



UNIVERSITI PUTRA MALAYSIA

***MOLECULAR DETECTION OF FELINE LEUKEMIA VIRUS (FeLV)
IN CLINICALLY ILL LOCAL CATS***

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MOLECULAR DETECTION OF FELINE LEUKEMIA VIRUS (FeLV)

IN CLINICALLY ILL LOCAL CATS

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CERTIFICATION

It is hereby certified that we have read this project entitled “Molecular detection of Feline leukemia virus (FeLV) in clinically ill local cats”, by Kunambiga Mummoorthy and in our opinion it is satisfactory in terms of scope, quality, and presentation as partial fulfilment of the requirement for the course of VPD 4999-Final Year Project

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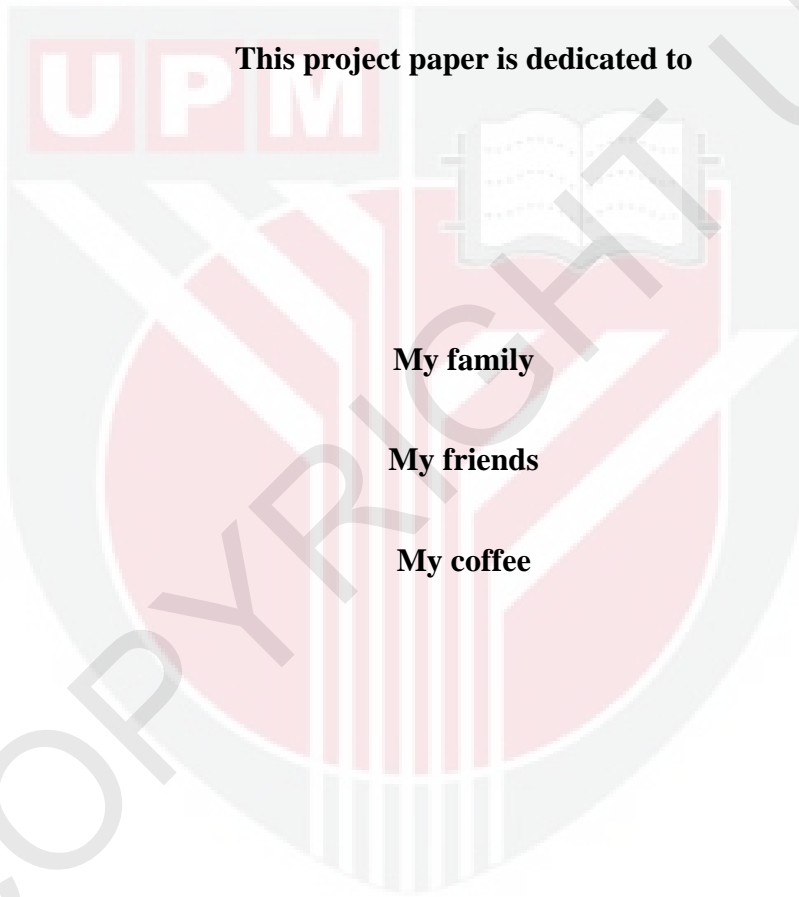
DEDICATIONS

This project paper is dedicated to

My family

My friends

My coffee



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LIST OF ABBREVIATIONS

%	Percent
μL	Microliter
μM	Micromolar
°C	Degree Celsius
ATCC	American Type-culture Collections
BLAST	Basic Local Alignment Search Tool
bp	Base pair
CD4+	Cluster of differentiation 4
CD8+	Cluster of differentiation 8
DNA	Deoxyribonucleic acid
DNase	Deoxyribonuclease
Dntp	Deoxyribonucleotide triphosphate
EDTA	Ethylenediaminetetraacetic acid
ELISA	Enzyme-linked immunosorbent Assay
FeLV	Feline leukemia virus
g	Gram
IACUC	Institutional Animal Care and Use Committee
IN	Integrase
Kb	Kilobase
LTRs	Long Terminal repeats
MEGA	Molecular Evolutionary Genetics Analysis

mL	Milliliter
mg	Milligram
mg/kg	Milligram per kilogram
MgSO ₄	Magnesium Sulphate
Mm	Millimeter
NCBI	National Center for Biotechnology Information
NJ	Neighbour-joining
PBS	Phosphate buffer solution
RNA	Ribonucleic acid
RNase	Ribonuclease
RT	Reverse transcriptase
SU	Surface protein
RT-PCT	Reverse Transcription Polymerase Chain Reaction
TAE	Tris-acetate-ethylenediaminetetraacetic acid
UPM	Universiti Putra Malaysia
USA	United States of America
UK	United Kingdom
UV	Ultraviolet
w/v	Weight per volume
xg	Relative centrifugal force

ABSTRAK

Abstrak daripada kertas projek yang dikemukakan kepada Fakulti Perubatan Veterinar untuk memenuhi sebahagian daripada keperluan kursus VPD 4999-Projek Ilmiah Tahun Akhir

PENGESANAN MOLEKUL FELINE LEUKEMIA VIRUS (FeLV) DALAM KUCING TEMPATAN YANG SAKIT SECARA KLINIKAL

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2017

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Feline leukemia virus (FeLV) tergolong di bawah genus Gammaretrovirus dan dikaitkan dengan pelbagai tanda-tanda klinikal di seluruh dunia. Setakat ini, pencirian molekul FeLV tempatan yang pertama dan satu-satunya dilakukan pada tahun 2014 telah

mendedahkan bahawa pencilan tempatan berkait rapat dengan pencilan UK. Oleh kerana liputan terhad kajian dan sifat semula jadi virus yang biasanya mengintegrasikan DNA ke dalam genom dan menjalani mutasi, kajian mengenai status semasa jangkitan FeLV adalah diperlukan. Oleh itu, kajian ini bertujuan untuk mengesan antigen FeLV pada kucing sakit secara klinikal dengan menggunakan kaedah RT-PCR dan untuk membandingkan persamaan varian yang dikenalpasti pada masa ini dengan pencilan virus terdahulu dari Malaysia dan taburan geografi yang lain. Dengan menggunakan kaedah pensampelan mudah, plasma dan air liur dikumpulkan dari 15 kucing sakit secara klinikal dan 5 kucing sihat dari Hospital Gasing Veterinar. Nukleik asid virus telah diasingkan dan tertakluk kepada One-Step RT-PCR dengan primer khusus yang mensasarkan kawasan U3LTR dan gag separa yang sangat terpelihara. Dua kucing telah diuji positif untuk antigen dari kumpulan sakit secara klinikal. Separa nukleotida penjujukan and analisis filogenetik mendedahkan bahawa varian semasa didapati 93-99% homolog kepada pencilan Malaysia sebelum ini dan masih berkait rapat dengan varian daripada UK. Menariknya, ia juga didapati berkait rapat dengan varian yang diasingkan dari Jepun. Kesimpulannya, kajian ini menonjolkan kemungkinan hubungan evolusi di antara FeLV dari Malaysia dengan FeLV dari UK dan Jepun.

Kata kunci: *Feline Leukemia Virus (FeLV), RT-PCR, Separa nukleotida, penjujukan, analisis filogenetik, varian*

ABSTRACT

Abstract of the project paper presented to the Faculty of Veterinary Medicine in partial requirement for the course VPD 4999-Final Year Project

**MOLECULAR DETECTION OF FELINE LEUKEMIA VIRUS IN
CLINICALLY ILL IN LOCAL CATS****By****Kunambiga Mummoorthy****2017****Supervisor: Dr Nor Yasmin Abd. Rahaman****Co-supervisors:****Professor Dr.Abd. Rahman Omar****Associate Professor Dr.Siti Suri Arshad****Dr. Nurfazila Saulol Hamid****Dr.Prem Anand**

Feline leukemia virus (FeLV) belongs under the genus Gammaretrovirus and is associated with a wide range of clinical signs worldwide. Sofar, the first and only molecular characterization of local FeLV isolates performed in 2014 revealed that local isolates to be closely related to UK isolate. Due to limited coverage of the study and the nature of the virus that typically integrates the DNA into the host genome and undergoes mutation, study on the current status of FeLV infection is necessary. Therefore, this study aim to detect FeLV antigen in clinically ill cats by RT-PCR and to compare the

currently identified variant similarity with previous related virus isolates from Malaysia and other geographical distribution. By using convenience sampling method, plasma and saliva were collected from 15 clinically ill cats and 5 healthy cats from Gasing Veterinary Hospital. Viral nucleic acid was extracted and subjected to One-Step RT-PCR with specific primer targeting the highly conserved U3LTR and partial gag regions. Two cats were tested positive for the antigen from the clinically ill group. Partial nucleotide sequencing and phylogenetic analysis revealed that the current variant are found to be 93-99% homologous to the previous Malaysian isolates and is still closely related to UK isolate. Interestingly, they were also found to be closely related to isolates from Japan. In conclusion, this study highlights the possibilities of evolutionary relations between FeLV from Malaysia with FeLV of UK and Japan.

Keywords: *Feline leukemia virus, RT-PCR, partial nucleotide sequencing, phylogenetic analysis*

1.0 INTRODUCTION

Feline leukaemia virus (FeLV) belongs to the Gammaretrovirus genus from the family of Retroviridae. It was first described in the year 1964 by Jarrett *et al.*, and is a naturally occurring gammaretrovirus that infects domestic cats and sporadically wild cats (Arjona *et al.*, 2007; Torres *et al.*, 2008; Bolin *et al.*, 2011). It is an enveloped, positive sense, single-stranded RNA virus (Boeke *et al.*, 2011). There are currently four FeLV subgroups of clinical importance namely FeLV-A, FeLV-B, FeLV-C or FeLV-T (Chandhasin *et al.*, 2005; Levy, 2008). Subgroup FeLV-A is found in all naturally infected cats meanwhile subgroup B arises from the recombination of endogenous FeLV with FeLV-A whereas FeLV subgroup C and T are the mutated forms of FeLV-A. (McCaw, 2010; Eiden *et al.*, 2010). FeLV is mostly transmitted via the oro-nasal route via saliva and nasal secretions through sharing of food and water dishes as well as the cat's grooming or aggressive behaviour. Occasionally, vertical transmissions could occur but is of little relative significance (Gomes-Keller *et al.*, 2006; McCaw, 2010).

FeLV related disorders are associated with the manifestation of immunosuppression, lymphoid or myeloid tumors, anaemia, reproductive problems, immune complexes, enteritis and certain other disorders (McCaw, 2010). FeLV infections are divided into four stages namely the abortive infection (regressor cats), regressive infection (transient viraemia followed by latent infection), progressive infection (persistent viraemia) and focal or atypical infection (Torres *et al.*, 2005, Hofmann-Lehmann *et al.*, 2007; 2008, Levy *et al.*, 2008). The disease is distributed worldwide in the feline population and the

prevalence varies greatly with geographical location and risk factors such as health status, age and population density (Gleich *et al.*, 2009, Bande *et al.*, 2012).

In most clinics and hospitals, the diagnosis of FeLV is usually done with the detection of p27 antigen with rapid test kits (Hartmann *et al.*, 2007). Conversely, the demonstration of p27 antigen is relatively difficult during the early viraemia and latent infections and studies conducted showed that FeLV viral RNA and provirus DNA are better predictors of progressive and latent infections respectively (Cattori *et al.*, 2009).

To date, the first and only molecular assay investigation of the clinical status of Malaysian cats was carried out in 2014 by Bande *et al.*, together with the first sequencing and phylogenetic characterizations of the Malaysian FeLV isolates that revealed Malaysian FeLV to be highly homologous to each other and showed a possible evolutionary relationship with FeLV in UK. The hypothesis for this study proposes that the current local FeLV variant would be highly homologous to previous Malaysian isolates and would likely be closely related to the UK strain. This study was undertaken to fulfil these following objectives:

1. To detect FeLV antigen in clinically ill cats by using RT-PCR method.
2. To compare currently detected FeLV with previously detected FeLV from Malaysia and other geographical distribution

REFERENCES

- Arjona, A., Barquero, N., Dome´nech, A., Tejerizo, G., Collado, V. M., Toural, D. M. & Go´mez-Lucia, E. (2007). Evaluation of a novel nested PCR for the routine diagnosis of feline leukemia virus (FeLV) and feline immunodeficiency virus (FIV). *J Feline Med Surg*, 9, 14–22.
- Al-Kappany, Y.M., Lappin, M.R., Kwok, O.C.H., AbuElwafa, S.A., Hilali, M & Dubey, J.P. (2011) “Seroprevalence of *Toxoplasma gondii* and concurrent *Bartonella* spp., Feline immunodeficiency virus, feline leukemia virus, and *Dirofilaria immitis* infections in Egyptian cats,” *Journal of Parasitology*, 97(2), 256–258.
- Bande, F., Arshad, S. S., Hassan, L., Zakaria, Z., Sopian, N. A., Rahman, N. A., & Alazawy, A. (2012). Prevalence and risk factors of feline leukaemia virus and feline immunodeficiency virus in peninsular Malaysia. *BMC veterinary research*, 8(1), 33.
- Bande, F., Arshad, S. S., Hassan, L., & Zakaria, Z. (2014). Molecular detection, phylogenetic analysis, and identification of transcription motifs in feline leukemia virus from naturally infected cats in Malaysia. *Veterinary medicine international*, 2014, 1-10
- Bolin, L. L., Chandhasin, C., Lobelle-Rich, P. A., Albritton, L. M. & Levy, L. S. (2011). Distinctive receptor binding properties of the surface glycoprotein of a natural feline leukemia virus isolate with unusual disease spectrum. *Retrovirology*, 8(1), 35-52.
- Bannert, N., Fiebig, U. & Hohn, O. (2010). Retroviral Particles, Proteins and Genomes. In Kurth, R., Bannert, N. (Eds.) *Retroviruses; Molecular Biology, Genomics and Pathogenesis* (pp. 71-106). Norfolk, UK: Caister Academic press
- Boeke, J.D., Eickbush, T., Sandmeyer, S.B. & Voylas, D.F. (2011). The Reverse Transcribing DNA and RNA viruses. In King, A. M., Lefkowitz, E., Adams, M. J., & Carstens, E. B. (Eds.), *Virus taxonomy: Ninth report of the International Committee on Taxonomy of Viruses* (pp. 427-496) London, UK: Elsevier

- Cattori, V., Tandon, R., Riond, B., Pepin, A. C., Lutz, H. & Hofmann-Lehmann, R. (2009). The kinetics of feline leukaemia virus shedding in experimentally infected cats are associated with infection outcome. *Veterinary microbiology*, 133(3), 292-296.
- Chew-Lim, M., Fong, N. & Chong, S. Y. (1989). "A survey of the feline leukaemia virus in Singapore," *Annals of the Academy of Medicine Singapore*, 18(6), 646-648.
- Chandhasin, C. P., Lobelle-Rich, & Levy, L. S. (2004). "Feline leukaemia virus LTR variation and disease association in ageographical and temporal cluster," *Journal of General Virology*, 85, 2937-2942.
- Chandhasin, C., Coan, P. N. & Levy, L. S. (2005). Subtle mutational changes in the SU protein of a natural feline leukemia virus subgroup A isolate alter disease spectrum. *Journal of virology*, 79(3), 1351-1360.
- Coelho, F. M., Bomfim, M. R. Q. & Caxito, F. D. A. (2008). "Naturally occurring feline leukemia virus subgroup A and B infections in urban domestic cats," *Journal of General Virology*, 89(11), 2799-2805.
- Donahue, P. R., Hoover, E. A., Beltz, G. A., Riedel, N., Hirsch, V. M., Overbaugh, J. & Mullins, J. I. (1988). Strong sequence conservation among horizontally transmissible, minimally pathogenic feline leukemia viruses. *Journal of Virology*, 62(3), 722-731.
- Dunham, S. P. & Graham, E. (2008). Retroviral infections of small animals. *Veterinary Clinics of North America: Small Animal Practice*, 38(4), 879-901.
- Eiden, M. V., Radke, K., Rovnak, J. & Quackenbush, S. L. (2010). Non-primate Mammalian and Fish Retroviruses. In Kurth, R., Bannert, N. (Eds.) *Retroviruses; Molecular Biology, Genomics and Pathogenesis* (pp. 371-394). Norfolk, UK: Caister Academic press
- Gomes-Keller, M. A., Gönczi, E., Tandon, R., Riondato, F., Hofmann-Lehmann, R., Meli, M. L. & Lutz, H. (2006). Detection of feline leukemia virus RNA in saliva from naturally infected cats and correlation of PCR results with those of current diagnostic methods. *Journal of clinical microbiology*, 44(3), 916-922.

- Gleich, S. E., Krieger, S. & Hartmann, K. (2009). Prevalence of feline immunodeficiency virus and feline leukaemia virus among client-owned cats and risk factors for infection in Germany. *Journal of Feline Medicine & Surgery*, 11(12), 985-992.
- K. Hartmann, P., Griessmayr, B. & Schulz (2007). "Quality of different in-clinic test systems for feline immunodeficiency virus and feline leukaemia virus infection," *Journal of Feline Medicine and Surgery*, 9(6), 439–445.
- Hartmann, K. (2012). Clinical aspects of feline retroviruses: a review. *Viruses*, 4(11), 2684-2710.
- Hofmann-Lehmann, R., Cattori, V., Tandon, R.; Boretti, F.S., Meli, M.L., Riond, B., & Lutz, H. (2008). How molecular methods change our views of felv infection and vaccination. *Vet Immunol Immunopathol*, 123, 119-123.
- Hofmann-Lehmann, R., Cattori, V., Tandon, R., Boretti, F. S., Meli, M. L., Riond, B. & Lutz, H. (2007). Vaccination against the feline leukaemia virus: outcome and response categories and long-term follow-up. *Vaccine*, 25(30), 5531-5539.
- Hofmann-Lehmann, R., Huder, J.B., Gruber, S., Boretti, F., Sigrist, B. & Lutz, H. (2001). Feline leukaemia provirus load during the course of experimental infection and in naturally infected cats. *J Gen Virol* 2001, 82, 1589-1596
- Hardy, W. D., Old, L. J., Hess, P. W., Essex, M. & Cotter, S. (1973). Horizontal transmission of feline leukaemia virus. *Nature*, 244(5414), 266-269.
- Jarrett, W., Crawford, E., Martin, W. & Davie, F., (1964). Leukemia in the cat: A virus-like particle associated with leukemia (lymphosarcoma). *Nature*, 202, 562-569.
- Kawamura, M., Watanabe, S., Odahara, Y., Nakagawa, S., Endo, Y., Tsujimoto, H. & Nishigaki, K. (2015). Genetic diversity in the feline leukemia virus gag gene. *Virus research*, 204, 74-81.
- Little, S., Sears, W., Lachtara, J. & Bienzle, D. (2009). "Seroprevalence of feline leukemia virus and feline immunodeficiency virus infection among cats in Canada," *Canadian Veterinary Journal*, 50(6), 644–648

- Lee, I.T., Levy, J.K., Gorman, S.P., Crawford, P.C. & Slater, M.R. (2002). "Prevalence of feline leukemia virus infection and serum antibodies against feline immunodeficiency virus in unowned free-roaming cats," *Journal of the American Veterinary Medical Association*, 220(5), 620–622.
- Levy, J. K., Scott, H. M., Lachtara, J. L. & Crawford, P. C. (2006). Seroprevalence of feline leukemia virus and feline immunodeficiency virus infection among cats in North America and risk factors for seropositivity. *Journal of the American Veterinary Medical Association*, 228(3), 371-376.
- Levy, L.S. (2008). "Advances in understanding molecular determinants in FeLV pathology," *Veterinary Immunology and Immunopathology*, 123(1-2), 14–22.
- Levy, J., Crawford, C., Hartmann, K., Hofmann-Lehmann, R., Little, S., Sundahl, E. & Thayer, V. (2008). 2008 American Association of Feline Practitioners' feline retrovirus management guidelines. *Journal of Feline Medicine and Surgery*, 10(3), 300-316.
- Lutz, H., Addie, D., Belák, S., Boucraut-Baralon, C., Egberink, H., Frymus, T. & Marsilio, F. (2009). Feline leukaemia. ABCD guidelines on prevention and management. *Journal of Feline Medicine & Surgery*, 11(7), 565-574.
- Lin, J.A., Cheng, M.C. & Inoshima, Y. (1995). "Seroepidemiological survey of feline retrovirus infections in cats in Taiwan in 1993 and 1994," *The Journal of Veterinary Medical Science*, 57(1), 161–163.
- Major, A., Cattori, V., Boenzli, E., Riond, B., Ossent, P., Meli, M.L., Hofmann-Lehmann, R. & Lutz, H. (2010). Exposure of cats to low doses of felv: Seroconversion as the sole parameter of infection. *Vet Research*, 1(2), 1-10
- Mansky, L. M. (1998). Retrovirus mutation rates and their role in genetic variation. *Journal of general Virology*, 79(6), 1337-1345.
- Maruyama, S., Kabeya, H. & Nakao, R. (2003). "Seroprevalence of Bartonella henselae, toxoplasma gondii, FIV and FeLV infections in domestic cats in Japan," *Microbiology and Immunology*, 47(2), 147–153.

- McCaw, D.L. (2010). Feline Leukemia Virus and Related Disease. In Kahn, C.M. *The Merck Veterinary Manual* (10th ed., pp. 718-722) New Jersey, U.S.A: Merck.
- Miyazawa, T., Ikeda, Y., & Maeda, K. (1998). "Seroepidemiological survey of feline retrovirus infections in domestic and leopard cats in Northern Vietnam 1997," *Journal of Veterinary Medical Science*, 60(11), 1273–1275.
- Miyazawa, T., & Jarrett, O. (1997). "Feline leukaemia virus proviral DNA detected by polymerase chain reaction in antigenaemic but non-viraemic ("discordant") cats," *Archives of Virology*, 142(2), 323–332.
- Miyazawa, T. (2002). Infections of feline leukemia virus and feline immunodeficiency virus. *Front Biosci*, 7, 504-518.
- Soe, L. H., Devi, B. G., Mullins, J. I. & Roy-Burman, P. (1983). Molecular cloning and characterization of endogenous feline leukemia virus sequences from a cat genomic library. *Journal of virology*, 46(3), 829-840
- Torres, A. N., Mathiason, C. K. & Hoover, E. A. (2005). Re-examination of feline leukemia virus: host relationships using real-time PCR. *Virology*, 332(1), 272-283.
- Torres, A. N., O'Halloran, K. P., Larson, L. J., Schultz, R. D. & Hoover, E. A. (2008). Development and application of a quantitative real-time PCR assay to detect feline leukemia virus RNA. *Veterinary immunology and immunopathology*, 123(1), 81-89.
- Pepin, A.C., Tandon, R., Cattori, V., Niederer, E., Riond, B., Willi, B., Lutz, H. & Hofmann-Lehmann, R. (2007). Cellular segregation of feline leukemia provirus and viral rna in leukocyte subsets of long-term experimentally infected cats. *Virus Res* 2007, 127, 9-16.
- Vobis, M., D'Haese, J., Mehlhorn, H. & Mencke, N. (2003). Evidence of horizontal transmission of feline leukemia virus by the cat flea (*Ctenocephalides felis*). *Parasitology research*, 91(6), 467-470.