



**UNIVERSITI PUTRA MALAYSIA**

***INDUCED RESISTANCE OF OIL PALM SEEDLINGS AGAINST BASAL  
STEM ROT USING CALCIUM, COPPER AND SALICYLIC ACID***

**RAHAMATH BIVI BINTI M SHAHUL HAMEED**

**FSPM 2013 4**



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**By**

**RAHAMATH BIVI BINTI M SHAHUL HAMEED**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Master Science**

**October 2013**

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**October 2013**

**Chairman : Khairulmazmi Ahmad, PhD**

**Faculty : Agriculture and Food Sciences (Bintulu)**

The oil palm (*Elaeis guineensis*) tree, which belongs to the Palma family, is one of the most versatile crops in tropical countries. Oil palm was the major commodity inside the Malaysian agriculture sector. Malaysia is presently the world's leading exporter of oil palm and it was second only to soybean as the major source of vegetable oil. However, a soil fungus pathogen, *Ganoderma boninense*, which causes basal stem rot (BSR) in the oil palm's trunk, destroys thousands of hectares of plantations in Southeast Asia every year. Thus far there was no conclusive method for early detection of BSR in order to control the disease from spreading in the plantation.

Plant nutrient and hormone are vital variables and modifying these as a control method may prevent *Ganoderma* attack. The limited nutrients and hormone of plants can trigger lignin degradation and conversely ensuring they are available for oil palm may limit the rot. These nutrients could conceivably be supplied by foliar and soil application with a view to control the disease in addition to them being used as fertilizers. Fertilizers are added which hold some of these minerals, but the point here is to control the disease and also improve the growth of the oil palm. The objective of the present work is to study calcium (Ca), copper (Cu) and salicylic acid (SA) application in oil palm and to determine their potential as fertilizers to control against *Ganoderma boninense* *in vitro* and *in vivo*.



The *in vitro* effects of single and combined application of Ca, Cu and SA were evaluated on growth and sporulation of *G. boninense*. In poison medium test, T7- (Ca+Cu+SA) showed effective control of *G. boninense in-vitro* with EC<sub>50</sub> and EC<sub>90</sub> values of 1500+150+150 ppm and 2000+200+200 ppm, respectively. However, in dipping test, T7- (Ca+Cu+SA) indicates effective control of *G. boninense* at low concentration, 500+50+50 ppm as shown by EC<sub>90</sub> analysis. Pre-treatment of *Ganoderma*-infected rubber wood block with 500 ppm Ca+50 ppm Cu+50 ppm SA reduced the number (20.14%) and (0.8 g) weight of basidiocarp compared to the control. This was followed by a significant reduction in weight loss of the *Ganoderma*-infected rubber wood block (41.85%) suggesting the inhibition of the degradative enzymatic activity of the fungus. The mixture of Ca, Cu and SA had potential to suppress growth of *G. boninense* under *in vitro* condition.

The effect of Ca, Cu and SA further tested in a glass house for about 9 months for their efficacy in controlling BSR in oil palm seedlings through foliar and soil drenching by control of water application. As tested in oil palm seedlings inoculated with *G. boninense*, both of them, alone and in mixture suppressed *G. boninense* compared to the control. Not only they delayed the onset of BSR symptoms, but also promoted the growth of the seedlings. The application of Ca, Cu and SA together reduced disease incidence most effectively and was recorded reduction at 81.0% and followed by BSR disease incidence reduced for mixture of Ca and SA; a mixture of Cu and SA; and Ca alone respectively with a 76.1% reduction. In addition, treatment with Ca+Cu+SA increased the levels of the defense related enzymes phenolics, peroxidase, lignin and hydrogen peroxide in oil palm seedling tissues. Plant growth and antibiosis, plant nutrient and hormone enhanced the resistance in plants through the induction of defense enzymes in the oil palm seedlings.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains.

**ARUHAN RINTANGAN ANAK BENIH KELAPA SAWIT TERHADAP  
REPUT PANGKAL BATANG MENGGUNAKAN KALSIUM, KUPRUM DAN  
ASID SALISILIK**

Oleh

**RAHAMATH BIVI BINTI M SHAHUL HAMEED**

**Oktober 2013**

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Pokok kelapa sawit (*Elaeis guineensis*), yang tergolong dalam famili Palma adalah salah satu tanaman yang paling versatil di negara-negara tropika. Kelapa sawit merupakan komoditi utama di dalam sektor pertanian Malaysia. Kini, Malaysia adalah pengeksport terkemuka di dunia minyak sawit dan ia adalah kedua selepas kacang soya sebagai sumber utama minyak sayuran. Walau bagaimanapun, patogen kulat tanah, *Ganoderma boninense*, yang menyebabkan reput pangkal pada batang pokok kelapa sawit, memusnahkan beribu-ribu hektar ladang di Asia Tenggara setiap tahun. Setakat ini tidak ada kaedah muktamad untuk pengesanan awal BSR untuk mengawal penyakit daripada merebak.

Nutrien dan hormon tumbuhan adalah pembolehubah penting dan mengubahsuai ianya sebagai kaedah kawalan boleh menghalang serangan *Ganoderma*. Mengehadkan nutrisi dan hormon tumbuhan boleh mencetuskan degradasi lignin dan sebaliknya memastikan ianya sedia ada untuk kelapa sawit boleh mengehadkan pereputan. Nutrien tersebut boleh digunakan untuk dibekalkan pada foliar dan tanah dengan tujuan untuk mengawal penyakit itu di samping digunakan sebagai baja. Baja mengandungi beberapa mineral bagi mengawal penyakit dan juga untuk meningkatkan pertumbuhan kelapa sawit. Objektif penyelidikan ini adalah untuk mengkaji aplikasi kalsium (Ca), kuprum (Cu) dan asid salisilik (SA) kepada anak benih kelapa sawit dan menentukan potensi mereka sebagai baja untuk mengawal jangkitan *G. boninense* secara *in vitro* dan *in vivo*.

Kesan *in vitro* tunggal dan campuran aplikasi Ca, Cu dan dan SA telah dinilai pada tumbesaran dan sporulasi *G. boninense*. Pada ujian media racun, T7- (Ca+Cu+SA) menunjukkan kawalan berkesan terhadap *G. boninense* secara *in vitro* dengan nilai  $EC_{50}$  dan  $EC_{90}$  pada 1500+150+150 ppm dan 2000+200+200 ppm, masing-masing. Walau bagaimanapun, pada ujian pencelupan, T7- (Ca+Cu+SA) menunjukkan kawalan yang berkesan terhadap *G. boninense* pada kepekatan rendah, 500+50+50 ppm seperti ditunjukkan pada analisis  $EC_{90}$ . Pra-rawatan pada blok kayu getah yang dijangkiti *Ganoderma* pada 500 ppm Ca+50 ppm Cu+50 ppm SA mengurangkan bilangan (20.14%), dan (0.8 g) berat basidiokap berbanding kawalan. Ini diikuti oleh pengurangan yang ketara dalam penurunan berat pada blok kayu getah yang dijangkiti *Ganoderma* (41.85%) menunjukkan perencatan aktiviti enzim degradasi daripada kulat. Campuran Ca, Cu dan SA mempunyai potensi untuk menghalang pertumbuhan *G. boninense* secara *in vitro*.

Kesan Ca, Cu dan SA diuji lagi dalam rumah kaca selama 9 bulan untuk keberkesanan mereka bagi mengawal RPB pada anak benih kelapa sawit melalui aplikasi pada foliar dan tanah dengan kawalan penyiraman air. Seperti yang diuji pada benih kelapa sawit yang disuntik dengan *G. boninense*, kedua-dua mereka, bersendirian dan campuran menghalang *G. boninense* berbanding kawalan. Bukan sahaja mereka melambatkan kemunculan simptom RPB tetapi juga menggalakkan pertumbuhan benih. Aplikasi Ca, Cu dan SA bersama-sama mengurangkan insiden penyakit paling berkesan dan merekod pengurangan 81.0% dan diikuti oleh insiden penyakit RPB dikurangkan bagi campuran Ca dan SA; campuran Cu dan SA; dan Ca sahaja masing-masing dengan penurunan 76.1%. Di samping itu, rawatan dengan Ca+Cu+SA meningkatkan tahap enzim yang berkaitan ketahanan seperti phenolik, peroxidase, lignin, dan hidrogen peroksida dalam tisu anak benih kelapa sawit. Pertumbuhan pokok dan antibiosis, nutrien tumbuhan dan hormon meningkatkan rintangan dalam tumbuhan melalui induksi enzim pertahanan dalam anak benih kelapa sawit.



## ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and the Most Merciful.

First of all, I must acknowledge and thank The Almighty Allah for blessing, protecting and guiding me throughout this period. I could never have accomplished this without the faith I have in the Almighty. It gives me great pleasure in conveying my gratitude to all those people who have supported me and had their contributions in preparing this thesis possible.

I express my profound sense of reverence to my supervisor Dr. Khairulmazmi bin Ahmad, for his constant guidance, support, motivation and assistance throughout the course of my Master. I as well express my deepest gratitude to committee members, especially Dr. Idris bin Abu Seman and Dr. Susilawati Kasim for boosting my morale during the course of research. They always cared, a source of wisdom and motivation. I also would like to thank to Associate Prof. Dr. Osumanu Haruna Ahmed for helping me in statistical analysis and being available to guide me in my projects. Being a statistical leader, I always find his comments, questions and suggestions in the manuscripts very challenging and I always felt very relaxed after answering his concerns.

My acknowledgement also goes to all the staffs of the Institute of Bioscience, UPM, Serdang for their co-operations and assistance. Sincere thanks to all staff in UPM Bintulu campus, Sarawak especially Kak Siti Fatimah Razali and others for their kindness and moral support during my research work. One person who has always been ready to accompany and help me was my best friend Siti Noor Farhana binti Md Daut.

*I can't imagine my current position without the care and support from my family. I thank my parents, M. Shahul Hameed and Halimah @ Sabina for striving hard to provide a good education for me and my siblings. I always fall short of words and felt impossible to describe their support in words and thanks to my siblings for their encouragement and care. To those who indirectly contributed to this research, your kindness means a lot to me. Thank you very much.*

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## LIST OF ABBREVIATIONS

AAS	Atomic Absorption Spectrophometer
ANOVA	Analysis of Variance
BSR	Basal Stem Rot
CRD	Completely Randomized Design
CWA	Cell Wall Apposition
CWDE	Cell Wall Degrading Enzymes
DI	Disease Incidence
DR	Disease Reduction
DSI	Disease Severity Index
EC <sub>50</sub>	Effective Concentration 50%
EC <sub>90</sub>	Effective Concentration 90%
ER	Epidemic Rate
E	Effective
LE	Least Effective
HR	Hypersensitive Reaction
IAA	Indole-3-Acetic Acid
MAI	Month After Inoculation
MEA	Malt Extract Agar
μL	Micro Liter
μm	Micrometer
mM	Milimolar
M	Molar
MIT	Methylisothiocyanate

MPOB	Malaysian Palm Oil Board
nm	Nanometer
NE	Not Effective
PAL	Phenylalanine Ammonia Lyase
PDA	Potato Dextrose Agar
PIRG	Percentage Inhibition of Radial Growth
POD	Peroxidase
ppm	Parts per million
PR	Pathogenesis Related
psi	Per square inch
RCBD	Randomised Completely Block Design
SA	Salicylic Acid
SAS	Statistical Analysis Software
SEM	Scanning Electron Microscopy
spp	Species
TEM	Transmission Electron Microscope
TPC	Total Phenolics Content
v/v	Volume/ volume
WAI	Week After Inoculation
WPL	Weight Percent Loss



## CHAPTER 1

### GENERAL INTRODUCTION

Oil palm (*Elaeis guineensis*) is the most significant commodity crop in the Malaysian agricultural sector. The rapid increase in area planted with oil palm from 300,000 ha in 1970 to 5.0 million ha in 2011 (Sime Darby, 2012) shows the economic importance of this crop. According to Sime Darby (2012), the export earnings from oil palm products reached about RM80.41 billion in 2011.

However, a soil borne fungus identified as *Ganoderma boninense*, which induced basal stem rot (BSR) disease in the oil palm's trunk, ruins thousands of hectares of plantations in Southeast Asia almost every year. A direct loss of oil palm trees when the disease causing infected palm trunks to break at the base. About (30 to 70%) losses of oil palm occurred from 4.85 million hectares of the total oil palm area, due to BSR disease which have an adverse effect on the oil palm industry. This disease is conceived to be the most serious oil palm disease in Malaysia and other parts of South East Asia (Susanto, 2009).

In Peninsular Malaysia, most of the oil palm estates discover this disease in the second or the third replanting. About 90% of the estates in West Malaysia have been infested with *G. boninense* (Khairuddin and Chong, 2008). In Sabah and Sarawak, most of the estates are in their first cycle of planting, allowing for alternative hypotheses for the lower relative incidence of pathogen attacks in these regions. Thus, *G. boninense* has a significant effect on the lifetime of affected trees and adverse effect on yield (Corley and Tinker, 2003).

The *G. boninense* was found to infect 4 to 5 years old palms more drastically compare than 1 to 2 years old (Ariffin *et al.*, 2000). The symptoms of oil palm disease are withering, yellowing of fronds followed by necrosis on one side of older fronds. Necrosis occurring in the lower leaves extend progressively to younger leaves of the crown. When the foliar symptoms were observed, at least one-half of the basal stem has been killed by the fungus. Young palms can take 6 months to 2 years to die after the first symptoms, but mature palms take 2 to 3 years.

Contact with living palm roots with colonized debris within the soil has been believed as a primary infection of palms by *Ganoderma* species (Idris *et al.*, 2002). Secondary spread of inoculum was assumed to be in contact with roots and roots of living palms (De Oliveira *et al.*, 2005). Palms between 7 to 15 years old are also infected, beside, the very old palms and this is the peak age of fruit production and effort should be made to control disease spread out and losses of trees. During its earliest point of entry very little information about the fungus was realized.

Unfortunately, the external symptoms become visible when the disease is too far in advanced. Thus, BSR disease seems a silent time bomb to oil palm trees.

Field controls of BSR by contact chemicals have not been very successful even in *vitro* efficacy of fungicides have been reported against *G. boninense* (Soepena *et al.*, 2000). In addition, drenching of fungicides failed to control effectively (Chung, 1990). Control by physical methods such as clean clearing and tree surgery has had but transient effects, although there is testify that BSR can be dealt if all the disease inoculum is removed before planting or replanting the crop. Besides, biological control of *Ganoderma* involves the use of antagonistic fungus, e.g. *Trichoderma* sp. and the use effective micro-organisms (Illias and Abdullah, 1999; Sariah, 2003; Siti Muslimah *et al.*, 2010). The results have been variable. To date, no commercial application of biocontrol agent is practiced. Therefore, treatment such as nutrient and plant hormone should be applied in the selected concentration at the seedling stage in order to make them resistant towards BSR disease. Thus, it is important to develop disease control methods that are good compatible with biological control, like using agents that elicit natural inducible plant defences at low concentrations.

The nutritional status of a plant has a major impact on disease susceptibility, and this has been responsible for suppressing a variety of diseases (Engelhard, 1989). Previous studies tested calcium nitrate suppressed BSR symptoms on clonal materials (Sariah and Zakaria, 2000) and it is found that copper played a significant role in organic and conventional systems for battling some fungal diseases. *Ganoderma* wilts disease caused by *G. applanatum* and *G. lucidum* in coconut trees has been controlled by copper-based fungicides (Nambiar *et al.*, 1992). Lately, salicylic acid a naturally occurring plant hormone had attention after it was found, it can induce resistance to pathogens and abiotic stress tolerance in plants (Gautam and Singh 2009; Pieterse *et al.*, 2009; Ramirez *et al.*, 2009).

Calcium, magnesium, copper, carbon, salicylic acid and nitrogen are decisive variables for plants and modifying these as a control method may keep *Ganoderma* attack. These nutrients and plant hormone could conceivably be added by soil application to control the disease at the same time utilized as fertilizers. Applied fertilizers comprise some of these minerals to control diseases and to improve the development of the oil palm. The existing literature is silent on controlling *G. boninense* although the uses of soil amendments have been suggested by Sariah and Zakaria (2000). Due to the increasing economic impact of this disease in Malaysia, effective and viable management strategies need to be established. In this research, first, an experiment about in *vitro* and block study, followed by glass house trials and eventually access the plant secondary metabolites. Hence the hypothesis of this study is; nutrients and plant hormone have the potential to suppress BSR development in oil palm seedlings.

This research is conducted with the following objectives:

1. To assess *in vitro* effect of salicylic acid, calcium and copper ions on growth and sporulation of *Ganoderma boninense*.
2. To investigate the induction of systemic acquired resistance (SAR) of oil palm seedlings after being treated with calcium, copper ions and salicylic acid.
3. To assess the plant secondary metabolites in oil palm seedlings after being treated with calcium, copper ions and salicylic acid.





## REFERENCES

- Abdullah, M.A., Alzate, O., Mohammad, M., McNall, R.J., Adang, M.J. and Dean, D.H. (2003). *Applied Environment of Microbiology*. 69: 5343-5353.
- Adaskaveg, J.E., Gilbertson, R.L., Blanchette, R.A. (1990). Comparative studies of delignification caused by *Ganoderma* species. *Applied Environment of Microbiology*. 56: 1932-1943.
- Adaskaveg, J.E., Blanchette, R.A., Gilbertson, R.L. (1991). Decay of date palm wood by white rot and brown rot fungi. *Canadian Journal of Botany*. 69: 615-629.
- Agarwal, S., Sairam, R.K., Srivastava, G.C., Tyagi, A. and Meena, R.C. (2005). Role of ABA, salicylic acid, calcium and hydrogen peroxide on antioxidant enzymes induction in wheat seedlings. *Plant Sciences*. 169: 559-570.
- Agrios, G.N. (2005). *Plant Pathology* (5th edition). San Diego: Elsevier-Academic Press.
- Aide, T.M. (1993). Patterns of leaf development and herbivory in a tropical understory community. *Ecology*. 74: 455-466.
- Ali, Y., Aslam, Z., Ashraf, M.Y. and Tahir, G.R. (2004). Effect of salinity on chlorophyll concentration, leaf area, yield and yield components of rice genotypes grown under saline environment. *International Journal of Environmental Science and Technology*. 1: 221-225.
- Ariffin, D., Idris, A.S. and Marzuki, A. In *Development of a Technique to Screen Oil Palm Seedlings for Resistance to Ganoderma*. Proceedings of the 1995 PORIM National Oil palm Conference. Jalani B.S., Ariffin D., Rajanaidu N., Tayeb M.D., Paranjothy K. and Basri M.W. (Eds.); Palm Oil Research Institute of Malaysia: Kuala Lumpur, 1995.
- Ariffin, D., Idris, A.S. and Singh, G. (2000). Status of *Ganoderma* on oil palm. In J. Flood. *Ganoderma Disease of Perennial Crops* (pp. 44-68). Wallingford: UK CAB International Publishing.
- Atkinson, D. and McKinlay, R.G. (1997). Crop protection and its integration within sustainable farming systems. *Agriculture Ecosystem and Environment Journal*. 64: 87-93.
- Baker, D.E. and Senef, J.P. (1995). Copper. In B.J. Alloway. *Heavy metals in soils* (pp. 179-205). London: Blackie Academic and Professional.
- Benjamin, J.G. (1993). Tillage effects on near-surface soil hydraulic properties. *Soil Tillage Residue*. 26: 277-288.
- Benjamin, M. and Chee, K.H. (1995). Basal stem rot of oil palm- a serious problem on inland soils. *MAPPS Newsletter*. 19(1): 3.

- Boerjan, W., Ralph, J., Baucher, M. (2003). Lignin biosynthesis. *Annual Review Plant Biology*. 54: 519-546.
- Breton, F., Hasan Y., Hariadi, Lubis Z. and de Franqueville H. In *Characterization of parameters for the development of an early screening test for Basal Stem Rot tolerance in oil palm progenies*. Proceedings of Agriculture, Biotechnology and Sustainability Conference. Technological Breakthroughs and Commercialization - The Way Forward, PIPOC 2005 MPOB International Palm Oil Congress, Sept. 25-29, 2005. Kuala Lumpur: Malaysia, 2005.
- Campbell, C.L. and Madden, L.V. (1990). *Introduction to plant disease epidemiology*. USA: John Wiley and Sons.
- Carpin, S., Crevecoeur, M., de Meyer, M., Simon, P., Greppin, H. and Penel, C. (2001). Identification of a  $\text{Ca}^{2+}$ -pectate binding site on an apoplastic peroxidase. *Plant Cell*. 13: 511-520.
- Carpita, N.C. and Gibeaut, D.M. (1993). Structural models of primary-cell walls in flowering plants - consistency of molecular-structure with the physical-properties of the walls during growth. *Plant Journal*. 3: 1-30.
- Carrillo-González, R., Šimůnek, J., Sauvé, S. and Adriano, D. (2006). Mechanisms and pathways of trace elements in soils. *Advanced Agronomy*. 91: 111-178.
- Castañeda, P. and Pérez, L.M. (1996). Calcium ions promote the response of Citrus lemon against fungal elicitors or wounding. *Phytochemistry*. 42: 595-598.
- Chen, Z., Silva, H. and Klessig, D.F. (1993). Active oxygen species in the induction of plant systemic acquired resistance by salicylic acid. *Science*. 262: 1883-1886.
- Chen, Z., Iyer, S., Caplan, A., Klessig, D.F. and Fan, B. (1997). Differential accumulation of salicylic acid and salicylic acid-sensitive catalase in different rice tissues. *Plant Physiology*. 114: 193-201.
- Chen, E. L., Chen, Y. A., Chen, L. M. and Liu, Z. H. (2002). Effect of copper on peroxidase activity and lignin content in *Raphanus sativus*. *Plant Physiology Biochemistry*. 40: 439-444.
- Chung, G F. Preliminary results on trunk injection of fungicides against *Ganoderma* basal stem rot in oil palm. Proceedings of *Ganoderma* Workshop, Palm Oil Research Institute of Malaysia. Kuala Lumpur: Malaysia, 1990.
- Clarkson, D.T. and Sanderson, J. (1969). The uptake of a polyvalent cation and its distribution in root apices of *Allium cepa*: tracer and autoradiographic studies. *Planta*. 89: 136-154.
- Clarkson, D.T. and Sanderson, J. (1974). The endodermis and its development in barley roots as related to radial migration of ions and water. In J. Kolek. *Structure and function of primary root tissues* (pp. 87-100). Bratislavia: Veda Publishing House of the Slovak Academy of Sciences.



- Cochard, H., Damour, G., Bodet, C., Tharwat, I., Poirier, M. and Améglio, T. (2005). Evaluation of a new centrifuge technique for rapid generation of xylem vulnerability curves. *Physiology of Plant*. 124: 410–418.
- Coley, P.D. (1983). Herbivory and defensive characteristics of tree species in a lowland tropical forest. *Ecological monographs*. 53: 209-233.
- Corina V.A., Dempsey D.A. and Klessig D.F. (2009). Salicylic acid, a multifaceted hormone to combat disease. *Annual Review of Phytopathology*. 47: 177-206.
- Corley, R.H.V and Tinker, P.B. (2003). *The Oil Palm* (Fourth Edition). (pp. 27-28; 507-413). Great Britain: Blackwell Publishing.
- Cox, V.S., Wallace, L.J. and Jessen, C.R. (1978). An anatomic and genetic study of canine cryptorchidism. *Teratology*. 18: 233-240.
- Dat, J.F., Lopez-Delgado, H., Foyer, C.H. and Scott, I.M. (1998). Parallel changes in  $H_2O_2$  and catalase during thermotolerance induced by salicylic acid or heat acclimation in mustard seedlings. *Plant Physiology*. 116: 1351-1357.
- De Oliveira, F.G.R., Candian, M., Lucchette, F.F. (2005). A technical note on the relationship between ultrasonic velocity and moisture content of Brazilian hardwood (*Goupia glabra*). *Building and Environment*. 40: 297-300.
- Delaney, T.P. (2004). Salicylic Acid. In J.P. Davies. *Plant Hormones: Biosynthesis, Signal Transduction Action*. (pp. 635-653). Dordrecht: Kluwer Academic Publishers.
- Dicko, M.H., Gruppen, H., Zouzouho, O.C., Traoré, A.S., Van Berkel, W.J.H. and Voragen, A.G.J. (2006). Effects of germination on the activities of amylases and phenolic enzymes in sorghum varieties grouped according to food end-use properties. *Journal Science of Food and Agriculture*. 86: 953-963.
- Dietz, K.J., Baier, M. and Kramer, U. (1999). Free radicals and reactive oxygen species as mediators of heavy metal toxicity in plants. In M.N.V. Prasad and J. Hagemeyer. *Heavy metal stress in plants: from molecules to ecosystems* (pp. 73-97). Berlin: Springer-Verlag.
- Dikin, A., Kamaruzaman, S., Zainal Abidin, M.A. and Idris, A.S. (2003). Biological Control of Seedborne Pathogen of Oil Palm, *Schizophyllum commune* Fr. with Antagonistic Bacteria. *International Journal of Agriculture and Biology*. 5(4): 507-512.
- El-Tayeb, M.A. (2005). Response of barley grains to the interactive effect of salinity and salicylic acid. *Plant Growth Regulation*. 45: 215-224.
- Elisashvili, V., Kachlishvili E. and Penninckx, M. (2008). Effect of growth substrate, method of fermentation, and nitrogen source on lignocellulose-degrading enzymes production by white-rot Basidiomycetes. *Journal of Industrial Microbiology and Biotechnology*. 35: 1531-1538.
- Engelhard, A.W. (1989). *Soilborne plant pathogens: Management of diseases with macro- and microelements*. St. Paul Minn: APS Press.



- Espelie, K.E. and Kolattukudy, P.E. (1986). Immunocytochemical localization and time course appearance of an anionic peroxidase associated with suberization in wound healing potato tuber tissue. *Plant Physiology*. 87: 487.
- Evans, I., Solberg, E. and Huber, D. M. (2007). Copper and Plant Disease. In L.E. Datnoff, W.H. Elmer and D.M. Huber. *Mineral Nutrition and Plant Disease* (pp. 177-188). USA: The American Phytopathological Society.
- Fang, W.C. and Kao, C.H. (2000). Enhanced peroxidase activity in rice leaves in response to excess iron, copper and zinc. *Plant Science*. 158(1-2): 71-76.
- Ferguson, I.B. and Bollard, E.G. (1976). The movement of calcium in woody stems. *Annals of Botany*. 40: 1057-1065.
- Flood, J. (2006). A review of *Fusarium* wilts of the oil palm caused by *Fusarium oxysporum* f. sp. *elaeidis*. *Phytopathology*. 96 (6): 660-662.
- Fodor, J., Gullner, G., Adam, A.L., Barna, B., Komives, T. and Kiraly, Z. (1997). Local and systemic responses of antioxidants to tobacco mosaic virus infection and to salicylic acid in tobacco. *Plant Physiology*. 114: 1443-1451.
- Ganesan, V. and Thomas, G. (2001). Salicylic acid response in rice: influence of salicylic acid on H<sub>2</sub>O<sub>2</sub> accumulation and oxidative stress. *Plant Science*. 160: 1095-1106.
- Gautam, S. and Singh, P.K. (2009). Salicylic acid induced salinity tolerance in corn grown under NaCl stress. *Acta Physiologiae Plantarum*. 31: 1185-1190.
- George, S.T., Chung, G.F. and Zakaria, K. In *Updated results (1990-1995) on trunk injection of fungicides for the control of Ganoderma basal stem rot*. Proceedings of the 1996 PORIM International Palm Oil Congress – Agriculture. Ariffin A., Mohn Basri D., Rajanaidu W., Mohd Tayeb N., Paranjothy D., Cheah K., Chang C.S. and Ravigadevi S. (Eds.); Kuala Lumpur, 1996.
- George, S.T., Chung, G.F. and Zakaria, K. (2000). Benefits of soil mounding tall palms in a high *Ganoderma* incidence areas in Lower Perak. In E. Pushparajah. *Plantation Tree Crops in the New Millenium* (pp. 565-576). Kuala Lumpur: The Incorporated Society of Planters.
- Graham, R.D. and Webb, M.J. (1991). Micronutrients and disease resistance and tolerance in plants. In J.J. Mortvedt, F.R. Cox, L.M. Shuman, and R.M. Welch. *Micronutrients in Agriculture* (2nd Edition) (pp. 329-370). USA: Soil Science Society of America.
- Gross, G.G. (1980). The biochemistry of lignifications. *Advances in Botanical Research*. 8: 25.
- Hall, J.L. (2002). Cellular mechanisms for heavy metal detoxification and tolerance. *Journal of Experimental Botany*. 53: 1-11.
- Hammerschmidt, R., Nuckles, E.M. and Kuc, J. (1982). Association of enhanced peroxidase activity with induced systemic resistance of cucumber to *Colletotrichum lagenarium*. *Physiology and Plant Pathology*. 20: 73-82.

- Harada, H., Cote, W.A. Jr. (1985). Structure of wood. In T. Higuchi. *Biosynthesis and biodegradation of wood components* (pp. 1-42). Orlando, FL: Academic Press.
- Harfouche, A.L., Rugini, E., Mencarelli, F., Botondi, R. and Muleo, R. (2008). Salicylic acid induces H<sub>2</sub>O<sub>2</sub> production and endochitinase gene expression but not ethylene biosynthesis in *Castanea sativa* *in vitro* model system. *Journal of Plant Physiology*. 165: 734-744.
- Hartley, C.W.S. (1988). *The Oil Palm* (3rd Edition). New York: John Wiley and Sons.
- Hayat, S. and Ahmad, A. (2007). Salicylic Acid: A Plant Hormone. *Springer*, UK. ISBN-13: 978-1-4020-5183-8.
- He, Y., Liu, Y., Cao, W., Huai, M., Xu, B. and Huang, B. (2005). Effects of salicylic acid on heat tolerance associated with antioxidant metabolism in Kentucky Blue grass. *Crop Science*. 45: 988-995.
- Higinbotham, N., Etherton, B. and Foster, R.J. (1967). Mineral ion contents and cell transmembrane electropotentials of pea and oat seedling tissue. *Plant Physiology*. 42: 37-46.
- Ho, Y.W. and Nawawi, A. (1985) *Ganoderma boninense* from basal stem rot in oil palm (*Elaeis guineensis*) in West Malaysia. *Pertanika Journal of Tropical Agricultural Science*. 8(3): 425-428.
- Ho, Y. W. and Khairuddin, H. (1995). Pathogenicity and histopathology of *Ganoderma boninense* on oil palm seedlings. *Journal of Bioscience*. 6: 155-164.
- Ho, C.T. and Khairudin, H. (1997) Usefulness of soil mounding treatments in prolonging productivity of prime-aged *Ganoderma* infected palms. *The Planter*. 73(854): 239-244.
- Huber, D.M. and Graham, R.D. (1999). The role of nutrition in crop resistance and tolerance to diseases. In Z. Rengel. *Mineral Nutrition of Crops: Fundamental Mechanisms and Implications* (pp. 169-204). London: Food Products Press.
- Idris, A.S., Ismail, S., Arifin, D. and Ahmad, H. (2002). Control of *Ganoderma* infected palm—development of pressure injection and field applications. MPOB Information Series MPOB TT No.148, No.131, Malaysia.
- Idris, A.S., Yamaoka, M., Hayakawa, S., Basri, I., Noorhashimah and Ariffin, D. (2003). PCR technique for detection of *Ganoderma*. MPOB Information Series MPOB TT No. 188, Malaysia.
- Idris, A.S. and Ariffin, D. In *Basal stem rot- biology, detection and control*. Proceedings of Invited paper presented at the International Conference on 'Pests and Disease of Importance to the Oil Palm Industry', May. 18-19, 2004. Kuala Lumpur: MPOB, Bangi, 2004.



- Idris, A.S., Ismail, S. And Ariffin, D. (2004a). Prolonging the productive life of *Ganoderma*-infected palms with hexacanozole. MPOB Information Series MPOB TT No.214, Malaysia.
- Idris, A.S., Kushairi, M.A., Ismail, S. And Ariffin, D. (2004b). Selection for partial resistance in oil palm progenies to *Ganoderma* basal stem rot. *Journal of Oil Palm Research*. 16: 12-18.
- Idris, A.S., Rajinder, S., Madihah, A.Z. and Wahid, M.B. (2010). Multiplex PCR-DNA kit for early detection and identification of *Ganoderma* species in oil palm. MPOB Information Series MPOB TT No.531, Malaysia.
- Illias, G.N.M. (2000). *Trichoderma* and its efficacy as a bio-control agent of basal stem rot of oil palm (*Elaeis guineensis* Jacq.). Doctoral dissertation. Universiti Putra Malaysia, Malaysia.
- Illias, G.M.N. and Abdullah, F. In *Effect of culture filtrates of Trichoderma harzianum and T. virens against Ganoderma boninense*. Proceedings of the 5th International Conference on Plant Protection in the Tropics, March 15-18, 1999. Kuala Lumpur, Malaysia, 1999.
- Jacobson, L., Hannapel, R.J., Moore, D.P. and Schaedle, M. (1961). Influence of calcium on selectivity of ion absorption process. *Plant Physiology*. 36: 58-61.
- Janda, T., Horvath, E., Szalai, G. and Paldi, E. (2007). Role of salicylic acid in the induction of abiotic stress tolerance. In S. Hayat, A. Ahmad. *Salicylic acid: A plant hormone* (pp. 91-150). Dordrecht, The Netherlands: Springer.
- Kauss, H. and Jeblick, W. (1995). Pretreatment of parsley suspension cultures with salicylic acid enhances spontaneous and elicited production of H<sub>2</sub>O<sub>2</sub>. *Plant Physiology*. 108: 1171-1178.
- Keltjens, W.G. (1981). Absorption and transport of nutrient cations and anions in maize roots. *Plant and Soil*. 63(1): 39-46.
- Khairudin, H. (1990). Basal Stem rot of oil palm: Incidence, etiology and control. Unpublished Master dissertation, Universiti Pertanian Malaysia, Malaysia.
- Khairudin, H. In *Basal stem rot of oil palm caused by Ganoderma boninense*. Proceedings of the 1993 PORIM International Palm Oil Congress. Jalani S. (Eds.); Palm Oil Research Institute of Malaysia: Bangi, Malaysia, 1993.
- Khairuddin, H. and Chong, T.C. (2008). An overview of the current status of *Ganoderma* basal stem rot and its management in a large plantation group in Malaysia. *The Planter*. 84: 469-482.
- Khodary, S.E.A. (2004). Effect of salicylic acid on the growth, photosynthesis and carbohydrate metabolism in salt stressed maize plants. *International Journal of Agriculture and Biology*. 6(1): 5-8.
- Kochian, L.V. (1991). Mechanisms of micronutrient uptake and translocation in plants. In J.J. Mortvedt. *Micronutrients in Agriculture* (pp. 251-270). Madison, WI: Soil Science Society of America.

- Lopez-Delgado, H., Dat, J.F., Foyer, C.H. and Scott, I.M. (1998). Induction of thermotolerance in potato microplants by acetylsalicylic acid and H<sub>2</sub>O<sub>2</sub>. *Journal of Experimental Botany*. 49: 713-720.
- Lautner, S., Ehrling, B., Windeisen, E., Rennenberg, H., Matyssek, R. and Fromm, J. (2007). Calcium nutrition has a significant influence on wood formation in poplar. *New Phytologist*. 173: 743-752.
- Lim, K.H. and Udin, W. *Management of Ganoderma in peat soil in Indonesia*. Paper presented in 2010 Second International Seminar Oil Palm Diseases, Advances in *Ganoderma* Research and Management. Sheraton Hotel - Yogyakarta, Indonesia. May, 2010.
- Lin, C. C., Chen, L. M., and Liu, Z.H. (2005). Rapid effect of copper on lignin biosynthesis in soybean root. *Plant Science*. 168: 855-861.
- Macklon, A.E.S. (1975). Corticol cell fluxes and transport to the stele in excised root segments of *Allium cepa* L. Calcium as affected by its external concentration. *Planta*. 152: 381-387.
- Malamy, J. and Klessing, D.F. (1992). Salicylic acid and plant disease resistance. *The Plant Journal*. 2: 643-654.
- Marscher, H. (1995). *Mineral nutrition of higher plants*. (2nd edition) (pp. 889). San Diego: Academic Press.
- Meister, A. (1992). On the antioxidant effects of ascorbic acid and glutathione. *Biochemistry and Pharmacology*. 44: 1905-1915.
- Møller, S.G. and McPherson, M.J. (1998). Developmental expression and biochemical analysis of the *Arabidopsis* *ataol* gene encoding an H<sub>2</sub>O<sub>2</sub> generating diamine oxidase. *Plant Journal*. 13: 781-791.
- Muller, C.H. (1969). The "co"- in coevolution. *Science*. 164: 197-198.
- Nambiar, K.K.N., Rethinam, P. and Varghese, M. (1992). Management of *Ganoderma* wilt disease of coconut in Kerala. *Indian Coconut Journal*. 22(10): 6-9.
- Nicholson, R.L. and Hammerschmidt, R. (1992). Phenolic compounds and their role in disease resistance. *Annual Review of Phytopathology*. 30: 369-389.
- Nie, X. (2006). Salicylic acid suppresses Potato virus Y isolate N: O induced symptoms in tobacco plants. *Phytopathology*. 96: 255-263.
- Nur Ain Izzati, M.Z. and Abdullah, F. (2008). Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*. 44: 101-107.
- Nur Sabrina, A.A., Sariah, M. and Zaharah, A.R. (2012). *Effects of calcium and copper on lignin biosynthesis and suppression of Ganoderma boninense infection in oil palm seedlings*. Unpublished master dissertation. Universiti Putra Malaysia, Malaysia.



- Nurrashyeda, R., Idris, A.S., Madihah, A.Z., Ramle, M. and Kushairi, A. (2011). *Hendersonia* GanoEF1 granules for the control of *G. boninense* in oil palm. MPOB Information MPOB TT Series No. 556, Malaysia.
- Oborn, I., Edwards, A.C., Witter, E., Oenema, O., Ivarsson, K., Withers, P.J.A., Nilsson, S.I. and Richert Stinzing, A. (2003). Element balances as a toll for sustainable nutrient management: a critical appraisal of their merits and limitations within an agronomic and environmental context. *European Journal of Agronomy*. 20: 211-225.
- Paterson, R.R.M., Sariah, M., Zainal Abidina M.A. and Lima, N. (2008). Prospects for inhibition of lignin degrading enzymes to control *Ganoderma* white rot of oil palm. *Current Enzyme Inhibition*. 4: 172-179.
- Pei, Z.M., Murata, Y., Benning, G., Thomine, S., Klusener, B., Allen, G.J., Grill, E. and Schroeder, J.I. (2000). Calcium channels activated by hydrogen peroxide mediate abscisic acid signalling in guard cells. *Nature*. 406: 731-734.
- Penel, C. and Greppin, H. (1996). Pectin binding proteins: Characterization of the binding and comparison with heparin. *Plant Physiology and Biochemistry*. 34: 479-488.
- Pieterse, C.M.J., Reyes, A.L., Ent, S.V.D. and Wees, S.C.M.V. (2009). Networking by small molecule hormones in plant immunity. *Nature Chemical Biology*. 5(5): 308-316.
- Plomion, C., Leprovost, G. and Stokes, A. (2001). Wood formation in trees. *Plant Physiology*. 127: 1513-1523.
- Raiskila S, Pulkkinen M, Laakso T, Fagerstedt K, Löija M, Mahlberg R, Paaanen L, Ritschkoff A-C, Saranpää P. (2007). FTIR spectroscopic prediction of Klason and acid soluble lignin variation in Norway spruce cutting clones. *Silva Fennica*. 41: 351-371.
- Ramirez, A.A. et al. (2009). Evidence for a role of gibberellins in salicylic acid-modulated early plant responses to abiotic stress in *Arabidopsis* seeds. *Plant Physiology*. 150: 1335-1344.
- Rao, M.V., Paliyaht, G., Ormrod, D.P., Murr, D.P. and Watkins, C.B. (1997). Influence of salicylic acid on H<sub>2</sub>O<sub>2</sub> production, oxidative stress, and H<sub>2</sub>O<sub>2</sub>-metabolizing enzymes. *Plant Physiology*. 115: 137-149.
- Rao, V., Lim, C.C., Chia, C.C. and Teo, K.W. (2003). Studies on *Ganoderma* spread and control. *The Planter*. 79: 367-83.
- Raskin, J. (1992). "Role of salicylic acid in plants". Annually. Review Plant Physiology. *Plant molecular Biology*. 43: 439-463.
- Rees, R. (2006). *Ganoderma stem rot of oil palm (Elaeis guineensis); mode of infection, epidemiology and biological control*. Doctoral dissertation, University of Bath, United Kingdom.
- Rees, R.W., Flood, J., Hasan, Y. and Cooper, R.M. (2007). Low soil temperature and root inoculum contact enhance *Ganoderma* infection of oil palm; implications

for late disease appearance in plantations and screening for disease resistance. *Plant Pathology*. 56: 862-870.

- Rees, R.W., Flood, J., Hasan, Y. and Cooper, R.M. (2009). Basal stem rot infection of oil palm; mode of root and lower stem invasion by *Ganoderma*. *Plant Pathology*. 58: 982-989.
- Robson, A.D., Hartley, R.D. and Jarvis, S.C. (1981). Effect of copper deficiency on phenolic and other constituents of wheat cell walls. *New Phytologist*. 89: 361-371.
- Rosenthal, G.A. and Berenbaum, M.R. (1991). *Herbivores: Their Interactions with secondary plant metabolites*. (2nd edition volume 1). San Diego: Academic Press.
- Rosenthal, G.A., Berenbaum, M.R. (1992). *Herbivores: Their Interactions with secondary plant metabolites*. (2nd edition volume 2). San Diego: Academic Press.
- Ruch, R.J., Cheng, S.J. and Klaunig, J.E. (1989). Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea. *Journal of Carcinogenesis*. 10: 1003-1008.
- Salmén, L., Olsson, A.M., Stevanic, J.S., Simonovic, J. and Radotic, K. (2012). Organisation of wood polymers. *Bioresources*. 7(1): 521-532.
- Sanderson, F.R. (2005). An insight into dispersal of *Ganoderma boninense* on oil palm. *Mycopathologia*. 159: 139-141.
- Sariah, M. (2003). The potential of biological management of basal stem rot of oil palm: issues, challenges and constraints. *Oil Palm Bulletin*. 47: 1-5.
- Sariah, M., Hussin, M.Z., Miller, R.N.G. and Holderness, M. (1994). Pathogenicity of *Ganoderma boninense* tested by inoculation of oil palm seedlings. *Plant Pathology*. 43: 507-10.
- Sariah, M., Joseph, H. and Zakaria, H. (1997). Suppression of basal stem rot (BSR) of oil palm seedlings by calcium nitrate. *The Planter*. 73: 359- 360.
- Sariah, M., Zakaria, H., Hendry, J., Shanji, G.T. and Chung, G.F. The potential use of soil amendments for the suppression of basal stem rots of oil palm seedlings. In *Second Workshop on Ganoderma Diseases of Perennial Crops*, Wallingford: UK CAB International Publishing. 1998.
- Sariah, M. and Zakaria, H. The use of soil amendments for the control of basal stem rot of oil-palm seedlings. In *Ganoderma Diseases of Perennial Crops*. (pp.89-99). Flood J., Bridge P.D., Holderness M. (Eds.); Wallingford: UK CAB International Publishing, 2000.
- Sauvé, S., McBride, M.B., Norvell, W.A. and Hendershot, W.H. (1997). Copper solubility and speciation of *in situ* contaminated soils: effects of copper level, pH and organic matter. *Water, Air and Soil Pollution*. 100: 133-149.



- Schafer, Z.T., Grassian, A.R. and Song, L. (2009). Antioxidant and oncogene rescue of metabolic defects caused by loss of matrix attachment. *Nature*. 461: 109-13.
- Shirasu, K., Nakajima, H., Rajasekhar, V.K., Dixon, R.A. and Lamb, C. (1997). Salicylic acid potentiates an agonist-dependent gain control that amplifies pathogen signals in the activation of defence mechanisms. *Plant Cell*. 9: 261-270.
- Sime Darby. Sustainability practices-Malaysia palm oil industry. Malaysia-Romania Palm Oil Trade Fair and Seminar 2012 (POTS), Malaysia, Sept. 17-19, 2012. Bucharest: Romania, 2012.
- Singh, G. *Ganoderma*- The scourge of oil palm in the coastal areas. In *Proceedings of the Ganoderma Workshop*. (pp.7-35). Ariffin D. and Jalani S. (Eds.); Palm Oil Research Institute of Malaysia, Bangi, Malaysia, 1990.
- Singh, G. (1991). *Ganoderma*- the scourge of oil palm in the coastal areas. *The Planter*. 67: 421-444.
- Singh, B. and Usha, K. (2003). Salicylic acid induced physiological and biochemical changes in wheat seedlings under water stress. *Plant Growth Regulation*. 39: 137-141.
- Siti Muslimah, W., Susanto, A. and Djoyobisono. Beneficial soil microbes to suppress *Ganoderma boninense* in oil palm plantations: concept and applications. Paper presented in 2010 Second International Seminar on Oil Palm Diseases: Advances in *Ganoderma* Research and Management. Sheraton Hotel- Yogyakarta, Indonesia, May 31, 2010.
- Skidmore, A.M. and Dickinson, C.H. (1976). Colony interactions and hyphal interference between *Septoria Nodorum* and phylloplane fungi. *Transactions of the British Mycological Society*. 66: 57-64.
- Smith, M. and Thurnston, C.F. (1997). Fungal laccases: Role in delignification and possible industrial application. In A. Messerschmidt. *Multi-Copper Oxidases* (pp. 253-259). Singapore, New Jersey, London, Hong Kong: World Scientific.
- Soepena, H., Purba, R.Y. and Pawirosukarto, S. (2000). A control strategy for basal stem rot (*Ganoderma*) on oil palm. In J. Flood, P.D. Bridge and M. Holderness (pp. 83-88). UK: CAB International.
- Susanto, A. Basal stem rot in Indonesia. In *Biology, economic importance, epidemiology, detection and control*. Proceedings of International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases, Kuala Lumpur Convention Centre, Malaysia. Universiti Putra Malaysia Press, 2009.
- Tasgin, E., Atici, O. and Nalbantoglu, B. (2003). Effects of salicylic acid and cold on freezing tolerance in winter wheat leaves. *Plant Growth Regulation*. 41: 231-236.
- Teh, C.L., Tey, C.C., Azlina, Z. and Hamdan, I. Integrated Pest Management in a Plantation Group - Sime Darby Experience. Handout of presentation slides during 8th ISP National Seminar 2010, June 21-23, 2010. Sabah: Shangrila Tanjung Aru Resort & Spa, Kota Kinabalu, Malaysia, 2010.

- Tey, C.C and Mohd Ahdly, A. In *Mitigating measures against Ganoderma basal stem rot of oil palm*. PIPOC 2007 International Palm Oil Congress of Agricultural Biotechnology and Sustain Conference - Vol 2 (pp.866-880). Ahmad Kushairi Din. (Eds.); Kuala Lumpur: Malaysian Palm Oil Board, Malaysia, 2007.
- Thompson, A. (1931). Stem-rot of the oil palm in Malaysia. Bulletin Department of Agriculture, Straits Settlements and F.M.S., Sciences Series, 6.
- Tohirrudin, L., Tandiono, J., Abner J Silalahi, Prabowo, N.E and Foster, H.L. (2010). *Journal of Oil Palm Research*. 22: 869-877.
- Turner, P.D. (1981). *Oil Palm Diseases and Disorders*. Oxford: Oxford University Press.
- Van Etten, H., Temporini, E. and Wasmann, C. (2001). Phytoalexin (and phytoanticipin) tolerance as a virulence trait: why is it not required by all pathogens? *Physiological and Molecular Plant Pathology*. 59: 83-93.
- Van Loon, L.C. and Bakker, P.A.H.M. (2004). Signalling in rhizobacteria-plant interactions. In: De Kroon, J. and E.J.W. Visser (eds.), *Ecological Studies. Journal of Root Ecology*. 168: 287-330.
- Vance, C.P., Kink, I.K. and Sherwood, R.T. (1980) Lignification as a mechanism of disease resistance. *Annual Review of Phytopathology*. 18: 259-288.
- Walter, M. H. (1992). Regulation of lignification in defense. In T. Boller and F. Meins. *Plant Gene Research: Genes involved in Plant defense* (pp. 327-352). Vienna: Springer.
- Waterhouse, A. (2005). Determination of total phenolics. In R.E. Wrolstad. *Handbook of Food Analytical Chemistry*. Hoboken, NJ: John Wiley and Sons.
- Willats, W.G.T., McCartney, L., Mackey, W., and Knox, J.P. (2001). Pectin, cell biology and prospects for functional analysis. *Plant Molecular Biology*. 47: 9-27.
- Yalpani, N., Leon, J., M.A. and Raskin I. (1993). "Pathway of salicylic biosynthesis in healthy and virus-inoculated tobacco". *Plant Physiology*. 103: 315-321.
- Yang, V.W. and Clausen, C.A. (2007). Antifungal effect of essential oils on southern yellow pine. *International Biodegradation and Biodeterioration*. 59: 302-306.
- Zaitun, S., S. Meon and Z.A.M.Ahmad, 2008. Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture and Biology*. 10: 127-132.
- Zeyen, R.J., Carver, T.L.W. and Lyngkjaer, M.F. (2002). Epidermal cell papillae. In R.R. Belanger and W.R. Bushnell. *The powdery mildews: a comprehensive treatise* (pp. 107-125). St Paul, Minnesota, USA: APS Press.