

UNIVERSITI PUTRA MALAYSIA

EDIBLE MOLLUSC (GASTROPOD AND BIVALVE) AT SELECTED DIVISIONS OF SARAWAK

HADI BIN HAMLI

FPSM 2013 1

EDIBLE MOLLUSC (GASTROPOD AND BIVALVE) AT SELECTED DIVISIONS OF SARAWAK

UPM

HADI BIN HAMLI

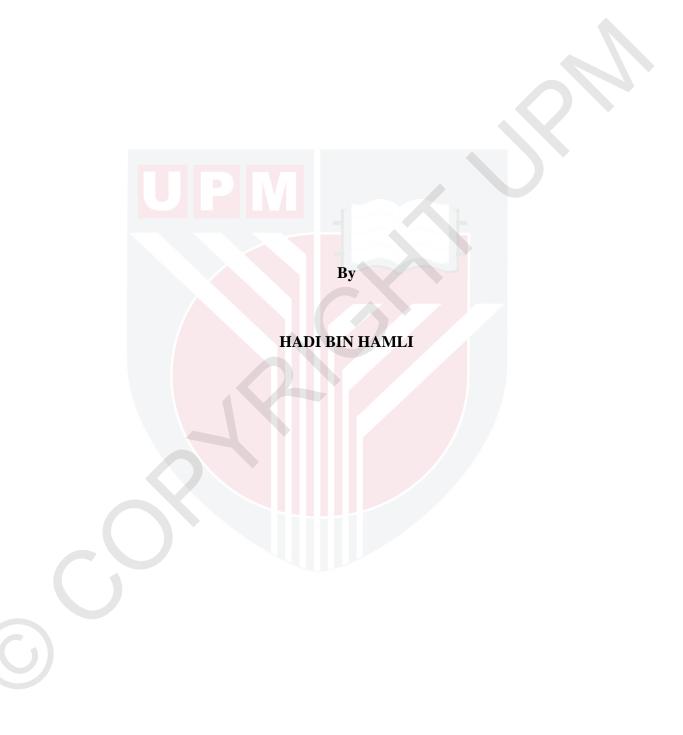
MASTER OF SCIENCE

MASTER SCIENCE UNIVERSITI PUTRA MALAYSIA

2013

2013

EDIBLE MOLLUSC (GASTROPOD AND BIVALVE) AT SELECTED DIVISIONS OF SARAWAK



Thesis Submitted to the School of Graduates Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

TABLE OF CONTENTS

		Page
DEDICATION		ii
ABSTARCT		iii
ABSTRAK		vi
ACKNOWLEDO	GEMENTS	ix
APPROVAL		X
DECLARATION	1	xii
LIST OF TABLE	ES	xvi
LIST OF FIGUR	ES	xviii
LIST OF PLATE	ES	XX
LIST OF ABBRI	EVIATIONS	xxiii
CHAPTER		
1	GENERAL INTRODUCTION	1
	1.1 Background Study	1
	1.2 Problem Statement	4
	1.3 Objectives	6
2	LITERATURE REVIEW	7
	2.1 Mollusca Morphology	7
	2.1.1 Classification	9
	2.1.2 Bivalve Morphology	10
	2.1.3 Gastropod Morphology	12
	2.2 Distribution	13
	2.3 Habitat	15
	2.4 Genetic Diversity Study	18
	2.5 Important of Mollusc in Malaysia	21
3	GENERAL METHODOLOGY	24
	3.1 Description of Study Area	24
	3.2 Collection and Identification of Edible mollusc	25
	3.3 Physico-chemical Parameters of <i>Polymesoda</i> spp. Habitat	26
	3.4 Genetic Distance Study	27
4	TAXONOMIC AND MORPHOLOGICAL STUDY OF EDIBLE BIVALVE AND	28
	GASTROPOD	• •
	4.1 Introduction	28
	4.2 Objectives	29
	4.3 Materials and Methods	29
	4.3.1 Similarity Index Study	30
	4.3.2 Morphological Identification	30

4.4	Resul	ts		33
	4.4.1	Bivalve		33
		4.4.1.1	Bivalve Distribution	35
		4.4.1.2		37
		4.4.1.3		38
		1.1.1.5	Characters Description	50
	4.4.2	Gastrop	±	99
	1. 1.2	4.4.2.1		101
		4.4.2.2		101
		4.4.2.3	•	103
		4.4.2.3	Characters Description	104
4.5	Discu	agion	Characters Description	165
4.3		Bivalve		165
				168
15	Concl	Gastrop	bod	
4.5	Conci	usion		170
MO	DDUO			171
			C CHARACTERS OF	171
		LIDAE	OF SARAWAK	
	GION			171
	Introd			171
	Objec			173
5.3		ials and l		173
	5.3.1	-	Preservation	175
		-	ometric Study	175
	5.3.3		ometric Data Analysis	178
		5.3.3.1	Proportion Ratio of	178
			Morphometric	
			Measurement	
		5.3.3.2	Cluster Analysis	178
		5.3.3.3	Principal Component	179
			Analysis (PCA)	
5.4	Resul			179
	5.4.1	Morpho	ometric Analysis of Three	179
		Polyme.	soda species	
		5.4.1.1	Proportion Ratio of Shell	182
			Length for Polymesoda	
			species	
	5.4.2	Morpho	ometric Analysis of	186
		Polyme.	soda Populations	
		5.4.2.1	Cluster Analysis	196
		5.4.2.2	Principal Component	198
			Analysis (PCA)	
5.5	Discu	ssion	-	201
	5.5.11	Differenc	es of Three Species of	201
		Polyme		
		5.5.1.1		202
			Morphometric	
			Characteristic	

5

 $\overline{(}$

		5.5.2		ntiation of <i>Polymeso</i>	oda	203
			Populat	Cluster Analysis of	2	205
			5.5.2.1	Polymesoda Popula		205
			5.5.2.2	Principal Compone		206
				Analysis of Polyme		
				Population (PCA)		
	5.6	Conc	lusion			206
6	GEN	NETIC	DISTA	NCE OF EDIBLE		208
			C (BIVA			
				AT SELECTED		
	DIV	ISION	IS OF SA	RAWAK		
	6.1	Intro	duction			208
		Objec				208
	6.3		rials and	Methods		211 211
	0.5	6.3.1		Collection		212
			DNA E			212
				uantitation and		213
			Electrpl	noresis		
				Gel preparation		213
			6.2.3.2	Sample loading and	b	214
				Electrophoresis		
		6.3.4	-	rase Reaction Chain		214
	<i>с</i> 1			Amplification		015
	6.4 6.5			ce Analysis		215
	0. <i>5</i> 6.6					216 224
	0.0 6.7	Concl				224
	0.7	Conci	1031011			221
7	GEN	NERAI	L DISCU	SSION AND		228
	CON	NCLUS	SION			
REFERENCE						235
APPENDICES						248
BIODATA OF STU	JDEN	T				257
LIST OF PUBLICA	ATIC	NS				258

DEDICATION



TO MY BELOVED FAMILY,

THEY GAVE A LOT OF INSPIRATION

TO ME

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EDIBLE MOLLUSC (GASTROPOD AND BIVALVE) AT SELECTED DIVISIONS OF SARAWAK

By

HADI BIN HAMLI

January 2013

Chairman: Mohd Hanafi Bin Idris, PhD Faculty: Agriculture and Food Sciences (Bintulu)

Diversity of edible gastropod and bivalve was investigated at seven selected coastal division of Sarawak namely Kuching, Sibu, Mukah, Bintulu, Miri, Limbang and Lawas from July 2010 to January 2012. The most common and widely distributed species i.e., *Polymesoda* spp. was observed in details for morphology and genetic variation among different populations from different divisions.

A total of 41 edible species comprising 11 malacological family of gastropod with 21 species and 12 malacological family of bivalve with 20 species was recorded and identified. Bivalve from Corbiculidae (*Polymesoda erosa*, *P. expansa* and *P. bengalensis*) family was widely distributed in all divisions compared to other bivalve species. Whilst gastropod from Potamididae (*Cerithidea obtusa*, *C. quadrata*, *C. rizophorarum* and *Telescopium telescopium*) family was commonly recorded at six from seven divisions. Edible species of gastropod and bivalve was found higher in

iii

Bintulu division (13 species for gastropod and 11 species for bivalve), while lower number of species was recorded from Sibu and Mukah divisions. Results on Jaccard's index showed high similarity for edible bivalves within Limbang versus Miri (0.75) and Limbang versus Lawas (0.75), while it was Sibu versus Mukah (0.50) for edible gastropod. This indicated species occurrence between two divisions is almost similar for higher similarity index while different species occurrence for lower similarity index.

A total of 15 morphometric characteristics (6 general and 9 additional characters) were used to differentiate three species of *Polymesoda* recorded from Sarawak. Significant differences (ANOVA, p<0.05) on 12 morphometric characteristics (SL; Shell Length, SW; Shell Width, SH; Shell Height, UL; Umbo Length, AL; Anterior Length, PL; Posterior Length, LPAS; Length of Posterior adductor scar to Anterior adductor Scar, LCT; Length of Cardinal Tooth, PVM; Palial line to Ventral Margin, AAAM; Anterior Adductor scar to Anterior Margin, PAPM; Posterior Adductor scar to Posterior adductor Margin and VPM; Ventral Posterior Margin) were found among three *Polymesoda* species collected from different divisions. Analysis showed that the proportion ratio for LCT/SL have had significant difference (ANOVA, p<0.05) between *P. bengalensis* and *P. expansa*, whilst VPM/SL significantly different (ANOVA, p<0.05) for *P. erosa* and *P. expansa* contrasted with *P. bengalensis*. However, the differences were not significant (ANOVA, p>0.05) between *P. erosa* and *bengalensis*, and *P. erosa* and *P. expansa* for LCT/SL and VPM/SL respectively.

Sixteen populations comprises of *P. erosa*, *P. expansa* and *P. bengalensis* recorded from different divisions were assessed (ANOVA) using morphometric characteristics to reveal differences among the species. Results showed significant differences (p<0.05) among populations of different divisions. Morphometric characteristic analysis on 16 populations of *Polymesoda* can be grouped based on cluster analysis which resulted in six groups at 98% similarity, namely Group 1 (*P. bengalensis* population from Limbang, Lawas, Miri, Bintulu and Kuching), Group 2 (*P. bengalensis* population from Sibu), Group 3 (*P. expansa* population from Miri and *P. erosa* population from Lawas and Miri), Group 4 (*P. erosa* population from Sibu, Mukah and Bintulu), Group 5 (*P. expansa* population from Limbang, Lawas and Bintulu) and Group 6 (*P. erosa* population from Limbang). These six groups of *Polymesoda* population were also supported by the results of Principal Component Analysis (PCA).

Genetic distance analysis through the 5S rDNA primer on *Polymesoda* species (*P. erosa, P. bengalensis* and *P. expansa*) revealed distinct banding patterns for 16 population samples from seven divisions (Limbang, Lawas, Miri, Bintulu, Mukah, Sibu and Kuching) of Sarawak. Band size for *P. erosa* and *P. bengalensis* were ranged from 100-1000 bp, while *P. expansa* 100-500 bp. Finding demonstrated that the most distant genetic was *P. bengalensis* from Bintulu (0.3365) while the close related genetic distance was *P. expansa* (Miri and Bintulu) and *P. bengalensis* (Sibu and Kuching) with 0.000 genetic distant. The results highlight the efficiency of 5S rDNA marker as identification tool which consent the identification of *Polymesoda* species.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Master Sains

MOLLUSKA BOLEH DIMAKAN (GASTROPODA DAN BIVALVIA) DI BAHAGIAN TERPILIH DI SARAWAK

Oleh

HADI BIN HAMLI

Januari 2013

Pengerusi: Mohd Hanafi Bin Idris, PhD

Fakulti: Sains Pertanian dan Makanan

Kepelbagaian siput gastropoda dan siput bivalvia yang boleh dimakan telah diselidik di tujuh bahagian pesisir pantai terpilih di Sarawak iaitu Kuching, Sibu, Mukah, Bintulu, Miri, Limbang dan Lawas dari Julai 2010 sehingga Januari 2012. Taburan spesis sebagai contoh, *Polymesoda* spp. adalah yang paling meluas dan umum telah diselidik terperinci untuk morfologi dan kepelbagaian genetik di antara populasi yang berbeza berdasarkan kawasan kajian.

Sejumlah 41 spesis yang terdiri 11 famili malakologi siput gastropoda dengan 21 spesis dan 12 famili malakologi siput bivalvia dengan 20 spesis telah direkodkan dan dikenal pasti. Bivalvia daripada Corbiculidae (*Polymesoda erosa*, *P. expansa* and *P. bengalensis*) mempunyai taburan yang meluas di mana terdapat pada semua kawasan kajian berbanding dengan spesis siput bivalvia yang lain. Manakala siput gastropoda dari famili Potamididae (*Cerithidia obtusa*, *C. quadrata*, *C. rizophorarum* and

vi

Telescopium telescopium) telah direkod secara umum pada enam daripada tujuh bahagian. Spesis siput gastropoda dan bivalvia dijumpai paling banyak di bahagian Bintulu (13 spesis untuk siput gastropoda dan 11 spesis untuk siput bivalvia), manakala bilangan spesis terendah direkodkan adalah bahagian Sibu dan Mukah. Keputusan Indeks Jaccrad telah menunjukkan kesamaan tertinggi untuk siput bivalvia di antara Limbang lawan Miri (0.75) dan Limbang lawan Lawas (0.75), sementara Sibu lawan Mukah (0.50) untuk siput gastropoda yang boleh dimakan.

Sejumlah 15 ciri morfometrik (6 umum dan 9 ciri tambahan) telah digunakan untuk membezakan tiga spesis *Polymesoda* yang telah direkod daripada Sarawak. Terdapat perbezaan yang ketara (ANOVA, p<0.05) untuk 12 ciri morfometrik (SL; Panjang Cengkerang, SW; Lebar Cengkerang, SH; Tinggi Cengkerang, UL; Panjang Umbo, AL; Panjang Anterior, PL; Panjang Posterior, LPAS; Panjang parut aduktor Posterior ke parut aduktor anterior, LCT; Panjang Gigi Kardinal, PVM; Garis pallial ke hujung, AAAM; Parut aduktor Anterior ke hujung Anterior, PAPM; Parut aduktor Posterior ke hujung Posterior dan VPM; Hujung Posterior ventral) telah dikenalpasti antara tiga spesis *Polymesoda* yang dikumpul dari bahagian yang berbeza. Analisis menunjukkan kadar nisbah untuk LCT/SL adalah ketara berbeza (ANOVA, p<0.05) antara *P. bengalensis* dan *P. expansa* manakala VPM/SL ketara berbeza (ANOVA, p<0.05) antara *P. erosa* dan *P. erosa* dan *P. erosa* dan *P. bengalensis*. Walau bagaimanapun, masing-masing tidak ada perbezaan ketara (ANOVA, p>0.05) antara VPM/SL.

Enam belas populasi yang terdiri *P. erosa, P. expansa* dan *P. bengalensis* direkod dari bahagian berbeza telah dinilai (ANOVA) melalui ciri-ciri morfometrik bagi menunjukkan perbezaan antara spesis. Keputusan menunjukkan perbezaan ketara (p<0.05) antara populasi dari pelbagai bahagian. Analisis ciri morfometrik 16 populasi *Polymesoda* boleh dikumpulkan berdasarkan analisis kluster di mana menghasilkan enam kumpulan pada 98% kesamaan seperti Kumpulan 1 (Populasi *P. bengalensis* dari Limbang, Lawas, Miri, Bintulu dan Kuching), Kumpulan 2 (Populasi *P. bengalensis* dari Sibu), Kumpulan 3 (Populasi *P. erosa* dari Miri dan populasi *P. erosa* dari Lawas dan Miri), Kumpulan 4 (Populasi *P. erosa* dari Sibu, Mukah dan Bintulu), Kumpulan 5 (Populasi *P. erosa* dari Limbang, Lawas dan Bintulu) dan Kumpulan 6 (Populasi *P. erosa* dari Limbang). Enam kumpulan populasi *Polymesoda* ini juga disokong melalui analisis komponen utama (PCA).

Analisis jarak genetik melalui penanda 5S rDNA terhadap spesis *Polymesoda* (*P. erosa, P. bengalensis* dan *P. expansa*) menunjukkan corak jalur yang berbeza untuk 16 sampel populasi dari tujuh bahagian (Limbang, Lawas, Miri, Bintulu, Mukah, Sibu dan Kuching). Saiz jalur untuk *P. erosa* dan *P. bengalensis* pada julat 100-1000 bp, sementara *P. expansa* pada julat 100-500 bp. Penemuan menunjukkan jarak genetic yang jauh adalah dari Bintulu (0.3365) sementara jarak genetik terdekat ditunjukkan oleh *P. expansa* (Miri dan Bintulu) dan *P. bengalensis* (Sibu dan Kuching) dengan jarak genetik 0.000. *P. bengalensis* dari Bintulu merupakan populasi yang unik berbanding populasi lain dari spesis yang sama. Keputusan menunjukkan kecekapan penanda 5S rDNA sebagai alat pengenalpastian yang membantu pengecaman spesis *Polymesoda*.

ACKNOWLEDGEMENTS

All praises be unto the Allah Almighty whose blessings and gave me strength to finish up my Master project and completing this thesis. I would like to thanks to my beloved family that gave me support and inspired me to be more spirited. They taught me to never give up in anything I do. Special thanks to supervisor community members Dr. Mohd. Hanafi Idris, Dr. Abu Hena Mustafa Kamal and Dr. Wong Sing King for their valuable advice and guidance during the completing this project. They gave high commitment to ensure I can finish my project smoothly. Also thanks to lab assistants Mr. Awangku Ahmad Nizam, Mr. Mohd. And Ezwandi Bin Mahrit for their help in water quality analysis and plankton observation. Special thanks to my friends Mohd. Hafizbillah Bin Zawawi, Azimah Binti Abdul Rahim, Lim Chin Tsong Norein Binti Rosli and Zulshafiq Bin Mohd Rubaei for their support. Thank you also to Faculty of Agriculture and Food Sciences, Universiti Putra Malaysia Bintulu Sarawak Campus for technical, logistic supports and laboratory facilities provided. Lastly special thank you Ministry of Higher Education, for the research grant (5523703 FRGS) which make this study possible. I certify that a Thesis Examination Committee has met on 10 January 2013 to conduct the final examination of Hadi Bin Hamli on his thesis entitled "Edible Mollusc (Gastropod and Bivalve) at Selected Divisions of Sarawak" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master Science.

Members of the Thesis Examination Committee were as Follows:

Seca Gandaseca, PhD

Associate Professor Faculty of Agriculture and Food Sciences Universiti Putra Malaysia (Chairman)

Hishamuddin Omar, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Annie Christianus, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Tan Shau Hwai Aileen,PhD

Associate Professor School of Biological Sciences Universiti Sains Malaysia (External Examiner)

SEOW HENG FONG, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd. Hanafi Idris, PhD

Senior Lecturer Faculty of Agriculture and Food Sciences Universiti Putra Malaysia (Chairman)

Abu Hena Mustafa Kamal, PhD

Senior Lecturer Faculty of Agriculture and Food Sciences Universiti Putra Malaysia (Member)

Wong Sing King, PhD

Senior Lecturer Faculty of Agriculture and Food Sciences Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citation which has been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



LIST OF TABLES

Table		Page
3.1	Details location of study area for year 2012 (Department of Statistics Malaysia, 2013)	25
3.2	Ecological parameter of <i>Polymesoda</i> from seven divisions of Sarawak	26
4.1	Species check list of edible bivalve from Sarawak, Malaysia	34
4.2	Distribution of edible bivalve at the selected divisions of Sarawak, Malaysia	36
4.3	Species check list of edible gastropod from Sarawak, Malaysia	100
4.4	Distribution of edible gastropod at the selected divisions of Sarawak	102
5.1	Morphometric characteristic used for bivalve identification with the abbreviation	177
5.2	Analysis of one way ANOVA with general linear model (GLM) for 15 morphometric characteristic of three <i>Polymesoda</i> species	180
5.3	Analysis of one way ANOVA with general linear model (GLM) for 14 morphometric characteristic proportion with shell length of three <i>Polymesoda</i> species	183
5.4	Morphometric characters analysed by ANOVA with GLM or <i>Polymesoda erosa</i> at six different populations	188
5.5	Morphometric characters analysed by ANOVA with GLM for <i>Polymesoda expansa</i> at four different populations	191
5.6	Morphometric characters analysed by ANOVA with GLM for <i>Polymesoda bengalensis</i> at six different populations	194
5.7	Loading of variables on the first two principal components edible bivalve	199
6.1	Matrix table of genetic distance among 16 populations of <i>Polymesoda</i> at Sarawak	220
1A	Tabe 1A: Result of One way ANOVA with GLM for three <i>Polymesoda</i> spp.	248

- 2A Table 2A: Result on one way ANOVA with GLM for proportion 251 analysis of *Polymesoda* spp
- 3A Table 3A: Result on one way ANOVA with GLM for 254 *Polymesoda* spp from different population

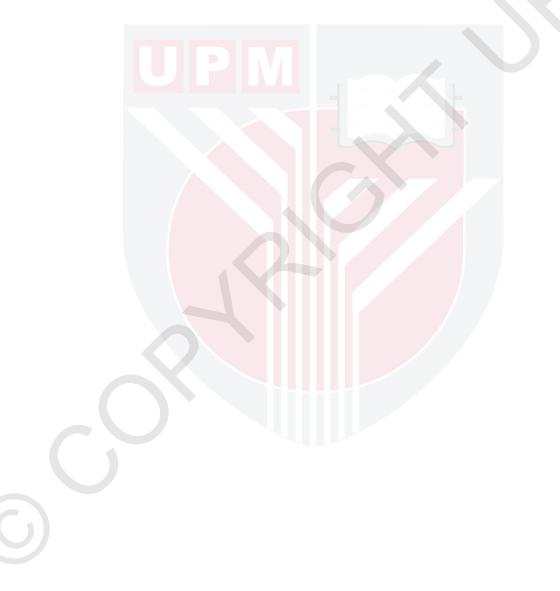


LIST OF FIGURES

Figure		Page
2.1	Previous study on gastropod and bivalve collection in Malaysia	14
2.2	Statistic of shellfish landing in Malaysia for 2011 (Source: Department of Fisheries Malaysia, 2013).	22
3.1	Sampling area location showing the selected divisions in Sarawak, Malaysia	24
4.1	Measurements of shell characters for bivalve and gastropod	32
4.2	Map of edible bivalve distribution at selected divisions of Sarawak	37
4.3	Similarities of collected edible bivalves between divisions using Jaccard's Index	38
4.4	Map of edible gastropod at selected divisions of Sarawak	103
4.5	Similarities of collected edible gastropods between divisions using Jaccard's Index	104
5.1	Map of <i>Polymesoda</i> spp. distribution at selected divisions of Sarawak	174
5.2	Photo on length of cardinal tooth	184
5.3	Photo and diagrammatic sketch of the morphometric ratio on ventral posterior margin (VPM/SL)	185
5.4	Cluster group of <i>Polymesoda</i> at seven divisions of Sarawak based on 15 morphometric characters	197
5.5	First two principal components for <i>Polymesoda</i> genus at selected divisions based on morphometric characteristics	200
6.1	Band appeared after PCR analysis and electrophoresis for six populations of <i>Polymesoda erosa</i>	216
6.2	Band appeared after PCR analysis and electrophoresis for six populations of <i>Polymesoda bengalensis</i>	217

6

- 6.3 Band appeared after PCR analysis and electrophoresis for 218 four populations of *Polymesoda expansa*
- 6.4 Hierarchical cluster based on coefficient of Nei (1978) and 223 unweighted pair group method with arithmetic averaging (UPGMA), modified from NEIGHBOR procedure of PHYLP Version 3.5



LIST OF PLATES

Plate		Page
1	Morphological structure of <i>Polymesoda expansa</i> (Mousson, 1849), A; sketch of inner, B; inner view, C; outer view	41
2	Morphological structure of <i>Polymesoda bengalensis</i> (Lamarck, 1818), A; sketch of inner view, B; inner view, C; outer view	44
3	Morphological structure of <i>Polymesoda erosa</i> (Lightfoot, 1786), A; sketch of inner, B; inner view, C; outer view	47
4	Morphological structure of <i>Meretrix meretrix</i> (Linnaeus, 1958), A; sketch of inner view, B; inner view, C; outer view	50
5	Morphological structure of <i>Meretrix lyrata</i> (Sowerby, 1851) A; sketch of inner view, B; inner view, C; outer view	53
6	Morphological structure of <i>Paphia undulata</i> (Born, 1778) A; sketch of inner view, B; inner view, C; outer view	56
7	Morphological structure of <i>Circe scripta</i> (Linnaeus, 1758) A; sketch of inner view, B; inner view, C; outer view	59
8	Morphological structure of <i>Solen regularis</i> (Dunker, 1862) A; sketch of inner view, B; inner view, C; outer view	62
9	Morphological structure of <i>Solen lamarckii</i> (Deshayes, 1839) A; sketch of inner view, B; inner view, C; outer view	65
10	Morphological structure of <i>Pharella acutidens</i> (Broderip and sowerby, 1828) A; sketch of inner view, B; inner view, C; outer view	68
11	Morphological structure of <i>Amusium pleuronectes</i> (Linnaeus, 1758) A; sketch of inner view B; inner view, C; outer view	71
12	Morphological structure of <i>Anadara granosa</i> (Linnaeus, 1758) A; sketch of inner view B; inner view, C; outer view	74
13	Morphological structure of <i>Pholas orientalis</i> (Gmelin, 1791) A; sketch of inner view, B; inner view, C; outer view	77
14	Morphological structure of <i>Glauconome virens</i> (Linnaeus, 1767) A; sketch of inner view B; inner view, C; outer view	80

15	Morphological structure of <i>Placuna placenta</i> (Linnaeus, 1758) A; sketch of inner view B; inner view, C; outer view	83
16	Morphological structure of <i>Crassostrea lugubris</i> (Sowerby, 1871) A; sketch of inner view, B; inner view, C; outer view	86
17	Morphological structure of <i>Isognomon ephippium</i> (Linnaeus, 1758) A; sketch of inner view, B; inner view, C; outer view	89
18	Morphological structure of <i>Arcuatula arcuatula</i> (Hanley, 1843)A; sketch of inner view, B; inner view, C; outer view	92
19	Morphological structure of <i>Anodonta woodiana</i> (Martens, 1874) A; sketch of inner view, B; inner view, C; outer view	95
20	Morphological structure of <i>Pilsbryoconcha exilis</i> (Lea, 1839) A; sketch of inner view, B; inner view, C; outer view	98
21	Morphological structure of <i>Cerithidea obtusa</i> (Lamarck, 1822), A; sketch of ventral view, B; ventral view, C; dorsal view	107
22	Morphological structure of <i>Cerithidea rizophorarum</i> (Adams, 1855), A; sketch of ventral view, B; ventral view, C; dorsal view	110
23	Morphological structure of <i>Cerithidea quadrata</i> (Sowerby, 1866), A; sketch of ventral view, B; ventral view, C; dorsal view	113
24	Morphological structure of <i>Telescopium telescopium</i> (Linnaeus, 1758), A; sketch of ventral view, B; ventral view, C; dorsal view	116
25	Morphological structure of <i>Nerita articulata</i> (Gould, 1847), A; sketch of ventral view, B; ventral view, C; dorsal view	119
26	Morphological structure of <i>Nerita chamaeleon</i> (Linnaeus, 1758), A; sketch of ventral view, B; ventral view, C; dorsal view	121
27	Morphological structure of <i>Nerita albicilla</i> (Linnaeus, 1758), A; sketch of ventral view, B; ventral view, C; dorsal view	123
28	Morphological structure of <i>Clithon retropictus</i> (Von Marterns, 1879), A; sketch of ventral view, B; ventral view, C; dorsal view	125
29	Morphological structure of <i>Trochus radiatus</i> (Gmelin, 1791), A; sketch of ventral view, B; ventral view, C; vertical view	128
30	Morphological structure of <i>Monodonta labio</i> (Linnaeus, 1758), A; sketch of ventral view, B; ventral view, C; dorsal view	131

31	Morphological structure of <i>Melo melo</i> (Lightfoot, 1786), A; sketch of ventral, B; ventral view, C; dorsal view	134
32	Morphological structure of <i>Planaxis sulcatus</i> (Born, 1778), A; sketch of ventral view, B; ventral view, C; dorsal view	137
33	Morphological structure of <i>Turbo crasus</i> (Wood, 1828), A; sketch of ventral view, B; ventral view, C; dorsal view	140
34	Morphological structure of <i>Thais aculeata</i> (Deshayes and Milne Edwards, 1844), A; sketch of ventral view, B; ventral view, C; dorsal view	143
35	Morphological structure of <i>Ellobium aurisjudae</i> (Linnaeus, 1758), A; sketch of ventral view, B; ventral view, C; dorsal view	146
36	Morphological structure of <i>Melanoides costellaris</i> (Lea, 1850), A; sketch of ventral view, B; ventral view, C; dorsal view	149
37	Morphological structure of <i>Blanocochlis glandiformis</i> (Schepman, 1896), A; sketch of ventral view, B; ventral view, C; dorsal view	152
38	Morphological structure of <i>Brotia costula</i> (Troschel, 1837), A; sketh of ventral view, B; ventral view, C; dorsal view	155
39	Morphological structure of <i>Tylomelania helmuti</i> (Von Rintelen and Glaubrecht, 2003), A; sketch of ventral view, B; ventral view, C; dorsal view	158
40	Morphological structure of <i>Pomacea bredgesii</i> (Reeve, 1856), A; sketch of ventral view, B; ventral view, C; dorsal view; dorsal view	161
41	Morphological structure of <i>Pomacea canaliculata</i> (D Orbigny,	164

41 Morphological structure of *Pomacea canaliculata* (D Orbigny, 164 1835), A; sketch of ventral view, B; ventral view, C; dorsal view

C.

LIST OF ABBREVIATIONS

	ANOVA	Analysis of Variance
	AL	Anterior length
	AW	Aperture width
	AAAM	Anterior adductor scar to anterior margin
	AAPM	Anterior adductor scar to posterior margin
	AW	Anterior adductor scar width
	bp	Base pair
	DNA	Deoxyribonucleic Acid
	FAO	Food and Agriculture Organization
	LCT	Length of cardinal tooth
	LL	Ligament length
	LPAS	Length of posterior adductor scar to anterior adductor scar
	NA	Not available
	ns	No significant difference
	ОР	Outer lip thickness
	PAPM	Posterior adductor scar to posterior margin
	PL	Posterior length
	PCA	Principal Component Analysis
	PCR	Polymerase Reaction Chain
	PRIMER	Plymouth Routines In Multivariate Ecological Research
	PVM	Pallial line to ventral margin
	PW	Posterior adductor scar width

SAS	Statistical Analysis of Software
SH	Shell height
SL	Shell length
SW	Shell Weight
UL	Umbone
UPGMA VPM	Unweighted Pair Group Method with Arithmetic averaging Ventral posterior margin length
ha μL	Hectare Microliters
mL	Mililiters
ng	nanogram
mm	Millimeter

CHAPTER 1

GENERAL INTRODUCTION

1.1 Background Study

Mollusc provides an important source of protein for human besides fish. It is found in many part of the world such as marine, brackish, freshwater and terrestrial area including Southeast Asia. It is believed that seasonal pattern in the Southeast Asia, for instance monsoonal rainfall provide nutrients enriched environment for these filter feeder organisms which eventually help to increase the number of mollusc in this area (Vermeij,1978).

Bivalvia and gastropoda classes are belonging to phylum mollusca. Both of these classes comprise of 290 families (Okutani, 2000). They can be found distributed at the Western Central Pacific area (Poutiers, 1998). In Southeast Asia, about 1211 species of bivalves was reported and it is the highest diversity for bivalves compared to 29 regions around the world (Crame, 2000). Distribution of gastropod and bivalve in Malaysia can be located from east coast (Terenganu and Pahang), west coast (Melaka), south (Johor) of Peninsular Malaysia, Sabah and Sarawak (Idris *et al.*, 2011; Yap and Edward 2010; Zaidi *et al.* 2010; Abdullah *et al.* 2007; Sallih, 2005; Supian and Ikhwanuddin 2002; Hashim, 1993).

In Peninsular Malaysia, gastropod is locally known as "Siput", whereas it is known as "Tekoyong" in Sarawak. This bivalve is locally known as many names depends on species such as "lokan, kepah, ambal, kunau and kerang" in Sarawak.Variable names given by local people are referring to several species of edible mollusc. In term of scientific nomenclature only few studies were recorded regarding non edible and edible mollusc in Indo-Pacific such as, edible bivalve, (Nateewathana, 1995), morphology of Pinnidae (Scheltema, 1983), taxonomic of Pinnidae (Rosewater, 1961), review of *Polymesoda* (Morton, 1984), occurrence on Opistobranch from Hong Kong (Rudman and Darvell, 1990) and embryos of giant clam (Soria-Dengg and Ochavillo, 1990).

Bivalve and gastropod can be differentiated based on shell morphological characteristic. Gastropod could be identified based on one piece, hard and usually coiled shells (Wye, 2007). Whilst bivalve have shells with two hinged and also called as Pelecypoda because of the battle axe foot shape (Feinberg, 2003). These external characters were the basic work on taxonomic to distinguish mollusc species (Rosewater, 1961).

The morphological method can be used with other method to clarify species and distribution (Araujo *et al.*, 1993). Other method is combination of morphological technique and supported with molecular genetics to asses mussel populations (Mass *et al.*, 1999). The molecular work itself as the strong method for species identification. Moreover genetic diversity is more diverse than morphology diversity since genetic variation in natural population come from mutational, genetic

recombination and sexual reproduction (Nguyen *et al.*, 2006). Besides that marine mollusc has gene flow ability which tend to be more homogenous in term of genetic even for large geographic area (Donrung *et al.*, 2011).

Gastropoda and bivalvia classes are the largest and abundance mollusca phylum throughout the world. Furthermore, mollusc from the marine habitat received more attention because of their aesthetic and gastronomic appeals (Subba Rao, 1993). Mollusc diversity, abundance and diversity are related to physico-chemical parameters, climatic condition and soil of the habitat area. This habitat area condition and characters were reported as strong factor influenced mollusc diversity and behaviour (Armitage and Fong, 2004; Chapperon and Seuront, 2011).

Different species is significantly depends on habitat types for instance high proportion of mud suitable for *Anadara granosa* and *Gafiarium tumidum* (Baron and Clavier, 1992; Broom, 1985). Habitat located at tropical water with sandy shallow and sandy reef flats are inhabit by Strombide and *Lambis* respectively (Ramadoss, 2010). Furthermore other gastropod and bivalve species can inhabit wide range habitat from mangrove, freshwater, seagrass bed, intertidal, coral reef and even most extreme habitat such as acidic, high temperature and high sulphide substance.

In Malaysia *Anadara granosa* is found abundantly at the West coast of Peninsular Malaysia due to their suitable habitat with extensive tidal mudflats. Another species that easily can be found is *Perna viridis* which spread to Western coast of Peninsular Malaysia especially Melaka (Sallih, 2005) and this species is native at Malaysian coast (Vakily, 1989). Razor clam (*Solen* spp.) that belongs to Solenidae family can be found at Kuching, Sarawak has high market demand since it found seasonally (Hassan and Hung, 2006).

1.2 Problem Statement

Little information was recorded regarding non edible and edible mollusc in Malaysia which included gastropod diversity around Malaysia (Abu Hena *et al.*, 2004), nursery habitat (Zaidi *et al.*, 2010) and source of edible marine food in Sabah (Hashim, 1993). Lack number of documented mollusc from Malaysia may directly relate on number of habitat preference.

In Sarawak, molluscs inhabit in wetland together with other flora and fauna to form tangible ecosystem that support each other. Commonly in Sarawak, large number of gastropod and bivalve species which derived from wetland is consumed by local people. These molluscs provide essential protein in daily diet of the local communities and some species play important role for fishery economy of the state. The increasing of development activity at Sarawak may expose the major wetland destruction, damages due to pollution, changes of habitat characteristics and condition. Major changes on wetland habitat will affect on the biodiversity including mollusc population which lastly lead toward the extinction of certain species. Besides, pollutants derived from sawmills, wood chip and sago factories may destroy the natural habitat of mollusc and other biodiversity (Davy and Graham, 1982). Over exploitation on edible mollusc by local people could also gradually contribute to species depletion.

To fill the gap of lack of information on edible mollusc diversity, present study was intended to collect information regarding diversity, distribution and morphology of edible mollusc from Sarawak. This study also includes some of their habitat characteristic. The most and widely distributed species was investigated for morphometric characteristics that could differentiate among the similar species. Genetic distance of this species population also was investigated if have had any differences among them.

Documentation of this edible mollusc could have scientific importance as a record that can be used as ready reference in future research activities toward aquaculture development, conservation and proper management of this natural resource in Sarawak.

1.3 Objectives

Objectives of the study are:

- 1. To investigate the diversity, distribution and morphology of edible mollusc at selected divisions of Sarawak.
- To examine the morphometric characteristics of the most common bivalve,
 Polymesoda spp. from the coastal area of Sarawak.
- 3. To investigate the genetic diversity and variation of the *Polymesoda* spp. of the study areas.

REFERENCES

- Abdullah, M.H., Sidi, J. and Aris, A.Z. 2007. Heavy Metal (Cd, Cu, Pb, Zn) in *Meretrix meretrix* Roding, Water and Sediments from Estuaries in Sabah, North Borneo. *International Journal of Environmental and Science Educational*, 2(3): 69–74.
- Abu Hena, M.K., Hsamuddin, O., Misri, K., Abdullah F., and Loo, K.K. 2004. Benthic Faunal Composition of *Penaeus monodon* Fabricius Culture Pond West Coast of Peninsular Malaysia. *Journal of Biology Science*, 4(5): 631–636.
- Akester, R.J. and Martel, A.L. 2000. Shell shape, dysodont tooth morphology, and hinge ligament thickness in the bay mussel *Mytilus trossulus* correlate with wave exposure. *Canadian Journal of Zoological*, 78: 240–253.
- Allen, J.A. 1976. Further evidence for apostatic selection by wild passerine birds-9:1 experiments. *Heredity*, 36:173–180.
- Alunno-Bruscia M., Bourget, E. and Frechette, M. 2001. Shell allometry and length mass density relationship for *Mytilus edulis* in an experimental food regulated situation. *Marine Ecology Progress Series*, 219:177–188.
- Araujo, R., Moreno, D. and Ramos, M.A. 1993. The Asiatic Clam Corbicula fluminea (Muller 1774) (Bivalvia: Corbiculidae) in Europe. American Malacological Bulletin, 10(1): 39–49.
- Armitage, A.R. and Fong, P. 2004. Gastropod Colonization of a Created Coastal Wetland: Potential Influences of Habitat suitability and Dispersal Ability. *Restoration Ecology*, 12(3): 391–400.
- Arneri, E., Giannnetti, G. and Antolini, B. 1998. Age determination and growth of Venus verrucosa L. (Bivalvia: Veneridae) in the southern Adriatic and Aegean Sea. Fisheries Research. 38:193–198.
- Aroon, P.S., Lohachit, C. and Harada, M. 2004. Survey of Brackish Water Snails in Eastern Thailand. *SouthEast Asian Journal Tropical Medical Public Health*, 35(1): 150–155.
- Aziz, A., S.B. Japar and Mutaharah, Z. 2001. Checklist of the shallow water intertidal invertebrates of Pulau Redang, Proceeding of the National Symposium on Marine Park and Terengganu Island: Toward sustainable Usage and Management of Islands, Kuala Terengganu, pp: 12–18.
- Babaei, M.M., Sahafi, H.H., Ardalan, A.A., Ghaffari, H. and Abdollahi, R. 2010.
 Morphometric Relationship and of Weight and Size of Clam *Amiantis umbonella* L., 1818 (Bivalvia: Veneridae) In the Eastern Coasts of Bandar Abbas, Persian Gulf. *Advances in Environmental Biology*, 4(3): 376-382.

- Bagatini, Y.M., Panarari, R.S., Higuti, J., Cecillo, E.B., Prioll, A.J. and Prioli, S.M.A.P. 2005. Morphological and molecular characterization of *Corbicula* (Mollusca, Bivalvia) at Rosana Resevoir, Brazil. *Acta Science Biology Science*, 27(4): 397–404.
- Baker, A.J. 2000. Molecular Methods in Ecology. Blackwell Science, Oxford. 337 pp.
- Barker, G.M. 2001. Gastropod on Land, Phylogeny, Diversity, and Adaptive Morphology: The Biology of Terrestrial Molluscs, G.M. Barker, Ed. CAB International, pp 1–146.
- Baron, J. and Clavier, J. 1992. Effects of Environmental Factors on the Distribution of the Edibles Bivalves *Atactodea striata*, *Gafrarium tumidum* and *Anadara scapha* on the coast of New Caledonia (SW Pacific). *Aquatic Living Resource*, 5:107–114.
- Beukema, J.J. and Meehan, B.W. 1985. Latitudinal variation in linear growth and other shell characteristic of Macoma balthic. *Marine Biology*, 90:27–33.
- Brandt, R.A.M. 1974. The Non Marine Aquatic Mollusc of Thailand. Archiv Fur Molluskenkunde, vol. 105(i-iv): 1–423.
- Braullio de Luna Sales, J., Rodrigues-Filho, L.F.S., Haimovichi, M., Sampaio, I. and Schneider, H. 2011. Molecular differentiation of the species of two squid families (*Loliginidae* and *Ommastrephidae*) based on PCR study of the 5S rDNA gene. *Food Control*, 22: 90–98.
- Broom, M. J. 1985. The Biology and Culture of Marine Bivalves Molluscs of Genus *Anadara*. ICLARM Studies and Reviews, 12:37.
- Brown, D.S. 1971. Ecology of Gastropoda in a South African Mangrove swamp. Proceeding of the Malacological society of Landon, 39: 263–279.
- Canapa, A., Barucca, M., Marinelli, A. and Olmo, E. 2001. A Molecular Phylogeny of Heterodonta (Bivalvia) Based on Small Ribosomal Subunit RNA Sequences. Molecular Phylogenetic and Evolution, 21(1): 156–161.
- Chapperon, C. and Seuront, L. 2011. Variability in the Motion Behavior of Intertidal Gastropod: Ecology and Evolutionary Perspectives. *Journal of the Marine Biology Association of the United Kingdom*, 91(1): 237–244.
- Clarke, B.C., Arthur, W., Horsley, D.T. and Parkin, D.T. 1978. Genetic variation and natural selection. in pulmonate mollusks. In *Pulmonates*, ed. V. Fretter, and J. Peake, Vol. 2A, pp. 219–270., *Systematic, Evolution and Ecology*. Academic press. New York.

Clarke, K.R., Gorley, R.N., 2001. Primer Version 5. Primer-E, Plymouth, UK.

- Claxton, W.T., Wilson, A.B., Mackie, G.L. and Boulding, E.G. 1998. A Genetic and Morphological comparison of shallow and deep water populations of the introduced dreissenid bivalve Dreissena bugensis. *Canadian Journal of Zoology*, 76:1269–1276.
- Claudi, R. and Mackie, G.L. 1994. Practical manual for zebra mussel monitoring and control. Lewis Publishers, Boca Raton.
- Costa, C., Aguzii, J., Menesatti, P., Antonucci, F., Rimatori, V. and Mattoccia, M. 2008. Shape analysis of different populations of clams in relation to their geographical structure. *Journal of Zoological*, 276: 71–80.
- Crame, J.A. 2000. Evolution of Taxonomic Diversity Gradients in the Marine Realm: Evidence from the Composition of Recent Bivalves Faunas. *Paleaobiology*, 26(2): 188–214.
- Creed, J.C. and Kinupp, M. 2011. Small scale change in mollusk diversity along a depth gradient in a seagrass bed off Cabo Frio, (Southeast Brazil). *Brazilian Journal of Oceanography*, 59(3): 267–276.
- Davy, F.B. and Graham, M. 1982. Bivalve Culture in Asia and the Pacific: Proceedings of Workshop held in Singapore. Ottawa, IDRC, pp 90.
- Department of Fisheries Malaysia. 2013. Landings of Marine fish by Month and state 2011. http:// http://www.dof.gov.my. Retrieved 20 January 2013
- Department of Statistic Malaysia. 2013. Statistic population of Sarawak 2010. http://www.statistics.gov.my. Retrieved 20 January 2013.
- Dewitt, T.J., Sih, A. and Hucko, J.A. 1999. Trail and compensation and cospecialization in a freshwater snail: size, shape and antipredator behavior. *Animal Behaviour*, 58:397–407.
- Donrung, P., Tunkijjanukij, S., Jarayabhand, P. and Poompuang, S. 2011. Spatial Genetic Structure of the Surf Clam *Paphia undulata* in Thailand Waters. *Zoological Studies* 50(2): 211–219.
- Endler, J.A. 1986. Natural Selection in the Wild. Princeton University Press, New Jersey.

Feinberg, H.S. 2003. Shells. Simon and Schuster Inc. pp. 512

Food and Agriculture Organization (FAO). 2011. National Aquaculture Sector Overview:Malaysia.Available:http://www.fao.org/fishery/countrysector/naso_m alayisa/en. Retrieved 25 January 2012

- Franz, D.R. 1993. Allometry of Shell and body weight in relation to shore level in the intertidal bivalves Geukensia demissa (Bivalvia: Mytilidae). *Journal of Experimental Marine Biology and Ecology*, 174:193–207.
- Frith, D.W., Tantanasiriwong, R. and Bhatia, O. 1976. Zonation of Macrofauna on a Mangrove Shore, Phuket Island. *Phuket Marine Biological Center Research Bulletin*, 10: 1–37.
- Fuiman, L.A., Gage, J.D. and Lamont, P.A. 1999. Shell Morphometry of deep sea protobranch bivalve Ledella pustulosa in the Rockall Trough, North-East Atlantic. *Journal of the Marine Biological Association of the United Kingdom*, 79:661–671.
- Gibbs, P.E. 1978. Macrofauna of the Intertidal Sand Flats on Low Wooded Islands Northen Great Barrier Reef. *Philosophical Transaction of the Royal Society*, 283:81-97.
- Gillespie, J.H. 1998. Population genetics: A concise Guide, John Hopkins Press. ISBN 0-8018-5755-4.
- Glaubrecht, M., Feher, Z. and Kohler, F. 2007. Inventorizing an invader: Annotated type catalogue of Corbiculidae Gray, 1847 (Bivalvia, Heterodonta, Veneroidea) including old world limnic *Corbicula* in the Natural History Museum Berlin. *Malacologia*, 49(2): 243–272.
- Grandjean, F., Bouchan, D. and Souty-Grosset, C. 2002. Systematics of the European Crayfish species *Austropotamobius pallipes* (Decapoda: Astacidae) with a re-examination of the status of *Austropotamobius bernhauseri*. *Journal of Crustacean Biology*, 22(3): 677–681.
- Han, W., Lui, J., He, X., Cai, Y., Ye, F., Xuan, L. and Ye, N. 2003. Shellfish and Fish Biodiversity of Mangrove Ecosystems in Leizhou Peninsula, China. *Journal of Coastal Development*, 7(1): 21–29.
- Hare, M.P., Palumbi, S.R. and Butman, C.A. 2000. Single-step species identification of bivalve larvae using multiplex polymerase chain reaction. Springer-Verlag, *Marine Biology*, 137: 953–961.
- Hasse, M. and Schilthuizen, M. 2007. A new Georissa (Gastropoda: Neritopsina: Hydrocenidae) from a limestone cave in Malaysian Borneo. *Journal of Molluscan*, 73: 215–221.
- Hashim, R. 1993.Sumber Makanan Laut Pesisiran Laut Sabah. Dewan Bahasa dan Pustaka, pp. 309.

- Hassan R. and Hung T.M. 2006. Razor Clam (*Solen* spp.) of Kuching bay: Priliminary work on morphological assessment and total genomic DNA isolation. Proceeding of Conference on Natural Resources in the Tropics: Development and commercialization of Tropical Natural Resource, Universiti Malaysia Sarawak, 141–143.
- Hinch, S.G. and Bailey, R.C. 1988. Within and among lake variation in shell morphology of the freshwater clam Elliptio complanata (Bivalvia: Unionidae) from south-central Ontario lakes. *Hydrobiologia* 157: 27–32.
- Hyman, L.H. 1992. The Invertebrates: Mollusca 1 Applacophora, Polyplacophora, Monoplacophora, Gatsropoda, The Coelolmate Bilateral, Vol. 6. MacGraw-Hill, pp.769.
- Idris, M.H., Arshad. A., Japar Sidik, B., Mazlan A.G. and Daud, S.K. 2009. Morphological Characteristics of *Pinna bicolor* Gmelin and *Pinna deltodes* Menke from Seagrass Bed of Sungai Pulai, Johor, Peninsular Malaysia. *Sains Malaysiana*, 38(3): 333–339.
- Idris, M.H., Arshad, A., Amin S.M.N., Japar Sidik B., Daud, S.K., Mazlan, A.G., Zakaria, M.S. and Yusoff, F.M. 2011. Age, Growth and Length-Weight Relationships of *Pinna bicolor* Gmelin (Bivalvia: Pinnidae) in the Seagrass Beds of Sungai Pulai Estuary, Johor, Peninsular Malaysia. *Journal Applied Ichthyology*, pp 1–4.
- Jan, A.P. 2005. Biology of the Invertebrates. Fifth edition, MacGrawhill.
- Jin, L. and Chakraborty, R. 1994. Estimation of Genetic and Coefficient of Gene Diversity from Single-Probe Multilocus DNA Fingerprinting Data. *Molecular Biology and Evolution*, 11(1): 120–127.
- Jiang, J.X. and Li, R.G. 1995. An Ecological Study on the Mollusc in Mangrove Areas in the Estuary of Jiulong River. *Hydrobiologia*, 295: 213–220.
- Jokinen, E.H. 1982. Cipangopaludina chinensis (Gastropoda: Viviparidae) in North America, review and update Nautilus 96: 89–95.
- Katoh, M. and Foltz, D.W. 1994. Genetic Subdivision and morphological variation in a freshwater snail species complex formerly referred to as Viviparus georgianus (Lea). *Biology Journal of the Linnean Society*, 53:73–95.
- Kanakaraju, D., Jios, C. and S.M., Long. 2008. Heavy metal concentration in the razor clams (*Solen* spp.) from Muara Tebas, Sarawak. The Malaysian Journal of Analytical Sciences, 12(1): 53–58.
- King, J.R. and Porter, S.D. 2004. Recommendation on the use of alcohols for preservation of ant specimens (Hymenoptera, Formicidae). *Insectes Sociaaux*, 51: 197–202.

- Klinbunga, S. Pripue, P., Khamnamtong, N., Punaglarp, N. Tassanakajon A., Jarayabhan, P. Hirono, I., Aoki, T. and Menasveta P. 2003. Genetic Diversity and Molecular Marker of the Tropical Abalone (*Haliotis asinina*) in Thailand. *Marine Biotechnology* 5: 505–517.
- Kohler, F. and Glaubrecht, M. 2001. Toward a Systematic Revision of the Southeast Asian Freshwater Gastropod *Brotia* H. Adams, 1866 (Cerithiodea: Pachychilidae): An Account of Species from Around the South East China Sea. *Journal of Molluscan studies*, 67: 281–318.
- Kohler, F. and Glaubrecht, M. 2002. Morphology and Reproductive Biology and Molecular Genetics of ovoviviparous freshwater gastropods (Ceritioidea, Pachychilidae) from the Philippines, with Description of a New Genus Jagora. Zoological Scripta, 32: 35–59.
- Kovitvadhi, S., Kovitvadhi, U., Swangwong, P., Trisaranuwatana, P. and Machado, J. 2009. Morphometric Relationship of Weigth and Size of Cultured Freshwater Pearl Mussel, *Hyriopsis (Limnoscapha) myersiana*, Under Laboratory Conditions and Earthen Pond Phases. *Aquaculture International*, 17: 57–67.
- Kovitvadhi, S., Kovitvadhi, U., Sawangwong, P., Thongpan, A. and Machado, J. 2006. Optimization of Diet and Culture Environment for Larvae and Juvenile Freshwater Pearl Mussels, *Hyriopsis (Limnoscapha) myersiana* Lea, 1856. *Invertebrate Reproduction and Development*, 49: 61–70.
- Kripa, V and Appukuttan, K K. 2003. Status of Exploited Marine Fishery Resources of India: *Marine Bivalves*, M. Mohan Joseph and A.A. Jayaprakash, Ed. CMFRI, Cochin, 211-220
- Lajtner, J., Marusic, Z., Klobucar, G.I.V., Maguire, I. and Erben, R. 2004. Comparative shell morphology of the zebra mussel, Dreissena polymorpha in the Drava River (Croatia). *Biologia*, 59:595–600.
- Laxmilatha, P., Sivadasan, M.P. and Surendranathan, V.G. 2007. Bivalve resources and its exploitation in Malabar. *Marine Fish Information Service Technical and Extension Series*, 192: 6-9.
- Ludwig, A.J. and Reynolds, J.F. 1988. Statistical Ecology: A PRIMER on Methods and Computing. A Wiley-Interscience publication Pp. 337.
- Macdonald, K.B. 1969. Quantitative study of salt Marsh Mollusc Faunas from the North American Pacific Coast, *Ecological Monograph*, 39(1): 33–60.
- Macintosh, D.J., Ashton, E.C. and Havanon, S. 2002. Mangrove Rehabilitation and Intertidal Biodiversity: A Study in the Ranong Mangrove Ecosystem Thailand. *Estuarine, Coastal and Shelf Science*, 55: 331–345.

- Mass, P.A.Y., O'Mullan, G.D., Lutz, R.A. and Vrijenhoek, R.C. 1999. Genetic and Morphometric Characterization of Mussels (Bivalvia: Mytilidae) from Mid-Atlantic Hydrothermal Vents. *Biological Bulletin*, 196: 265–272.
- Marin, S.A., Haye, P.A., Marchant, S. and Winkler, F.M. 2007. Molecular Markers used to Analyze Species-specific status in Abalones with Ambiguous Morphology. *Journal of Shellfish Research* 26(3): 833–837.
- Marlina, Radu, S., Kqueen, C.Y., Napis, S. Zakaria, Z., Mutalib, S.A. and Nishibushi, M. 2007. Detection of TDH and TRH Gene in *Vibrio Parahaemolyticus* Isolated from *Corbicula Moltkiana* Prime in West Sumatera, Indonesia. *Southeast Asian Journal of Tropical Medicine and Public Health*, 38(2): 349–355.
- Marwoto, R.M. and Djajasasmita, M. 1994. Dimorfisme Seksual Pada Keong Gondang Pila ampulacea (Linnaeus, 1758). Masyarakat Zoologi Indonesia, 24: 1–8.
- Metabos, M., Bris, N.L., Pendlebury, S. and Thiebaut, E. 2008. Role of Physicochemical environment on Gastropod assembladges at hydrothermal vents on the East Pacific Rise (13°N/EPR), *Journal of the Marine Biological Association of the United Kingdom*, 88(5): 995–1008.
- Moghraby, A.I.E and Adam, M.E. 1984. Ring formation and annual growth in *Corbicula consobrina*, 1827 (Bivalvia, Corbiculidae). *Hydrobiologia*, 110: 219–225.
- Morris, S., and R.D., Purchon. 1981. The Marine Shelled Mollusca of West Malaysia and Singapore. II. Polyplacophora and Gatsropoda. *Journal of Molluscan Studies*, 47:313–321.
- Morton, B. 1976. The biology and functional morphology of the Southeast Asian mangrove bivalve, *Polymesoda (Geloina) erosa* (Solander 1786) (Bivalvia: Corbiculidae). *Canadian Journal of Zoology*. 54: 482–500.
- Morton, B. 1984. Review of *Polymesoda* (*Geloina*) Gray 1842 (Bivalvia: Corbiculacea) from Indo-Pacific Mangroves. *Asian Marine Biology*, 1: 77–86.
- Mujiano, N. 2009. Mudwhelks (Gastropoda: Potamididae) from Mangroves of Ujung Kulon National Park, Banten. *Jurnal Biologi*, 13(2): 51–56.
- Myers, P., Espinosa, R., Parr, C.S., Jones, T., Hammond, G.S. and Dewey, T.A. 2013. The animal diversity web. http://animaldiversity.org. Retrieved 20 January 2013.
- Nakao, S., Nomura, H. and. Satar, M.K.B.A. 1989. Macrobenthos and Sedimentary Environments in a Malaysian Intertidal Mudflat of the Cockle bed. *Bulletin of the Faculty of Fisheries Hokkaido University*, 40(4): 203s–213.

- Narayanan, B. and Sivadas, P. 1986. Studies on the Intertidal Macrofauna of the Sandy Beach at Kavaratti Atoll (Lakshadweep). *Mahasagar Bulletin National Institute Oceanography*, 19:84–98.
- Nateewathana, A. 1995.Taxanomic Accaount of Commercial Edible Molluscs, Excluding Cephalopodas, of Thailand. *Phuket Marine Biological Center*, 20: 93–116.
- Newell, C.R. and Hidu, H. 1982. The effects of sediment type on growth rate and shell allometry in the soft shelled clam Mya arenaria (L). *Journal of Experimental Marine Biology and Ecology*, 65: 285–295.
- Nguyen, T.T.T., Hurwood, D., Mather, P., Na-Nakorn, U., Kamonrat, W. and Bartley, D. 2006. Manual on Applications of Molecular Tools in Aquaculture and Inland Fisheries Management, Part 1: Conceptual basis of population genetic approaches NACA Monograph No. 1, pp 80.
- Okutani, T. 2000. Marine Mollusks in Japan. Tokai University Press. pp 1173.
- Ong, C.C., Yusoff, K., Yap, C.K. and Tan, S.G. 2009 .Genetic characterization of *Perna viridis* L. in peninsular Malaysia using microsatellite markers. *Journal Genetic* 88, 153–163.
- Ostend. 2006. Marine Biodiversity Data Mobilisation Workshop on Molluscs. UNESCO.
- Othman, A.S. 2004. Teknik- Teknik asas Penanda DNA. In Penanda DNA untuk Kajian Kepelbagaian Genetik, ed A.S. Othman, pp. 44–60. Universiti Sains Malaysia, Pulau Pinang.
- Page, S., 2011. Biodiversity Information on Peat swamp Forest in S.E. Asia. Available:http://www.strapeat.alterra.nl/download/6%20biodiversity%20inform ation.pdf. Retrieved 20 August 2011.
- Palmer, A.R. 1985. Quantum Changes in Gastropod Shell Morphology Need Not Reflect Speciation. *Evolution*, pp 699–705.
- Pathansali, D. 1966. Notes on The Biology Cockle Anadara granosa L. *Proceeding*. *Indo-Pacific Fish Council*, 11:84-98.
- Pattikawa, J.A. 2003. Random Amplified Polymorphic DNA (RAPD) Analysis of the Cockle, *Cerastoderma edule* Population. *Ichthyos*, 2(1): 33–36.
- Paul, C.P.K. 2011. Management of Mangrove Forest of Sarawak. http://www.sarwakforestry.com/pdf/hj7-wetland10.pdf. Retrieved 20 August 2011.

- Perez, K.E., Clark, S.A. and Lydeard, C. 2004. A Primer to Freshwater Gastropod Identification. In Freshwater Gastropod Identification Workshop. pp 1–61.
- Pigneur, L.M., Risterucci, A.M., Dauchot, N., Li, X. and Doninck, K.V. 2011. Development of novel microsatellite markers to identify the different invasive lineages in the *Corbicula* complex and to assess androgenesis. *Molecular Ecology Resources*, 11: 573–577.
- Pinhal, D., Yoshimura, T.S. Araki, C.S and Martins, C. 2011. The 5S rDNA family evolves through concerted and birth-and-death evolution in fish genomes: an example from freshwater stingrays. *BMC Evolutionary Biology*, 11: 151
- Piskur, J. and Rupprecht, A. 1995. Aggregated DNA in ethanol solution. *FEBS Letters* 375: 174–178.
- Poutiers, M. 1998. The Living Marine Resources of the Western Central Pacific: FAO Species Identification Guide for Fishery Purpose, K.E. Carpenter and V.H. Niem, Ed. Rome, Italy: Food and Agriculture Organization of the United Nations, 1: 123–362.
- Printrakoon, C., Wells, F.E. and Chitramyong, Y. 2008. Distribution of Molluscs in Mangrove at Six Sites in the Upper Gulf of Thailand. *The Raffles Bulletin of Zoology*, 18: 247–257.
- Purchon, R.D., Purchon, D.E.A. 1981. The Marine Shelled Mollusca of West Malaysia and Singapore. I. General Introduction and an account of Collections. *Journal of Molluscan Studies*, 47:290–312.
- Ramadoss, K. 2010. Gastropods. In Status of Exploited Marine Fishery Resource of India. Ed. M. M. Joseph and A. A. Jayaparaksh. *Central Marine Fisheries Research Institute*, pp. 203–210.
- Ramesh, R. and Ravichandran, S. 2008. A Statistical Approach of the Length-Weight Relationship and Allometry of *Turbo brunneus*. *Research Journal of Environmental Science*, 2(2): 124–131.
- Rice, E.L. 1990. Nucleotide sequence of the 18S ribosomal RNA gene from the Atlantic sea scallop *Placopecten magellanicus* (Gmelin, 1791). *Nucleic Acid Research*, 18(18): 5551.
- Riddoch, B.J. 1993. The adaptive significance of electropheretic mobility in phosphoglucose isomerase (PGI). *Biological Journal of the Linnean Society* 50:1–17.
- Rosewater, J. 1961. The Family Pinnidae in the Indo-Pacific. *Indo-Pacific Mollusca*, 1(4): 175-226.

- Rudman, W.B. and Darvell, B.W. 1990. Opisthobranch mollusc of Hong Kong: Part
 1. Goniodorididae, Onchidorididae, Triophidae, Gymnodorididae, Chromodorididae (Nudibranchia). Asian Marine Biology, 7:31–79.
- Sallih, K. 2005. Mussel Farming in the State of Sarawak, Malaysia: A Feasibility Study. *Fisheries Training Programme*. pp. 44.
- SAS® Institute Inc. 2004. SAS ODBC Driver 9.1: User's Guide and Programmer's Reference. Cary, NC: SAS Institute Inc.
- Scheltema, A. 1983. *Pinna deltodes* Menke newly described and differentiated from *P. Bicolor* Gmelin (Bivalvia, Ptreioida). Journal Malacological Society of Australia, 6(1–2): 37-52.
- Schilthuizen, M., Chai, H.N., Kimsin, T.E. and Vermeulen, J.J. 2003. Abundance and Diversity and Land-Snails (Mollusca: Gastropoda) on Limestone Hills in Borneo. *The Raffles Bulletin of Zoology*, 51(1): 35–42.
- Sergeyev, V.G., Mikhailenko, S.V. Pyshkina, O.A. Yaminsky I.V. and Yoshikawa, K. 1999. How does alcohol dissolve the complex of DNA with cationic surfactant? *Journal of the American Chemical Society*, 121: 1780–1785.
- Singh, M., Kumar R., Nagpure, N.S., Kushhawa B., Mani, I., Chauhan, U.K. and Lakra, W.S. 2009. Population distribution of 4S and 5S rDNA in golden mahseer, *Tor putitora*: population-specific FISH marker. *Journal of Genetics*, 88(3): 315–320.
- Siti, A.M.N. 2004. Tindak Balas Rantaian Polimerase (PCR) dan Polimorfisme DNA Rawak Teramplifikasi (RAPD). In *Penanda DNA untuk Kajian Kepelbagaian Genetik*, ed A.S. Othman, pp. 61–72. Universiti Sains Malaysia, Pulau Pinang.
- Somchai, B. 1995. The Market Value of Rare and Common Molluscs, Phuket Island, Thailand. *Phuket Marine Biological Center*, 20: 35–38.
- Soria-Dengg, S. and Ochavillo, D. 1990. Comparative toxicities of trace metals on embryos of giant clam, *Tridacna derasa*. Asian Marine Biology 7: 161–166.

Spencer, B.E. 2002. Molluscan Shellfish Farming. Fishing News Books. pp. 274.

- Subba Rao, N.V. 1993. Freshwater mollusc of India. In Recent Advances in Freshwater.
- Supian, Z. and Ikhwanuddin, A.M. 2002. Population Dynamic of Freshwater Molluscs (Gastropod: *Melanoides Tuberculata*) in Croker Range Park, Sabah. ASEAN Review of Biodiversity and Environmental Conservation (ARBEC).

- Supriyantini, E., Ambariyanto and Widowati, I. 2007. The Influence of Natural Food *Tetraselmis chuii* dan *Skeletonema costatum* on the profile of the Unsaturated Fatty Acid in Marsh Clam *Polymesoda erosa. Jurnal Pasir Laut*, 3(1): 46–60.
- Suresh, M., Arularasan, S. and Ponnusamy, K. 2012. Distribution of molluscan fauna in the artificial mangrove of Pazhayar back water canal, southeast Coast of India. *Advanced in Applied Science Research*, 3(3): 1795–1798.
- Tan, K.S. and Sigurdsson, J.B. 1990. A new species of *Thais* (Gastropoda: Muricidae), from Singapore and Peninsular Malaysia. *Raffles Bulletin of Zoology*, 38: 205–211.
- Tan, K.S. and Kastoro, W.W. 2004. A Small Collection of Gastropod and Bivalves from the Anambas and Natura Islands, South China Sea. *Raffles Bulletin of Zoology*, 11: 47-54.
- Tan, S.K. and Clements, R. 2008. Taxonomy and Distribution of Neritidae (Mollusca: Gastropoda) in Singapore. *Zoological Studies* 47(4): 481–494.
- Tang, S., Popongviwat, A., Klinbunga S., Tassanakajon, A. Jarayabhand, P. and Menasveta, P. 2005. Genetic Heterogenity of the Tropical Abalone (*Haliotis* asinia) Revealed by RAPD and Microsatellite Analyses. Journal of Biochemistry and Molecular Biology, 38(2): 182–190.
- Teusch, K.P., Jones, D.S. and Allmon, W.D. 2002. Morphological variation in Turritellid Gastropods from the Pleistocene to Recent of Chile: Association with Upwilling Intensity, *PALAIOS*, 17: 366–377.
- Toral-Barza, L. and Gomez, E.D. 1985. Reproductive cycle of the cockle Anadara aniquata L. in Catalangas. Batangas, Philippines. Journal of Coastal Research, 1:141–245.
- Ubukata, T. 2003. A morphometric study on morphological plasticity of shell form in crevice-dwelling Pterioida (Bivalvia). *Biological Journal of the Linnean Socciety*, 79: 285–297.
- Vakily, J.M. 1989. The biology and culture of mussels of the Genus *Perna*. International Center for Living Aquatic Resources Management, Manila, Philippines, pp. 1–63.
- Vermeij, G.J. 1978. Biogeography and adaption. Patterns of Marine Life. Harvard University Press, Cambridge, Massachusetts.
- Vierna, J., Jensen, K.T., Martinez-Lage, A. and Gonzalez-Tizon, A.M. 2011. The linked units of 5S rDNA and U1 snDNA of razor shells (Mollusca: Bivalvia: Pharidae). *Heredity*, 107: 127–142.

- Vizoso, M. Vierna, J. Gonzalez-Tizon, A.M. and Martinez-Lage A. 2011. The 5S rDNA Gene Family in Mollusks: Characterization of Transcriptional Regulatory Regions, Prediction of Scondary Structure, and Long-Term Evolution, with Specieal Attention to Mytillidae Mussels. *Journal of Heredity*, 102(4): 433–447.
- Volye, V. 1975. The Habit and Functional anatomy of the West African Bloody cockle Anadara senilis (L) Proceeding Malacological Society of London, 41:277–299.
- Von Cosel, R., Comtet, T.and Krylova, E. 1997. Gene flow and genetic diversity in naturally fragmented metapopulations of deep-sea hydrothermal vent animals. *Journal of Heredity*, 88:285–293.
- Wang, L. Zhang, H., Song, L. and Guo, X. 2007. Loss of allele diversity in introduction of the hermaphroditic bay scallop *Argopecten irradians*. *Aquaculture*, 271: 252–259.
- Way, K., and Purchon, R.D. 1981. The Marine Shelled Mollusca of West Malaysia and Singapore. III. Bivalvia. *Journal of Molluscan Studies*, 47:322–327.
- Webb, J.E., Wallwork, J.A. and Elgood, J.H. 1978. Guide to Invertebrate Animals, Second Edition, Macmillan Press Ltd. pp 310.
- Wells, F.E. 1986. Distribution of Molluscs Across a Pneumatophore Boundary in a Small Bay in North-Western Australia. *Journal of Molluscan Studies*, 52: 83–90.
- Wye, K. 2007. Pocket Guide to Shell. Silverdale Books, pp. 240.
- Wong, N.L.W.S., Alfian, A.A.K., Affendi, Y.A., Ooi, J.L.S., Badrul H.T. and Yusri, Y. 2008. The diversity of marine mollusc in the Southwestern Reefs of Pulau Tioman, Malaysia. Proceedings of the NAGISA Western Pacific Conference, Jakarta, Indonesia.
- Yang, J., Li, Q., Kong, L., Zheng, X. and Wang, R. 2008. Genetic Structure of the Veined Rapa Whelk (*Rapana venosa*) Populations along the Coast of China. Biochemical Genetic, 46: 539–548.
- Yap, C.K. and Edward, F.B. 2010. Distribution of Heavy Metals in the Different Parts of *Cerithidea obtusa* and the Relationhips between Metal Distribution and Allometric Parameters of the Snail. *Environment Asia*, 3(2): 38–48.
- Yap, C.K., Hisyam, M.N.D., Edward, F.B., Cheng, W.H. and Tan, S.G. 2010. Concentration of Heavy Metal in Different Parts of the gastropod, *Faunus ater* (Linneaus), Collected from Intertidal Areas of Peninsular Malaysia. *Pertanika Journal Tropical Agriculture Science*, 33(1): 45–60

- Yuh-Wen, C., Hon-Cheng, C., Sin-Che, L. and Chaolun, A. C. 2001. Morphometric Analysis of Shell and Operculum Variation in Viviparid Snail, *Cipangopaludina chinensis* (Mollusca: Gastropoda), in Taiwan. *Zoological Studies*, 41(3): 321–331.
- Zaidi, C.C., Arshad, A., Bujang, J.S., Muda W.L.W. and Ghaffar M.A. 2010. Metamorphosis Induction of the Dog *Strombus canarium* (Gastropoda: Strombdae) Using Cues Associated with Nursery Habitat. *Journal of Applied Sciences*, pp 1–8.
- Zaidi, C.C., Aziz, A., Wan-Lofti W.M., and Mazlan, A.G. 2008. Gastropod and bivalve molluscs associated with the seagrass bed at Merambong Shoal, Johor Straits, Malaysia. Malaysia marine ecosystem: The studies of Johor Darul Takzim East Coasts. Marine Ecosystem Research Centre (EKOMAR), National University of Malaysia, pp. :89–99.

LIST OF PUBLICATIONS

- Hamli, H., Idris, M. H., Abu Hena, M. K. and Wong, S. K. (2012). Taxonomic Study of Edible Bivalve from Selected Division of Sarawak, Malaysia. *International Journal of Zoological Research*, ISSN, 1811-9778, v(8): 52-58.
- Hamli, H., Idris, M. H., Abu Hena, M. K. and Wong, S. K. (2012). Diversity of Edible Mollusc (Gastropoda and Bivalvia) at Selected Division of, Sarawak. *International Journal on Advanced Science Engineering Information Technology*, ISSN, 2088-5334, v(2): 5-7
- Hamli, H., Idris, M. H., Abu Hena, M. K., Wong, S. K. and Arshad, A. (2013) Checklist and Habitat description of edible gastropods from Sarawak, Malaysia. *Journal of Fisheries and Aquatic Science*. ISSN, 1816-4927, 8(2): 412–418.