

UNIVERSITI PUTRA MALAYSIA

CHARACTERISATION OF CELLULOLYTIC ACTIVITY AND BIOCONVERSION OF PALM KERNEL CAKE BY LACTIC ACID BACTERIA

WAN SUET YING

FBSB 2015 48

CHARACTERISATION OF CELLULOLYTIC ACTIVITY AND BIOCONVERSION OF PALM KERNEL CAKE BY LACTIC ACID BACTERIA



WAN SUET YING 161407

Thesis submitted to the Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia in fulfilment of the requirement for the Bachelor of Science (Hons.) Biotechnology

UNIVERSITI PUTRA MALAYSIA 2015

FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES UNIVERSITI PUTRA MALAYSIA

Date : 2 June 2015

LETTER OF PERMISSION

It is hereby to approve that I, **Wan Suet Ying (Matric No: 161407)** have done my final year project entitled **"Characterisation of cellulolytic activities and bioconversion of palm kernel cake by lactic acid bacteria"** under supervision of Assoc. Professor. Dr. Foo Hooi Ling from Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia. I hereby give the permission to my supervisor to write and prepare manuscript from the results of my research to be published in any form if I did not do it in six (6) months from the above date. The addition of my name in the manuscript depends on the supervisor herself.

Yours sincerely,

(Wan Suet Ying)

FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES UNIVERSITI PUTRA MALAYSIA

Date : 2 June 2015

APPROVAL SHEET

This attached project report entitled **"Characterisation of cellulolytic activities and bioconversion of palm kernel cake by lactic acid bacteria"** was prepared by **Wan Suet Ying (Matric No: 161407)** in fulfilment of the requirement for Degree of Bachelor of Science (Honour) Biotechnology in the Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia.

Approved by,

Assoc. Prof. Dr. Foo Hooi Ling Supervisor, Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Unversiti Putra Malaysia.

ABSTRACT

Abstract of thesis presented to the Faculty of Biotechnology and Biomolecular Sciences in fulfilment of the requirement for the Degree of Bachelor of Science (Hons.) Biotechnology

CHARACTERISATION OF CELLULOLYTIC ACITIVITIES AND BIOCONVERSION OF PALM KERNEL CAKE BY LACTIC ACID BACTERIA

By:

Wan Suet Ying June 2015

Supervisor : Assoc. Professor Dr. Foo Hooi Ling Faculty : Faculty of Biotechnology and Biomolecular Sciences

Cellulolytic enzyme is becoming more important due to its application in agro-wastes, which appear to be a potential substitution for production of bioenergy and biofuel. Lactic acid bacteria (LAB) isolated from Malaysian fermented food (Lactobacillus plantarum RG14, RG11, RI11, RS5, TL1, I-UL4 and B4) were characterised for their cellulolytic activities (CMCase, FPase, β-glucosidase, xylanase and mannanase assays) under different pH conditions (pH 5, 6.5 and 8). Two quantification methods (Bradford and modified Lowry methods) of protein concentration were conducted for the determination of specific celluloytic activity. Specific cellulolytic activities were 45 to 52 times higher when the protein concentration of cell-free supernatant was determined by Bradford method as compared to modified Lowry method. L. plantarum RG14, RG11 and RI11 were observed with higher scoring in overall cellulolytic activity and hence they were selected for biotransformation of palm kernel cake (PKC) via solid state fermentation (SSF) for 14 days with 2-day sampling intervals. The PKC extract was collected from fermented PKC to determine the cellulolytic activities and LAB population. The highest LAB cell population was observed at day 2 PKC extract treated by L. plantarum RG14 and RG11 with 8.15 log CFU/mL and 8.11 log CFU/mL, respectively. As for *L. plantarum* RI11, the highest cell population was noted at day 4 with 8.13 log CFU/mL. The cell population of LAB was maintained throughout the SSF. The solubilised protein concentration of PKC extract determined using modified Lowry method increased



according to the SSF period, whereas a decrement in solubilised protein concentration was observed when Bradford method was employed. This might due to lower sensitivity and linearity in Bradford method in comparison to modified Lowry method. Hence, the specific cellulolytic activity of PKC extract was determined using modified Lowry method under pH condition at 5. The CMCase activity was enhanced at day 6, followed by a drastic decline. The β -glucosidase activity was increased during the SSF with the highest activity observed at day 12 when PKC was treated by L. plantarum RG14 and day 14 by L. plantarum RG11 and RI11respectively. A significant enhancement in specific mannanase activity was exhibited at the 8th day of incubation. Unfortunately, no FPase activity was detected throughout the fermentation period. Interestingly, the specific cellulolytic activity obtained through submerged fermentation in MRS broth was higher as compared to PKC extract obtained through SSF. This was due to MRS broth provided sufficient moisture and nutrient to support the growth and enzyme production in LAB isolates. In conclusion, the LAB isolates were able to produce versatile cellulolytic enzymes to degrade various forms of polysaccharides and PKC. Thus, the cellulolytic enzymes of LAB possessed vast potential for biotransformation of biomass.

ABSTRAK

Abstrak tesis dihasilkan bagi Fakulti Bioteknologi and Sains Biomolekul untuk memenuhi keperluan Ijazah Sarjana Muda Sains (Hons) Bioteknologi

PENCIRIAN AKTIVITI CELLULOLYTIC DAN PENUKARAN BIO UNTUK HAMPAS ISIRONG KELAPA SAWIT OLEH BAKTERIA ASID LAKTIK

Oleh:

Wan Suet Ying Jun 2015

Penyelia : Profesor Madya Dr. Foo Hooi Ling Fakulti : Fakulti Biotekologi and Sains Biomolekul

Enzim cellulolytic menjadi lebih penting kerana penggunaannya dalam bahan buangan pertanian, yang berpotensi menjadi penngganti dalam pengeluran biotenaga dan biofuel. Aktivity cellulolytic bakteria asid laktik (LAB) yang diperolehi daripada makanan peraman Malaysia (Lactobacillus plantarum RG14, RG11, RI11, RS5, TL1, I-UL4 dan B4) ditentukan melalui ujian CMCase, FPase, β-glucosidase, xylanase dan mannanase pada pH 5, 6,5 dan 8. Dua kaedah kuantifikasi bagi kepekatan protein larut (kaedah Bradford dan Lowry yang diubahsuai) digunakan sebagai penentuan aktiviti cellulolytic khusus. Aktiviti cellulolytic khusus adalah 45 hingga 52 kali lebih tinggi apabila kepekatan protein larut bagi supernatan tiada sel ditentukan dengan kaedah Bradford berbanding kaedah Lowry yang diubahsuai. Lactobacillus plantarum RG14, RG11 dan RI11 dengan pemarkahan yang lebih tinggi dalam aktiviti cellulolytic keseluruhan dipilih untuk menjalani biotransformasi selama 14 hari dengan teknologi fermentasi pepejal (SSF) menggunakan hampas isirong kelapa sawit (PKC) sebagai substrat dengan sampel ujian diambil setiap dua hari. Ekstrak PKC yang difermentasi telah dikumpulkan untuk menentukan prestasi aktiviti cellulolytic dan pertumbuhan sel LAB. Pertumbuhan sel tertinggi diperhatikan pada hari ke-2 oleh L. plantarum RG14 and RG11 masing-masing dengan 8.15 log CFU/mL dan 8.11 log CFU/mL. Pada hari ke-4, L. plantarum RI11 menunjukkan pertumbuhan sel tertinggi iaitu 8.13 log CFU/mL. Pertumbuhan sel dalam PKC adalah stabil sepanjang proses teknologi fermentasi pepejal. Kepekatan protein larut daripada ekstrak PKC yang telah difermentasi ditentukan dengan



menggunakan kaedah Lowry yang diubahsuai meningkat manakala kaedah Bradford menunjukkan penurunan kepekatan protein larut. Hal ini mungkin disebabkan oleh tahap sensitiviti dan kelinearan yang rendah dalam kaedah Bradford berbanding kaedah Lowry yang diubahsuai. Oleh itu, aktiviti cellulolytic khusus bagi ekstrak PKC ditentukan dengan menggunakan kaedah Lowry yang diubahsuai pada pH 5. Aktiviti CMCase telah menunjukkan peningkatan pada hari ke-6 diikuti dengan penurunan drastik. Aktiviti β-glucodidase bagi ekstrak PKC meningkat semasa SSF dengan aktiviti tertinggi pada hari ke-12 oleh PKC yang dirawat oleh L. plantarum RG14 dan hari ke-14 oleh L. plantarum RG11 and RI11. Aktiviti mannanase khusus menunjukkan peningkatan pada hari ke-8 fermentasi. Aktiviti FPase tidak menunjukkan sebarang aktiviti sepanjang tempoh SSF. Aktiviti enzim khusus bagi ekstrak PKC yang diperolehi melalui fermentasi tenggelam menggunakan media MRS adalah lebih tinggi berbanding proses teknologi fermentasi pepejal. Hal ini disebabkan media MRS menyediakan kelembapan dan nutrien yang mencukupi untuk menyokong pertumbuhan sel dan penghasilan enzim dalam LAB. Kesimpulannya, LAB mampu menghasilkan enzim yang mempunyai aktiviti cellulolytic dan dapat meleraikan pelbagai bentuk polisakarida dan PKC. Oleh itu, enzim cellulolytic LAB berpotensi untuk biotransformasi bagi biomass.

ACKNOWLEDGEMENT

The accomplishment of this thesis in its current form is due to the assistance and guidance of several people. Here I would like to express my deepest gratitude and appreciation to my final year project supervisor, Assoc. Professor Dr. Foo Hooi Ling, for her supervision and warm encouragement along the process of research and thesis writing.

Not forgotten my sincere thanks to my senior, Muziana, Huey Kheng, Fu Haw, Cui Jin, Ye Heng and May Foong, for all the help and guidance provided during my research and thesis writing. They have given a lot of excellent advice, patience, caring and assistance with various problems during my research work. I am very grateful for the scientific advice, knowledge and insightful discussion they shared with me throughout my final year project duration. I warmly thank to all my friends and lab mates who have given a great emotional atmosphere to motivate me to continue with my project.

Last but not least, I would be forever thankful to my family, especially my beloved parents, Mr. Wan Kwee Loy and Mrs. Yong Lee Hong for giving me unconditional love can care. I would also like to thank my family members for their constant support and prayers. Finally, I just want to contribute my acknowledgment to all those who contributed directly or indirectly to the success of this project. All of your kindness means a lot to me. Thank you very much.

TABLE OF CONTENT

Title	Pages
LETTER OF PERMISSION	I
APPROVAL SHEET	Ű.
ABSTRACT	III
ABSTRAK	V
ACKNOWLEDGEMENT	VII
TABLE OF CONTENT	VIII
LIST OF TABLES	Х
LIST OF FIGURES	XI
LIST OF ABBREVIATIONS	XIII
CHAPTER 1	1
1.0 INTRODUCTION	1
1.1 Objectives	3
CHAPTER 2	4
2.0 LITERATURE REVIEW	4
2.1 Lactic Acid Bacteria	4
2.1.1 Sugar Fermentation Pathway	5
2.1.2 Classification of Lactic Acid Bacteria	6
2.2 Cellulosic Materials	7
2.2.1 Cellulose	8
2.2.2 Hemicellulose	9
2.4 Cellulolytic and Hemicellulolytic Enzyme	11
2.4.1 Cellulase	11
2.4.2 Hemicellulase	15
2.5 Enzymatic Assays	16
2.5.1 FPase	17
2.5.2 Exoglucanase	18
2.5.3 Endoglucanase	18
2.5.4 β-glucosidase	18
2.5.5 Xylanase	19
2.5.6 Mannanase	19
2.6 Solid State Fermentation	20
CHAPTER 3	21
3.0 MATERIALS AND METHODS	21
3.1 Microorganism and Culture	21
3.2 Preparation of cell-free supernatant	21

3.3 Buffer preparation	21
3.4 Preparation of substrates	21
3.4.1Carboxymethylcellulose	21
3.4.2 Xylan from Birchwood	22
3.4.3 Locust Bean Gum	22
3.4.4 Preparation of Dinitrosalicylic (DNS) reagent	22
3.4.5 Measurement of reducing sugar	22
3.5 Enzyme Activity Assays	22
3.5.1 CMCase activity	22
3.5.2 FPase activity	23
3.5.3 β-glucosidase activity	23
3.5.4 Xylanase activity	23
3. <mark>5.5 Mannanase activity</mark>	24
3.6 Protein content determination	24
3.6.1 Bradford method	24
3.6.2 Modified Lowry method	24
3.7 Solid state fermentation with palm kernel cake	25
3.8 Preparation of palm kernel cake extract	25
3.9 Viable cell count	25
3.10 Enzyme activity assay for palm kernel cake extract	25
3.11 Statistical analysis	25
CHAPTER 4	26
4.0 RESULTS AND DISCUSSION	26
4.1 Characterisation of cellulolytic activity	26
4.2 Solid state fermentation with palm kernel cake	45
CHAPTER 5	59
5.0 CONCLUSION	59
5.1 RECOMMENDATION	61
REFERENCES	62
APPENDICES	68
Appendix I - Standard curves	68
Appendix II - Data for specific enzyme activity from CFS of LAB	71
Appendix III - Data for cell population of Lactic acid bacteria	76
Appendix IV - Data for solubilised protein concentration of PKC extract	77
Appendix V - Data for specific enzyme activity of PKC extract	78

LIST OF TABLES

Table	Pages
Table 4.1: Protein concentration of cell-free supernatant produced by different LAB isolates using Bradford and modified Lowry method.	27
Table 4.2: The summary of the specific enzyme activity in different pH conditions.	40
Table 4.3: The summary of the specific enzyme activity in different pH conditions (Continued).	41
Table 4.4: The summary of the specific enzyme activity in different pH conditions (Continued).	42
Table 4.5: The summary of specific enzyme activity at pH 5.	43

C

LIST OF FIGURES

Figure	Pages
Figure 2.1: Major fermentation pathway of glucose.	6
Figure 2.2: Representation of cell wall structure in a fiber cell from wood.	8
Figure 2.3: The structural formula of cellulose.	8
Figure 2.4: The plant cell wall structure with lignin, cellulose and hemicellulose.	9
Figure 2.5: The flow chart for palm oil extraction process.	10
Figure 2.6: The mechanism of cellulose hydrolydid with cellulase complex.	11
Figure 2.7: The schematic diagram of cellulase complex hydrolytic action on amorphous and microcrystalline cellulose.	12
Figure 2.8: The mechanism of cellulase action.	13
Figure 2.9: The redox reaction between aldehyde group and 3,5- dinitrosalicylic acid reagent.	16
Figure 4.1: Specific CMCase activity of cell-free supernatant produced by LAB isolates in different pH conditions.	29
Figure 4.2: Specific FPase activity of cell-free supernatant produced by LAB isolates in different pH conditions.	31
Figure 4.3: Specific β-glucosidase activity of cell-free supernatant produced by LAB isolates in different pH conditions.	33
Figure 4.4: Specific xylanase activity of cell-free supernatant produced by LAB isolates in different pH conditions.	35
Figure 4.5: Specific mannanase activity of cell-free supernatant produced by LAB isolates in different pH conditions.	37
Figure 4.6: Cell population of lactic acid bacteria isolated from palm kernel cake treated with <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 via solid state fermentation.	46
Figure 4.7: Solubilised protein concentration of palm kernel cake extract produced by <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 for 14 days via solid state fermentation.	48

Figure 4.8: Specific CMCase activity of palm kernel cake extract produced by <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 for 14 days via solid state fermentation.	50
Figure 4.9 : Specific FPase activity of palm kernel cake extract produced by <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 for 14 days via solid state fermentation.	51
Figure 4.10: Specific β-glucosidase activity of palm kernel cake extract produced by <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 for 14 days via solid state fermentation.	53
Figure 4.11: Specific xylanase activity of palm kernel cake extract produced by <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 for 14 days via solid state fermentation.	55
Figure 4.12: Specific mannanase activity of palm kernel cake extract produced by <i>Lactobacillus plantarum</i> RG14, RG11 and RI11 for 14 days via solid state fermentation.	57

 \bigcirc

LIST OF ABBREVIATIONS

Abbreviation used in this text:

g	gram
log	logarithm with base 10
Μ	Molarity
h	hour
min	minute
mL	millilitre
nmol	nanomole
рН	exponent of hydrogen ion
rpm	rotation per minute
hà	microgram
μ	microlitre
μm	micrometer
°C	degree celsius
%	percentage
СМС	carboxymethylcellulose
FPase	Filter paperase
LAB	Lactic acid bacteria
NSP	Non starch polysaccharides
РКС	Palm kernel cake
SSF	Solid state fermentation



CHAPTER 1

1.0 INTRODUCTION

The explosive increment of global population has been a popular topic for recent decades due to the consequences it brings along. Problems caused by exponentially increment of human population are given great attention and well publicized. Environmental pollution and food demand are the problems we are facing today. With limited resources available such as non renewable fossil fuel and inconvenience living condition, it could be a challenge for the living condition of human beings.

Scientists are putting a lot of efforts in exploring potential solutions for the problems we are facing nowadays. It was found that cellulosic materials, the most abundant renewable resource on earth (Robson & Chambliss, 1984), could be a possible solution for the state of difficulties we are facing. These biological resources have a great potential to serve as an alternative for diminishing fossil energy resources (Yin *et al.*, 2010). The complete utilization of cellulosic materials has become increasingly important due to the global demand of resources.

According to Malaysia Palm Oil Board (MPOB) (2014), Malaysia produced around 1.7 million tonnes of crude palm oil and was known to be the second largest export in the year of 2013. Production of agro-wastes from oil palm industry is a common problem for environmental pollution issue. Agro-wastes or agricultural wastes refer to the waste produces from various agricultural processes, usually from farms, poultry houses and plantation sites. Hence, the production of biofuel and bioenergy from agro-wastes has gained vast attention from the public, instead of consumable crops.

Palm oil industry is one of the major economy engines of Malaysia with the production of around 19 million tonnes of crude palm oil in 2014, as reported by MPOB (2014). The wealth generated by palm oil industry comes along with the bulk amount of residual wastes or biomass which produces as the by-product that contributes to environmental pollution. There are 4.7 million arches of oil palm plantation in Malaysia, with the potential to produce 77.2 million tonnes of palm biomass, which is around 94% of the biomass produce in Malaysia, as reported by Sin Chew Daily News (2014).

1

Palm kernel cake (PKC) is one of the by-products results from palm kernel extraction process (Tuan Lah *et al.*, 2012). Raw PKC with low economic value is usually disposed as waste and creates environmental issue. Therefore, recent research suggests that PKC can be used as substrate for solid state fermentation (SSF) in order to produce value-added products such as enzymes. In addition, SSF of PKC with microorganism can be used to enhance the nutritive quality of agrowastes that can be applied as animal feed. The low moisture content of SSF is favourable because a lower chance of contamination by other microorganism and higher porosity to provide aeration (Sindhu *et al.*, 2006).

Waste transformation is a trend to overcome the environmental issue as well as sustaining the food security worldwide. The presence of cellulolytic and hemicellulolytic enzymes is essential for the degradation of PKC due to the characteristic of PKC which mainly consists of non starch polysaccharides (NSPs) and crude fibre, with the composition of 78% mannan, 3% arabinoxylan, 3% glucuronoxylan and 12% cellulose (Alimon, 2004).

Synergetic reaction of cellulase complex: endo- β -1,4-glucanase (EC 3.2.1.4), exo- β -1,4-glucanase (EC 3.2.1.91) and β -D-glucosidase (EC 3.2.1.21) provide effective biological hydrolysis β -1,4 linked of cellulose into glucose (Yin *et al.*, 2010). Apart from cellulose, hemicellulose is another component found mainly in primary and secondary layer of plant cell wall that closely associated with cellulose and lignin. The presence of hemicellulolytic enzyme such as xylanase and mannanase are required to degrade the xylan and mannan in PKC to ensure effective biodegradation of PKC.

Fungus is a common producer for cellulase, yet the cost of production is high due to the substrates use and its slow growth (Ariffin *et al.*, 2006). Hence, the focus has directed to bacteria with ability to produce cellulase as a potential substitution for fungus in the production of cellulase enzyme due to its faster growth rate compared to fungus. In this study, the effort was put forward to investigate the potential of lactic acid bacteria (LAB) to produce cellulolytic enzyme.

Simple fermentative nature of LAB which involves one or a few fermentation end products is considerable academic interest and provides an excellent model system for the study of energy transduction, solute transport and membrane biology (Axelsson, 1998). On the other hand, the diverse metabolic capacity that enables them to adapt to a variety of conditions has made LAB more applicable in wide range of industrial fields. Varieties of food products incorporated with LAB produce in commercial scale due to their positive health aspects reported in the recent decades (Melgar *et al.*, 2012). The general purpose of this study was directed toward the cellulolytic activity of LAB isolated from Malaysian fermented food. Cellulolytic activity of isolated LAB was characterised through different cellulolytic enzyme assays. The selected cellulolytic activity of LAB isolates were then employed for SSF of PKC for 14 days of incubation was determined.

1.1 Objectives

The specific objectives of this project are:

- 1. To characterise cellulolytic activities of LAB isolated from Malaysian fermented food.
- 2. To biotransform palm kernel cake by cellulolytic LAB isolates via SSF.

REFERENCES

- Abdeshahian, P., Samat, N., & Yusoff, W. M. W. (2010). Utilization of palm kernel cake for production of β-glucosidase by *Aspergillus niger* FTCC 5003 in solid substrate fermentation using an aerated column bioreactor. *Biotechnology*, *9*(1), 17-24.
- Alimon, A. R. (2004). The nutritive value of palm kernel cake for animal feed. *Palm Oil Dev, 40*(1), 12-14.
- Alshelmani, M. I., Loh, T. C., Foo, H. L., Lau, W. H., & Sazili, A. Q. (2013). Characterization of cellulolytic bacterial cultures grown in different substrates. *The Scientific World Journal, 2013*.
- Alshelmani, M. I., Loh, T. C., Foo, H. L., Lau, W. H., & Sazili, A. Q. (2014). Biodegradation of palm kernel cake by cellulolytic and hemicellulolytic bacterial cultures through solid state fermentation. *The Scientific World Journal, 2014*.
- Álvarez, C. J., Hernández, D. E. M., Arana, C. A., Díaz, G. G., & Mercado, F. Y. (2013). Purification and characterization of xylanase SRXL1 from *Sporisorium reilianum* grown in submerged and solid-state fermentation. *BioResources, 8*(4), 5309-5318.
- Ariffin, H., Abdullah, N., Umi Kalsom, M. S., Shirai, Y., & Hassan, M. A. (2006). Production and Characterization of Cellulase by *Bacillus pumilus* EB3. *International Journal of Engineering and Technology*, *3*(1), 47-53.
- Axelsson, L. (1998). Lactic Acid Bacteria: Classification and Physiology. In S. Salminen & V. A. Wright (Eds.), *Lactic Acid Bacteria: Microbiology and Functional Aspects*: Marcel Dekker Inc.
- Axelsson, L. (2004). Lactic acid bacteria: Classification and physiology Lactic acid bacteria: Microbiology and functional aspects (pp. 1-66).
- Babu, K. R., & Satyanarayana, T. (1996). Production of bacterial enzymes by solid state fermentation. *Journal of scientific & industrial research*, *55*(5-6), 464-467.
- Baraldo Junior, A., Borges, D. G., Tardioli, P. W., & Farinas, C. S. (2014). Characterization of β-glucosidase produced by *Aspergillus niger* under solidstate fermentation and partially purified using MANAE-agarose. *Biotechnology research international, 2014.*
- Berges, J. A., Fisher, A. E., & Harrison, P. J. (1993). A comparison of Lowry, Bradford and Smith protein assays using different protein standards and protein isolated from the marine diatom *Thalassiosira pseudonana*. *Marine Biology*, *115*(2), 187-193.
- Bhargav, S., Panda, B. P., Ali, M., & Javed, S. (2008). Solid-state fermentation: an overview. *Chemical and Biochemical Engineering Quarterly*, 22(1), 49-70.
- Biddle, A. D., & Warner, P. J. (1993). Induction of 6-phosphogluconate pathway is correlated with change of colony morphology in *Lactobacillus acidophilus*. *FEMS Microbiology Rev.*, *12*, 47.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical biochemistry*, 72(1), 248-254.
- Camassola, M., & Dillon, A. J. P. (2012). Cellulase Determination: Modification to Make the Filter Paper Assay Easy, Fast, Practical and Efficient. *Open Access Scientific Reports, 1*(1), 1-4.
- Catte, M., Gancel, F., Dzierszinski, F., & Tailliez, R. (1999). Effects of water activity, NaCl and smoke concentrations on the growth of *Lactobacillus plantarum* ATCC 12315. *International journal of food microbiology*, *52*(1), 105-108.
- Cave, I. D., & Walker, J. F. C. (1994). Stifness of wood in farown plantation softwood: influence of microfibril angle. *Forest Product Journal*.

Chaplin, M. F., & Bucke, C. (1990). *Enzyme technology*: CUP Archive.

- Chee, K. L., & Ayob, M. K. (2013). Optimization of hexametaphosphate-assisted extraction and functional characterization of palm kernel cake protein. *Food Science and Technology International, 19*(2), 109-122.
- Chen, W. L., Liang, J. B., Jahromi, M. F., Ho, Y. W., & Abdullah, N. (2013). Optimization of multi-enzyme production by fungi isolated from palm kernel expeller using response surface methodology. *BioResources, 8*(3), 3844-3857.
- Chin, F. Y. (2001). *Palm kernel cake (PKC) as a supplement for fattening and dairy cattle in Malaysia*. Paper presented at the 7th Meet. of FAO Regional Working Group on Grazing and Feed Resources for SE Asia Manado, Indonesia.
- da Silva, R., Lago, E. S., Merheb, C. W., Macchione, M. M., Park, Y. K., & Gomes, E. (2005). Production of xylanase and CMCase on solid state fermentation in different residues by *Thermoascus aurantiacus* miehe. *Brazilian Journal of Microbiology*, 36(3), 235-241.
- Dashtban, M., Maki, M., Leung, K. T., Mao, C., & Qin, W.S. (2010). Cellulase activities in biomass conversion: measurement methods and comparison. *Critical reviews in biotechnology*, *30*(4), 302-309.
- Dave, R. I., & Shah, N. P. (1996). Evaluation of media for selective enumeration of *Streptococcus thermophilus, Lactobacillus delbrueckii ssp. bulgaricus, Lactobacillus acidophilus, and Bifidobacteria. Journal of Dairy Science, 79*(9), 1529-1536.
- Dawson, R. M. C., Elliot, D. C., Elliot, W. H., & Jones, K. M. (1974). pH, buffers and physiological media. *Data for biochemical research*, 484-485.
- De Gussem, R. L., Aerts, G. M., Claeyssens, M., & De Bruyne, C. K. (1978). Purification and properties of an induced β-D-glucosidase from *Stachybotrys atra*. *Biochimica et Biophysica Acta (BBA) - Enzymology*, *525*(1), 142-153.
- Donohue, D. C., Salminen, S., & Marteau, p. (2004). Safety of Probiotic Bacteria. In S. Salminen & V. A. Wright (Eds.), *Lactic Acid Bacteria: Microbiology and Functional Aspects* (pp. 369-381): Maecel Dekker, Inc.
- El-Ghonemy, D. H. I., Ali, T. H., & Moharam, M. E. (2014). Optimization of culture conditions for the production of extracellular cellulases via solid state fermentation. *British Microbiology Research Journal*, *4*(6), 698-714.
- Esau, K. (1953). Plant anatomy. Soil Science, 75(5), 407.
- Espejo, J., Alcala, M., Esteban, M. A., Gomez, R., & Slik, S. (1994). Influence of sodium chloride concentration on the growth and survival of microorganisms isolated from Cabrales cheese. *MAN Microbiologie, aliments, nutrition, 12*(3), 251-254.
- Eze, J. M. O., & Dumbroff, E. B. (1982). A comparison of the Bradford and Lowry methods for the analysis of protein in chlorophyllous tissue. *Canadian Journal of Botany, 60*(7), 1046-1049.
- Fernández, E. M. T., Ramón, D., Piñaga, F., & Vallés, S. (1992). Xylanase production by *Aspergillus nidulans*. *FEMS microbiology letters*, *91*(2), 91-96.
- Ghose, T. k., & Bisaria, V. S. (1987). Measurement of Hemicellulase Activities Part 1: Xylanases. *International Union of Pure and Applied Chemistry, 59*(12), 1739-1752.
- Ghoshal, S. (2009). *Fundamentals of bioanalytical techniques and instrumentation*: PHI Learning Pvt. Ltd.
- Gueimonde, M., Jalonen, L. H., Hiramatsu, M., & Salminen, S. (2006). Adhesion and competitive inhibition and displacement of human enteropathogens by selected *Lactobacilli*. *Food Research International*, *39*, 467-471.
- Gupta, U., & Kar, R. (2009). Xylanase production by a thermo-tolerant *Bacillus* species under solid-state and submerged fermentation. *Brazilian Archives of Biology and Technology*, *52*(6), 1363-1371.

- Gupte, A., & Madamwar, D. (1997). Solid state fermentation of lignocellulosic waste for cellulase and β-Glucosidase production by cocultivation of *Aspergillus ellipticus* and *Aspergillus fumigatus*. *Biotechnology Progress*, *13*(2), 166-169.
- Gusakov, A. V., Kondratyeva, E. G., & Sinitsyn, A. P. (2011). Comparison of Two methods for Assaying Reducing Sugars in the Determination of Carbohyrase Activities. *International Journal of Analytical Chemistry*, 1-4.
- Hägglund, P. (2002). *Mannan-hydrolysis by hemicellulases: enzyme-polysaccharide interaction of a modular beta-mannanase*. Lund University.
- Haigler, C. H. (1990). Biosynthesis and biodegradation of cellulose: CRC Press.
- Jernejc, K., Cimerma, A., & Perdih, A. (1986). Comparison of different methods for protein determination in *Aspergillus niger* mycelium. *Applied microbiology and biotechnology*, *23*(6), 445-448.
- Jones, A. M., Reed, R. H., Weyers, J., & Weyers, J. D. B. (2007). *Practical skills in biology*: Pearson Education.
- Kandler, O., & Weiss, N. (1986). Regular, non-sporing Gram-positive rods. In P. H. A. Sneath, N. S. Mair, M. E. Sharpe & J. G. Holt (Eds.), (Vol. 2, pp. 1208-1234): Williams and Wilkins.
- Karnchanatat, A., Petsom, A., Sangvanich, P., Piaphukiew, J., Whalley, A. J. S., Reynolds, C. D., & Sihanonth, P. (2007). Purification and biochemical characterization of an extracellular β-glucosidase from the wood-decaying fungus *Daldinia eschscholzii* (Ehrenb.: Fr.) Rehm. *FEMS microbiology letters*, 270(1), 162-170.
- Khalil, M. I., Hoque, M. M., Basunia, M. A., Alam, N., & Khan, M. A. (2011). Production of cellulase by *Pleurotus ostreatus* and *Pleurotus sajor-caju* in solid state fermentation of lignocellulosic biomass. *Turkish Journal of Agriculture and Forestry*, 35(4), 333-341.
- King, K. W., & Vessal, M. I. (1986). Enzymes of the Cellulase Cmplex. In R. F. Gould (Ed.), *Cellulase and Their Application* (pp. 7-25): American Chemical Society.
- Kinoshita, H., Wakahara, N., Watanabe, M., Kawasaki, T., Matsuo, H., Kawai, Y., Horii, A. (2008). Cell surface glyceraldehyde-3-phosphate dehydrogenase (GAPDH) of *Lactobacillus plantarum* LA 318 recognizes human A and B blood group antigens. *Research in microbiology*, *159*(9), 685-691.
- Kormelink, F. J. M., Searle-van Leeuwen, M. J. F., Wood, T. M., & Voragen, A. G. J. (1993). Purification and characterization of three endo-(1, 4)- β -xylanases and one β -xylosidase from *Aspergillus awamori*. *Journal of Biotechnology*, 27(3), 249-265.
- Korotkova, O. G., Semenova, M. V., Morozova, V. V., Zorov, I. N., Sokolova, L. M., Bubnova, T. M., Sinitsyn, A. P. (2009). Isolation and properties of fungal βglucosidases. *Biochemistry (Moscow),* 74(5), 569-577.
- Krishna, C. (1999). Production of bacterial cellulases by solid state bioprocessing of banana wastes. *Bioresource Technology*, *69*(3), 231-239.
- Lee, S. N. (2011). The production of fungal mannanase, cellulase and xylanase using palm kernel meal as a substrate. *Walailak Journal of Science and Technology (WJST), 4*(1), 67-82.
- Leeber, S., Vanserleyden, J., & De Keersmaecker, S. C. J. (2008). Genes and Molecules of *Lactobacilli* supporting probiotic action. *Microbiology and Molecular Biology Reviews*, 72(4), 728-764.
- Lim, Y. S., Foo, H. L., Raha, A. R., Loh, T. C., Bejo, M. H., & Gulam Rusul, R. A. (2006). *The probiotic characteristics of Lactobacillus plantarum strains isolated from local foods.* Paper presented at the Proceeding of the 28th Symposium of Malaysian Society for Microbiology, Melaka, Malaysia.
- Lucarini, A. C., & Kilikian, B. V. (1999). Comparative study of Lowry and Bradford methods: interfering substances. *Biotechnology techniques, 13*(2), 149-154.

- Lusis, A. J., & Becker, R. R. (1973). The β-glucosidase system of the thermophilic fungus *Chaetomium thermophile* var. *Coprophile* n. var. *Biochimica et Biophysica Acta (BBA) General Subjects, 329*(1), 5-16.
- Lynd, L. R., Weimer, P. J., Van Zyl, W. H., & Pretorius, I. S. (2002). Microbial cellulose utilization: fundamentals and biotechnology. *Microbiology and molecular biology reviews*, *66*(3), 506-577.
- Malaysia Palm Oil Board: Statistic. (2014). *Economics & Industry Development Division: Malaysia Palm Oil Board*. Retrieved Oct, 2014 from http://bepi.mpob.gov.my/index.php/statistics/production/118-production-2013/601-annual-forecast-production-of-crude-palm-oil-2012-2013.html
- Mandels, M., & Reese, E. T. (1960). Induction of cellulase in fungi by cellobiose. Journal of Bacteriology, 79(6), 816.
- Mayer, L., Schick, L. L., & Setchell, F. W. (1986). Measurement of protein in near shore marine sediments. *Marine Ecology-Progress Series, 30*(40577), 159.
- Medeiros, R. G., Ceolho, L. A., & Filho, E. X. F. (2008). Agriculturral Residues as Source for Production of Hemicellulases from *Humicola grisea* var. *thermoidea*. *Dynamic Biochemistry, Process Biotechnology and Molecular Biology, 2*(1), 30-33.
- Melgar, L. G., Rivera, E. Y., & Hernández, S. H. (2012). Lactobacillus plantarum: An Overvoew with Emphasis in Biochemical and Healthy Properties. In A. I. P. Compos & A. L. Mena (Eds.), Lactobacillus: Classification, Uses and Health Implications (pp. 1-34). New York: Nova Science Publishers, Inc.
- Miller, G. L. (1959a). Protein determination of large numbers of samples. *Analytical chemistry*, *31*(5), 964-964.
- Miller, G. L. (1959b). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical chemistry*, *31*(3), 426-428.
- Miller, G. L., Blum, R., Glennon, W. E., & Burton, A. L. (1960). Measurement of carboxymethylcellulase activity. *Analytical Biochemistry*, *1*(2), 127-132.
- Moghadam, M. S., Foo, H. L., Leow, T. C., Rahim, R. A., & Loh, T. C. (2010). Novel bacteriocinogenic *Lactobacillus plantarum* strains and their differentiation by sequence analysis of 16S rDNA, 16S-23S and 23S-5S intergenic spacer regions and randomly amplified polymorphic DNA analysis. *Food Technology and Biotechnology*, 48(4), 476-483.
- Murray, P., Aro, N., Collins, C., Grassick, A., Penttilä, M., Saloheimo, M., & Tuohy, M. (2004). Expression in *Trichoderma reesei* and characterisation of a thermostable family 3 β-glucosidase from the moderately thermophilic fungus *Talaromyces emersonii*. Protein expression and purification, 38(2), 248-257.
- Nigam, P. S. N., & Pandey, A. (2009). *Biotechnology for agro-industrial residues utilisation: utilisation of agro-residues*: Springer Science & Business Media.
- Niklaus, M., Jakob, H., Mats, K., Roberto, R., & Ferdi, S. (2013, 23 May 2015). [Catalytic milling: A new entry point for lignocellulose biorefineries].
- Okutucu, B., Dinçer, A., Habib, O., & Zihnioglu, F. (2007). Comparison of five methods for determination of total plasma protein concentration. *Journal of biochemical and biophysical methods, 70*(5), 709-711.
- Olaniyi, O. O., Igbe, F. O., & Ekundayo, T. C. (2013). Optimization studies on mannanase production by *Trichosporonoides oedocephalis* in submerged state fermentation. *E3 Journal of Biotechnology and Pharmacrutical Research*, *4*(7), 110-116.
- Oskoueian, E., Abdullah, N., Idrus, Z., Ebrahimi, M., Goh, Y. M., Shakeri, M., & Oskoueian, A. (2014). Palm kernel cake extract exerts hepatoprotective activity in heat-induced oxidative stress in chicken hepatocytes. *BMC complementary and alternative medicine*, *14*(1), 368.
- Othman, M. F., Kalil, M. S., & Sahri, M. M. (2013). Solid state fermentation of palm kernel cake (PKC) by newly isolated *Rhizopus oryzae* Me01. *Asian Journal of Experimental Biological Sciences, 4*(1), 84-88.

- Palm Biomass with Great Potential in Producing Renewable Energy. (2014). *Sin Chew Daily News*. Retrieved Oct, 2014, from http://news.sinchew.com.my/node/393482
- Prior, B. A., Du Preez, J. C., & Rein, P. W. . (1992). Environmental parameters *Solid-substrate cultivation* (pp. 65-86).
- Rapp, P, & Beermann, A. (1991). Bacterial Cellulases. In C. H. Haigler & P. J. Weimwe (Eds.), *Biosynthesis and Biodegradation of Cellulose* (pp. 535-597): Marcel Dekker, Inc.
- Rashid, A. S., Ibrahim, D., & Omar, I. C. (2012). Mannanase production by *Aspergillus niger* USM F4 via solid substrate fermentation in a shallow tray using palm kernel cake as a substrate.
- Redmile-Gordon, M. A., Armenise, E., White, R. P., Hirsch, P. R., & Goulding, K. W. T. (2013). A comparison of two colorimetric assays, based upon Lowry and Bradford techniques, to estimate total protein in soil extracts. *Soil Biology and Biochemistry*, *67*, 166-173.
- Rees, D. A. (1977). Polysaccharides Shapes: Chapman and Hall: Science Press.
- Richmond, P. A. (1991). Occurrence and functions of native cellulose. In C. H. Haigler (Ed.), *Biosynthersis and Biodegradation of Cellulose* (pp. 5-23): Marcel Dekker, Inc. New York, USA.
- Robson, L. M., & Chambliss, G. H. (1984). Characterization of the Cellulolytic Activity of a *Bacillus* Isolate. *Applied and Environmental Microbiology*, 47(5), 1039-1046.
- Royer, J. C., & Nakas, J. P. (1991). Purification and characterization of two xylanases from *Trichoderma longibrachiatum*. *European Journal of Biochemistry*, 202(2), 521-529.
- Ryu, D. D. Y., & Mandels, M. (1980). Cellulases: Biosynthesis and applications. *Enzyme and Microbial Technology*, 2(2), 91-102.
- Salminen, S., Beighton, M. A., Benno, Y., & Gorbach, S. L. (2004). Lactic Acid Bacteria in Health and Disease. In S. Salminen & V. A. Wright (Eds.), *Lactic Acid Bacteria: Microbiology and Functional Aspects* (pp. 211-253): Marcel Dekker, Inc.
- Samanta, A. K., Kolte, A. P., Senani, S., Sridhar, M., & Jayapal, N. (2011). A simple and efficient diffusion technique for assay of endo β-1, 4-xylanase activity. *Brazilian Journal of Microbiology*, *42*(4), 1349-1353.
- Schurz, J., Billiani, J., Honel, A., Eigner, W. D., Janosi, A., Hayn, M., & Esterbauer, H. (1985). Reaction-mechanism and structural-changes at enzymatic degradation of cellulose by *Trichoderma reesei* cellulase. *Acta Polymerica*, 36(2), 76-80.
- Seo, J, Park, T. S., Kim, J. N., Ha, J. K., & Seo, S. (2014). Production of endoglucanase, Beta-glucosidase and xylanase by *Bacillus licheniformis* grown on minimal nutrient medium containing agriculture residues. *Asian-Australasian Journal of Animal Sciences*, 27(7), 946.
- Shah, N. P. (2000). Probiotic Bacteria: Selective Enumeration and Survival in Dairy Foods. *Journal of Dairy Science*, *83*(4), 894-907.
- Sindhu, I., Chhibber, S., Capalash, N., & Sharma, P. (2006). Production of cellulasefree xylanase from *Bacillus megaterium* by solid state fermentation for biobleaching of pulp. *Current microbiology*, *53*(2), 167-172.
- Sivashanmugam, A., Murray, V., Cui, C., Zhang, Y., Wang, J., & Li, Q. (2009). Practical protocols for production of very high yields of recombinant proteins using *Escherichia coli*. *Protein Science*, *18*(5), 936-948.
- Sivasothy, K. (2000). *Advances in Oil Palm Research* (Y. Basiron, B. S. Jalani & K. W. Chan Eds. Vol. 1): Malaysian Palm Oil Board, Kuala Lumpur.
- Subramaniyam, R., & Vimala, R. (2012). Solid state and submerged fermentation for the production of bioactive substances: A comparative study. *International Journal of Scencei and Nauret, 3*, 480-486.

- Subramaniyan, S., Prema, P., & Ramakrishna, S. V. (1997). Isolation and screening for alkaline thermostable xylanases. *Journal of basic microbiology*, *37*(6), 431-437.
- Sukumaran, R. K. (2009). Bioethanol form Lignocellulosic Biomass: Part II Production of Cellulases and Hemicellulases. In P. A. (Ed.), *Handbook of Plant-Based Biofuels* (pp. 141-158): Taylor & Francis Group.
- Sukumaran, R. K., Singhania, R. R., & Pandey, A. (2005). Microbial cellulasesproduction, applications and challenges. *Journal of Scientific and Industrial Research*, 64(11), 832.
- Sunde, E. P., Setlow, P., Hederstedt, L., & Halle, B. (2009). The physical state of water in bacterial spores. *Proceedings of the National Academy of Sciences*, 106(46), 19334-19339.
- Talbot, G., & Sygusch, J. (1990). Purification and characterization of thermostable beta-mannanase and alpha-galactosidase from *Bacillus stearothermophilus*. *Applied and Environmental Microbiology, 56*(11), 3505-3510.
- Thomas, L., Larroche, C., & Pandey, A. (2013). Current developments in solid-state fermentation. *Biochemical Engineering Journal, 81*, 146-161.
- Tuan Lah, T. N., Rahman, N. N. N. A., Hasnan, N. J., Ben Name, M. M., Nagao, H., & Kadir, M. O. A. (2012). Cellulase Activity in solid state fermentation of palm kernel cake with *Trichoderma* sp. *Malaysian Journal of Microbiology*, 8(4), 235-241.
- Weimer, P. J. (1991). Quantitative and Semiquatitative Measurement of Cellulose Biodegradation. In C. H. Hailger & P. J. Weimer (Eds.), *Biosynthesis and Biodegradation of Cellulose* (pp. 263-291): Marcel Dekker, Inc.
- Wood, T. M. (1991). Fungal Cellulase. In C. H. Haigler & P. J. Weimer (Eds.), Biosynthesis and Biodegradation of Cellulose (pp. 491-533): Marcel Dekker, Inc.
- Woodward, J., & Wiseman, A. (1982). Fungal and other β-d-glucosidases Their properties and applications. *Enzyme and Microbial Technology*, *4*(2), 73-79.
- Yin, L. J., Lin, H. H., & Xiao, Z. R. (2010). Purification and characterization of a cellulase from *Bacillus subtilis* YJ1. *Journal of Marine Science and Technology*, 18(3), 466-471.