

UNIVERSITI PUTRA MALAYSIA

EXTRACTION, FRACTIONATION AND CHARACTERIZATION OF DURIAN (Durio zibethenius M.) LEAF EXTRACT AS POTENTIAL SOURCE OF SQUALENE AND ITS APPLICATION IN GELATIN-BASED FILM

KAM WAI YEE

FSTM 2019 1



EXTRACTION, FRACTIONATION AND CHARACTERIZATION OF DURIAN (Durio zibethenius M.) LEAF EXTRACT AS POTENTIAL SOURCE OF SQUALENE AND ITS APPLICATION IN GELATIN-BASED FILM

By

KAM WAI YEE

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

November 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

EXTRACTION, FRACTIONATION AND CHARACTERIZATION OF DURIAN (Durio zibethenius M.) LEAF EXTRACT AS POTENTIAL SOURCE OF SQUALENE AND ITS APPLICATION IN GELATIN-BASED FILM

By

KAM WAI YEE

November 2017

Chair : Associate Professor Seyed Hamed Mirhosseini, PhD Faculty : Food Science and Technology

The utilization of extract from durian leaf by traditional practioners has indicated the presence of phytochemical compounds with anti-inflammatory as well as antioxidant properties. However, there is no updated scientific reports on phytochemical compound recovery from durian leaf via extraction methods. The main objective of this study was to investigate the most efficient extraction method on durian leaf, proceeded by recovery of high squalene content and antioxidant activity via fractionation of the crude leaf extract. Two extraction methods that were applied in this study were ultrasound-assisted extraction (UAE) and accelerated solvent extraction (ASE). The effectiveness of both extraction methods were investigated on the yield of crude leaf extract, DPPH free radical scavenging activity, ferric reducing antioxidant power (FRAP), total phenolic content (TPC), total flavonoid content (TFC) and squalene content. The results revealed that UAE was more efficient than ASE in obtaining natural extract from durian leaf waste with optimum antioxidant activity and squalene content. In addition, UAE was reported to require shorter processing time and less energy required at lower temperature condition. Results from the optimization process revealed that the optimum condition for UAE was 5 min extraction time under continuous mode with the amplitude of 66% (power intensity 261 W/cm2), using 100% hexane at the solvent-sample ratio of 13:1. Under this optimum condition, a crude yield extract of 6.63% was obtained, and possessing antioxidant activity at 6.63 mg/mL TE per 100 mg DW in DPPH assay, 55.96 mg/mL TE per 100 mg DW in FRAP assay, 69.61 g GAE / 100 g DW in TPC and 1210.8 mg QE / 100 mg DW in TFC. Squalene content was reported at 20.56 %. The crude ^¢dæ&o4ræ{] |^Á+[{ ÁNOEÒq Á[] œã ~ { Á&[} åããã[} Á was preceded for further fractionation. The objective was to achieve fractions that were high in antioxidant activity and/or squalene content. Four fractions were obtained from this experiment and labeled as foll [] a * A Coal } and a compare the second s æ; [ĭ} α/i -Áĭĭæ;^}^ÁQ/i QHÏ G/i *Á§i ÁF€€/i *ÁÖY DŹEÓ α/i⊗]} αæ§i ^å Ác@ Á@*i @• α/ij αæ)Á] @} [| J&ÁSI } c^} cAcat[™] ^ ÁCFÌ ÈCGÁ* ÁÕ OEÒÁÐÁF€€ÉÁ* ÁÖY DĚAÔcÁSI } cast ^ å Ác@ Á@t @• cÁ ÖÚÚPÁ¦æåå&æ‡Á•&æç^} * ā * Áæ&æçãc ÁÇ È GÁ{ * ₽) ŠÁVÒÁÐÁF€€Á{ * ÁÖY DÁæ} åÁ-ÖqÁ contained highest FRAP value (108.15 mg/mL TE / 100 mg DW) and total flavonoid content (527.27 mg QE / 100 mg DW). This experiment revealed that the non-modified (both physical and chemical) alumina was efficient in the process of recovering squalene from the crude extract. n-hexane was the most efficient eluent for squalene recovery during fractionation. Fractions with high antioxidant activity were efficiently recovered by alumina and silica gel with modification using 10% AgNO3 and 10% NaCl impregnation respectively. Chloroform had efficiently recovered fraction with high antioxidant activity when using non-modified adsorbent material. The four fractions were subsequently incorporated into a gelatin-based packaging film at two usage levels (0.2% and 0.5%). All film samples were tested and compared on physicochemical properties against a negative and a positive control. The present study shows that the incorporation of fraction D from the crude leaf extract was able to enhance the radical scavenging activity in gelatin-based film. Results from the water vapor permeability test revealed that the film samples with added leaf extract fractions were not significantly (p > 0.05) improved compared to the positive control. In addition to the physicochemical tests, the potential use of the film samples were also tested on their final application use as a packaging film for minimally processed durian fruit pulp. All durian fruit pulps wrapped in the gelatin film samples were evaluated on physical changes after storage under refrigerated condition for 4 weeks. The finding of the study revealed that gelatin film with added leaf extracts were not able to control weight loss and color changes of the fruit during storage. Sensory evaluation was carried out by trained panel and the results revealed that the overall acceptance of the durian fruit after storage for three weeks under refrigerated condition were not significantly (p > 0.05) affected by the addition of leaf extract in gelatin film samples. In general, film samples with added leaf extract fractions that were high in antioxidant activity demonstrated improvement in physicochemical properties especially in DPPH radical scavenging activity. Finally, all gelatin film samples with added leaf extract did not bring prominent impact towards the physical changes in durian fruit pulp for the shelf-life extension. This study had revealed that UAE is more appropriate for obtaining a crude extract with maximum antioxidant activity and squalene content from durian leaf waste. Physicochemical properties of gelatin films were significantly improved in antioxidant activity after incorporated with leaf extract. The film samples with added leaf extract also resulted in negligible physical and sensory changes to the durian fruit pulp during storage...

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGEKSTRAKAN, PEMERINGKATAN DAN PENCIRIAN DAUN DURIAN (Durio zibethenius M.) SEBAGAI SUMBER SKUALENE SEMULAJADI DAN PENGGUNAANNYA DI DALAM FILEM GELATIN

Oleh

KAM WAI YEE

November 2017

Pengerusi : Profesor Madya Seyed Hamed Mirhosseini, PhD Fakulti : Sains dan Teknologi Makanan

Penggunaan jus daun durian pada zaman dahulu telah menunjukkan kehadiran bahan fitokimia yang berfungsi sebagai bahan anti-keradangan serta antipengoksidaan. Walau bagaimanapun, tiada laporan saintifik terkini mengenai kaedah pengekstrakan bahan fitokimia daripada daun durian. Objektif utama penyelidikan ini adalah mengkaji kaedah pengekstrakan daun durian yang paling berkesan diikuti dengan kaedah pemulihan semula yang tinggi kandungan skualene serta aktiviti pengoksidaan ekstrak daun. Dua kaedah pengekstrakan yang dikaji adalah pengekstrakan menerusi gelombang-ultra (UAE) dan pemecutan pelarut (ASE). Keberkesanan kaedah pengekstrakan ini dikaji dari segi hasilan ekstrak daun kasar, aktiviti anti-pengoksidaan dari segi pembuangan radikal bebas DPPH serta kuasa penurunan ferik (FRAP), jumlah kandungan fenolik (TPC), jumlah kandungan flavonoid (TFC) serta kandungan skualene. Hasil kajian menunjukkan bahawa UAE lebih berkesan daripada ASE dalam pemulihan ekstrak daripada daun durian yang mengandungi aktiviti antipengoksidaan serta kandungan skualene yang optimum. Di samping itu, UAE juga menggunakan masa pemprosesan yang lebih pendek dan kurang memerlukan tenaga suhu yang rendah. Hasil keputusan daripada pengoptimuman proses menunjukkan masa pengekstrakan pada keadaan operasi optimum untuk UAE adalah 5 min dengan amplitud 66% (intensiti kuasa 261 W/cm2), menggunakan 100% heksana dengan nisbah pelarut kepada sampel 13:1. Pada keadaan operasi optimum ini, sebanyak 6.63% ekstrak kasar dihasilkan dan aktiviti anti-pengoksidaan DPPH dilaporkan sebanyak 6.63 mg/mL TE / 100 mg DW, FRAP adalah 55.96 mg/mL TE / 100 mg DW, TPC dilaporkan sebanyak 69.61 g GAE / 100 g DW dan TFC dilaporkan 1210.8 mg QE / 100 mg DW. Kandungan skualene dilaporkan sebanyak 20.56 mg di dalam 100 mg ekstrak. Hasil ekstrak kasar daripada UAE kemudian dilanjutkan ke proses pemeringkatan seterusnya. Objektif proses ini adalah untuk pemulihan aktiviti anti-pengoksidaan serta / atau kandungan skualene yang lebih tinggi. Sebanyak empat bahagian telah diperolehi daripada eksperimen. Setiap

bahagian masing-masing telah dilab/\\ æ) Á^àæ ﷺ (^}*æ) å`}*æ`\`æ^\^^A c^¦ca]**ã4QîQEÏGÁÃDÊA-Óq{(^}*a)å*}*ã4)ãaaãA/ÚÔÁc^¦ca]**ã4QFÌÈOGÁ*ÁÕOEÒÁDAF€€Á*Á ÖY DÉALÔqÁ(^}*a) a`}*aÁ, afaaán ÖÜÚUPÁ k^¦ca, **a40(ÈECA(*EP)ŠÁ /ÒADAF∈€A(*AÖY DA&aa) A ±ÖqÁ{^}*æ}å`}*ãÅ}āpæãÁ2ÜŒÚÁc∿¦cāj**ãÁÇF€ÌÈFÍÁ{*È₽ ŠÁVÒÁÐÁF€€Á{*ÁDW) serta TFC tertinggi (527.27 mg QE / 100 mg DW). Eksperimen ini menunjukkan bahawa alumina yang tidak terubahsuai (secara kimia atau fizikal) adalah berkesan dalam penghasilan skualene daripada ekstrak kasar. Di samping itu, n-heksana merupakan eluen yang paling berkesan dalam menghasilkan skualene sewaktu proses pemeringkatan. Bahagian yang menunjukkan aktiviti anti-pengoksidaan tertinggi dilaporkan oleh alumina dan gel silika masingmasing diubahsuai menggunakan 10% AgNO3 and 10% NaCl. Kloroform dilaporkan berkesan dalam penghasilan aktiviti anti-pengoksidaan tertinggi apabila diguna bersama ajen penjerap yang tidak diubahsuai. Empat bahagian ekstrak yang diperolehi daripada proses pemeringkatan kemudian ditambahkan ke filem pembungkusan berasaskan gelatin pada dua tahap (0.2% dan 0.5%). Semua sampel filem telah diuji dan dibandingkan dengan sampel kawalan positif dan negatif masing-masing dari segi ciri-ciri fizikokimia. Hasil kajiselidik ini {^}`}b`\\æ}Á àæ@æç æÁ]^}**`}ææ)Á àæ@æt ãæ)Á ^\•dæbÁ ±ÖqÁ à^\`]æîæA meningkatkan tahap aktiviti anti-pengoksidaan DPPH dalam filem berasaskan gelatin. Walau bagaimanapun, penggunaan ekstrak daun durian tidak menujukkan kesan yang signifikan (p > 0.05) dalam filem gelatin dari segi ketelapan wap air apabila dibanding dengan sampel kawalan positif. Selain daripada ujikaji fizikokimia, penggunaan filem gelatin sebagai pembungkus durian juga telah dikaji. Semua isi durian yang dibungkus dengan filem gelatin disimpan dalam peti-sejuk selama 4 minggu dan perubahaan fizikal durian dikaji. Hasil keputusan menunjukkan bahawa filem gelatin yang ditambah ekstrak daun tidak berupaya mengawal isi durian dari segi kehilangan berat serta perubahan warna sepanjang masa penyimpanan. Ujian sensori telah dijalankan dengan menggunakan panel terlatih. Hasil kajian menunjukan penerimaan keseluruhan isi durian tidak dipengaruhi secara signifikan (p > 0.05) oleh penambahan ekstrak dalam filem gelatin. Secara keseluruhannya, penggunaan ekstrak daun yang tinggi aktiviti anti-pengoksidaan telah meningkatkan keupayaan filem gelatin terutamanya dari segi aktiviti pembebasan radikal DPPH. Walau bagaimanapun, tiada hasil keputusan yang signifikan dari segi pemanjangan tempoh hayat penyimpanan isi durian. Kesimpulan daripada kaji-selidik ini ialah UAE diikuti dengan proses pemeringkatan merupakan kaedah pengekstrakan yng lebih sesuai untuk hasilan ekstrak yang tinggi kandungan skualene serta mengandungi aktivit anti-pengoksidaan yang tinggi daripada daun durian yang gugur. Ciri-ciri fizikokimia filem gelatin dipertingkatkan dengan signifikan dari segi aktiviti anti-pengoksidaan apabila ditambah ekstrak daun. Sampel filem gelatin ditambah dengan ekstrak daun menyebabkan perubahan fizikal dan deria yang minimum terhadap isi buah durian semasa penyimpanan.

ACKNOWLEDGEMENTS

I would like to express sincere gratitude to my supervisor, Associate Professor Dr. Seyed Hamed Mirhosseiniand for his valuable time & energy in providing endless support throughout my study. His help and guidance have always been motivating and inspiring which encouraged me to complete the study on time with the best of my capability.

I would like to extend my appreciation to my co-supervisors Prof. Madya Dr. Faridah binti Abas and Dr. Norhayati bt. Hussain as well as all laboratory staffs. They had generously made themselves available and responded whenever I needed their help and support during the period of my study.

Finally, I would like to express my deepest appreciation to my family members especially my beloved mother, Tham Yoke Ying and my late father, Kam Ng Hoong, who had and still have been very supportive to all that I have been doing until today.

Last but not least, my sincere appreciation to my dearest friends Florence and Simin who never failed in offering helps whenever I needed during my study. It has been such great pleasure to have known and worked with them in the same laboratory. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Seyed Hamed Mirhosseini, PhD

Associate Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Chairman)

Faridah Abas, PhD

Associate Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

Norhaytai Hussain, PhD

Associate Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No.:	

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xvi
LIST OF FIGURES	xix
LIST OF ABBREVIATIONS	xxii

CHAPTER

1	INTE	RODUC	TION		1
2	LITE 2.1 2.2 2.3 2.4 2.4 2.5	ERATUF Durian Phytoo Plant a 2.3.1 2.3.2 Squale Phytoo	RE REVIEN (Durio zik chemical c antioxidant Phenolic Terpenol ene as a n chemical c	N pethinus M.) ompounds in durian leaf s compounds id compounds atural antioxidant ompounds extraction from	3 3 4 5 6 7 9
		plant 2.5.1	Ultrasou	nd-assisted extraction	10
			(UAE) 2.5.1.1	Factors affecting bioactive compounds extraction from plant materials using	11
		2.5.2	Accelera	ted Solvent Extraction	13
			2.5.2.1	Factors affecting bioactive compounds extraction from plant materials using	13
	2.6	Fractic extrac	onation and t	d purification of crude	15
		2.6.1	Column extract fr	Chromatography in crude actionation	16
	2.7	Comm 2.7.1	ion packag Developi cut duria	ging for fresh cut fruits ment of packaging for fresh- n	17 18
	2.8	Biodeg 2.8.1 2.8.2	gradable F Gelatin-t Gelatin-t with add	ilm based Biodegradable Film based Biodegradable Film ed plant extract	19 20 21

		2.8.3	End-use a active film	application of antioxidant is	22
3	MAT 3.1	ERIALS Materia	S AND ME	THODS / METHODOLOGY	24 24
		3.1.1 3.1.2	Chemical Raw Mate	s, Standards and Reagents erial	24
	3.2	Experir 3.2.1	nental Pro Prelimina	cedures ry Study	24 25
			3.2.1.1	Sample Preparation of Durian Leaf Wastes	25
			3.2.1.2	Solvent Selection for Durian Leaf Extraction	26
			3.2.1.3	Identification of Major Compounds in Durian	27
				Leaf Extracts Using Gas Chromatography-Mass Spectrometry (GC-MS)	
		3.2.2	Optimizat Durian Le Assisted	tion of Leaf Extraction from af Waste using Ultrasound- Extraction (UAE)	27
			3.2.2.1	Experimental Design and Data Analysis	27
			3.2.2.2	Ultrasound-Assisted Extraction (UAE)	29
			3.2.2.3	UAE Optimization and Validation	29
			3.2.2.4	Determination of Crude Extract Yield	30
			3.2.2.5	Determination of Antioxidant Activity	30
				3.2.2.5.1 DPPH Assay 3.2.2.5.2 FRAP Assay	30 31
			3.2.2.6	Determination of Total Phenolic Content (TPC)	31
			3.2.2.7	Determination of Total Flavonoid Content (TFC)	32
			3.2.2.8	Squalene Content Analysis	32
			3.2.2.9	Cytotoxicity Test on crude leaf extract from optimized UAE condition	33
			3.2.2.10	Non-volatile compound identification in crude leaf extract from optimized UAE condition via liquid chromatography . mass spectrometry (LC-MS)	33
		3.2.3	Optimizat Durian Le	tion of Leaf Extraction from eaf Wastes using	34

	Accelerat	ed Solvent Extraction	
	(ASE)		
	3.2.3.1	Experimental Design and Data Analysis	34
	3.2.3.2	Accelerated Solvent	35
	3.2.3.3	ASE Optimization and	36
	3.2.3.4	Validation Determination of Crude	37
	3.2.3.5	Extract Yield Determination of	37
		Antioxidant Activity 3.2.3.5.1 DPPH Assav	37
		32352 FRAP Assav	37
	3.2.3.6	Determination of Total	37
	3.2.3. <mark>7</mark>	Determination of Total	37
	3.2.3.8	Squalene Content	37
	3.2.3.9	Cytotoxicity Test on crude	37
		optimized ASE condition	
	3.2.3.10	Non-volatile compound identification in crude leaf	38
		extract from optimized ASE condition via liquid	
		chromatography . mass spectrometry (LC-MS)	
3.2.4	Column C Leaf Extr	Chromatography for Crude action Fractionation	38
	3.2.4.1	Experimental Design and Data Analysis	38
	3.2.4.2	General procedure for	40
		fractionation	40
	3.2.4.3	Antioxidant Activity	40
		3.2.4.3.1 DPPH Assay	40
		3.2.4.3.2 FRAP Assay	41
	3.2.4.4	Determination of Total Phenolic Content (TPC)	41
	3.2.4.5	Determination of Total Flavonoid Content (TFC)	41
	3.2.4.6	Squalene Content Analysis	41
3.2.5	Preparati	on of gelatin film with	41

				gelatin film	with added	
			3.2.5.2	Preparation	n of film sample	43
				aliquot		
			3.2.5.3	Determinat Vapor Perr	tion of Water neability of	43
				gelatin film	-	
			3.2.5.4	Physical qu	uality	44
				evaluation	on durian fruit	
				film	ed in gelaun	
				3.2.5.4.1	Color	44
				32542	Weight loss	44
				3.2.5.4.3	Texture	45
				000	profile	
					analysis	
				3.2.5.4.4	Sensory evaluation	45
			3.2.5.5	Statistical /	Analysis	46
4	RES			USSION		47
-	4.1	Prelir	ninary Study			47
		4.1.1	Solvent Se	election for Du	urian Leaf	47
			Extraction			
		4.1.2	Identificatio	on of Major C	ompounds in	49
	12	Ontim	Crude Du	trasound Ass		54
	7.2	Extrac	ction for Sau	alene and Ar	ntioxidant-	54
		Enrich	ned Extract f	rom Durian L	eaf (Durio	
		zibeth	ninus M.)			
		4.2.1	Effect of U/	AE Variables	on Extraction	54
		422	Yield Effort of LL		on Antiovident	57
		4.2.2	Activity			57
		4.2.3	Effect of UA	AE variables	on Lotal	61
			Flavonoid C	Content (TPC)		
		4.2.4	Effect of U/	AE variables	, on squalene	63
		4.2.5	UAE Optim	ization and v	alidation	64
			process			•
		4.2.6	Cytotoxicity from optim	Test on cruc	de leaf extract n of UAE	65
		4.2.7	Non-volatile	e compound i	identification in	66
			crude leaf condition v	extract from o	optimized UAE matography.	
			mass spec	trometry (LC	-MS)	
	4.3	Optin Extra	nization of A action for Sa	ccelerated Solution	olvent Intioxidant-	67
			•			

6

	Enric		
	4.3.1	67	
	122	Yield Effect of LIAE Veriables on Antioxident	70
	4.3.2	Activity	70
		4.3.2.1 DPPH assay	70
		4.3.2.2 FRAP assay	72
	4.3.3	Effect of ASE variables on Total	73
		Phenolic Content (TPC)	
	4.3.4	Effect of ASE variables on Total	74
	125	Filavonoid Content (TFC)	75
	4.3.5	content	75
	436	Ontimization and validation of ASE	76
	4.0.0	condition	
	4.3.7	Cytotoxicity Test on crude leaf extract	78
		from optimized condition of ASE	
	4.3.8	Non-volatile compound identification in	78
		crude leaf extract from optimized ASE	
		condition via liquid chromatography.	
		mass spectrometry (LC-MS)	
	4.3.9	Result comparison of crude leaf	81
		extract using optimized condition of	
11	Facto	one and ASE	82
4.4	extra	ct using column chromatography	02
	4.4.1	Effects of fractionation variables on	84
		antioxidant activity	
	4.4.2	Effects of fractionation variables on	87
		total phenolic content (TPC)	
	4.4.3	Effects of fractionation variables on	89
		total flavonoid content (TFC)	
	4.4.4	Effects of fractionation variables on	89
	445	Solution of fractions for application in	03
	4.4.5	gelatin-based biodegradable film	93
45	Physi	cochemical properties of gelatin film	93
1.0	4.5.1	Antioxidant Activity via DPPH assav	94
		and FRAP assay	
	4.5.2	Water vapor permeability of gelatin-	96
		based film with added leaf extract	
	4.5.3	Changes in physical quality of durian	97
		fruit pulp after storage using gelatin-	
		based film with added leaf extract	07
		4.5.3.1 Weight IOSS	97 100
		4.5.3.2 Color properties	100
		4.5.3.4 Sensory attributes	112
		· · · · · · · · · · · · · · · ·	

 \bigcirc

5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	119
REFERE	INCES	121
APPEND	DICES	138
BIODAT	A OF STUDENT	144
LIST OF	PUBLICATIONS	145



 \bigcirc

LIST OF TABLES

Table		Page
2.1	Nutritional composition of durian aril (per 100a)	3
2.2	Antioxidant capacity and bioactive compounds detected in durian aril at different ripening stages	4
2.3	Classes of Terpenoids	7
2.4	Extraction techniques that were applied for squalene extraction from plant sources	10
2.5	Applications of UAE in the extraction of antioxidant compounds from plant materials	12
2.6	Applications of ASE in the extraction of antioxidant compounds from plant materials	15
2.7	Classical and modern chromatographic methods	15
2.8	Studies carried out to date on the manufacture of gelatin-based biodegradable / edible films and coatings with enhanced antioxidant capacity	21
2.9	Application of gelatin-based packaging film on food product	23
3.1	Matrix design of ultrasound-assisted extraction treatments	28
3.2	Matrix of accelerated solvent extraction treatments	35
3.3	Input variables and experimental design matrix for the fractionation of crude leaf extract from durian leaf waste via UAE	39
3.4	Label description of gelatin-based film samples	43
3.5	Sensory attributes, definition and endpoint labels used for the evaluation of the durian fruit samples	45
4.1	Secondary metabolites detected using GC. MS, with relative retention time (RT) and molecular weight (MW) in the mass spectra of the crude extract from durian leaf waste	50
4.2	The p-value and F-ratio of ultrasound-assisted extraction (UAE) variables in the final reduced models fitted for crude extract from durian leaf wastes	55
4.3	R ² and p-value for Regression of the final reduced model fitted for crude extract from durian leaf wastes via UAE	56

4.4	Optical density value and percentage of cell viability in various concentrations of durian leaf extract (LAE) exposed to the 3T3 cells	65
4.5	Tentative non-volatile compound of crude leaf extract from optimized UAE condition	66
4.6	The p-value and F-ratio of accelerated solvent extraction (ASE) variables in the final reduced models fitted for crude extract from durian leaf	69
4.7	R ² and p-value for Regression of the final reduced model fitted for crude extract from durian leaf wastes via ASE	70
4.8	Optical density value and percentage of cell viability in various concentrations of durian leaf extract (ASE) exposed to the 3T3 cells	78
4.9	Tentative non-volatile compound of crude leaf extract from optimized ASE condition	79
4.10	Comparison between crude extract from durian leaf waste using UAE and ASE at optimized condition	81
4.11	The p-value and F-ratio of fractionation variables in the final reduced models fitted for crude extract from durian leaf wastes	83
4.12	Fractions selected from 36 experiments with highest value for each response	93
4.13	p-value and F-ratio of weight loss in durian fruit] ĭ]] Á ãœa Á Á ^^\ • oA € d	98
4.14	p-value and F-ratio of weight loss in durian fruit pulp wrapped in different film samples	98
4.15	Mean value of weight loss (%) in durian fruit pulp during storage over 4 weeks in gelatin- based film with added	99
4.16	p-value <mark>and F-ratio</mark> of color properties in durian ⊣്ã⁄h,́`]] Á ão@a Á Á ^^\ • ɗ ɗ ¦æt ^	102
4.17	p-value and F-ratio of color properties in durian fruit pulp wrapped in different film samples	103
4.18	Mean value of L* value (lightness) in durian fruit pulp during storage over 4 weeks in gelatin-based film with added leaf extract	103
4.19	Mean value of a* value (redness) in durian fruit pulp during storage over 4 weeks in gelatin- based film with added leaf extract	105
4.20	Mean value of b* value (yellowness) in durian fruit pulp during storage over 4 weeks in gelatin-based film with added leaf extract	106
4.21	p-value and F-ratio on texture parameters in å ĭ ¦ææ) Á¦ĭ ãoÁĭ [] Á ão@ã Á Á ^^\•oÁtd ¦æ≛^	108
4.22	p-value and F-ratio of texture parameters for durian in gelatin-àæ^åÁą̃{ Á ãœ̃, ÁFÁ ^^\•q́ storage	109

- 4.23 Mean value of hardness (mg) in durian fruit pulp during storage over 4 weeks in gelatinbased film with added leaf extract
- 4.24 Mean value of resilience in durian fruit pulp during storage over 4 weeks in gelatin-based film with added leaf extract
- 4.25 p-value and F-ratio of sensory parameters for durian fruit pulp in gelatin-based film within 1 ,^^\ • m (¦ ☆ ^
- 4.26 p-value and F-ratio on sensory parameters in å ĭ ¦ãæ) Á¦ĭ ãŕA ĭ [] Á ãr@ ÁrÍÁ ^^\ • qÁt d ¦æ≛^
- 4.27 Mean score of the Quantitative Descriptive Analysis (QDA) attributes of durian fruit pulp in gelatin-based film with added leaf extract after storage of 1 and 3 weeks respectively

111

110

112

114

115

LIST OF FIGURES

Figure		Page
2.1	Durian leaves in combination colours of	4
22	green and olive green Strive (And All 2011) (And And And And And And And And And And	7
2.2	Phenolic diternenes	8
2.0	$\dot{U}d^{*} \& \dot{A} \dot{A} \dot{A} [] [\& [] A \\ A \dot{A}] $	8
2.5	Chemical structure of Squalene	9
2.6	Adsorption and hydrogen bonding between compound and sorbent face	16
2.7	Examples of packaging used for fresh-cut	18
2.8	Example of packaging for fresh-cut durian in local market	18
2.9	Edible and non-edible type biopolymer for the potential use in biodegradable packaging	19
	films	
3.1	General flow of methodology	25
3.2	Flow diagram of durian leaf sample preparation	26
3.3	A complete setup of ultrasound-assisted extraction (a), a sonotrode (probe) with a tip diameter of 13mm (b) and a processor unit	29
3.4	Thermo Scientific Dionex ASE 150	36
35	Film Forming Solution (EES) preparation step	42
0.0 4 1	Gas chromatogram of crude leaf extract from	48
7.1	durian leaf waste using ethanol (a) hexane	40
	(b) via maceration and secondary metabolites	
	detected using GC/SIM, MS conditions, with	
	relative retention time (rt)	
4.2	Gas chromatogram of crude leaf extract from	49
	durian leaf waste using hexane via	
	ultrasound assisted extraction and secondary	
	metabolites detected using GC/SIM. MS	
	conditions, with relative retention time (rt)	
4.3	Interaction effects of UAE variables between	54
	amplitude and solvent mixture ratio (a) pulsed	
	time (b) on the yield	
4.4	Interaction effects of UAE variables between	58
	extraction time and amplitude on the DPPH	
	scavening activity (a) FRAP assay (b)	
4.5	Interaction effects of UAE variables between	59
	amplitude and pulsed time (a) solvent mixture	
4.0	ratio (b) on the DPPH scavenging activity	~~
4.0	extraction time and pulsed time (a) solvent :	60

 \bigcirc

	sample ratio (b) on the DPPH scavenging	
4.7	Interaction effects of UAE variables between solvent : sample ratio and solvent mixture	61
4.8	Interaction effects of UAE variables between solvent mixture ratio and amplitude on Total Phenolic Content (a) : Total Flavonoid	62
4.9	Content (b) Interaction effects of UAE variables between solvent mixture ratio and extraction time on	63
4.10	Numerical multiple optimization plots demonstrating the optimum UAE of crude	65
4.11	LC-MS Chromatogram of non-volatile compounds in crude leaf extracts from	66
4.12	Interaction effects of ASE variables between solvent mixture ratio and temperature on the	68
4.13	Interaction effects of ASE variables between temperature and solvent : sample ratio (a) solvent mixture ratio (b) on DPPH scavenging	71
4.14	Interaction effects of ASE variables between solvent : sample ratio and temperature on the	73
4.15	Interaction effects of ASE variables between temperature and solvent : ratio (a) solvent mixture ratio (b) on TEC	75
4.16	Interaction effects of ASE variables between extraction time and temperature on squalene content	76
4.17	Numerical multiple optimization plots demonstrating the optimum ASE of crude extract from durian leaf waste	77
4.18	LC-MS Chromatogram of non-volatile compounds in crude leaf extracts from optimized ASE condition	78
4.19	Main effect plot of fractionation variables on antioxidant activity via DPPH assay (a) FRAP assay (b)	85
4.20	Interaction effects of fractionation variables between eluent and chemical treatment on antioxidant activity via DPPH assay (a) FRAP	86
4.21	Interaction effects of fractionation variables between absorbent type and chemical treatment on antioxidant activity via DPPH assay (a) FRAP assay (b)	87

C

4.22	Main effect plots of chemical treatment (a) eluent (b) on total phenolic content	88
4.23	Interaction effects plot of fractionation variables between eluent and chemical treatment on total phenolic content	88
4.24	Main effect plots of chemical treatment (a) eluent (b) on TFC	89
4.25	Interaction effects of fractionation variables between adsorbent and eluent on squalene content	90
4.26	Interaction effects of fractionation variables between chemical treatment and eluent on squalene content	91
4.27	Interaction effects of fractionation variables between heat treatment and eluent on squalene content	92
4.28	Interaction effects of fractionation variables between adsorbent and chemical treatment on squalene content	92
4.29	Antioxidant activity (Free radical scavenging activity) of gelatin-based film with added leaf extract	94
4.30	Antioxidant activity (Reducing power) of gelatin-based film with added leaf extract	95
4.31	Water vapor permeability of gelatin-based film with added leaf extract	96
4.32	Weight loss (%) in durian fruit after 4 weeks	100

 (\mathbf{G})

LIST OF ABBREVIATIONS

Redness / greeness		
Analysis of variance		
Silver nitrate		
Accelerated solvent extraction		
Yellowness / blueness		
Central composite design		
2,2-diphenyl-1-picrylhydrazyl		
dried weight		
Ferric reducing antioxidant power		
gram		
Gallic acid equivalent		
Gas chromatography . mass spectrometer		
Gas chromatography . flame ion detector		
Hydrochloric Acid		
kilogram		
Lightness		
Sodium chloride		
Milligram		
Milligram per mililiter		
minute		
mililiter		
milimeter		
milimolar		
milimole		
Correlation coefficient		
Response surface methodology		
second		
Squalene		
Total flavonoid content		
Total phenolic content		
Ultrasound-assisted extraction		
Volume per volume		
Volume per weight		
Watt / square centimeter		
Water vapor permeability		

xxii

CHAPTER 1

INTRODUCTION

Natural antioxidant has now become one of the major subjects proceeding with growing demand for natural food ingredients in the food industry. The safety concern about using synthetic antioxidants (such as butylatedhydroxytoluene and butylatedhydroxyanisole) has motivated the food industry to seek natural alternatives (Brewer, 2011). Plant extract is generally a good source of natural antioxidant. The plant-based extracts contain the active compounds (such as flavonoids, phenolic acids, carotenoids, and tocopherols) with a potential antioxidant activity. They can inhibit Fe3+/AA induced oxidation, scavenge free radicals, and act as reductants (Khanduja & Bhardwaj, 2003; Ozsov, Candoken, & Akey, 2009). Previous researchers reported different natural antioxidants in various plant leaves (Fu et al., 2016; Nowak, Czyzowska, Efenberger, & Krala, 2016; Romero-García et al., 2016; Tahir et al., 2015). However, the quality, antioxidant activity and composition of the plant-based extract depend on the extraction type and condition. Squalene was reported to be possessing antioxidant effect (Conforti et al., 2005). Plant-based squalene has been sourced extensively with the intention to replace animal origined squalene, which is commonly available from shark liver oil. During the preliminary investigation of this study, squalene was identified as a major compound in durian leaf waste when extracted using n-hexane.

Various extraction methods have been developed for the optimum recovery of extraction yield from plants. In this case where squalene extraction was of concerned, the commonly reported extraction method was supercritical fluid extraction (Kraujalis & Venskutonis, 2013; Suleiman, Baharin, Mirhosseini, Sarker, & Islam, 2012). To date, ultrasound-assisted extraction (UAE) and accelerated solvent extraction (ASE) are the modern extraction methods reported to be efficient for the recovery of phytochemical compounds via optimization (Kang, Kim, & Moon, 2016; Kazemi, Karim, Mirhosseini, & Hamid, 2016). Accelerated solvent extraction was specifically efficient for the extraction of non-polar compounds even when polar solvents along with high pressure are applied (Zaghdoudi *et al.*, 2015). This has become the advantage of this technique in extracting natural compounds of various polarities from the same plant materials in comparison to other modern techniques. However, limited report available on the recovery of squalene from plant materials using these two extraction methods.

Durian (*Durio zibethinus* M.) leaf is known used by traditional practitioners for therapeutic purposes. Tate (1999) reported a few findings on pharmacological properties and therapeutic effects of durian leaves. It was believed that decoction from durian leaves and root possessing anti-inflammatory effect, which helped to relief patient who was suffering from high fever. Phytochemical properties in durian have been reviewed by Ho and Bhat (2015), but mainly emphasizing a $\frac{1}{3}$ and $\frac{1}{3}$ and

Hence, this study has been carried out to report on optimal extraction condition for crude extract from durian leaf waste using different modern solvent extraction methods, which are ultrasound-assisted extraction (UAE) and accelerated solvent extraction (ASE). The main goal and novelty of this study was to report the most efficient extraction technique on durian leaf, which ultimately recovered the highest antioxidant activity as well as squalene content from the leaf extract. At the final part of the study, the leaf extract was applied into gelatin-based film. This was to identify the potential use of the leaf extract as a functional ingredient in the gelatin-based film. Film samples with added leaf extracts were evaluated on antioxidant properties as well as functional use in protecting durian fruit pulp from physical changes during storage (4 weeks). Physcial changes in durian fruit pulp that were investigated including water loss, color and texture. Therefore the specific objectives of this research were as follow:

- 1. To optimize the ultrasound-assisted extraction (UAE) for durian leaf extract with the highest yield, antioxidant activity and squalene content.
- 2. To optimize the accelerated solvent extraction (ASE) for durian leaf extract with the highest yield, antioxidant activity and squalene content.
- 3. To investigate the effect of purification/fractionation condition on the yield and antioxidant activity of durian leaf extract
- 4. To examine the potential use of durian leaf extract as a functional ingredient in a gelatin-based film for controlling physical changes in durian fruit pulp during storage period.

REFERENCES

- Afsar, T., Khan, M. R., Razak, S., Ullah, S., & Mirza, B. (2015). Antipyretic, antiinflammatory and analgesic activity of Acacia hydaspica R. Parker and its phytochemical analysis. *BMC Complementary and Alternative Medicine*, *15*(1), 136.
- Ahmad, M., Benjakul, S., Prodpran, T., & Agustini, T. W. (2012). Physicomechanical and antimicrobial properties of gelatin film from the skin of unicorn leatherjacket incorporated with essential oils. *Food Hydrocolloids*, *28*(1), 189. 199.
- Ahmad, M., Benjakul, S., Sumpavapol, P., & Nirmal, N. P. (2012). Quality changes of sea bass slices wrapped with gelatin film incorporated with lemongrass essential oil. *International Journal of Food Microbiology*, *155*(3), 171. 178.
- Akgün, N. A. (2011). Separation of squalene from olive oil deodorizer distillate using supercritical fluids. *European Journal of Lipid Science and Technology*, *113*(12), 1558. 1565.
- CF ¢åæctÅr Å? HÅQæàæàæææævÅv HÅQF@eq æåÅr Å? Å? HÅQF-Mahasneh, M. A., Almajwal, A., Õæq { [@ÅUHÅõ ÁQF|åÅÆÅQAGEFÏ]. A review of phenolic compounds in oilbearing plants: Distribution, identification and occurrence of phenolic compounds. Food Chemistry, 218, 99. 106.
- Amornputti, S., Ketsa, S., & van Doorn, W. G. (2014). Effect of 1methylcyclopropene (1-MCP) on storage life of durian fruit. *Postharvest Biology and Technology*, 97, 111. 114.
- Anbinder, P. S., Peruzzo, P. J., Martino, M. N., & Amalvy, J. I. (2015). Effect of antioxidant active films on the oxidation of soybean oil monitored by Fourier transform infrared spectroscopy. *Journal of Food Engineering*, 151, 43. 50.
- Arancibia-Avila, P., Toledo, F., Park, Y.-S., Jung, S.-T., Kang, S.-G., Heo, B. Õ⊞Ãõ ÁÕ[¦ậ•ởậ ÂÙĚAÇ3€€Ì DĂŒ; đặ ¢ãàæ) óÅ] ¦[]^¦æ?•Á[-Áåč¦ãæ) Á-¦čãtéæ Á influenced by ripening. *LWT-Food Science and Technology*, *41*(10), 2118. 2125.
- ASTM. (1989). Standard test methods for water vapor transmission of materials. Annual Book of ASTM Standards. Designation E96-E80, 730. 739.
- Atarés, L., & Chiralt, A. (2016). Essential oils as additives in biodegradable films and coatings for active food packaging. *Trends in Food Science & Technology*, 48, 51. 62.
- CE àæ c^A ¦É\$ ÉÉQ^A ÉÓ ÉÁ æ@; ÉÚ ÉÉA Å^O { åÉÔ ÉQEF HDÁU] œí ã ææi } Á Á |dæ[} & assisted extraction of antioxidant compounds from blackberry leaves using response surface methodology. *Industrial Crops and Products*, 44, 558.

565.

- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, ØĚĂõ ÁU{ æÊAŒĂSEĂT ĚÁŒ€FHDĂV^&@ã`^•Á-{ \Á^¢dæ&æ] \ Á[-Áàā] æ&æj.^Á compounds from plant materials: a review. *Journal of Food Engineering*, 117(4), 426. 436.
- Bai-Ngew, S., Therdthai, N., Dhamvithee, P., & Zhou, W. (2014). A study of the effect of the drying process on the composition and physicochemical properties of flours obtained from durian fruits of two ripening stages. *International Journal of Food Science & Technology*, 49(1), 230. 237.
- Baldwin, E. A., Hagenmaier, R., & Bai, J. (2011). *Edible coatings and films to improve food quality*. CRC Press.
- Barbosa-Pereira, L., Cruz, J. M., Sendón, R., de Quirós, A. R. B., Ares, A., Castro-Š5] A: ÉAT ÉA õ Á Úæ Aã[-Losada, P. (2013). Development of antioxidant active films containing tocopherols to extend the shelf life of fish. *Food Control*, 31(1), 236. 243.
- Battisti, R., Fronza, N., Júnior, Á. V., da Silveira, S. M., Damas, M. S. P., & Quadri, M. G. N. (2017). Gelatin-coated paper with antimicrobial and antioxidant effect for beef packaging. *Food Packaging and Shelf Life*, *11*, 115. 124.
- Bellumori, M., Innocenti, M., Binello, A., Boffa, L., Mulinacci, N., & Cravotto, G. (2016). Selective recovery of rosmarinic and carnosic acids from rosemary leaves under ultrasound-and microwave-assisted extraction procedures. *Comptes Rendus Chimie*, *19*(6), 699. 706.
- Benzie, I. F. F., & Strain, J. J. (1999). [2] Ferric reducing/antioxidant power assay: Direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methods in Enzymology*, 299, 15. 27.
- Besbes, S., Drira, L., Blecker, C., Deroanne, C., & Attia, H. (2009). Adding value to hard date (Phoenix dactylifera L.): Compositional, functional and sensory characteristics of date jam. *Food Chemistry*, *112*(2), 406. 411.
- Bonilla, J., & Sobral, P. J. A. (2016). Investigation of the physicochemical, antimicrobial and antioxidant properties of gelatin-chitosan edible film mixed with plant ethanolic extracts. *Food Bioscience*, *16*, 17. 25.
- Booncherm, P., & Siriphanich, J. (1991). Postharvest physiology of durian pulp and husk. *Kasetsart J*, *25*, 119. 125.
- Brannegan, D., Lee, C., Wang, J., & Taylor, L. (2011). Extraction Techniques Leveraging Elevated Temperature and Pressure. In *Sample Preparation of Pharmaceutical Dosage Forms* (pp. 93. 128). Springer.

- Brewer, M. S. (2011). Natural antioxidants: sources, compounds, mechanisms of action, and potential applications. *Comprehensive Reviews in Food Science and Food Safety*, *10*(4), 221. 247.
- Carrizo, D., Taborda, G., Nerín, C., & Bosetti, O. (2016). Extension of shelf life of two fatty foods using a new antioxidant multilayer packaging containing green tea extract. *Innovative Food Science & Emerging Technologies*, *33*, 534. 541.
- Cazzonelli, C. I. (2011). Carotenoids in nature: insights from plants and beyond. *Functional Plant Biology*, *38*(11), 833. 847.
- Chang, C.-C., Yang, M.-H., Wen, H.-M., & Chern, J.-C. (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *Journal of Food and Drug Analysis*, *10*(3).
- Chang, C. J., Chang, Y.-F., Lee, H., Lin, J., & Yang, P.-W. (2000). Supercritical carbon dioxide extraction of high-value substances from soybean oil deodorizer distillate. *Industrial & Engineering Chemistry Research*, *39*(12), 4521. 4525.
- Cheah, E. L. C., Chan, L. W., & Heng, P. W. S. (2006). Supercritical carbon dioxide and its application in the extraction of active principles from plant materials. *Asian Journal of Pharmaceutical Sciences*, *1*, 59. 71.
- Chemat, F., Rombaut, N., Sicaire, A. G., Meullemiestre, A., Fabiano-Tixier, A. S., & Abert-Vian, M. (2017). Ultrasound assisted extraction of food and natural products. Mechanisms, techniques, combinations, protocols and applications. A review. Ultrasonics Sonochemistry, 34, 540. 560. https://doi.org/10.1016/j.ultsonch.2016.06.035
- Chemat, S., Aissa, A., Boumechhour, A., Arous, O., & Ait-Amar, H. (2017). Extraction mechanism of ultrasound assisted extraction and its effect on higher yielding and purity of artemisinin crystals from Artemisia annua L. leaves. *Ultrasonics Sonochemistry*, 34, 310.316. https://doi.org/10.1016/j.ultsonch.2016.05.046
- CHEN, X., ZHAO, X., & WANG, W. (2012). Separation and Purification of Solanesol from Tobacco Using Silver Ion Complexation. *Journal of Hubei University of Science and Technology (Medical Sciences)*, *5*, 1.
- Choi, H.-S., Song, H. S., Ukeda, H., & Sawamura, M. (2000). Radicalscavenging activities of citrus essential oils and their components: detection using 1, 1-diphenyl-2-picrylhydrazyl. *Journal of Agricultural and Food Chemistry*, *48*(9), 4156. 4161.
- Conforti, F., Statti, G., Loizzo, M. R., Sacchetti, G., Poli, F., & Menichini, F. (分€€Í DĂQ 検護[Á\$) @ 〈義 ~^ & 〈\$) å 為 @ ゐ ãã (} / Á · Á - amylase of two varieties of Amaranthus caudatus seeds. *Biological and Pharmaceutical Bulletin*, *28*(6), 1098. 1102.

- Corrales, M., García, A. F., Butz, P., & Tauscher, B. (2009). Extraction of anthocyanins from grape skins assisted by high hydrostatic pressure. *Journal of Food Engineering*, *90*(4), 415. 421.
- Cuq, B., Gontard, N., & Guilbert, S. (1998). Proteins as agricultural polymers for packaging production. *Cereal Chemistry*, 75(1), 1. 9.
- Davalos, A., Miguel, M., Bartolome, B., & Lopez-Fandino, R. (2004). Antioxidant activity of peptides derived from egg white proteins by enzymatic hydrolysis. *Journal of Food Protection*, *67*(9), 1939. 1944.
- de Dicastillo, C. L., Rodríguez, F., Guarda, A., & Galotto, M. J. (2016). Antioxidant films based on cross-linked methyl cellulose and native Chilean berry for food packaging applications. *Carbohydrate Polymers*, *136*, 1052. 1060.
- De Vries, B. (1964). Separation of triglycerides by column chromatography on silica impregnated with silver nitrate. *Journal of the American Oil Chemists Society*, *41*(6), 403. 406.
- Deng, G.-F., Shen, C., Xu, X.-R., Kuang, R.-D., Guo, Y.-J., Zeng, L.-ÙÉÁÓ ÁvãæÁ E.-Q. (2012). Potential of fruit wastes as natural resources of bioactive compounds. *International Journal of Molecular Sciences*, *13*(7), 8308. 8323.
- Dessi, M. A., Deiana, M., Day, B. W., Rosa, A., Banni, S., & Corongiu, F. P. (2002). Oxidative stability of polyunsaturated fatty acids: effect of squalene. *European Journal of Lipid Science and Technology*, 104(8), 506. 512.
- Dicko, M. H., Gruppen, H., Traoré, A. S., Voragen, A. G. J., & van Berkel, W. J. H. (2006). Phenolic compounds and related enzymes as determinants of sorghum for food use. *Biotechnology and Molecular Biology Review*, 1(1), 20. 37.
- Dumont, M.-J., & Narine, S. S. (2007). Characterization of flax and soybean soapstocks, and soybean deodorizer distillate by GC-FID. *Journal of the American Oil Chemists' Society*, *84*(12), 1101. 1105.
- Falguera, V., Quintero, J. P., Jiménez, A., Muñoz, J. A., & Ibarz, A. (2011). Edible films and coatings: structures, active functions and trends in their use. *Trends in Food Science & Technology*, 22(6), 292. 303.
- Feuereisen, M. M., Barraza, M. G., Zimmermann, B. F., Schieber, A., & Schulze-Kaysers, N. (2017). Pressurized liquid extraction of anthocyanins and biflavonoids from Schinus terebinthifolius Raddi: A multivariate optimization. *Food Chemistry*, 214, 564. 571.
- Flannigan, D. J., & Suslick, K. S. (2010). Inertially confined plasma in an imploding bubble. *Nature Physics*, *6*(8), 598. 601.

- Fonseca, S. C., Oliveira, F. A. R., & Brecht, J. K. (2002). Modelling respiration rate of fresh fruits and vegetables for modified atmosphere packages: a review. *Journal of Food Engineering*, 52(2), 99. 119.
- Foti, M. C., & Ingold, K. U. (2003). Mechanism of inhibition of lipid peroxidation à Â-terpinene, an unusual and potentially useful hydrocarbon antioxidant. *Journal of Agricultural and Food Chemistry*, *51*(9), 2758. 2765.
- FreshPlaza. (2017). Thai fresh-cut durian arrives on the Chinese market (webpage article). Retrieved May 2, 2017, from http://www.freshplaza.com/article/174254/Thai-fresh-cut-durian-arrives-on-the-Chinese-market
- Fu, C., Tian, H., Li, Q., Cai, T., & Du, W. (2006). Ultrasound-assisted extraction of xyloglucan from apple pomace. *Ultrasonics Sonochemistry*, *13*(6), 511. 516.
- Fu, Z., Tu, Z., Zhang, L., Wang, H., Wen, Q., & Huang, T. (2016). Antioxidant activities and polyphenols of sweet potato (Ipomoea batatas L.) leaves extracted with solvents of various polarities. *Food Bioscience*, 15, 11. 18.
- Ghebreyessus, K. Y., Schiltz, H., & Angelici, R. J. (2006). Partial separation of polyunsaturated fatty acid esters from FAMEs mixtures by adsorption on silver nitrate-impregnated silica gel. *Journal of the American Oil Chemists' Society*, 83(7), 645. 652. https://doi.org/10.1007/s11746-006-1252-5
- Gibbons, S. (2012). An Introduction to Planar Chromatography and Its Application to Natural Products Isolation. *Natural Products Isolation*, 117. 153.
- Gironi, F., & Piemonte, V. (2011). Temperature and solvent effects on polyphenol extraction process from chestnut tree wood. *Chemical Engineering Research and Design*, *89*(7), 857. 862.
- Gomes, S. V. F., Portugal, L. A., dos Anjos, J. P., de Jesus, O. N., de Oliveira, E. J., David, J. P., & David, J. M. (2017). Accelerated solvent extraction of phenolic compounds exploiting a Box-Behnken design and quantification of five flavonoids by HPLC-DAD in Passiflora species. *Microchemical Journal*.
- Gómez-Guillén, M. C., Giménez, B., López-Caballero, M. E. al, & Montero, M. P. (2011). Functional and bioactive properties of collagen and gelatin from alternative sources: A review. *Food Hydrocolloids*, 25(8), 1813. 1827.
- Gontard, N., Duchez, C., CUQ, J., & Guilbert, S. (1994). Edible composite films of wheat gluten and lipids: water vapour permeability and other physical properties. *International Journal of Food Science & Technology*, *29*(1), 39. 50.

Gorinstein, S., Poovarodom, S., Leontowicz, H., Leontowicz, M., Namiesnik, J.,

X^{*}æðæð ð ÉÚÉÉő Á/æ @ æÉZÉÇGEFFDÉOE dð ¢ãaæ) of [] ^{*} æð• Áæ) å Ábð ð æsæð Á constituents of some rare exotic Thai fruits and comparison with conventional fruits: in vitro and in vivo studies. *Food Research International*, 44(7), 2222. 2232.

- Grassmann, J. (2005). Terpenoids as plant antioxidants. *Vitamins & Hormones*, 72, 505. 535.
- Grigoriadou, D., Androulaki, A., Psomiadou, E., & Tsimidou, M. Z. (2007). Solid phase extraction in the analysis of squalene and tocopherols in olive oil. *Food Chemistry*, *105*(2), 675. 680.
- Halvorsen, B. L., & Blomhoff, R. (2011). Validation of a quantitative assay for the total content of lipophilic and hydrophilic antioxidants in foods. *Food Chemistry*, 127(2), 761. 768.
- Hanani, Z. A. N., Roos, Y. H., & Kerry, J. P. (2012). Use of beef, pork and fish gelatin sources in the manufacture of films and assessment of their composition and mechanical properties. *Food Hydrocolloids*, *29*(1), 144. 151.
- Hanani, Z. A. N., Roos, Y. H., & Kerry, J. P. (2014). Use and application of gelatin as potential biodegradable packaging materials for food products. *International Journal of Biological Macromolecules*, *71*, 94. 102.
- Hemwimol, S., Pavasant, P., & Shotipruk, A. (2006). Ultrasound-assisted extraction of anthraquinones from roots of Morinda citrifolia. *Ultrasonics Sonochemistry*, *13*(6), 543. 548.
- Hernández-Ledesma, B., Dávalos, A., Bartolomé, B., & Amigo, L. (2005). Ú!^] ææað \ Á. Áæ) að çã çãæ) að : `{ ææð Á@ å![|`•æ* • Á![{ Á -lactalbumin and -lactoglobulin. Identification of active peptides by HPLC-MS/MS. Journal of Agricultural and Food Chemistry, 53(3), 588. 593.
- Herrero, M., Castro-Puyana, M., Mendiola, J. A., & Ibañez, E. (2013). Compressed fluids for the extraction of bioactive compounds. *TrAC Trends in Analytical Chemistry*, *43*, 67. 83.
- Ho, L. H., & Bhat, R. (2015). Exploring the potential nutraceutical values of durian (Durio zibethinus L.) An exotic tropical fruit. *Food Chemistry*, *168*, 80. 89. https://doi.org/10.1016/j.foodchem.2014.07.020
- Huang, Z.-R., Lin, Y.-K., & Fang, J.-Y. (2009). Biological and pharmacological activities of squalene and related compounds: potential uses in cosmetic dermatology. *Molecules*, *14*(1), 540. 554.
- Ibrahim, H., Sim, K. S., Syamsir, D. R., Nor, N. R. M., Nurestri, A. M. S., & Awang, K. (2010). Cytotoxic activity of leaf and rhizome extracts of Alpinia scabra (Blume) Nves, a wild ginger from Peninsular Malaysia. *African Journal of Pharmacy and Pharmacology*, *4*(10), 708. 711.

- Isabelle, M., Lee, B. L., Lim, M. T., Koh, W.-P., Huang, D., & Ong, C. N. (2010). Antioxidant activity and profiles of common fruits in Singapore. *Food Chemistry*, 123(1), 77. 84.
- ISO 8586-2. (1994). Sensory Analysis: General Guidance for the Selection, Training and Monitoring of Assessors. ISO.
- Izadiyan, P., & Hemmateenejad, B. (2016). Multi-response optimization of factors affecting ultrasonic assisted extraction from Iranian basil using central composite design. *Food Chemistry*, 190, 864. 870.
- James, J. B., Ngarmsak, T., & Rolle, R. S. (2011). Processing of fresh-cut tropical fruits and vegetables: a Technical Guide. FAO Agricultural Service Bulletin, 1. 86.
- Jaswir, I., MAN, C. H. E., YAAKOB, B., SELAMAT, J., AHMAD, F., & SUGISAWA, H. (2008). Retention of volatile components of durian fruit leather during processing and storage. *Journal of Food Processing and Preservation*, *32*(5), 740. 750.
- Jongjareonrak, A., Benjakul, S., Visessanguan, W., & Tanaka, M. (2008). Antioxidative activity and properties of fish skin gelatin films incorporated ac@OPV/As) aA -tocopherol. *Food Hydrocolloids*, 22(3), 449. 458.
- Juan, C., González, L., Soriano, J. M., Moltó, J. C., & Mañes, J. (2005). Accelerated solvent extraction of ochratoxin A from rice samples. *Journal of Agricultural and Food Chemistry*, 53(24), 9348. 9351.
- Kader, A. A., & Saltveit, M. E. (2003). Respiration and gas exchange. Postharvest Physiology and Pathology of Vegetables, 2, 7. 29.
- Kamalian, N., Mirhosseini, H., Mustafa, S., & Manap, M. Y. A. (2014). Effect of alginate and chitosan on viability and release behavior of Bifidobacterium pseudocatenulatum G4 in simulated gastrointestinal fluid. *Carbohydrate Polymers*, *111*, 700. 706.
- Kang, J.-H., Kim, S., & Moon, B. (2016). Optimization by response surface methodology of lutein recovery from paprika leaves using accelerated solvent extraction. *Food Chemistry*, *205*, 140. 145.
- Kazemi, M., Karim, R., Mirhosseini, H., & Hamid, A. A. (2016). Optimization of pulsed ultrasound-assisted technique for extraction of phenolics from pomegranate peel of Malas variety: Punicalagin and hydroxybenzoic acids. *Food Chemistry*, 206, 156. 166.
- Kelly, G. S. (1999). Squalene and its potential clinical uses. *Alternative Medicine Review: A Journal of Clinical Therapeutic, 4*(1), 29. 36.
- Khamtache-Abderrahim, S., Lequart-Pillon, M., Gontier, E., Gaillard, I., Pilard, ÙĦ æ@[] ĐĐĂ Ấ ﷺ æBenabdesselam, F. (2016). Isoquinoline alkaloid

fractions of Fumaria officinalis: Characterization and evaluation of their antioxidant and antibacterial activities. *Industrial Crops and Products*, *94*, 1001. 1008.

- Khanduja, K. L., & Bhardwaj, A. (2003). Stable free radical scavenging and antiperoxidative properties of resveratrol compared in vitro with some other bioflavonoids.
- Khemakhem, I., Ahmad-Qasem, M. H., Catalán, E. B., Micol, V., García-Pérez, J. V., Ayadi, M. A., & Bouaziz, M. (2017). Kinetic improvement of olive [^æç^• qaã æšaç^ Á&[{] [` } å• Á^¢dæšaā] } Aà ´A` •ā] * Á] [, ^r ultrasound in a wide temperature range. Ultrasonics Sonochemistry, 34, 466. 473.
- Kim, S.-K., Kim, Y.-T., Byun, H.-G., Nam, K.-S., Joo, D.-S., & Shahidi, F. (2001). Isolation and characterization of antioxidative peptides from gelatin hydrolysate of Alaska pollack skin. *Journal of Agricultural and Food Chemistry*, 49(4), 1984. 1989.
- Kim, S.-K., Kim, Y.-T., Byun, H.-G., Park, P.-J., & Ito, H. (2001). Purification and characterization of antioxidative peptides from bovine skin. *BMB Reports*, *34*(3), 219. 224.
- Kimbaris, A. C., Siatis, N. G., Daferera, D. J., Tarantilis, P. A., Pappas, C. S., & Polissiou, M. G. (2006). Comparison of distillation and ultrasound-assisted extraction methods for the isolation of sensitive aroma compounds from garlic (Allium sativum). *Ultrasonics Sonochemistry*, *13*(1), 54. 60.
- Kowalczyk, D., & Biendl, M. (2016). Physicochemical and antioxidant properties of biopolymer/candelilla wax emulsion films containing hop extract. A comparative study. *Food Hydrocolloids*, *60*, 384. 392.
- Slæibæjå) ÉXEÁÚukalskas, A., Kraujalis, P., & Venskutonis, P. R. (2016). Biorefining of Bergenia crassifolia L. roots and leaves by high pressure extraction methods and evaluation of antioxidant properties and main phytochemicals in extracts and plant material. *Industrial Crops and Products*, *89*, 390. 398.
- Kraujalis, P., & Venskutonis, P. R. (2013). Supercritical carbon dioxide extraction of squalene and tocopherols from amaranth and assessment of extracts antioxidant activity. *The Journal of Supercritical Fluids*, *80*, 78. 85.
- Słæi bajár ÉŹÚÉÁX^} •\` (‡) ár ÉŹÚÉÁÜÉÁÚČ \ ap+\ær ÉŹOÉÉÁBÁSæ ^¦} aga a c ÉŹÜÉÁÇJEFHDÉÁ Accelerated solvent extraction of lipids from Amaranthus spp. seeds and characterization of their composition. LWT-Food Science and Technology, 54(2), 528. 534.
- Kruk, J., Holländer-Czytko, H., Oettmeier, W., & Trebst, A. (2005). Tocopherol as singlet oxygen scavenger in photosystem II. *Journal of Plant Physiology*, *16*2(7), 749. 757.

- Lee, J. H., Ku, C. H., Baek, N., Kim, S.-H., Park, H. W., & Kim, D. K. (2004). Phytochemical constituents from Diodia teres. *Archives of Pharmacal Research*, *27*(1), 40. 43.
- Li, H., Chen, B., & Yao, S. (2005). Application of ultrasonic technique for extracting chlorogenic acid from Eucommia ulmodies Oliv.(E. ulmodies). *Ultrasonics Sonochemistry*, 12(4), 295. 300.
- Li, H., Pordesimo, L., & Weiss, J. (2004). High intensity ultrasound-assisted extraction of oil from soybeans. *Food Research International*, *37*(7), 731. 738.
- Li, H., Zhang, D., Tan, L.-H., Yu, B., Zhao, S.-P., & Cao, W.-G. (2017). Comparison of the antioxidant properties of various solvent extracts from Dipsacus asperoides and identification of phenolic compounds by LC-ESI-QTOF-MS. MS. South African Journal of Botany, 109, 1.8.
- Li, J.-H., Miao, J., Wu, J.-L., Chen, S.-F., & Zhang, Q.-Q. (2014). Preparation and characterization of active gelatin-based films incorporated with natural antioxidants. *Food Hydrocolloids*, *37*, 166. 173.
- Li, Q.-M., Luo, J.-G., Wang, X.-B., Yang, M.-H., & Kong, L.-Y. (2013). Sesquiterpenes from the rhizomes of Alpinia japonica and their inhibitory effects on nitric oxide production. *Fitoterapia*, *86*, 29. 34.
- López, D., Márquez, A., Gutiérrez-Cutiño, M., Venegas-Yazigi, D., Bustos, R., & Matiacevich, S. (2017). Edible film with antioxidant capacity based on salmon gelatin and boldine. *LWT-Food Science and Technology*, 77, 160. 169.
- Marcuse, R. (1960). Antioxidative effect of amino-acids. *Nature*, *186*(4728), 886. 887.
- Mendis, E., Rajapakse, N., & Kim, S.-K. (2005). Antioxidant properties of a radical-scavenging peptide purified from enzymatically prepared fish skin gelatin hydrolysate. *Journal of Agricultural and Food Chemistry*, *53*(3), 581. 587.
- Mirhosseini, H., & Amid, B. T. (2012). Influence of chemical extraction conditions on the physicochemical and functional properties of polysaccharide gum from durian (Durio zibethinus) seed. *Molecules*, *17*(6), 6465. 6480.
- Mirhosseini, H., & Tan, C. P. (2009). Response surface methodology and multivariate analysis of equilibrium headspace concentration of orange beverage emulsion as function of emulsion composition and structure. *Food Chemistry*, *115*(1), 324. 333.
- Mirhosseini, H., Tan, C. P., Yusof, S., & Hamid, N. S. A. (2008). Solid-phase microextraction for determining twelve orange flavour compounds in a model beverage emulsion. *Phytochemical Analysis*, 19(5), 429. 437.

- Mohebi, E., & Shahbazi, Y. (2017). Application of chitosan and gelatin based active packaging films for peeled shrimp preservation: A novel functional wrapping design. *LWT-Food Science and Technology*, *76*, 108. 116.
- Moreno, E., Reza, J., & Trejo, A. (2007). Extraction of polycyclic aromatic hydrocarbons from soil using water under subcritical conditions. *Polycyclic Aromatic Compounds*, *27*(4), 239. 260.
- Moudache, M., Nerín, C., Colon, M., & Zaidi, F. (2017). Antioxidant effect of an innovative active plastic film containing olive leaves extract on fresh pork meat and its evaluation by Raman spectroscopy. *Food Chemistry*, *229*, 98. 103.
- Munne-Ó[•&@ÂÙÈQC€€Í DÀV@Á'[|^Á[-Á -tocopherol in plant stress tolerance. Journal of Plant Physiology, 162(7), 743. 748.
- Mussatto, S. I., Ballesteros, L. F., Martins, S., & Teixeira, J. A. (2011). Extraction of antioxidant phenolic compounds from spent coffee grounds. *Separation and Purification Technology*, *83*, 173. 179.
- Mustafa, A., & Turner, C. (2011). Pressurized liquid extraction as a green approach in food and herbal plants extraction: A review. *Analytica Chimica Acta*, *703*(1), 8. 18.
- Mutsokoti, L., Panozzo, A., Tongonya, J., Kebede, B. T., Van Loey, A., & Hendrickx, M. (2017). Carotenoid stability and lipid oxidation during storage of low-fat carrot and tomato based systems. *LWT*-Food Science and Technology, 80, 470. 478.
- Nerín, C., Tovar, L., Djenane, D., Camo, J., Salafranca, J., Beltrán, J. A., & Roncalés, P. (2006). Stabilization of beef meat by a new active packaging containing natural antioxidants. *Journal of Agricultural and Food Chemistry*, *54*(20), 7840. 7846.
- Nerin, C., Tovar, L., & Salafranca, J. (2008). Behaviour of a new antioxidant active film versus oxidizable model compounds. *Journal of Food Engineering*, *84*(2), 313. 320.
- Nimse, S. B., & Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *Rsc Advances*, *5*(35), 27986. 28006.
- Niponsak, A., Laohakunjit, N., & Kerdchoechuen, O. (2015). Contribution to Volatile Fingerprinting and Physico-chemical Qualities of Minimally Úl[&^••^åÁÖč ¦ 論 Á&; 密[] c@] * +Öč ¦ 導 * ÁÙ﴿ ¦ 證 ^ K@A } cãa&æậ } Á , Á∞f>[ç^ | Á Chemical Ripeness Marker. *Food and Bioprocess Technology*, 8(6), 1229. 1243.
- Noronha, C. M., de Carvalho, S. M., Lino, R. C., & Barreto, P. L. M. (2014). Ô@طæ&c^¦ãæāā, Á[Áæ] هُوْ مُعْمَى اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّ tocopherol nanocapsules. *Food Chemistry*, *159*, 529. 535.

- Nowak, A., Czyzowska, A., Efenberger, M., & Krala, L. (2016). Polyphenolic extracts of cherry (Prunus cerasus L.) and blackcurrant (Ribes nigrum L.) leaves as natural preservatives in meat products. *Food Microbiology*, *59*, 142. 149.
- Ojagh, S. M., Rezaei, M., Razavi, S. H., & Hosseini, S. M. H. (2010). Development and evaluation of a novel biodegradable film made from chitosan and cinnamon essential oil with low affinity toward water. *Food Chemistry*, *122*(1), 161. 166.
- Ozsoy, N., Candoken, E., & Akev, N. (2009). Implications for degenerative disorders: Antioxidacăç^Áæcăçăc Éd (catÁ) @} [|• Édaç[] [ã • Éde & [à ã Áæcã É -&æd [c^} ^ Áæ) å Á -tocopherol in Aloe vera. Oxidative Medicine and Cellular Longevity, 2(2), 99. 106.
- Palma, M., Piñeiro, Z., & Barroso, C. G. (2002). In-line pressurized-fluid extraction. solid-phase extraction for determining phenolic compounds in grapes. *Journal of Chromatography A*, 968(1), 1. 6.
- Pan, Z., Qu, W., Ma, H., Atungulu, G. G., & McHugh, T. H. (2011). Continuous and pulsed ultrasound-assisted extractions of pomegranate peel. *Ultrasonics Sonochemistry*, 18(5), 1249. 1257.
- Úæç [[çã ÊÁÖÈÁT ÈÁÓæàã ÊÁÙÈÁP [¦çæ ÉÁOÈÁRĚÁT ÈÁBÁSæzc^]æ] Macan, M. (2007). Sample preparation in analysis of pharmaceuticals. *TrAC Trends in Analytical Chemistry*, 26(11), 1062. 1075.
- Pereda, M., Ponce, A. G., Marcovich, N. E., Ruseckaite, R. A., & Martucci, J. F. (2011). Chitosan-gelatin composites and bi-layer films with potential antimicrobial activity. *Food Hydrocolloids*, 25(5), 1372. 1381.
- Perry, R. H., & Green, D. W. (1999). *Perry's chemical engineers' handbook*. McGraw-Hill Professional.
- Pihlström, T., Isaac, G., Waldebäck, M., Österdahl, B.-G., & Markides, K. E. (2002). Pressurised fluid extraction (PFE) as an alternative general method for the determination of pesticide residues in rape seed. *Analyst*, *127*(4), 554. 559.
- Piñeiro, Z., Palma, M., & Barroso, C. G. (2004). Determination of catechins by means of extraction with pressurized liquids. *Journal of Chromatography A*, 1026(1), 19. 23.
- Pinelo, M., Del Fabbro, P., Manzocco, L., Nuñez, M. J., & Nicoli, M. C. (2005). Optimization of continuous phenol extraction from Vitis vinifera byproducts. *Food Chemistry*, 92(1), 109. 117.
- Praditdoung, S. (1986). Cold storage of durian (English abstract). *Kasetsart J*, 20, 44. 49.

- Priego-Capote, F., & Mari´a Del Pilar, D. D. L. T. (2013). Chapter 5 Accelerated Liquid Extraction. In M. A. Rostagno & J. M. Prado (Eds.), *Natural product extraction: principles and applications* (pp. 157. 195). Royal Society of Chemistry.
- Psomiadou, E., & Tsimidou, M. (1999). On the role of squalene in olive oil stability. *Journal of Agricultural and Food Chemistry*, *47*(10), 4025. 4032.
- Rezaei, S., Rezaei, K., Haghighi, M., & Labbafi, M. (2013). Solvent and solvent to sample ratio as main parameters in the microwave-assisted extraction of polyphenolic compounds from apple pomace. *Food Science and Biotechnology*, 22(5), 1. 6.
- Ribeiro-Santos, R., Andrade, M., de Melo, N. R., & Sanches-Silva, A. (2017). Use of essential oils in active food packaging: recent advances and future trends. *Trends in Food Science & Technology*.
- Richter, B. E., Jones, B. A., Ezzell, J. L., Porter, N. L., Avdalovic, N., & Pohl, C. (1996). Accelerated solvent extraction: a technique for sample preparation. *Analytical Chemistry*, *68*(6), 1033. 1039.
- Rodov, V., Ben-Yehoshua, S., Aharoni, N., & Cohen, S. (2010). 5 Modified Humidity Packaging of Fresh Produce. *Horticultural Reviews*, *37*, 281.
- Romero-García, J. M., Lama-Muñoz, A., Rodríguez-Gutiérrez, G., Moya, M., Ruiz, E., Fernández-Bolaños, J., & Castro, E. (2016). Obtaining sugars and natural antioxidants from olive leaves by steam-explosion. *Food Chemistry*, *210*, 457. 465.
- Rostagno, M. A., Palma, M., & Barroso, C. G. (2003). Ultrasound-assisted extraction of soy isoflavones. *Journal of Chromatography A*, *1012*(2), 119. 128.
- Saitoh, T., Matsushima, S., & Hiraide, M. (2005). Concentration of polyaromatic hydrocarbons in water to sodium dodecyl sulfate- -alumina admicelle. *Journal of Chromatography A*, *1069*(2), 271. 274.
- Saleh, I. A., Vinatoru, M., Mason, T. J., Abdel-Azim, N. S., Aboutabl, E. A., & Hammouda, F. M. (2016). A possible general mechanism for ultrasoundassisted extraction (UAE) suggested from the results of UAE of chlorogenic acid from Cynara scolymus L.(artichoke) leaves. *Ultrasonics Sonochemistry*, *31*, 330. 336.
- Salvo, A., La Torre, G. L., Di Stefano, V., Capocchiano, V., Mangano, V., Saija, OHAO ÁO * [AO AQ EFI DA a dWUSÔĐUOCA d^c' { a action of squalene in Sicilian PDO pistachio from Bronte: Optimization of oil extraction method and analytical characterization. *Food Chemistry*, 221, 1631. 1636.
- Sánchez-González, L., González-Martínez, C., Chiralt, A., & Cháfer, M. (2010). Physical and antimicrobial properties of chitosan. tea tree essential oil

composite films. Journal of Food Engineering, 98(4), 443. 452.

- Sarker, S. D., Latif, Z., & Gray, A. I. (2005). *Natural products isolation* (Vol. 20). Springer Science & Business Media.
- Seppanen, C. M., Song, Q., & Csallany, A. S. (2010). The antioxidant functions of tocopherol and tocotrienol homologues in oils, fats, and food systems. *Journal of the American Oil Chemists' Society*, *87*(5), 469. 481.
- Sharmila, G., Nikitha, V. S., Ilaiyarasi, S., Dhivya, K., Rajasekar, V., Kumar, N. T HÁO ÁT č@\`{ aba} HÓEQEFÎ DÁWdæ[`}åÁæ•ã chåÁ¢da&cā]} Á[Á[caþÁ phenolics from Cassia auriculata leaves and evaluation of its antioxidant activities. Industrial Crops and Products, 84, 13. 21.
- Shirsath, S. R., Sonawane, S. H., & Gogate, P. R. (2012). Intensification of extraction of natural products using ultrasonic irradiations. a review of current status. *Chemical Engineering and Processing: Process Intensification*, 53, 10. 23.
- Sivakumar, V., Vijaeeswarri, J., & Anna, J. L. (2011). Effective natural dye extraction from different plant materials using ultrasound. *Industrial Crops and Products*, 33(1), 116. 122.
- Smuleac, V., Butterfield, D. A., Sikdar, S. K., Varma, R. S., & Bhattacharyya, D. (2005). Polythiol-functionalized alumina membranes for mercury capture. *Journal of Membrane Science*, 251(1), 169. 178.
- Soliman, E. M., Saleh, M. B., & Ahmed, S. A. (2006). Alumina modified by dimethyl sulfoxide as a new selective solid phase extractor for separation and preconcentration of inorganic mercury (II). *Talanta*, *69*(1), 55. 60.
- Soradech, S., Nunthanid, J., Limmatvapirat, S., & Luangtana-anan, M. (2017). Utilization of shellac and gelatin composite film for coating to extend the shelf life of banana. *Food Control*, *73*, 1310. 1317.
- Stankovic, M. S. (2011). Total phenolic content, flavonoid concentration and antioxidant activity of Marrubium peregrinum L. extracts. *Kragujevac J Sci*, *33*(2011), 63. 72.
- Stone, H., Sidel, J., Oliver, S., Woolsey, A., & Singleton, R. C. (2008). Sensory evaluation by quantitative descriptive analysis. *Descriptive Sensory Analysis in Practice*, 23. 34.
- Suleiman, N., Baharin, B. S., Mirhosseini, H., Sarker, M., & Islam, Z. (2012). Squalene recovery from palm fatty acid distillate using supercritical fluid extraction. *International Food Research Journal*, *19*(4), 1661. 1667.
- Væ@IÉSIEÆPæ IEÂUIEÆSIÆÓIEÆS @) ÉQUEÁVIEÆS @) ÉZ EVVEÆP EZŐ[}* ÉÚEŸ EÆS ÁŒ@ æ ÉA A. (2015). Nerium oleander leaves extract mediated synthesis of gold nanoparticles and its antioxidant activity. *Materials Letters*, 156, 198. 201.

- Talapatra, S. K., & Talapatra, B. (2015). Natural Products Chemistry: A General Treatment. In *Chemistry of Plant Natural Products* (pp. 243. 316). Springer.
- Tao, Y., & Sun, D.-W. (2015). Enhancement of food processes by ultrasound: a review. Critical Reviews in Food Science and Nutrition, 55(4), 570. 594.
- Tate, D. (1999). *Tropical fruit*. Editions Didier Millet.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., & Byrne, D. H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(6), 669. 675.
- Tiwari, B. K. (2015). Ultrasound: A clean, green extraction technology. *TrAC Trends in Analytical Chemistry*, *71*, 100. 109.
- Tongnuanchan, P., Benjakul, S., & Prodpran, T. (2012). Properties and antioxidant activity of fish skin gelatin film incorporated with citrus essential oils. *Food Chemistry*, 134(3), 1571. 1579.
- Tongnuanchan, P., Benjakul, S., & Prodpran, T. (2013). Physico-chemical properties, morphology and antioxidant activity of film from fish skin gelatin incorporated with root essential oils. *Journal of Food Engineering*, *117*(3), 350. 360.
- Toubane, A., Rezzoug, S. A., Besombes, C., & Daoud, K. (2017). Optimization of Accelerated Solvent Extraction of Carthamus Caeruleus L. Evaluation of antioxidant and anti-inflammatory activity of extracts. *Industrial Crops and Products*, *97*, 620. 631.
- Trinh, K. T., & Glasgow, S. (2012). On the texture profile analysis test. *Chemeca* 2012: Quality of Life through Chemical Engineering: 23-26 September 2012, Wellington, New Zealand, 749.
- Tsimidou, M. (2010). Squalene and tocopherols in olive oil: importance and methods of analysis. *Olives and Olive Oil in Health and Disease Prevention*, 561. 567.
- Tuberoso, C. I. G., Kowalczyk, A., Sarritzu, E., & Cabras, P. (2007). Determination of antioxidant compounds and antioxidant activity in commercial oilseeds for food use. *Food Chemistry*, *103*(4), 1494. 1501.
- Uddin, M. N., Afrin, R., Uddin, M. J., Uddin, M. J., Alam, A., Rahman, A. A., & Sadik, G. (2015). Vanda roxburghii chloroform extract as a potential source of polyphenols with antioxidant and cholinesterase inhibitory activities: identification of a strong phenolic antioxidant. *BMC Complementary and Alternative Medicine*, *15*(1), 195.
- Udem, S. C., Anyanwu, M. U., Obidike, R. I., & Udem, N. D. (2011). The effects of Psidium guajava Linn.(Myrtaceae) leaf chloroform extract on some

hematological and biochemical parameters in mice. *Comparative Clinical Pathology*, *20*(1), 47. 51.

- Vangani, V., Joseph, R., Devi, S., & Rakshit, A. K. (1991). Thermodynamics of adsorption of polybutadiene on alumina and silica gel: effect of temperature and solvent. *Colloid & Polymer Science*, 269(3), 242. 247.
- Vázquez, L., Torres, C. F., Fornari, T., Señoráns, F. J., & Reglero, G. (2007). Recovery of squalene from vegetable oil sources using countercurrent supercritical carbon dioxide extraction. *The Journal of Supercritical Fluids*, *40*(1), 59. 66.
- Vinatoru, M. (2001). An overview of the ultrasonically assisted extraction of bioactive principles from herbs. *Ultrasonics Sonochemistry*, *8*(3), 303. 313.
- Voon, Y. Y., Hamid, N. S. A., Rusul, G., Osman, A., & Quek, S. Y. (2006). Physicochemical, microbial and sensory changes of minimally processed durian (Durio zibethinus cv. D24) during storage at 4 and 28 C. *Postharvest Biology and Technology*, 42(2), 168. 175.
- Voon, Y. Y., Hamid, N. S. A., Rusul, G., Osman, A., & Quek, S. Y. (2007). Volatile flavour compounds and sensory properties of minimally processed durian (Durio zibethinus cv. D24) fruit during storage at 4 C. Postharvest Biology and Technology, 46(1), 76. 85.
- Waldebäck, M., Señoráns, F. J., Fridström, A., & Markides, K. E. (2006). Pressurized fluid extraction of squalene from olive biomass. In C. Turner (Ed.), *Modern Extraction Techniques for food and agricultural samples* (pp. 96. 106). ACS Publications. https://doi.org/10.1021/bk-2006-0926.ch007
- Wang, L., Auty, M. A. E., Rau, A., Kerry, J. F., & Kerry, J. P. (2009). Effect of pH and addition of corn oil on the properties of gelatin-based biopolymer films. *Journal of Food Engineering*, 90(1), 11. 19.
- Wang, L., & Li, X. (2011). Antioxidant Activity of Durian (Durio zibethinus Murr.) Shell In vitro. Asian Journal of Pharmaceutical & Biological Research (AJPBR), 1(4).
- Y a) * ÉXY ÉÁT æÉXÝ ÉÁY ÉÁY ÉÁY ÉÁY ÉÁQ ÉÁY ÉÁRa) * ÉZ ÉÁQ 3 * ÉXVÉÁO ÁŠã ÉÁO ÉÁQ EFÍ DÁ Ultrasound-assisted heating extraction of pectin from grapefruit peel: Optimization and comparison with the conventional method. Food Chemistry, 178, 106. 114.
- Wang, X., Wang, X., & Wang, T. (2013). Enrichment of arachidonic acid for the enzymatic synthesis of arachidonoyl ethanolamide. *Journal of the American Oil Chemists' Society*, *90*(7), 1031. 1039.
- Watada, A. E., Ko, N. P., & Minott, D. A. (1996). Factors affecting quality of freshcut horticultural products. *Postharvest Biology and Technology*, 9(2), 115. 125.

- Watada, A. E., & Qi, L. (1999). Quality of fresh-cut produce. *Postharvest Biology* and *Technology*, *15*(3), 201. 205.
- Wei, X., Chen, C., Yu, Q., Gady, A., Yu, Y., Liang, G., & Gmitter, F. G. (2014). Comparison of carotenoid accumulation and biosynthetic gene expression between Valencia and Rohde Red Valencia sweet oranges. *Plant Science*, 227, 28. 36.
- Wisutiamonkul, A., Ampomah-Dwamena, C., Allan, A. C., & Ketsa, S. (2017). Carotenoid accumulation in durian (Durio zibethinus) fruit is affected by ethylene via modulation of carotenoid pathway gene expression. *Plant Physiology and Biochemistry*.
- Wisutiamonkul, A., Ketsa, S., & van Doorn, W. G. (2015). Endogenous ethylene regulates a&& { ` |æ] } Á Á -æ) åÁ-carotene in the pulp of harvested durian fruit. *Postharvest Biology and Technology*, *110*, 18. 23.
- Wisutiamonkul, A., Promdang, S., Ketsa, S., & van Doorn, W. G. (2015). Carotenoids in durian fruit pulp during growth and postharvest ripening. *Food Chemistry*, *180*, 301. 305.
- Wu, J., Chen, S., Ge, S., Miao, J., Li, J., & Zhang, Q. (2013). Preparation, properties and antioxidant activity of an active film from silver carp (Hypophthalmichthys molitrix) skin gelatin incorporated with green tea extract. *Food Hydrocolloids*, 32(1), 42. 51.
- Xiao, H., Yao, Z., Peng, Q., Ni, F., Sun, Y., Zhang, C. X., & Zhong, Z. X. (2016). Extraction of squalene from camellia oil by silver ion complexation. Separation and Purification Technology, 169, 196. 201.
- Yen, G.-C., Duh, P.-D., & Chuang, D.-Y. (2000). Antioxidant activity of anthraquinones and anthrone. *Food Chemistry*, *70*(4), 437. 441.
- Zaghdoudi, K., Pontvianne, S., Framboisier, X., Achard, M., Kudaibergenova, R., Ayadi-V¦æà^|• ÂA Há hố ấng cá & ố carotenoids from: Tunisian Kaki (Diospyros kaki L.), peach (Prunus persica L.) and apricot (Prunus armeniaca L.). *Food Chemistry*, *184*, 131. 139.
- Zhang, C.-F., Zhang, S.-L., He, X., Yang, X.-L., Wu, H.-T., Lin, B.-Û∰ Â Â à * Â Z.-L. (2014). Antioxidant effects of Genkwa flos flavonoids on Freund s adjuvant-induced rheumatoid arthritis in rats. *Journal of Ethnopharmacology*, 153(3), 793. 800.

- Zhang, H.-F., Yang, X.-H., Zhao, L.-D., & Wang, Y. (2009). Ultrasonic-assisted extraction of epimedin C from fresh leaves of Epimedium and extraction mechanism. *Innovative Food Science & Emerging Technologies*, *10*(1), 54. 60.
- Zhang, H. F., Yang, X. H., Zhao, L. D., & Wang, Y. (2009). Ultrasonic-assisted extraction of epimedin C from fresh leaves of Epimedium and extraction mechanism. *Innovative Food Science and Emerging Technologies*, 10(1), 54. 60. https://doi.org/10.1016/j.ifset.2008.09.007

