



UNIVERSITI PUTRA MALAYSIA

***ISOLATION, SCREENING AND CHARACTERIZATION OF
BIOSURFACTANT PRODUCING MICROORGANISMS***

SITI ZULAIKHA SURYATI HALIM

FBSB 2015 43

**ISOLATION, SCREENING AND CHARACTERIZATION OF BIOSURFACTANT
PRODUCING MICROORGANISMS**

By:

SITI ZULAIKHA SURYATI HALIM

Thesis Submitted to the Faculty of Biotechnology and Biomolecular Sciences,
Universiti Putra Malaysia, in fulfilment of the requirement for the Degree of Bachelor
of Science (Honours) Biotechnology

2015

**FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES
UNIVERSITI PUTRA MALAYSIA**

Date:

LETTER OF PERMISSION

It is thereby to state that I, SITI ZULAIKHA SURYATI HALIM (Matric No: 162562) have done a final year project entitled "**Isolation, Screening and Characterization of Biosurfactant Producing Microorganisms**" under supervision of Professor Dr. Suraini Abd. Aziz from the Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

I hereby give permission to my supervisor to write and prepare manuscript from the results of this research to be published in any form, if I do not do so in six (6) months from the date above, in condition that my name is also added as one of the article's authors. The arrangement of the name depends on the supervisor herself.

Yours sincerely,

.....
(SITI ZULAIKHA SURYATI HALIM)

FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES
UNIVERSITI PUTRA MALAYSIA

APPROVAL SHEET

This thesis entitled “**Isolation, Screening and Characterization of Biosurfactant Producing Microorganisms**” is submitted by SITI ZULAIKHA SURYATI HALIM (Matric No: 162562) in fulfilment of the requirement for the Degree of Bachelor of Science (Honours) Biotechnology in Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

Approved by,

.....
(Professor Dr. Suraini Abd. Aziz)
Project Supervisor
Department of Bioprocess Technology
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia

Date:

ABSTRACT

Abstract of thesis presented to the Faculty of Biotechnology and Biomolecular Sciences in fulfilment of the requirement for the Degree of Bachelor Science of (Honours) Biotechnology

ISOLATION, SCREENING AND CHARACTERIZATION OF BIOSURFACTANT PRODUCING MICROORGANISMS

By:

SITI ZULAIKHA SURYATI HALIM

June 2015

Supervisor: Professor Dr. Suraini Abd. Aziz

Faculty: Faculty of Biotechnology and Biomolecular Sciences

Biosurfactants are amphiphilic compounds comprising of a non-polar (hydrophobic) group and a polar (hydrophilic) group which are produced by microorganisms. Biosurfactants play an important role in bioremediation processes due to their effectiveness as dispersion and bioremediation agents as well as their environmentally characteristics, such as low toxicity and high biodegradability than chemical surfactants. However, the high production cost of biosurfactants is a major drawback in the industry. Used cooking oil is one of the alternative cheap substrate for biosurfactant production. The aim of this study is to isolate, screen and characterize biosurfactant-producing bacteria from the sources of used cooking oil, contaminated soil, and palm oil mill effluent (POME). Four strains which designated as HIO 1, HIP 1, HIP 2 and HIP 3 were able to produce biosurfactant from used cooking oil. Identification using 16S rRNA reveals that HIP 1 and HIP 2 are *Lysinibacillus* strain while HIP 3 and HIO 1 are strains of *Bacillus*. HIP 3 has the biggest oil displacement area with 92.12 cm² when tested using oil spreading assay. All the bacterial strains were identified as the biosurfactants producer.

ABSTRAK

Abstrak tesis yang dikemukakan kepada Fakulti Bioteknologi dan Sains Biomolekul sebagai memenuhi sebahagian daripada keperluan untuk Bacelor Sains (Kepujian) Bioteknologi

PENGASINGAN, PENYARINGAN DAN PENCIRIAN MIKROORGANISMA YANG MENGHASILKAN BIOSURFAKTAN

Oleh:

SITI ZULAIKHA SURYATI HALIM

Jun 2015

Penyelia: Professor Dr. Suraini Abd. Aziz

Fakulti: Fakulti Bioteknologi dan Sains Biomolekul

Biosurfaktan adalah sebatian amfifilik yang terdiri daripada kumpulan bukan kutub (hidrofobik) dan kumpulan kutub (hidrofilik) yang dihasilkan oleh mikroorganisma. Biosurfaktan memainkan peranan penting dalam proses bioremediasi disebabkan oleh keberkesanannya sebagai agen penyebaran dan bioremediasi dan juga disebabkan oleh ciri-ciri alam seperti kurang toksik dan tinggi biodegradasi berbanding surfaktan kimia. Namun, kos penghasilan biosurfaktan yang tinggi menjadi kelemahan utama untuk diaplikasikan di dalam industri. Minyak masak terpakai adalah satu substrat alternatif yang murah untuk penghasilan biosurfaktan. Tujuan kajian ini adalah untuk mengasing, menyaring dan mencirikan bakteria yang menghasilkan biosurfaktan dari sumber minyak masak terpakai, tanah yang tercemar dan efluen kilang minyak kelapa sawit (POME). Empat strain yang dinamakan sebagai HIO 1, HIP 1, HIP 2 dan HIP 3 mampu menghasilkan biosurfaktan dari minyak masak terpakai. Pengenalpastian menggunakan 16S rRNA menunjukkan bahawa HIP 1 dan HIP 2 adalah strain *Lysinibacillus* manakala HIP 3 dan HIO 1 adalah strain *Bacillus*. HIP 3 menunjukkan kawasan minyak tersesar yang paling besar, iaitu 92.12 cm² apabila diuji dengan penilaian minyak merebak. Semua strain bakteria dikenal pasti sebagai penghasil biosurfaktan.

ACKNOWLEDGEMENTS

Alhamdulillah, praise to Allah SWT, whom with His willing I was able to complete this final year project. In this opportunity, I would like to express my insightful appreciation to my project supervisor, Professor Dr. Suraini Abd. Aziz for her non-stop guidance, observing and constant encouragement throughout the course of this thesis. A mountain of thanks to my co-supervisor, Miss Nurul Hanisah Md Badrul Hisham for her genuine support, valuable information and guidance in which helping me in completing this task through various phases. I am grateful to all Master and PhD students of Biomass Technology Centre (BTC) and laboratory assistants of Makmal Pengajaran Pusat, who had given full cooperation and assistance in using the equipments and apparatus in the laboratory. I am also grateful for their readiness to assist me in improving my laboratory skills and laboratory management practices. Lastly, I would like to thank all my colleagues and family for their absolute support from the beginning until the end of the project.

TABLE OF CONTENTS

	Page
LETTER OF PERMISSION	i
APPROVAL SHEET	ii
ABSTRACT	iii
ABSTRAK	iv
ACKNOWLEDGEMENTS	v
TABLE OF CONTENTS	vi
LIST OF TABLES	ix
LIST OF FIGURES	x
LIST OF ABBREVIATIONS	xi
CHAPTER	
1.0 INTRODUCTION	1
2.0 LITERATURE REVIEW	
2.1 Biosurfactants	4
2.1.1 Properties of biosurfactant	4
2.1.2 Comparison between biosurfactant and chemical surfactant	6
2.2 Type of biosurfactants and microorganisms involved	8
2.2.1 Glycolipid	10
2.2.2 Lipopeptides and lipoprotein	13
2.2.3 Phospholipids and fatty acid biosurfactants	15
2.2.4 Polymeric biosurfactants	15
2.2.5 Particulate biosurfactants	16
2.3 Biosynthesis of biosurfactant	16
2.3.1 Type of substrates	19
2.4 Applications of biosurfactant	21
2.4.1 Food industry	21
2.4.2 Health care, cosmetic and therapeutic industry	21
2.4.3 Environmental application	22
2.5 Market demand	23

2.6	Concluding remarks	24
3.0	MATERIALS AND METHODS	
3.1	Samples collection	25
3.2	Isolation of potential microorganisms for biosurfactant production	25
3.3	Screening of microorganisms producing biosurfactant	26
3.3.1	Bushnell and Haas agar screening	26
3.3.2	Oil spreading technique	27
3.4	Characterization of the microorganisms producing biosurfactant	29
3.4.1	Gram staining and morphology analysis	29
3.4.2	16S rRNA sequence analysis	29
3.4.2.1	Genomic DNA Extraction	29
3.4.2.2	Polymerase Chain Reaction (PCR) amplification of 16S rRNA gene	29
3.4.2.3	Agarose gel electrophoresis	30
3.4.2.4	16S rRNA gene sequence analysis	31
3.4.3	Growth profiling	31
3.5	Analytical method	31
3.5.1	Cell optical density (OD)	31
4.0	RESULTS AND DISCUSSION	
4.1	Isolation and screening of biosurfactant producing microorganisms	32
4.1.1	Isolation of biosurfactant producing microorganisms	32
4.1.2	Bushnell and Haas agar screening	32
4.1.3	Oil spreading assay	35
4.2	Characterization of bacteria producing biosurfactant	37

4.2.1	Gram staining technique	37
4.2.2	Identification of bacteria using 16S rRNA sequence analysis	39
4.2.3	Growth profiling of the bacterial isolates	43
5.0	CONCLUSION AND RECOMMENDATIONS	
5.1	Conclusion	48
5.2	Recommendations	49
	REFERENCES	50
	APPENDICES	59



LIST OF TABLES

Table	Page
2.1 Biosurfactants classification and microorganisms involved	9
3.1 Locations of samples collection	25
3.2 The composition of Bushnell and Haas agar	26
3.3 The composition of mineral salt medium	27
3.4 The composition of trace element	28
3.5 The reaction mixture composition for the Polymerase Chain Reaction (PCR)	30
3.6 The PCR conditions and the time taken	30
4.1 Microorganisms screened using Bushnell and Haas agar	33
4.2 Oil spreading assay of potentially biosurfactant producing bacteria	36
4.3 Gram staining result of bacterial isolates	37
4.4 Identification of the bacteria producing biosurfactant isolates by 16S rRNA sequence analysis	42

LIST OF FIGURES

Figure	Page
2.1 Mechanism of hydrocarbon solubilization within biosurfactant micelles	5
2.2 The chemical structure of trehalolipid	11
2.3 Chemical structure of rhamnolipid	12
2.4 Chemical structure of sophorolipids and derivatives	13
2.5 The chemical structure of surfactin	14
2.6 The chemical structure of lichenysin A	15
2.7 Potential biosurfactant biosynthetic pathways in microorganisms	17
4.1 Positive Bushnell and Haas agar screening	34
4.2 Oil displacement assay	35
4.3 Gram positive	38
4.4 Phylogenetic tree for HIP 1	39
4.5 Phylogenetic tree for HIP 2	40
4.6 Phylogenetic tree for HIP 3	40
4.7 Phylogenetic tree for HIO 1	41
4.8 Time courses of cell growth for HIP 1	43
4.9 Time courses of cell growth for HIP 2	44
4.10 Time courses of cell growth for HIP 3	45
4.11 Time courses of cell growth for HIO 1	46

LIST OF ABBREVIATIONS

μL	Microlitre
πr^2	Pi Radius Squared
BLAST	Basic Local Alignment Search Tool
BS	Biosurfactant
BTC	Biomass Technology Centre
CAGR	Compound Annual Growth Rate
CMC	Critical Micelle Concentration
dH ₂ O	Distilled Water
DNA	Deoxyribonucleic acid
g	Gram
g/cm	Gram per Centimetre
g/L	Gram per Litre
h	Hour
HLB	Hydrophilic and Lipophilic Balance
IgG	Human Immunoglobulin G
min	Minute
kg	Kilogram
MEOR	Microbial Enhanced Oil Recovery
MES	Methyl Ester Sulfonate
mg	Milligram
mg/L	Milligram per Litre
mL	Millilitre

MSM	Mineral Salt Medium
NCBI	National Center for Biotechnology Information
nm	Nanometre
OD	Optical Density
OOME	Olive Oil Mill Effluent
PAHs	Polycyclic Aromatic Hydrocarbons
PCR	Polymerase Chain Reaction
PDA	Potato Dextrose Agar
POME	Palm Oil Mill Effluent
RoW	Rest of the World
rRNA	Ribosomal Ribonucleic Acid
sp.	Species
USD	United States Dollar
UPM	Universiti Putra Malaysia
v/v	Volume over Volume
YPD	Yeast Potato Dextrose

CHAPTER 1

INTRODUCTION

Oil spillage and oil pollution in marine environment have been the main risk to the ecology including the ocean life as well as to the human being through the transmission of toxic organic materials including polycyclic aromatic hydrocarbons (PAHs) into the food chain (Dasgupta *et al.*, 2013). Deziel *et al.* (1996) stated that most of PAHs are carcinogenic, thus giving a potential threat to the marine animals and also human health. To encounter this problem, bioremediation technology has become a global phenomenon. Mueller *et al.* (1996) defines bioremediation as the process of degrading organic wastes biologically under controlled conditions to a harmless state, or to the points below concentration limits recognized by regulatory authorities. From the definition, bioremediation is the utilization of living organisms, predominantly microorganisms, to reduce the environmental contaminants into less toxic forms.

Principally, Vidali (2001) describes that bioremediation uses naturally occurring bacteria and fungi or plants in order to degrade or detoxify hazardous substances that can endanger human health and the environment. As being studied by Saravanan & Vijayakumar (2012), hydrocarbons degradation using microorganisms has a really important role in the environmental pollution treatment because most of the hydrocarbons are insoluble in water. Hydrocarbon degrading microorganisms produce biosurfactants which are surface active compounds with different chemical nature and molecular size. This compound has a very unique property in which it can reduce the surface tension of the hydrophobic water-insoluble substrates, thus improve their bioavailability as well as the rate of bioremediation.

Nitschke & Costa (2007) found that the world attention is triggered to microbial-derived surface-active compounds basically because of their low toxicity and biodegradable nature compared to synthetic surfactants which are produced chemically derived from petroleum that possess toxic property. Biosurfactants are amphiphilic compounds, consisting of two parts, a non-polar (hydrophobic) group and a polar (hydrophilic) moiety which are produced by microorganisms, such as

bacteria, yeast and fungi. The hydrophobic moiety comprises of saturated, unsaturated, and hydroxylated fatty acids whereas hydrophilic group contains mono-, oligo-, or polysaccharides, proteins or peptides.

From the previous studies, many potential microorganisms have been found to be producing biosurfactants. However, the abundant part of biosurfactants is produced by bacteria and only few are produced by fungi and yeast (Kitamoto *et al.*, 2002). Microbial surfactants are categorized by their chemical composition and microbial origin (Nitschke & Costa, 2007) and as being suggested by Rosenberg & Ron (1999), biosurfactants can be grouped into low sub-atomic mass particles which proficiently lower surface and interfacial strain; and high sub-atomic polymers which are more compelling as emulsion-balancing out agents. Cameotra *et al.* (2010) and Desai & Banat (1997) described that low-sub-atomic mass particles are including glycolipids, lipopeptides and phospholipids, and fatty acids whereas high-mass surfactants incorporate polymeric surfactants and particulate surfactants.

Generally, Makkar & Cameotra (1999) stated that hydrocarbons have been the substrates of choice for the production of biosurfactants. As being stated by Carey (2014), hydrocarbons are any class of organic chemical compounds that comprised of carbon (C) and hydrogen (H) elements only. The carbon atoms grouped to form the compound framework while the hydrogen atoms attached to them in diverse configurations. The examples of hydrocarbons include alkanes, alkenes, alkynes and arenes. However, many industries such as cosmetics, food and pharmaceutical industries cannot accept hydrocarbon as the substrate. Therefore, the usage of non-hydrocarbon substrates such as oils, fats, carbohydrates and glycerol are explored widely. Cameotra & Makkar (1998) suggested that the success of biosurfactant production relies on the improvement of economical processes and the utilization of cheaper raw materials in which make up for 10–30% of the overall cost. As being reported by Makkar & Cameotra (1999), some research had been conducted in finding the best way to reduce the cost and the solutions found is by using wastes either free or carry a cost credit to the environmental benefit.

Vegetable oils are used in the worldwide abundantly for many cooking processes, for instance, in Europe, about 17 million tons of vegetable oils are used annually and for each passing year, the amount raises approximately by 2% (Agriculture and Food Development, 2000). Hanisah *et al.* (2013) reported that the waste cooking oil produced by Europe was approximately 700,000-1,000,000 tonnes per annum, inclusive of the oils from snack food and French fries. As being studied by Ismail (2005), it was estimated that Asia countries such as China, Malaysia, Indonesia, Thailand, Hong Kong and India produced 40,000 tonnes waste cooking oil per year. Based on these facts, it can be concluded that there are a huge amount of waste frying oil resources will be produced in the next few years.

From the study conducted by Khalisanni *et al.* (2008), used cooking oils possess high fixed carbon about 99.92%, indicating that it is rich in carbon content. The high carbon content of used cooking oils makes it the best candidate to be the raw material in the production of biosurfactants. Other than that, used cooking oils possess long chain of palmitic acid and oleic acid which is potential to be cracked by thermal cracking or catalytic cracking for possible formation of hydrocarbon chain. As from these chemical characteristics of used cooking oil, it is greatly potential to be used as the sole substrate for the production of biosurfactant that can reduce the overall production cost because the substrate can be obtained without any charge. Besides, used cooking oil is chosen because it is a renewable feedstock as being proposed by Mulligan *et al.* (2014).

The objectives of the study are:

1. To isolate and screen the potential biosurfactants producing microorganisms
2. To characterize the potential biosurfactants producing microorganisms

REFERENCES

- Abouseoud, M., Maachi, R. and Amrane, A. (2007). Biosurfactant production from olive oil by *Pseudomonas fluorescens*. In *Communicating Current Research and Educational Topics and Trends in Applied Microbiology* (pp. 340–347).
- Agricultural and Food Development Authority (2000). *Waste oils and fats as biodiesel feedstocks: An assessment of their potential in the EU, ALTENER Program NTB-NETT Phase IV, Task 4, Final Report*.
- Al-Bahry, S. N., Al-Wahaibi, Y. M., Elshafie, A. E., Al-Bemani, A. S., Joshi, S. J., Al-Makhmari, H. S. and Al-Sulaimani, H. S. (2013). International biodeterioration & biodegradation biosurfactant production by *Bacillus subtilis* B20 using date molasses and its possible application in enhanced oil recovery. *International Biodeterioration and Biodegradation*, 81, 141–146.
- Asselineau, C. and Asselineau, J. (1978). Trehalose-containing glycolipids. *Progress in the Chemistry of Fats and Other Lipids*, 16, 59–99.
- Atlas, R. M. (2010). Atlas Oil Agar. In *Handbook of Microbiological Media, Fourth Edition* (Fourth Edi.). United States, US: CRC Press.
- Batista, S., Mounteer, A., Amorim, F. and Totola, M. (2005). Isolation and characterization of biosurfactant/bioemulsifier-producing bacteria from petroleum contaminated sites. *Bioresource Technology*, 97(2005), 73–82.
- Beveridge, T. J. (2001). Use of the Gram stain in microbiology. *Biotechnology and Biochemistry*, 76(3), 111-118.
- Boström, C. E., Gerde, P., Hanberg, A., Jernström, B., Johansson, C., Kyrklund, T., Rannug, A., Tornqvist, M., Victorin, K. and Westerholm, R. (2002). Cancer risk assessment, indicators, and guidelines for polycyclic aromatic hydrocarbons in the ambient air. *Environmental health perspectives*, 110(Suppl 3), 451.
- Bruckner, M. Z. (2012). Gram staining. *Microbial Life Educational Resources*. Retrieved from http://serc.carleton.edu/microbelife/research_methods/micro on 26/05/15
- Brusseau, M. L., Oleen, J.K., Santamaria, J., Cheng, L., Orosz-Coghlan, P., Chetochine, A. S., Blanford, W. J., Rykwald, C.P. and Gerba, P. (2005). Transport of *Microsporidium encephalitozoon* intestinales spores in sandy porous media. *Water Resolution*, 39, 3636–3642.
- Busscher, H. J., van Hoogmoed, C. G., Geertsema-Doornbush, G. I., van der Kuij-Booij, M. and van der Mei, H. C. (1996). Biosurfactants from Thermophilic Dairy Spectrococci and Their Potential Role in the Fouling Control of Heat Exchanger Plates. *Journal of Industrial Microbiology and Biotechnology*, 16(1), 15–21.
- Carey, F. A. (2014). Hydrocarbon: Chemical compound. Retrieved from <http://global.britannica.com/EBchecked/topic/278321/hydrocarbon> on 27/4/15.

- Cameotra, S. C., Makkar, R. S., Kaur, J. and Mehta, S. K. (2010). Chapter 20: Synthesis of biosurfactants and their advantages to microorganisms and mankind. In *Biosurfactants*, India: Landes Bioscience.
- Cameotra, S. S. and Makkar, R. S. (1998). Synthesis of biosurfactants in extreme conditions. *Applied Microbiology Biotechnology*, 50, 520–529.
- Cameotra, S. S. and Makkar, R. S. (2004). Recent application of biosurfactants as a biological and immunological molecules. *Current Opinion in Microbiology*, 7, 262–266.
- Converti, A., Aliakbarian, B., Dominguez, J. M., Bustos Vasquez, G. and Perego, P. (2010). Microbial production of biovanillin. *Brazilian Journal of Microbiology*, 41, 519–530.
- Dasgupta, D., Ghosh, R. and Sengupta, T. K. (2013). Biofilm-mediated enhanced crude oil degradation by newly isolated *Pseudomonas* species. *ISRN Biotechnology*, 2013, 121-124.
- Dehority, B. A. and Tirabasso, P. A. (2000). Antibiosis between ruminal bacteria and ruminal fungi. *Applied Environmental Microbiology*, 66, 2921–2927.
- Desai, J. D. and Banat, I. M. (1997). Microbial production of surfactants and their commercial potential. *Microbiology and Molecular Biology Reviews*, 61(1), 47–64.
- Deziel, E., Paquette, G., Villemur, R., Lepine, F. and Bisailon, J. (1996). Biosurfactant production by a soil *Pseudomonas* strain growing on polycyclic aromatic hydrocarbons. *Applied and Environmental Microbiology*, 62(6), 134–136.
- Domingues, P. M., Louvado, A., Oliveira, V., Coelho, F. J. C. R., Almeida, A., Gomes, N. C. M. and Cunha, A. (2013). Selective cultures for the isolation of biosurfactant producing bacteria: Comparison of different combinations of environmental inocula and hydrophobic carbon sources. *Preparative Biochemistry and Biotechnology*, 37–41.
- Edwards, K. R., Lepo, J. E. and Lewis, M. A. (2003). Toxicity comparison of biosurfactants and synthetic surfactants used in oil spill remediation to two estuarine species. *Marine Pollution Bulletin*, 46, 1309–1316.
- Feliciano, T. (2009). Quarterly Chemical Report: Lichenysin A and Surfactin. *American Chemical Society Division of Chemical Toxicology*.
- Fu, S. L., Wallner, S. R., Bowne, W. B., Hagler, M. D., Zenilman, M. E., Gross, R. A. and Bluth, M. H. (2008). Sophorolipids and their derivatives are lethal against human pancreatic cancer cells. *Journal of Surgical Research*, 148(1), 77–82.
- Gan, B. S., Kim, J., Reid, G., Cadieux, P. and Howard, J. C. (2002). *Lactobacillus fermentum* RC-14 inhibits *Staphylococcus aureus* infection of surgical implants in rats. *Journal of Infectious Diseases*, 185(9), 1369–1372.

- Gautam, K. K. and Tyagi, V. K. (2005). Microbial surfactants: A review. *Journal of Oleo Science*, 55, 155–156.
- George, S. and Jayachandran, K. (2013). Production and characterization of rhamnolipid biosurfactant from waste frying coconut oil using a novel *Pseudomonas aeruginosa* D. *Journal of applied microbiology*, 114(2), 373-383.
- Gnanaprakasam, A., Sivakumar, V. M., Surendhar, A., Thirumarimurugan, M. and Kannadasan, T. (2013). Recent strategy of biodiesel production from waste cooking oil and process influencing parameters: A review. *Journal of Energy*, 2013, 243-256.
- Grand View Research (2014). Biosurfactants market analysis by product (rhamnolipids, sophorolipids, MES, APG, sorbitan esters, sucrose esters) and segment forecast to 2020. Retrieved from <http://www.grandviewresearch.com/> on 26/05/15.
- Hamzah, A., Sabturani, N. S. and Radiman, S. (2013). Screening and optimization of biosurfactant production by the hydrocarbon-degrading bacteria. *Universiti Sains Malaysia*, 42(5), 615–623.
- Hanisah, K., Kumar, S. and Tajul, A. Y. (2013). The management of waste cooking oil: A preliminary survey. *Health and The Environmental Journal*, 4(1), 76-81.
- Horowitz, S. and Currie, J. K. (1990). Novel dispersants of silicon carbide and aluminium nitride. *Journal Dispersion Science Technology*, 11, 637–659.
- Hu, Y. and Ju, L.-K. (2001). Purification of lactonic sophorolipids by crystallization. *Journal of Biotechnology*, 87(3), 263–272.
- Innis, M. A., Gelfand, D. H., Sninsky, J. J. and White, T. J. (1990). Molecular biology. In *Amplifications or Applications*. Academic Press, 156-166.
- Ismail, R. (2005). Palm oil and palm olein frying applications. *Asia Pacific Journal of Clinical Nutrition*, 14(6), 414–419.
- Ismail, W., Al-Rowaihi, I. S., Al-Humam, A. A., Hamza, R. Y., El Nayal, A. M. and Bououdina, M. (2013). Characterization of a lipopeptide biosurfactant produced by a crude-oil-emulsifying *Bacillus* sp. I-15. *International Biodeterioration and Biodegradation*, 84, 168-178.
- Isoda, H., Kitamoto, D., Shinmoto, H., Matsumura, M. and Nakahara, T. (1997). Microbial extracellular glycolipid induction of differentiation and inhibition of the protein kinase C activity of human promyelocytic leukemia cell line HL60. *Bioscience Biotechnology Biochemistry*, 61, 609–614.
- Jarvis, F. G. and Johnson, M. J. (1949). A glycolipid produced by *Pseudomonas aeruginosa*. *Journal American Chemistry Society*, 71, 4124–4126.
- Joshi, S. J. and Desai, A. J. (2010). Biosurfactant's role in bioremediation of naphthalene and fermentative production. In *Biosurfactants* (pp. 222-235). New York, NY: Springer.

- Juwarkar, A. A., Dubey, K. V., Nair, A. and Singh, S. K. (2008). Bioremediation of Multi-metal contaminated soil using biosurfactant- A novel approach. *Indian Journal Microbiology*, 48, 142–146.
- Kapadia Sanket, G. and Yagnik, B. N. (2013). Current trend and potential for microbial biosurfactants. *Asian Journal of Experimental Biological Sciences*, 4(1), 1–8.
- Karanth, N. G. K., Deo, P. G. and Veenanadig, N. K. (1999). Microbial production of biosurfactants and their importance. *Current Science*, 77(1), 116–126.
- Khalisanni, K., Khalizani, K., Rohani, M. S. and Khalid, P. O. (2008). Analysis of waste cooking oil as raw material for biofuel production. *Global Journal of Environmental Research*, 2(2), 81–83.
- Khire, J. M. and Khan, M. T. (1994). Microbially enhanced oil recovery (MEOR), importance and mechanism of MEOR. *Enzymatic Microbiology Technology*, 16, 170–172.
- Kitamoto, D., Isoda, H. and Nakahara, T. (2002). Functions and potential applications of glycolipid biosurfactants- From energy-saving materials to gene delivery carriers. *Journal of Bioscience and Bioengineering*, 94(3), 187–201.
- Kitamoto, D., Morita, T., Fukuoka, T., Konishi, M. and Imura, T. (2009). Self-assembling properties of glycolipid biosurfactants and their potential applications. *Current Opinion in Colloid & Interface Science*, 14(5), 315–328.
- Kosaric, N. (2001). Biosurfactants and their application for soil bioremediation. *Food Technology and Biotechnology*, 39(4), 295–304.
- Lang, S. and Philip, J. C. (1998). Surface-active lipids in rhodococci. *Antonie van Leeuwenhoek*, 74, 59–70.
- Lang, S. and Wulbrandt, D. (1999). Rhamnolipids biosynthesis, microbial production and application potential. *Applied Microbiology and Biotechnology*, 51, 22–32.
- Lima, T. M. S., Procópio, L. C., Brandão, F. D., Carvalho, A. M. X., Tótola, M. R. and Borges, A. C. (2011). Biodegradability of bacterial surfactants. *Biodegradation*, 22(3), 585–92. doi:10.1007/s10532-010-9431-3
- Lin, S. C. (1996). Review biosurfactants: Recent advances. *Journal Chemical Technology Biotechnology*, 66, 109–120.
- Maier, R. M. and Soberon-Chavez, G. (2000). *Pseudomonas aeruginosa* rhamnolipids: Biosynthesis and potential environmental applications. *Applied Microbiology Biotechnology*, 54, 625–633.
- Makkar, R. S. and Cameotra, S. S. (1999). Biosurfactant production by microorganisms on unconventional carbon sources. *Journal of Surfactants and Detergents*, 2(2), 237–241.

- Makkar, R. S. and Cameotra, S. S. (2002). An update on the use of unconventional substrates for biosurfactant production and their new applications. *Application Microbiology Biotechnology*, 58, 428–434.
- McInerney, M. J., Javaheri, M. and Nagle, D. P. J. (1990). Properties of the biosurfactant produced by *Bacillus licheniformis* strain JF-2. *Journal of Industrial Microbiology*, 5, 95–102.
- Mehta, S. K., Sharma, S., Mehta, N. and Cameotra, S. S. (2010). Biomimetic amphiphiles: Properties and potential use. In *Biosurfactants* (pp. 102-120). New York, NY: Springer.
- Morikawa, M., Hirata, Y. and Imanaka, T. (2000). A study on the structure-function relationship of lipopeptides biosurfactants. *Biochimica et Biophysica Acta* 1488, 211–218.
- Mueller, J. G., Cerniglia, C. E. and Pritchard, P. H. (1996). Bioremediation of environments contaminated by polycyclic aromatic hydrocarbons. *Bioremediation: Principles and Applications* (pp. 125–194). Cambridge University Press.
- Mukherjee, A. K. and Das, K. (2005). Correlation between diverse cyclic lipopeptides production and regulation of growth and substrate utilization by *Bacillus Subtilis* Strains in a particular habitat. *FEMS Microbiology Ecology*, (54), 479–489.
- Mulligan, C. N. (2005). Environmental applications for biosurfactants. *Environmental Pollution*, 133, 183–198.
- Mulligan, C. N., Sharma, S. K. and Mudhoo, A. (Eds.) (2014). *Biosurfactants: Research trends and applications*. United State, US: CRC Press.
- Neves, L. C. M., Oliveira, K. S., Kobayashi, M. J., Penna, T. C. V. and Converti, A. (2007). Biosurfactant production by cultivation of *Bacillus Atrophaeus* ATCC 9372 in semidefined glucose/casein-based media. *Applied Biochemistry and Biotechnology*, 137, 539–554.
- Nitschke, M. and Costa, S. G. V. A. O. (2007). Biosurfactants in food industry. *Trends in Food Science and Technology*, 18, 252–259.
- Nitschke, M., Ferraz, C. and Pastore, G. M. (2004). Selection of microorganisms for biosurfactant production using agroindustrial wastes. *Brazilian Journal of Microbiology*, 35, 81–85.
- Obayari, O. S., Ilori, M. O., Adebuseye, S. A., Oyetibo, G. O., Omotayo, A. E. and Amund, O. O. (2009). Degradation of hydrocarbons and biosurfactant production by *Pseudomonas* sp. strain LP1. *World Journal Microbiology Biotechnology*, 25, 1615–1623.
- Pacwa-Plociniczak, M., Płaza, G. A., Piotrowska-seget, Z. and Cameotra, S. S. (2011). Environmental applications of biosurfactants: Recent advances. *International Journal of Molecular Sciences*, 12, 633–654.

- Panagou, E. Z., Schillinger, U., Franz, C. M. and Nychas, G. J. E. (2008). Microbiological and biochemical profile of cv. Conservolea naturally black olives during controlled fermentation with selected strains of lactic acid bacteria. *Food Microbiology*, 25(2), 348-358.
- Pandey, A. and Anis, R. (2013). Optimization and characterization of biosurfactant producing microbes isolated from oil contaminated soil and expression of biosurfactant genes in *E. coli*. *International Journal of Pharmaceutical Research and Bio-science*, 2(2), 46-67.
- Peng, L., Huihui, L., Rong, L., Shunpeng, L. and Xing, H. (2009). Biodegradation of fomesafan by *Lysinibacillus* sp.ZB-1 isolated from soil. *Chemosphere*, 77, 1614–1619.
- Perfumo, A., Smyth, T. J. P., Marchant, R. and Banat, I. M. (2010). 47: Production and roles of biosurfactants and bioemulsifiers in accessing hydrophobic substrates. In *Handbook of Hydrocarbon and Lipid Microbiology*.
- Peypoux, F., Bonmatin, J. M. and Wallach, J. (1999). Recent trends in the biochemistry of surfactin. *Applied Microbiology Biotechnology*, (51), 553–563.
- Prakash, B. and Irfan, M. (2011). *Pseudomonas aeruginosa* is present in crude oil contaminated sites of Barmer region (India). *Journal of Bioremediation and Biodegradation*, 2 (129). 135-167.
- Rambaut, A. (2013). How to read a phylogenetic tree. Retrieved from http://epidemic.bio.ed.ac.uk/how_to_read_a_phylogeny on 01/014/15.
- Ramli, N. (2012). Development of a local bacterial isolate expressing cyclodextrin glycosyltransferase through molecular cloning approaches. Universiti Putra Malaysia.
- Rau, U., Hammen, S., Heckmann, R., Wray, V. and Lang, S. (2001). Sophorolipids: a source for novel compounds. *Industrial Crops Production*, 13:85–92.
- Raza, Z. A., Khan, M. S., Khalid, Z. M. and Rehman, A. (2006). Production of biosurfactant using different hydrocarbons by *Pseudomonas aeruginosa* EBN-8 mutant. *Zeitschrift Fur Naturforschung C-Journal of Biosciences*, 61, 87–94.
- Reid, G., Beuerman, D., Heinemann, C. and Bruce, A. W. (2001). Effect of oral probiotic *Lactobacillus* therapy on the vaginal flora and susceptibility to urogenital infections. *FEMS Immunological Medic Microbiology*, 32, 37–41.
- Robert, M., Mercade, M. E., Bosch, M. P., Parra, J. L., Espuny, M. J., Manresa, M. A. and Guinea, J. (1989). Effect of the carbon source on biosurfactant production by *Pseudomonas aeruginosa* 44T. *Biotechnology Letters*, 11, 871–874.
- Rodrigues, L., Banat, I. M., Teixeira, J. and Rosario, O. (2006). Biosurfactants : Potential applications in medicine. *Journal of Antimicrobial Chemotherapy*, (57), 609–618.
- Rollins, D. M. and Joseph, S. W. (2000). The gram stain. Retrieved from <http://life.umd.edu/classroom/bsci424> on 26/05/15.

- Rosenberg, E. and Ron, E. Z. (1999). High- and low-molecular-mass microbial surfactants. *Applied and Environmental Microbiology*, 52, 154–162.
- Rufino, R. D., de Luna, J. M., de Campos Takaki, G. M. and Sarubbo, L. A. (2014). Characterization and properties of the biosurfactant produced by *Candida lipolytica* UCP 0988. *Electronic Journal of Biotechnology*, 17(1), 34–38.
- Sanli, H., Canakci, M. and Alptekin, E. (2011). Characterization of waste frying oils obtained from different facilities. In *World Renewable Energy Congress* (pp. 479–485).
- Saravanan, V. and Vijayakumar, S. (2012). Isolation and screening of biosurfactant producing microorganisms. *Journal Academic Industrial Research*, 1(5), 1–5.
- Satpute, S. K., Bhawsar, B. D., Dhakephalkar, P. K. and Chopade, B. A. (2008). Assessment of different screening methods for selecting biosurfactant producing marine bacteria. *Indiana Journal Marine Science*, 37, 243–250.
- Satpute, S. K., Bhuyan, S. S., Pardesi, K. R., Mujumdar, S. S., Dhakephalkar, P. K., Shete, A. M. and Chopade, B. A. (2010). Molecular genetics of biosurfactant synthesis in microorganisms. In *Biosurfactants* (pp. 14-41). New York, NY: Springer.
- Sen, R. (2010). *Biosurfactants*. Edited by Ramkrishna Sen. Landes Bioscience and Springer + Business Media, LLC, USA.
- Sen, R. and Swaminathan, T. (2005). Characterization of concentration and purification parameters and operating conditions for the small-scale recovery of surfactin. *Process Biochemistry*, 40, 2953–2958.
- Shivaji, S., Srinivas, T. N. R. and Reddy, G. S. N. (2014). The Family Planococcaceae. *The Prokaryotes: Firmicutes and Tenericutes*, 303-351.
- Shmaefsky, B. R. (2004). Rhamnolipid expression: The potential for edible phytoremediation crops and beyond. *ISB News Report*, 1-3.
- Singh, P. and Cameotra, S. S. (2004). Potential applications of microbial surfactants in biomedical sciences. *Trends in Biotechnology*, 22(3), 142–146.
- Sudo, N., Aiba, Y., Takaki, A., Yu, X., Oyama, N., Koga, Y. and Kubo, C. (2000). Dietary nucleic acids promote a shift in Th1/Th2 balance toward Th1-Dominant immunity. *Clinical and Experimental Allergy*, 30, 979–987.
- Sutton, S. (2011). Measurement of microbial cells by optical density. *Journal of Validation Technology*, 17, 47-49.
- Syldatk, C. and Wagner, F. (1987). Production of biosurfactants. In *Biosurfactants and Biotechnology* (In N. Kosa., pp. 89–120). New York: Marcel Dekker Incorporation.

- Tarntip, R. and Sirichom, T. (2011). Isolation of proteolytic , lipolytic , and bioemulsifying bacteria for improvement of the aerobic treatment of poultry processing wastewater. *African Journal of Microbiology Research*, 5(30), 5493–5497.
- Thavasi, R., Jayalakshmi, S., Balasubramanian, T. and Banat, I. M. (2008). Production and characterization of a glycolipid biosurfactant from *Bacillus megaterium* using economically cheaper sources. *World Journal Microbiology Biot*, 24, 917–925.
- Thavasi, R., Sharma, S. and Jayalakshmi, S. (2011). Evaluation of screening methods for the isolation of biosurfactant producing marine bacteria. *Journal Petroleum and Environmental Biotechnology* 2011, 1–6.
- Transparency Market Research (2015). Specialty surfactants market and biosurfactants market: global scenario, raw material and consumption trends, industry analysis, size, share and forecast 2010 - 2018. Retrieved from <http://www.transparencymarketresearch.com/specialty-and-biosurfactants-mark> at 26/05/15.
- Urum, K., Grigson, S., Pekdemir, T. and McMenemy, S. (2006). A comparison of the efficiency of different surfactants for removal of crude oil from contaminated soils. *Chemosphere*, 62(9), 1403–1410.
- Van Bogaert, I. N. A., De Maeseneire, S. L., De Schampelaire, W., Develter, D., Soetaert, W. and Vandamme, E. J. (2007). Cloning, characterisation and functionality of the orotidine-5'-phosphate decarboxylase gene (URA3) of the glycolipid producing yeast *Candida bombicola*. *Yeast*, 24, 201–208.
- Van Haesendonck, I. P. H. and Vanzeveren, E. C. A. (2004). Rhamnolipids in Bakery Products. W.O. 2004/040984. *International Application Patent (PCT)*.
- Vater, J., Kablitz, B., Wilde, C., Franke, P., Mehta, N. and Cameotra, S. S.(2002). Matrix-assisted laser desorption ionization-time of flight mass spectrometry of lipopeptide biosurfactants in whole cells and culture filtrates of *Bacillus subtilis* C-1 isolated from petroleum sludge. *Applied Environmental Microbiology*, 68, 6210–6219.
- Vidali, M. (2001). Bioremediation . An overview *. *Pure Applied Chemistry*, 73(7), 1163–1172.
- Wayman, M., Jenkins, A. D. and Kormady, A. G. (1984). Biotechnology for oil and fat industry. *Journal American Oil Chemistry Society*, 61, 129–131.
- Wu, S. J., Hu, Z. H., Zhang, L. L., Yu, X. and Chen, J. M. (2009). A novel dichloromethane-degrading *Lysinibacillus sphaericus* strain Wh22 and its degradative plasmid. *Applied Microbiology Biotechnology*, 82, 731–740.
- Yakimov, M. M., Fredrickson, H. L. and Timmis, K. N. (1996). Effect of heterogeneity of hydrophobic moieties on surface activity of lichenysin A, a lipopeptide biosurfactant from *Bacillus licheniformis* BAS50. *Biotechnology Applied Biochemistry*, 23, 13–18.

Youssef, N. H., Duncan, K. E., Nagle, D. P., Savage, K. N., Knapp, R. M. and McInernery, M. J. (2004). Comparison of methods to detect biosurfactant production by diverse microorganisms *Journal Microbiology Methodology*, 56(3), 339–347.

Zhang, J., Gorkovenko, A., Gross, R. A., Allen, A. L. and Kaplan, D. L. (1997). Incorporation of 2-hydroxyl fatty acids by *Acinetobacter calcoaceticus* RAG-1 to tailor emulsan structure. *International Journal Biology Macromolecule*, 20, 9–21.

