

UNIVERSITI PUTRA MALAYSIA

OPTIMIZATION ON ENZYMATIC SACCHARIFICATION OF PRETREATED SAGO HAMPAS FOR FERMENTABLE SUGAR PRODUCTION

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OPTIMIZATION ON ENZYMATIC SACCHARIFICATION OF PRETREATED SAGO HAMPAS FOR FERMENTABLE SUGAR PRODUCTION



By

ROSFARIZA MD REJAB



Thesis submitted to the Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, as a fulfillment of the requirement for the degree of Bachelor of Science (Honours) Biotechnology

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FAKULTI BIOTEKNOLOGI DAN SAINS BIOMOLEKUL

UNIVERSITI PUTRA MALAYSIA

DATE :

LETTER OF PERMISSION

It is thereby to state that I, ROSFARIZA MD REJAB (Matric No: 162632) have done a final year project entitled "**Optimization on Enzymatic Saccharification of Pretreated Sago Hampas for Fermentable Sugar Production**" under supervision of Dr. Mohamad Faizal Ibrahim from the Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

I hereby give permission to my supervisor to write and prepare manuscript from the results of this research to be published in any form, if I do not do so in six (6) months from the date above, in condition that my name is also added as one of the article's authors. The arrangement of the name depends on the supervisor himself.

Yours sincerely,

(ROSFARIZA MD REJAB)

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APPROVAL SHEET

This thesis entitled "Optimization on Enzymatic Saccharification of Pretreated Sago Hampas for Fermentable Sugar Production" is submitted by ROSFARIZA MD REJAB (Matric No: 162632) in fulfillment of the requirement for the Degree of Bachelor of Science (Honours) Biotechnology in Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

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(Dr. Mohamad Faizal Ibrahim) Project Supervisor Department of Bioprocess Technology Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia

ABSTRACT

Abstract of thesis presented to the Faculty of Biotechnology and Biomolecular Sciences in fulfillment of the requirement for the Degree of Bachelor of Science (Honours) Biotechnology

OPTIMIZATION ON ENZYMATIC SACCHARIFICATION OF PRETREATED SAGO HAMPAS FOR FERMENTABLE SUGAR PRODUCTION



Supervisor : Dr. Mohamad Faizal Ibrahim

Faculty : Faculty of Biotechnology and Biomolecular Sciences

Nowadays, lignocellulosic feedstocks such as corn stover, oil palm biomass, rice straw, sugarcane bagasse and also sago biomass are used for the production of many valuable bioproducts. Sago biomass is a starch and lignocellulosic-based biomass produce from the processing of sago palm. There are three majors sago biomass produced which are sago bark (24 t), sago hampas (50 to 110 t) and sago waste water (300 to 1000 m³) generated daily in the sago processing mills. The waste generated usually discharged to the river without any treatment and this has contributed to high Biological Oxygen Demand (BOD), Chemical Oxygen demand (COD) and suspended solid in the river causing a serious environmental pollution.

Sago hampas contained 58% starch, 23% cellulose, 9.2% hemicellulose and also 4% of lignin. In this study, sago hampas were pretreated using the steaming process (autoclave) to remove the starch before proceeded with the enzymatic saccharification to produce fermentable sugar. The saccharification was conducted using a commercial cellulase enzyme (Acremonium Cellulase) and the optimization study was conducted using a two-level factorial design with four parameters which are substrate concentration (1-10%), enzyme concentration (10-50 FPU/g), agitation speed (50-250 rpm) and incubation period (24-96 h). The optimization of enzymatic saccharification of cellulose and hemicellulose of pretreated sago hampas by a

cellulase enzyme (10 FPU/g) had produced the highest fermentable sugar (5.94 g/L) at parameter : 10% of substrate concentration, 10 FPU/g of enzyme concentration at 250 rpm of agitation speed in 24 h of incubation period. Through the optimization, the production of fermentable sugar was increased up to 35.2% as the sugar produced before the optimization was about 3.85 g/L.



ABSTRAK

Abstrak tesis yang dikemukakan kepada Fakulti Bioteknologi dan Sains Biomolekul sebagai memenuhi sebahagian daripada keperluan untuk Bacelor Sains (Kepujian) Bioteknologi

PENGOPTIMUMAN SAKARIFIKASI ENZIM DARI HAMPAS SAGU TERAWAT UNTUK PENGHASILAN GULA FERMENTASI



Fakulti : Fakulti Bioteknologi dan Sains Biomolekul

Pada masa kini, bahan mentah lignoselulosa seperti batang jagung, biomas kelapa sawit, jerami padi, hampas tebu dan biomas sagu digunakan untuk pengeluaran banyak bioproduk berharga. Biomas sagu adalah kanji dan lignoselulosa berasaskan biomas hasil dari pemprosesan pokok sagu. terdapat tiga jenis biomas utama sagu dihasilkan iaitu kulit kayu sagu (24 t), hampas sagu (50 hingga 11 t) dan air sisa sagu (300 hingga 1000 m³) yang dihasilkan setiap hari di kilang-kilang pemprosesan sagu. Sisa yang dihasilkan biasanya dilepaskan ke sungai tanpa sebarang rawatan dan ini telah menyumbang kepada permintaan oksigen biologi (BOD), permintaan oksigen kimia (COD) dan pepejal terampai yang tinggi di dalam sungai menyebabkan pencemaran alam sekitar yang serius.

Hampas sagu mengandungi kanji 58%, 23% selulosa, hemiselulosa 9.2% dan 4% lignin. Di dalam pembelajaran ini, hampas sagu dirawat dengan menggunakan proses pengukusan (autoklaf) untuk mengeluarkan kanji sebelum diteruskan dengan pengoptimuman melalui sakarifikasi enzim menjadi gula fermentasi. Sakarifikasi ini dijalankan dengan menggunakan enzim selulase komersial (Acremonium Sellulase) dan kajian pengoptimuman telah dijalankan dengan menggunakan reka bentuk dua tingkat faktorial dengan empat parameter iaitu kepekatan substrat (1-10%), kepekatan enzim (10-50 FPU/g), kelajuan pergolakan (50-250 rpm) dan tempoh pengeraman (24-

96 h). Pengoptimuman sakarifikasi enzim daripada selulosa dan hemiselulosa daripada hampas sagu terawat oleh enzim selulase (10 FPU/g) telah menghasilkan gula beragi tertinggi (5.94 g/L) pada parameter : 10% daripada kepekatan substrat, 10 FPU/g kepekatan enzim pada 250 rpm kelajuan pergolakan dalam 24 jam tempoh pengeraman. Melalui pengoptimuman, penghasilan gula fermentasi telah meningkat kepada 35.2% di mana gula yang terhasil sebelum pengoptimuman adalah 3.85 g/L.



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Alhamdulillah, praise to Allah S.W.T, by the blessing of Him and His willing, I was able to complete the final year project and this thesis entitled 'Optimization on enzymatic saccharification of pretreated sago hampas for fermentable sugar production' for my bachelor degree. In completing this project, I would like to take this opportunity to express my deep appreciation to my project supervisor, Dr. Mohamad Faizal Ibrahim for his willing to guide, critic, monitoring and also gives a constant encouragement to me throughout of this project and writing this thesis. Besides, I would like to express my special thanks to my co-supervisor, Ms. Hazwani Husin for her valuable information, support and guide which helped me a lot in completing this project through the semester.

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LIST OF ABBREVIATIONS

ANN	Artificial neural network
ANOVA	Analysis of variance
BOD	Biological oxygen demand
COD	Chemical oxygen demand
DNS	Dinitrosalicyclic acid
FPU/g	Filter paper unit per gram
g/L	Gram per litre
h	Hour
Kg	Kilogram
М	Molar
m ³	Cubic metre
mg/L	Milligram per litre
min	Minute
MJ/Kg	Megajoule per kilogram
mL	Mililitre
MSG	Monosodium glutamate
OPEFB	Oil Palm Empty Fruit Bunch
rpm	Revolutions per minute
RSM	Response surface methodology
spp	several species
SPR	Sago pith residue
t	Tonnes
TSS	Total suspended solid
w/v	Weight per Volume
w/w	Weight per Weight

CHAPTER 1

INTRODUCTION

A carbohydrate, which also known as saccharides are one of the most important biomolecules in the world. The main function of these sugar biomolecules is to provide a basic backbone towards the energy storage, act as a fuel in the cellular functions and also render the metabolic processes stable. Besides, the carbohydrate also plays an important role in controlling the interaction of cells themselves (Flitsch *et al.*, 2003). Carbohydrate can be divided into three classes, there are consist of monosaccharides, disaccharides and polysaccharides. Monosaccharides refers to a single and simple sugar such as glucose, fructose and galactose that generally formed a basic unit for the carbohydrate, whereas disaccharides are referring to two unit monomers of a monosaccharide which combined and joined together with the removal of the water molecules. The combination of these two monosaccharides formed a sugar called maltose, lactose and sucrose. The complexity classes of sugar are called polysaccharides (Horton, 2008).

Polysaccharides consist of a long and complex chain of monosaccharides which bound together by a linkage called glycosidic bond. Basically, polysaccharides structure may be in a form of linear or highly branched. Starch and glycogen are the examples of linear polysaccharides, while cellulose and chitin are in the branched formed. In plants, there are a lot of polysaccharide insides which has a function in a plant metabolic process and it also provides a lot of advantages. Nowadays, polysaccharides that are broken down into sugars and undergo a several process such as fermented or chemically altered tends to produce a valuable product such as in biofuel and chemical industries. In the production of fuel and chemical, the biological conversion of cellulosic biomass is frequently used. According to Wayman *et al.* (1999), the conversion of cellulosic biomass in the meantime has become economically competitive as it gives an important strategy, environmental and economic advantages in industries. Thus, the biological conversion of these structures may give an advantage by producing a valuable product.

In recent years, there is an increasing towards the uses of agriculture residue for the production of a valuable product. It has been a lot of sustainable feedstocks that has been obtained from a variety of resources in order to extract it into a fermentable sugar through a bioconversion. A feedstock which contained a lignocellulosic material such as corn stover, oil palm empty fruit bunch (OPEFB), rice straw, sugarcane bagasse and also sago biomass (Linggang *et al.*, 2012; Awg-Adeni *et al.*, 2012; Jenol *et al.*, 2014) has widely been used for the production of the fermentable sugar. Nowadays, an abundant presence of sago biomass in the state of Sarawak (Linggang *et al.*, 2012) giving advantages to the residue to serve as an alternative substrate for the production of fermentable sugar. Sago biomass is a starch and lignocellulosicbased biomass produce from the processing of sago palm for the production of sago starch in industries. Currently, Malaysia is the world's biggest exporter of sago starch. Malaysia's exporting of sago starch annually is about 44 000 t, mainly in peninsular Malaysia, Japan, Singapore and other countries (Awg-Adeni *et al.*, 2012).

Sago is a starch extracted in the spongy center or pith of various tropical palms stems. Sago which also known as *Metroxylon* sagu can be found in tropical lowland forest and freshwater swamps across Southeast Asia and New Guinea. The plant has the ability to thrive in the harsh swampy peat environment. Generally in the production of sago, several types of waste have been generated as a byproduct such as sago bark, sago hampas and sago wastewater (Awg-Adeni *et al.*, 2012). Currently, the waste produced had been dumped into nearby river which then caused an increasing of the river pollution (Linggang *et al.*, 2013). Thus, it has contributed to an environmental pollution. A study by Ozawa *et al.* (1996), waste such as sago hampas had contains 58% starch, 23% cellulose, 9.2% hemicellulose and 3.9% lignin. According to Awg-Adeni *et al.* (2012), sago hampas consists of a starchy lignocellulosic by-product which is generated from the pith after the starch extraction.

There are two types of the saccharification process that commonly being used in converting the biomass into a fermentable sugar. The process can be done either by using the enzyme (enzymatic saccharification) or by using an acid (acid saccharification) as a catalyst. An enzymatic saccharification is a process involves specific enzymes that facilitate the cleavage of bond in molecules in the presence of water (Broda *et al.*, 1996). There are three major enzymes usually involves which are exoglucanase, endoglucanase and also β -glucosidase. During the hydrolysis process, a physical barrier of the plant (sago) cell wall is distrupted, thus, resulting the degrading of the biomass structure (Wyman, 1999). Nowadays, the production of fermentable sugar is started to be important as it is able to be further used for the production of biofuel, bio-based, bioenergy and chemical products in industries.

Therefore, the objective of this study is to optimize the conditions of enzymatic saccharification of a pretreated sago hampas as a substrate by using two-level factorial design in order to produce a fermentable sugar.

REFERENCES

- Abd-Aziz, S. (2002). Sago starch and its utilization. *Journal of Bioscience Bioengineering*, 9(6):525-529.
- Anita, S., Namita, S. and Narsi, R. (2009). Production of cellulases by *Aspergillus heteromorphus* from wheat straw under submerged fermentation. *Journal Environmental and Science Engineering*, 1:23-26.
- Awg-Adeni, D.S., Abd-Aziz, S., Bujang, K. and Hassan, M.A. (2010). Bioconversion of sago residue into value added products. *African Journal Biotechnology*, 9(14):2016-2021.
- Awg-Adeni, D.S., Bujang, K.B., Hassan, M.A. and Abd-Aziz, S. (2012). Recovery of glucose from residual starch of sago hampas for bioethanol production. *BioMed Research International*, 1-8.
- Bahrin, E.K., Baharuddin, A.S., Ibrahim, M.F., Razak, M.N.A., Sulaiman, A., Abd-Aziz, S., Hassan, M.A., Shirai, Y. and Nishida, H. (2012). Physiochemical property changes and enzymatic hydrolysis enhancement of oil palm empty fruit bunches treated with superheated steam. *Bioresources*, 7(2):1784-1801.
- Bakri, Y.P., Jacques, J. and Thonart, P. (2003). Xylanase production by *Penicillum* canescens 10-10c in solid state fermentation. *Applied Biochemistry and Biotechnology*, 108(1-3):737-748.
- Bertoft, E., Piyachomkwan, K., Chatakanonda, P. and Sriroth, K. (2008). Internal unit chain composition in amylopectin. *Carbohydrate Polymers*, 74:527-543.
- Bezerraa, M.A., Santelli, R.E., Oliveiraa, E.P., Villara, L.S. and Escaleira, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76:965-977.
- Broda, P., Birch, P.R.J., Brooks, P.R. and Sims, P.F.G. (1996). Molecular Microbiology. *Microbiology*, 19(5):923-932.
- Bujang, K. (2006). Potential of bioenergy from the sago industries in Malaysia. *Biotechnology*, 4.
- Bujang, K. (2010). Production and processing of sago : a food and fuel alternative. International Seminar on Sago & Spices for Food Security.
- Bujang, K.B. and Ahmad, F.B. (2000). Country report : Production and utilization of sago starch in Malaysia. International Sago Seminar. pp:1-8.
- Bujang, K.B., Apun, K. and Salleh, M.A. (1996). A study in the production and bioconversion of sago waste. Sago: The Future Source of Food and Feed. *Proceeding of the 6th International Sago Symposium*, Universitas Riau, Pekan Baru, Sumatra. Republic of Indonesia. 9-12 Dec, 1996.

- Bujang, K.B., Yusop, M.A. and Lopez-Real, J.M. (2004). Effects of aeration and agitation in the systematic treatment of sago effluent. *EXPO Malaysian Science and Technology Exhibition, COSTAM, Kuala Lumpur.* 5-7 October, 2004.
- Chew, T.Y. and Shim Y.L. (1993). Management of sago processing wastes, in : waste management in Malaysia current status and prospects for bioremediation. In B.G. Yeoh, K.S. Chee, S.M. Phang, Z. Isa, A. Idris and M. Mohamed (Eds.), *Ministry of Science, Technology and the Environment, Kuala Lumpur*.
- Chong, K.H., Law, P.L., Rigit, A.R.H., Baini, R. and Shanti, F.S. (2014). Sago bark as renewable energy. *Journal of civil engineering*, 29-34.
- Dimarogona, M., Topakas, E. and Christakopoulos, P. (2012). Cellulose degradation by oxidative enzymes. *Computational and Structural Biotechnology Journal*, 2(3): e201209015.
- Ebringerova, A., Hromadkova, Z. and Heinze, T. (2005). Hemicellulose. Advance in *Polymer Sciences*, 2193-2198.
- Ezeji, T.C. (2006). Biobutanol production by solventogenic *Clostridium* species : recent progress and challenges ahead. 19-22.
- Fan, L.T., Lee, Y.H. and Beardmore, D.R. (1981). The influence of major structural features of cellulose on rate of enzymatic hydrolysis. *Biotechnology and Bioengineering*, 23:419-424.
- Flach, M. (1983). Sago palm : *Metroxylon sagu* Rottb promoting the conservation and use of underutilized and neglected crops. FAO Plant Production and Protection paper 47. Rome : Bioversity International.
- Flitsch., Sabine, L., and Ulijn, R.V. (2003). Sugars tied to the spot. *Nature*, 421(6920):219-20.
- Gharpuray, M.M., Lee, Y-H. and Fan, L.T. (1983). Structural modification of lignocellulosics by pretreatments to enhance enzymatic hydrolysis. *Biotechnology and Bioengineering*, 25:157-172.
- Harmsen, P.F.H., Huijgen, W.J.J., Lopez, L.M.B. and Bakker, R.R.C. (2010). Literature Review of physical and chemical pretreatment processes for lignocellulosic biomass. *Food and Biobased Research*.
- Harris, J.H. (1990). Plant cell wall structure and development, in : Microbial and plant opportunities to improve lignocellulose utilization by ruminants. In D.E. Akin, L.G. Ljungdahl, J.R. Wilson and P.J. Harris (Eds.), pp 71-90. London, England.
- Held, P. (2012). Part II: Optimization of polymer digestion and glucose production in microplates. *Biofuel Research*.
- Hendriks, A.T.W.M. and Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*, 100(1):10-18.

- Horton, D. (2008). The Development of Carbohydrate Chemistry and Biology. *Carbohydrate Chemistry, Biology and Medical Applications*, 1-28.
- Ibrahim, M.F., Abd-Aziz, S., Razak, M.N.A., Phang, L.Y. and Hassan, M.A. (2012). Oil palm empty fruit bunch as alternative substrate for acetone–butanol–ethanol production by *Clostridium butyricum* EB6. *Applied Biochemistry and Biotechnology*, 166:1615-1625.
- Immanual, G., Dhanusha, R., Prema, P. and Palavesan. (2006). Effect of different growth parameters on endoglucanase enzyme activity by bacteria isolated from coir retting effluents of estuarine environment. *International Journal of Environmental Science and Technology*, 3(1):25-34.
- Jalil, S.N.J. (2013). Characterization and nutritive value of fermented sago by *Bacillus amyloliquefaciens* for aquaculture and livestock feed. Bachelor Thesis, Universiti Malaysia Sarawak.
- Jane, J-L. (2009). Structural features of starch granules II. *Chemistry and Technology*, 193-236.
- Janggu, U. (2011). Pretreatment of sago fibre for maximum fermentable sugars production. Master Thesis. Universiti Malaysia Sarawak.
- Janggu, U. and Bujang, K.B. (2009). Development of pre-treatment of sago fibres for recovery of bio-convertible polysaccharides (cellulose) for ethanol fermentation. *Proceedings of the 2nd Biotechnology Colloquium*. Universiti Malaysia Sarawak, Kota Samarahan, Sarawak, Malaysia. 14-15 April, 2009.
- Jenol, M.A., Ibrahim, M.F., Phang, L.Y., Md Salleh, M. and Abd-Aziz, S. (2014). Sago biomass as a sustainable source for biohydrogen production by *Clostridium butyricum* A1. *Bioresources*, 9(1):1007-1026.
- Johnson, D. (1977). Distribution of sago making in the old world. In Sago-76: Papers of the First International Sago Symposium, Ed. M.A. Tankooling, pp 65-75. Kuala Lumpur : Kemajuan Kanji Sdn. Bhd.
- Kamal, S.M.M., Mahmud, S.N., Hussain, S.A. and Ahmadun, F.R. (2007). Improvement on sago flour processing. *Journal of Chemical and Environmental Engineering*, 4:8-14.
- Khuri, A.I. and Mukhopadhyay, S. (2010). Response surface methodology. *WIREs Computational Statistic*, 2:128-149.
- Kirk-Otmer. (2001). Encyclopedia of chemical technology (4th ed.). Volume 5. ISBN: 978-0-471-15158-6.
- Kristensen, J.B., Felby, C. and Jorgensen, H. (2009). Determining yields in high solids enzymatic hydrolysis of biomass. *Applied Biochemistry and Biotechnology*, 156:127-132.
- Kumar, R., Singh, S. and Singh, O.V. (2008). Bioconversion of lignocellulosic biomass: Biochemical and molecular perspectives. *Journal of Industrial Microbiology and Biotechnology*, 35:377-391.

- Kumaran, S., Sastry, C.A. and Vikineswary, S. (1997). Laccase, cellulase and xylanase activities during growth of *Pleurotus sajor-caju* on sago hampas. *World Journal* of *Microbiology and Biotechnology*, 13:43-49.
- Kumoro, A.C., Ngoh, G.C., Hasan, M., Ong, C.H. and Teoh, E.C. (2008). Conversion of fibrous sago (*Mextroxylon Sagu*) waste into fermentables sugar via acid and enzymatic hydrolysis. *Asian Journal of Scientific Research*, 1:412-420.
- Kuo, C-K. and Lee, C-K. (2009). Enhancement of enzymatic saccharification of cellulose by cellulose dissolution pretreatments. *Carbohydrate Polymers*, 77:41-46.
- Laureano-Perez, L., Teymouri, F., Alizadeh, H. and Dale, B.E. (2005). Understanding factors that limit enzymatic hydrolysis of biomass. *Applied Biochemistry and Biotechnology*, 1081-1099.
- Lavarack, B.P., Griffin, G.J. and Rodman, D. (2002). The acid hydrolysis of sugarcane bagasse hemicellulose to produce xylose, arabinose, glucose and other products. *Biomass Bioenergy*, 23:367-380.
- Lenihin, P., Orozco, A., O'Neill, E., Ahmad, M.N.M., Rooney, D.W. and Walker, G.M. (2010). Dilute acid hydrolysis of lignocellulosic biomass. *Chemical Engineering Journal*, 156(2):395-403.
- Linggang, S., Phang, L.Y., Wasoh, M.H. and Abd-Aziz, S. (2012). Sago pith residue as an alternative cheap substrate for fermentable sugars production. *Applied Biochemistry and Biotechnology*, 16(1):122-31.
- Linggang, S., Phang, L.Y., Wasoh, M.H. and Abd-Aziz, S. (2013). Acetone-Butanol-Ethanol Production by *Clostridium acetobutylicum* ATCC 824 using sago pith residue hydrolysate. *Bioenergy Resources*, 6:321-328.
- Lynd, L.R., Weimer, P.J., Van Zyl, W.H. and Pretorius, I.S. (2002). Microbial cellulose utilization : fundamentals and biotechnology. *Microbiology*, 66:506-577.
- Mahamud, M.R. and Gomes, D.J. (2012). Enzymatic saccharification of sugarcane by the crude enzyme from indigenous fungi. *Journal of Scientific Research*, 1:227-238.
- Miller, G.L. (1959). Use of Dinitrosalycylic acid reagent for determination of reducing sugars. *Analytical chemistry*, 313:426-428.
- Nagata, Y. and Chu, K.H. (2003). Optimization of a fermentation medium using neural networks and genetic algorithms. *Biotechnology Letters*, 25:1837-1842.
- Nakamura, L. K. (1981). *Lactobacillus amylovorus*, a New Starch-Hydrolyzing Species from Cattle Waste-Corn Fermentation. *International Journal of Systematic Bacteriology*, 58.
- O'Connor, R.P., Woodley, R., Kolstad, J.J, Kean, R., Glassner, D.A., Mastel, B., Ritzenthaler, J.M., John, H., Warwick, J., Hettenhaus, J.R. and Brooks, R.K. (2007). Process for fractionating lignocellulosic biomass into liquid and solid products. US patent, WO 2007120210.

- Odeniyi, O.A., Anthony, A.O. and Maria, A.A. (2012). Characteristics of a β-1,4-d endoglucanase from *Trichoderma virens* wholly applied in a palm-fruit husk-based diet for poultry layers. *Brazilian Journal of Microbiology*, 1467-1475.
- Ouyang, J., Li, Z., Li, X., Ying, H. and Yong, Q. (2009). Enhancement enzymatic conversion and glucose production via two-step enzymatic hydrolysis of corncob residue from xylo-oligosaccharides producers waste. *Bioresources*, 4(4):1586-1599.
- Ozawa, T., Takahiro, O. and Osama, N. (1996). Hemicelluloses in the fibrous residue of sago palm. *Proceeding of 6th International Sago Symposium*, Universitas Riau, Pekan Baru, Sumatra. Republic of Indonesia. 9-12 Dec, 1996.
- Palmqvist, B., Wiman, M. and Liden, G. (2011). Effect of mixing on enzymatic hydrolysis of steam-pretreated spruce: a quantitative analysis of conversion and power consumption. *Biotechnology and Biofuels*, 4:10.
- Perez, S., Baldwin, P.M. and Gallant, D.J. (2009). Structural features of starch granules I. *Chemistry and Technology*, 149-192.
- Praytno, T.A. (1991). Palm Wood Utilization: Sago Properties and its Utilization. University of Yogyakarta : IDRC-GMU Project Report.
- Preiss, J. (2009). Biochemistry and molecular biology of starch biosynthesis. *Chemistry* and Technology, 83-148.
- Qi, B., Chen, X., Shen, F., Su, Y. and Wan, Y. (2009). Optimization of enzymatic hydrolysis of wheat straw pretreated by alkaline peroxide using response surface methodology. *Institute of Proceeding Engineering*, 48:15.
- Ratanakhanokchai, K., Waeonukul, R., Pason, P., Tachaapaikoon, C., Kyu, K.L., Sakka, K., Kosugi, A. and Mori, Y. (2013). *Paenibacillus curdlanolyticus* Strain B-6 multienzyme complex : a novel system for biomass utilization, biomass now-cultivation and utilization. In M.D. Matovic, (Ed.), ISBN: 978-953-51-1106-1.
- Razak, M.N.A., Ibrahim, M.F., Phang, L.Y., Hassan, M.A. and Abd-Aziz, S. (2013). Statistical Optimization of Biobutanol Production from Oil Palm Decanter Cake hydrolysate. *Bioresources*, 8(2):1758-1770.
- Rodrigues, A.L., Cavalett, A. and Lima, A.O.S. (2010). Enhancement of *Esterichia coli* cellulolytic activity by co-production of beta-glucosidase and endoglucanase enzymes. *Electronic Journal of Biotechnology*, 13(5):1-9.
- Ruddle, K.R. (1977). Sago in the new world. In *Sago-76: Papers of the first international sago symposium*, Ed. M.A. Tankooling, pp 53-64. Kuala Lumpur : Kemajuan Kanji Sdn. Bhd.
- Samaniuk, J.R., Scott, C.T., Root, T.W. and Klingenberg, D.J. (2011). The effect of high intensity mixing on the enzymatic hydrolysis of concentrated cellulose fiber suspensions. *Bioresources Technology*, 102:4489-4494.

- Sarao, L.K., Arora, M. and Sehgal, V.K. (2010). Use of Scopulariopsis acremonium for the production of cellulose and xylanase through submerged fermentation. African Journal of Microbiology Research, 4(14):1506-1510.
- Schlesinger, W.H. (1991). Biogeochemistry: an analysis of global change. San Diego : Academic.
- Shahrim, Z. (2006). Production of sugars from sago hampas by *Trichoderma* sp. during solid substrate fermentation. Master Thesis, Universiti Putra Malaysia.
- Singh, A. and Hayashi, K. (1995). Microbial cellulases, protein architecture molecular properties and biosynthesis. *Advance Applied Microbiology*, 40:1-44.
- Sridevi, A., Narasimha, G. and Rajasekhar Reddy, B. (2008). Production of Cellulase by *Aspergillus niger* on natural and pretreated lignocellulosic wastes. *Journal of Microbiology*, 7:1.
- Swatloski, R.P., Spear, S.K.H. and Rogers, R.D. (2002). Dissolution of cellose with ionic liquids. *Journal of the American chemical society*, 124:4974-4975.
- Tie, Y.L. and Lim, C.P. (1991). Lowland peat soils for sago-growing in Sarawak. In Sago-76: Papers of the first international sago symposium, pp 187-189. Kuala Lumpur : Kemajuan Kanji Sdn. Bhd.
- Valdeir, A. and Jack, N.S. (2011). Cellulose accessibility limits the effectiveness of minimum cellulase loading on the efficient hydrolysis of pretreated lignocellulosic substrates. *Biotechnology for Biofuels*, 4:3.
- Vamadevan, V. and Bertoft, E. (2015). Structure-function relationship of starch components. *Starch/Starke*, 67:55-68.
- Verardi, A., De Bari, I., Ricca, E. and Calabro, V. (2012). Hydrolysis of Lignocellulosic Biomass: Current Status of Processes and Technologies and Future Perspectives, Bioethanol. In M.A.P Lima, (Ed.), ISBN: 978-953-51-0008-9.
- Wada, M., Nishiyama, Y., Chanzy, H., Forsyth, T. and Langan, P. (2008). The structure of cellulose. ISSN 1097-0002.
- Wang, L., Luo, Z. and Shahbazi, A. (2013). Optimization of simultaneous saccharification and fermentation for the production of ethanol from sweet sorghum (Sorghum bicolor) bagasse using response surface methodology. *Industrial Crops and Products*, 42:280-291.
- Wyman, C.E. Lynd, L.R. and Gemgross, T.U. (1999). Biocommodity engineering. *Biotechnology*, 15:777-793.
- Yean, C.T. and Lan, S.Y. (1993). Sago processing wastes. In Yeoh et al. (Eds.) Waste Management in Malaysia : current status and prospects for bioremediation. *Ministry of Science, Technology and Environment of Malaysia*, pp. 159-167.

- Yunus, N., Jahim, J.M., Anuar, N., Abdullah, S.R.S. and Kofli, N.T. (2014). Feasibility of fermentative hydrogen production from sago (*mextroxylon* sp.) mill effluent seeded with mixed microbial consortia: effect of controlled and uncontrolled fermentation medium pH. *International Journal of Chemical and Environmental Engineering*, 5(4).
- Yusop, M.A. (2007). Development of treatment system for wastewater discharged from sago processing industry. Master Thesis, Universiti Malaysia Sarawak.
- Zhou, X., Chen, H. and Li, Z. (2004). CMCase activity assay as a method for cellulose adsorption analysis. *Enzyme and Microbial Technology*, 35:455-459.

