



UNIVERSITI PUTRA MALAYSIA

***PHENOL DEGRADATION AND MOLECULAR VALIDATION OF
PHENOL HYDROXYLASE GENE OF *Alcaligenes faecalis****

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PENGESAHAN

Dengan ini adalah disahkan bahawa tesis projek yang bertajuk “Phenol degradation and molecular validation of phenol hydroxylase gene of *Alcaligenes faecalis*” telah disiapkan serta dikemukakan kepada Jabatan Biokimia oleh Nur Muhamad Syahir Bin Abdul Habib (161060) sebagai syarat untuk kursus BCH4999 Projek.

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ABSTRACT

Currently, phenol pollution has caused some environmental concerns as it causes severe toxicity towards human health and environmental conditions. Intensive efforts to reduce the contamination of pollutants have been done especially in bioremediation techniques. Many microbial species have been introduced to be utilised for contamination clean-up and one of them is *Alcaligenes faecalis*. A study on the phenol degrading ability and molecular analysis of *A. faecalis* was conducted. To study the phenol degrading ability of *A. faecalis*, the bacteria was incubated in six different concentrations of phenol – 0.1, 0.4, 0.5, 0.9, 1.0, and 1.5 g/L. The growth of bacteria and phenol degradation in each phenol concentration were monitored. Among all the concentrations studied, phenol concentration of 0.9 g/L showed the highest degradation rate. Meanwhile, the molecular analysis of the bacteria was carried out by isolating the phenol hydroxylase gene which is responsible for degrading phenol by using the designated primer. The gene was amplified by using PCR technique with an annealing temperature of 56.4°C. The expected size of the gene was between 300 - 400 bp. After DNA sequencing, molecular analysis was done and the DNA fragment obtained had a length of 337 bp. Next, BLAST search was used to confirm the sequence obtained was phenol hydroxylase gene isolated from *A. faecalis*. The BLAST result showed that the phenol hydroxylase gene was successfully amplified from the bacteria. These studies showed the hypothetical use of this bacteria to treat phenol-contaminated environment.

ABSTRAK

Kini, pencemaran fenol telah meningkatkan keprihatinan manusia terhadap alam sekitar kerana fenol telah menyebabkan kesan toksik yang teruk terhadap kesihatan manusia dan alam sekitar. Langkah-langkah intensif telah dilakukan untuk mengurangkan pencemaran terutamanya melalui teknik bioremediasi. Pelbagai spesis mikroorganisma telah diketengahkan untuk pembersihan bahan cemar tersebut dan salah satu daripada mikroorganisma tersebut adalah bakteria *Alcaligenes faecalis*. Satu kajian mengenai keupayaan menguraikan fenol dan analisis molekular oleh *A. faecalis* telah dijalankan. Bagi mengkaji keupayaan menguraikan fenol, bakteria ini telah diletakkan di dalam kepekatan fenol yang berbeza - 0.1, 0.4, 0.5, 0.9, 1.0, dan 1.5 g/L. Pertumbuhan bakteria dan tahap penguraian fenol oleh setiap kepekatan fenol telah dicatat. Berdasarkan hasil kajian, kepekatan fenol 0.9 g/L menunjukkan kadar penguraian yang tertinggi. Sementara itu, untuk menganalisis bakteria ini dari segi molekular, gen fenol hidroksilase yang bertanggungjawab menguraikan fenol telah diasingkan menggunakan primer gen yang telah direka. Seterusnya, gen itu diamplyfikan menggunakan kaedah PCR bersama dengan suhu perlekatan iaitu 56.4°C. Saiz anggaran gen adalah diantara 300 - 400 bp. Selepas penjujukan DNA, analisis molekular telah dijalankan dan fragmen DNA yang diperolehi adalah 337 bp. Seterusnya, carian BLAST telah dibuat bagi mengesahkan jujukan DNA adalah gen fenol hidroksilase daripada *A. faecalis*. Hasil BLAST menunjukkan gen fenol hidroksilase berjaya diasingkan daripada bakteria ini. Kajian ini menunjukkan kebolehan penggunaan bakteria ini untuk merawat alam sekitar yang tercemar dengan fenol.

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LIST OF ABBREVIATIONS

% (w/v)	Percent concentration weight / volume
°C	Degree celsius
(NH ₄) ₂ SO ₄	Ammonium sulphate
μl	Microlitre
4-AAP	4-aminoantiprene
bp	Base pair
DNA	Deoxyribonucleic acid
dNTP	Deoxyribonucleotide triphosphate
g	Gram
kb	Kilobase
K ₂ Fe(CN) ₆	Potassium ferric cyanide
K ₂ HPO ₄	Di-potassium hydrogen phosphate
KH ₂ PO ₄	Potassium dihydrogen phosphate
L	Litre
mg	Milligram
MgCl ₂	Magnesium chloride
MgSO ₄	Magnesium sulphate
min	Minutes
MSM	Mineral salt media
ml	Millilitre
mM	Millimolar
NaCl	Sodium chloride
nm	Nanometer
PCR	Polymerase chain reaction
rpm	Revolution per minute
TAE	Tris-acetate-EDTA
UV	Ultraviolet
V	Voltage

CHAPTER 1

INTRODUCTION

Phenol and phenolic compounds are being widely distributed within the environment partly because of natural processes and non-natural processes, due to human and industrial activities. These aromatic pollutants originated mainly from the industrial processes such as resin manufacturing, oil refineries, petrochemicals, steel, pharmaceuticals, dyes, textiles, plastic, as well as the pulp and paper industries (Annadurai *et al.*, 2002; Duan, 2011; Ahmad *et al.*, 2012; Mahiudddin *et al.*, 2012).

Phenol is classified as a highly hazardous chemical for being a persistent compound. This is due to its toxicity, carcinogenic and mutagenic characteristics toward human being, fish, and environment as well as to several biochemical functions (Duan, 2011; Ahmad *et al.*, 2012). The pollution effects of phenolic effluent have been reported worldwide and removing them has become a major environmental concern (Mahiudddin *et al.*, 2012).

The removal of contaminants from the polluted area has been widely applied using different physical, chemical and biological methods. The biological technique for phenol reduction has been chosen as the technique that poses relatively low cost and offers complete mineralisation (Basha *et al.*, 2010; Nawawi *et al.*, 2010).

Microorganisms are heavily involved in the bioremediation process (Krastanov *et al.*, 2013). In recent years, the number of research on microbial degradation has increased due to its sustainability ways of cleaning up

contaminated areas. Most microbes could quickly adapt and grow at extreme condition using those hazardous compounds as both their carbon and energy sources (Mahiuddin *et al.*, 2012). Vast variety of microbes is known to be capable of degrading phenol under both aerobic and/or anaerobic conditions that are regulated by the action of enzymes and the microbial metabolism itself (Mahiuddin *et al.*, 2012; Sridevi *et al.*, 2012; Nawawi *et al.*, 2014).

One of the phenol degrading bacteria that has been studied is the *Alcaligenes faecalis* (Rehfuss and Urban, 2005). Many present studies have shown that *A. faecalis* has the property as a resourceful phenol-degrading bacterium. The effectiveness of a certain catabolic pathway usually depends on the properties of the involved key enzyme (Jiang *et al.*, 2007).

Phenol hydroxylase is the key enzyme that responsible for the conversion of phenol to catechol, which is the initial and rate-limiting step in phenol degradation pathways. Both single component and multi-component types of this enzyme have been identified with the latter recognised as the prevalent one in natural environments (Zhang *et al.*, 2004).

Recent studies have shown that *A. faecalis* has the ability to degrade phenol using phenol hydroxylase enzyme. However, further research and studies are needed in obtaining a more specific information on the degrading mechanism of the enzyme.

Thus, a study was carried out with the following objectives:

1. To evaluate the ability of *A. faecalis* to degrade phenol.
2. To determine the phenol degradation rate by *A. faecalis* using different concentration of phenol.
3. To isolate and analyse the phenol hydroxylase gene from *A. faecalis*.



REFERENCES

- Abd-Elsalam, K. A. 2003. Bioinformatic tools and guideline for PCR primer design. *African Journal of Biotechnology*, 2(5): 91-95.
- Ahmad, S. A., Shamaan, N. A., Arif, N. M., Koon, G. B., Shukor, M. Y. A. and Syed, M. A. 2012. Enhanced phenol degradation by immobilized *Acinetobacter* sp. Strain AQ5NOL 1. *World Journal of Microbiology and Biotechnology*, 2(2): 347–352.
- Al-Thani, R. F., Abd-El-Haleem, D. A. M. and Al-Shammri, M. 2007. Isolation, biochemical and molecular characterization of 2-chlorophenol-degrading *Bacillus* isolates. *African Journal of Biotechnology*, 6(23): 2675–2681.
- Anderson, G. L., Love, M. and Zeider, B. K. 2003. Metabolic energy from arsenite oxidation in *Alcaligenes faecalis*. *Journal of Physics IV France*, 107: 49-52.
- Annadurai, G., Juang, R. S. and Lee, D. J. 2002. Microbiological degradation of phenol using mixed liquors of *Pseudomonas putida* and activated sludge. *Waste Management*, 22(7): 703–710.
- APHA (American Public Health Association). (1998). Standard methods for the examination of water and wastewater. 20th edition., Washington DC, USA, 540-544.
- Awan, Z. U. R., Shah, A. H. and Amjad, M. 2013. Microbial degradation of phenol by locally isolated soil bacteria. *International Journal of Modern Cellular and Molecular Biology*, 2(1): 1-13.
- Bai, J., Wen, J. P., Li, H. M. and Jiang, Y. 2007. Kinetic modelling of growth and biodegradation of phenol and *m*-cresol using *Alcaligenes faecalis*. *Process Biochemistry*, 42: 510-517.
- Bak, F. and Widdel, F. 1986. Anaerobic degradation of phenol and phenol derivatives by *Desulfobacterium phenolicum* sp. *Microbiology*, 146: 177–180.
- Balamurugan, P., Preetha, B. and Virithagiri, T. 2012. Study on effect of operating parameters on biodegradation of phenol by *Aspergillus fumigatus*. *International Journal of Engineering Research and Applications*, 2(2): 981-986.
- Basha, K. M., Rajendran, A. and Thangavelu, V. 2010. Recent advances in the biodegradation of phenol: a review. *Asian Journal of Experimental Biological Sciences*, 1(2): 219–234.
- Bizet, J. and Bizet, C. 1997. Strains of *Alcaligenes faecalis* from clinical material. *Journal of Infection*, 35(2): 167-169.

- Donkor, E. S., Dayie, N. T. K. D. and Adiku, T. K. 2014. Bioinformatics with basic local alignment tool (BLAST) and fast alignment (FASTA). *Journal of Bioinformatics and Sequence Analysis*, 6(1): 1-6.
- Duan, Z. 2011. Microbial degradation of phenol by activated sludge in a batch reactor. *Environment Protection Engineering*, 37(2): 53-63.
- El-Sayed, W. S., Ibrahim, M. K. and Ouf, S. A. 2014. Molecular characterization of the alpha subunit of multicomponent phenol hydroxylase from 4-chlorophenol-degrading *Pseudomonas* sp. strain PT3. *Journal of Microbiology (Seoul, Korea)*, 52(1): 13–9.
- Jiang, Y., Wen, J., Bai, J., Jia, X. and Hu, Z. 2007. Biodegradation of phenol at high initial concentration by *Alcaligenes faecalis*. *Journal of Hazardous Material*, 147: 672-676.
- King, R. J., Short, K. A. and Seidler, R. J. 1991. Assay for detection and enumeration of genetically engineered microorganisms which based on the activity of a deregulated 2,4-dichlorophenoxyacetate monooxygenase. *Applied and Environmental Microbiology*, 57(6): 1790-1792.
- Krug, M., Ziegler, H. and Straube, G. 1985. Degradation of phenolic compounds by the yeast *Candida tropicalis* HP 15. I. physiology of growth and substrate utilization. *Journal of Basic Microbiology*, 25: 103–110.
- Krastanov, A., Alexieva, Z. and Yemendzhiev, H. 2013. Microbial degradation of phenol and phenolic derivatives. *Engineering in Life Sciences*, 13(1): 76–87.
- Ma, H., Li, G., Fang, P., Zhang, Y. and Xu, D. 2010. Identification of phenol-degrading *Nocardia* sp. strain C-14-1 and characterization of its ring-cleavage 2,3-dioxygenase. *International Journal of Biology*, 2(1): 79–83.
- Mailin, M. and Firdausi, R. 2006. High performance phenol degrading microorganisms isolated from wastewater and oil-contaminated soil. *Malaysian Journal of Microbiology*, 2(2): 32-36.
- Mahiuddin, M., Fakhrudin, A. N. M. and Abdullah-Al-Mahin. 2012. Degradation of phenol via meta cleavage pathway by *Pseudomonas fluorescens* PU1. *International Scholarly Research Network (ISRN) Microbiology*, 1: 1-6.
- Michałowicz, J. and Duda, W. 2007. Phenols – sources and toxicity. *Polish Journal of Environmental Study*, 16(3): 347–362.
- Mohite, B. V, Jalgaonwala, R. E., Pawar, S. and Morankar, A. 2010. Isolation and characterization of phenol degrading bacteria from oil contaminated soil. *Innovative Romanian Food Biotechnology* 7: 61–65.

- Nawawi, N. M., Ahmad, S. A., Shukor, M. Y., Syed, M. A. and Ibrahim, L. 2014. Isolation and characterization of phenol-degrading microorganism: Recent advances. *Journal of Environmental Bioremediation and Toxicology*, 2(1): 11–22.
- Nordlund, I., Powlowski, J., Hagström, A. and Shingler, V. 1993. Conservation of regulatory and structural genes for a multi-component phenol hydroxylase within phenol-catabolizing bacteria that utilize a meta-cleavage pathway. *Journal of General Microbiology*, 139(11): 2695–703.
- Reh fuss, M. and Urban, J. 2005. *Alcaligenes faecalis* subsp. *phenolicus* subsp. nov. a phenol-degrading, denitrifying bacterium isolated from a graywater bioprocessor. *Systematic and Applied Microbiology*, 28: 421-429.
- Shingler, V., Franklin, F.C.H., Tsuda, M., Holroyd, D. and Bagdasarian, M. 1989. Molecular analysis of a plasmid-encoded phenol hydroxylase from *Pseudomonas* CF600. *Journal of General Microbiology*, 135: 1083–1092.
- Sridevi, V., Lakshmi, M. V. V. C., Manasa, M. and Sravani, M. 2012. Metabolic pathways for the biodegradation of phenol. *International Journal of Engineering Science and Advanced Technology*, 2(3): 695–705.
- Usman, A. D. and Ali, S. 2009. An overview on the application of Polymerase Chain Reaction (PCR) in the diagnosis of bacterial infections. *Bayero Journal of Pure and Applied Sciences*, 2(1): 109-114.
- Valones, M. A. A., Guimarães, R. L., Brandão, L. A. C., De Souza, P. R. E., Carvalho, A. A. T. and Crovela, S. 2009. Principles and applications of Polymerase Chain Reaction in medical diagnostic fields: a review. *Brazilian Journal of Microbiology*, 40: 1-11.
- Zhang, X., Gao, P., Chao, Q., Wang, L., Senior, E. and Zhao, L. 2004. Microdiversity of phenol hydroxylase genes among phenol-degrading isolates of *Alcaligenes* sp. from an activated sludge system. *FEMS Microbiology Letters*, 237(2): 369–75.
- Zhu, C., Zhang, L. and Zhao, L. 2008. Molecular cloning, genetic organization of gene cluster encoding phenol hydroxylase and catechol 2,3-dioxygenase in *Alcaligenes faecalis* IS-46. *World Journal of Microbiology and Biotechnology*, 24: 1687-1695.