



**UNIVERSITI PUTRA MALAYSIA**

***EXPERIMENTAL INTRAOCULAR INFECTION OF JAPANESE QUAILS  
(*Coturnix coturnix japonica*) WITH INFECTIOUS BURSAL DISEASE  
VIRUS (IBDV) INTERMEDIATE STRAIN***

**SITI NOR AZIZAH BINTI MAHAMUD**

**FPV 2016 59**

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(IBDV) INTERMEDIATE STRAIN**

**SITI NOR AZIZAH BINTI MAHAMUD**

A Project paper submitted to the  
Faculty of Veterinary Medicine, Universiti Putra Malaysia  
In partial fulfilment of the requirement for the  
DEGREE OF DOCTOR OF VETERINARY MEDICINE  
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## CERTIFICATION

It is hereby certified that we have read this project paper entitled “Experimental Intraocular Infection of Japanese Quails (*Coturnix coturnix japonica*) with Infectious Bursal Disease (IBDV) Intermediate strain”, by Siti Nor Azizah binti Mahamud and in our opinion it is satisfactory in terms of scope, quality and presentation as partial fulfilment of the requirement for the course VPD4999 – Project.

---

**DR. MOHD HEZMEE MOHD NOOR**

**DVM (UPM), PhD (QUEENSLAND),**

Senior Lecturer,

Faculty of Veterinary Medicine,

Universiti Putra Malaysia

(Supervisor)

---

**PROF.DR.ABDUL RAHMAN OMAR**

**DVM (UPM), PhD (CORNELL),**

Professor

Faculty of Veterinary Medicine,

Universiti Putra Malaysia

(Co-Supervisor)

---

**DR.LOKMAN HAKIM IDRIS**

**DVM (UPM), PhD (UPM),**

Senior Lecturer,

Faculty of Veterinary Medicine,

Universiti Putra Malaysia

(Co-Supervisor)

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## DEDICATION

This project paper is dedicated to Allah SWT, who had created me and made all things possible,

To my family,

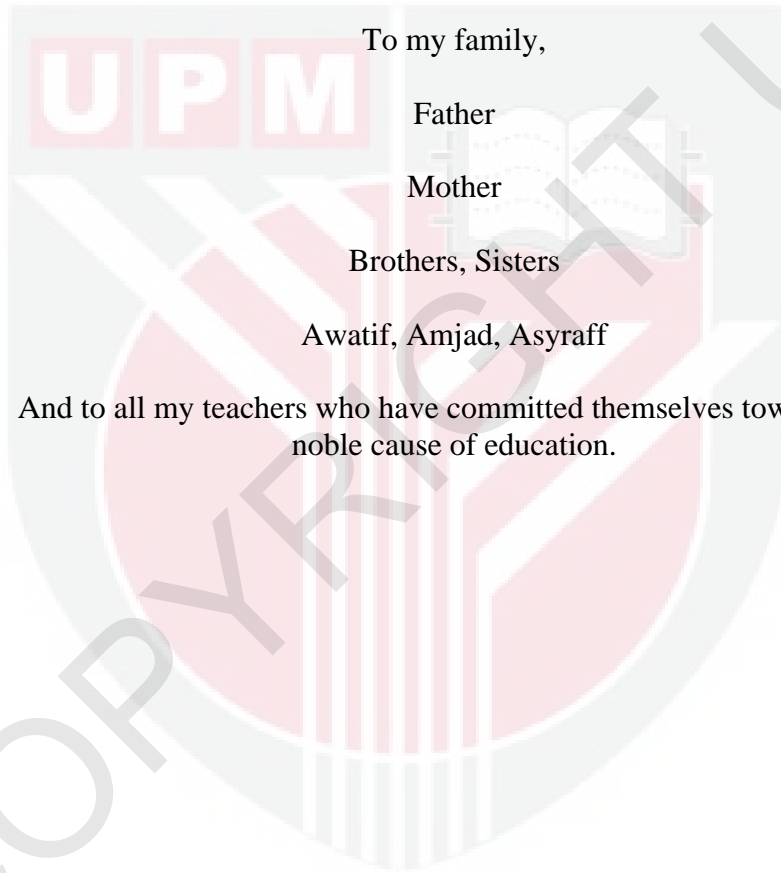
Father

Mother

Brothers, Sisters

Awatif, Amjad, Asyraff

And to all my teachers who have committed themselves towards the noble cause of education.



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## LIST OF ABBREVIATION

%	Percentage
$^{\circ}\text{C}$	Degree celcius
$\mu\text{l}$	Micrometer
ml	Mililiter
H&E	Haematoxylin and Eosin
IACUC	Institutional Animal Care and Use Committee
IBD	Infectious bursal disease
IBDV	Infectious bursal disease virus
MLV	Modified-live vaccine
PBS	Phosphate buffered saline
RNA	Ribonucleic acid
rpm	Rotation per minute
RT-PCR	Reverse transcriptase polymerase chain reaction
SD	Standard deviation
$\text{TCID}_{50}$	Median tissue culture infective dose
VP	Viral protein

## ABSTRAK

Abstrak daripada kertas projek yang dikemukakan kepada Fakulti Perubatan Veterinar untuk memenuhi sebahagian daripada keperluan kursus VPD 4999 – Projek

INFEKSI PERCUBAAN INTRAOKULAR KEPADA PUYUH JEPUN (*Coturnix coturnix japonica*) MENGGUNAKAN VIRUS PENYAKIT BERJANGKIT BURSAL (IBDV) STRAIN PERANTARAAN.

Oleh

Siti Nor Azizah binti Mahamud

2016

Penyelia: Dr. Mohd Hezmee bin Mohd Noor

Penyelia bersama: Prof. Dr. Abdul Rahman bin Omar

Dr. Lokman Hakim bin Idris

Penemuan pertama penyakit berjangkit bursal (IBD) ialah pada tahun 1957 di Gumboro, Delaware, USA dan pertama kali diterangkan di Malaysia pada tahun 1991. IBD merupakan penyakit bawaan virus yang membawa kerugian ekonomi yang besar kepada industri poltri kerana menyebabkan imunitas dan kadar kematian yang tinggi. Eksperimen ini dijalankan bertujuan mempercepatkan jangkitan kepada puyuh jepun menggunakan vaksin hidup dilemahkan strain perantara IBD

secara intraokular. Parameter yang diperolehi adalah pemerhatian tanda klinikal, lesi post-mortem, pengesanan antigen menggunakan transcriptase membalik reaksi berantai polimerase konvensional (RT-PCR) dan perubahan histopatologikal pada puyuh dari group A, B dan C. Spesifik primer telah direka untuk mensasarkan protein major luar kapsid iaitu gen protein virus 2 (VP2). Parameter ini diukur selepas vaksin diberikan dan bilangan puyuh tertentu dikorbankan pada hari ke 5, 9, dan 15 pos-infeksi dan 24 bursa telah dikumpulkan. Hasil eksperimen menunjukkan pengurangan limfoid minor pada kumpulan A, tanda klinikal yang jelas dan pengurangan limfoid sederhana pada kumpulan B dan tiada lesi post-mortem yang ketara pada semua kumpulan. Analisis transcriptase membalik reaksi berantai polimerase menunjukkan hasil negatif bagi semua sampel yang diuji. Kesimpulannya, ujian klinikal, patalogikal dan molekular menunjukkan IBDV strain perantaraan tidak menghasilkan immun respons yang cukup pada puyuh untuk menjadikan mereka pembawa atau perumah.

Kata Kunci: IBDV, intraokular, puyuh jepun (*Coturnix coturnix japonica*), RT-PCR, histopathologi.

ABSTRACT

An abstract of the project paper presented to the Faculty of Veterinary Medicine in partial requirement for the course VPD 4999 – Project

EXPERIMENTAL INTRAOCULAR INFECTIONS OF JAPANESE QUAILS  
(*Coturnix coturnix japonica*) WITH INFECTIOUS BURSAL DISEASE VIRUS  
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By

Siti Nor Azizah binti Mahamud

2016

Supervisor: Dr. Mohd Hezmee bin Mohd Noor

Co-supervisor: Prof. Dr. Abdul Rahman bin Omar

Dr. Lokman Hakim bin Idris

© Infectious Bursal Disease (IBD) was first discovered in 1957 at Gumboro, Delaware, USA and was first described in Malaysia in 1991. IBD became an important viral disease in poultry industry due to its significant economic losses with high mortality and profound immunosuppression. This experiment was conducted to induce the Japanese quails with IBD modified-live vaccine intermediate strain intraocularly.

The parameters obtained were observation of clinical signs, post-mortem lesions, antigen detection using conventional reverse transcriptase PCR and histopathological changes in quails from group A, B and C respectively. Specific primer was designed to target the major outer capsid protein which is viral protein 2 gene (VP2). These parameters are measured after vaccine administration and selected number of quails from each group were euthanized at day 5, 9 and 15 post-infection and the total of 24 bursas were collected. The result reveals minor lymphoid depletion in Group A, prominent clinical signs and mild lymphoid depletion for Group B and no significant post-mortem findings in all groups. RT-PCR analysis gave negative findings in all samples tested. In conclusion, clinical, pathological and molecular results indicate that IBDV intermediate strain does not produce sufficient immune response in quails to warrant them as carrier or host.

Keywords: IBDV, intraocular, Japanese quails (*Coturnix coturnix japonica*), RT-PCR, histopathology.

## 1.0 INTRODUCTION

Infectious Bursal Disease is often referred to as Gumboro disease, was discovered in 1957 in Gumboro, Delaware, USA (Khan *et al.*, 2005). The outbreak of IBD was first described in Malaysia in 1991 by Hair-Bejo (1992). According to Washington Disease Diagnostic Laboratory (2015), the natural host of IBDV is the domestic fowl including chickens and turkeys and young chickens within the age of 3 to 6 weeks are the most susceptible to clinical diseases. The wild birds such as healthy ducks, guinea fowl, quail and pheasants, have been found to be naturally infected with IBDV (Washington Disease Diagnostic Laboratory, 2015). The most recent survey of international poultry specialists, conducted by *World Poultry*, highlighted continuing concern in the sector over the sanitary status of poultry. Gumboro diseases topped the list of the most serious poultry diseases (Tsukamoto *et al.*, 1999).

IBD is a highly contagious viral disease that affects mainly young chickens and is economically important to the poultry industry (Van den Berg, 2000) due to significant economic losses as it lead to high mortality and morbidity, impaired growth and profound immunosuppression. According to IDERIS (1999), IBDV causes severe inflammation of the bursa of Fabricius that leads to immunosuppression due to destruction of immature B-lymphocytes within the bursa of Fabricius and finally leads to lymphoid depletion and significant depression of the humoral antibody response.

The indirect economic impact of the disease is also considerable, due to virus-induced immunosuppression and potential interactions between IBDV and other viruses, bacteria or parasites. These indirect losses are due to secondary infections, growth retardation and condemnation of carcasses at the slaughterhouse (Van den Berg, 2000). Moreover, the increased use of antibiotics against secondary infections constitutes a growing public health concern. The birds infected with IBD became susceptible to get infection of other diseases that can result in an increase of occurrence of disease caused by opportunistic pathogens and prevents young chickens from responding optimally to vaccination (Ojeda *et al.*, 1997).

The objectives of this study are:

- 1) To observe the clinical signs shown by challenged quails.
- 2) To examine the post-mortem lesions on challenged quails.
- 3) To examine the histopathological changes on bursa of Fabricius of challenged quails.
- 4) To demonstrate the presence of antigen of IBDV in challenged quails via molecular method which is RT-PCR.

Therefore, the hypotheses for this study are:

- 1) There are significant clinical signs shown by challenged quails.
- 2) There are significant post-mortem lesion on challenged quails.
- 3) There are significant histopathological changes on challenged quails.
- 4) There are presence of antigen in challenged quails upon tested with RT-PCR.



## 8.0. REFERENCES

1. Abao, E. S., Barro, J. R. D., Gonato, R. P. L., Simpao, K. C., & Ybañez, A. P. Negative Sero-occurrence of Infectious Bursal Disease, Newcastle Disease and Infectious Bronchitis in Japanese Quail.
2. Access RT-PCR System quick protocol. (2000-2009). Promega technical bulletin # TB2200. 27<sup>th</sup> January 2016. Retrieved from: [www.promega.com](http://www.promega.com).
3. Akter S.H., Khan M.Z.I., Jahan M.R., Karim M.R. and Islam M.R. (2006). Histomorphological study of the lymphoid tissues of broiler chickens. *Bangl. J. Vet. Med.* 4 (2): 87–92.
4. Ashraf, S. (2005). *Studies on infectious bursal disease virus* (Doctoral dissertation, The Ohio State University).
5. Bayliss C.D., Spies U., Shaw K., Peters R.W., Papageorgiou A., Muller H. & Boursnell M.E.G. (1990). A comparison of the sequences of segment A of four infectious bursal disease virus strains and identification of a variable region in VP2.J. *gen. Virol.*, 71, 1303-1312.
6. Bolis, D. A., Paganini, F. J., Simon, V. A., Zuanaze, M. F., Scanavini Neto, H., Correa, A. R. A., & Ito, N. M. K. (2003). Gumboro disease: evaluation of serological and anatomopathological responses in vaccinated broiler chickens challenged with very virulent virus strain. *Revista Brasileira de Ciência Avícola*, 5(2), 137-146.
7. Cardoso, T. C., Rosa, A. C., Astolphi, R. D., Vincente, R. M., Novais, J. B., Hirata, K. Y., & Luvizotto, M. C. R. (2008). Direct detection of infectious bursal disease virus from clinical samples by in situ reverse transcriptase-linked polymerase chain reaction. *Avian Pathology*, 37(4), 457-461.

8. Chin, P. H. (1993). List of approved animal vaccines and biological importation, sales and use in West Malaysia. *First education. Veterinary Association, Malaysia.*
9. Dobos, P., Hill, B. J., Hallett, R., Kells, D. T., Becht, H., & Teninges, D. (1979). Biophysical and biochemical characterization of five animal viruses with bisegmented double-stranded RNA genomes. *Journal of Virology*, 32(2), 593-605.
10. Eterradosi N., Toquin D., Rivallan G. & Guittet M. (1997). Modified activity of a VP2-located neutralizing epitope on various vaccine, pathogenic and hypervirulent strains of infectious bursal disease vims. *Arch. Virol.*, 142, 255-270.
11. Faragher J.T. (1972). - Infectious bursal disease of chicken. *Vet. Bull*, 42, 361-369.
12. Khan, C. M. (2005). The Merck Veterinary Manual 9th ed. Merck and C. Inc. NJ. USA, 2125-2136.
13. Hair-Bejo, M. (1992). An outbreak of infectious bursal disease in broilers. *J. Vet. Malaysia*, 4(168), 124-128.
14. Hair-Bejo, M., Salina, S., Hafiza, H., & Julaida, S. (2000). In ovo vaccination against infectious bursal disease in broiler chickens. *J. Vet.-Malaysia*, 2, 63-69.
15. Hair-Bejo, M., Ng, M. K., & Ng, H. Y. (2004). Day old vaccination against infectious bursal disease in broiler chickens. *International Journal of Poultry Science*, 3(2), 124-128.
16. Hoque, M. M., Omar, A. R., Hair-Bejo, M., & Aini, I. (2002). Sequence and phylogenetic analysis of the VP2 gene of very virulent infectious bursal disease virus isolates. *Journal of biochemistry, molecular biology, and biophysics: JBMBB: the official journal of the Federation of Asian and Oceanian Biochemists and Molecular Biologists (FAOBMB)*, 6(2), 93-99.
17. IDERIS, D. A. (1999). Faculty Veterinary Medicine Universiti Putra Malaysia.

18. Infectious Bursal Disease. College of veterinary medicine. Washington Animal Disease Diagnostic Lab. 10<sup>th</sup> March 2016. Retrieved from: <http://waddl.vetmed.wsu.edu/animal-disease-faq/infectious-bursal-disease>
19. Jackwood D.J. (1990). Development and characterization of nucleic acid probes to infectious bursal disease viruses. *Vet. Microbiol*, 24, 253-260.
20. Lee, L. H., Ting, L. J., Shien, J. H., & Shieh, H. K. (1994). Single-tube, noninterrupted reverse transcription-PCR for detection of infectious bursal disease virus. *Journal of clinical microbiology*, 32(5), 1268-1272.
21. Luengo, A., Butcher, G., Kozuka, Y., & Miles, R. (2001). Histopathology and transmission electron microscopy of the bursa of Fabricius following IBD vaccination and IBD virus challenge in chickens. *Revista Científica*, 11(6).
22. Luque, D., Rivas, G., Alfonso, C., Carrascosa, J. L., Rodríguez, J. F., & Castón, J. R. (2009). Infectious bursal disease virus is an icosahedral polypliod dsRNA virus. *Proceedings of the National Academy of Sciences*, 106(7), 2148-2152.
23. Mizutani, M. (2003). The Japanese quail. *Age*, 80, 90.
24. Moraes, H. L. D. S., Salle, C. T. P., Padilha, A. P., Nascimento, V. P. D., Souza, G. F. D., Pereira, R. A., ... & Salle, F. D. O. (2004). Infectious bursal disease: evaluation of pathogenicity of commercial vaccines from Brazil in specific pathogen free chickens. *Revista Brasileira de Ciência Avícola*, 6(4), 243-247.
25. Muskett, J. C., Hopkins, I. G., Edwards, K. R., & Thornton, D. H. (1979). Comparison of two infectious bursal disease vaccine strains: efficacy and potential hazards in susceptible and maternally immune birds. *The Veterinary Record*, 104(15), 332-334.
26. Ojeda, F., Skardova, I. A., Guarda, M. I., Ulloa, J., & Folch, H. U. G. O. (1997). Proliferation and apoptosis in infection with infectious bursal disease virus: a flow cytometric study. *Avian diseases*, 312-316.
27. Randall, M., & Bolla, G. (2008). Raising Japanese quail. *Primefacts*, 602, 1-5.

28. Recommended Agarose Gel Percentages for Resolution of Linear DNA. 3<sup>rd</sup> February 2016. Retrieved from: [http://www.genomics.agilent.com/files/Mobio/Nucleic%20Acids\\_Gel\\_Electrophoresis.pdf](http://www.genomics.agilent.com/files/Mobio/Nucleic%20Acids_Gel_Electrophoresis.pdf)
29. Romao, J. M., de Moraes, T. G. V., Salles, R. P. R., Cardoso, W. M., & Buxade, C. C. (2011). Efeito dos procedimentos de vacinação in ovo sobre embriões de codorna japonesa (*Coturnix japonica*) e desempenho da incubação. *Ciência Animal Brasileira*, 12(4), 584-592.
30. Schering-Plough Animal Health.(2004). A Comparison of IBD Vaccines Used to Control vvIBD: Histopathology – Bursa-Vac® Provides Protection with Comparable Effect on the Bursa. Schering- Plough Animal Health technical service bulletin.
31. Seet, C. P., & Azizah, M. D. (1987). Growth performance and carcass characteristics of Japanese quail [*Coturnix coturnix japonica*]. *MARDI Research Bulletin (Malaysia)*.
32. Sharma J.M., Dohms J., Walser M. & Snyder D.B. (1993). Presence of lesions without virus replication in the thymus of chickens exposed to infectious bursal disease virus. *Avian Dis.*, 37 (3), 741-748.
33. Sjaak J.J de Wit. (2014). Volvac IBD MLV: update on safety and efficacy studies. GD animal health service.
34. Sonfada M.L., Kwari H.D., Rabo J.S., Wiam I.M., and Hena S.A., (2014) Observations on The Quail's Bursa of Fabricius Under Normal and Experimental Infectious Bursal Disease Conditions. *African Journal of Cellular Pathology*. 2(1):29-34.
35. Stuart J.C. (1989). Acute infectious bursal disease in poultry. *Vet. Rec.*, 125 (10), 281.
36. Tabeekh, M. A., & Al-Mayah, A. A. S. (2009). Morphological investigation of bursa of fabricius of imported broilers and local chicks vaccinated with two types of ibd vaccines. In *Iraqi Journal of Veterinary Sciences* (Vol. 23, No. Suppl. 2, pp. En201-En206). College of Veterinary Medicine, University of Mosul.

37. Tsukamoto, K., Kojima, C., Komori, Y., Tanimura, N., Mase, M., & Yamaguchi, S. (1999). Protection of chickens against very virulent infectious bursal disease virus (IBDV) and Marek's disease virus (MDV) with a recombinant MDV expressing IBDV VP2. *Virology*, 257(2), 352-362.
38. Van den Berg, T. P., Eterradossi, N., Toquin, D., & Meulemans, G. (2000). Infectious bursal disease (Gumboro disease). *Revue scientifique et technique (International Office of Epizootics)*, 19(2), 509-543.
39. Volvac® IBD MLV. (2016). Dose and administration route. Boehringer Ingelheim Animal Health.

