



**UNIVERSITI PUTRA MALAYSIA**

***BIOLOGICAL CONTROL OF ROOT KNOT NEMATODES (*Meloidogyne incognita*) ON BLACK PEPPER (*Piper nigrum* L.) USING *paecilomyces lilacinus* and *bacillus thuringiensis****

**PAU CHEN GUAN**

**FSPM 2012 5**

**PAU CHEN GUAN**

**MASTER OF SCIENCE**

**2012**

**BIOLOGICAL CONTROL OF ROOT KNOT  
NEMATODES (*Meloidogyne incognita*)  
IN BLACK PEPPER (*Piper nigrum* L.) USING  
*Paecilomyces lilacinus* and *Bacillus thuringiensis***



**PAU CHEN GUAN**

**MASTER OF SCIENCE**

**UNIVERSITI PUTRA MALAYSIA**

**2012**

**BIOLOGICAL CONTROL OF ROOT KNOT NEMATODES (*Meloidogyne incognita*)  
IN BLACK PEPPER (*Piper nigrum* L.) USING  
*Paecilomyces lilacinus* and *Bacillus thuringiensis***

**By**

**PAU CHEN GUAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Master of Science**

**August 2012**

**BIOLOGICAL CONTROL OF ROOT KNOT  
NEMATODES (*Meloidogyne incognita*) IN  
BLACK PEPPER (*Piper nigrum* L.) USING  
*Paecilomyces lilacinus* and *Bacillus thuringiensis***



**MASTER OF SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

**2012**

## TABLE OF CONTENTS

	Page
<b>DEDICATION</b>	i
<b>ABSTRACT</b>	ii
<b>ABSTRAK</b>	iv
<b>ACKNOWLEDGEMENTS</b>	vi
<b>APPROVAL</b>	viii
<b>DECLARATION</b>	x
<b>LIST OF TABLES</b>	xi
<b>LIST OF FIGURES</b>	xii
<b>LIST OF ABBREVIATIONS</b>	xiv
<b>CHAPTER</b>	
1	<b>INTRODUCTION</b> 1
2	<b>LITERATURE REVIEW</b> 4
2.1	Black Pepper 4
2.2	Plant Parasitic Nematodes 4
2.2.1	Introducing Plant Parasitic Nematodes 4
2.2.2	Incidence of Root-Knot Nematode Infestation on Black Pepper 5
2.2.3	Life Cycle of Root-Knot Nematode 7
2.3	Nematode Control 8
2.3.1	Resistance Variety 8
2.3.2	Chemical Control 10
2.3.3	Crop Rotation 11
2.3.4	Organic Amendments 13
2.3.5	Biological control 15
2.3.5.1	Nematophagous Fungi 16
2.3.5.1.1	<i>Paecilomyces lilacinus</i> 17
2.3.5.2	Nematode Antagonistic Bacteria 19
2.3.5.2.1	<i>Bacillus thuringiensis</i> 22
	Nematicidal effect of <i>Bacillus thuringiensis</i> 25
3	<b>MATERIALS AND METHODS</b>
3.1	Establishment of Pure <i>Meloidogyne incognita</i> Culture and Perineal Pattern Characterization of <i>Meloidogyne incognita</i> 27
3.1.1	Establishment of Pure <i>Meloidogyne incognita</i> Culture 27
3.1.2	Perineal Pattern Characterization 28
3.2	Isolation, Morphology and Molecular Identification of <i>Paecilomyces lilacinus</i> 28
3.2.1	Isolation and Morphological Identification of <i>Paecilomyces lilacinus</i> 28
3.2.2	Primers for Molecular Identification of <i>Paecilomyces lilacinus</i> 29
3.2.3	DNA Extraction of <i>Paecilomyces lilacinus</i> 30

3.2.4	Polymerase Chain Reaction	31
3.2.5	Agarose Gel Electrophoresis	31
3.2.6	DNA Purification	32
3.2.7	Sequence Analysis	32
3.3	Growth Performance, Sporulation and Formulation of <i>Paecilomyces lilacinus</i>	33
3.3.1	Growth Performance and Sporulation of <i>Paecilomyces lilacinus</i>	33
3.3.2	Mass Production/Formulation of <i>Paecilomyces lilacinus</i>	34
3.4	Female Nematode Bioassay	34
3.5	<i>Paecilomyces lilacinus</i> Colonization Rate on Female and Egg Mass	35
3.6	<i>Paecilomyces lilacinus</i> Parasitism on Eggs	35
3.7	Effects of Spore Suspension of <i>Paecilomyces lilacinus</i> on Egg Hatch and J2	36
3.8	Isolation of <i>Bacillus thuringiensis</i> , Parasporal Crystal Protein Staining and Detection of Cry Gene with Molecular Method	37
3.8.1	Isolation of <i>Bacillus thuringiensis</i>	37
3.8.2	Parasporal Crystal Protein Staining	37
3.8.3	Detection of Cry Gene with Molecular Method	38
3.9	<i>Bacillus thuringiensis</i> 's Parasporal Crystal Toxicity on 2 <sup>nd</sup> Stage Juvenile	38
3.9.1	Production and Harvesting of Parasporal Crystals	38
3.9.2	Bioassay on the Toxicity of Parasporal Crystal on 2 <sup>nd</sup> Stage Juvenile	39
3.10	Biocontrol efficacy of <i>Paecilomyces lilacinus</i> and <i>Bacillus thuringiensis</i> against root-knot nematode, <i>Meloidogyne incognita</i> on black pepper ( <i>Piper nigrum</i> ) in Pots	40
a.	Rooting of Black Pepper Cuttings in Nursery and Nematode Inoculation	40
b.	Treatments with Microbial Control Agents	40
3.10.1	Effects of MCAs Treatments on Growth Parameters and Root Gall Index of Black Pepper	43
3.10.2	Nematode Enumeration in the Root of Black Pepper	44
a.	Staining of Root for Nematode Enumeration	44
b.	Nematode Enumeration after Root Staining	45
3.11	Percentage of Collected Egg Masses and Female Nematode Infected by <i>Paecilomyces lilacinus</i>	45
3.12	Enumeration of <i>Paecilomyces lilacinus</i> and <i>Bacillus thuringiensis</i> in the Rhizosphere	46
3.12.1	Enumeration of <i>Paecilomyces lilacinus</i>	46
3.12.2	Enumeration of <i>Bacillus thuringiensis</i>	47
3.13	Endophytic Colonisation by <i>Paecilomyces lilacinus</i>	48
3.14	Endophytic Colonisation by <i>Bacillus thuringiensis</i>	48
3.15	Growth Diameter of <i>Paecilomyces lilacinus</i> A at 36°C	49

3.16	<i>In Vitro</i> Antagonism between <i>Paecilomyces lilacinus</i> and <i>Bacillus thuringiensis</i>	49
3.16.1	Dual Inoculation	49
3.16.2	Pairing	50
3.17	The Effect of Temperature on the Shelf life of Formulated <i>Paecilomyces lilacinus</i> A	50
3.18	Statistics Analysis	51

#### 4

### RESULTS AND DISCUSSION

4.1	Establishment of Pure <i>Meloidogyne incognita</i> Culture Perineal Pattern Characterization of <i>Meloidogyne Incognita</i>	53
4.2	Isolation, Morphology and Molecular Identification of PL	54
4.3	Growth Performance, Sporulation and Formulation of PL	56
4.4	Female Nematode Bioassay	61
4.5	PL Colonization Rate on Female and Egg Mass	64
4.6	PL Parasitism on Eggs	66
4.7	Effect of PL Spore Suspension on Egg Hatch and J2 mortality	70
4.7.1	Effect of PL Spore Suspension on Egg Hatch	70
4.7.2	Effect of PL Spore Suspension on J2 mortality	71
4.8.	Isolation of BT, Crystal Protein Staining and Molecular Detection of Cry Gene	72
4.9	<i>Bacillus thuringiensis</i> 's Parasporal Crystal Toxicity on 2 <sup>nd</sup> Stage Juvenile	75
4.10	Biocontrol efficacy of <i>Paecilomyces lilacinus</i> and <i>Bacillus thuringiensis</i> against <i>Meloidogyne incognita</i> on black pepper ( <i>Piper nigrum</i> ) in Pots	79
4.10.1	Effects of MCAs Treatments on Growth Parameters and Root Gall Index of Black Pepper	79
4.10.1.1	Shoot Length	79
4.10.1.2	Root Fresh Weight	80
4.10.1.3	Shoot Fresh Weight	82
4.10.1.4	Root Length	83
4.10.1.5	Relative Leaf Chlorophyll Content	85
4.10.1.6	Root Gall Index, Mean Diameter and Total Root Gall	87
4.10.2	Total Egg+J2, Female Nematode and Egg Mass	92
4.10.2.1	Total Egg+J2	92
4.10.2.2	Total Female	93
4.10.2.3	Total Egg Mass	95
4.10.2.4	Reproduction Factor of Nematode	97
4.10.3	Efficacy of PL in Nematode Suppression	98
4.10.4	Growth Promoting Effect of PLA	100
4.10.5	Efficacy of BT in Nematode Suppression	102
4.10.6	Growth Promoting Effect of BT	103
4.10.7	Concomitant treatment versus single treatment	105

4.11	Percentage of Collected Egg masses and Female Infected by PL	108
4.12	Enumeration of <i>Paecilomyces lilacinus</i> and <i>Bacillus thuringiensis</i> in the Rhizosphere	110
4.12.1	Enumeration of PL in the Rhizosphere	110
4.12.2	Enumeration of Spore, Vegetative Cell and Total BT in the Rhizosphere	115
4.13	Endophytic Colonization by PL	118
4.14	Endophytic Colonization by BT	119
4.15	Growth Diameter of PLA at 36°C	120
4.16	<i>In Vitro</i> Antagonism between BT and PLA	122
4.17	The Effect of Temperature on the Shelf life of Formulated <i>Paecilomyces lilacinus</i> A	123
5	<b>SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH</b>	127
	<b>REFERENCES</b>	136
	<b>APPENDICES</b>	157
	<b>BIODATA OF STUDENT</b>	166
	<b>LIST OF PUBLICATIONS</b>	167
	<b>AWARDS</b>	168



## DEDICATION

“HE chose the lowly things of this world and the despised things – and the things that are not – to nullify the things that are, so that no one may boast before HIM.”

1 Corinthians 1: 28-29.

I would like to dedicate this thesis to my late mother and late father for the love with which they brought me up. To my late mother I owe my deepest gratitude for her invaluable advice and many life’s lessons which I utilize to my advantage. To my late father, I would like to thank him for the love and freedom he had given me to pursue my full time study. It is sad that he couldn’t wait long enough to see the completion of my thesis and graduation. I miss them all and believe they will share the joy of my accomplishment from above. I also wish to dedicate this thesis to all my siblings, especially to my sisters, and thank them for their encouragement and moral support.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

**BIOLOGICAL CONTROL OF ROOT KNOT NEMATODES  
(*Meloidogyne incognita*) IN  
BLACK PEPPER (*Piper nigrum* L.) USING  
*Paecilomyces lilacinus* and *Bacillus thuringiensis***

By

**PAU CHEN GUAN**

**August 2012**

**Chairman: Stephen Leong Chan Teck, PhD**

**Faculty: Faculty of Agriculture and Food Sciences (Bintulu)**

Black pepper (*Piper nigrum* L.) is an important cash crop of Sarawak. However the plantations are facing challenges due to widespread infestation of root-knot nematode. Development of biological control methods to address this problem is important for sustainable farming as well as consumer health. *Paecilomyces lilacinus* (PL), a saprophytic soil fungus has drawn many research attentions owing to its promising effect in parasitizing and controlling population of phytonematodes. *Bacillus thuringiensis* (BT), a spore forming bacterium with well known insecticidal property also has been reported in demonstrating toxicity towards root-knot nematodes. Ten indigenous strains of PL and a strain of BT carrying Cry6 and Cry14 gene sequences were isolated as an initiative to combat root-knot nematode (RKN) problem. In female nematode bioassay on water agar, PL demonstrated high significant colonization (>90%, P<0.01) on female. In egg parasitism test, spore suspension ( $10^5$  spore/ml) of PLA exhibited 78.8% parasitism on eggs. Meanwhile,

hatching rates of nematode eggs incubated in spore suspension of PLA for seven days were significantly reduced; 89% of eggs were hatch-inhibited as compared to control (26%). Pot trials were conducted to evaluate the efficacy of PL and BT in managing RKN infestation on black pepper cuttings in single (PL alone, BT alone) and concomitant treatment (BT + PL) under opened house condition. All treatments manifested significant reduction in root gall index as compared to control in the 70 days treatment. In 140 days treatment, fenamiphos and PL were recorded to produce the lowest number of gall per root system (14.6 and 71.9 galls/root system respectively). The percentages of reduction in nematode reproduction factor ( $R_f$ ) for these two treatments were at the greatest, 99.8% and 99.2% respectively. BT and concomitant treatments exhibited no significant difference in term of the number of gall per gram of root as compared to control in both 140 days and 180 days of treatments. In experiment II and III, PLA alone appeared to demonstrate better suppression of RKN per root system than in dual combination treatment (BT+PLA). For 180 days treatments, RKN reproduction factor for fenamiphos, PLA, BT and BT+PLA treatments were 0, 47, 113 and 108 respectively. The above investigation provides opportunity to further evaluate PLA's efficacy in field trial with integrated management.

Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Master Sains

**KAWALAN BIOLOGI TERHADAP NEMATOD BENGKAK AKAR  
(*Meloidogyne incognita*) PADA  
LADA HITAM (*Piper nigrum* L.) DENGAN MENGGUNAKAN  
*Paecilomyces lilacinus* DAN *Bacillus thuringiensis***

Oleh

**PAU CHEN GUAN**

**Ogos 2012**

**Pengerusi: Stephen Leong Chan Teck, PhD**

**Fakulti: Fakulti Sains Pertanian dan Makanan (Bintulu)**

Lada hitam (*Piper nigrum* L.) merupakan tanaman penting yang mendatangkan sumber pendapatan kepada Sarawak. Namun, tanaman ini menghadapi cabaran disebabkan jangkitan penyakit nematod-bengkak-akar yang merebak dengan meluas. Perkembangan kaedah kawalan biologi terhadap masalah ini adalah penting demi keamanan pertanian serta kesihatan pengguna. *Paecilomyces lilacinus* (PL), sejenis kulat saprofitik yang berasal dari tanah telah menarik banyak perhatian dalam penyelidikan kerana kesan positif dalam menjangkiti dan mengawal populasi fitonematod. *Bacillus thuringiensis* (BT), sejenis bakteria pembentuk spora yang terkenal dengan nilai keracunan terhadap serangga, juga telah dilaporkan bahawa ia menunjukkan ketoksikan terhadap nematod-bengkak akar. Sepuluh PL baka tempatan dan sejenis baka BT pembawa urutan gen Cry6 dan Cry14 telah dipencilkan sebagai langkah inisiatif untuk memerangi masalah penyakit ulat-bengkak akar. Dalam bioassai pada nematod betina di atas agar-agar berair, PL

menunjukkan pengkolonian yang bererti dan tinggi ( $> 90\%$ ,  $P < 0.01$ ) pada nematod betina. Dalam ujian parasitisme telur, ampaian spora ( $10^5$  spora /ml) PLA mempamerkan 78.8% pamparasitan pada telur. Sementara itu, kadar penetasan telur nematod yang dieram dalam ampaian spora PLA selama tujuh hari telah mengalami pengurangan yang ketara; 89% telur telah terhalang daripada menetas berbanding dengan ujian kawalan (26%). Ujian berpasu telah dijalankan untuk menilai keberkesanan PL dan BT dalam mengurangkan infestasi RKN pada keratan lada hitam di bawah pondok terbuka melalui rawatan tunggal (PL sahaja, BT sahaja) dan rawatan seiring (BT + PL). Semua jenis rawatan menunjukkan penurunan indeks bengkak akar yang bererti berbanding dengan ujian kawalan dalam rawatan selama 70 hari. Dalam rawatan selama 140 hari, fenamifos dan PL mencatat bilangan bengkak akar per sistem akar yang terendah (masing-masing 14.6 dan 71.9). Peratus peunuranan dalam faktor pembiakan nematod untuk kedua-dua rawatan ini adalah yang tertinggi masing-masing pada 99.8% dan 99.2%. BT dan rawatan seiring (PL+BT) menunjukkan perbezaan yang tidak bererti dalam aspek purata bengkak per gram akar berbanding dengan ujian kawalan bagi kedua-dua jenis rawatan selama 140 hari dan 180. Dalam eksperimen I dan II, PLA menunjukkan penindasan RKN per sistem akar yang lebih baik daripada rawatan seiringan (BT + PLA). Bagi rawatan selama 180 hari, faktor pembiakan RKN untuk rawatan fenamifos, PLA, BT dan rawatan seiring (BT + PLA) adalah masing-masing 0, 47, 113 dan 108. Penyelidikan di atas telah membuka peluang untuk menilai keberkesanan PLA dalam percubaan ladang secara pengurusan bersepadu.

## ACKNOWLEDGEMENTS

First of all, I wish to thank my supervisor Dr Stephen Leong for his academic support, patience and help. I also wish to extend my gratitude to my co-supervisors, Dr Wong Sing King and Dr Osumanu Haruna Ahmed for their guidance, advice and insight. Many thank to all members of my supervisory committee for their time, patience and input in reviewing and correcting my manuscript.

I also wish to extend my appreciation to Dr Lily Eng for her help and invaluable advice about some of the laboratory skills on nematology. I am grateful to her for teaching me the technique of preparing perineal pattern, sending me a photocopy of her literature and lending me her Bridge counting disks.

My profound gratitude to Prof Dato Dr Nik Muhammad Majid, black pepper research team leader, for his insight and constructive comment. I would also like to register my thanks for all lecturers and members in the black pepper research team for their input during monthly meeting and progress presentation. Gratitude is also expressed to the Dean and Deputy Dean of Faculty of Agriculture and Food Science, Prof. Dr. Japar Sidik Bin Bujang and Dr. Mohd. Hanafi Bin Idris, and all UPM lecturers who have contributed constructive idea to my studies. Many thanks for their friendship and moral support.

Also, my deep appreciation to Malaysia Pepper Board for funding this study and to UPM Graduate School for the scholarship that I received and for the financial support during my research and studies in UPM Bintulu. My gratitude to Bahagian

Tajaan of Kementerian Pelajaran Malaysia for granting me with full time unpaid study leave so that I could concentrate on research.

I would also like to thank our lab assistants, Mohammad Ezwandi and Maini Chee for their technical assistance on some of our laboratory equipments. To all my laboratory mates and coursemates, Sharon Wong, Amelia Tang, Hsien Loong, Chin Tsong, Latifah, Kona, Latip, thank you for your friendship, encouragement and assistance. Thank you also to Dr Yiu Pang Hong for allowing me to use your homogenizer in your chemistry laboratory. My deepest appreciation extended to English lecturer in IPG, Mr Tay Nguong Yong who carefully corrected the grammatical errors in my thesis.

Also, my heartfelt respect and gratitude to all my examiners, Prof. Dr Sepiah Muid, Assoc. Prof. Dr. Radziah Othman, Assoc. Prof. Dr. Kamaruzaman Sijam and chairperson of viva voce, Asso. Prof. Dr. Joseph Bong Choon Fah for their reviews, constructive comments and suggestions in improving the content of my thesis.

Lastly, thanks to all my siblings and relatives for their moral support and help during this course of study. No part of this research was conducted as a sole endeavor and you all have contributed at different levels in different ways. It is a pleasure for me to thank those who made this master thesis possible. Lastly, my best regards and blessings to all who have directly or indirectly supported me in any aspect during the completion of this thesis.

I certify that a Thesis Examination Committee has met on 13 August 2012 to conduct the final examination of PAU CHEN GUAN on his thesis entitled “Biological control of root-knot nematodes (*Meloidogyne incognita*) in black pepper (*Piper nigrum* L.) using *Paecilomyces lilacinus* and *Bacillus thuringiensis*” in accordance with Universities and University Colleges Act 1971 and the Constitution of the University Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Joseph Bong Choon Fah, PhD**

Associate Professor  
Faculty of Agriculture and Food Science  
Universiti Putra Malaysia  
(Chairman)

**Kamaruzaman bin Sijam, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Radziah binti Othman, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Sepiah Muid, PhD**

Professor  
Faculty of Resource Science and Technology  
Universiti Malaysia Sarawak  
(External Examiner)

---

**SEOW HENG FONG, PhD**

Professor and Deputy Dean  
School of Graduate Studie  
Universiti Putra Malaysia

Date: 26 February 2013



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Stephen Leong Chan Teck, PhD**

Senior Lecturer  
Faculty of Agriculture and Food Science  
Universiti Putra Malaysia  
(Chairman)

**Wong Sing King, PhD**

Senior Lecturer  
Faculty of Agriculture and Food Science  
Universiti Putra Malaysia  
(Member)

**Osumanu Haruna Ahmed, PhD**

Associate Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Member)

---

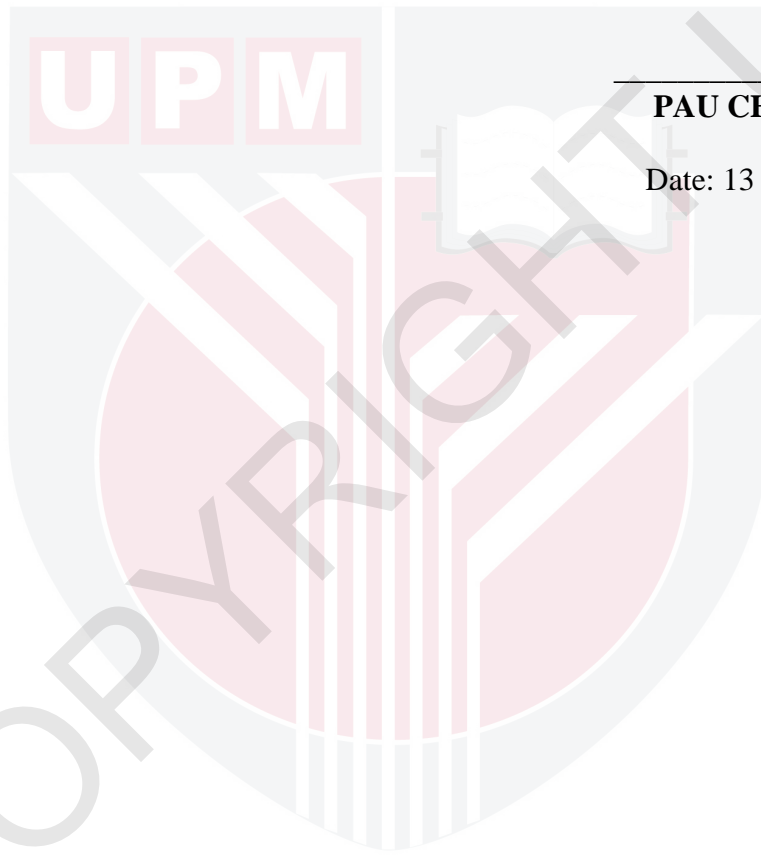
**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



---

**PAU CHEN GUAN**

Date: 13 August 2012

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
1.1	Mean diameter growth of PLM and PLA on PDA, PDA + 6% molasses and 6% molasses.	57
1.2	Comparison between spore production of PLM and PLA on PDA, PDA + 6% Molasses, 6% Molasses Agar after 20 days of incubation.	58
1.3	Mass Production of PLA, PLB and PLM in rice husk + 10% molasses + moisture content 25.97%.	58
2.1	Effect of PL spore suspension ( $10^5$ spore/ml) on egg parasitism, egg hatch inhibition and J2 mortality.	71
2.2	Percentage of paralyzed J2 of MI after 24, 48 and 77 hours exposed to extract of spore-crystal mixtures of BT.	76
3.1.0	Shoot Length of black pepper in Experiment I, II and III	80
3.1.1	Root Fresh Weight of black pepper in Experiment I, II and III	81
3.1.2	Shoot Fresh Weight of black pepper in Experiment I, II and III	83
3.1.3	Root Length of black pepper in Experiment I, II and III	84
3.1.4	Relative Leaf Chlorophyll Content of black pepper in Experiment I, II and III	86
3.1.5	Root Gall Index of black pepper in Experiment I, II and III	88
3.1.6	Mean Diameter of Root Gall in Experiment II and III	89
3.1.7	Total Root Gall (per gram of root and per root system) of black pepper in Experiment II and III	91
3.1.8	Total Egg plus Juvenile (per gram of root and per root system) in Experiment II and III	94
3.1.9	Total Female (per gram of root and per root system) in Experiment II and III	95
3.1.10	Total Egg Mass (per gram of root and per root system) in Experiment II and III	97
4.1	CFUs (total count, spores & vegetative cells) of BT in soils under BT and BT+PLA treatments.	116
4.2	Endophytic colonization of BT on root of black pepper	119

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
4.1 a-u	Perineal Pattern of female nematodes.	53
4.2 a	Growth morphology of PL colonies on PDA agar plate.	55
4.2 b	Hypha of <i>Paecilomyces lilacinus</i> bear phialides (P) with huge numbers of conidia (C) attached loosely in long divergent chains.	55
4.2 c	Detection of PCR amplification of ITS genes from <i>Paecilomyces lilacinus</i>	56
4.3	Growth of PLM and PLA on three different media: Molasses (6%) agar, PDA and PDA+6% molasses	57
4.4 a	Percentage of female nematodes colonized by <i>Paecilomyces lilacinus</i>	63
4.4 b-c	Colonization on female nematode by PLM & PLA	63
4.4 d-e	Colonization on female nematode by PLA.	63
4.5 a	Colonization rate of <i>Paecilomyces lilacinus</i> on female nematodes under <i>in vitro</i> condition.	65
4.5 b	Colonization rate of <i>Paecilomyces lilacinus</i> on egg masses under <i>in vitro</i> condition	65
4.6 a-f	Parasitism of <i>Paecilomyces lilacinus</i> on eggs	69
4.7 a	Sporulation and crystal producing colonies of BT	73
4.7 b	Photomicrograph of BT culture stained with Coomassie brilliant blue	73
4.7 c	Detection of PCR amplification of Cry6 and Cry14 genes of three BT isolates	74
4.8	Effect of treatments on <i>Meloidogyne incognita</i> infested roots of black pepper	92
4.9	Fushsin acid staining of Root, Eggs and Female Nematode	94
4.10	Percentage of female and egg mass infected by <i>Paecilomyces lilacinus</i> .	109
4.11	A dissected big root gall of inoculated control	110

4.12	Total CFUs of PL in PLA alone, PLB alone and PLA+BT treated soil	111
4.13	<i>In vitro</i> growth profiles (mean $\pm$ SE) of PL on PDA+ plate incubated at 28°C(Ж) and 36°C	121
4.14	Antagonism between PL and BT on PDA	122
4.15	The influence of storage temperature on the shelf-life of formulated PL.	125



## LIST OF ABBREVIATIONS

1	BT	<i>Bacillus thuringiensis</i>
2	CFUs	Colony Forming Units
3	CRD	Complete Randomized Design
4	Fig	Figure
5	J2	2 <sup>nd</sup> stage Juveniles
6	MCAs	Microbial Control Agents
7	PCR	Polymerase Chain Reaction
8	MI	<i>Meloidogyne incognita</i>
9	PDA	Potato Dextrose Agar
10	PDA+	Potato Dextrose Agar amended with 0.01% (w/v) Chloramphenicol and 3% (w/v) Sodium Chloride
11	PDB	Potato Dextrose Broth
12	PGPR	Plant Growth Promoting Rhizobacteria
13	PL	<i>Paecilomyces lilacinus</i>
14	RCBD	Randomized Complete Block Design
15	R <sub>f</sub>	Reproduction factor
16	RGI	Root Gall Index
17	RKN	Root Knot Nematode
18	RLCC	Relative Leaf Chlorophyll Content
19	SDS	Sodium Dodecylsulphate
20	Tab	Table
21	TAE	Tris Acetic acid EDTA
22	TE	Tris EDTA
23	VC	Vegetative Cell

## CHAPTER 1

### INTRODUCTION

Black pepper (*Piper nigrum L.*), the king of spices, is an important cash crops of Sarawak. Sarawak exported about 19,748 tonnes of pepper in 2004 and 22,218 tonnes in 2008, valued at RM 113.2 million and RM170.0 million respectively (Lim, 2009). About 6,125 tonnes of black pepper were exported from January to June 2010, valued at RM77.889 million (Anonymous, 2010). Today, pepper is one of the important cash crops supporting the livelihood of about 67,000 rural dwellers in upland areas of Sarawak (Lim, 2009).

However, most of the black pepper farms in Sarawak are being infested by root-knot nematodes, *Meloidogyne* spp. (Kueh, 1978,; Leong, 1986; Ramana and Eapen, 2000; Eng, 2001) which has always been a major constraint to the black pepper production. In a survey conducted by Eng (2001) on 43 black pepper farms in Sarawak, root-knot nematodes (RKN) were reported presence in all the farms. Badly infested vines, with yellowish speckle especially at the interveinal areas of the leaves and galls on their roots, are stunted and vulnerable to moisture or temperature pressure as well as other pathogenic infections such as *Phytophthora* (Winoto, 1972). Subsequently, the vines became unproductive and abandoned, resulting in substantial economic losses to farmers each year.

Currently, no RKN resistant black pepper cultivar is available (Eng, 2001). For the past decades, chemical nematicides have been used in managing nematodes infestation on crops but gradually are being reappraised in respect of health and environmental concern and limited availability in developing nations. On the other hand, microbial control agents (MCAs) are gaining popularity in integrated nematodes management due to their promising results in nematodes control and are safer than synthetic nematicides (Mukhtar & Pervaz, 2003; Dong & Zhang, 2006).

*Paecilomyces lilacinus* (PL), a saprophytic soil fungus, commonly known as natural facultative egg parasite of root-knot and cyst nematodes (Rumbos & Kiewnick, 2006) has drawn many research attentions for the past decade owing to its efficacy in parasitizing and controlling population of phytonematodes (Jatala, 1986; Dube & Smart, 1987; Hewlett *et al.*, 1988; Freitas *et al.*, 1995; Nagesh *et al.*, 1997; Khan *et al.*, 2006a; Kiewnick & Sikora, 2006a; Kiewnick & Sikora, 2006b; Brand *et al.*, 2010). It was reported with high frequency of occurrence in tropics and subtropic (Morgan *et al.*, 1984; Chen *et al.*, 1996) and can be found in most of agricultural soils (Brand *et al.*, 2010). According to the report of Eng (2001), 82.9% of the total forty one surveyed farms in Sarawak contained PL despite intensive application of fungicide in the farm. Cabanillas *et al.* (1989) observed maximum growth of PL at temperature ranged from 24°C–30°C. They claimed that PL was able to grow on a wide range of common organic substrates and remain competitive with other microbes in soil. Besides, it also tolerated broad range of soil pH and was able to grow well at 15°C–30°C. Since PL has high adaptability in its life strategy, it is competitive in a broad spectrum of range adaptability.



Meanwhile, *Bacillus thuringiensis* (BT), a well-known entomopathogenic bacteria used for pest insects control since four decades ago (Brar *et al.*, 2006) has also drawn intensive studies on its nematicidal effects against economically important phyto-parasitic nematodes (Deviddas and Rehberger, 1992; Siddiqui and Mahmood, 1994; Carneiro *et al.*, 1998; Mozgovaya *et al.*, 2002; El-Nagdi and Youssef, 2004; El-Sherif *et al.*, 2007; Mohammed *et al.*, 2008; Khan *et al.*, 2010). Carneiro *et al.* (1998) claimed that culture of BT was efficacious in killing freshly hatched 2<sup>nd</sup> stage juvenile (J2) of *Meloidogyne javanica*. Mozgovaya *et al.* (2002) reported 80% mortality of nematodes after *in vitro* treatment with BT. El-Nagdi and Youssef (2004) commented that soaking faba beans with BT reduced the population density of *M. incognita* but increased the plant growth. According to El-Sherif *et al.* (2007), BT applied alone improved the growth parameters of egg plant and reduced nematode development. Mohammed *et al.* (2008) reported that the spore/crytal proteins of BT showed highest nematicidal activities against *Meloidogyne incognita*. However, no current report was found on the positive effect of BT in controlling RKN development in black pepper. Therefore, an effort is initiated to explore the use of BT and PL as alternatives to the chemical nematicides currently in use for the control of root-knot nematodes in black pepper. The objectives of this study were:

1. To isolate and evaluate the local isolates of *Paecilomyces lilacinus* and *Bacillus thuringiensis* for their nematicidal properties against *Meloidogyne incognita* in the laboratory bioassay.
2. To produce, formulate and evaluate the selected strains of *Paecilomyces lilacinus* and *Bacillus thuringiensis* for their biological control of *Meloidogyne incognita* on black pepper in pot trial.

## REFERENCES

- Ahmad, S.F. and Khan, T.A. 2004. Management of root-knot nematode, *Meloidogyne incognita*, by integration of *Paecilomyces lilacinus* with organic materials in chilli. *Archives of Phytopathology and Plant Protection* 37: 35–40.
- Ahrens D. and Tunlid, A. 2003. Evolution of parasitism in nematode-trapping fungi. *Journal of Nematology* 35: 194–197.
- Akhtar, M. 2000. Nematicidal potential of the neem tree *Azadirachta indica* (A. Juss). *Integrated Pest Management Reviews* 5: 57–66.
- Akhtar, M. and Malik, A. 2000. Role of organic soil amendments and soil organisms in the biological control of plant-parasitic nematodes: a review. *Bioresource Technology* 74: 35-47.
- Al Kader, M.A.A. 2008. *In Vitro Studies on Nematode Interactions with their Antagonistic Fungi in the Rhizosphere of Various Plants*, PhD Thesis, Albert-Ludwigs Universität, Freiburg im Breisgau, Germany.
- Amir, H. and Alabouvette, C. 1993. Involvement of soil abiotic factors in the mechanisms of soil suppressiveness to *Fusarium* wilts. *Soil Biology and Biochemistry* 25: 157-164.
- Anastasiadis, I.A., Giannakou, I.O., Prophetou-Athanasiadou, D.A. and Gowen, S.R. 2008. The combined effect of the application of a biocontrol agent *Paecilomyces lilacinus*, with various practices for the control of root-knot nematodes. *Crop Protection* 27: 352-361
- Anonymous. 2010. Market Review January-June 2010. In: *Malaysia Pepper Industry Bullentin Jan-Jun 2010*, ed. T.H.S. Rapae, pp. 10-16. Kuching, Sarawak: Malaysian Pepper Board.
- Anver, S., Khan, A.A. and Alam, M.M. 2001. Integrated management of root-knot and reniform nematodes with neem cake and biocontrol fungus *Paecilomyces lilacinus* on chickpea and pigeonpea. *Archiv für Phytopathologie und Pflanzenschutz* 34:255-264
- Asano, S.-I., Yamashita, C., Iuzika, T., Takeuchi, K., Yamanaka, S., Cerf, D. and Yamamoto. T. 2003. A strain *Bacillus thuringiensis* subsp. *galleriae* containing a Novel *cry8* Gene Highly Toxic to *Anomala cuprea* (Coleoptera: Scarabaeidae). *Biological Control* 28: 191-196.
- Ashraf, M.S. and Khan, T.A. 2005. Effect of opportunistic fungi on the life cycle of the root-knot nematode (*Meloidogyne javanica*) on brinjal. *Archives of Phytopathology and Plant Protection* 38: 227-233.
- Atkins, S. D., Clark, I.M., Pande, S., Hirsch, P.R. and Kerry, B.R. 2005. The use of real-time PCR and species-specific primers for the identification and monitoring of *Paecilomyces lilacinus*. *FEMS Microbiology and Ecology* 51: 257–264.

- Bakker, P.A.H.M., Pieterse, C.M.J. and Van Loon, L.C. 2007. Induced systemic resistance by fluorescent *Pseudomonas* spp. *Phytopathology* 97: 239-243.
- Bansal, R.K., Walia, R.K. and Bhatti, D.S. 1988. Evaluation of some agro-industrial wastes for mass propagation of the nematode parasitic fungus, *Paecilomyces lilacinus*. *Nematologia Mediterranea* 16: 135-136.
- Bansal, R.K., Walia, R.K. and Bhatti, D.S. 1992. Wood charcoal powder, a carrier of *Paecilomyces lilacinus* spores. *Nematologia Mediterranea* 20: 5-7.
- Bonants, P.J.M., Fitters, P.F.L., Thijs, H., Belder, E.D., Waalwijk, C. and Henfling, J.W.D.M. 1995. A basic serine protease from *Paecilomyces lilacinus* with biological activity against *Meloidogyne hapla* eggs. *Microbiology* 141: 775-784.
- Bourne, J.M., Kerry, B.R. and De Leij F.A.A.M. 1994. Methods for the study of *Verticillium chlamydosporium* in the rhizosphere. *Journal of Nematology* 26: 587-591.
- Brand, D., Soccol, C.R., Sabu, A. and Roussos, S. 2010. Production of fungal biological control agents through solid state fermentation: A case study on *Paecilomyces lilacinus* against root-knot nematodes. *Micologia Aplicada International* 22: 31-48.
- Brar, S.K., Verma, M., Tyagi, R.D., Val e'ro, J.R. and Surampalli, R.Y. 2006. Efficient centrifugal recovery of *Bacillus thuringiensis* biopesticides from fermented wastewater and wastewater sludge. *Water Research* 40: 1310-1320.
- Bridge, J. and Page, S.L.J., 1980. Estimation of root-knot nematodes infestation levels using a rating chart. *Tropical Pest Management* 26: 296-298.
- Brooks, F.E. 2008. University of Hawaii, Burrowing nematode disease: *The Plant Health Instructor*. [http://www.apsnet.org/edcenter/intropp/lessons/Nematodes/Pages/Burrowing\\_nematode.aspx](http://www.apsnet.org/edcenter/intropp/lessons/Nematodes/Pages/Burrowing_nematode.aspx). Retrieved 14 Jan 2012.
- Butko, P., Huang, F., Pusztai-Carey, M. and Surewicz, K.W. 1997. Membrane permeabilization induced by cytolytic delta-endotoxin CytA from *Bacillus thuringiensis* var. *israelensis* with lipid membranes. *Biochemistry* 35: 11355-11360.
- Cabanillas, E., and Barker, K. R. 1989. Impact of *Paecilomyces lilacinus* inoculum level and application time on control of *Meloidogyne incognita* on tomato. *Journal of Nematology* 21:115-120.
- Cabanillas E., Barker, K.R. and Daykin, M.E. 1988. Histology of the interactions of *Paecilomyces lilacinus* with *Meloidogyne incognita* on tomato. *Journal of Nematology* 20: 362-365.

- Cabanillas, E., Barker, K.R. and Nelson, L.A. 1989. Growth of isolates of *Paecilomyces lilacinus* and their efficacy in biocontrol of *Meloidogyne incognita* on tomato. *Journal of Nematology* 21:164-172
- Cáceres, T., Megharaj, M., Venkateswarlu, K., Sethunathan, N. and Naidu, R. 2010. Fenamiphos and related organophosphorus pesticides: environmental fate and toxicology. *Reviews of Environmental Contamination and Toxicology* 205: 117-162.
- Cannayane, I. and Sivakumar, C.V. 2001. Nematode egg-parasitic fungus 1: *Paecilomyces lilacinus*-A review. *Agricultural Reviews* 22: 79-86.
- Carey, J., D'Amico, R., Sutton, D A. and Rinaldi, M. G. 2003. *Paecilomyces lilacinus* vaginitis in an immunocompetent patient. *Emerging Infectious Diseases* 9: 1155-1158.
- Carneiro, R.M.D.G., Souza, I.S.D. and Belarmino, L.C. 1998. Nematicidal activity of *Bacillus* spp. strains on juveniles of *Meloidogyne javanica*. *Nematologia Brasileira* 22: 12-19.
- Chen, Z.X., Dickson, D.W., McSorley, R., Mitchell, D.J. and Hewlett, T.E. 1996. Suppression of *Meloidogyne arenaria* race 1 by soil application of endospores of *Pasteuria penetrans*. *Journal of Nematology* 28: 159-168.
- Chen, S.Y., Dickson, D. W. and Mitchell, D. J. 2000. Viability of *Heterodera glycines* exposed to fungal filtrates. *Journal of Nematology* 32: 190-197.
- Chen, Z.X., Chen, S.Y. and Dickson, D. W. 2004. *Nematology: Advances and Perspectives. Vol. 2: Nematode Management and Utilization*. Wallingford, UK: CAB International.
- Chen, Z. X. and Dickson, D. W. 1998. Review of *Pasteuria penetrans*: biology, ecology, and biological control potential. *Journal of Nematology* 30: 313-340.
- Chen, Z. X., Dickson, D. W., McSorley, R., Mitchell, D. J. and Hewlett, T. E. 1996. Suppression of *Meloidogyne arenaria* race 1 by soil application of endospores of *Pasteuria penetrans*. *Journal of Nematology* 28: 159-168.
- Chitwood, D.J., 2002. Phytochemical based strategies for nematode control. *Annual Review of Phytopathology* 40: 221-249.
- Connick, W.J., Daigle, D.J., Boyette, C.D., and Williams, K.S. 1996. Water activity and other factors that affect the viability of *Colletotrichum truncatum* conidia in wheat flourkaolin granules ('Pesta')'. *Biocontrol Science and Technology* 6: 277-284.
- Cook, R. and Starr, J.L. 2006. Resistant Cultivars. In *Plant Nematology*, ed. R.N. Perry, and M. Moens, pp. 370-391. Wallingford, UK: CABI.

- Costa, M.J.N., Campos, V.P., Pfenning, L.H. and Oliveira, D.F. 2001. Toxicity of filtered fungals to *Meloidogyne incognita*. *Fitopatologia Brasileira* 26: 749-755.
- Crickmore, N., Zeigler, D.R., Feitelson, J., Schnepf, E., Van-Rie, J., Lereclus, D., Baum, J. and Dean, D.H. 1998. Revision of nomenclature for the *Bacillus thuringiensis* pesticidal crystal proteins. *Microbiology and Molecular Biology Reviews* 62: 807-813.
- Crickmore, N. 2005. Using worms to better understand how *Bacillus thuringiensis* kills insects. *Trends in Microbiology* 13: 347-350.
- Crickmore, N., Zeigler, D.R., Schnepf, E., Van Rie, J., Lereclus, D., Baum, J, Bravo, A. and Dean, D.H. 2012. *Bacillus thuringiensis* toxin nomenclature. [http://www.lifesci.sussex.ac.uk/Home/Neil\\_Crickmore/Bt/](http://www.lifesci.sussex.ac.uk/Home/Neil_Crickmore/Bt/)
- Datta, S.C. 2006. Effects of *Cina* on root-knot disease of mulberry. *Homeopathy* 95: 98-102.
- Davide, R. G. 1988. Nematode problems affecting agriculture in the Philippines. *Journal of Nematology* 20: 214-218.
- Dawar, S. and A. Ghaffar. 2003. Effect of inorganic fertilizers on the efficiency of *Paecilomyces lilacinus* in the control of soil borne root infecting fungi on mung bean. *Pakistan Journal of Botany* 35: 479-482.
- Delfosse, E. S. 2005. Risk and ethics in biological control. *Biological Control* 35: 319-329.
- Devidas, P. and Rehberger, L.A. 1992. The effects of exotoxin (Thuringiensin) from *Bacillus thuringiensis* on *Meloidogyne incognita* and *Caenorhabditis elegans*. *Plant and Soil* 145: 115-120.
- Diedhiou, P.M., Hallmann, J., Oerke, E.C., Dehne, H.W. 2003. Effects of arbuscular mycorrhizal fungi and a non-pathogenic *Fusarium oxysporum* on *Meloidogyne incognita* infestation of tomato. *Mycorrhiza* 13: 199-204.
- Djian, C., Pijarowski, L., Ponchet, M., Arpin, N. and Favre-Bonvinm, J. 1991. Acetic acid: a selective nematicidal metabolite from culture filtrates of *Paecilomyces lilacinus* (Thom) Samson and *Trichoderma longibrachiatum* Rifai. *Nematologica* 37: 101-112.
- Domenech, J., Reddy, M.S., Kloepper, J.W., Ramos, B. and Gutierrez-Mañero. 2006. Combined application of the biological product LS213 with *Bacillus*, *Pseudomonas* or *Chryseobacterium* for growth promotion and biological control of soil-borne diseases in pepper and tomato. *BioControl* 51: 245-258.
- Dong, L. Q. and K. Q. Zhang. 2006. Microbial control of plant-parasitic nematodes: a five-party Interaction. *Plant and Soil* 288: 31-45.

- Du, J., Knowles, B.H., Li, J. and Ellar, D.J. 1999. Biochemical characterization of *Bacillus thuringiensis* cytolytic toxins in association with phospholipids bilayer. *Biochemical Journal* 338: 185-193.
- Duan, W., Yang, E., Xiang, M. and Liu, X. 2008. Effect of storage conditions on the survival of two potential biocontrol agents of nematodes, the fungi *Paecilomyces lilacinus* and *Pochonia chlamydosporia*. *Biocontrol Science and Technology* 18: 613-620.
- Dube, B. and Smart, G.C.Jr2. 1987. Biological control of *Meloidogyne incognita* by *Paecilomyces lilacinus* and *Pasteuria penetrans*. *Journal of Nematology* 19: 222-227.
- Eapen, S.J., Beena, B. and Ramana, K.V. 2005. Tropical soil microflora of spice-based cropping systems as potential antagonists of root-knot nematodes. *Journal of Invertebrate Pathology* 88: 218-225.
- Eisenback, J.D. and Triantaphyllou, H.H. 1991. Root-knot nematodes: *Meloidogyne* species and races. In *Manuel of Agricultural Nematology*, ed. W.R. Nickle, pp. 191-274. New York: Marcel Dekker, Inc.
- El-Nagdi, W.M.A. and Youssef, M.M.A. 2004. Soaking faba bean seed in some bio-agents as prophylactic treatment for controlling *Meloidogyne incognita* root-knot nematode infection. *Journal of Pest Science* 77: 75-78.
- El-Sherif, A.G., Refaei, A.R., El-Nagar, M.E. and Salem, H.M.M. 2007. Integrated management of *Meloidogyne incognita* infecting egg plant by certain organic amendments, *Bacillus thuringiensis* and Oxamyl with reference to N P K and total chlorophyll status. *Plant Pathology Journal*. 6: 147-152.
- Elbadri, G.A.A., Lee, D.W., Park, J.C. and Ho., Y.C. 2009. Nematicidal efficacy of herbal powders on *Meloidogyne incognita* (Tylenchida: Meloidogynidae) on potted watermelon. *Journal of Asia-Pacific Entomology* 12: 37-39.
- Elbadri, G.A., Lee, D.W., Park, J.C., Yu, H.B. and Choo, H.Y. 2008. Evaluation of various plant extracts for their nematicidal efficacies against juveniles of *Meloidogyne incognita*. *Journal of Asia-Pacific Entomology* 11: 99-102.
- Elzein, A.E.M. 2003. Development of a granular mycoherbicidal formulation of *Fusarium oxysporum* "Foxy 2" for the biological control of *Striga hermonthica*. In *Tropical Agriculture 12– Advances in Crop Research (2)*, ed. J. Kroschel, pp 190. Weikersheim, Germany: Margraf Verlag.
- Elzein, A., Kroschel, J., and Müller-Sto"ver, D. 2004. Optimization of storage conditions for adequate shelf-life of 'Pesta' formulation of *Fusarium oxysporum* 'Foxy 2', a potential mycoherbicide for *Striga*: Effects of temperature, granule size and water activity. *Biocontrol Science and Technology* 16: 89-98
- Elzein, A., Kroschel, J., and Müller-Sto"ver, D. 2004b. Effects of inoculum type and propagule concentration on shelf life of 'Pesta' formulations containing

*Fusarium oxysporum* Foxy 2, a potential mycoherbicide agent for *Striga* spp. *Biological Control* 30: 203-211.

- Eng, L. 2001. Biological control of root-knot nematodes (*Meloidogyne* species) on black pepper (*Piper nigrum* L.) in Sarawak, *PhD Thesis*, University of Reading, United Kingdom.
- Esfahani, M. N. and Pour, B. A. 2006. The effect of *Paecilomyces lilacinus* on the pathogenesis of *Meloidogyne javanica* and tomato plant growth parameters. *Iran Agricultural Research* 24/25: 67-75.
- Filippi, C., Bagnoli, G., Treggi, G. and Picci, G. 1984. Antagonistic effects of soil bacteria on *Fusarium oxysporum* Schlecht f.sp. *dianthii* (Prill and Del.) Snyd. and Hans. *Plant and Soil* 80: 119-125.
- Freitas, L.G., Ferraz, S. and Muchovej, J.J. 1995. Effectiveness of different isolates of *Paecilomyces lilacinus* and an isolate of *Cylindrocarpon destructans* on the control of *Meloidogyne javanica*. *Nematropica* 25: 109-115.
- Friesen, T.J., Holloway, G., Hill, G.A. and Pugsley, T. S. 2006. Effect of conditions and protectants on the survival of *Penicillium bilaiae* during storage. *Biocontrol Science and Technology* 16: 89-98.
- Fuller, V.L., Lilley, C.J. and Urwin, P.E. 2008. *Tansley review* Nematode resistance. *New Phytologist* 180: 27-44
- Gautam, A., Siddiqui, Z.A., Mahmood, I. 1995. Integrated management of *Meloidogyne incognita* on tomato. *Nematologia Mediterranea* 23: 245-247.
- Giannakou, I.O., Karpouzas, D.G. and Prophetou-Athanasidou, D. 2004. A novel non-chemical nematicide for the control of root-knot nematodes. *Applied Soil Ecology* 26: 69-79.
- Gill, S.S., Cowles, E.A. and Pietrantonio, P.V. 1992. The mode of action of *Bacillus thuringiensis* endotoxins. *Annual Review of Entomology* 37: 615-636.
- Glass, N.L. and Donaldson, G.C. 1995. Development of primers set designed for use with the PCR to amplify conserved genes from filamentous Ascomycetes. *Applied and Environmental Microbiology* 61: 1323-1330.
- Goettel, M.S., Hajek, A.E., Siegel, J.P., Evans, H.C. 2001. Safety of fungal biocontrol agents. In: *Fungi as Biocontrol Agents: Progress, Problems and Potentials*, ed. T.M. Butt, C. Jackson, N. Magan, pp. 347-375. Wallingford, UK: CABI Publishing.
- Gomez, K.A. and Gomez, A.A. 1984. *Statistical Procedures for Agricultural Research*, 2<sup>nd</sup> edition. New York: John Wiley and Sons.

- Gortari, M.C. and Hours, R.A. 2008. A review: Fungal chitinases and their biological role in the antagonism onto nematode eggs. *Mycological Progress* 7: 221–238.
- Goswami, B.K., Pandey, R. K., Rathour, K.S., Bhattacharya, C. and Singh, L. 2006. Integrated application of some compatible biocontrol agents along with mustard oil seed cake and furadan on *Meloidogyne incognita* infecting tomato plants. *Journal of Zhejiang University Science B* 7: 873-875.
- Gray, E.J., Lee, K.D., Souleimanov, A.M., Di Falco, M.R., Zhou, X., Ly, A., Charles, T.C., Driscoll, B.T. and Smith, D.L. 2006. A novel bacteriocin, thuricin 17, produced by plant growth promoting rhizobacteria strain *Bacillus thuringiensis* NEB17: isolation and classification. *Journal of Applied Microbiology* 100: 545-554.
- Gucalp, R., Carlisle, P., Gialanella, P., Mitsudo, S., McKittrick, J., Dutcher, J. 1996. *Paecilomyces* sinusitis in an immunocompromised adult patient: case report and review. *Clinical Infectious Diseases* 23: 391–393.
- Guerchicoff, A., Delecluse, A. and Rubinstein, C.P. 2001. The *Bacillus thuringiensis* cyt genes for haemolytic endotoxins constitute a gene family. *Applied and Environmental Microbiology*. 67: 1090-1096.
- Gul, A.S. and Shah, S.F.A. 1990. Control of root-knot nematodes in tomato through organic amendments and NPK. *Sarhad Journal of Agriculture* 6: 95–97.
- Gul, A.S. and Zulfiqar, M. 1990. Promising control of root-knot nematodes (*Meloidogyne*) of tomato through organic amendments. *Sarhad Journal of Agriculture* 6: 417-420.
- Guo, G., Zhang, L., Zhou, Z., Ma, Q., Liu, J.P., Zhu, C., Zhu L., Yu, Z.N., Sun, M. 2008. A new group of parasporal inclusions encoded by the S-layer gene of *Bacillus thuringiensis*. *FEMS Microbiology Letters* 282: 1-7.
- Gutierrez-Rodero, F., Moragon, M., Ortiz de la Tabla, V., Mayol, M.J., Martin, C. 1999. Cutaneous hyalohyphomycosis caused by *Paecilomyces lilacinus* in an immunocompetent host successfully treated with itraconazole: case report and review. *European Journal of Clinical Microbiology & Infectious Disease* 18: 814–818.
- Hartman, K.M. and Sasser, J.N. 1985. Identification of *Meloidogyne* species on the basis of differential host test and perineal pattern morphology. In *An Advanced Treatise on Meloidogyne, Volume II: Methodology*, ed. K.R. Barker, C.C. Carter and J.N. Sasser, pp.69-77. Raleigh, USA: North Carolina State University.
- Hashem, M., and Abo-Elyousr, K. A. 2011. Management of the root-knot nematode *Meloidogyne incognita* on tomato with combinations of different biocontrol organisms. *Crop Protection* 30: 285-292.



- Hewlett, T.E., Dickson, D.W., Mitchell, D.J. and Kannwischer-Mitchell, M.E. 1988. Evaluation of *Paecilomyces lilacinus* as biocontrol agents of *Meloidogyne javanica* on tomato. *Journal of Nematology* 20: 578-584.
- Holland, R.J., Gunasekera, T.S., Williams, K.L. and Nevalainen, K.M.H. 2002. Ultrastructure and properties of *Paecilomyces lilacinus* spores. *Canadian Journal of Microbiology* 48: 879-885.
- Holland, R.J., Williams, K.L. and Khan, A. 1999. Infection of *Meloidogyne javanica* by *Paecilomyces lilacinus*. *Nematology* 2: 131-139.
- Holland, R.J., Williams, K.L. and Nevalainen, H.K.M. 2003. *Paecilomyces lilacinus* strain Bioact251 is not a plant endophyte. *Australasian Plant Pathology* 32: 473-478.
- Huang, X., Zhao, N. and Zhang, K. 2004. Extracellular enzymes serving as virulence factors in nematophagous fungi involved in infection of the host. *Research in Microbiology* 155: 811-816.
- Hussey, R.S., and Janssen, G.J.W. 2002. Root-knot nematodes: *Meloidogyne* species. In *Plant Resistance to Parasitic Nematodes*. ed. J.L. Starr, R. Cook, and J. Bridge, pp. 43-70. London, England: CAB International.
- Ichinohe, M. 1984. Integrated control of the root-knot nematode, *Meloidogyne incognita*, on black-pepper plantations in the Amazonian region. *Agriculture, Ecosystems and Environment* 12: 271-283.
- Ignoffo, C.M. and Dropkin, V.H. 1977. Deleterious effects of the thermostable toxins of *Bacillus thuringiensis* on species of soil inhabiting, myceliophages, and plant parasitic nematodes. *Journal of Kansas Entomological Society* 50: 394-398.
- Inglis, P.W. and Tigano, M.S. 2006. Identification and taxonomy of some entomopathogenic *Paecilomyces* spp. (Ascomycota) isolates using rDNA-ITS Sequences. *Genetics and Molecular Biology* 29: 132-136.
- Irving, F. and Kerry, B.R. 1986. Variation between strains of the nematophagous fungus, *Verticillium chlamyosporium* Goddard. II. Factors affecting parasitism of cyst nematode eggs. *Nematologica* 32: 474-485 [abstract].
- Jacob, H., Gray, S.N. and Crump, D.H. 2003. Interactions between nematophagous fungi and consequences for their potential as biological agents for the control of potato cyst nematodes. *Mycological Research* 107: 47-56.
- Jansson, H.-B. and Lopez-Llorca, L.V. 2001. Biology of nematophagous fungi. In: *Trichomycetes and other fungal groups: Robert W. Lichtwardt commemoration volume*, ed. J.K. Misra and B.W. Horn, pp. 144-173. Enfield, NH: Science Publishers Inc.
- Jansson, H.-B., Tunlid, A. and Nordbring-Hertz, B. 1997. Nematodes. In *Fungal Biotechnology*, ed. T. Anke, pp. 38-48. Weinheim, Germany: Chapman and Hall.

- Jatala, P. 1986. Biological control of plant parasitic nematodes. *Annual Review of Phytopathology* 24: 453-489.
- Javed, N., Gowen, S. R., El-Hassan, S. A., Inam-ul-Haq, M., Shahina, F., and Pembroke, B. 2008. Efficacy of neem (*Azadirachta indica*) formulations on biology of root-knot nematodes (*Meloidogyne javanica*) on tomato. *Crop Protection* 27: 36-43.
- Jonathan, E.I., Padmanabhan, D. and Ayyamperumal, A. 1995. Biological control of root-knot nematode on betelvine, *Piper Betle*, by *Paecilomyces Lilacinus*. *Nematologia Mediterranea* 23: 191-193
- Jonathan, E.I., Padmanabhan, D. and Ayyamperumal, A. 2000. Field Application of *Paecilomyces lilacinus* for the control of *Meloidogyne incognita* on Betelvine, *Piper betle*. *Nematologia Mediterranea* 28: 131-133.
- Katsvairo, T.W., Rich, J.R. and Dunn, R.A. 2006. Perennial grass rotation: an effective and challenging tactic for nematode management with many other positive effects. *Pest Management Science* 62: 793-796.
- Kenney, D.S. 1986. DeVine-the way it was developed-a industrialist's view. *Weed Science* 34: 15-16.
- Kerry, B.R. 2001. Exploitation of the nematophagous fungal *Verticillium chlamydosporium* Goddard for the Biological control of root-knot nematodes (*Meloidogyne spp.*). In *Fungi as Biocontrol Agents: Progress, Problems and Potentials*. ed. T.M. Butt, C. Jackson, N. Magan, pp. 155-167. Wallingford, UK: CABI Publishing.
- Khan, A., Williams, K.L. and Nevalainen, H.K.M. 2004. Effects of *Paecilomyces lilacinus* protease and chitinase on the eggshell structures and hatching of *Meloidogyne javanica* juveniles. *Biological Control* 31: 346-352.
- Khan, A., Williams, K.L. and Nevalainen, H.K.M. 2006a. Control of plant-parasitic nematodes by *Paecilomyces lilacinus* and *Monacrosporium lysipagum* in pot trials. *BioControl* 51: 643-658.
- Khan, A., Williams, K.L. and Nevalainen, H.K.M. 2006b. Infection of plant-parasitic nematodes by *Paecilomyces lilacinus* and *Monacrosporium lysipagum*. *BioControl* 51: 659-678.
- Khan, M.Q., Abbasi, M.W., Zaki, M.J. and Khan, S.A. 2010. Evaluation of *Bacillus thuringiensis* isolates against root-knot nematodes following seed application in okra and mungbean. *Pakistan Journal of Botany* 42: 2903-2010.
- Khan, Z., Kim, S. G., Jeon, Y. H., Khan, H. U., Son, S. H., and Kim, Y. H. 2008. A plant growth promoting rhizobacterium, *Paenibacillus polymyxa* strain GBR-1, suppresses root-knot nematode. *Bioresource Technology* 99: 3016-3023.
- Khyami-Horani, H., M. Hajaij, J.-F. Charles. 2003. Characterization of *Bacillus thuringiensis* ser. Jordanica (Serotype H71), a novel serovariety isolated in Jordan. *Current Microbiology* 47: 26-31.

- Kiewnick, S. and Sikora, R.A. 2004. Optimising the biological control of plant parasitic nematodes with *Paecilomyces lilacinus* strain 251. *Phytopathology* 94: S51.
- Kiewnick, S. 2006. Effect of temperature on growth, germination, germ tubes extension and survival of *Paecilomyces lilacinus* strain 251. *Biocontrol Science and Technology* 16: 535-546.
- Kiewnick, S. and Sikora, R.A. 2006a. Evaluation of *Paecilomyces lilacinus* strain 251 for the biological control of the northern root-knot nematode *Meloidogyne hapla* Chitwood. *Nematology* 8: 69-78.
- Kiewnick, S., and Sikora, R. A. 2006b. Biological control of the root-knot nematode *Meloidogyne Incognita* by *Paecilomyces lilacinus* strain 251. *Biological Control* 38: 179-187.
- Kiewnick, S., Lauritzen, A.J. and Eilenberg, J. 2006a. Investigations on the colonization of root-knot nematode galls and egg masses by the antagonistic fungus *Paecilomyces lilacinus* strain 251. *Phytopathology* 96: 60 [abstract].
- Kiewnick, S., Neumann, S. and Sikora, R. 2006b. Importance of nematode inoculum density and antagonist dose for biocontrol efficacy of *Paecilomyces lilacinus* strain 251. *Phytopathology* 96:60 [abstract].
- Koenning, S.R., Overstreet, C., Noling, J.W., Donald, P.A., Becker, J.O., Fortnum, B.A. 1999. Survey of crop losses in response to phytoparasitic nematodes in the United States for 1994. *Journal of Nematology* 31: 587-618.
- Koshy, P.K., Santhosh, J., Eapen, S.J. and Pandey, R. 2005. Nematode parasites of spices, condiments and medicinal plant. In: *Plant Parasitic Nematode in Subtropical and Tropical Agriculture*, ed. M. Luc, R.A. Sikora and J. Bridge, pp. 751-792. Wallingford, UK: CAB International.
- Kueh, T.K. 1990. Major diseases of black pepper and their management. *The Planter* 66: 59-69.
- Kueh, T.K. and Sim, S.L. 1992. Slow decline of black pepper caused by root knot nematodes. In *Proceedings International Workshop on Black Pepper Disease*, ed. P. Wahid, D. Sitepu, S. Deciyanto and U. Suparman, pp. 198-206. Bogor, Indonesia: Research Institute for Spice and Medicinal Crops,
- Kueh, T.K and Teo, C.H. 1978. Chemical control of root-knot nematodes in *Piper nigrum*. *The Planter* 54: 237-245.
- Lacey, L.A. and Siegel, J.P. 2000. Safety and ecotoxicology of entomopathogenic bacteria. In *Entomopathogenic Bacteria: From Laboratory to Field Application*, ed. J.F. Charles, A. Delecluse and C. Nielsen, pp. 253-273. Dordrecht: Kluwer Academic.
- Ladner, D.C., P.B. Tchounwou and G.W. Lawrence. 2008. Evaluation of the effect of Ecologic on root knot nematode, *Meloidogyne incognita*, and tomato plant,

*Lycopersicon esculenum*. *International Journal of Environmental Research and Public Health* 5: 104-110.

- Lawrie J., Down, V.M., Greaves, M.P. 2001. Effects of storage on viability and efficacy of granular formulations of the microbial herbicides *Alternaria alternata* and *Terematophoma lignicola*. *Biocontrol Science and Technology* 11: 283-295.
- Lee, M.K., Walters, F.S., Hart, H., Palekar, N. and Chen, J.S. 2003. The mode of action of the *Bacillus thuringiensis* vegetative insecticidal protein Vip3A differs from that of Cry1Ab  $\delta$ -endotoxin. *Applied Environmental Microbiology* 69: 4648-4657.
- Leena, M.D., Easwaramoorthy, S. and Nirmala, R. 2003. *In vitro* production of entomopathogenic fungi *Paecilomyces farinosus* (Hotmskiold) and *Paecilomyces lilacinus* (Thoms.) Samson using byproducts of sugar industry and other agroindustrial byproducts and wastes. *Sugar Tech* 5: 231-236.
- Leong, C.T.S. 1986. Pepper nematodes. In: *Annual Report for the Year 1984*, pp. 74-78. Sarawak, Borneo: Department of Agriculture.
- Li, X.Q., Wei, J.Z., Tan, A. and Aroian., R.V. 2007. Resistance to root-knot nematode in tomato roots expressing a nematicidal crystal protein. *Plant Biotechnology* 5: 455-464.
- Lim, H.P. 2009. Bright outlook for pepper despite economic turmoil. *The Borneo Post*, March 27, B12.
- Liu, D., Coloe, S., Baird, R. and Pedersen, J. 2000. Rapid mini-preparation of fungal DNA for PCR. *Journal of Clinical Microbiology* 38: 471.
- Liu, Ya-Jun, Chong-Yan Zhai, Yi Liu and Ke-Qin Zhang. 2009. Nematicidal activity of *Paecilomyces* spp. and isolation of a novel active compound. *The Journal of Microbiology* 47: 248-252.
- Lopez-Llorca, L.V. and Duncan, G.H. 1991. Effects of fungal parasites on cereal cyst nematodes (*Heterodera avenae* Woll.) from naturally infested soil—a scanning electron microscopy study. *Canadian Journal of Microbiology* 37: 218–225 [abstract].
- Lopez-llorca, L.V., MaciÁ-Civente, J.G. and Jansson, H.-B. 2008. Mode of action and interactions of nematophagous fungi. In: *Ciancio, A. and K.G. Mukerji. eds. Integrated Management and Biocontrol of Vegetable and Grain Crops Nematodes*, pp. 51–76. Springer: The Netherlands.
- Lopez-Llorca, L.V., Olivares-Bernabeu, C., Salinas, J., Jansson, H.-B., and Kolattukudy, P.E. 2002. Prepenetration events in fungal parasitism of nematode eggs. *Mycological Research* 106: 499–506.
- Lugtenberg, B. and Kamilova, F. 2009. Plant-growth-promoting rhizobacteria. *Annual Review of Microbiology* 63: 541-556.

- Mac Innes, T.C. and Bouwer, G. 2009. An improved bioassay for the detection of *Bacillus thuringiensis* b-exotoxin. *Journal of Invertebrate Pathology* 101: 137-139.
- Mai, W.F. and Abawi, G.S. 1987. Interactions among root-knot nematodes and *Fusarium* wilt fungi on host plants. *Annual Review of Phytopathology* 25: 317-338.
- Marroquin, L.D., Elyassnia, D., Griffiths, J.S., Feitelson, J.S., Aroian, R.V. 2000. *Bacillus thuringiensis* toxin susceptibility and isolation of resistance mutants in the nematode *Caenorhabditis elegans*. *Genetics* 155: 1693-1699.
- Mateille, T., Dabiré, K.R., Fould, S. and Diop, M.T. 2010. Host-parasite soil communities and environmental constraints: Modelling of soil functions involved in interactions between plant-parasitic nematodes and *Pasteuria penetrans*. *Soil Biology & Biochemistry* 42: 1193-1199
- McClatche, G.V., Moore, D., Bateman, R.P. and Prior, C. 1994. Effects of temperature on the conidia of *Metarhizium flavoviride* in oil formulations. *Mycological Research* 9: 749-756.
- McInroy, J.A. and Kloepper, J.W. 1995. Survey of indigenous bacterial endophytes from cotton and sweet corn. *Plant and Soil* 173: 337-342.
- Melakeberhan, H., Xu, A., Kravchenko, A., Mennan, S. and Riga, E. 2006. Potential use of arugula (*Eurica sativa* L.) as a trap crop for *Meloidogyne hapla*. *Nematology* 8: 793-799.
- Mensink, B.J.W.G. and Scheepmaker, J.W.A. 2007. How to evaluate the environmental safety of microbial plant protection products: A Proposal. *Biocontrol Science and Technology* 17: 3-20.
- Meyer, S.L.F., Roberts, D.P., Chitwood, D.J., Carta, L.K., Lunsden, R.D. and Mao, W. 2001. Application of *Burkholderia cepacia* and *Trichoderma virens* alone and in combinations, against *Meloidogyne incognita* on bell pepper. *Nematropica* 31: 75-86.
- Mikami, Y., Yazawa, K., Fukushima, K., Arai, T., Udagawa, S.I. and Samson, R.A. 1989. Paecilotoxin production in clinical or terrestrial isolates of *Paecilomyces lilacinus* strains. *Mycopathologia* 108: 195-199.
- Mittal, N., Saxena, G. and Mukerji, K.G. 1995. Integrated control of root-knot disease in three crop plants using chitin and *Paecilomyces lilacinus*. *Crop Protection* 8: 647-651.
- Mohammed, S. H., Saedy, A.E.M., Enan, M.R., Ibrahim, N.E., Ghareeb, A. and Moustafa, S.A. 2008. Biocontrol efficiency of *Bacillus thuringiensis* toxins against root-knot nematode, *Meloidogyne incognita*. *Journal of Cell Molecular Biology* 7: 57-66.

- Mohandas, C. and Ramana, K.V. 1991. Pathogenicity of *Meloidogyne incognita* and *Radopholus similis* on black pepper (*Piper nigrum* L.). *Journal of Plantation Crops* 19: 41-43.
- Mondal, S.N., Kageyama, K. and Hyakumachi, M. 1995. Germinability, viability and virulence of chlamydospores of *Fusarium solani* f. sp. phaseoli as affected by the loss of endogenous carbon. *Phytopathology* 85: 1238-1244.
- Morgan-Jones, G., White, J.F. and Rodriguez-Kabana, R. 1984. Phytonematode pathology: Ultrastructural studies II. Parasitism of *Meloidogyne arenaria* eggs and larvae by *Paecilomyces lilacinus*. *Nematropica* 14: 57-71.
- Morton, C.O., Hirsch, P.R. and Kerry, B.R. 2004. Infection of plant-parasitic nematodes by nematophagous fungi –a review of the application of molecular biology to understand infection processes and to improve biological control. *Nematology* 6: 161-170.
- Mozgovaya, I.N., Byzov, B.A., Ryabchenko, N.F., Romanenko, N. D. and Zvyagintsev, D.G. 2002. Nematicidal effects of entomopathogenic bacteria *Bacillus thuringiensis* in soil. *Pedobiologia* 46: 558-572.
- Mukhtar, T. and Pervaz, I. 2003. *In vitro* evaluation of ovicidal and larvicidal effects of culture filtrate of *Verticillium chlamydosporium* against *Meloidogyne javanica*. *International Journal of Agriculture and Biology* 4: 576–579
- Mustika, I. 1990. *Studies on the interaction of Meloidogyne incognita, Radopholus similis and Fusarium solani on Balck Pepper (Piper nigrum L.)*, PhD Thesis, Wageningen Agricultural University, Wageningen, The Netherlands.
- Nagesh, M., Parvatha Reddy, P. and Rao, M. S. 1997. Integrated management of *Meloidogyne incognita* on tuberose using *Paecilomyces lilacinus* in combination with plant extracts. *Nematologia Mediterranea* 25: 3-7.
- Nitao, K.N., Meyer, S.L.F. and Chitwood, D.J. 1999. *In vitro* assays of *Meloidogyne incognita* and *Heterodera glycines* for detection of nematode-antagonistic fungal compounds. *Journal of Nematology* 1: 172-183.
- Nordbring-Hertz, B., Jansson, H.-B., and Tunlid, A. 2006. Nematophagous fungi. In *Encyclopedia of life sciences*, pp.1-11. Chichester: John Wiley & Sons, Ltd.
- Oclarit, E.L., and Cumagun, C.J.R. 2009. Evaluation of efficacy of *Paecilomyces lilacinus* as biological control agent of *Meloidogyne incognita* attaching tomato. *Journal of Plant Protection* 49: 337-340.
- Ohba, M. and Aizawa, K. 1986. Distribution of *Bacillus thuringiensis* from soils of Japan. *Journal of Invertebrate Pathology* 47: 277–282.
- Oka, Y. 2010. Mechanisms of nematode suppression by organic soil amendments— A review. *Applied Soil Ecology* 44: 101-115

- Oka, Y., Shapira, N. and Fine, P. 2007. Control of root-knot nematodes in organic farming systems by organic amendments and soil solarization. *Crop Protection* 26: 1556–1565.
- Oka, Y., Tkachi, N., Shuker, S. and Yerumiyahu, U. 2007b. Enhanced nematocidal activity of organic and inorganic ammonia-releasing amendments using neem extracts. *Journal of Nematology* 39: 9–16.
- Okhravi, N., Dart, J.K., Towler, H.M., Lightman, S. 1997. *Paecilomyces lilacinus* endophthalmitis with secondary keratitis: a case report and literature review. *Archives Ophthalmology* 115:1320–1324.
- Orion, D., Kritzman, G., Meyer, S.L.F., Erbe, E.F. and Chitwood, D.J. 2001. A role of the gelatinous matrix in the resistance of root-knot nematode (*Meloidogyne* spp.) eggs to microorganisms. *Journal of Nematology* 33: 203–207.
- Ornat, C. and Sorribas, J. 2008. Integrated management of root-knot nematodes in Mediterranean horticultural crops. In *Integrated Management and Biocontrol of Vegetable and Grain Crops Nematodes*. ed. A. Ciancio and K.G. Mukerji, pp 295–319. Dordrecht, The Netherlands: Springer.
- Ornat, C., Sorribas, F.J., Verdejo-Lucas, S. and Galeano, M. 2001. Effect of planting date on development of *Meloidogyne javanica* on lettuce in northeastern Spain. *Nematropica* 31: 148–149.
- Paau, A.S. 1988. Formulation useful in applying beneficial microorganisms to seeds. *Trends in Biotechnology* 6: 276-279.
- Padgham, J.L. and Sikora, R.A. 2007. Biological control potential and modes of action of *Bacillus megaterium* against *Meloidogyne graminicola* on rice. *Crop Protection* 26: 971-977.
- Page, S.L.J. and Bridge, J. 1985. Observations on *Pasteuria penetrans* as a parasite of *Meioidogyne arenaria*. *Nematologica* 31: 238-240.
- Padgham, J.L. and Sikora, R.A. 2007. Biological control potential and modes of action of *Bacillus megaterium* against *Meloidogyne graminicola* on rice. *Crop Protection* 26: 971-977.
- Park, J.-O., Hargreaves, J.R., McConville, E.J., Stirling, G.R., Ghisalberti, E.L. and Sivasithamparam, K. 2004. Production of leucinostatins and nematocidal activity of Australian isolates of *Paecilomyces lilacinus* (Thom) Samson. *Letters in Applied Microbiology* 38: 271–276.
- Paulus, A.D., Eng, L., Teo, C.H. and Sim, S.L. 1993. Screening black pepper genotypes and *Piper* spp. for resistance to root knot nematode. In *The Black Pepper Industry – Problems and Prospects*, ed. M.Y. Ibrahim, C.F.J. Bong and I.P. Ipor, pp. 132-139. Sarawak: Universiti Pertanian Malaysia Bintulu Campus.

- Pepper Ian, L. 2004. Filamentous fungi. In *Environmental Microbiology: A Laboratory Manual*, ed. Pepper Ian, L., pp. 27-36. Burlington, MA, USA: Academic Press.
- Persson, C. and Jansson, H.B. 1999. Rhizosphere colonization and control of *Meloidogyne* spp by nematode-trapping fungi. *Journal of Nematology* 31: 164–171.
- Prabhu, S., Kumar, S. and Subramanian, S. 2008. Mass production and commercial formulation of *Paecilomyces Lilacinus*. *Madras Agricultural Journal* 95: 415-417.
- Prasad, S.S.V., Tilsk, K.V.R. and Gollakota, K.G. 1972. Role of *Bacillus thuringiensis* var *thuringiensis* on the larval survivability and egg hatching of *Meloidogyne* spp. The causative agent of root-knot disease. *Journal of Invertebrate Pathology* 20: 377-378.
- Racke, J. and R.A. Sikora. 1992. Isolation, formulation and antagonistic activity of rhizobacteria toward the potato cyst nematode *Globodera pallida*. *Soil Biology and Biochemistry* 24: 521-526.
- Ramana, K.V. and Mohandas, C. 1986. Reaction of black pepper germplasm to root knot nematode *Meloidogyne incognita*. *Indian Journal of Nematology* 16: 138-139.
- Ramana, K.V. 1994. Efficacy of *Paecilomyces lilacinus* (Thom.) Samson in suppressing nematode infestations in black pepper (*Piper nigrum* L.). *Journal of Spices & Aromatic Crops* 3: 130-134.
- Ramana, K.V. and Eapen, S.J. 2000. Nematode induced diseases of black pepper. In *Black Pepper Piper nigrum*, ed. P. Ravindran, pp. 269-295. Malaysia: Harwood academic publishers.
- Rang, C., Gil, P., Neisner, N., Van Rie, J. and Frutos, R. 2005. Novel Vip3-related protein from *Bacillus thuringiensis*. *Applied and Environmental Microbiology* 71: 6276–6281.
- Rao, M.S., Parvatha, Reddy, P.P. and M. Nagesh. 1999. Bare root dip treatment of tomato seedlings in calotropis or castor leaf extracts mixed with *Paecilomyces lilacinus* spores for the management of *Meloidogyne incognita*. *Nematologia Mediterranea* 27: 323-326.
- Rhodes, D.J. 1990. Formulation requirements for biological control agents. *Aspects of Applied Biology* 24: 145-153.
- Rich, J.R., Dunn, R. and Noling, J. 2004. Nematicides: Past and present uses. In: *Nematology: Advances and Perspectives, Vol 2. Nematode Management and Utilization*, ed. Z.X. Chen, S.Y. Chen and D.W. Dickson, pp. 1041-1082. Wallingford, UK: CABI Publishing.



- Roberts, P. A. 1995. Concept and practical aspects of variability in root-knot nematodes related to host plant resistance. *Annual Review Phytopathology* 33: 199-221.
- Robl, D., Sung, L.B., Novakovich, J.H., Marangoni, P.R.D., Zawadneak, M.A.C., Dalzoto, P.R., Gabardo, J., Pimentel, I.C. 2009. Spore production in *Paecilomyces lilacinus* (THOM.) Samsons Strain on Agro-industrial residue. *Brazilian Journal of Microbiology* 40: 296-300.
- Rumbos, C. and Kiewnick, S. 2006. Effect of plant species on persistence of *Paecilomyces lilacinus* strain 251 in soil and on root colonization by the fungus. *Plant and Soil* 283:25-31.
- Rumbos, C., Mendoza, A., Sikora, R. and Kiewnick, S. 2008. Persistence of the nematophagous fungus *Paecilomyces lilacinus* strain 251 in soil under controlled conditions. *Biocontrol Science and Technology* 18: 1041-1050.
- Rumbos, C., Reimann, S., Kiewnick, S. and Sikora, R.A. 2006. Interaction of *Paecilomyces lilacinus* strain 251 with mycorrhizal fungus *Glomus intraradices*: implications for *Meloidogyne incognita* control on tomato. *Biocontrol Science and Technology* 16: 981-986.
- Safdar, A. 2002. Progressive cutaneous hyalohyphomycosis due to *Paecilomyces lilacinus*: rapid response to treatment with caspofungin and itraconazole. *Clinical Infectious Disease* 34: 1415–1417.
- Salehi Jouzani, G., Seifinejad, A., Saedizadeh, A., Nazarian, A., Yousefloo, M., Soheilvand, S., Mousivand, M., Jahangiri, R., Yazdani, M., Amiri, R.M. and Akbari, S. 2008. Molecular detection of nematocidal crystalliferous *Bacillus thuringiensis* strains of Iran and evaluation of their toxicity on free-living and plant-parasitic nematodes. *Canadian journal of Microbiology* 54: 812-822.
- Samson, R. A. 1975. *Paecilomyces* and some allied hyphomycetes. Studies in Mycology 6: 1-119. *The Netherlands: Centraalbureau voor Schimmelcultures, Baarn.*
- Sasanelli N., Ciccicarese, F. and Papajova, I. 2008. *Aphanocladium album* by via subirrigation in the control of *Pyrenochaeta lycopersici* and *Meloidogyne incognita* on tomato in a plastic-house. *Helminthologia* 45: 137–142.
- Sasser, J.N. and Freckman, D.W. 1987. World perspective on nematology. The role of the society. In: *Vistas on Nematology: A Commemoration of the 25th Anniversary of the Society of Nematologists*, ed. J.A. Veech and D.W. Dickson, pp. 7–14. Inc., Hayattsville, MD: Society of Nematologists.
- Schnepf, E., Crickmore, N., Van Rie, J., Lereclus, D., Baum, J., Feitelson, J., Zeigler, D.R. and Dean, D.H. 1998. *Bacillus thuringiensis* and its pesticidal crystal proteins. *Microbiology and Molecular Biology Reviews* 62:775–806.

- Sebesta, K. and Horska, K. 1970. Mechanism of inhibition of DNA dependent RNA polymerase by exotoxin of *Bacillus thuringiensis*. *Biochimica et Biophysica Acta* 209: 357-376.
- Sewell, G.W.F. 1965. The effect of altered physical conditions of soil on biological control. In *Ecology and Soil Borne Plant Pathogens*, ed. K.F. Baker and W.C. Snyder, pp. 479-494. U.S.A: University of California Press, Berkeley.
- Shabana Y.M., Muller-Stover D. and Sauerborn, J. 2003. Granular pest formulation of *Fusarium oxysporum* f. sp. *orthoceras* for biological control of sunflower broomrape: efficacy and shelf-life. *Biological Control* 26: 189-201.
- Sharif, F. A. and N. G. Alaeddinoğlu. 1998. A rapid and simple method for staining of the crystal protein of *Bacillus thuringiensis*. *Journal of Industrial Microbiology* 3: 227-229.
- Sharma, P. and Pandey, R. 2009. Biological control of root-knot nematode; *Meloidogyne incognita* in the medicinal plant; *Withania somnifera* and the effect of biocontrol agents on plant growth. *African Journal of Agricultural Research* 4: 564-567.
- Sharma, R.D. 1994. *Bacillus thuringiensis*: a biocontrol agent of *Meloidogyne incognita* on Barley. *Nematologia Brasileira* 18: 79-84.
- Sharon, E., Chet, I., Viterbo, A., Bar-Eyal, M., Nagan, H., Samuels, G.J. and Spiegel, Y. 2007. Parasitism of *Trichoderma* on *Meloidogyne javanica* and role of the gelatinous matrix. *European Journal of Plant Pathology* 118: 247-258.
- Siddiqui, I.A., and Shahid Shaukat, S. 2003. Suppression of root-knot disease by *Pseudomonas fluorescens* CHA0 in tomato: Importance of bacterial secondary metabolite, 2,4- diacetylphloroglucinol. *Soil Biology and Biochemistry*. 35: 1615-1623.
- Siddiqui, I.A., Qureshi, S.A., Sultana, V., Ehteshamul-Haque, S. and Ghaffar, A. 2000. Biological control of root rot-root knot disease complex of tomato. *Plant and soil* 227: 163-169
- Siddiqui, Z.A. 2005. PGPR: Prospective Biocontrol Agents of Plant Pathogens. In *PGPR: Biocontrol and Biofertilization*, ed. Z.A. Siddiqui, pp. 111-142. The Netherlands: Springer.
- Siddiqui, Z.A. and Akhtar, M.S. 2008. Synergistic effects of antagonistic fungi and a plant growth promoting rhizobacterium, an arbuscular mycorrhizal fungus, or composted cow manure on populations of *Meloidogyne incognita* and growth of tomato. *Biocontrol Science and Technology* 3: 279-290.
- Siddiqui, Z.A. and Akhtar, M.S. 2009. Effect of plant growth promoting rhizobacteria, nematode parasitic fungi and root-nodule bacterium on root-knot nematodes *Meloidogyne javanica* and growth of chickpea. *Biocontrol Science and Technology* 19: 511-521.

- Siddiqui, Z. A. and Mahmood, I. 1994. Culture of *Paecilomyces lilacinus* on leaf extracts and leaf residues for nematode control. *Bioresource Technology* 49: 187–189.
- Siddiqui, Z.A., and Mahmood, I. 1999. Role of bacteria in the management of plant parasitic nematodes: A review. *Bioresource Technology* 69: 167-179.
- Siegel, J.P. 2001. Minireview: The mammalian safety of *Bacillus thuringiensis* based insecticides. *Journal of Invertebrate Pathology* 77: 13-21.
- Sijmons, P.C. 1994. Parasitic strategies of root nematodes and associated host cell responses. *Annual Review of Phytopathology* 32: 235-259
- Singh, P and Z. A Siddiqui. 2010. Biocontrol of root-knot nematode *Meloidogyne incognita* by the isolates of *Bacillus* on tomato. *Archives of Phytopathology and Plant Protection* 43: 552-561.
- Singh, S and Mathur, N. 2010. *In vitro* studies of antagonistic fungi against the root-knot nematode, *Meloidogyne incognita*. *Biocontrol Science and Technology* 20: 275-282.
- Sipes, V.S., Caswell-Chen, E.P., Sarah, J.-L. and Apt, W.J. 2005. Nematode parasites of pineapple. In *Plant Parasitic Nematodes in Subtropical and Tropical Agriculture*, ed. M. Luc, R.A. Sikora, and J. Bridge, pp. 723-724. Oxfordshire, UK: CABI.
- Siti Hajijah, A.S. 1993. Observation of root knot infestation on pepper (*Piper nigrum* L.) in Sarawak. In *The Pepper Industry-Problem and Prospects*. ed. M.Y. Ibrahim, C.F.J. Bong and I.P. Ipor, pp 140-147. Sarawak, Malaysia: Universiti Pertanian Malaysia.
- Studdert J.P. and Kaya, H.K. 1990. Effect of water potential, temperature, and clay-coating on survival of *Beauveria bassiana* conidia in a loam and peat soil. *Journal of Invertebrate Pathology* 55: 417–427.
- Sun, M.H., Gao, L., Shi, Y.X., Li, B.J. and Liu, X. Z. 2006. Fungi and actinomycetes associated with *Meloidogyne spp.* eggs and females in China and their biocontrol potential. *Journal of Invertebrate Pathology* 93: 22-28.
- Tabashnik, B.E., Patin, A.L., Dennehy, T.J., Liu, Y.B., Carrière, Y., Sims, M.A. and Antilla, L. 2000. Frequency of resistance to *Bacillus thuringiensis* in field populations of pink bollworm. *PNAS* 97: 12980-12984.
- Tadayyon A., Hill, G.A., Ingledew, W.M. and Sokhonsanj, S. 1997. Contact-sorption drying of *Penicillium bilaii* in a fluidized bed dryer. *Journal of Chemical Technology and Biotechnology* 68: 277-282.
- Tan, T.Q., Ogden, A.K., Tillman, J., Demmler, G.J., Rinaldi, M.G. 1992. *Paecilomyces lilacinus* catheter-related fungemia in an immunocompromised pediatric patient. *Journal of Clinical Microbiology* 30: 2479–2483.

- Terefe, M., Tefera, T., and Sakhuja, P. K. 2009. Effect of a formulation of *Bacillus firmus* on root-knot nematode *Meloidogyne incognita* infestation and the growth of tomato plants in the greenhouse and nursery. *Journal of Invertebrate Pathology*, 100: 94-99.
- Thuy, T.T.T. 2010. *Incidence and effect of Meloidogyne incognita (Nematoda: Meloidogyninae) on black pepper plants in Vietnam*, PhD Thesis, University of Agriculture, Vietnam.
- Travers, R.S., Martin, P.A.W. and Reichelderfer, C.F. 1987. Selective process for efficient isolation of soil *Bacillus* spp. *Applied and Environmental Microbiology* 53: 1263–1266.
- Trudgill, D. L. 1991. Resistance and tolerance of plant parasitic nematodes in plants. *Annual Review Phytopathology* 29: 167-192.
- Tunlid, A. and Jansson, S. 1991. Proteases and their involvement in the infection and immobilization of nematodes by nematophagous fungus *Arthrobotrys oligospora*. *Applied and Environmental Microbiology* 57: 2868–2872.
- Tzortzakakis, E. A., Trudgill, D. L., and Phillips, M. S. 1998. Evidence for a dosage effect of the Mi gene on partially virulent isolates of *Meloidogyne javanica*. *Journal of Nematology* 30: 76–80.
- van Frankenhuyzen, K. 2009. Insecticidal activity of *Bacillus thuringiensis* crystal proteins. *Journal of Invertebrate Pathology* 101: 1-16.
- Van Lenteren, J.C., Babendreier, D., Bigler, F., Burgio, G., Hokkanen, H.M.T., Kuske, S., Loomans, A.J.M., Menzler-Hokkanen, I., Van Rijn, P.C.J., Thomas, M.B., Tommasini, M.G., and Zeng, Q.-Q. 2003. Environmental risk assessment of exotic natural enemies used in inundative biological control. *Biocontrol* 48: 3-38.
- Van Peer, R., Niemann, G.J. and Schippers, B. 1991. Induced resistance and phytoalexin accumulation in biological control of *Fusarium* wilt of carnation by *Pseudomonas* sp. strain WCS417r. *Phytopathology* 81: 728–34.
- Vanaja, T., Neema, V.P., Rajesh, R. and Mammooty, K.P. 2007. Graft recovery of *Piper nigrum* L. runner shoots on *Piper colubrinum* Link. rootstocks as influenced by varieties and month of grafting. *Journal of Tropical Agriculture* 45: 61-62.
- Vilas-Bôas, G.T., Peruca, A.P.S. and Arantes, O.M.N. 2007. Biology and taxonomy of *Bacillus cereus*, *Bacillus anthracis*, and *Bacillus thuringiensis*. *Canadian Journal of Microbiology* 53: 673-87.
- Walia, R.K., Nandal, S.N. and Bhatti, D.S. 1999. Nematicidal efficacy of plant leaves and *Paecilomyces lilacinus*, alone or in combination, in controlling *Meloidogyne incognita* on okra and tomato. *Nematologia Mediterranea* 27: 3-8.

- Walia, R.K., Sharma, S.B. and Vats., R. 2000. Bacterial antagonists of phytonematodes. In *Biocontrol Potential and Its Exploitation in Sustainable Agriculture. Volume 1: Crop Diseases, Weeds, and Nematodes*, ed. R.K. Upadhyay, K.G. Mukerji and B.P. Chamola, pp. 25-37. Spring Street, New York: Kluwer Academic/Plenum Publisher.
- Walters, S.A., Wehner, T.C. and Barker, K.R. 1997. A single recessive gene for resistance to the root-knot nematode (*Meloidogyne javanica*) in *Cucumis sativus* var. *hardwickii*. *The Journal of Heredity* 88: 66-69.
- Wang, Q., Klassen, W. and Handoo, Z. 2007. Influence of cover crops and soil amendments on okra (*Abelmoschus esculentus* L.) production and soil nematodes. *Renewable Agriculture and Food Systems* 22: 41-53
- Wei, J.Z., Hale, K., Carta, L., Platzer, E., Wong, C., Fang, S.C. and Aroian, R.V. 2003. *Bacillus thuringiensis* crystal protein that target nematodes. *Proceedings of the National Academy of Sciences of the USA* 100: 2760-2765.
- Weller, D.M. 1988. Biological control of soilborne plant pathogens in the rhizosphere with bacteria. *Annual Review of Phytopathology* 26: 379-407.
- Wesemael, W. 2007. Biology and management of the root-knot nematode *Meloidogyne chitwoodi* in field vegetable crops, *PhD Thesis*, Ghent University, Belgium.
- Wessolossky, M., Haran, J.P. and Bagchi, K. 2008. *Paecilomyces lilacinus* olecranon bursitis in an immunocompromised host: case report and review. *Dignostic Microbiology and Infectious Disease* 61: 354-357.
- White, T.J., Bruns, T., Lee, S. and Taylor, J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In *PCR protocols, a guide to methods and applications*, ed. M.A. Innis, D.H. Gelfand, J.J. Sninsky, T.J. White, pp 315-322. San Diego: Academic Press.
- Williamson, V. and Hussey, R.S. 1996. Nematode pathogenesis and resistance in plants. *The Plant Cell* 8: 1735-1745.
- Williamson, V.M., and Kumar, A. 2006. Nematode resistance in plants: The battle underground. *Trends in Genetics* 22: 396-403
- Winoto, R.S. 1972. Effect of *Meloidogyne* species on the growth of *Piper nigrum* L. *Malaysia Agricultural Research* 1: 86-89.
- Wu, W., Shen, H., and Yang, W. 2009. Sources for heat-stable resistance to southern root-knot nematode (*Meloidogyne incognita*) in *Solanum lycopersicum*. *Agricultural Sciences in China* 8: 697-702.
- Xavier, R., Reena Josephine, C.M. and Sreeramanan, S. 2007. Environmental distribution and diversity of insecticidal proteins of *Bacillus thuringiensis* Berliner. *Malaysian Journal of Microbiology* 3: 1-6.

- Yu, Y.M., Cho, M.R., Zhu, Y.Z., Park, D.H., Hur, J.H. and Lim, C.K. 2003. Suppression of *Meloidogyne incognita* in Lettuce and Oriental Melon by *Pasteuria penetrans* KW1. *The Plant Pathology Journal* 19: 177-180
- Yu, Z.Q., Zhou, Y., Sun, M. and Yu, Z.N. 2004. Progress of research on activity of *Bacillus thuringiensis* against plant-parasitic nematodes. *Acta Phytophylacica Sinica* 31: 418-424.
- Yu, Z.Q., Wang, Q.L., Liu, B., Zou, X., Yu, Z.N. and Sun, M. 2008. *Bacillus thuringiensis* crystal protein toxicity against plant-parasitic nematodes. *Chinese Journal of Agricultural Biotechnology* 5: 13-17.
- Zakeel, M.C.M., Dissanayake D.M.D. and Weerasinghe P.A. 2009. Molecular characterization of *Bacillus thuringiensis* strains isolated from a selected site in nochchiyagama, Anuradhapura in Sri Lanka. *Tropical Agricultural Research and Extension* 12: 31-34.
- Zaki F.A. and Bhatti, D.S. 1991. Effect of culture media on sporulation of *Paecilomyces lilacinus* and its efficacy against *Meloidogyne javanica* in tomato. *Nematologia Mediterranea* 19: 211-212.
- Zasada, I. A., Halbrendt, J. M., Kokalis-Burelle, N., LaMondia, J., McKenry, M. V., and Noling, J. W. 2010. Managing nematodes without methyl bromide. *Annual Review of Phytopathology* 48: 311-328.
- Zulfiqar, M. and Gul, A.S. 1990. Organic amendments as control of root-knot nematodes. *Int. Nematol. Network*. 7: 22-24.

## BIODATA OF STUDENT

Pau Chen Guan, born in 18 May 1974, originated from Sri Aman, Sarawak. He completed his primary school in Sekolah Rendah Kebangsaan Stampin in 1986, secondary school in SMK Gapor Stampin, Kuching in 1992 and high school in SMK Green Road, Kuching in 1994. He successfully obtained Bachelor of Science (Hons) in Microbiology with second class upper (CGPA 3.149) from Universiti Putra Malaysia, Serdang in 1999. After that he worked as a temporary teacher in SMB St Thomas, Kuching for a year and as an education officer in Sarawak Biodiversity Centre for six months under contract before he proceeded for diploma in education in Maktab Perguruan Keningau, Sabah for nine months. After completed his diploma in education, he was posted to SMK Simanggang, Sri Aman in January 2002 as a form six biology teacher for three years before transferred back to Kuching. He taught SPM biology and chemistry in SMK Lumba Kuda for four and a half year. In July 2009, he was granted with two year unpaid study leave from Bahagian Tajaan, Kementerian Pelajaran Malaysia to pursue Master of Science (Plant Pathology) which he is undertaking now in Faculty of Agriculture and Food Sciences, Universiti Putra Malaysia Bintulu Sarawak Campus. In July 2011, he was posted back to his former school, SMK Green Road, Kuching to teach biology at STPM and SPM level while waiting for his viva voice and final thesis correction.

## LIST OF PUBLICATIONS

1. Pau, C.G., C.T.S. Leong, S.K. Wong, L. Eng, M. Jiwan, F.R. Kundat, Z.F.B.A. Aziz, O.H. Ahmed and N.M. Majid, 2012. Isolation of indigenous strains of *Paecilomyces lilacinus* with antagonistic activity against *Meloidogyne incognita*. *Int. J. Agric. Biol.* 14: 197–203 [Published]
2. Pau, C.G., C.T.S. Leong, S.K. Wong, L. Eng, M. Jiwan, F.R. Kundat, Z.F.B.A. Aziz, O.H. Ahmed and N.M. Majid. 2012. Evaluation of *Paecilomyces lilacinus* and *Bacillus thuringiensis* as microbial control agents for *Meloidogyne incognita* in black pepper (*Piper nigrum* L.) planted in pots under open house condition. *Biocontrol Science and Technology* (Under review).