciful, praise be to Him for giving me the strength, courage and guidance during completing this thesis. First and foremost, I would like to express my deepest gratitude to my supervisor, Dr. Hafandi Ahmad for his guidance, suggestions, encouragement and giving me plenty of freedom and space to conduct my study as well as completing this thesis. My sincere appreciation is also extended to my co-supervisors, Assoc. Prof. Dr. Goh Yong Meng and Prof. Dr. Mohamed Ali Rajion for their help and valuable ideas shared throughout my research project. I would like to express my very great appreciation to my ex-supervisor during Final Year Project (UiTM), Mdm. Nurdiana Samsulrizal for her continuous support, motivation, and immense knowledge. Besides that, I would like to offer my special thanks Dr. Mokrish for his time and constructive suggestions during performing the research work. His knowledge offered to me was greatly appreciated.

I am particularly grateful for the assistance given by the laboratory staffs, Mdm. Zainab, Mdm. Rosmawati, Mr. Hafiz and Mr. Jamil, for their kindness to allow me to use all the laboratory apparatus and equipment freely while completing this project. Besides, I would also like to thank Dr. Hj. Ahmad Zuhaidi Yahya and En. Mohd Noor Mahat from Forest Research Institute Malaysia (FRIM) for their generosity in helping me collecting samples needed for my study. Acknowledgments are also extended to Mdm. Fazila Sulaiman, a research officer from Agro-Bioteknologi Institut (ABI), MARDI for her guidance during handling the blood pressure machine.

My warmest appreciation also goes to all of my beloved friends, Nurul 'Izzati Shafie, Nur Ain Amin, Noraini Shamsudin, Ummu Afiqah Hassan and to all my friends who those involve directly or indirectly in supporting and helping me undoubtedly to run this project successfully. Last but not least, my dearest appreciation and thanks to my beloved mother, Salmah binti Ismail and all my family members for their pray and support that strengthen me to complete this project. All of the contributions are really appreciated and only Allah S.W.T could give something as return. Thank you very much.

I certify that a Thesis Examination Committee has met on 8th August 2017 to conduct the final examination of Nur Farhana Binti Ahmad Sopian on her thesis entitled “Effects of Ethanolic Azadirachta excelsa (Jack) Leaf Extract on Blood Pressure and Oxidative Stress in the Spontaneously Hypertensive Rat Model” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**RASEDEE @ MAT BIN ABDULLAH, PHD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**MOHD HEZMEE BIN MOHD NOOR, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Internal Examiner)

**SULAIMAN BIN MD DOM, PhD**

Associate Professor  
Faculty of Health Sciences  
Universiti Teknologi Mara Selangor  
(External Examiner)

---

**NOR AINI AB. SHUKOR, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 26 October 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Hafandi bin Ahmad, PhD**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Goh Yong Meng, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Mohamed Ali bin Rajion, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: \_\_\_\_\_

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of member of  
Supervisory  
Committee: \_\_\_\_\_

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF APPENDICES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xvi
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	
1.1 Background of study	1
1.2 Significance of study	2
1.3 Objectives of study	3
1.4 Hypotheses	3
1.5 Justifications of the study	3
<b>2 LITERATURE REVIEW</b>	
2.1 Hypertension	5
2.1.1 Essential hypertension	6
2.1.2 Secondary hypertension	7
2.1.3 Stages of hypertension	9
2.1.4 The impact of hypertension	10
2.2. Pathogenesis of hypertension	10
2.2.1 Reactive oxygen species (ROS) in blood vessels	11
2.2.2 Renin-Angiotensin Aldosterone System (RAAS)	20
2.3 Medication used for hypertension	22
2.3.1 Captopril	22
2.4 Hypertensive animal model	23
2.5 Plant remedies in hypertension treatment	24
2.6 <i>Azadirachta excelsa</i>	25
2.6.1 Geographical and botanical Description	26
2.6.2 Phytochemicals in <i>Azadirachta excelsa</i>	26
2.6.3 Biological activities of <i>Azadirachta excelsa</i>	27
2.6.4 Antidiabetic effect of <i>Azadirachta excelsa</i>	27
<b>3 MATERIALS AND METHOD</b>	
3.1 Preparation of <i>Azadirachta excelsa</i> leaf powder and ethanolic crude extract	29
3.2 Experimental location and animal housing	29
3.3 Experimental design	29
3.4 Collection of Blood and Organ Samples	30

<b>4</b>	<b>QUANTITATIVE ANALYSIS AND ANTIOXIDANT ACTIVITIES OF <i>Azadirachta excelsa</i> ETHANOLIC LEAF EXTRACT</b>	
4.1	Introduction	33
4.2	Objective	33
4.3	Materials and methods	34
4.3.1	Chemicals and raw materials	34
4.3.2	Preparation of leaves powder and ethanolic extract	34
4.3.3	Quantitative analysis	34
4.3.4	<i>In- vitro</i> antioxidant activities	35
4.4	Results	36
4.4.1	Quantitative analysis of phytochemicals on phenolics and flavonoids content	36
4.4.2	<i>In-vitro</i> antioxidant study of <i>Azadirachta excelsa</i> leaf extract (DPPH assay)	36
4.5	Discussion	38
4.5.1	Quantitative analysis of phytochemicals on TPC and TFC	38
4.5.2	<i>In- vitro</i> antioxidant study of <i>Azadirachta excelsa</i> leaf extract	39
4.6	Conclusions	40
<b>5</b>	<b>ANTIHYPERTENSIVE EFFECTS OF ETHANOLIC <i>Azadirachta excelsa</i> LEAF EXTRACT ON SERUM ENZYMES</b>	
5.1	Introduction	41
5.2	Objectives	42
5.3	Materials and methods	42
5.3.1	Measurement of blood pressure on single oral dose and 28 days administration of <i>Azadirachta excelsa</i> ethanolic leaf extract	42
5.3.2	Measurement of body weight and organs weight (somatic index)	43
5.3.3	Serum analysis	43
5.4	Data analysis	44
5.5	Results	44
5.5.1	Single oral dose administration on systolic blood pressure of SHR	44
5.5.2	Blood pressure in SHR during 28 days administration	47
5.5.3	Body weight during treatment	51
5.5.4	Organ weight over body weight ratios	53
5.5.5	Serum analysis of LDH and CPK	55
5.6	Discussion	57
5.6.1	Single oral dose of <i>Azadirachta excelsa</i> leaf extract administration on blood pressure of SHR	57
5.6.2	Blood pressure level in SHR during 28 days administration	57
5.6.3	Body weight	58
5.6.4	Organ weight over body weight ratios	59
5.6.5	Serum analysis	59
5.7	Conclusion	60

<b>6</b>	<b>ASSESSMENT ON MALONDIALDEHYDE (MDA) AND ANTIOXIDANT ENZYME ACTIVITIES LEVEL IN KIDNEY, LIVER AND HEART.</b>	
6.1	Introduction	61
6.2	Objectives	62
6.3	Materials and methods	62
6.3.1	Processing of tissues	62
6.3.2	Protein assay	62
6.3.3	Measurement of malondialdehyde (MDA) level	62
6.3.4	Antioxidant enzyme activity	63
6.4	Data analysis	64
6.5	Results	64
6.5.1	Measurement of malondialdehyde (MDA) level	64
6.5.2	Antioxidant enzyme activity in organs	66
6.5.3	Glutathione peroxidase (GPx) activities	68
6.6	Discussion	70
6.6.1	Malondialdehyde (MDA) level	70
6.6.2	Antioxidant enzyme activity in organs	71
6.7	Conclusions	72
<b>7</b>	<b>EVALUATION ON POTENTIAL TOXICITY CHANGES ASSOCIATED WITH THE ADMINISTRATION OF ETHANOLIC <i>Azadirachta excelsa</i> EXTRACT</b>	
7.1	Introduction	73
7.2	Objectives	74
7.3	Materials and methods	74
7.3.1	Serum analysis on toxicity	74
7.3.2	Tissue collection and histopathology	74
7.3.3	Lesions scoring for liver, kidney, and heart	75
7.4	Data analysis	75
7.5	Results	76
7.5.1	Serum level of AST and ALT	76
7.5.2	Histopathological findings of organs	78
7.5.3	Scoring index of liver	90
7.5.4	Scoring index of kidney	90
7.5.5	Scoring index of heart	91
7.6	Discussion	92
7.7	Conclusions	95
<b>8</b>	<b>GENERAL DISCUSSION AND CONCLUSION</b>	<b>96</b>
	<b>REFERENCES</b>	<b>99</b>
	<b>APPENDICES</b>	<b>116</b>
	<b>BIODATA OF STUDENT</b>	<b>121</b>
	<b>LIST OF PUBLICATIONS</b>	<b>122</b>



## LIST OF TABLES

Table		Page
1	Stages of hypertension	9
2	Total phenolic and flavonoid content of <i>Azadirachta excelsa</i>	36
3	Single oral dose administration on blood pressure of SHR	45
4	Heart rate changes (bpm) of rats during treatment	50
5	Body weight changes (g) during treatment	52
6	Somatic index (% of body weight) of SHR's organ	54
7	Superoxide dismutase (SOD) enzyme activities	67
8	Catalase (CAT) enzyme activities	67
9	Scoring index of liver	90
10	Scoring index of kidney	90
11	Scoring index of heart	91

## LIST OF FIGURES

Figure		Page
1	Interaction of genetic and environmental factors in the development of primary hypertension	7
2	The mechanism in the development of hypertension	8
3	Sources of ROS in vascular cells	13
4	ROS and the enzymes that regulate their level	15
5	Combination of several mechanisms leading to hypertension	18
6	Angiotensin-Converting Enzyme (ACE) mechanism	21
7	<i>Azadirachta excelsa</i> tree	26
8	Summary of research approach for plant study	31
9	Summary of experimental design for animal study	32
10	Percentage of DPPH scavenging activity against samples	37
11	Percentage of DPPH scavenging activity at 500 µg/ml	37
12	Systolic blood pressure (SBP) level (mmHg) at different doses of <i>A. excelsa</i>	46
13	Systolic blood pressure (SBP) level (mmHg) of rats	48
14	Diastolic blood pressure (DBP) level (mmHg) of rats	49
15	CPK level (U/L) after 4weeks treatment	56
16	LDH level (U/L) after 4weeks treatment	56
17	MDA level (nmol/mg) in liver	65
18	MDA level (nmol/mg) in kidney	65
19	MDA level (nmol/mg) in heart	66
20	GPx level (nmol/mg) in liver	68
21	GPx level (nmol/mg) in kidney	69
22	GPx level (nmol/mg) in heart	69
23	Serum level of AST (U/L)	77
24	Serum level of ALT (U/L)	77
25	Photomicrographs of liver of rats of each group	79-81
26	Photomicrographs of kidney of rats of each group	83-85
27	Photomicrographs of heart of rats of each group	87-89

## LIST OF APPENDICES

<b>Appendix</b>		<b>Page</b>
1	Animal ethic form	119
2	Standard curve of BCA protein and MDA	120
3	Standard curve of CAT and SOD enzyme	121
4	Standard curve of GPx and gallic acid	122
5	Standard curve of rutin and DPPH assay	123



## LIST OF ABBREVIATIONS

AAPH	2,2'-azobis (2-amidinopropane)
ACE	Angiotensin converting enzyme
ACEI	Angiotensin converting enzyme inhibitors
ADH	anti-diuretic hormone
AGT	Angiotensinogen
AHA	American Heart Association
ALT	Alanine aminotransferase
ANG I	Angiotensin I
ANG II	Angiotensin II
ANOVA	Analysis of variance
AST	Aspartate aminotransferase
BBB	blood-brain barrier
BPM	beats per minute
CAT	Catalase
CI	cardiosomatic index
CVD	Cardiovascular disease
CVO	circumventricular organs
DBP	Diastolic blood pressure
DPPH	2,2-diphenyl-1-picrylhydrazyl
eNOS	endothelial nitric oxide synthase
GR	glutathione reductase
GSSG	oxidized glutathione
GPx	Glutathione peroxidase
H <sub>2</sub> O <sub>2</sub>	hydroxyl peroxide
HSI	Hepatosomatic index
IC <sub>50</sub>	half maximal inhibitory concentration
ICV	intracerebroventricular
KSI	kidney somatic index
LDLs	Low density lipoproteins
MDA	Malondialdehyde
MmHg	Millimeter per mercury
NADPH	Nicotinamide adenine dinucleotide phosphate
NO	nitric oxide
PKD	polycystic kidney disease
PUFA	Polyunsaturated fatty acid
RAAS	Renin angiotensin aldosterone system
ROS	Reactive oxygen species
SBP	Systolic blood pressure
SEM	Standard error of mean
SHR	Spontaneously hypertensive rat
SOD	Superoxide dismutase
VSMC	vascular smooth muscle cell
WHO	World Health Organization
WKY	Wistar-Kyoto rat

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of study

Hypertension is a silent killer disease manifested as the major contributor to the development of many cardiovascular diseases (CVD). It was reported that CVD is the main cause of mortality and morbidity rate in a worldwide population contributed to 16.7 million deaths in worldwide each year (WHO, 2003). Back in 2009, it was estimated about 12.8% of total deaths per year in which 7.5 million from the total deaths is accredited to hypertension (WHO, 2009). However, because of population growth and aging, the number had increased to 16.5% of annual deaths worldwide where hypertension is recognized as leading cause associated with CVD (WHO, 2013). According to WHO (2013), the annual death toll is estimated to reach 23.5 million people by 2030. Commonly, people with high blood pressure are at high risk of CVD such as heart attack, stroke, and atherosclerosis. Other than that, hypertension also lead to renal damage, dementia, or blindness (August, 2004; Freedman and Cohen, 2016). Thus, alleviating hypertension can reduce the risk of mortality associated with CVD and others complications in hypertensive patients (Chobanian et al., 2003).

In current medical practice, there are many types of antihypertensive drugs prescribed for hypertension although sometimes the exact etiology of hypertension remains unknown. Despite this, there is increasing concern about the quality of patient's life receiving treatment for hypertension as these regimes may be associated with various adverse effects. Specifically, elderly hypertensive patients are little more prone to side-effects. Most of the synthetic antihypertensive drugs often cause undesired side effects such as dehydration, vomiting, and cough. As an example, beta-blockers and diuretics drugs only reduce blood pressure but unable to prevent organ damage (Clinical Practice Guidelines for the Management of Hypertension, 2013). In order to achieve an optimal effect of hypotensive action, the efficiency of the drugs are only 34% when two or more antihypertensive drugs from different categories were combined (August, 2004; Yeh et al., 2009; Wang and Xiong, 2012). Major concerns often delay the treatment are represented by their availability and accessibility (Wang and Xiong, 2012), unwanted side effects (Susalit et al., 2011; Wang and Xiong, 2012) and higher costs of antihypertensive drugs (Susalit et al., 2011) when combination of two or more pills are needed. Thus, researchers revealed many potential plant species that possessed antihypertensive properties. For instance, high intake of natural flavonoids from medicinal plants in the diet was reported to have an inverse relationship to risk of CVD (Knekt et al., 2002). They play a protective role against the development of pathological conditions such as hypertension, diabetes, and obesity which associated to oxidative stress. Oxidative stress emerges as a key regulator of blood pressure regulation by

balancing vasoconstrictor and vasodilator mechanisms. Disruption of the mechanism leads to a persistent vasoconstriction and resulted in an elevation of blood pressure. As hypertension is one of the most common metabolic disorders characterized by oxidative stress, there is a growing interest of inhibiting oxidative stress by using natural antioxidants in plants such as phenolic and flavonoid compounds.

Commonly, the hypertensive patients from low-income group, especially rural dwellers in developing countries are seeking alternative approaches such as herbal remedies to control hypertension and its concomitant complications in face of limited socioeconomic resources (Chockalingam et al., 2006). Therefore, plant-based remedies are being widely investigated by researchers to provide alternatives in treating hypertension. Currently, various species of plants have been identified to exert antioxidant associated with antihypertensive properties (Nahida and Feroz, 2011). However, not all of them have received scientific scrutiny and therefore scientific research needs to be done to verify their effectiveness and elucidate safety profile of such herbal remedies.

*Azadirachta excelsa* or locally known as “sentang” belongs to Meliaceae family where it is a plant family that recognizable with various benefits associated with medical properties. A number of studies have investigated the biological activities of *A. excelsa* such as antiseptic, anti-inflammatory, antimicrobial, antifeedant and insecticidal agent (Ng et al., 2003; Akhtar et al., 2008; Mustafa and Al-Khazraji, 2008). Meanwhile, our previous finding on animal studies had proven that *A. excelsa* acts as an antidiabetic agent by reducing blood glucose level in type 1 and type 2 of diabetes mellitus (Nurdiana et al., 2013; Nurdiana et al., 2014; Nurul ‘Izzati 2016). However, *A. excelsa* has not been studied extensively regarding its antihypertensive activity. Thus, the current study was designed primarily to determine its effect on blood pressure in genetic hypertension animal model. This study may also provide an alternative source of natural herbs potential namely *A. excelsa* as antihypertensive agents with probably fewer or no adverse side effect.

## 1.2 Significance of study

The study was designed primarily to evaluate the antioxidant activities of *A. excelsa* leaf extract on Spontaneously Hypertensive Rat (SHR) model. This study allows the plant to be scientifically prove as remedies for hypertension as well as its ability in scavenging free radicals. As encouraged by World Health Organization (WHO), the present study can be a turning point in enhancing more research involving natural herbs as drugs of tomorrow to improve our quality life.

### 1.3 Objectives of study

Main objective;

To determine the antioxidant activities of *A. excelsa* leaf extract on blood pressure and oxidative stress in SHR animal model.

Specific objectives;

1. To quantify the total phenolic, total flavonoids content and the IC<sub>50</sub> value of *A. excelsa*.
2. To investigate the effects of *A. excelsa* leaf extract on elevated blood pressure in SHR model and serum level of cardiac injury.
3. To evaluate the effects of *A. excelsa* leaf extract on oxidative stress marker and antioxidant enzyme activities in SHR's organs.
4. To determine the effects of *A. excelsa* leaf extract on serum level of liver toxicity and histopathological changes of liver, kidney and heart in SHR model.

### 1.4 Hypotheses

It was hypothesized that ;

Antioxidant activities of *A. excelsa* exhibited an antihypertensive effect on SHR model by ameliorating serum level of cardiac injury.

*A. excelsa* mitigates ROS level and lipid peroxidation process in SHR's organs by altering the activities of superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx) antioxidant enzymes.

Administration of *A. excelsa* ethanolic extract for 28 days reduced serum liver toxicity as well as improve the histopathological structure and the extent of oxidative stress damage.

### 1.5 Justifications of the study

Recently, several studies were reported on the antidiabetic effect of *A. excelsa* in type I and type II diabetic rats (Nurdiana et al., 2013 and 2014; Nurul 'Izzati, 2016). However, the effect of *A. excelsa* on oxidative stress, specifically in hypertensive rats remains elusive and yet to be investigated. On those studies, the approach used to induce diabetes was by an injection of streptozotocin (STZ). The injection generates free radicals from  $\beta$ -cells of pancreatic islet that are vulnerable and sensitive to oxidative stress (Robertson et al., 2003). As the main underlying cause of diabetes and hypertension is oxidative stress, the current

study was done to investigate effects of *A. excelsa* on oxidative stress in genetic hypertension of SHR model.

The uses of SHR as an animal model to study the pathogenesis of hypertension and screening any potential antihypertensive plants are well documented. While, an elevation and progression of blood pressure in this genetic animal model were attributable by oxidative stress (Hirooka et al., 2011; Fanelli and Zatz, 2011). Therefore, this study was conducted to evaluate the effect of *A. excelsa* on blood pressure in SHR while examining oxidative stress level in organs.

Quercetin was chosen at 10 mg/kg based on the average of human intake in the diet (Duarte et al., 2001b). It was included in the current study to compare between the treatment of quercetin with crude extract from *A. excelsa*, since quercetin was the purified plant phenols found in *Azadirachta* sp (Sithisarn et al., 2007). Meanwhile, captopril was chosen as a positive control in the treatment because it causes little side effects and possessed antioxidant as well as renoprotective effects (Dzau et al., 2001).



## REFERENCES

- Abdollahi, M., Zuki, A. B. Z., Goh, Y. M., Rezaeizadeh, A. and Noordin, M. M. (2010). The effects of *Momordica charantia* on the liver in streptozotocin-induced diabetes in neonatal rats. *African Journal of Biotechnology*, 9(31): 5004–12.
- Afsana Khatun, M., Harun-Or-Rashid, M. and Rahmatullah, M. (2011). Scientific validation of eight medicinal plants used in traditional medicine systems of Malaysia: A review. *American-Eurasian Journal of Sustainable Agriculture*, 5(1): 67-75.
- Ajay, M., Chai, H. J., Mustafa, A. M., Gilani, A. H. and Mustafa, M. R. (2007). Mechanisms of anti-hypertensive effect of *Hibiscus sabradiffa* L. calyces. *Journal of Ethnopharmacology*, 109: 388-393.
- Akanji, M. A., Nafiu, M. O. and Yakubu, M. T. (2008). Enzyme activities and histopathology of selected tissues in rats treated with potassium bromated. *African Journal of Biomedical Research*, 11: 87-95.
- Akhtar, Y., Yeoung, Y. R. and Isman, M. B. (2008). Comparative bioactivity of selected extracts from Meliaceae and some commercial botanical insecticides against teo noctuid caterpillars, *Trichoplusia ni* and *Pseudaletia unipuncta*. *Phytochemical Reviews*, 7(1): 77-88.
- Al Disi, S. S., Anwar, M. A. and Eid, A. H. (2016). Anti-hypertensive Herbs and their Mechanisms of Action : Part I. *Frontiers in Pharmacology*, 323 (6) ; 1-24.
- Andriansitohaina, R., Auger, C., Chataigneau, T., Etienne-Selloum, N., Li, H., Martinez, M. C., Schini-Kerth, V. B. and Laher, I. (2012). Molecular mechanisms of the cardiovascular protective effects of polyphenols. *British Journal of Nutrition*, 1:1-18.
- August, P. (2004). Overview: mechanisms of hypertension: cells, hormones, and the kidney. *Journal of American Society Nephrology*, 15: 1971–1973.
- Bartosz, M., Kedziora, J. and Bartosz, G. (1997). Antioxidant and pro-oxidant properties of captopril and enalapril. *Free Radical Biology and Medicine*, 23:729–35.
- Bernatova, I., Pechanova, O., Pelouch, V. and Simko, F. (2000). Regression of chronic L-NAME-treatment-induced left ventricular hypertrophy: effect of captopril. *Journal Molecular Cell Cardiology*, 32: 177-185.
- Berra, K. and Miller, N. H. (2009). Inhibiting the renin-angiotensin system: why and in which patients. *Journal American Academy Nurse Practice*, 21(1):66–75.

- Bolterman, R. J., Manriquez, M. C., Ortiz Ruiz, M. C., Juncos, L. A. and Romero, J. C. (2005). Effects of captopril on the renin angiotensin system, oxidative stress, and endothelin in normal and hypertensive rats. *Hypertension*, 46: 943-947.
- Bonetti, P. O., Lerman, L. O. and Lerman, A. (2003). Endothelial dysfunction a marker of atherosclerotic risk. *Arteriosclerosis Thrombosis and Vascular Biology*, 23:168-175.
- Botelho-Ono, M.S., Pina, H. V. Sousa, K. H. F., Nunes, F. C., Medeiros, I. A. and Braga, V. A. (2011). Acute superoxide scavenging restores depressed baroreflex sensitivity in renovascular hypertensive rats. *Autonomic Neuroscience*, 159: 38-44.
- Braga, V. A. (2012). Depressed Baroreflex Sensitivity in Hypertensive Rats: A Role for Reactive Oxygen Species. *Journal Hypertension*, 1, e103.
- Brandes, R. P., Miller, F. J., Beer, S., Haendeler, J., Hoffmann, J., Ha, T., Holland, S. M., Gorlach, A. and Busse, R. (2002). The vascular NADPH oxidase subunit p47phox is involved in redox-mediated gene expression. *Free Radical Biology and Medicine*, 32:1116-1122.
- Campbell, N. A., Reece, J. B., Urry, L. A., Cain, M. L., Wasserman, S. A., Minorsky, P. V. and Jackson, R. B. (2002). *Biology* (Edition 8th). (pp 952-972). Pearson Benjamin Cummings Publishers.
- Carretero, O. A. and Oparil, S. (2000). Essential hypertension. Part I: definition and etiology. *Circulation*, 101: 329-35.
- Cediël, E., Sanz-Rosa, D., Oubina, M. P., de las, H. N., Gonzalez, P. F. R., Vegazo, O., Jimenez, J., Cachofeiro, V. and Lahera, V. (2003). Effect of AT1 receptor blockade on hepatic redox status in SHR: possible relevance for endothelial dysfunction? *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*, 85: 674-681.
- Chabrashvili, T., Tojo, A., Onozato, M. L., Kitiyakara, C., Quinn, M. T., Fujita, T., Welch, W. J. and Wilcox, C. S. (2002). Expression and cellular localization of classic NADPH oxidase subunits in the spontaneously hypertensive rat kidney. *Hypertension*, 39:269-274.
- Chade, A. R., Rodriguez-Porcel, M. and Herrmann, J. (2004). Antioxidant intervention blunts renal injury in experimental renovascular disease. *Journal of American Society of Nephrology*, 15: 958-66.
- Chahal, C. A. A. and Somers, V. K. (2015). Secondary hypertension: Obstructive sleep apnea. *Journal society of Hypertension*, 9(3): 244-247.

- Chakraborty, T., Uerotta, L. and Poddar, G. (1989). Evaluation of *Azadirachta indica* leaf extract for hypoglycemic activity in rats. *Phytotherapy Research*, 3: 30-32.
- Chan, K. W., Khong, N. M. H., Iqbal, S., Mansor, S. M. and Ismail, M. (2013). Defatted kenaf seed meal (DKSM): Prospective edible flour from agricultural waste with high antioxidant activity. *Food Science and Technology*, 53(1): 308-313.
- Chattopadhyay, R. R. and Bandyopadhyay, M. (2005). Effects of *Azadirachta indica* leaf extract on serum lipid profile changes in normal and streptozotocin induced diabetic rats. *African Journal of Biomedical Research*, 8:101-104.
- Chen, Q., Xuan, G., Fu, M., He, G., Wang, W., Zhang, H., and Ruan, H. (2007). Effect of angiotensin I-converting enzyme inhibitory peptide from rice dregs protein on antihypertensive activity in spontaneously hypertensive rats. *Asian Pacific Journal of Clinical Nutrition*, 16(1): 281-285.
- Chobanian, A. V., Bakris, G. L., Black, H. R., Cushman, W. C., Green, L. A., Izzo, J. L., Jones, D. W., Materson, B. J., Oparil, S. and Wright, J. T. (2003). Seventh report of the joint national committee on prevention, detection, evaluation, and treatment of high blood pressure. *Hypertension*, 42: 1206–1252.
- Chockalinga, A., Campbell, N.R. and Fodor, J.G. (2006). Worldwide epidemic of hypertension. *Canada Journal Cardiology*, 22(7): 553-555.
- Choi, H. S., Cho, H. Y., Yang, H. C., Ra, K. S. and Suh, H. J. (2001). Angiotensin I-converting enzyme inhibitor from *Grifola frondosa*. *Food Research International*, 34: 177-182.
- Clinical practice guidelines for the management of hypertension. (2013). Retrieved from [http://www.moh.gov.my/attachments/CPG\\_Management\\_of\\_Hypertension\\_4th\\_Edition.pdf](http://www.moh.gov.my/attachments/CPG_Management_of_Hypertension_4th_Edition.pdf)
- Crowley, S. D., Gurley, S. B., Herrera, M. J., Ruiz, P., Griffiths, R., Kumar, A. P., Kim, H-S., Smithies, O., Le, T. H. and Coffman, T. M. (2006). Angiotensin II causes hypertension and cardiac hypertrophy through its receptors in the kidney. *Proceedings of the National Academy of Sciences of the United States of America*, 103(47): 17985-17990.
- Csonka, C., Pataki, T., Kovacs, P., Muller, S. L., Schroeter, M. L., Tosaki, A., and Blasig, I. E. (2000). Effects of oxidative stress on the expression of antioxidative defense enzymes in spontaneously hypertensive rat hearts. *Free Radical Biology and Medicine*, 29(7): 612-619.

- Cui, B., Chai, H., Constant, H. L., Santisuk, T., Reutrakul, V., Beecher, C. W. W., Farnsworth, N. R., Cordell, G. A., Pezzuto, J. M. and Kinghorn, A. D. (1998). Limonoids from *Azadirachta excelsa*. *Phytochemistry*, 47: 1283-1287.
- Dawood, T. and Schilaich, M. P. (2009). Mediators of target organ damage in hypertension: Focus on obesity associated factors and inflammation. *Minerva Cardioangiologica*, 57: 687-703.
- Deanfield, J., Donald, A., Ferri, C., Giannattasio, C., Halcox, J., Halligan, S., Lerman, A., Mancina, G., Oliver, J. J., Pessina, A.C., Rizzoni, D., Rossi, G. P., Salvetti, A., Schiffrini, E. L., Taddei, S. and Webbe, D. J. (2005). Endothelial function and dysfunction. Part I: Methodological issues for assessment in the different vascular beds : A statement by the Working Group on Endothelin and Endothelial Factors of the European Society of Hypertension. *Journal of Hypertension*, 23(1) :7-17.
- de Cavanagh, E.M., Insera, F., Ferder, L. and Fraga, C.G., (2000). Enalapril and captopril enhance glutathione-dependent antioxidant defenses in mouse tissues. *American Journal Physiology Regulation Integrative Comparative Physiology*, 278: R572-R577.
- De Haan, J. B., Bladier, C., Griffiths, P., Kelner, M., O'Shea, R. D., Cheung, N. S., Bronson, R. T., Silvestro, M. J., Wild, S., Zheng, S. S., Beart, P. M., Hertzog, P. J. and Kola, I. (1998). Mice with a homozygous null mutation for the most abundant glutathione peroxidase, GPx1, show increased susceptibility to the oxidative stress-inducing agents paraquat and hydrogen peroxide. *The Journal of Biological Chemistry*, 273(35): 22528-22536.
- Dey, A. and Cederbaum, A. I. (2006). Alcohol and oxidative liver injury. *Hepatology*, 43(2-1): 63-74.
- DiBona, G. F. (2004). The sympathetic nervous system and hypertension: recent developments. *Hypertension*, 43:147-150.
- Duarte, J., Galisteo, M., Ocete, M. A., Perez-Vizcaino, F., Zarzuelo, A. and Tamargo, J. (2001a). Effects of chronic quercetin treatment on hepatic oxidative status of spontaneously hypertensive rats. *Molecular and Cellular Biochemistry*, 221: 155-160.
- Duarte, J., Perez-Palencia, R., Vargas, F., Ocete, M. A., Perez-Vizcaino, F., Zarzuelo, A. and Tamargo, J. (2001b). Antihypertensive effects of the flavonoid quercetin in spontaneously hypertensive rats. *Journal of Pharmacology*, 133: 117-124.
- Dzau, V. J., Bernstein, K., Celermajer, D., Cohen, J., Dahlof, B. and Deanfield, J. (2001). The relevance of tissue angiotensin-converting enzyme: manifestations in mechanistic and endpoint data. *American Journal of Cardiology*, 88:1L-20L.

- Ellis, G. R., Anderson, R. A. and Lang, D. (2000). Neutrophil superoxide anion-generating capacity, endothelial function and oxidative stress in chronic heart failure: effects of short- and long-term vitamin C therapy. *Journal American College of Cardiology*, 36:1474-82.
- Erdos, B., Broxson, C. S., King, M. A., Scarpace, P. J. and Tumer, N. (2006). Acute pressor effect of central angiotensin II is mediated by NAD(P)H-oxidase-dependent production of superoxide in the hypothalamic cardiovascular regulatory nuclei. *Journal Hypertension*, 24:109–16.
- Erejuwa, O. O., Siti, A. S., Mohd, S. A. W., Kuttulebbai, N. S. S., Salzihan, S. and Sunil, G. (2012). Honey supplementation in Spontaneously Hypertensive Rats elicits antihypertensive effect via amelioration of renal oxidative stress. *Oxidative Medicine and Cellular Longevity*, 1-14.
- Eser, I., Khorshid, L., Gunes, U. Y. and Demir, Y. (2007). The effect of different body positions on blood pressure. *Journal of Clinical Nursing*, 16: 137-140.
- Evans, M. D., Dizdaroglu, M. and Cooke, M. S. (2004). Oxidative DNA damage and disease: induction, repair and significance. *Mutation Research*, 567:1-61.
- Ezzati, M., Lopez, A. D., Rodgers, A., Vandern Hoorn, S. and Murray, C. J. (2002). Selected major risk factors and global and regional burden of disease. *Lancet*, 360: 1347–1360.
- Fanelli, C. and Zatz, R. (2011). Linking oxidative stress, the renin-angiotensin system, and hypertension. *Hypertension*, 57(3): 373-374.
- Feng, M., Whitesall, S., Zhang, Y., Beibel, M., D'Alecy, L. and DiPetrillo, K. (2008). Validation of volume-pressure recording tail-cuff blood pressure measurements. *American Journal of Hypertension*, 21(12): 1288-1291.
- Freedman, B. I. and Cohen, A. H. (2016). Hypertension-attributed nephropathy: what's in a name? *Nature Reviews Nephrology*, 12: 27–36.
- Fujii, S., Zhang, L., Igarashi, J. and Kosaka, H. (2003). L-arginine reverses p47phox and gp91phox expression induced by high salt in Dahl rats. *Hypertension*, 42: 1014–1020.
- Galisteo, M., García-Saura, M. F., Rosario Jiménez, R., Villar, I. C., Antonio Zarzuelo, A., Vargas, F. and Duarte, J. (2004). Effects of chronic quercetin treatment on antioxidant defence system and oxidative status of deoxycorticosterone acetate-salt-hypertensive rats. *Molecular and Cellular Biochemistry*, 259: 91–99.

- Gomez-Amores, L., Mate, A., Revilla, E., Santa-Maria, C. and Vazquez, C. M. (2006). Antioxidant activity of propionyl-L-carnitine in liver and heart of spontaneously hypertensive rats. *Life Sciences*, 78: 1945-1952.
- Gomez-Amores, L., Mate, A., Miguel-Carrasco, J. L., Jimenez, L., Jos, A., Camean, A. M., Revilla, E., Santa-Maria, C. and Vazquez, C. M. (2007). L-carnitine attenuates oxidative stress in hypertensive rats. *Journal of Nutritional Biochemistry*, 18: 533-540.
- Gospodaryov, D. and Lushchak, V. (2012). Oxidative Stress: Cause and Consequence of Diseases, *Oxidative Stress and Diseases*, Dr. Volodymyr Lushchak (Ed.), ISBN: 978-953-51-0552-7, InTech, Available from: <http://www.intechopen.com/books/oxidative-stress-and-diseases/oxidative-stress-as-a-cause-and-consequence-of-diseases>
- Guimarães, D. D., Carvalho, C. C. and Braga, V. A. (2012). Scavenging of NADPH oxidase-derived superoxide anions improves depressed baroreflex sensitivity in spontaneously hypertensive rats. *Clinical and Experimental Pharmacology and Physiology*, 39: 373–378.
- Hall, J. E. (2003). The kidney, hypertension and obesity. *Hypertension* 41: 625–633.
- Halliwell, B. and Gutteridge, J. M. C. (1989). *Free Radicals in Biology and Medicine*. Clarendon Press Oxford.
- Harrison, D. G., Gongora, M. C., Guzik, T. J. and Widder, J. (2007). Oxidative stress and hypertension. *Journal of the American Society of Hypertension*, 1(1): 30-44.
- Heitzer, T., Schlinzig, T., Krohn, K., Meinertz, T. and Münzel, T. (2001). Endothelial Dysfunction, Oxidative Stress, and Risk of Cardiovascular Events in Patients With Coronary Artery Disease. *Circulation*, 104:2673-2678
- Hirunpanich, V., Utaipat, A., Morales, N. P., Bunyaphatsara, N., Sato, H., Herunsalee, A. and Suthisisang, C. (2005). Antioxidant effects of aqueous extracts from dried calyx of *Hibiscus sabdariffa* LINN (Roselle) in vitro using rat low-density lipoprotein (LDL). *Biological and Pharmaceutical Bulletin*, 28: 481–484.
- Hirooka, Y. (2011). Oxidative stress in the cardiovascular center has a pivotal role in the sympathetic activation in hypertension. *Hypertension Research*, 34(4): 407-412.
- Hodgson, J. M. and Croft, K. D. (2006). Review. Dietary flavonoids: effects on endothelial function and blood pressure. *Journal of Science of Food and Agriculture*, 86: 2492-2498.

- Hoetzel, A., Welle, A., Schmidt, R., Loop, T., Humar, M., Ryter, S. W., Geiger, K. K., Choi, A. M. K. and Pannen, B. H. J. (2008). Nitric oxide-deficiency regulates hepatic heme oxygenase-1. *Nitric Oxide Biology and Chemistry*, 18: 61–69.
- Houston, M. C. (2010). The role of cellular micronutrient analysis, nutraceuticals, vitamins, antioxidants and minerals in the prevention and treatment of hypertension and cardiovascular disease. *Therapeutic Advances in Cardiovascular Disease*, 4(3): 165-183.
- Hropot, M., Langer, K. H., Wiemer, G., Grottsch, H. and Linz, W. (2003). Angiotensin II subtype AT1 receptor blockade prevents hypertension and renal insufficiency induced by chronic NO-synthase inhibition in rats. *Naunyn Schmiedeberg's Archives of Pharmacology*, 367: 312-317.
- Husain, K., Scott, B. R., Reddy, S. K. and Somani, S. M. (2001). Chronic ethanol and nicotine interaction on rat tissue antioxidant defense system. *Alcohol*, 25: 89–97.
- Isman, M. B., Towers, N., Gunning, P., Spollen, K. and Ng, L.T. (1997). An environmentally safe pesticide composition. *Malaysia Patent P970207*.
- Jendekova, L., Kojsova, S., Andriantsitohaina, R. and Pechanova, O. (2006). The time-dependent effects of Provinols on brain NO synthase activity in L-NAME-induced hypertension. *Physiology Research*, 55: 31-37.
- Joker, D. (2000). *Azadirachta excelsa* (Jack) M. Jacobs. Danida Forest Seed Centre No. 13.
- Joshi, U. H., Ganatra, T. H., Desai, T. R. and Tirgar, P. R. (2012). Evaluation of antihypertensive activity of *Evolvulus alsinoides* in adrenaline induced hypertensive rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4(4): 0975-1491.
- Juliana, M. J., Suhaila, M., Intan, N. A., Noordin, M. S., Yazid, A. M. and Nordaniel, R. (2011a). Effects of catechin-rich palm leaf extract on normal and hypertensive rats' kidney and liver. *Food Chemistry*, 128:433-441.
- Juliana, M. J., Suhaila, M., Norrdaniel, R., Intan, N. A., Mustapha, M. N. and Yazid, A. M. (2011b). Antihypertensive and cardiovascular effects of catechin-rich oil palm (*Elais guineensis*) leaf extract in nitric oxide-deficient rats. *Journal of Medicinal Food*, 14(7/8): 775-783.
- Karpha, M. and Lip, G. V. (2006). The pathophysiology of target organ damage in hypertension. *Minerva Cardioangiologica*, 54: 417–429.

- Khalil, M. I. and Sulaiman, S. A. (2010). The potential role of honey and its polyphenols in preventing heart diseases: a review. *African Journal Traditional Complementary Alternative Medicine*, 7(4): 315- 321.
- Kijkar, S. (1995). Part II - Species descriptions: *Azadirachta excelsa* (Jack) Jacobs. Association of South-East Asian Nations (ASEAN), Forest Tree Seed Centre, Thailand, pp. 318-320.
- Kim, M. J., Lee, H. J., Wiryowidagdo, S. and Kim, H. K. (2006). Antihypertensive effects of *Gynura procumbens* extract in spontaneously hypertensive rats. *Journal of Medicinal Food*, 9(4): 587-590.
- Kim, H., Moon, J. Y., Kim, H., Lee, D. S., Cho, M., Choi, H. K., Kim, Y. S., Mosaddik, A. and Cho, S. K. (2010). Antioxidant and antiproliferative of mango (*Mangifera indica* L.) flesh and peel. *Food Chemistry*, 117:429-436.
- Kizhakekuttu, T. J. and Widlansky, M. E. (2010). Natural antioxidants and hypertension: promise and challenges. *Cardiovascular Therapeutics*, 28(4): 20-32.
- Klopfleisch, R. (2013). Multiparametric and semiquantitative scoring systems for the evaluation of mouse model histopathology - a systematic review. *BioMedCentral (BMC) Veterinary Research*, 9(123): 1-15.
- Knekt, P., Kumpulainen, J., Järvinen, R., Rissanen, H., Heliövaara, M., Reunanen, A., Hakulinen, T. and Aromaa, A. (2002). Flavonoid intake and risk of chronic disease. *American Journal Clinical Nutrition*, 76: 560-568.
- Kopp, U. C., Cicha, M. A. and Smith, L. A. (2003). Dietary sodium loading increases arterial pressure in afferent renal-denervated rats. *Hypertension*, 42: 968–973.
- Kotsis, V., Stabouli, S., Papakatsika, S., Rizos, Z., and Parati, G. (2010). Mechanism of obesity-induced hypertension. *Hypertension Research*, 33: 386-393.
- Koya, D., Hayashi, K., Kitada, M., Kashiwagi, A., Kikkawa, R. and Haneda, M. (2003). Effects of antioxidants in diabetes-induced oxidative stress in the glomeruli of diabetic rats. *Journal American Society of Nephrology*, 14(8): 250–253.
- Kundu, S. and Rao, J. P. (2008). The story of spontaneously hypertensive rat (SHR): A review. *Journal Medicinal Science*, 1(1): 65-66.
- Kurose, K. and Yatagai, M. (2005). Components of the essential oils of *Azadirachta indica* A. Juss, *Azadirachta siamensis* Velton, and *Azadirachta excelsa* (Jack) Jacobs and their comparison. *Journal of Wood Science*, 51: 185-188.



- Kyriakides, Z. S., Kremastinos, D. T., Bofilis, E., Tousoulis, D., Antoniadis, A. and Webb, D. J. (2000). Endogenous endothelin maintains coronary artery tone by endothelin type A receptor stimulation in patients undergoing coronary arteriography. *British Heart Journal*, 84:176–182.
- Laursen, J. B., Somers, M. and Kurz, S. (2001). Endothelial regulation of vasomotion in apoE-deficient mice: implications for interactions between peroxynitrite and tetrahydrobiopterin. *Circulation*, 103:1282-1288.
- Luo, Y., Owens, D., Mulder, G., McVey, A. and Fisher T. (2008). Blood Pressure Characterization of Hypertensive and Control Rats for Cardiovascular Studies. American Health Association, Atlanta: Charles River.
- Madamanchi, N. R., Vendrov, A. and Runge, M. S. (2005). Oxidative stress and vascular disease. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 1:29-38.
- Maisuthisakul, P., Suttajit, M. and Pongsawatmanit, R. (2007). Assessment of phenolic content and free radical scavenging capacity of some Thai indigenous plants. *Food Chemistry*, 100(4): 1409-1418.
- Maizura, M. Z., Zaiton, Z. and Nor Anita, M. M. N. (2015). The use of *Piper sarmentosum* leaves aqueous extract (Kadukmy™) as antihypertensive agent in spontaneous hypertensive rats. *BMC Complementary and Alternative Medicine*, 15(54): 1-10.
- Malkoff, J. (2005). Non- invasive blood pressure for mice and rats. 1-8
- Marathe, G. K., Zimmerman, G. A., Prescott, S. M. and McIntyre, T. M. (2002). Activation of vascular cells by PAF-like lipids in oxidized LDL. *Vascular Pharmacology*, 38:193-200.
- Maytin, M. and Colucci, W. S. (2002). Molecular and cellular mechanisms of myocardial remodeling. *Journal of Nuclear Cardiology*, 9: 319-27.
- McArdavey, D. and Robertson, J. I. S. (1990). Angiotensin Converting Enzyme Inhibitor and Moderate Hypertension. *Drugs*, 40(3):326-345.
- Meng, S., Cason, G. W., Gannon, A. W., Racusen, L. C. and Manning, R. D. (2003). Oxidative stress in Dahl salt-sensitive hypertension. *Hypertension*, 41:1346–52.
- Middleton, E., Kandaswami, C. and Theoharides, T. C. (2000). The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer. *Pharmacology Review*, 52: 673–751.
- Miguel-Carrasco, J. L., Monserrat, M. T., Mate, A. and Vazquez, C. M. (2010). Comparative effects of captopril and L-carnitine on blood pressure an

antioxidant enzyme gene expression in the heart of spontaneously hypertensive rats. *European Journal of Pharmacology*, 632: 65-72.

Milne, F. J. and Pinkney-Atkinson, V. J. (2004). Hypertension Guideline 2003 update. *Southern African Medical Journal*, 94 (3): 205-226.

Mistry, H. D. Wilson, V., Ramsay, M. M., Symonds, M. E. and Pipkin, F. B. (2008). Reduced selenium concentrations and glutathione peroxidase activity in preeclamptic pregnancies. *Hypertension*, 52: 881-888.

Miyajima, K., Minatoguchi, S. and Ito, Y. (2007). Reduction of QTc dispersion by the angiotensin II receptor blocker valsartan may be related to its anti-oxidative stress effect in patients with essential hypertension. *Hypertension Research*, 30: 307– 313.

Mohamed Saleem, T. S. , Lokanath,N., Prasanthi, A., Madhavi, M., Mallika, G. and Vishnu, M. N. (2013). Aqueous extract of *Saussurea lappa* root ameliorate oxidative myocardial injury induced by isoproterenol in rats. *Journal of Advanced Pharmaceutical Technology and Research*, 4(2): 94-100.

Mon, T.A. (2006) Omega 3 fatty acids and hypertension in humans. *Clinical Experimental Pharmacology Physiology*, 33: 842-846.

Morawietz, H., Weber, M., Rueckschloss, U., Lauer, N., Hacker, A. and Kojda, G. (2001). Biochemical and Biophysical. *Research Communications*, 285: 1130–1135.

Mordue (Luntz), A.J. and Blackwell, A. (1992). Azadirachtin: an update. *Journal of insect Physiology*, 39: 903-924.

Morgan, E. D. (2009). Azadirachtin, a scientific gold mine. *Bioorganic and Medicinal Chemistry*, 17: 4096-4105.

Mori, T. A., Burke, V., Puddey, I. and Irish, A. (2009). The effects of omega 3 fatty acids and coenzyme Q 10 on blood pressure and heart rate in chronic kidney disease: a randomized controlled trial. *Journal of Hypertension*, 27: 1863-1872.

Motshakeri, M., Ebrahimi, M., Goh, Y. M., Othman, H. H., Hair-Bejo, M. and Mohamed, S. (2014). Effects of brown seaweed (*Sargassum polycystum*) extracts on kidney, liver, and pancreas of type 2 diabetic rat model. *Evidence- Based Complementary and Alternative Medicine*, 1-11.

Moss, A. J., Zareba, W. and Hall, W. J. (2002). Prophylactic implantation of a defibrillator in patients with myocardial infarction and reduced ejection fraction. *The New England Journal of Medicine*, 346:877-83.

- Munzel, T., Daiber, A. and Ullrich, V. (2005). Vascular consequences of endothelial nitric oxide synthase uncoupling for the activity and expression of the soluble guanylyl cyclase and the cGMP-dependent protein kinase. *Arteriosclerosis Thrombosis and Vascular Biology*, 25(8):1551–1557.
- Mustafa, M. A. and Al-Khazraji, A. (2008). Effect of some plant extracts on the *Culex pipiens molestus* Forskal larvae. *Iraqi Journal of Veterinary Sciences*, 22(1): 9- 12.
- Nahida, T. and Feroz, A. (2011). Role of natural herbs in the treatment of hypertension. *Pharmacognosy Review*, 5 (9); 30-40.
- Natarajan, S. (2016). Resistant hypertension. *Journal of Indian College of Cardiology*, 30(3): 1-3.
- Nayak, D. U., Karmen, C., Frishman, W. H. and Vakili, B. A. (2001) Antioxidant vitamins and enzymatic and synthetic oxygen-derived free radical scavengers in the prevention and treatment of cardiovascular disease. *Heart Disease*, 3: 28-45.
- Ng, L. T., Yuen, P. M., Loke, W. H. and Azizol, A. K. (2003). Effects of *Azadirachta excelsa* on feeding behavior, body weight and mortality of *Crociodomia binotalis* Zeller (Lepidoptera: Pyralidae). *Journal of the Science of Food and Agriculture*, 83: 1327-1330.
- Nurdiana, S., Mohamad Shukri, K., Elizabeth Jega, J. and Nurul 'Izzati, S. (2013). Lowering blood glucose effect of *Azadirachta excelsa* leaves extract. *Natural products: An Indian Journal*, 9: 363-66.
- Nurdiana, S., Nor Haziqah, A. S., Nur Ezwa Khairunnisa, M. K., Nurul 'Izzati, S., Siti Amna, Y., Norashirene, M. J. and Nur Hilwani, I. (2014). Attenuation of pancreatic histology, hematology and biochemical parameters in type 2 diabetic rats treated with *Azadirachta excelsa*. *International Journal of Medical, Health, Pharmaceutical and Biomedical Bioengineering*, 8(9): 600-603.
- Nur syimal'ain, A. (2014). Oral subacute toxicity study of ethanolic extract of *Azadirachta excelsa* leaf in sprague dawley rats. (degree dissertation). Universiti Teknologi Mara, Malaysia.
- Nurul 'Izzati, S. (2016). Hypoglycemic, hypolipidemic and histopathological changes of selected organs in type II diabetic rats treated with *Azadirachta excelsa* leaf extract. (Master dissertation). Universiti Putra Malaysia, Malaysia.
- Nurul 'Izzati, S. , Nurdiana, S., Nur Farhana, A. S., Rajion, M. A., Goh, Y. M., Mokrish, A. and Hafandi, A. (2015). Qualitative phytochemical screening and GC-MS profiling of *Azadirachta excelsa* leaf extract. *Malaysian Applied Biology*, 44(3): 87-92.

- Nyman, U., Joshi, P., Madsen, L. B., Pedersen, T. B., Pinstrup, M., Rajasekharan, S., George, V. and Pushpangadan, P. (1998). Ethnomedical information and in vitro screening for angiotensin-converting enzyme inhibition of plants utilized as traditional medicines in Gujarat, Rajasthan and Kerala (India). *Journal of Ethnopharmacology*, 60: 247–263.
- Obiefuna, I. and Young, R. (2005). Concurrent administration of aqueous *Azadirachta indica* (Neem) leaf extract with DOCA-salt prevents the development of hypertension and accompanying electrocardiogram changes in the rat. *Phytotherapy Research*, 19 (9): 792-795.
- OECD. (2008). Guidelines for the testing of chemicals, 1-13.
- Ohkawa, H., Ohishi, N. and Yagi, K. (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical Biochemistry*, 95(2): 351-358.
- Okamoto, K. and Aoki, K. (1963). Development of a strain of spontaneously hypertensive rats. *Japanese Circulation Journal*. 27; 282-293.
- Ondetti, M. A., Williams, N. J., Sabo, E. F., Pluvec, J., Weaver, E. R. and Kocy, O. (1971). Angiotensin-converting enzyme inhibitors from the venom of *Bothrops jararaca*: isolation, elucidation of structure and synthesis. *Biochemistry*, 10: 4033-4042.
- Orwa, C., Mutua, A., Kindt, R. and Jamnadass, R. (2009). Agroforestry database: a tree reference and selection guide version 4.0. World Agroforestry Centre, Kenya.
- Pechanova, O., Matuskova, J., Capikova, D., Jendekova, L., Paulis, L., Simko, F. (2000). Effect of spironolactone and captopril on nitric oxide and S-nitrosothiol formation in kidney of L-NAME-treated rats. *Kidney International*, 70: 170-176.
- Peer, P. A., Trivedi, P. C., Nigade, P. B., Mahesh, M., Ghaisas, M. M. and Deshpande, A. D. (2008). Cardioprotective effect of *Azadirachta indica* A. Juss. on isoprenaline induced myocardial infarction in rats. *International Journal of Cardiology*, 126: 123-126.
- Perlini, S. and Grassi, G. (2013). Hypertension-related target organ damage: Is it a continuum? *Journal of Hypertension*, 31: 1083–1085.
- Pisoschi, A. M. and Negulescu, G. P. (2011). Methods for total antioxidant activity determination: A review. *Biochemistry and Analytical Biochemistry*, 1(1): 1-10.
- Power, S. K. and Jackson, M. J. (2008). Exercise-Induced Oxidative Stress: Cellular Mechanisms and Impact on Muscle Force Production. *Physiology Review*, 88: 1243–1276.

- Rad, A. (2006). Renin-angiotensin-aldosterone system. Retrieved from [https://commons.wikimedia.org/wiki/File%3ARenin-angiotensin-aldosterone\\_sy stem.png](https://commons.wikimedia.org/wiki/File%3ARenin-angiotensin-aldosterone_sy stem.png)
- Rajasekar, P., Palanisamy, N. and Anuradha, C. V. (2007). Increase in nitric oxide and reductions in blood pressure, protein kinase C beta II and oxidative stress by L-carnitine: a study in the fructose-fed hypertensive rat. *Clinical Experimental Hypertension*, 29:517–530.
- Rates, S. M. K. (2001). Plants as source of drugs. *Toxicon*, 39: 603-613.
- Reule, S. E. and Drawz, P. (2012). Heart rate and blood pressure: Any possible implications for management of hypertension? *Current Hypertension Reports*, 14(6): 478–484.
- Roberto de, B. S. (2012). Antihypertensive Drugs, (Ed.), Hypertension and renin-angiotensin system (pp. 85-94). Croatia: InTech Publishers.
- Robertson, R. P., Harmon, J., Tran, P. O., Tanaka, Y. and Takahashi, H. (2003). Glucose toxicity in  $\beta$ -cells: type 2 diabetes, good radicals gone bad, and the glutathione connection. *Diabetes*, 52: 581-587.
- Rodrigo, R., Gonzalez, J. and Paoletto, F. (2011). The role of oxidative stress in the pathophysiology of hypertension. *Hypertension Research*, 34(4): 431-40.
- Rodrigo, R., Passalacqua, W., Araya, J., Orellana, M. and Rivera, G. (2003). Implications of oxidative stress and homocysteine in the pathophysiology of essential hypertension. *Journal of Cardiovascular Pharmacology*, 42: 453–461.
- Rubattu, S., Pagliaro, B., Pierelli, G., Santolamazza, C., Di Castro, S., Mennuni, S., and Volpe, M. (2015). Pathogenesis of Target Organ Damage in Hypertension: Role of Mitochondrial Oxidative Stress. *International Journal of Molecular Sciences*, 16: 823-839.
- Sanchez, M., Galisteo, M., Vera, R., Villar, I. C., Zarzuelo, A., Tamargo, J., Perez-Vizcaino, F. and Duarte, J. (2006). Quercetin downregulates NADPH oxidase, increases eNOS activity and prevents endothelial dysfunction in spontaneously hypertensive rats. *Journal Hypertension*, 24:75-84.
- Sawyer, D. B., Siwik, D. A. Xiao, L., Pimentel, D. R., Singh, K. and Colucci, W. S. (2002). Role of oxidative stress in myocardial hypertrophy and failure. *Journal Molecular Cell Cardiology*, 34: 379-388.
- Schini-Kerth, V. B. Etienne-Selloum, N., Chataigneau, T. and Auger, C. (2011). Vascular protection by natural product- derived polyphenols: *In Vitro* and *In Vivo* evidence. *Planta Medica*, 77: 1161-1167.

- Schumutterer, H. and Doll, M. (1993). The marrango on philippine neem tree, *Azadirachta excelsa* (*A. intergrifoliola*): A new source of insecticides with growth regulating properties. *Phytoparasitica*, 21: 79-86.
- Selvan, V. T., Manikandan, L., Senthil Kumar, G. P., Suresh, R., Kakoti, B. B., Gomathi, P., Kumar, D. A., Saha, P., Gupta, M. and Mazumder, U. K. (2008). Antidiabetic and antioxidant effect of methanol extract of *Artanema sesamoides* in streptozotocin-induced diabetic rats. *International Journal of Applied Research in Natural Products*, 1(1): 25-33
- Serafini, M., Bellocco, R, Wolk, A. and Ekstrom, A. M. (2002). Total antioxidant potential of fruit and vegetables and risk of gastric cancer. *Gastroenterology*, 123: 985-999.
- Sherma, Z., Reza, F., Satoru, M., Yinrong, L., Lai, Y. F., Pooya, M. T., Judith, B. U. and Ronald, R. W. (2007). Oral administration of purple passion fruit peel extract attenuates blood pressure in female spontaneously hypertensive rats and humans. *Nutrition Research*, 27: 408-416.
- Shori, A. B. and Baba, A. S. (2013). Antioxidant activity and inhibition of key enzymes linked to type-2 diabetes and hypertension by *Azadirachta indica*-yogurt. *Journal of Saudi Chemical Society*, 17: 295-301.
- Shukor, N. A., Camp, J. V., Gonzales, G. B., Staljanssens, D., Struijs, K., Zotti, M. J., Raes, K. and Smagghe, G. (2013). Angiotensin-converting enzyme inhibitory effects by plant phenolic compounds : a study of structure activity relationships. *Journal of Agricultural and Food Chemistry*, 1-9.
- Singh, M., Mensah, G. A. and Bakris, G. (2010). Pathogenesis and clinical physiology of hypertension. *Cardiology Clinical*. 28:545-559.
- Singh, S. N., Vats, P., Suri, S., Shyam, R., Kumria, M. M. L., Ranganathan, S. and Sridharan, K. (2001). Effect of an antidiabetic extract of *Catharanthus roseus* on enzymic activities in streptozotocin induced diabetic rats. *Journal of Ethnopharmacology*, 76: 269-277.
- Sithisarn, P., Carlsen, C. U. Andersen, M. L., Gritsanapan, W., and Skibsted, L. H. (2007). Antioxidative effects of leaves from *Azadirachta* species of different provenience. *Food Chemistry*, 104: 1539-1549.
- Siti Suriani, A., Norhaizan, M. E. and Hazilawati, H. (2014). Histology Histopathologic Changes in Liver and Kidney Tissues from Male Sprague Dawley Rats Treated with *Rhaphidophora Decursiva* (Roxb.) Schott Extract. *Journal Cytology Histology*, 1-6.
- Sporkova, A., Reddy, N. R., Falck, J. R., Imig, J. D., Kopkan, L., Sadowski, J. and Cervenka, L. (2016). Interlobular arteries from two-kidney, one-clip Goldblatt hypertensive rats exhibited impaired vasodilator response to

- epoxyeicosatrienoic acids. *American Journal Medical Sciences*, 351 (5): 513-519.
- Stevens, K. R. and Gallo, M. A. (1989). Practical considerations in the conduct of chronic toxicity studies. In *Principles and Methods of Toxicology*, 2nd ed., A. W. Hayes. (pp 237-250). New York: Raven Press.
- Susalit, E., Agus, N., Effendi, I., Tjandrawinata, R. R., Nofiarny, D., Perrinjaquet-Moccetti, T., et al. (2011). Olive (*Olea europaea*) leaf extract effective in patients with stage-I hypertension: comparison with Captopril. *Phytomedicine*, 18: 251–258.
- Susanta, K. R. Dutta., S., Sengupta, M. Das, S. and Rout, B. (2010). Antihypertensive therapy: the concepts of management with herbal and synthetic agents for pulmonary hypertension. *International Journal of Pharmaceutical Sciences Review and Research*, 3(2): 72-79.
- Thomas, G.D. (2011). Neural control of the circulation. *Advance in Physiology Education*, 35: 28–32.
- Tian, N., Thrasher, K. D., Gundy, P. D., Hughson, M. D. and Manning, R. D. (2005). Antioxidant treatment prevents renal damage and dysfunction and reduces arterial pressure in salt-sensitive hypertension. *Hypertension*, 45:934–939.
- Touyz, R. M. (2004). Reactive oxygen species, vascular oxidative stress, and redox signaling in hypertension. What is the clinical significance. *Hypertension*. 44: 248-252.
- Touyz, R. M. and Schiffri, E. L. (2004). Reactive oxygen species in vascular biology: implications in hypertension. *Histochemistry Cell Biology*, 122: 339-352.
- Ueshima, H., Stamler, J., Elliot, B. and Brown, C.Q. (2007) Food omega 3 fatty acid intake of individuals (total, linolenic acid, long chain) and their blood pressure: INTERMAP study. *Hypertension*, 50: 313-319.
- Valko, M., Leibfritz, D., Moncola, J., Cronin, M. T. D., Mazura, M. and Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *International Journal Biochemistry Cell Biology*, 39: 44-84.
- Viridis, A., Neves, M. F., Amiri, F., Touyz, R. M. and Schiffrin, E. L. (2004). Role of NAD(P)H oxidase on vascular alterations in angiotensin II infused mice. *Journal Hypertension*, 22:535–542.
- Wang, J. and Xiong, X. (2012). Outcome measures of chinese herbal medicine for hypertension:an overview of systematic reviews. *Evidence Based Complementary Alternaternative Medicine*, 697237.

- WHO World Health Report, (2003). Retrieved from [http://www.who.int/whr/2003/en/whr03\\_en.pdf](http://www.who.int/whr/2003/en/whr03_en.pdf)
- World Health Organization, (WHO) (2009). Global health risks: mortality and burden of disease attributable to selected major risks. Geneva.
- World Health Organization, (WHO) (2013). Cardiovascular Diseases (CVDs). Geneva: WorldHealth Organization.FactsheetNo317
- World Health Organization, (WHO) (2016). Cardiovascular Diseases (CVDs). Retrieved from <http://www.who.int/mediacentre/factsheets/fs317/en/>.
- Wilcox, C. S. (2003). Redox regulation of the afferent arteriole and tubuloglomerular feedback. *Acta Physiologica Scandinavica*,179: 217–223.
- Wilcox, C. S. (2005). Oxidative stress and nitric oxide deficiency in the kidney: a critical link to hypertension? *American Journal of Physiology Regulation and Integrative Comparative Physiology*, 289: 913–935.
- Wilson, D. O. and Johnson, P. (2000). Exercise modulates antioxidant enzyme gene expression in rat myocardium and liver. *Journal of Applied Physiology*, 88: 1791–1796.
- Wolfe, K. W. X. and Liu, R. H. (2003). Antioxidant activity of apple peels. *Journal of Agricultural and Food Chemistry*, 51(3): 609-614.
- Wu, L., Noyan Ashraf, M. H, Facci, M., Wang, R., Paterson, P. G., Ferrie, A. and Juurlink, B. H. J. (2004). Dietary approach to attenuate oxidative stress, hypertension, and inflammation in the cardiovascular system. *PNAS*, 101(18); 7094-7099.
- Xin-Fang, L., Mohd Rais, M. and Kamsiah, J. (2013). Nigella sativa and its protective role in oxidative stress and hypertension. *Evidence-Based Complementary and Alternative Medicine*, 1: 1-9.
- Yakobson, G. S., Antonov, A. R., Golovatyuk, A. V., Markel., A. L. and Yakobson, M. G. (2001). Selenium content and blood antioxidant activity in rats with hereditary arterial hypertension during experimental myocardial infarction. *Experimental Biology and Medicine*, 132(7): 38-41.
- Yang, Q., Sun, Y-H., Zhang, L., Xu, L., Hu, M-Y., Liu, X-Y., Shi, F-Y. and Gu, Z-Y. (2014). Antihypertensive Effects of Extract from Flower Buds of *Coreopsis tinctoria* on Spontaneously Hypertensive Rats. *Chinese Herbal Medicine*, 6(2); 103-109.
- Yeh, C. T., Huang, W. H. and Yen, G. C. (2009). Antihypertensive effects of Hsian-tsao and its active compound in spontaneously hypertensive rats. *Journal of Nutritional Biochemistry*, 20: 866-875.



- Zafar, M., Naqvi, S. N. H., Ahmed, M. and Kaimkhani, Z. A. (2009). Altered liver morphology and enzymes in streptozotocin induced diabetic rats. *International Journal of Morphology*, 27(3): 719-725.
- Zalba, G., Beaumont, F. J., San Jose, G., Fortuno, A., Fortuno, M. A., Etayo, J. C. and Diez, J. (2000). *Hypertension*, 35:1055–1061.
- Zhou, Y-J., Zhang, S-P., Liu, C-W. and Cai, Y-Q. (2009). The protection of selenium on ROS mediated-apptosis by mitochondria dysfunction in cadmium-induced LLC-PK1 cells. *Toxicology in Vitro*, 23: 288-294.
- Zimmerman, M. C., Lazartigues, E. and Lang, J. A. (2002). Superoxide mediates the actions of angiotensin II in the central nervous system. *Circulation Research*, 91:1038–45.
- Zimmerman, M. C., Dunlay, R. P. and Lazartigues, E. (2004). Requirement for Rac1-dependent NADPH oxidase in the cardiovascular and dipsogenic actions of angiotensin II in the brain. *Circulation Research*, 95:532–9.