

UNIVERSITI PUTRA MALAYSIA

DETERMINATION OF POST-MORTEM INTERVAL (PMI) VIA IMMUNOHISTOCHEMICAL LOCALIZATION AND EXPRESSION OF BIOGENIC AMINE (CADAVERINE)

YEOW MEI JUAN

FPV 2017 86

DETERMINATION OF POST-MORTEM INTERVAL (PMI) VIA IMMUNOHISTOCHEMICAL LOCALIZATION AND EXPRESSION OF BIOGENIC AMINE (CADAVERINE)

YEOW MEI JUAN

A project paper submitted to the Faculty of Veterinary Medicine, University Putra Malaysia In partial fulfilment of the requirement for the DEGREE OF DOCTOR OF VETERINARY MEDICINE

University Putra Malaysia Serdang, Selangor Darul Ehsan

MARCH 2017

CERTIFICATION

It hereby certified that I have read this project paper entitled "Determination of Postmortem Interval (PMI) via Immunohistochemical Localization and Expression of Biogenic Amine (Cadaverine)", by Yeow Mei Juan and in my opinion it is satisfactory in terms of scope, quality, and presentation as partial fulfilment of the requirement for the course VPD 4999 – Final Year Project.

PROF. DR. NOORDIN MOHAMED MUSTAPHA DVM (UPM), MSc (UPM), PHD (Murdoch)

Lecturer, Faculty of Veterinary Medicine University Putra Malaysia (Supervisor) This project is dedicated to my beloved family for being my tower of strength;

To all the animals that have suffered injustice;



DEDICATION

ACKNOWLEDGEMENTS

First and foremost, I would like to thank my supervisor, Prof. Dr. Noordin Mohamed Mustapha for giving me this opportunity to conduct this project. It was indeed one of the most unforgettable and interesting experiences throughout my DVM years.

A billion thank you I would like to bid to Dr. Mazlina Mazlan, for lending a warm, helping hand during the most difficult times, and also to the staff members of histopathology lab. All of your guidance and assistance will be remembered.

To my ever-supportive family and White White, thank you for the positive encouragements and support in everything I pursue, especially throughout the FYP period. Not forgetting my high school friends and best friends who are always there for me despite being far apart. Also my college roommate, Ginny Lim for being patient and spontaneously fun throughout these 5 years. A billion thank you to my rotation mates for 3 semesters (Gam Gajah) and my beloved DVM 2012/17 classmates for the physical and moral support whenever I needed the most.

Last but not least, I would like to thank myself for the perseverance, the courage and the undying spirit to strive despite all possible challenges. Never forget to stay humble, positive and firm in everything you pursue. Have faith, and always stay strong!

III

TABLE OF CONTENTS

CERTIFICATIONI
DEDICATIONII
ACKNOWLEDGEMENTSIII
TABLE OF CONTENTS IV
LIST OF TABLES
LIST OF PLATES
LIST OF FIGURES
LIST OF ABBREVIATIONS IX
ABSTRAKX
ABSTRACTXII
1.0 INTRODUCTION1
2.0 LITERATURE REVIEW
3.0 MATERIALS AND METHOD9
3.1 Tissue sampling
3.2 Hematoxylin-eosin staining
3.3 Immunohistochemistry staining method10
3.4 Image acquisition12
3.5 Immunohistochemistry data analysis – Expression of cadaverine
3.6 Statistical analysis13
4.0 RESULTS AND DISCUSSION14
4.1 Mean expression of cadaverine in brain, liver, muscle and testis14

 \bigcirc

IV

4.2 Morphometric analysis of cells	
4.3 Correlation between mean cadaverine expression and morphometric analysis	
of cells in brain, liver and testis21	
4.4 Correlation between arbitrary temperature changes and mean cadaverine	
expression in brain and liver21	
5.0 CONCLUSION	
6.0 RECOMMENDATIONS	
REFERENCES	

 \bigcirc

V

LIST OF TABLES

Table 1. Mean expression of cadaverine over the four time points post-mortem in the		
brain, liver, muscle and testis and muscle14		
Table 2. Mean morphometric measurement of area of the hepatocytes, neurons and		
seminiferous tubules over the four time points post-mortem		
Table 3. Correlation between temperature changes against mean expression of		
cadaverine in the brain and liver.		

G



LIST OF PLATES

Plate 2. Photomicrograph of periodic expression of cadaverine in the liver.......18

Plate 3. Photomicrograph of periodic expression of cadaverine in the muscle......19

VII

VIII

LIST OF FIGURES

 Figure 1. The exponential curve for mean expression of cadaverine in the brain, liver,

 muscle and testis over the 24 hours post-mortem period.
 .15

 Figure 2. The linear curve for mean expression of cadaverine in the brain, liver,

 muscle and testis over the 24 hours post-mortem period.
 .15

 Figure 3. The mean surface area of neurons, hepatocytes and testicular tubules during

 the experimental period
 .21

 Figure 4. Linear correlation between mean expression of cadaverine and temperature

 changes in brain and liver at different time points post-mortem
 .22

LIST OF ABBREVIATIONS

%	percent
°C	degree Celcius
μΙ	microliter
BSA	bovine serum albumin
cm	centimeter
DPX	dibutyl phthalate xylene
H&E	Hematoxylin and Eosin
ІНС	immunohistochemical
kg	kilogramme
mL	mililiter
mM	milimolar
PBS	phosphate buffered solution
PMI	post-mortem interval

IX

ABSTRAK

Abstrak kertas projek yang dikemukakan kepada Fakulti Perubatan Veterinar

untuk memenuhi sebahagian daripada keperluan kursus

VPD 4999 - Projek Tahun Akhir

PENGANGGARAN TEMPOH KEMATIAN MELALUI PEWARNAAN IMUNOHISTOKIMIA DAN KEPEKATAN AMINA BIOGENIK

(KADAVERINA)

Oleh

Yeow Mei Juan

2017

Penyelia: Prof. Dr. Noordin Mohamed Mustapha

Kekurangan pengetahuan mengenai sela masa pasca kematian (PMI) masih wujud disebalik penyelidikan meluas termasuklah amina biogenik (kadaverina) belum pernah dinilai sebagai petunjuk PMI. Maka, kajian ini bertujuan untuk mencadangkan parameter baru untuk penganggaran PMI melalui imunohistokimia dan taburan kepekatan kadaverina. Sampel otak, hati, otot dan testis yang diperolehi daripada tiga ekor anjing pada jam ke-0, ke-12, ke-18, dan ke-24 pasca eutanasia diawet dalam formalin 10% dan diproses untuk histologi dan imunohistokimia kadaverina (IHC). Sampel pewarnaan H&E juga dijalankan dan didedahkan kepada analisis morfometri. Kepekatan setanding kadaverina didapati pada jam ke-18 dan ke-24 pasca kematian adalah lebih tinggi daripada yang lain (p <0.01). Kepekatan kadaverina hepatik melebihi tisu-tisu lain, kecuali kepada otak (p<0.01). Walau bagaimanapun, kesan

bergantungan kepada masa didapati pada semua organ diuji (p<0.01). Walau bagaimanapun, kesan bergantungan kepada masa didapati pada semua organ diuji (p<0.01). Ukuran morfometri hepatosit, neuron dan tubul seminiferous berbeza secara ketara (p<0.01). Kesimpulannya, ternyata bahawa ungkapan kadaverina boleh berfungsi sebagai penganggar PMI. Walau bagaimanapun, kajian yang sama perlu tertakluk kepada amina biogenik yang berbeza dengan kuantifikasi tepat.

Kata kunci: Amina biogenik, imunohistokimia, kadaverina, sela masa pasca kematian

XI

ABSTRACT

XII

Abstract of the project paper presented to the Faculty of Veterinary

Medicine in partial requirement for the course

VPD 4999 – Final Year Project

DETERMINATION OF POST- MORTEM INTERVAL (PMI) VIA IMMUNOHISTOCHEMICAL LOCALIZATION AND EXPRESSION OF BIOGENIC AMINE (CADAVERINE)

by

Yeow Mei Juan

2017

Supervised by: Prof. Dr. Noordin Mohamed Mustapha

A dearth of knowledge still exists despite extensive research being conducted on a reliable post mortem interval (PMI) indicator. Biogenic amine has never been attempted as an indicator of PMI and is evaluated in this study based on immunologically expressed cadaverine. Samples of the brain, liver, muscle and testis obtained from three dogs at 0, 12th, 18th, and 24th hour post-euthanasia were immediately fixed in 10% formalin and routinely processed for histology (Haematoxylin and Eosin) and cadaverine immunohistochemistry (IHC). The H&Estained samples were also subjected to morphometric analysis. Comparable cadaverine expressions found at 18th and 24th hour post-mortem were higher than 0 and 12th hour (p<0.01). Hepatic cadaverine expression surpassed other tissues, except

XIII

of the brain (p<0.01). However, a time-dependent effect was found on all organs tested (p<0.01). The morphometry of hepatocytes, neurons and seminiferous tubules were significantly different (p<0.01). In conclusion, it appears that cadaverine expression may serve as a PMI indicator. However, similar studies should be subjected to different biogenic amines along with its exact quantification.

Key words: Biogenic amine, cadaverine, immunohistochemistry, post-mortem interval



1.0 INTRODUCTION

The study of the time since death, or the post-mortem interval (PMI) has been one of the most popular fields of research in human forensic medicine. However, in a veterinary context, published data on the estimation of the PMI is scarce, especially studies on companion animals and domestic livestock (Erlandsson and Munro, 2007). Furthermore, most of the data were from finding under temperate conditions which may be incompatible to those found in the tropics. Accurate estimation of the time of death is crucial to the investigation of death via eliminating possible suspects or events, especially in cases of alleged offences related to neglect or abuse of companion animals, deaths of high number of animals, or animal deaths during transportation. Besides, in accordance with the Malaysian Government Act 999 (2009) and Animal Welfare Act (2015), the establishment of a legal framework which primarily aims for animal welfare warrants the need of a veterinary forensic knowledge. Hence, a reliable medico-legal death time estimation would allow evidence-based prosecutions to be carried out.

Despite some improvements which have been made for the past 30 years, according to Swift (2010), the existence of any single, reliable and accurate method in estimating the time since death during the early PMI remains debatable. Munro and Munro (2012) stated two basic approaches to the estimation of time of death: (1) the measurement of change that takes place at a known rate, and (2) the comparison of the occurrence of events known to have taken place at a specific time with the time of death. A list of techniques adopted in veterinary forensic investigations for the

estimation of time of death was stated by Munro and Munro (2012), namely temperature-based methods, post-mortem chemistry, electrical stimulation of muscle and nerves, gross appearance of body (rigor mortis, decomposition, shape, colour and luminosity of the eye etc.), histopathology and electron microscopy, radiology, DNA and RNA analyses, entomology and environmental and associated evidence. However, the lack of validation of many of these methods due to failure in demonstrating quantitative measurement and inclusion of a mathematical description causes the results open to challenge which may lead to ambiguity in legal cases.

Besides, it is worth noting that veterinary pathologists often estimate the time since death based on their experience of gross post-mortem changes such as autolysis, livor mortis, rigor mortis and putrefaction. However, the vast range of species covered and the variety of circumstances in which they are found renders these estimations to be questionable. Similarly, experienced pathologists in the field of human forensic pathology frequently underestimate PMI (James and Knight, 1965).

Forensic entomology can be of considerable value in a veterinary context, although the identification of the types and stages of maggots and beetles requires a prerequisite of knowledge on insect fauna (Anderson and Huitson, 2004). However, it is deemed necessary for veterinary pathologists to acquire appropriate skills and knowledge in the collection of entomological evidence.

On the other hand, various studies on the pattern of temperature drop (algor mortis) in estimating PMI showed important findings. These include the thumb rule of postmortem rectal temperature fall of 1.5 ^oF per hour (Baccino *et al..*, 1996) and the

development of nomogram based on a single rectal measurement (Henssge, 1988), whereby the latter is currently the most widely accepted and practical method on time of death estimation. Body weight, varying degrees of ambience, the effect of wind, surface conductance and irradiation were among the confounding factors which have to be taken into consideration when applying temperature-based methods. A study by Abdulazeez and Noordin (2010) showed the rate of cooling under tropical conditions was less consistent than in temperate climates, also the absence of lag phase in the cooling curve which was derived from temperature measurements of organs in dogs as compared to human (Erlandsson, 2003; Abdulazeez and Noordin, 2010; Okene, 2010).

Thus, careful consideration and attention are required during the application of temperature-based methods in different regions, also during extrapolation between species. Researches on other methods in the determination of post-mortem interval, such as post-mortem chemistry, microscopic and ultrastructural changes and post-mortem radiology, however may serve as means of support and refinement, or as alternatives for further investigation and independent validation which are of relevance to forensic veterinary pathology (Munro and Munro, 2012).

Three main sources in which investigators could derive additional information during the determination of PMI would be evidences i. obtained from the carcass, ii. associated with the environment and iii. anamnestic which is based on ante-mortem movements or daily activities of an individual (Swift, 2010). Studies on methods extolling the vast number of identifiable changes which may occur during the PMI

should therefore being constantly conducted, in order to attribute a temporal value for the time since death.



REFERENCES

Abdulazeez, I. O., & Noordin, M. M. (2010). Algor mortis pattern in dogs, a guide to estimation of time of death. Pertanika J trop Agric Sci, 33, 105-111.

Barron, S., Mulholland, P. J., Littleton, J. M., & Prendergast, M. A. (2008). Age and gender differences in response to neonatal ethanol withdrawal and polyamine challenge in organotypic hippocampal cultures. *Alcoholism: Clinical and Experimental Research*, *32*(6), 929-936.

Bastida, C. M., Cremades, A., Castells, M. T., López-Contreras, A. J., López-García, C., Sánchez-Mas, J., & Penafiel, R. (2007). Sexual dimorphism of ornithine decarboxylase in the mouse adrenal: influence of polyamine deprivation on catecholamine and corticoid levels. *American Journal of Physiology-Endocrinology and Metabolism*, 292(4), E1010-E1017.

Casero, R. A., & Pegg, A. E. (2009). Polyamine catabolism and disease. *Biochemical Journal*, *421*(3), 323-338.

Chen, G. G., Fiori, L. M., Moquin, L., Gratton, A., Mamer, O., Mechawar, N., & Turecki, G. (2010). Evidence of altered polyamine concentrations in cerebral cortex of suicide completers. *Neuropsychopharmacology*, *35*(7), 1477-1484.

Chen, G. G., Turecki, G., & Mamer, O. A. (2009). A quantitative GC-MS method for three major polyamines in postmortem brain cortex. *Journal of mass spectrometry*, *44*(8), 1203-1210.

Chin, H. C., Marwi, M. A., Salleh, A. M., Jeffery, J., & Omar, B. (2007). A preliminary study of insect succession on a pig carcass in a palm oil plantation in Malaysia. Tropical Biomedicine, 24(2), 23-27.

Dent, B., Forbes, S. & Stuart, B. 2004. Review of human decomposition processes in soil. *Environmental Geology* 45(4): 576-585.

Erlandsson, M., & Munro, R. (2007). Estimation of the post-mortem interval in beagle dogs. Science & Justice, 47(4), 150-154.

Farn, G., & Sims, G. G. (1987). Chemical indices of decomposition in tuna. In Seafood quality determination: proceedings of the International Symposium on Seafood Quality Determination, coordinated by the Univ. of Alaska Sea Grant College Program, Anchorage, AK/edited by DE Kramer, J. Liston. Amsterdam: Elsevier, 1987.

Ferioli, M. E., Pinotti, O., & Pirona, L. (1999). Gender-related differences in polyamine oxidase activity in rat tissues. *Amino acids*, *17*(2), 139-148.

Gilad, G. M., & Gilad, V. H. (2002). Stress-induced dynamic changes in mouse brain polyamines. Role in behavioral reactivity. *Brain research*, *943*(1), 23-29.

Igarashi, K., Hara, K., Watanabe, Y., Hirose, S., & Takeda, Y. (1975). Polyamine and magnesium contents and polypeptide synthesis as a function of cell growth. *Biochemical and biophysical research communications*, 64(3), 897-904.

Igarashi, K., & Kashiwagi, K. (2010). Modulation of cellular function by polyamines. *The international journal of biochemistry & cell biology*, 42(1), 39-51.

Janaway, R.C., Percival, S.L. & Wilson, A.S. 2009a. Decomposition of human remains. In *Microbiology and Aging*, edited by Percival, S.L. New York: The Humana Press. pp. 313-334.

Marton, L. J., & Pegg, A. E. (1995). Polyamines as targets for therapeutic intervention. *Annual review of pharmacology and toxicology*, *35*(1), 55-91.

Nelson, E. L. (2000). Estimation of short-term postmortem interval utilizing core body temperature: a new algorithm. Forensic science international, 109(1), 31-38.

Paczkowski, S. & Schütz, S. 2011. Post-mortem volatiles of vertebrate tissue. *Applied Microbiology and Biotechnology* 91(4): 917-935.

Pegg, AE; McCann, PP (1982). "Polyamine metabolism and function". <u>American</u> Journal of Physiology. **243**: 212–21. PMID 6814260..

Pegg, A. E. (2009). Mammalian polyamine metabolism and function. *IUBMB life*, 61(9), 880-894.

Santos, M. S. (1996). Biogenic amines: their importance in foods. *International journal of food microbiology*, 29(2), 213-231.

Seiler, N., & Atanassov, C. L. (1994). The natural polyamines and the immune system. In *Progress in Drug Research/Fortschritte der Arzneimittelforschung/Progrès des recherches pharmaceutiques* (pp. 87-141). Birkhäuser Basel.

Seiler, N., & Raul, F. (2005). Polyamines and apoptosis. *Journal of cellular and molecular medicine*, 9(3), 623-642.

Tamim, N. M., & Doerr, J. A. (2003). Effect of putrefaction of poultry carcasses prior to rendering on biogenic amine production. *The Journal of Applied Poultry Research*, *12*(4), 456-460.

Tamime, A. Y. (Ed.). (2017). Microbial Toxins in Dairy Products. John Wiley & Sons.

Vass, A. A. (2001). Beyond the grave-understanding human decomposition. *Microbiology today*, 28, 190-193.

Wallace, H. M., Fraser, A. V., & Hughes, A. (2003). A perspective of polyamine metabolism. *Biochemical Journal*, *376*(1), 1-14.