



UNIVERSITI PUTRA MALAYSIA

***ENHANCEMENT OF SPORE PRODUCTION OF ANTAGONISTIC
Penicillium oxalicum T3.3 VIA SOLID STATE FERMENTATION***

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ENHANCEMENT OF SPORE PRODUCTION OF ANTAGONISTIC *Penicillium oxalicum* T3.3 VIA SOLID STATE FERMENTATION

By:

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Thesis submitted to the Faculty of Biotechnology and Biomolecular Sciences,
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FACULTY OF BIOTECHNOLOGY AND BIOMOLECULAR SCIENCES
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LETTER OF PERMISSION

It is thereby to state that I, AIN NAJIHAH AZMI (Matric No: 163161) have done a final year project entitled “**Enhancement of spore production of antagonistic *Penicillium oxalicum* T3.3 via solid state fermentation**” under supervision of Assoc. Prof. Dr. Umi Kalsom Md. Shah from the Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

I hereby give permission to my supervisor to write and prepare manuscript from the results of this research to be published in any form, if I do not do so in six (6) months from the date above, in condition that my name is also added as one of the article’s authors. The arrangement of the name depends on the supervisor herself.

Yours sincerely,

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APPROVAL SHEET

This thesis entitled “**Enhancement of spore production of antagonistic *Penicillium oxalicum* T3.3 via solid state fermentation**” is submitted by AIN NAJIHAH AZMI (Matric No: 163161) in fulfilment of the requirement for the Degree of Bachelor of Science (Hons) Biotechnology in Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

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ABSTRACT

Abstract of thesis presented to the Faculty of Biotechnology and Biomolecular Sciences in fulfilment of the requirement for the Degree of Bachelor of Science (Hons.) Biotechnology

ENHANCEMENT OF SPORE PRODUCTION OF ANTAGONISTIC *Penicillium oxalicum* T3.3 VIA SOLID STATE FERMENTATION

By:

AIN NAJIHAH AZMI

JUNE 2015

Supervisor : Assoc. Prof. Dr. Umi Kalsom Md. Shah

Faculty : Faculty of Biotechnology and Biomolecular Sciences

Anthrachnose disease is a disease in which cause extensive pre- and postharvest damage to chilli crops caused by pathogen, *Colletotrichum gloeosporioides*. Recently, rather than using chemical control, spores produced from *Penicillium oxalicum* T3.3 raised high attention as biocontrol agent against *C. gloeosporioides*. Hence, this study was conducted in order to prove the antagonistic activity of *P. oxalicum* T3.3 toward *C. gloeosporioides* and to study the effect of cultural conditions on the production of *P. oxalicum* T3.3 spore in solid state fermentation. In this study, antagonistic activity of *P. oxalicum* T3.3 against *C. gloeosporioides* was tested in vitro by dual culture assay. *P. oxalicum* T3.3 was found to exhibit strong antagonistic activity against *C. gloeosporioides* with percent growth inhibition (PGI) of 55.86%. Then, solid state fermentation of *P. oxalicum* T3.3 was carried out in 250 mL flask containing 12 g of peat/vermiculite/soya bean meal (1:1:1, wt/wt/wt), sterilized and then inoculated with 4 discs of agar plugs (5mm in diameter) from 7 days old *P. oxalicum* culture and incubated at 28°C for 12 days. *P. oxalicum* T3.3 produced the highest spore concentration on 8th day of incubation time with 2.19×10^9 spores ml⁻¹ recorded. After that, effect of culture conditions on *P. oxalicum* T3.3 spore production was determined. The highest production of *P. oxalicum* T3.3 spore was obtained when mycelia was used as inoculum type, 4 discs (5 mm) agar plug as inoculum size, media (peat/vermiculite/soya bean meal) at proportion of 1:1:1 (wt/wt/wt), soya bean meal as nitrogen source, temperature at 28°C, and pH 6

as initial medium pH. From the findings, it can be concluded that *P. oxalicum* T3.3 was able to possess antagonistic activity against *C. gloeosporioides* and spore production can be significantly affected by culture condition factors in solid state fermentation.

Keywords: Biocontrol, *Penicillium oxalicum*, *Colletotrichum gloeosporioides*, spore, dual culture.



ABSTRAK

Abstrak tesis diserahkan kepada Fakulti Bioteknologi dan Sains Biomolekul bagi memenuhi keperluan untuk Ijazah Bacelor Sains (Kepujian) dalam Bioteknologi.

PENINGKATAN PENGHASILAN SPORA BERMUSUHAN *Penicillium oxalicum* T3.3 MELALUI FERMENTASI FASA PEPEJAL.

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Penyakit anthracnose adalah sejenis penyakit yang menyebabkan kerosakan pra dan pasca tuaian yang teruk terhadap tanaman cili, disebabkan oleh patogen *Colletotrichum gloeosporioides*. Baru-baru ini, daripada menggunakan kawalan kimia, spora yang dihasilkan daripada *Penicillium oxalicum* T3.3 telah menarik perhatian yang tinggi sebagai agen kawalan biologi terhadap *C. gloeosporioides*. Oleh itu, satu kajian telah dijalankan untuk membuktikan aktiviti antagonistik di antara *P. oxalicum* T3.3 dan *C. gloeosporioides* dan untuk mengkaji kesan keadaan kultur terhadap pengeluaran spora *P. oxalicum* T3.3 dalam fermentasi fasa pepejal. Dalam kajian ini, aktiviti antagonistik antara *P. oxalicum* T3.3 dan *C. gloeosporioides* diuji secara in vitro menggunakan kultur dual. *P. oxalicum* T3.3 didapati mempamerkan aktiviti antagonistik yang kuat terhadap *C. gloeosporioides* dengan peratus perencatan pertumbuhan (PGI) sebanyak 55.86%. Kemudian, fermentasi fasa pepejal *P. oxalicum* T3.3 telah dijalankan dalam kelalang 250 mL mengandungi 12 g gambut / vermikulit / kacang soya (1: 1: 1, berat / berat / berat), disterilkan dan kemudian disuntik dengan 4 cakera agar (5mm diameter) dari kultur *P. oxalicum* yang sudah ditumbuhkan selama 7 hari dan diinkubasi pada suhu 28°C selama 12 hari. *P. oxalicum* T3.3 menghasilkan spora yang tertinggi pada hari ke-lapan tempoh inkubasi dengan 2.19×10^9 spora ml⁻¹ direkodkan. Selepas itu, kesan keadaan kultur terhadap penghasilan spora daripada *P. oxalicum* T3.3 dikaji. Didapati,

penghasilan spora yang tertinggi dihasilkan apabila mycelia digunakan sebagai jenis inokulum, 4 cakera (5 mm) agar sebagai saiz inokulum, nisbah medium (gambut/vermikulit/kacang soya) pada 1:1:1 (berat/berat/berat), kacang soya sebagai sumber nitrogen, suhu 28°C, dan pH 6 sebagai pH awal media. Merujuk kepada hasil kajian, dapat disimpulkan bahawa *P. oxalicum* T3.3 boleh mempunyai aktiviti antagonistik terhadap *C. gloeosporioides* dan pengeluaran spora boleh dipengaruhi dengan ketara oleh faktor-faktor keadaan kultur di dalam fermentasi fasa pepejal.

Kata kunci: Agen kawalan biologi, *Penicillium oxalicum*, *Colletotrichum gloeosporioides*, spora, kultur dual.

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LIST OF ABBREVIATION

SSF:	Solid state fermentation.
SmF:	Submerged fermentation.
CVMV:	Chilli veinal mottle virus.
CMV:	Cucumber mosaic virus.
Maneb:	Manganese ethylenebisdithiocarbamate.
PDA:	Potato dextrose agar.
PGI:	Percent growth inhibition.
ANOVA:	Analysis of variance.
GIC:	Growth inhibition category.
LSD:	Least- significant difference.

CHAPTER 1

INTRODUCTION

Through the centuries, plant diseases have had profound effects on mankind. Based on scientific research on crop protection, it is projected that insects, diseases, weeds and animal pests remove almost half of the foodstuff produced in the world (Ellis & Boehm, 2008). Chilli (*Capsicum* spp) is a crucial economic crop worldwide especially in tropical and subtropical regions. Chilli is important to the culinary in term of enhancing flavor, aroma, texture, colour as well as contributing to the nutritional value of foods. However, in recent years, chilli production faced a great challenge in which almost up to 50% of the production yield is losses due to the infection with anthracnose disease (Pakdeevaporn *et al.*, 2005). According to Manandhar *et al.* (1995), anthracnose disease affected chilli fruits by causing extensive pre- and postharvest damage, hence led to anthracnose lesions. This disease is one of the major constraints to the chilli production worldwide caused by plant pathogens, *Colletotrichum* spp. Among *Colletotrichum* spp., *Colletotrichum gloeosporioides* have been recognized as one of the pathogens that can cause anthracnose disease in chilli (Than *et al.*, 2008).

Due to the emerging concern for this disease, several strategies have been established in order to minimize the incidence and severity of diseased chilli crops. The effective control of anthracnose disease is the use of either one or combination of these approaches: resistant cultivar, cultural control and chemical control (Narayanasamy, 2013). Conventionally, the uses of chemical biocides or pesticides have been long used to control microbial, fungal and insect plant pests. They play a significant part in the stabilization and increment of agricultural yield. Unfortunately, these pesticides may contribute to the atmospheric pollution as well as toxic to mammals and wildlife mainly in developing country (Santamarina *et al.*, 2002).

With growing concern for environmental pollution and toxicity effect of the chemical pesticides issues, biological control approach is receiving an intense research effort in order to lessen the utilization of chemical pesticides in the plant disease treatment

(Johnson, 1994). The term 'biological control' or 'biocontrol' is to indicate the control of "one organism by another organism" (Beirner, 1967). The pathogens can be effectively controlled by other beneficial microorganisms either by competition, parasitism or other modes of action. Moreover, by using microorganism as biocontrol agent, it provides efficient, effective and inexpensive control measures for a range of pests and disease (Zwieten *et al.*, 2007). However, until now, methods for biological control for chilli anthracnose disease have not received a lot of attention (Than *et al.*, 2008).

In view of this problem, a fungal species, *Penicillium oxalicum* has raised high attention as potential biocontrol agent against anthracnose disease. As reported, *P. oxalicum* Thom is a promising biocontrol agent against vascular wilts caused by *Verticillium* spp. and increase resistance against *Fusarium oxysporum* in tomato plants (De Cal *et al.*, 2009). The potential of *P. oxalicum* to produce antagonistic activity can be applied to control or suppress *C. gloeosporioides*, hence decrease the possibility of chilli fruits to be severely infected with anthracnose disease.

In this study, *P. oxalicum* T3.3 was used. The in vitro antagonistic activity interaction between *P. oxalicum* T3.3 and *C. gloeosporioides* can be proved by a dual culture method. The interaction between these two fungi is expected to produce inhibition zone. Application of *P. oxalicum* as biocontrol agent is based on the spore produced from this fungus. Spore of *P. oxalicum* T3.3 can be produced either from solid fermentation or liquid fermentation. For effective control of plant disease, fermentation mode that is able to produce high spore concentration is much desirable. Hence, solid state fermentation (SSF) is more preferable because it produce more spores compared to submerged fermentation (SmF) Larena *et al.*, (2002) and allows fungi to produce healthy spores and mycelia Bartlett & Jaronski (1988).

As mention above, SSF is more desirable to produce spore. Yet, to produce spore in masses, a reliable method must be developed. Hence, the effects of six parameters towards production of *P. oxalicum* T3.3 spore will be observed. First, the effect of inoculum type can be determined by inoculating fermentation media with agar plug in which referring to the mycelia, and spore suspensions. Second, the effect of

inoculum size can be obtained by varying the number of agar plugs to be inoculated into the fermentation media. Third, the effect of medium proportion is determined by changing the ratio of the medium. Fourth, the effect of nitrogen source is investigated by observing the outcomes of the spore production in relation to the presence and absence of soya bean meal in the medium. Last but not least, temperature as well as pH of media effects on spore production can be determined by varying the range of incubation temperature and initial pH of the media. The highest amount of spore concentration recorded in each parameter will be considered as the optimum condition for *P. oxalicum* T3.3 to produce spore in SSF.

1.1 Objective of the study

The aims of this study were:

1. To prove the antagonistic activity of *P. oxalicum* T3.3 toward *C. gloeosporioides*
2. To study the effect of cultural conditions; inoculum type, inoculum size, medium proportion, nitrogen source, temperature as well as pH on spore production of *P. oxalicum* T3.3 in solid state fermentation.

REFERENCES

- Abubakar, A., Suberu, H. A., Bello, I. M., Abdulkadir, R., Daudu, O. A., and Lateef, A.A. (2013). Effect of pH on mycelial growth and sporulation of *Aspergillus parasiticus*. *Plant Sciences*, 1(4): 64-67.
- Agosin, E., Volpe, D., Munoz, G., San Martin, R., and Crawford, A. (1997). Effect of culture conditions on spore shelf life of the biocontrol agent *Trichoderma harzianum*. *Microbiology & Biotechnology*, 13(2): 225-232.
- Ajay Kumar, G. (2014). *Colletotrichum gloeosporioides*: Biology, pathogenicity and management in India. *Plant Physiology and Pathology*, 2(2): 1-11.
- ASTM D4442 (American Society for Testing Materials in Publication). (1992). Standard test method for direct moisture content measurement of wood and wood-based. Retrieved from <ftp://law.resource.org/pub/us/cfr/ibr/003/astm.d4442.1992.pdf>. on 27 May 2015.
- Bailey, J.A., and Jeger, M.J. (1992). *Colletotrichum*: Biology, pathology and control (pp. 388). Wallingford, United Kingdom: Commonwealth Mycological Institute.
- Bartlett, M.C., and Jaronski, S.T. (1988). Mass production of entomogenous fungi for biological control of insects. In M.N. Gurge (ed.), *Fungi in biological control systems*. (pp. 61-85). Manchester, UK: Manchester University Press.
- Batta, Y.A. (2007). Control of postharvest diseases of fruit with an invert emulsion formulation of *Trichoderma harzianum* Rifai. *Postharvest Biological Technology*, 43:143–150.
- Beirner, B.P. (1967). Biological control and its potential. *World Review of Pest Control*, 6:7–20.
- Benítez, T., Rincón, A.M., Limón, M.C., and Codón, A.C. (2004). Biocontrol mechanisms of *Trichoderma* strains. *International Microbiology*, 7: 249–260.
- Berbee, M., Yoshimura, A., and Sugiyama, J. (1995). Is *Penicillium* monophyletic? An evaluation of phylogeny in the family Trichocomaceae from 18S, 5.8S and ITS ribosomal DNA sequence data. *Mycologia*, 87: 210–222.
- Bitton, G., and Boylan, R.A. (1985). Effect of acid precipitation on soil microbial activity: I. Soil core studies. *Journal of Environmental Quality*, 14: 66-69.
- Boddy, L. (2000). Interspecific combative interactions between wood-decaying basidiomycetes. *FEMS Microbiol Ecology*. 31, 185–194.
- Bosland, P.W., and Votava, E.J. (2003). Peppers: Vegetable and spice capsicums (pp. 23). England: CAB International.
- Brimner, T.A., and Boland, G.J. (2003). A review of the non-target effects of fungi used to biologically control plant diseases. *Agricultural Ecosystem Environment*, 100:3–16.

- Cannon, P.F., Buddie, A.G., Bridge, P.D., de Neergard, E., and Lübeck, M. (2012). *Lectera*, a new genus of the Plectosphaerellaceae for the legume pathogen *Volutella colletotrichoides*. *Myckeys*, 3: 23–36.
- Cano, J., Guarro, J., and Gené, J. (2004). Molecular and morphological identification of *Colletotrichum* species of clinical interest. *Clinical Microbiology* 42: 2450–2454.
- Carlile, M.J., Watkinson, S.C., and Goody, G.W. (2001). *The Fungi* (2nd ed.) (pp. 475). London: Academic Press.
- Chowdappa, P., Chethana, C.S., Bhargavi, R., Sandhya, H., and Prasad, R. (2012). Morphological and molecular characterization of *Colletotrichum gloeosporioides* (Penz) Sac. isolates causing anthracnose of orchids in India. *Biotechnology, Bioinformatic and Bioengineering*, 2(1):567-572.
- Cook, R. J. (1985). Biological control of plant pathogens: Theory to application. *Phytopathology*, 75: 25-29.
- Cook, R.J. (1987). Research briefing panel on biological control in managed ecosystems (pp. 12). Washington, DC: Committee of Science, Engineering and Public Policy, National Academy of Sciences, National Academy of Engineering, Institute of Medicine, and National Academies Press.
- Corda, A.C.I. (1831). Die Pilze Deutschlands. In J. Sturm (ed.) *Deutschlands Flora in Abbildungen nach der Natur mit Beschreibungen*, vol. 3, abt. 12 (pp. 21-32).Sturm: Nürnberg.
- Currie, J.N., and Thom, C. (1915). An oxalic acid producing *Penicillium*. *biology and chemical*, 22: 287-293.
- De Cal, A., and Melgarejo, P. (2001). Repeated applications of *Penicillium oxalicum* prolongs biocontrol of Fusarium wilt of tomato plants. *Plant Pathology*, 107:805-811.
- De Cal, A., García-Lepe, R., Pascual, S., and Melgarejo, P. (1999). Effects of timing and method of application of *Penicillium oxalicum* on efficacy and duration of control of Fusarium wilt of tomato. *Plant Pathology*, 48: 260-266.
- De Cal, A., Pascual, S., Larena, I., and Melgarejo, P. (1995). Biological control of *Fusarium oxysporum f.sp. lycopersici*. *Plant Pathology*, 44: 909-917.
- De Cal, A., Szejnberg, A., Sabuquillo, P., and Melgarejo, P. (2009). Management Fusarium wilt on melon and watermelon by *Penicillium oxalicum*. *Biological Control*, 51: 480-486.
- De Hoog, G.S., Guarro, J., Gene, J., and Figueras, M.J. (2000). *Atlas of Clinical Fungi* (2nd ed), vol.1. Utrecht, The Netherlands: Centraal bureau voor Schimmelcultures.
- Dean, R., Van Kan, J.A.L., Pretorius, Z.A., Hammond-Kosack, K.E., and Di Pietro, A. (2012). The top 10 fungal pathogens in molecular plant pathology. *Molecular Plant Pathology*, 13: 414–430.

- Dybas, R.A. (1984). Avermectins their chemistry and pesticidal activities. In J. Miyamoto, & P.C., Keaney (Eds.), *Pesticide chemistry* (pp. 83-90). New York: Pergamon Press.
- El- Enshasy, H.A., Farid, M.A., and El-Sayed, E.A. (2000). Influence of inoculum type and cultivation conditions on natamycin production by *Streptomyces natalensis*. *Basic Microbiology*, 40(5-6): 333-342.
- Elad, Y. (1995). Mycoparasitism. In K. Kohmoto, U.S. Singh & R.P. Singh (Eds.), *Pathogenesis and host specificity in plant diseases: Histopathological, biochemical, genetic and molecular basis, eukaryotes*, vol. 2 (pp. 289-307). Oxford, UK: Pergamon, Elsevier.
- Ellis, S.D., and Boehm, M.J. (2008). Plant Get Sick Too! An introduction to plant diseases (pp.1-5). Agriculture and Natural Resource, The Ohio State University.
- FAO (Food and Agriculture Organization of the United Nations). (2003). In FAO Production Yearbook , 2001. Rome: FAO, 55(170):333.
- Feng, K.C, Liu, B.L., and Tzeng, Y.M. (2000). *Verticillium lecanii* spore production in solid state and liquid-state fermentations. *Bioprocess Engineering*, 23: 25-29.
- Ferron, P. (1978). Biological control of insect pests by entomogenous fungi. *Annual Review of Entomology*, 23: 409-442.
- Fleming, A. (1929). On the antibacterial action of cultures of *Penicillium* with special reference to their use in the isolation of *B. influenzae*. *Review of Infectious Disease*, 2:129–139.
- Foda, M.S., El-Bendary, M.A. and Moharam, M.E. (2003). Salient parameters involved in mosquitocidal toxins production from *Bacillus sphaericus* by semi-solid substrate fermentation. *Egypt Microbiology*, 38: 229-246.
- Freeman, S., Katan, T., and Shabi, E. (1998). Charac-terization of *Colletotrichum* species responsible for anthracnose diseases of various fruits. *Plant Disease*, 82:596-605.
- Frisvad, J.C. (1993). Modifications on media based on creatine for use in *Penicillium* and *Aspergillus* taxonomy. *Letter of Applied Microbiology*, 16: 154–157.
- Frisvad, J.C., and Samson, R.A. (2004). Polyphasic taxonomy of *Penicillium* subgenus *Penicillium*. A guide to identification of food and air-borne Terverticillate Penicillia and their mycotoxins. *Studies in Mycology*, 49: 1–174.
- Frisvad, J.C., Smedsgaard, J., and Larsen, T.O. (2004). Mycotoxins, drugs and other extrolites produced by species in *Penicillium* subgenus *Penicillium*. *Studies in Mycology*, 49: 201–241.
- Fungal Biology. (2004). Growth of hyphae & development of fungi. Retrieved from http://bugs.bio.usyd.edu.au/learning/resources/Mycology/Growth_Dev/hyphalGrowth.shtml on 4 May 2015.
- García, F.J., Hinojosa, A., Real, M.C., and Santamarina, M.P. (2001). Estudio de la actividad bactericida y fungicida del hongo *Penicillium oxalicum* Currie et Thom. *Phytoma*, 132: 134–137.

- Gomori, G. (1955). Preparation of buffers for use in enzyme studies. *Methods in Enzymology*, 1: 138-146.
- Haggag, W.M., and El Soud, M.A. (2013). Pilot-scale production and optimizing of cellulolytic *Penicillium oxalicum* for controlling of mango malformation. *Agricultural Sciences*, 4(4) :165-174.
- Halsted, B.D. (1890). A new anthracnose of pepper. Torrey Botanical. Club – Bulletin, 18:14-15.
- Handelsman, J., and Stabb, E.V. (1996). Biocontrol of soilborne plant pathogens. *Plant Cell*, 8:1855–1869.
- Harman, G.E. (2000). Myths and dogmas of biocontrol: changes in perceptions derived from research on *Trichoderma harzianum* T-22. *Plant Disease*, 84: 377–393.
- Houbraken, J., Visagie, C.M., Frisvad, J.C., Hong, S.B., Klaassen, C.H.W., Perrone, G., Seifert, K.A., Varga, J., Yaguchi, T., and Samson, R.A. (2014). Identification and nomenclature of the genus *Penicillium*. *Studies in Mycology*, 78: 343-371.
- Hubballi, M., Nakkeeran, S., and Raguchander, T. (2012). First report of anthracnose on noni caused by *Colletotrichum gloeosporioides* in India. *Archive of Phytopathology and Plant Protection*, 45: 276-279.
- Hubballia, M., Nakkeerana, S., Raguchandera, T., Ananda, T., and Renukadevi, P. (2011). Physiological characterisation of *Colletotrichum gloeosporioides*, the incitant of anthracnose disease of noni in India. *Archive of Phytopathology and Plant Protection*, 44:1105-1114.
- Ikotun, T. (1984). Cell wall degrading enzymes produced by *Penicillium oxalicum* Curie et Thom. *Mycopathologia*, 88:15–21.
- Intanoo, W., and Chamswarnng, C. (2007). *Effect of antagonistic bacterial formulations for control of anthracnose on chilli fruits* (pp. 309-322). Proceeding of the 8th National Plant Protection Conference. Naresuan University, Phisanulok, Thailand.
- Isaac, I. (1989). What factors determine the duration of the dormancy of fungus prior to germination? (pp. 38-39). Crown Street, Liverpool: School of Biological Sciences, University of Liverpool.
- Isaac, S. (1992). *Fungal Plant Interaction* (pp. 115). London: Chapman and Hall Press.
- Jackson, A.M., Whipps, J.M., Lynch, J.M. and Bazin, M.S. (1991). Effects of some carbon and nitrogen sources on spore germination, production of biomass and antifungal metabolites by species of *Trichoderma* and *Gliocladium virens* antagonistic to *Sclerotium cepivorum*. *Biocontrol Science and Technology*, 1: 43-51.
- Jeffries, P., and Koomen, I. (1992). Strategies and prospects for biological control of diseases caused by *Colletotrichum*. In J.A. Bailey & M.J. Jeger (Eds.),

Colletotrichum: Biology, pathology and control (pp. 337-357). Wallingford, United Kingdom: Commonwealth Mycological Institute.

Jeger, M.J., and Jeffries, P. (1988). Alternative to chemical usage for disease management in the post-harvest environment. *Aspects of Applied Biology*, 17:47-57.

Johnson, K.B. (1994). Dose–response relationships and inundative biological control. *Phytopathology*, 84: 780–784.

Johnston, P.R., and Jones, D. (1997). Relationships among *Colletotrichum* isolates from fruit-rots assessed using rDNA sequences. *Mycologia*, 89(3):420-430.

Kamala, T., and Indira, S., (2011). Evaluation of indigenous *Trichoderma* isolates from Manipur as biocontrol agent against *Pythium aphanidermatum* on common beans. *3 Biotech*, 1(4),217-225.

Kim, K.K., Yoon, J.B., Park, H.G., Park, E.W., and Kim, Y.H. (2004). Structural modifications and programmed cell death of chilli pepper fruits related to resistance responses to *Colletotrichum gloeosporioides* infection. *Genetics and Resistance*, 94:1295-1304.

Kiss, L. (2003). A review of fungal antagonists of powdery mildews and their potential as biocontrol agents. *Pest Management Science*, 59:475–483.

Korsten, L., De-Jager, E.S., De-Villers, E.E., Lourens, A., Kotze, J.M., and Wehner, F.C. (1995). Evaluation of bacterial epiphytes isolated from avocado leaf and fruit surfaces for biocontrol of avocado postharvest diseases. *Plant Disease*. 79: 1149–1156.

Kumm, J. (2009). General characteristics of *Clostridium difficile*. Retrieved from http://bioweb.uwlax.edu/bio203/s2009/kumm_jakl/growth&adapt.htm on 19 May 2015.

Larena, I., Melgarejo, P., and De Cal, A. (2002). Production, survival and evaluation of solid substrate inoculum of *Penicillium oxalicum*, a biocontrol agent against *Fusarium wilt* of tomato. *Phytopathology*, 92:863-869.

Larena, I., Melgarejo, P., and De Cal, A. (2003). Drying of conidia of *Penicillium oxalicum*, a biological control agent against *Fusarium wilt* of tomato. *Phytopathology*, 151:600-606.

Lareo, C., Sposito, A.F., Bossio, A.L., and Volpe, D.C. (2006). Characterization of growth and sporulation of *Mucor bacilliformis* in solid state fermentation on an inert support. *Enzyme and Microbial Technology*, 38, 391-399.

Larone, D. H. (1995). *Medically Important Fungi - A Guide to Identification* (3rd ed.) Washington, D.C.: ASM Press.

Larroche, C., and Gros, J.B. (1992). Characterization of the growth and sporulation behaviour of *Penicillium roquefortii* in solid substrate fermentation by material and bioenergetic balances. *Biotechnology and Bioengineering*, 39: 815-827.

- Lenné, J.M., and Parbery, D.G. (1976). Phyllosphere antagonists and appressoria formation in *Colletotrichum gloeosporioides*. *Transactions of the British Mycological Society*, 66:334-336.
- Lewis, J. A., and Papavizas, G. C. (1991). Biocontrol of plant diseases: The approach for tomorrow. *Crop Protection*, 10:95-105.
- Lewis, K., Whipps, J.M., and Cooke, R.C. (1989). Mechanisms of biological diseases control with special reference to the case study of *Pythium oligandrum* as an antagonist. In J.M. Whipps & R.D. Lumsden (Eds), *Biotechnology of fungi for improving plant growth* (pp. 191-217). Cambridge: Cambridge University Press.
- Limón, M.C., Chacón, M.R., Mejías, R., Delgado-Jarana, J., Rincón, A.M. (2004). Increased antifungal and chitinase specific activities of *Trichoderma harzianum* CECT 2413 by addition of a cellulose binding domain. *Applied Microbiology and Biotechnology*, 64: 675-685.
- Link, H.F. (1809). Observationes in ordines plantarum naturales. Dissertatio 1ma. *Magaziner Gesellschaft Naturforschenden Freunde Berlin*, 3: 3–42.
- Mamat, S. (2014). Isolation and characterisation of antifungal activity of endophytic *Penicillium oxalicum* T3.3 for anthracnose biocontrol in dragon fruit (*Hylocereus* sp.). Master's Thesis, University Putra Malaysia, Serdang, Malaysia.
- Manandhar, J.B., Hartman, G.L., and Wang, T.C. (1995). Anthracnose development on pepper fruits inoculated with *Colletotrichum gloeosporioides*. *Plant Disease*, 79: 380-383.
- Manpreet, S., Sawraj, S., Sachin, D., Pankaj, S., and Banerjee, U.C. (2005). Influence of process parameters on the production of metabolites in solid-state fermentation. *Malaysian Journal of Microbiology*, 1(2): 1-9.
- Maymon, M., Minz, D., Barbul, O., Zveibil, A., Elad, Y., and Freeman, S. (2004). Identification to species of *Trichoderma* biocontrol isolates according to ap-PCR and ITS sequence analyses. *Phytoparasitica*, 32: 370–375.
- McCoy, C.W., Stamper, D.H. and Tuveson, R.W. (1984). Conidiogenous cell difference among mutant and wild type pathotypes of *Hirsutella thompsonii* var. *thompsonii*. *Invertebrate Pathology*, 43: 414-421.
- McQuiken, M. P., and Whipps, J.M. (1995). Production, survival and evaluation of solid substrate inoculum of *Coniothyrium minitans* against *Sclerotinia sclerotiorum*. *European Plant Pathology*, 101, 101-110.
- McQuilken, M.P., Budge, S.P., and Whipps, J.M. (1997). Production, survival and evaluation of liquid culture produced inoculum of *Coniothyrium minitans* against *Sclerotinia sclerotiorum*. *Biocontrol Science and Technology*, 7: 23-26.
- Mei, C., Zhang, Y., Mao, X., Jiang, K., Cao, L., Yan, X., Han, R. (2013). The effects of culture parameters on the conidial germination and yields of *Ophiocordyceps sinensis*. *Yeast and Fungal Research*, 4(4): 44-51.

- Method 9045D. (2004). Soil and waste pH. Retrieved from <http://www.epa.gov/osw/hazard/testmethods/sw846/pdfs/9045d.pdf>. on 1 May 2015.
- Moksia, J., Larroche, C., and Gros, J.B. (1996). Gluconate production by spores of *Aspergillus niger*. *Biotechnology Letters*, 18: 1025-1030.
- Monga, D. (2001). Effect of carbon and nitrogen sources on spore germination, biomass production and antifungal metabolites by species of *Trichoderma* and *Gliocladium*. *Indian Phytopathology*, 54(4): 435-437.
- Moreaux, C. (1980). Moisissures Toxiques dans (pp. 297). Paris: Alimentation Masson and Co.
- Moskowitz, G.J. (1979). Inoculum for blue-veined chesses and blue cheese flavour. In A.J. Peppier & D. Perlman (Eds), *Microbial Technology* (2nd ed.), vol.2 (pp. 201-210). New York & London : Academic.
- Mukherjee, P.K., and Raghu, K. (1997). Effect of temperature on antagonistic and biocontrol potential of shape *Trichoderma* sp. on *Sclerotium rolfsii*. *Mycopathologia*, 139(3): 151-155.
- Mukhtar, H., Nawaz, A., and Haq, I. (2014). Effect of volume of inoculum and fermentation medium on the production of protease by *Penicillium chrysogenum*. *New York Science*, 7(7): 34-36.
- Munoz, G.A., Agosin, E., Cotoras, M., San Martin, R. and Volpe, D. (1995). Comparison of aerial and submerged spore properties for *Trichoderma harzianum*. *FEMS Microbiology Letters*, 125: 63-70.
- Narayanasamy, P. (2013). Biological management of disease of crops, progress in biological control, Volume 2: Integration of biological control strategies with crop disease management system. New York, London: Springer
- Nawar, L.S. (2008). Control of root-rot of green bean with composted rice straw fortified with *Trichoderma harzianum*. *American – Eurasian Agricultural and Environmental Science*, 3(3): 370-379.
- Oostra, J., Tramper, J., and Rinzema, A. (2000). Model-based bioreactor selection for large scale solid state cultivation of *Coniothyrium minitans* spores on oats. *Enzyme and Microbial Technology*, 27:652-663.
- Pakdaman, B.S., Goltapeh, E.M., Soltani, B.M., Talebi, A.A., and Nadeppoor. (2013). Toward the quantification of confrontation (dual culture) test: A case study on the biological control of *Pythium aphanidermatum* with *Trichoderma asperelloides*. *Biofertilizer and Biopesticide*, 4: 137.
- Pakdeevaporn, P., Wasee, S., Taylor, P.W.J., and Mongkolporn, O. (2005). Inheritance of resistance to anthracnose caused by *Colletotrichum capsici* in *Capsicum*. *Plant Breeding*, 124(2):206-208.
- Pandey, A. (1992). Recent developments in solid-state fermentation. *Process Biochemistry*, 27: 109-117.

- Pandey, A. (2003). Solid-state fermentation. *Biochemical Engineering*, 13(2-3): 81-84.
- Pandey, A., Soccol, C.R., and Mitchell, D. (2000). New developments in solid-state fermentation: I-bioprocesses and products. *Process Biochemistry*, 35(10): 1153-1169.
- Pandey, R.R., Arora, D.K., and Dubey, R.C. (1993). Antagonistic interactions between fungal pathogens and phyllophane fungi of guava. *Mycopathologia*, 124: 31-39.
- Park, H.G. (2007). Problems of anthracnose in pepper and prospects for its management. In D.G. Oh & K.T. Kim (Eds.), *Abstracts of the first international symposium on chilli anthracnose* (pp.19). Republic of Korea: National Horticultural Research Institute, Rural Development of Administration.
- Pascual, S., De Cal, A., Magan, N., and Melgarejo, P. (2000). Surface hydrophobicity, viability and efficacy in biological control of *Penicillium oxalicum* spores produced in aerial and submerged culture. *Applied Microbiology*, 89:847-853.
- Pascual, S., Melgarejo, P., and Magan, N. (1997). Induction of submerged conidiation of the biocontrol agent *Penicillium oxalicum*. *Applied Microbiology and Biotechnology*, 48: 389-392.
- Penzig, A.G.O. (1882). Funghi agrumicoli. Contribuzione allo studio dei funghi parassiti degli agrumi. *Michelia*, 2: 385-508.
- Pickersgill, B. (1997). Genetic resources and breeding of *Capsicum* spp. *Euphytica*, 96(1):129-133.
- Pitt, J.I., and Hocking, A.D. (1999). *Fungi and Food Spoilage* (2nd ed.). New York: Aspen Publishers, Inc.
- Poonpolgul, S., and Kumphai, S. (2007). Chilli pepper anthracnose in Thailand. Country Report. In D.G. Oh & K.T. Kim (Eds.), *Abstracts of the first international symposium on chilli anthracnose* (pp. 23). Republic of Korea: National Horticultural Research Institute, Rural Development of Administration.
- Prusky, D., Kobler, I., Aridi, R., Beno-Moalem, D., Yakoby, N., and Keen, N.T. (2000). Resistance mechanisms of subtropical fruits to *Colletotrichum gloeosporioides*. In J.A. Bailey & M.J. Jeger (Eds.), *Colletotrichum: Biology, Pathology, and Control* (pp. 232-244). Wallingford, United Kingdom: CAB International.
- Ramachandran, S., Fontanille, P., Pandey, A. and Larroche, C. (2008). Stability of glucose oxidase activity of *A. niger* spores produced by solid-state fermentation and its role as biocatalyst in bioconversion reaction. *Food Technology and Biotechnology*, 46(2): 190-194.
- Ramachandran, S., Larroche, C., and Pandey, A. (2005). Production of spores. In A. Pandey, C.R. Soccol & C. Larroche (Eds), *Current development in solid-state fermentation* (pp. 230-246). New Delhi: Asiatech Publisher, Inc.

- Ramirez-Peralta, A., Zhang, P., Li, Y.Q., and Setlow, P. (2012). Effects of sporulation conditions on the germination and germination protein levels of *Bacillus subtilis* spores. *Applied and Environmental Microbiology*, 78: 2689-2697.
- Rodríguez-León, Jose A., Domenech, F., León, M., Méndez, T., Rodríguez, D. E., and Pandey, A. (1999). Production of spores of *Trichoderma harzianum* on sugar cane molasses and bagasse pith in solid state fermentation for biocontrol. *Brazilian Archives of Biology and Technology*, 42(1) Retrieved from http://www.scielo.br/scielo.php?script=sci_arttext&pid=S151689131999000100010&lng=en&tlng=en. 10.1590/S1516-89131999000100010 on 27 May 2015.
- Roselló, J., Serna, R., and Santamarina, P. (2002). Interacciones in vitro de *Trichoderma harzianum* Rifai frente a *Verticillium dahliae* Klebahn. *Revista Iberoamericana de Micología*, 19: 27.
- Rossi, V., Patteri, E., Ravanetti, A., and Giosuè, S. (2002). Effect of constant and fluctuating temperature regimes on sporulation of four fungi causing head blight of wheat. *Plant Pathology*, 84(2): 95-105.
- Sabuquillo, P., De Cal, A., and Melgarejo, P. (2006). Biocontrol of tomato wilt by *Penicillium oxalicum* formulations in different crop conditions. *Biological Control*, 37:256–265.
- Sambrook, J., and Russell, D.W. (2001). *Molecular Cloning: A laboratory manual*, (3rd ed.). New York: Cold Spring Harbor Laboratory Press.
- Samson, R. A., and Gams, W. (1984). The taxonomic situation in the fungal genera *Penicillium*, *Aspergillus* and *Fusarium*. *Antonie van Leeuwenhoek*, 50: 815-824.
- Santa, H.S. D., Santa, O.R.D., Brand, D., Vandenberghe, L.P.S. and Soccol, C.R. (2005). Spore production of *Beauveria bassiana* from agro-industrial residues. *Brazilian Archives of Biology and Technology*, 48: 51-60.
- Santamarina, M.P., Rosello, J., Llacer, R., and Sanchis, V. (2002). Antagonistic activity of *Penicillium oxalicum* Currie and Thom, *Penicillium decumbens* Thom and *Trichoderma harzianum* Rifai isolates against fungi, bacteria and insects in vitro. *Revista Iberoamericana de Micología*, 19: 99-103.
- Saucedo-Castañeda, G., Lonsane, B.K., Krishnaiah, M.M., Navarro, J.M., Roussos, S. and Raimbault, M. (1992). Maintenance of heat and water balances as a scale-up criterion for the production of ethanol by *Schwanniomyces castellii* in a solid state fermentation system. *Process Biochemistry*, 27: 97-107.
- Sempere, F., and Santamarina, M.P. (2008). Suppression of *Nigrospora oryzae* (Berk. & Broome) Petch by an aggressive mycoparasite and competitor, *Penicillium oxalicum* Currie & Thom. *Food Microbiology*, 122: 35-43.

- Sharma, A., Behere, A.G., Padwal-Desai, S.R., and Nadkarni, G.B. (1980). Influence of inoculum size of *Aspergillus parasiticus* spores on aflatoxin production. *Applied and Environmental Microbiology*, 40(6): 989-993.
- Shi, J., Li, Y., Zheng, Y., Zhu, Y., Zhang, X., Du, G., and Chen, J. (2008). Tryptophan supplementation and pH adjustment for optimizing the sporulation of *Coniothyrium minitans*. *Biotechnol Lett*, 30(2): 259-262.
- Shi, Y., Xu, X., and Zhu, Y. (2009). Optimization of *Verticillium lecanii* spore production in solid-state fermentation on sugarcane bagasse. *Applied Microbiology and Biotechnology*, 82(5): 921-927.
- Shishkoff, N., and McGrath, M.T. (2002). AQ10 biofungicide combined with chemical fungicides or AddQ spray adjuvant for control of cucurbit powdery mildew in detached leaf culture. *Plant Disease*, 86:915-918.
- Singh, A., Shahid, M., Srivastava, M., Pandey, S., Sharma, A., and Kumar, V. (2014). Optimal physical parameters for growth of *Trichoderma* species at varying pH, temperature and agitation. *Virology and Mycology*, 3(1): 1-7.
- Smith, K.L. (2000). Peppers. In R.J. Precheur (Ed.), *Ohio Vegetable Production Guide* (pp. 166-173). Columbus, Ohio: Ohio State University Extension.
- Soytong, K., and Soyong, K. (1997). Chaetomium as a new broad spectrum mycofungicide (pp. 124-132). Proceeding of first international symposium on biopesticide.
- Soytong, K., Srinon, W., Rattanacherdchai, K., Kanokmedhakul, S. and Kanokmedhakul, K. (2005). Application of antagonistic fungi to control anthracnose disease of grape. *Agricultural Biotechnology*, 1: 33-41.
- Sreenivasaprasad, S., and Talhinhas, P. (2005). Genotypic and phenotypic diversity in *Colletotrichum acutatum*, a cosmopolitan pathogen causing anthracnose on a wide range of hosts. *Molecular Plant Pathology*, 6: 361-378.
- Srivastava, S., and Thakur, I.S. (2006). Evaluation of bioremediation and detoxification potentiality of *Aspergillus niger* for removal of hexavalent chromium in soil microcosm. *Soil Biology and Biochemistry*, 38: 1904-1911.
- Staub, T. (1991). Fungicide resistance: practical experience and antiresistance strategies and the role of integrated use. *Annual Review of Phytopathology*, 29(1):421-442.
- St-Germain, G., and Summerbell, R. (1996). Identifying Filamentous Fungi - A Clinical Laboratory Handbook (1st ed.). Belmont, California: Star Publishing Company.
- Sutton, D. A., Fothergill, A. W., and Rinaldi, M. G. (1998). Guide to Clinically Significant Fungi (1st ed.). Baltimore: Williams & Wilkins.
- Szczepanowska, H. M. and Moomaw, W. R. (1994). Laser stain removal of fungus-induced stains from paper. *Journal of the American Institute for Conservation*, 33(1): 25-32.

- Tarocco, F., Lecuona, R.E., Couto, A.S., and Arcas, J.A. (2005). Optimization of erythritol and glycerol accumulation in conidia of *Beauveria bassiana* by solid-state fermentation using response surface methodology. *Applied Microbiology and Biotechnology*, 68: 481-488.
- Tee, E.S., Noor, M.I., Azudin, M,N and Idris, K. (1988). Nutrient Composition of Malaysian Foods. (pp. 50 & 89). Kuala Lumpur, Malaysia: ASEAN Food Handling Project.
- Than, P.P., Prihastuti, H., Phoulivong, S., Taylor, P.W.J., and Hyde, K.D. (2008). Chilli anthracnose disease caused by *Colletotrichum* species. *Zhejiang University SCIENCE B*, 9(10): 764-778.
- Thom, C. (1906). Fungi in cheese ripening: Camembert and Roquefort. U.S. Department of Agriculture, Bureau of Animal Industry – Bulletin, 82: 1–39.
- Tiwari, K.L., Jadhav, S.K., and Kumar, A. (2011). Morphological and molecular study of different *Penicillium* Species. *Middle-East Scientific Research*, 7(2): 203-210.
- Verdin, A., Sahraoui, A.L., and Durand, R. (2004). Degradation of benzo[a]pyrene by mitosporic fungi and extracellular oxidative enzymes. *International Biodeterioration and Biodegradation*, 53: 65-70.
- Vermiculite. (2013). Retrieved from <http://www.ipijaya.com/index.php/produk/vermiculite> on 18 May 2015.
- Vimala, R., and Subramaniam, R. (2012). Solid state and submerged fermentation for the production of bioactive substances: a comparative study. *International Science Nature*, 3(3): 480-486.
- Voorrips, R.E., Finkers, R., Sanjaya, L., and Groenwold, R. (2004). QTL mapping of anthracnose (*Colletotrichum* spp.) resistance in a cross between *Capsicum annum* and *C. chinense*. *Theoretical and Applied Genetic*, 109(6): 1275–1282.
- Weber, F.J., Tramper, J., and Rinzema, A. (1999). A simplified material and energy balance approach for process development and scale-up of *Coniothyrium minitans* conidia production by solid-state fermentation cultivation in a packed bed reactor. *Biotechnology and Bioengineering*, 65(4): 447-458.
- Whipps ,J.M. (1994). Advances in Biological Control in Protected Crops. Brighton Crop Protection Conference, Pests and Diseases (pp. 1259-1264). Farnham, UK: British Crop Protection Council.
- Winding, A., Binnerup, S.J., and Pritchard, H. (2004). Non-target effects of bacterial biological control agents suppressing root pathogenic fungi. *FEMS Microbiology and Ecology*, 47(2): 129-141.
- Wolken, W.A.M., Tramper, J., and van de Werf, M.J. (2002). Toxicity of terpenes to spores and mycelium of *Penicillium digitatum*. *Biotechnology and Bioengineering*, 80: 685-690.

Yoon, J.B. (2003). Identification of genetic resource, interspecific hybridization and inheritance analysis for breeding pepper (*Capsicum annuum*) resistant to anthracnose. PhD Thesis, Seoul National University, Korea.

Živković, S., Stojanović, S., Ivanović, Ž. 1, Gavrilović, V., Popović, T., and Balaž, J. (2010). Screening of antagonistic activity of microorganisms against *Colletotrichum acutatum* and *Colletotrichum gloeosporioides*. *Archives of Biological Sciences*, 62(3): 611-623.

Zwieten, M.V., Stovold, G., and Zwieten, L.V. (2007). Alternatives to copper for disease control in the Australian organic industry. A report for the rural industries research and development corporation, Australia.

