

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF FLAVOKAWAIN A ON INTERLEUKIN-6 AND MELANIN PRODUCTIONS IN HISTAMINE-INDUCED HaCaT KERATINOCYTES CELLS AND MOUSE B16-F10 MELANOMA CELLS

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FBSB 2015 54

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PENGESAHAN

Dengan ini adalah disahkan bahawa laporan projek bertajuk " Effects of Flavokawain A on Interleukin-6 and Melanin Productions in Histamine-induced HaCaT Keratinocytes Cells and Mouse B16-F10 Melanoma Cells" telah disiapkan serta dikemukakan kepada Jabatan Biokimia oleh KIM JI MAN sebagai syarat untuk kursus BCH 4999 projek.

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ACKNOWLEGEMENTS

First and foremost, I would like to express my deepest gratitude to my dedicated supervisor, Dr Syahida Ahmad for allowing me to have this chance to step in into the animal cell culture world. Hereby, I express my appreciation for her support, enthusiasm, optimism and guidance throughout this project.

It is undeniable that I have gain delightful experience and grown from this final year project. Besides that, I have discovered determination and strength that I never even knew I have through this project too. All glory goes back to god, on whose support and renew my strength throughout this project.

A special thank you also forwarded to my mentor, Miss Nurul Atika Razali for all her guidance and scarification of time on teaching me all the things that I have to learn for my project. I also owe my deepest appreciation to all my lab colleagues, Miss Naimah Latif, Miss Amiza, Mr Ibrahim and Mr Yakubu for all their help and cooperation given. Thanks to all my friends especially, Wong Jo Yen, Kristie, Wong Sim Yee, Wong Sook Yee and Lim Yi who have always stand by my side and given their support to me.

Not forgetful, a special thanks dedicated to my parents, Kim Toh Kwang and Lim Faa Yin, my brother Kim Ji Hao and my sister Kim Ji Fang for their support and confidence giving to me.

Last but not least, thank you for the department of Biochemistry and all the staff for providing all the instruments, facilities and cooperation which helping me on completion of this project.

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LISTS OF ABBREVATIONS

2-S-CD	: 2-S-cysteinyldopa
5-S-CD	: 5-S-cysteinyldopa
6-S-CD	: 6-S-cysteinyldopa
AC	: Adenylyl cyclase
AD	: Atoptic dermatitis
cAMP	: cyclic adenosine monophosphate
COX-2	: Cyclooxygenase-2
CRE	: cAMP-responsive elements
CREB	: cAMP-responsive-element binding
DAG	: Diacylglycerol
DCT	: Dopachrome tautomerase
DHI	: Dihydroxyindole
DHICA	: Dihydroxyindole-2-carboxylic acid
DQ	: Dopaquinone
ERK	: Extracellular signal-regulated kinases
ET-1	: Endothelin-1
FLA	: Flavokawain A
H1R	: Histamine H1 receptor
ІКК	: I kappa B kinase
L	: Litre
IL-1α	: Interleukin-1a
IL-1	: Interleukin-1
IL-1β	: Interleukin-1ß
IL-6	: Interleukin-6
IL-8	: Interleukin-8

C

	IP ₃	: Inositol-1,4,5-trisphosphate
	LPS	: Lipopolysaccharide
	LTB 4	: Leukotriene B4
	MAPK	: Mitogen-activated protein kinase
	MC1R	: Melanocortin-1 receptor
	mg	: Milligram
	μg	: Microgram
	ml	: Millilitre
	NF-kB	: Nuclear factor- kappaB
	NO	: Nitric oxide
	PGD 2	: Prostaglandins D2
	PGE 2	: Prostaglandin E2
	PIH	: Post-inflammatory hyperpigmentation
	PKA	: Phosphokinase A
	РКС	: Phosphokinase C
	PLC	: Phospholipase C
	TNF-α	: Tumor necrosis factor-alpha
	TYR	: Tyrosinase
	TYRP 1	:Tyrosinase-related protein 1
	α-MSH	: Alpha melanocyte stimulating hormone

ABSTRACT

Chronic skin inflammatory diseases affecting large populations in Malaysia and this condition is getting serious globally. Post-inflammatory hyperpigmentation (PIH) is one of chronic skin inflammatory diseases which usually resulted after inflammation process raises health, cosmetic and psychosocial concerns. Resultantly, antiinflammatory and skin depigmenting agents have been developed to alleviate the effects thereof. However, current treatments on these diseases have potential toxicity and carcinogenicity. Therefore, there is a need to develop new anti-inflammatory and skin depigmenting agents with minimal side effects and greater efficacy. Chalcone derivatives is a group of compound belong to flavonoid family and has been reported with antiinflammatory and anti-melanogenic properties. Thus study ought to determine the antiinflammatory and anti-melanogrnic effects of the chalcone compound 1-(2-Hydroxy-4,6dimethoxyphenyl)-3-(4-methoxyphenyl)prop-2-en-1-one (Flavokawain A, FLA) on cytokine production (Interleukin-6) in 10 µM histamine-induced HaCaT cells and melanin content in 10 µM histamine-induced mouse B16-F10 melanoma cells. The results showed that 50 µM of FLA suppressed the IL-6 production by 44% with HaCaT cell viability of 75%. Nevertheless, 50 µM of FLA was successfully reduced the melanin content by 73% with the B16-F10 melanoma cell viability of 78%. FLA showed high inhibitory activity towards melanin production in histamine-induced B16-F10 melanoma cells. This has indicates that FLA possess potential anti-inflammatory and antimelanogenic activities. Thus, FLA has shown potential anti-inflammatory and depigmenting activities in vitro. Therefore, FLA might has the potential to be developed as an agent or alternative medicine to treat PIH. However, further studies need to be done to evaluate its efficacy and safety.

ABSTRAK

Kulit radang penyakit kronik telah menjejas populasi yang besar di Malaysia dan keadaan ini juga menjadi semakin serius di seluruh dunia. Hiperpigmentasi yang berlaku selepas proses radang merupakan salah satu penyakit kulit radang kronik yang biasanya berlaku selepas proses keradangan dan hal ini telah menimbulkan kebimbangan terhadap kesihatan, kosmetik dan psikososial. Berikutan perkara tersebut, agen antiradang dan penyahpigmen pada kulit telah dihasilkan untuk mengurangkan kesan-kesan tersebut. Walau bagaimanapun, rawatan semasa mengenai penyakit-penyakit ini mempunyai ketoksikan yang berpotensi dan kekarsinogenan. Oleh itu, terdapat keperluan untuk memajukan ejen penyahpigmen dan anti-radang kulit baru dengan kesan sampingan yang minimum dan keberkesanan yang lebih tinggi. Derivatif Chalcone adalah kumpulan sebatian berasal dari keluarga flavonoid dan telah dilaporkan dengan ciri-ciri anti-radang dan penyahpigmen. Oleh itu, kajian ini bertujuan untuk menentukan kesan anti-radang dan anti-melanogenik kompaun chalcone 1- (2-Hydroxy-4,6dimethoxyphenyl) -3- (4-methoxyphenyl) prop-2-en-1-satu (Flavokawain A, FLA) mengenai pengeluaran cytokine (Interleukin-6) sel HaCaT yang telah dirangsang dengan $10 \,\mu\text{M}$ histamine dan juga pengeluaran melanin pada sel B16-F10 melanoma yang telah dirangsang dengan 10 µM histamine. Hasil kajian menunjukkan bahawa 50 µM FLA telah mengurangkan pengeluaran IL-6 sebanyak 44% dengan kadar hidup sel HaCaT sebanyak 75% manakala 50 µM FLA telah berjaya mengurangkan pengeluaran melanin sebanyak 73% dengan kadar hidup sel B16-F10 melanoma sebanyak 78%. FLA menunjukkan aktiviti perencatan terhadap pengeluaran melanin dalam sel B16-F10 melanoma yang telah dirangsang dengan histamine. Hal ini telah menunjukkan bahawa FLA mempunyai potensi aktiviti anti-radang dan anti-melanogenik in vitro. Oleh itu, FLA berpotensi untuk dimajukan sebagai agen atau perubatan alternatif untuk merawat PIH. Walau bagaimanapun, kajian lanjut perlu dilakukan untuk menilai keberkesanan dan keselamatan FLA.

CHAPTER 1

INTRODUCTION

Inflammation is a process in which an organism used to remove injurious stimuli and is the starting point of healing process in order to get rid of foreign particles (Marone *et al.*, 2003). However, prolonged inflammation will lead to chronic skin inflammatory diseases. In the inflammation process, histamine (β imidazolylethylamine) act as inflammatory mediator (Mizuguchi *et al.*, 2012).

Atopic dermatitis (AD), lichen planus, psoriasis and allergic contact dermatitis are chronic skin inflammatory diseases that are commonly found (Lynde *et al.*, 2006). There are about 8% to 25% of the population worldwide being affected by AD, and the incidence is increasing, especially in developed countries (Weston and Howe, 2008).

During inflammation, various cytokines and chemokines will be released via NF- κ B pathway. This will results in transcription of pro-inflammatory cytokines genes like IL-6, IL-1 β and TNF- α . The continued releasing of these pro-inflammatory cytokines will lead to chronic skin inflammatory diseases. For example, IL-6 exerts stimulatory effects on T- and B-cells, thus favoring chronic inflammatory responses. So, strategies targeting IL-6 and IL-6 signaling led to effective prevention and treatment of models on chronic inflammatory diseases (Gabay, 2006).

In addition, hyperpigmentation are usually accompanied with inflammation and this condition can be explained by accumulation of histamine secreted from mast cells during inflammation and this will stimulate melanocytes to produce increased amounts of pigment (Ruzicka *et al.*, 1983) as well as increases tyrosinase activity in melanocytes (Yoshida *et al.*, 2000). The hyperpigmentation that occurred after the inflammation process is known as post-inflammatory hyperpigmentation (PIH).

PIH is defined as an acquired condition with excess of pigment deposited in the skin attributed to preceding skin inflammatory disease and is occurs at the site of skin injury after the inflammation has subsided. This acquired excess of pigment can be attributed to various preceding disease processes that affect the skin like infections, allergic reactions, mechanical injuries, reactions to medications, phototoxic eruptions, trauma and inflammatory diseases such as lichen planus, lupus erythematosus, atopic dermatitis). The patches that occurred may results from both increase in melanin production by individual melanocytes as well as increase in total number of melanocytes (Mak, 2010). Although PIH is not life-threatening, but patients who consider it a social embarrassment may lowered confidence of individual and eventually affected their social life.

Topical corticosteroids remain as the mainstay of anti-inflammatory treatment, it showing efficacy in the control of both acute and chronic skin inflammation. However, it will induce various cutaneous adverse effects in locally applied sites, such as telangiectasia, skin atrophy, hypertrichosis, hypopigmentation, perioral dermatitis and atrophic cutaneous striae (Furue *et al.*, 2003).

In the case of hyperpigmentation, hydroquinone (HQ) or 1,4-dihydroxybenzene has been used successfully to treat many forms of epidermal hyperpigmentation like melasma, freckles and PIH (Hu *et al.*, 2009). It is a phenolic bleaching compound which is also the gold-standard therapy for PIH. However, long-term used of HQ will bring

about side effect like 'hydroquinone halo'. This condition is characterized by a 'halo' which surrounding the dark macule due to the bleaching of the surrounding normal skin (Woolery-Lloyd and Kammer, 2011). Besides that, there is report on patients who develop exogenous ochronosis (EO) on area where HQ has been applied although it was in low concentrations, (2%) and for short periods of time (6 months) (Ribas *et al.*, 2010). Concern regarding the side effects of HQ has arisen because the benzene metabolites of hydroquinone *in vivo* also seem to be involved in bone marrow toxicity and even carcinogenesis when extensively used (Levitt, 2007; Nordlund *et al.*, 2006). Moreover, hydroquinone also considered to be cytotoxic to melanocytes (Briganti *et al.*, 2003; Hermanns *et al.*, 2000).

In recent years, extensive researches have been done for development of more effective and safer drugs in the treatment of chronic skin inflammatory skin disease as well as the alternative treatment on PIH. One of the bioactive compounds that researchers interested in is chalcones. Chalcones are compounds that mostly found in plants and its anti-inflammatory and anti-melanogenic characteristics maybe playing role in inflammation process and melanogenesis and thus might be an alternative to cure chronic skin inflammatory disease and PIH. Furthermore, chalcones are abundant in edible plants (Nowakowska, 2007) and this is believed to have less side effects than the current medication in treating of both chronic skin inflammatory disease and PIH.

Inhibition of cytokines production by chalcones could be an important therapeutic intervention in skin inflammatory diseases and this may lead to the discovery of new drugs as alternative in treating chronic skin inflammatory disease and PIH. The objectives of this study were :

- 1. To evaluate the effects of synthesized chalcones 1-(2-Hydroxy-4,6dimethoxyphenyl)-3-(4-methoxyphenyl)prop-2-en-1-one (FLA) on cytokines production (Interleukin-6) in histamine-induced HaCaT keratinocytes cells.
- 2. To evaluate the effects of synthesized chalcones 1-(2-Hydroxy-4,6dimethoxyphenyl)-3-(4-methoxyphenyl)prop-2-en-1-one (FLA) on melanin production in histamine-induced B16-F10 murine melanoma cells.

References

- Ahmad, S., Israf, D. A., Hj. Lajis, N., Shaari, K., Mohamed, H., Wahab, A. A., Ariffin, K. T., Hoo, W. Y., Aziz, N. A., Kadir, A. A., Sulaiman, M. R., Somchit, M. N. (2006). Cardamonin, inhibits pro-inflammatory mediators in activated RAW 264.7 cells and whole blood. *European Journal of Pharmacology*, 538, 188–194.
- Akdis, C. A., Simons F. E. R. (2006). Histamine receptors are hot in immunopharmacology. *European Journal of Pharmacology*, 533, 69–76.
- Akhtar, M. N., Sakeh, M. N., Zareen, S., Gul, S., Lo, K. M., Ul-Haq, Z., Ali Shah, S. A., Ahmad, S. (2015). Design and synthesis of chalcone derivatives as potent tyrosinase inhibitors and their structural activity relationship. *Journal of Molecular Structure*, 1085, 97-103.
- Anto, R. J., Sukumaran, K., Kuttan, G., Rao, M. N. A., Subbaraju, V., Kuttan, R. (1995). Anticancer and antioxidant activity of synthetic chalcones and related compounds. *Cancer Letters*, 97, 33–37.
- Aoki, Y., Qiu, D., Zhao, G. H., Kao, P. N. (1998). Leukotriene B4 mediates histamine induction of NF-kappa B and IL-8 in human bronchial epithelial cells. *American Journal of Physiology*, 274, 1030-1039.
- Arkansas Laser Solutions. (2013). Facial spider veins or telangiectasia. Available at http://www.arkansaslasersolutions.com/facial_spider_veins.html. Accessed March 20 (2015).
- Bandgara, B. P, Gawande, S. S., Bodade, R. G., Totre, J. V., Khobragade, C. N. (2010). Synthesis and biological evaluation of simple methoxylated chalcones as anticancer, anti-inflammatory and antioxidant agents. *Bioorganic and Medicinal Chemistry*, 18(3), 1364-1370.
- Barfod, A., Kemp, K., Hansen, M., Kharazmi, A. (2002). Chalcones from Chinese liquorice inhibit proliferation of T cells and production of cytokines. *International Immunopharmacology*, 2, 545–555.
- Bharate, S. B., Mahajan, T. R., Gole, Y. R. (2008). Synthesis and evaluation of pyrazolo[3,4-β]pyridines and its structural analogues as TNF-alpha and IL-6 inhibitors. *Bioorganic and Medicinal Chemistry*, 16(15), 7167–7176.
- Briganti, S., Camera, E., Picardo, M. (2003). Chemical and instrumental approaches to treat hyperpigmentation. *Pigment Cell and Melanoma Research*, 16, 101–110.
- Busca, R., and Ballotti, R. (2000). Cyclic AMP a key messenger in the regulation of *skin pigmentation. Pigment Cell Research*, 13, 60–69.
- Chan, Y. Y., Kim, K. H., Cheah, S. H. (2011). Inhibitory effects of *Sargassum* polycystum on tyrosinase activity and melanin formation in B16F10 murine melanoma cells, *Journal of Ethnopharmacology*, 137, 1183-1188.

- Chang T-S. (2009). An Updated Review of Tyrosinase Inhibitors. *International Journal* of Molecular Scences, 10(6), 2440-2475.
- Cole, G. W. (2013). Eczema pictures, causes, symptoms and treatment. Available at http://www.webmd.com/skin-problems-andtreatments/psoriasis/ss/types-of-psoriasis. Acessed April 13 (2015).
- Davis, E. C., Callender, V. D. (2010). Postinflammatory hyperpigmentation. A review of epidemiology, clinical features, and treatment options in skin of colour. *Journal of Clinical Aesthetic Dermatology*, 3, 20-31.
- DeCaprio, A. P. (1999). The toxicology of hydroquinone relevance to occupational and environmental exposure. *Critical Reviews in Toxicology*, 29(3), 283-330.
- Drucker, C., Gewiese, J., Malchow, S., Scheller, J., Rose-John S. (2010). Impact of interleukin-6 classcic- and trans-signaling on liver damage and regeneration. *Journal Of Autoimmunity*, 34, 29-37.
- Folmer, F., Blasius, R., Morceau, F., Tabudravu, J., Dicato, M., Jaspars, M., Diederich, M. (2006). Inhibition of TNF-α induced activation of nuclear factor kB by kava (*Piper methysticum*) derivatives. *Biochemical pharmacology*,7(1), 1206 1218.
- Foresti, R., Hoque, M., Monti, D., Green, C. J., Motterlini, R. (2005). Differential activation of heme oxygenase-1 by chalcones and rosolic acid in endothelial cells. *Journal of Pharmacological and Experimental Therapeutics*, 312, 686 693.
- Furue, M., Terao1, H., Rikihisa, W., Urabe, Q., Kinukawa, N., Nose, Y., Koga, T. (2003). Clinical dose and adverse effects of topical steroids in daily management of atopic dermatitis. *British Journal of Dermatology*, 148(1), 128–133.
- Gabay, C. (2006). Interleukin-6 and chronic inflammation. Arthritis Research and Therapy, 8(Suppl 2), S3.
- Giustizieri, M. L., Albanesi, C., Fluhr, J., Gisondi, P., Norgauer, J., Girolomoni, G. (2004). H1 histamine receptor mediates inflammatory responses in human keratinocytes. *Journal of Allergy and Clinical Immunology*, 114(5), 1176-1182.
- Gordon, P. R., and Gilchrest. B.A. (1989). Human melanogenesis is stimulated by diacylglycerol. *Journal of Investigative Dermatology*, 93, 700–702.
- Haas, H. and Panula, P. (2003). The role of histamine and the tuberomamillary nucleus in the nervous system. *Nature Reviews Neuroscience*, 4, 121-130.
- Halder, R. M. The role of retinoids in the management of cutaneous conditions in blacks. (1998). *Journal of American Academy of Dermatology*. 39(2 Pt 3), 98-103.
- Halder, R. M., Nootheti, P.K. (2003). Ethnic skin disorders review. *Journal of American Academy Dermatology*, 48(6 Suppl), 143-148.

- Harvima, I. T, Nilsson, G., Suttle, M. M., Naukkarinen, A. (2008) Is there a role for mast cells in psoriasis? Archieves of Dermatological Research, 300, 461–478.
- Hatziieremia, S., Gray, A. I., Ferro, V. A., Paul, A., Plevin, R. (2006). The effects of cardamonin on lipopolysaccharide-induced inflammatory protein production and MAP kinase and NFκB signalling pathways in monocytes/ macrophages. *British Journal of Pharmacology*, 149(2), 188–198.
- Hermanns, J. F., Petit, L., Martalo, O., Pierard-Franchimont, C., Cauwenbergh, G., Pierard, G. E. (2000). Unraveling the patterns of subclinical pheomelanin enriched facial hyperpigmentation: effect of depigmenting agents. *Dermatology* 201, 118–122.
- Hill, S. J., Ganellin, C. R., Timmerman, H., Schwartz, J. C., Shankley, N. P., Young, J. M. (1997). Classification of histamine receptors. *Pharmacology Review*, 49, 253–278.
- Hsieh, H. K., Tsao, L. T., Wang, J. P., Lin, C. N. (2000). Synthesis and antiinflammatory effect of chalcones. *Journal of Pharmarceutical Pharmacology*, 52, 163-171.
- Hu, Z. M., Zhou, Q., Lei, T. C., Ding, S. F., Xu, S. Z. (2010). Effects of hydroquinone and its glucoside derivaties on melanogenesis and antioxidation: biosafety as skin whitening agents. *Journal of Dermatological Science*, 55(3), 179-184.
- Jain, A., Pai, S., Shenoi, S. (2013). Exogenous ocronosis. Indian Journal of Dermatology, Venereology and Leprology, 79(4), 522-523.
- Kanda, N., Watanabe, S. (2003). Histamine enhances the production of nerve growth factor in human keratinocytes. *Journal of Investigate Dermatology*, 121, 570 577.
- Kanda, N., Watanabe, S. (2004). Histamine enhances the production of granulocyte macrophage colony-stimulating factor via protein kinase C and extracellular signal-regulated kinase in human keratinocytes. *Journal of Investigate Dermatology*, 122, 863-872.
- Khatib, S., Nerya, O., Musa, R., Tamir, S., Peter, T., Vaya, J. (2007). Enhanced substituted resorcinol hydrophobicity augments tyrosinase inhibition potency. *Journal of Medicinal Chemistry*, 50, 2676–2681.
- Khatib, S., Neryaa, O., Musaa, R., Shmuela, M., Tamira, S., Vayaa, J. (2005). Chalcones as potent tyrosinase inhibitors: the importance of a 2,4-substituted resorcinol moiety. *Bioorganic & Medicinal Chemistry*, 13(2), 433–441.
- Kim N. H. and Lee A. Y. (2010). Histamine effect on melanocyte proliferation and vitiliginous keratinocyte survival. *Experimental Dermatology*, 19, 1073–1079.

- Kim, K. J, Lee, J. S, Kwak, M. K. (2009). Anti-inflammatory action of mollugin and its synthetic derivatives in HT-29 human colonic epithelial cells is mediated through inhibition of NF-kappa B activation. *European Journal of Pharmacology*, 622(1-3), 52–57.
- Kohda, F., Koga, T., Uchi, H., Urabe, K., Furue, M. (2002). Histamine-induced IL-6 and IL-8 production are differentially modulated by IFN-g and IL-4 in human keratinocytes. *Journal of Dermatological Science*, 28, 34-41.
- Kolbe, L., Mann, T., Gerwat, W., Batzer, J., Ahlheit, S., Scherner, C., Wenck, H., Sta b, F. (2013). 4-n-butylresorcinol, a highly effective tyrosinase inhibitor for the topical treatment of hyperpigmentation. *Journal of the European Academy of Dermatology and Venereology*, 27(1), 19-23.
- Kwon, D. J., Ju, S. M., Youn, G. S., Choi, S. Y., Park, J. (2013). Suppression of iNOS and COX-2 expression by flavokawain A via blockade of NF-κB and AP-1 activation in RAW 264.7 macrophages. *Food and Chemical Toxicology*, 58, 479-486.
- Lee, A. Y. and Noh, M. (2013). The regulation of epidermal melanogenesis via cAMP and/or PKC signaling pathways: insights for the development of hypopigmenting agents. *Archieves of Phamarlogical Research*, 36, 792-801.
- Lee, J. J. (2006). Blockade of nuclear factor-κB signaling pathway and antiinflammatory activity of cardamomin, a chalcone analog from *Alpinia* onchigera. The Journal of Pharmacology and Experimental Therapeutics, 316(1), 271–278.
- Lerner, A. B., Moellmann, G., Varga, V. L, Halaban, R., and Pawelek, J. (1979). Action of melanocyte-stimulating hormone on pigment cells. Cold Spring Harbor Conferences of Cell Proliferation, 6, 187–197.
- Levitt, J. (2007). The safety of hydroquinone: a dermatologist's response to the 2006 Fedral Register. *Journal of the American Academy of Dermatology*, 57, 854 872.
- Lin, C. T., Kumar, K. J. S., Tseng, Y. S., Wang, Z. J., Pan, M. Y., Xiao, J. H., Chien, S. C., Wang, S. Y. (2009). Anti-inflammatory activity of Flavokawain B from *Alpinia pricei Hayata*. *Journal of Agricultural and Food Chemistry*, 57(14), 6060-6065.
- Lynde, C. B., Kraft, J. N., Lynde, C. W. (2006). Topical treatments for melasma and postinflammatory hyperpigmentation. *Skin Therapy Letter*, 11, 1-12.
- Mak, F. (2010). Post inflammatory hyperpigmentation: causation predisposing factors, histopathology, physiology and management options. *Journal of Cosmetic Surgery & Medicine*, 5(1), 44-48.

- Marone, G., Granata, F., Spadaro, G., Genovese, A., Triggiani, M. (2003). The histamine-cytokine network in allergic inflammation. *Journal of Allergy and Clinical Immunology*, 112, 83-88.
- Matsubara, M., Tamura, T., Ohmori, K., Hasegawa, K. (2005). Histamine H1 receptor antagonist blocks histamine-induced proinflammatory cytokine production through inhibition of Ca2+- dependent protein kinase C, Raf/MEK/ERK and IKK/IκB/NF-κB signal cascades. *Biochemical Pharmacology*, 69, 433–449.
- Mendis, E., Kim, M. M, Rajapakse, N., Kim, S. K. (2008). Suppression of cytokine production in lipopolysaccharide-stimulated mouse macrophages by novel cationic glucosamine derivative involves down-regulation of NF-kappaB and MAPK expressions. *Bioorganic & Medicinal Chemistry*, 16(18), 8390–8396.
- Mizuguchi, H., Miyagi, K., Terao, T., Sakamoto, T., Yamawaki, Y., Adachi, T., Ono, S., Sasaki, Y., Yoshimura, Y., Kitamura, Y., Takeda, N., Fukui, H. (2012). PMA induced dissociation of Ku86 from the promoter causes transcriptional up regulation of histamine H1 receptor. *Scientific Reports*, 2, 916-926.
- Nerya, O., Musa, R., Khatib, S., Tamir, S., Vaya, J. (2004). Chalcones as potent tyrosinase inhibitors: the effect of hydroxyl positions and numbers. *Phytochemistry*, 65 1389–1395.
- Nordlund, J. J., Grimes, P. E., Ortonnes, J. P. (2006). The safety of hydroquinone. Journal of the European Academy of Dermatology and Venereology, 20, 781-787.
- Nowakowska, Z. (2007). A review of anti-infective and anti-inflammatory chalcones. *European Journal of Medicinal Chemistry*, 42, 125-137.
- Oakley, A. (2014). Dermatitis and corticosteroids. Available at: http://dermnetnz.org/doctors/dermatitis/corticosteroids.html. Accessed April 16 (2015).
- Oda, T., Morikawa, N., Saito, Y., Masuho, Y., Matsumoto, S. (2000). Molecular cloningand characterization of a novel type of histamine receptor preferentially expressed in leukocytes. *Journal of Biological Chemistry*, 275, 36781-36786.
- Park, H. Y., and Gilchrest, B. A. (1999). Signaling pathways mediating melanogenesis. *Cellular and Molecular Biology*, 45, 919–930.
- Park, H. Y., Russakovsky, V., Ohno, S., Gilchrest. B. A. (1993). The beta isoform of protein kinase C stimulates human melanogenesis by activating tyrosinase in pigment cells. *Journal of Biological Chemistry* 268, 11742–11749.
- Parveen, I., Threadgill, M. D., Moorby, J. M., Winters, A. (2010). Oxidative phenols in forage crops containing polyphenol oxidase enzymes. *Journal of Agricultural* and Food Chemistry, 58, 1371–1382.

- Parvez, S., Kang, M., Chung, H. S., Cho, C., Hong, M. C., Shin, M. K., Bae. H. (2006). Survey and mechanism of skin depigmenting and lightening agents. *Phytotherapy Research*, 20(11), 921-934.
- Parvez, S., Kang, M., Chung, H-S., Bae. H. (2007). Naturally occurring tyrosinase inhibitors: mechanism and applications in skin health, cosmetics and agriculture industries. *Phytotherapy Research*, 20(9), 805-816.
- Pawelek, J. M. (1976). Factors regulating growth and pigmentation of melanoma cells. *Journal of Investigative Dermatology*, 66, 201–209.
- Pawelek, J., Wong, G., Sansone, M., Morowitz, J. (1973). Molecular biology of pigment cells. Molecular controls in mammalian pigmentation. *Yale Joutnal of Biology and Medicine*, 46, 430–443.
- Rahman, M. A. (2011). Chalcone: a valuable insight into the recent advances and potential pharmacological activities. *Chemical Sciences Journal*, Volume 2011, CSJ-29, 1-16.
- Ribas, J., Schettini, A. P. M., Cavalcante, M. S. M. (2010). Exogenous ochronosis induced by hydroquinone: report of four cases.. *Brazilian Annals of Dermatology*, 85(5), 699-703.
- Ruzicka T, Gluck S. (1983). Cutaneous histamine levels and histamine releasability from the skin in atopic dermatitis and hyper-IgE-syndrome. *Archieves of Dermatological Research*, 275, 41–44.
- Saito, H., Yasumoto, K., Takeda, K., Takahashi, K., Yamamoto, H., Shibahara, S. (2003). Microphthalmia-associated transcription factor in the wnt signaling pathway. *Pigment Cell and Melanoma Research*, 16, 261–265.
- Sakai, C., Ollmann, M., Kobayashi, T., Abdel-Malek, Z., Muller, J., Vieira, W. D., Imokawa, G., Barsh, G. S., Hearing, V. J. (1997). Modulation of murine melanocyte function in vitro by agouti signal protein. *EMBO Journal*, 16, 3544 3552.
- Savory, S. A., Pandya, A. G. (2014). Post-inflammatory hyperpigmentation. *Dermatology Atlas for Skin of Colour*, 2014, 21-25.
- Scheller, J., Chalaris, A., Schmidt-Arras, D., Rose-John, S. (2011). The pro- and anti inflammatory properties of the cytokine interleukin-6. *Biochimica et Biophysica Acta*, 878–888.
- Schwabe, K. P. (1994). *Kava-kava extract, process for the production thereof and use thereof.* In: Edited by Patent US, vol. 5296224. United States: Dr. Wilmar Schwabe GmbH & Co.

- Shibahara, S., Takeda, K., Yasumoto, K., Udono, T., Watanabe, K., Saito, H., Takahashi, K. (2001). Microphthalmia-associated transcription factor (MITF): multiplicity in structure, function, and regulation. *Journal of Investigative Dermatology Symposium Proceedings*, 6, 99–104.
- Shibata, T., Yamada, T., Ishii, T., Kumazawa, S., Nakamura, H., Masutani, H. (2003). Thioredoxin as a molecular target of cyclopentenone prostaglandins. *Journal of Biological Chemistry*, 278, 26046–26054.
- Srinivasan, B., Johnson, T. E., Lad, R., Xing, C. (2009). Structure-activity relationship studies of chalcone leading to 3-hydroxy-4,3',4',5'-tetramethoxychalcone and its analogues as potent nuclear factor kappa B inhibitors and their anticancer activities. *Journal of Medicinal Chemistry*, 52, 7228–7235.
- Tak, P. P., Firestein, G. S. (2001). NF-kappaB: a key role in inflammatory disease. The *Journal of Clinical Investigation*, 107(1), 7-11.
- Weston, W. L. and Howe, W. Epidemiology, clinical manifestations, and diagnosis of atopic dermatitis (eczema). Up To Date 2007. Available at: http://www.uptodate.com. Accessed January 16 (2015).
- White, M. V. (1990). The role of histamine in allergic diseases. *Journal of Allergy and Clinical Immunology*, 86(4), 599–605.
- Whitehead, R. J., Taylor, D.J., Evanson, J. M., Hart, I. R., Woolley, D. E. (1988). Demonstration of histamine H_2 receptors on human melanoma cells. *Biochemical and Biophysical Research Communications*, 151, 518-523.
- Widlund, H. R and Fisher, D. E. (2003). Microphthalamia-associated transcription factor: a critical regulator of pigment cell development and survival. *Oncogene*, 22, 3035–3041.
- Woolery-Llyod, H. and Kammer, J. N. (2011). Treatment of hyperpigmentation. Seminars in Cutaneous Medicine and Surgery, 30, 171-175.
- Wu, J., Lee, J., Cai, Y., Pan, Y., Ye, F., Zhang, Y., Zhao, Y., Yang, S., Li, X., Liang, G. (2011). Evaluation and discovery of novel synthetic chalcone derivatives as antiinflammatory agents. *Journal of Medicinal Chemistry*, 54, 8110–8123.
- Yadav, V. R., Prasad, S., Sung, B., Aggarwal, B. B. (2011). The role of chalcones in suppression of NF-κB-mediated inflammation and cancer. *International Immunopharmacology*, 11, 295–309.
- Yoshida, M., Takahashi, Y., Inoue, Shintaro. (2000). Histamine induces melanogenesis and morphologic changes by protein kinase A activation via H_2 receptors in human normal melanocytes. The Society for Investigative Dermatology, 114(2), 334-342.

- Zhang, X. W., Zhao, D. H., Quan, Y. C., Sun, L. P., Yin, X. M., Guan, L. P. (2010). Synthesis and evaluation of anti-inflammatory activity of substituted chalcone derivative. *Medicinal Chemistry Research*, 19, 403-412.
- Zhu, X. F., Xie, B. F., Zhou, J. M., Feng, G. K., Liu, Z. C., Wei, X. Y., Zhang, F. X., Liu, M. F., Zeng, Y. X. (2005). Blockade of vascular endothelial growth factor receptor signal pathway and antitumor activity of ON-III (2,4-Dihydroxy- 6 methoxy-3,5-dimethylchalcone), a component from Chinese herbal medicine. *Molecular Pharmacology*, 67, 1444–1450.

