

# **UNIVERSITI PUTRA MALAYSIA**

# THE ASSESSMENT OF CHOLINESTERASE FROM GILLS OF ANABUS TESTUDINEUS AS DETECTION OF METAL IONS

LOW WEINI

FBSB 2015 59

### THE ASSESSMENT OF CHOLINESTERASE FROM GILLS OF ANABUS TESTUDINEUS AS DETECTION OF METAL IONS



LOW WEINI

165125

DEPARTMENT OF BIOCHEMISTRY FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES UNIVERSITI PUTRA MALAYSIA

### THE ASSESSMENT OF CHOLINESTERASE FROM GILLS OF ANABUS TESTUDINEUS AS DETECTION OF METAL IONS



LOW WEINI

165125

Thesis Submitted in Partial Fulfillment of the Requirement For the course BCH 4999 (Project) in the Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia

June 2015

#### PENGESAHAN

Dengan ini adalah disahkan bahawa tesis projek yang bertajuk "The Assessment of Cholinesterase from Gills of *Anabus testudineus* as Detection of Metal Ions" telah disiapkan serta dikemukakan kepada Jabatan Biokimia oleh Low Weini (165125) sebagai syarat untuk kursus BCH4999 Projek.

Disahkan oleh,

.....



Tarikh : .....

(Dr. Siti Aqlima Ahmad) Penyelia projek Jabatan Biokimia Fakulti Bioteknologi dan Sains Biomolekul Universiti Putra Malaysia

Tarikh : .....

(Prof. Dato' Dr. Abu Bakar Salleh) Ketua Jabatan Biokimia Fakulti Bioteknologi dan Sains Biomolekul Universiti Putra Malaysia

#### ABSTRACT

Cholinesterase from the gill of *Anabus testudineus* contains mostly AChE. It was partially purified by ammonium sulphate precipitation and DEAE cellulose using ion exchange chromatography with 4.890U of specific activity, 11.7 of purification fold and 3.0% yield. Optimum pH for AChE in gill of *A. testudineus* is 8 using Tris-HCl buffer at temperature 25°C. The optimum acetylthiocholine iodide concentration is 2.5 mM with  $V_{max}$  2.533 and  $K_m$  0.8802 in which the catalytic efficiency of 2.88. For metal ion inhibition, 10 metal ions were tested and AChE in gill of *Anabus testudineus* showed a critically low enzyme activity towards mercury in which the activity is inhibited by 99.05%. However, cobalt and silver had no inhibitory effect on AChE. From IC<sub>50</sub>, only 0.0123 ppm of Hg was required to reduce the enzyme activity by half.

#### ABSTRAK

Kolinesteres dari insang Anabus testudineus mengandungi asetilkolinesteres (AChE). Ia ditulenkan secara separa melalui aplikasi kerpasan ammonium sulfat dan kromatografi pertukaran ion dengan menggunakan selulosa DEAE sebagai matriks dengan aktiviti spesifik bernilai 4.89U, faktor penulenan bernilai 11.7 dan hasil perolehan sebanyak 3.0%. pH optima untuk AChE dari insang Anabus testudineus ialah 8 dengan menggunakan penimbal Tris-HCl pada suhu 25°C. Optima kepekatan substrat asetilkolinesteres iodide ialah 2.5 mM dengan nilai V<sub>max</sub> sebanyak 2.533 dan nilai K<sub>m</sub> sebanyak 0.8802 yang menunjukkan kecekapan pemangkin sebanyak 2.88. Sebayak 10 jenis ion logam berat telah diuji dengan AChE dari insang *Anabus testudineus* dan ia menunjukkan aktiviti enzim yang rendah apabila diuji dengan mekuri di mana aktiviti enzim telah direncat 99.05%. Walau bagaimanapun, kobalt dan perak tidak memberi kesan negatif kepada aktiviti enzim. Dari IC50, hanya 0.0123 ppm merkuri diperlukan untuk menurunkan aktiviti enzim ke 50%.

#### ACKNOWLEDGEMENT

Firstly, I would like to express my greatest gratitude to several people who contributed and helped me throughout this project directly and indirectly, especially Dr. Siti Aqlima Ahamd as the supervisor of this project, together with her postgraduate students namely Mr. Mohd Khalizan Sabullah and Ms. Sabrina Hayat who have taught and supervised me to complete this project. Not to forget my beloved laboratory mates especially Mr. Wong Yoong Fei who assisted me a lot in conducting the project and thanks for the free car ride.

I would also like to express my grateful thanks to my parents Mr. Low Boon San and Mrs. Low Ah Bee who always support and motivate me by showering me with love and care so that I can focus on my project. Without them, I will not achieve what I am today.

Last but not least, I would like to thank Mr. Wong Ming Hong who always encourages me and never fail to cheer me up when I was in one of those days where I just couldn't get the idea to write for the thesis. Thank you for assisting me to complete the thesis indirectly.

"There's nothing you can't do, it's all about the willingness."

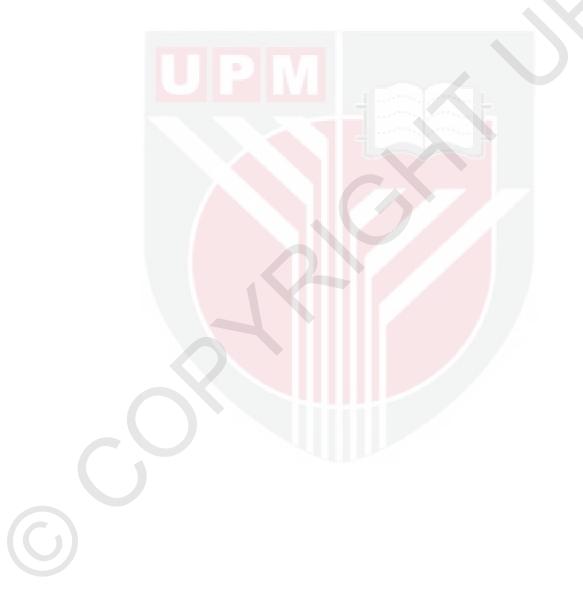
Low Weini, 2015

## TABLE OF CONTENTS

Page

ABS ABS ACK TAB LIST LIST	GESAHAN FRACT FRAK NOWLEDGEMENTS LE OF CONTENTS OF TABLES OF FIGURES OF FIGURES TOF ABBREVIATIONS	i iii iv v vi vii viii
1.0	INTRODUCTION	1
2.0	LITERATURE REVIEW         2.1 Cholinesterase         2.1.1 Acetylcholinesterase         2.2 Anabus testudineus         2.3 Metal Ions         2.3.1 Arsenic         2.3.2 Cadmium         2.3.3 Copper         2.3.4 Mercury         2.3.5 Other Metal Ions	3 4 5 6 7 7 7 7 8
3.0	MATERIALS AND METHODS	
	<ul> <li>3.1 Materials <ul> <li>3.1.1 Chemicals</li> <li>3.1.2 Equipments</li> <li>3.1.3 Specimen</li> </ul> </li> <li>3.3 Methodology <ul> <li>3.3.1 Extraction and purification</li> <li>3.3.2 Cholinesterase activity and protein content</li> <li>3.3.3 Substrate specificity</li> <li>3.3.4 pH and temperature profile</li> <li>3.3.5 The effect of metal ion</li> <li>3.3.6 Statistical analysis</li> </ul> </li> </ul>	10 11 12 12 12 13 14 14 14 15
4.0	<ul> <li>RESULTS AND DISCUSSIONS</li> <li>4.1 After extraction</li> <li>4.2 After ammonium sulphate precipitation</li> <li>4.3 After ion exchange chromatography <ul> <li>4.3.1 Purification table</li> </ul> </li> <li>4.4 Native PAGE</li> <li>4.5 Optimisation profile <ul> <li>4.5.1 Specific substrate concentration optimisation</li> <li>4.5.2 pH optimisation</li> </ul> </li> </ul>	16 17 18 19 20 23 24

	4.6 4.7	<ul><li>4.5.3 Temperature optimisation</li><li>Metal ion inhibition</li><li>Half maximal inhibitory concentration</li></ul>	26 27 29
5.0	CO	NCLUSION	30
	EREN ENDI		31 35



LIST	OF	<b>TABLES</b>
------	----	---------------

Table		Page
1	List of Chemicals	10
2	List of Equipments	11
3	Purification table	20
4	Table of kinetic study of enzyme activity	24
5	Summary of optimum conditions LIST OF FIGURES	30
<b>Figure</b> 1	Anabus testudineus bought from Pasar Borong Selangor.	<b>Page</b> 12
2	The graph of enzyme activity of the crude extract when three different substrate were used, namely acetylthiocholine iodide, butyrylthiocholine iodide and propionylthiocholine iodide.	16
3	The enzyme activity of the crude extract after ammonium sulphate precipitation by fractions ranging from 0% to 80%.	17
4	Enzyme activity with respective to protein amount obtained from Ellman method using acetylthiocholine iodide as substrate.	19
5	Figure above shows the bands of cholinesterase of gill obtained by Native PAGE.	21
6	The $log_{10}$ graph for broad range of protein molecular weight.	22
7	Substrate concentration optimisation using 3 different substrate, i.e acetylthiocholine iodide, butyrylthiocholine iodide and propynylthiocholine iodide at different concentration, ranging from 0.1 mM to 2.5 mM.	23
8	pH optimisation using 3 different buffers.	25
9	Temperature optimistation using ATC as substrate and Tris- HCl at pH 8.	26
10	Metal ion inhibition profile using 10 different metal ions at 10ppm to study the effect on cholinesterase.	28
11	$IC_{50}$ using mercury with different concentration, ranging from 0.01 ppm to 10 ppm.	29

6

## LIST OF ABBREVIATIONS

°C	Degree Celsius
%	Percent
AChE	Acetylcholinesterase
ACh	Acetylcholine
ATC	Acetylthiocholine iodide
BuChE	Butyrylcholinesterase
BTC	Butyrylthiocholine iodide
ChE	Cholinesterase
DTNB	5,5'-dithiobis(2-nitrobenzoic acid)
DDT	Dichlorodiphenyltrichloroethane
HCl	Hydrochloric acid
IC <sub>50</sub>	Half maximal inhibitory concentration
kDa	Kilo Dalton
Km	Michaelis-Menten constant
М	Molar
mg	Milligram
mM	Millimolar
μΙ	Microlitre
nm	Nanometre
NaOH	Sodium hydroxide
рН	-log concentration of H+ ion
РТС	Propionylthiocholine iodide
PAGE	Polyacrylamide gel electrophoresis
SDS	Sodium dodecyl sulphate
TEMED	N,N,N',N'-tetramethyl ethylenediamine
Vmax	Maximum velocity
Zn	Zinc
Pb	Lead
Ni	Nickel
Hg	Mercury
Co	Cobalt

Cu	Copper
Cr	Chromium
Cd	Cadmium
As	Arsenic
Ag	Silver



#### **CHAPTER 1**

#### **INTRODUCTION**

Nowadays, metal pollution of aquatic environment is a rising issue in Malaysia. There are numbers of contributors that account for metal pollution, for example, oil and gas industries, shipping and cargo, and agricultural activities. However, the major cause of pollution is due to the manufacturing sectors, for instance, metal manufacturer such as electroplating, etching and etc which contains a high level of Cadmium, Copper, and Zinc, which is harmful to human (Shazili *et al.*, 2007).

In the context of aquatic environment pollution, fish are the most affected and can be considered as the biomarker to measure the level of water pollution (Law and Singh, 1991). Other marine organisms such as microflora and algae have the ability to accumulate metal ions in their cells and as a result, the small fish that these microflora and algae will be enriched with these accumulated metal substances and consequently, human will be affected through the bioaccumulation occurs in food web (Jaffar *et al.*, 1998; Ashraf *et al.*, 2012). Fish is known as a good biomarker to determine the level of water pollution because it is easily reachable, able to corporate metals and has an ideal size to be sampled (Batvari *et al*, 2007).

Anabas testudineus, commonly known as climbing perch, is a kind of freshwater fish that can survive in the water with very low oxygen level and polluted water. It can also survive without water for 6 to 10 hours (Hughes *et al.*, 1970). Due to its ability to survive in polluted aquatic environment, it is suitable to be used as a biomarker to indicate the status of pollution in aquatic environment.

In the context of cholinesterase, it is commonly used in biomarker for determination of contamination due to its effectiveness, especially by pesticides and insecticides. Cholinesterase functions as detoxifying enzyme in human. Thus, the inhibition of cholinesterase is very crucial to be studied. Therefore, the objectives of this project are:

- 1. To extract and purify cholinesterase from the gill of Anabas Testudineus.
- 2. To identify the optimum temperature and pH for cholinesterase activity.
- 3. To determine the in-vitro effect of cholinesterase activity towards heavy metals exposure.



#### REFERENCES

- Andreescu, S., & Marty, J. L. 2006. Twenty years research in cholinesterase biosensors: from basic research to practical applications. *Biomolecular engineering*, 23(1), 1-15.
- Ashraf, M. A., Maah, M.J., Yusoff I. 2012. Bioaccumulation of Heavy Metals in Fish Species Collected from Former Tin Mining Catchment. *International Journal Environment Resources*, 6(1): 209-218.
- Batvari, B. P. D., Kamala-Kannan, S., Shanthi, K., Krishnamoorthy, R., Lee, K. J. and Jayaprakash, M. 2007. Heavy metals in two fish species (*Carangoidel malabaricus* and *Belone stronglurus*) from Pulicat Lake, North of Chennai, Southeast Coast of India. *Environment Monitoring Assessment*, 145(1-3): 167-175.
- Binoy, V. V., & Thomas, K. J. 2004. The climbing perch (Anabas testudineus Bloch), a freshwater fish, prefers larger unfamiliar shoals to smaller familiar shoals. *Current Science*, 86(1), 207-211.
- Bulbring, E., Burn, J. H., & Shelley, H. J. 1953. Acetylcholine and ciliary movement in the gill plates of Mytilus edulis. *Proceedings of the Royal Society of London. Series B-Biological Sciences*, 141(905), 445-466.
- Carroll, R. T., Grimm, J. L., Hepburn, T. W., & Emmerling, M. R. 1995. Purification of acetylcholinesterase by tacrine affinity chromatography. *Protein expression and purification*, 6(4), 389-393.
- Dass, P., Mejia, M., Landes, M., Jones, R., Stuart, B., & Thyssen, J. 1994. Cholinesterase: review of methods. *Clinical Chemistry*, 34, 135-157.
- Duong-Ly KC and Gabelli S. 2014. Salting out of proteins using ammonium sulphate precipitation. Methods in Enzymology, 541, 85-94.
- Ebrahimpour, M., & Mushrifah, I. 2008. Heavy metal concentrations in water and sediments in Tasik Chini, a freshwater lake, Malaysia. *Environmental monitoring and assessment*, 141(1-3), 297-307.
- Ellman, G. L., Courtney, K. D., Andres, V., & Featherstone, R. M. 1961. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical pharmacology*, 7(2), 88-95.
- Elumalai, M., Antunes, C., & Guilhermino, L. 2007. Enzymatic biomarkers in the crab Carcinus maenas from the Minho River estuary (NW Portugal) exposed to zinc and mercury. *Chemosphere*, *66*(7), 1249-1255.
- Escartín, E., & Porte, C. 1997. The use of cholinesterase and carboxylesterase activities from Mytilus galloprovincialis in pollution monitoring. *Environmental Toxicology and Chemistry*, *16*(10), 2090-2095.

- Fleming, W. R., Scheffel, K. G., & Linton, J. R. 1962. Studies on the gill cholinesterase activity of several cyprinodontid fishes. *Comparative biochemistry and physiology*, 6(3), 205-213.
- Forget, J., Livet, S., & Leboulenger, F. 2002. Partial purification and characterization of acetylcholinesterase (AChE) from the estuarine copepod Eurytemora affinis (Poppe). *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 132(1), 85-92.
- Frasco, M. F., Fournier, D., Carvalho, F., & Guilhermino, L. 2008. Does mercury interact with the inhibitory effect of dichlorvos on Palaemon serratus (Crustacea: Decapoda) cholinesterase?. Science of the total environment,404(1), 88-93.
- Fulton, M. H., & Key, P. B. 2001. Acetylcholinesterase inhibition in estuarine fish and invertebrates as an indicator of organophosphorus insecticide exposure and effects. *Environmental Toxicology and Chemistry*, 20(1), 37-45.
- Galgani, F., & Bocquené, G. 2000. Molecular biomarkers of exposure of marine organisms to organophosphorus pesticides and carbamates. Use of Biomarkers for Environmental Quality Assessment. Elsevier Science Publisher, 113-137.
- Gill, T. S., Tewari, H., & Pande, J. 1990. Use of the fish enzyme system in monitoring water quality: effects of mercury on tissue enzymes. *Comparative Biochemistry and Physiology Part C: Comparative Pharmacology*, 97(2), 287-292.
- Graham, J. B. (Ed.). 1997. Air-breathing fishes: evolution, diversity, and adaptation. Academic Press.
- Green, A. A., & Hughes, W. L. 1955. Protein fractionation on the basis of solubility in aqueous solutions of salts and organic solvents. *Methods in enzymology*, 1, 67-90.
- Holden, A. V. 1973. Mercury in fish and shellfish A review. *International Journal* of Food Science and Technology, 8(1), 1-25.
- Hughes ,G.M and Singh , B.N 1970. Respiration in an air breathing fish, the climbing perch, Anabas testudineus (Bloch) I .Oxygen uptake and carbon dioxide release in to air and water. *J Expt Biology*, 53: 265-280.
- Jaffar, M., Ashraf, M. and Rasool, A. 1988. Heavy metal contents in some selected local freshwater fish and relevant waters. *Pak. J. Sci. Ind. Res.*, 31 (3): 189–193.
- Law, A. T., & Singh, A. 1991. Relationships between heavy metal content and body weight of fish from the Kelang estuary, Malaysia. *Marine Pollution Bulletin*,22(2), 86-89.

- Leticia, A. G., & Gerardo, G. B. 2008. Determination of esterase activity and characterization of cholinesterases in the reef fish Haemulon plumieri. *Ecotoxicology and environmental safety*, 71(3), 787-797.
- Mora, P., Michel, X., & Narbonne, J. F. 1999. Cholinesterase activity as potential biomarker in two bivalves. *Environmental Toxicology and Pharmacology*, 7(4), 253-260.
- Rao, J. V., Shilpanjali, D., Kavitha, P., & Madhavendra, S. S. 2003. Toxic effects of profenofos on tissue acetylcholinesterase and gill morphology in a euryhaline fish, Oreochromis mossambicus. *Archives of toxicology*, 77(4), 227-232.
- Romani, R., Corsi, I., Bonacci, S., Focardi, S., De Medio, G. E., De Santis, A., ... & Rosi, G. 2006. Organophosphate-resistant forms of acetylcholinesterases in two scallops—the Antarctic Adamussium colbecki and the Mediterranean Pecten jacobaeus. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 145(2), 188-196.
- Scopes, R. K. 1994. Protein purification: principles and practice. Springer Science & Business Media.
- Shazili, N. A. M., Yunus, K., Ahmad, A. S., Abdullah, N., & Rashid, M. K. A. 2006. Heavy metal pollution status in the Malaysian aquatic environment. Aquatic Ecosystem Health & Management, 9(2), 137-145.
- Stefano, B., Ilaria, C., & Silvano, F. 2008. Cholinesterase activities in the scallop Pecten jacobaeus: Characterization and effects of exposure to aquatic contaminants. *Science of the Total Environment*, 392(1), 99-109.
- Suresh A, Sivaramakrishna B, Victoriamma P.C. and Radhakrishnaiah K. 1992. Comparative study on the inhibition of acetylcholinesterase activity in the freshwater fish *Cyprinus carpio* by mercury and zinc. *Biochemistry International*, 26(2): 367-375.
- Talesa, V., Romani, R., Antognelli, C., Giovannini, E., & Rosi, G. 2002. Different expressions of organophosphate-resistant acetylcholinesterases in the bivalve mollusk Scapharca inaequivalvis living in three different habitats. *Environmental toxicology and chemistry*, 21(1), 102-108.
- Tecles, F., & Ceron, J. J. 2001. Determination of whole blood cholinesterase in different animal species using specific substrates. *Research in Veterinary Science*, 70(3), 233-238.
- Tiller, K. G. 1992. Urban soil contamination in Australia. *Soil Research*, 30(6), 937-957.
- Toutant, J. P., Massoulié, J., & Bon, S. 1985. Polymorphism of pseudocholinesterase in Torpedo marmorata tissues: comparative study of the catalytic and molecular properties of this enzyme with acetylcholinesterase. *Journal of neurochemistry*, 44(2), 580-592.

- Tulasi, S. J., Reddy, P. U. M., & Ramana Rao, J. V. 1989. Effects of lead on the spawning potential of the fresh water fish, Anabas testudineus. *Bulletin of environmental contamination and toxicology*, 43(6), 858-863.
- Vieira, L. R., Gravato, C., Soares, A. M. V. M., Morgado, F., & Guilhermino, L. 2009. Acute effects of copper and mercury on the estuarine fish Pomatoschistus microps: linking biomarkers to behaviour. *Chemosphere*,76(10), 1416-1427.
- Voegborlo, R. B., El-Methnani, A. M., & Abedin, M. Z. 1999. Mercury, cadmium and lead content of canned tuna fish. *Food Chemistry*, 67(4), 341-345.
- Wittig, I., Braun, H. P., & Schägger, H. 2006. Blue native PAGE. Nature Protocols, 1, 418-428.
- Zarcinas, B. A., Ishak, C. F., McLaughlin, M. J., & Cozens, G. 2004. Heavy metals in soils and crops in Southeast Asia. *Environmental Geochemistry* and Health, 26(4), 343-357.